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**Phytoplankton Community Structure  
in Northeastern Coastal Waters  
of the United States.**

**II. November 1978**

U.S. DEPARTMENT OF COMMERCE  
National Oceanic and Atmospheric Administration  
National Marine Fisheries Service  
Northeast Fisheries Center  
Woods Hole, Massachusetts

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## NOAA Technical Memorandum NMFS-F/NEC-9

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# Phytoplankton Community Structure in Northeastern Coastal Waters of the United States. II. November 1978

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## ABSTRACT

The phytoplankton populations observed in coastal waters between Narragansett Bay and the Gulf of Maine during a November 1978 cruise are described and discussed. Diatoms (43%) and dinophyceans (40%) composed the majority of the total species. Diatoms and nannoplankton were more dominant at near-shore stations; Leptocylindrus danicus and Nitzschia pungens were found in high concentrations over Georges Bank. A total of 248 phytoplankton species was observed.

## CONTENTS

	<u>Page</u>
INTRODUCTION .....	1
METHODS .....	1
RESULTS AND DISCUSSION .....	2
ACKNOWLEDGEMENTS .....	11
LITERATURE CITED .....	12
FOOTNOTES .....	14

## FIGURES

1. Phytoplankton community structure station locations for cruise BEL-78-04 .....	15
2. Salinity (ppt) and temperature (°C) determinations for cruise BEL-78-04 .....	16
3. Some phytoplankters identified in areas of high chlorophyll a determined by Christine Evans, NMFS .....	17

## TABLES

1. Various observations and measurements for stations on <u>cruise Belogorsk 78-04</u> .....	18
2. Phytoplankton compositions observed at near and far shore stations off New England coastal waters between Rhode Island and Maine in November 1978. Numbers refer to average station concentrations in cells/liter .....	19

## APPENDIX I

Concentrations of phytoplankton by station .....	25
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## INTRODUCTION

This is the second in a series of reports on phytoplankton community structure for northeastern United States coastal and continental shelf waters. A total of 33 surface samples for phytoplankton analysis was collected during the cooperative MARMAP research program aboard the Soviet research vessel Belogorsk (cruise 78-04) between 15-30 November 1978. The vessel occupied standard stations located over Georges Bank, in the Gulf of Maine, and Nantucket Shoals-southern New England waters (Fig. 1).

The first report in the series discussed the phytoplankton populations for October 1978, from Delaware Bay to the Gulf of Maine. Following papers will discuss data obtained from six subsequent cruises of National Oceanic and Atmospheric Administration (NOAA) vessels made in March, May, June, August and December 1979 and February 1980. This intensive, long-term, cooperative study is designed to investigate phytoplankton dynamics in the area during a 17 month period, assessing the standing stock, identifying seasonal norms of distribution and noting dominant species of particular regions. The series of reports will delineate seasonal and geographic changes in the phytoplankton and will lead to the establishment of a phytoplankton composition base for these waters.

## METHODS

Coordinates for all stations, with observations on cloud cover, wind direction and speed, wave height and sea surface temperatures are listed in Table 1. The near-shore stations are defined as those within 35 km of the shore, and those beyond this distance as the far-shore stations.

Phytoplankton collection and analysis procedures used in this study were those previously reported by Marshall and Cohn (1981). The study was made in association with other investigators from the National Marine Fisheries Service concerned with chlorophyll concentrations<sup>3</sup>, primary production<sup>4</sup> and nutrient analysis<sup>5</sup> to provide a synoptic overview.

#### RESULTS AND DISCUSSION

A total of 248 phytoplankters was identified on this cruise, with representation from the Bacillariophyceae (109), Pyrrhophyceae (100), Haptophyceae (12), Cyanophyceae (4), Chrysophyceae (6), Cryptophyceae (6), Chlorophyceae (2), Euglenophyceae (4), Xanthophyceae (1) and Prasinophyceae (4). A species list is given in Table 2 with average concentrations for species given at near- and far-shore stations. The composition and concentration for the phytoplankters at each station are given in Appendix I.

The Bacillariophyceans and the Pyrrhophyceans (dinophyceans) composed the majority of the total species (43 and 40 percent respectively) noted in the samples, with the diatoms having the highest concentrations of cells. In contrast to the areal dominance exhibited by Skeletonema costatum at stations in the previous month during the Belogorsk 78-03 cruise (Marshall and Cohn, 1981), other diatoms and Nannochloris atomus were the more dominant species at near-shore stations, with Leptocylindrus danicus and Nitzschia pungens found in high concentrations over Georges Bank. Since this cruise did not occupy the stations off New Jersey and New York shores, month-to-month population comparisons for this region is not possible. The most noticeable change in the dominant species from October to November

was the general absence of a Skeletonema costatum pulse in the waters sampled and its replacement as a dominant species by several other phytoplankters. Increased significantly from the October cruise were the concentrations of the chlorophycean nanoplankter, Nannochloris atomus. This species was the most abundant phytoplankter found during the cruise, with near- and far-shore station averages being 11,673 and 1,776 cells per liter respectively. Its presence was more characteristic at stations closest to shore; numbers diminished rapidly seaward. The high counts of N. atomus for the far-shore stations given in Table 2 are more characteristic of the far-shore stations closest to the 35 km cut-off line than at stations farther out over the shelf.

Several diatoms were also co-dominants at the near-shore stations. These included Coscinodiscus lineatus (111 cells/l), Guinarida flaccida (106 cells/l), Rhizosolenia imbricata (130 cells/l), Rhizosolenia stolterfothii (106 cells/l), Thalassionema notzschiooides (434 cells/l), Thalassiosira nordenskioldii (442 cells/l), Thalassiosira rotula (109 cells/l), and Thalassiothrix frauenfeldii (533 cells/l). A general pattern of decreased concentrations seaward occurred for diatoms abundant at near-shore stations. Total cell numbers were also considerably higher at station #146 (79,500 cells/l) over Georges Bank in comparison to surrounding stations. Here the diatom Leptocylindrus danicus was dominant among 32 species found at the station. This station and the adjacent station #147 (36 species) were represented by numerous diatom species, with a general representation of neritic and oceanic forms. Several of the diatoms that had distinct higher average concentrations over the shelf than at the near-shore stations were

Chaetoceros decipiens, Corethron criophilum, Coscinodiscus radiatus, Guinardia flaccida, Leptocylindrus danicus, Nitzschia pungens, Rhizosolenia alata, and Rhizosolenia styliformis.

Pyrrophyceans common to both near- and far-shore stations included Ceratium fusus, Ceratium lineatum, Ceratium tripos, Ceratium tripos var. atlanticum, Gymnodinium dissimile, Prorocentrum micans, and Protoperidinium cerasus. Prorocentrum micans was the dominant dinophycean at both near- and far-shore stations, having average counts at these stations of 187 and 459 cells/l, respectively. Sixty-two of the 100 species in this group were noted only at the near-shore stations. Although the pyrrhophyceans were generally widespread over the near-shore waters, their concentrations were basically low. The coccolithophore most typical of the samples was Emiliania huxleyi, which was found widely distributed, but in low concentrations. A greater variety of coccolithophores was noted at near-shore stations on this cruise, with Cyclococcolithus leptoporus having the highest average concentrations for the far-shore locations. The most prominent chrysophyceans were the silicoflagellates, Dictyocha fibula and Distephanus speculum, which were numerous at far-shore stations. The cyanophyceae were not abundant, being represented by four species, all noted at far-shore stations. Most numerous was Anacystis marina. The euglenophyceans and cryptophyceans were found predominantly in the near-shore stations, with the only xanthophycean, Monodus guttula, reported at far-shore stations. The prasinophyceans were generally noted in low concentrations near-shore, with higher concentrations reported for Pyramimonas grossi at several far-shore stations.

In general, the species diversity at near-shore stations (where the phytoplankton was dominated by one species, Nannochloris atomus) was lower than

at far-shore stations. Near-shore, the values for species diversity ranged from 0.170 to 3.074, at station nos. 99 and 138. Each of these stations had 36 species, and were both within the Gulf of Maine. At station no. 99, however, Nannochloris atomus composed 97.8% of the phytoplankton composition with no other species having a count greater than 184 cells/l. In contrast, all the phytoplankton at station no. 138 had low concentrations, with the majority having 16 or less cells/l and the most abundant species having 208 cells/l. This latter type of distribution, numerous species present but in low concentrations, was more characteristic away from shore and over the far shelf. Exceptions to this pattern at far-shore stations occurred when single species reached high cell concentrations (e.g., station nos. 82, 88, 114, 146). The species diversity range for far-shore stations was 0.138 to 2.882, occurring at station nos. 88 and 116, respectively. Station no. 88 is one of the inner shelf stations south of Martha's Vineyard whereas station 116 is located beyond the shelf break, southwest of Georges Bank. Low species diversity often accompanies high concentrations of cells, usually during a pulse period for a single species, and indicates a high productivity potential for that station. However, high concentrations of cells will not always be accompanied by low diversity values, especially when multiple dominants are present. The highest cell concentrations during this cruise were found at stations directly beyond Narragansett Bay (station nos. 77 and 78), where cell counts were 73,000 and 145,492 cells/l, respectively. The only other station where cell counts were comparable to these was over Georges Bank (station no. 146) where there were 79,500 cells/l.

During October, high concentrations of Skeletonema costatum were found at near-shore stations between Cape Henlopen, Delaware and the coastal

waters of Maine. Other small sized diatoms were codominant in these near-shore waters. They included Leptocylindrus danicans, Asterionella glacialis, Chaetoceros simplex, and Rhizosolenia delicatula. Cell concentrations exceeded several million cells/l at some of the near-shore stations. Nannochloris atomus was a common nanoplankter, showing large concentrations along the New York-New Jersey coastline.

Since the October 1978 cruise, there has been a marked change in the dominant species off southern New England to the Gulf of Maine and over Georges Bank. The November 1978 cruise indicates large concentrations of Nannochloris atomus at several of the near-shore stations with the general loss of Skeletonema costatum as a major constituent of the phytoplankton populations. The largest development of Nannochloris atomus was off the Narragansett Bay area, with its presence noted along the coastal areas into the Gulf of Maine. Other common phytoplankters for this month include Coscinodiscus lineatus, Guinardia flaccida, Rhizosolenia imbricata, Rhizosolenia stolterfothii, Thalasionema nitzschiodes, Thalassiosira nordenskioldii, Thalassiosira rotula, and Thalassiothrix frauenfeldii. In general, the overall cell concentrations were lower than in the previous month. The two areas with greatest cell concentrations were found off Narragansett Bay and at a station over Georges Bank. In both of these areas small sized cells (nanoplankters) were most abundant. Concentrations of cells were generally higher near-shore, with lower counts over the shelf. However, patchy areas of low cell concentrations were found at both near- and far-shore locations.

The phytoplankton for this period is characterized as predominantly diatoms and pyrrhophyceans with the chlorophycean Nannochloris atomus

prominent at specific near-shore stations. Codominants were common, with the majority of samples having low concentrations of cells, moderate to high levels of species diversity, and with scattered stations where one or several species had high cell concentrations. Patchiness also occurred at several of the near-shore stations (e.g., station nos. 98 and 99) and over Georges Bank, with the potential for significant shifts in species development evident in comparison to October dominants. The use of average counts for the numerous stations over the near- and far-shore area (as presented in Table 2) is intended to reduce some of the problems associated with evaluating the various concentration levels in these waters. These values should be considered in relation to the actual locations of higher and lower regions of productivity that are being identified over the completed study period.

Due to their high concentrations and wide distribution in the samples, more comment is necessary regarding the significance of the ultraplankton components, such as Nannochloris atomus. In recent years, the ultraplankton constituents in both estuarine and marine waters have received increased attention (McCarthy et al., 1974; Waterbury et al., 1979; Johnson and Sieburth, 1979; Marshall, 1981a). In most cases these cells have been identified as either chlorophycean, haptophycean, or cyanophycean (Cyanobacterium) species. Their small size (<10 microns) and lack of distinct morphological characteristics often have led to misidentification or simply their placement in an unidentified category. However, the importance of this group in estuaries has been emphasized by McCarthy et al. (1974). In a two year study they found the ultraplankters "responsible for a substantial fraction of both the phytoplankton biomass and phytoplankton productivity in the Chesapeake Bay."

For this period they attributed 89.6% of phytoplankton productivity to the size fraction that passes through a 35 um mesh net. The ultraplankton constituents within Chesapeake Bay and coastal marine waters include representation by a wide variety of taxonomic groups. These include the bacillariophyceae, pyrrhophyceae, cyanophyceae, chlorophyceae, haptophyceae, chrysophyceae, xanthophyceae, and prasinophyceae (Marshall, 1980). In waters of the northeastern continental shelf the most commonly mentioned ultraplankton component has been the coccolithophores (haptophyceae) and specific diatoms (Hulbert, 1963, 1970). Hulbert (1970) further mentions the importance of several chlorophyceans (Chlorella, Selenastrum, Nannochloris) in estuaries. He generalizes ocean and estuary species as characteristically solitary and globular species, smaller in diameter than coastal species, sinking more slowly than species found along the coast and in the ocean. Large concentrations of these ultraplankton have been reported along the northeast coast. O'Reilly et al. (1976) noted that Nannochloris atomus was responsible for most of the nanoplankton reduction in the lower New York Bay estuary, with the nanoplankton (<20) outproducing the netplankton by a factor of 3.7:1. This study, as others in the New York Bay and near-shore areas (Patten, 1961; Hulbert, 1963; Malone, 1977), indicates distinct seasonal variations in the dominance of the ultraplankton component, with it being more prominent during summer months. However, this pattern does not exclude major population fluctuations of this group throughout the year over areas of the shelf or at specific stations (Marshall, 1981b).

During the October and November cruises discussed, as well as those reported by Marshall and Cohn (1981), high concentrations of ultraplankton species have been reported at many of the stations closest to the shore, with numbers decreasing

seaward. A patchiness in ultraplankton coastal distribution has also been evident, both along the coast and over transects directed seaward. It is speculated that this pattern is characteristic of the area, which is itself subject to fluctuating environmental conditions of both short and long duration. The background flora, larger in size, remains stable, being composed of forms less responsive or slower in growth response time to environmental changes, and is generally classified as more characteristic of the seasonal and regional assemblages for this area. The ultraplankton component responds rapidly to fluctuations of the milieu, often reaching very high concentrations. O'Reilly et al. (1976) discuss the rapid response and high growth potential for Nannochloris atomus in nutrient rich areas of the Raritan-Lower Hudson estuary. Such reactions occur in relation to fluctuating nutrient levels and separate specific environmental requirements and growth regulatory conditions. They are further augmented by the nature of the regional shelf area where there is a dynamic water system, influenced by various currents, wind action, upwelling, and changing seasonal conditions. The distribution and growth of phytoplankton is influenced by these factors differently throughout the year. The result of the changing milieu includes the high concentrations of the ultraplankton component and its sporadic appearance throughout the shelf area, with greatest development at near-shore stations, i.e., stations receiving the nutrient rich effluents of the estuaries. An unknown factor is the influence of grazers on this system and a full understanding of the seasonal periodicity of numbers.

Future reports in this series will present a more seasonal evaluation of the phytoplankton observed in the northeastern coastal waters. The

importance of the ultraplankton component will be further addressed in relation to the total phytoplankton flora for this region.

#### ACKNOWLEDGMENTS

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#### FOOTNOTES

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<sup>4</sup>O'Reilly, J. E. and D. Busch. 1979. Summary of measurements of primary productivity made during MARMAP surveys (Belogorsk 78-01, 78-03, 78-04).

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<sup>5</sup>Draxler, A. F. J., R. Waldhauer, and A. Matte. 1979. Nutrient data from Belogorsk cruise 78-04. Report No. SHL 79-07, Sandy Hook Laboratory, Highlands, N. J. 16 p.

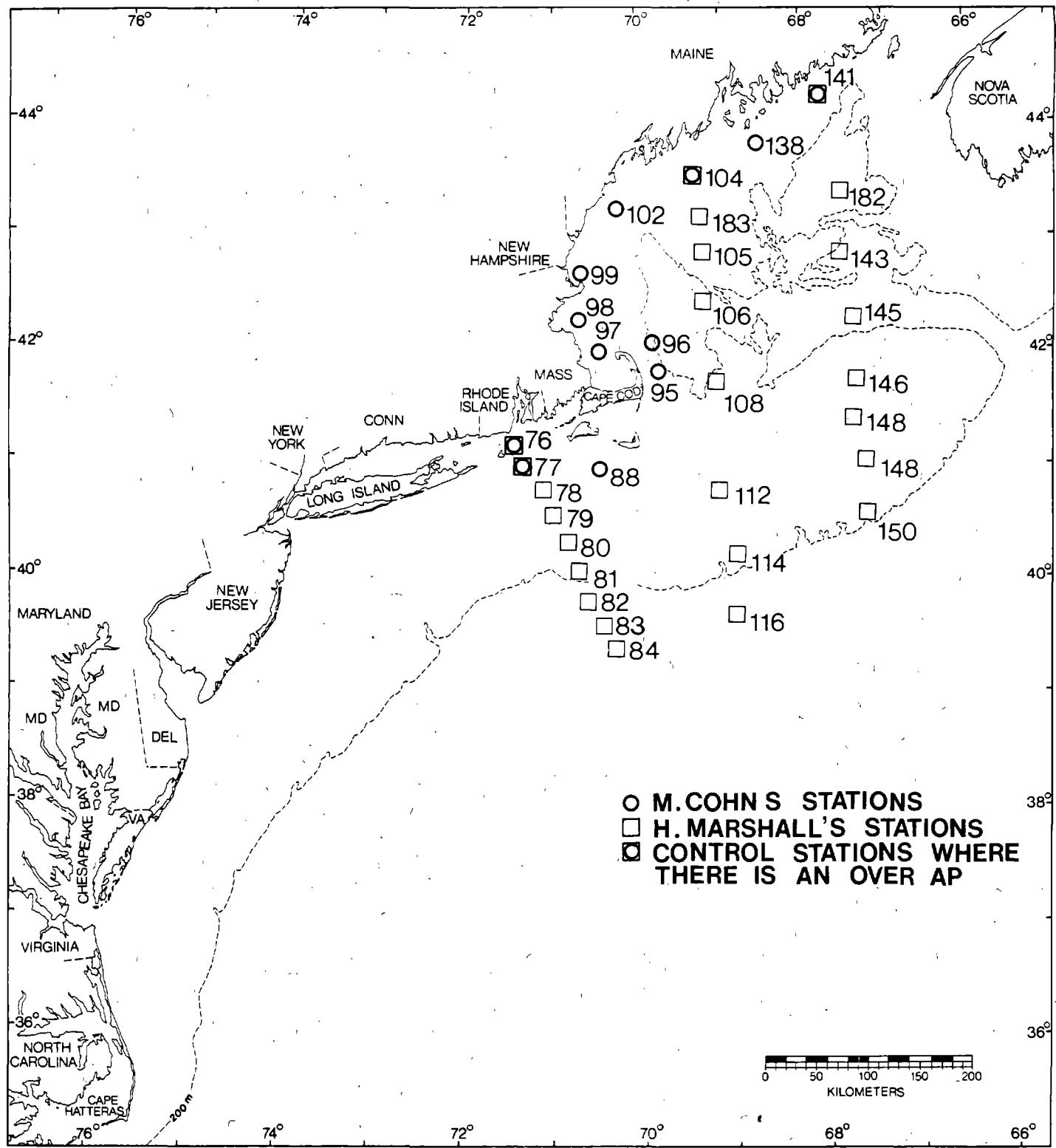


Figure 1. Phytoplankton community structure station locations for cruise BEL-78-04.

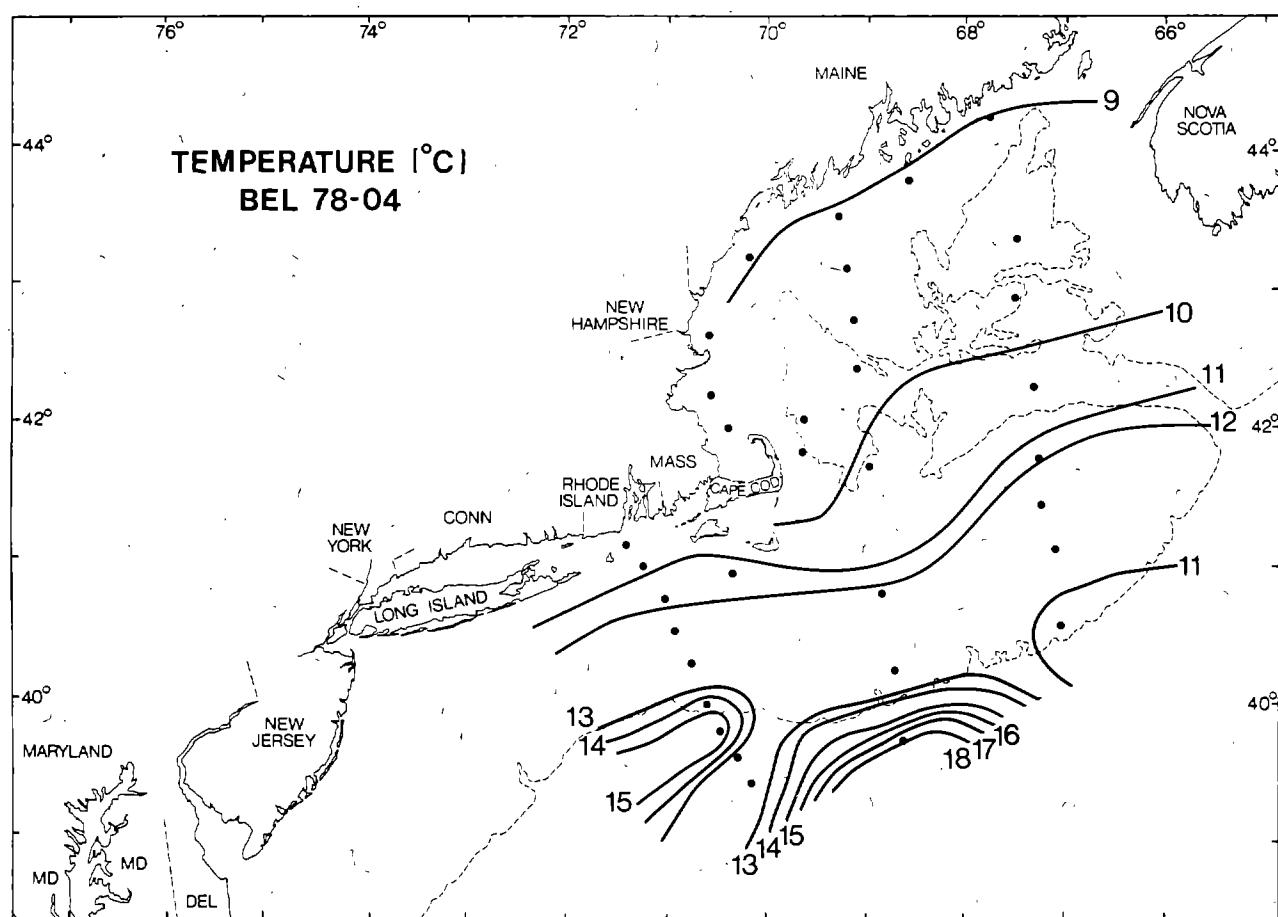
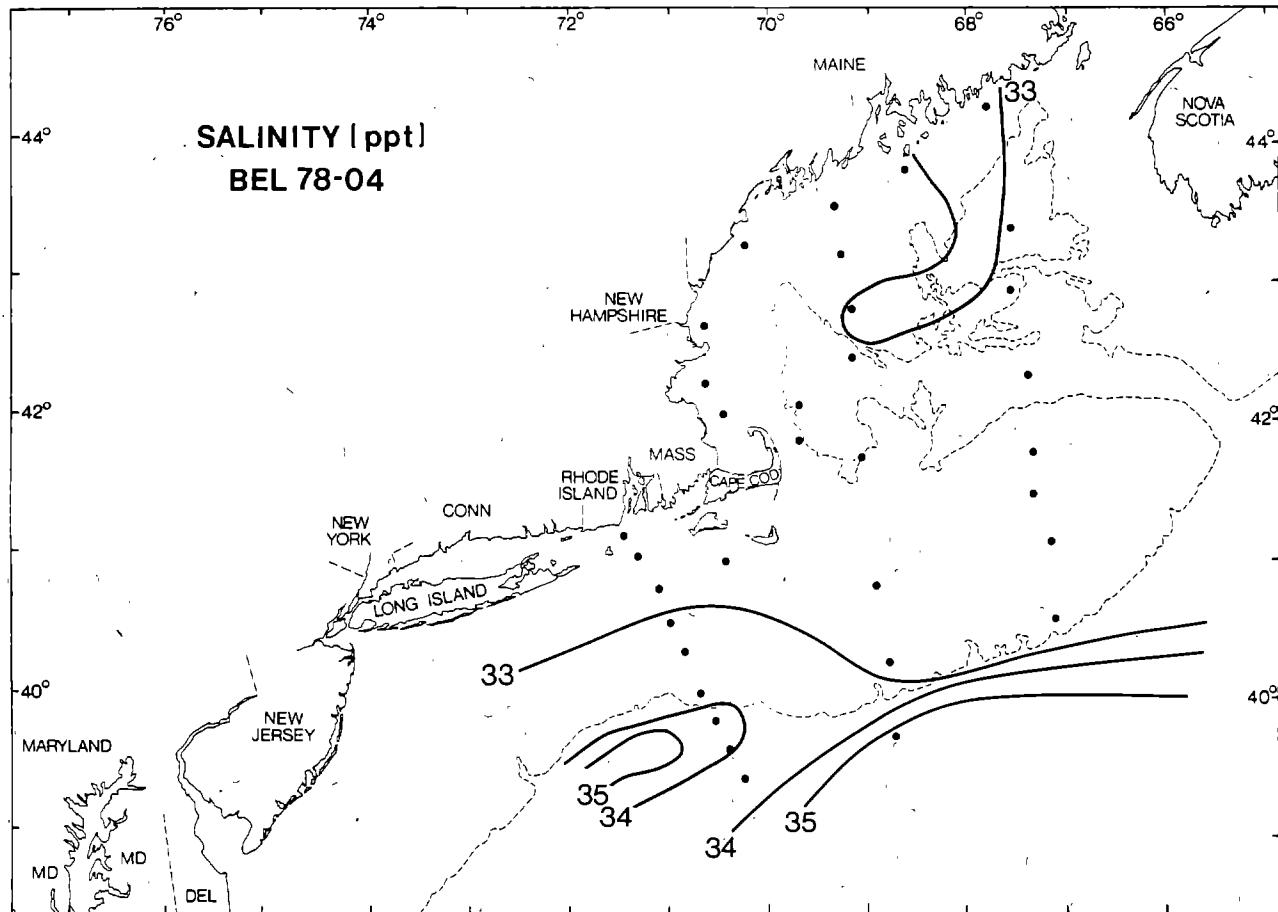


Figure 2. Salinity (ppt) and temperature ( $^{\circ}\text{C}$ ) determinations for cruise BEL-78-04.

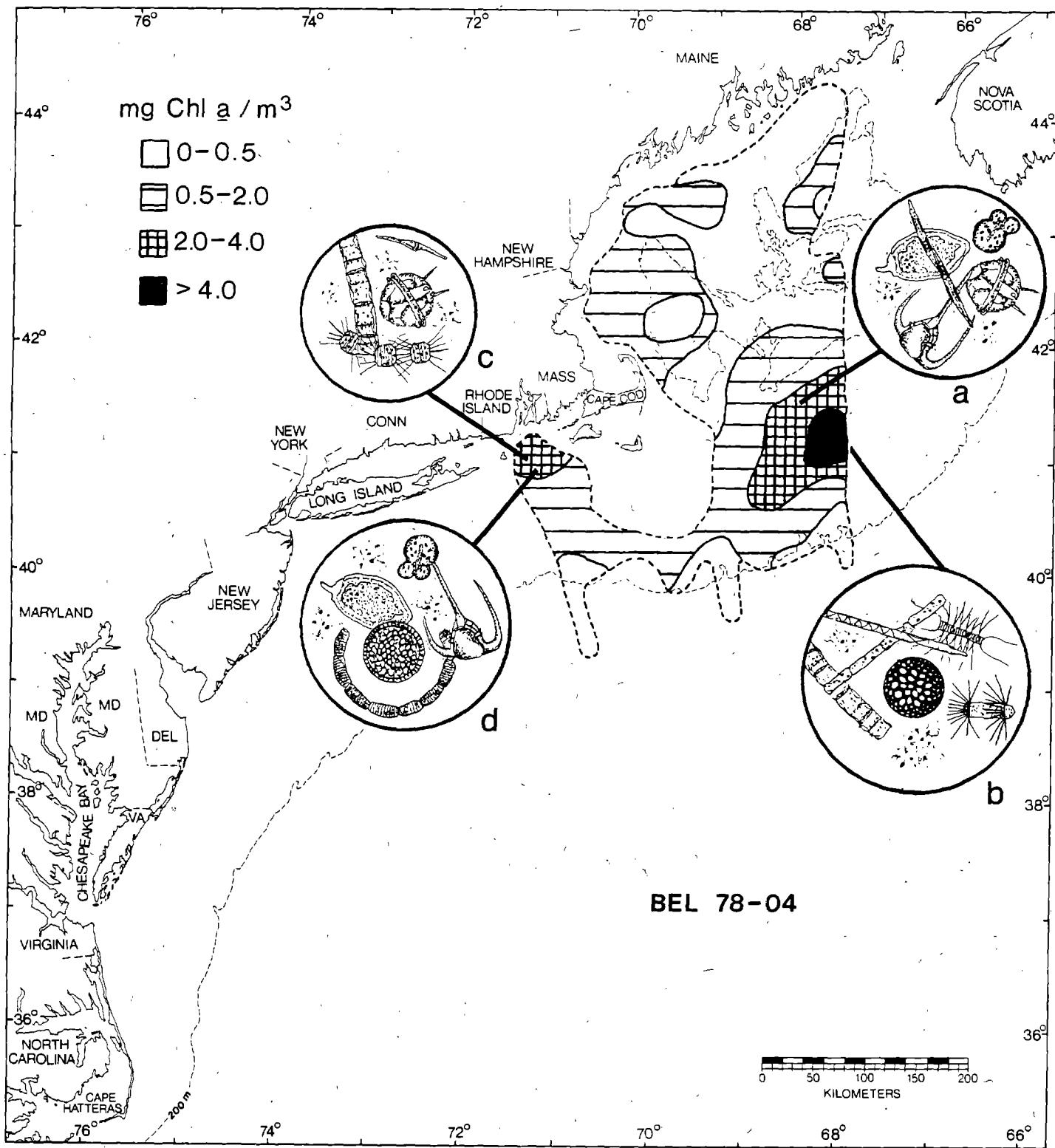


Figure 3. Some phytoplankters identified in areas of high chlorophyll a (determined by Christine Evans, NMFS): (a) Nitzschia pungens, Gymnodinium dissimile, Protoperidinium sp., Ceratium tripos, Prorocentrum micans, nanoplankton; (b) Rhizosolenia alata, R. delicatula, R. stolterfothi, Corethron criophilum, Chaetoceros decipiens, Guinardia flaccida, Coscinodiscus radiatus, nanoplankton; (c) Nitzschia pungens, Protoperidinium sp., Thalassiosira sp., nanoplankton; (d) Gymnodinium dissimile, Ceratium tripos, Rhizosolenia stolterfothi, Coscinodiscus lineatus, Prorocentrum micans, nanoplankton.

Table 1. Various data, species diversity and coordinates for stations in the Belogorsk 78-04 cruise.

S T A T I O N S

	<u>76</u>	<u>77</u>	<u>78</u>	<u>79</u>	<u>80</u>	<u>81</u>	<u>82</u>	<u>83</u>	<u>84</u>	<u>88</u>	<u>95</u>
Depth (m)	26	36	52	60	86	132	320	1600	2300	43	80
Wind speed (k)	23	24	15	15	20	27	13	7	4	13	27
Wind direction	90	15	30	245	270	280	320	46	40	110	320
Surf. temp. °C	10.68	10.78	11.30	12.8	12.4	12.6	15.5	12.1	12.5	11.08	8.85
Cloud cover	8	8	8	8	8	8	4	8	8	7	8
Wave ht. (m)	2.5	2.8	3.0	3.0	4.0	3.8	1.8	2.0	1.6	0.9	2.5
Salinity	32.36	32.51	32.70	33.6	33.5	33.5	35.0	34.0	33.6	32.44	32.51
Sp. diversity	1.485	1.449	1.993	2.478	2.291	2.757	0.427	1.133	2.337	0.138	0.322
Latitude (N)	41.19.0	41.08.8	40.57	40.40	40.21	40.10	39.59	39.46	39.37	41.02.5	41.57.4
Longitude (W)	71.19.9	71.14.9	71.11	71.02	70.51	70.46	70.40	70.36	70.30	70.31.9	69.49.8

96      97      98      99      102      104      105      106      108      112      114

Depth (m)	216	60	70	126	95	90	152	208	210	74	78
Wind speed (k)	33	21	18	12	10	11	5	16	16	17	18
Wind direction	340	330	345	60	130	100	60	30	115	50	50
Surf. temp. °C	9.10	8.97	9.02	8.54	8.80	8.86	9.14	9.02	10.15	12.10	12.13
Cloud cover	8	6	8	8	9	8	-	4	8	7	8
Wave ht. (m)	2.5	2.0	2.7	3.0	1.0	0.5	1.0	1.5	1.8	1.8	2.0
Salinity	32.73	32.33	32.21	32.24	32.68	32.90	32.05	33.01	32.42	32.56	32.75
Sp. diversity	2.420	1.722	2.615	0.170	0.394	0.579	1.509	2.610	1.951	2.172	0.773
Latitude (N)	42.15.5	42.07.0	42.26.3	42.47.6	43.24.0	43.39.4	42.58.1	42.34.8	41.54.3	40.55.7	40.24.6
Longitude (W)	69.43.3	70.21.2	70.38.0	70.32.2	70.13.3	69.22.2	69.17.4	69.13.4	69.10.2	69.05.7	69.03.4

116      138      141      143      145      146      147      148      150      182      183

Depth (m)	1900	80	70	210	228	28	29	29	75	240	158
Wind speed (k)	17	8	10	19	13	18	16	12	12	14	6
Wind direction	30	20	360	360	340	300	290	290	300	340	100
Surf. temp. °C	18.60	8.92	8.86	9.74	10.31	12.04	12.10	12.66	11.20	9.70	9.06
Cloud cover	9	4	5	8	0	0	0	1	3	4	6
Wave ht. (m)	1.5	.4	.8	1.8	2.1	1.5	1.4	1.5	1.2	1.5	0.5
Salinity	35.47	33.03	33.09	32.84	35.52	32.48	32.46	32.36	32.60	32.84	32.98
Sp. diversity	2.883	3.074	0.186	2.372	1.768	0.970	1.753	1.279	1.626	1.517	2.417
Latitude (N)	39.51.8	43.56.3	44.20.0	42.46.5	42.18.3	41.48.3	41.28.1	41.15.9	40.37.0	42.23.0	43.17.9
Longitude (W)	68.59.8	68.31.2	67.41.5	67.42.2	67.42.1	67.43.7	67.42.5	67.41.4	67.40.5	67.51.4	69.20.0

Table 2. Phytoplankton composition observed at near and far shore stations off New England coastal waters between Rhode Island and Maine in November 1978. Numbers refer to average station concentrations in numbers of cells per liter.

	Near Shore	Far Shore
<u>Bacillariophyceae</u>		
<i>Achnanthes</i> sp.	.4	---
<i>Actinoptychus senarius</i> Ehrenberg	61.6	43.8
<i>Actinoptychus splendens</i> Ralfs	2.4	---
<i>Amphora arenaria</i> Donkin	---	.4
<i>Amphora crassa</i> Gregory	13.2	---
<i>Asterionella glacialis</i> Castracane	38.4	---
<i>Asterolampra marylandica</i> Ehrenberg	.8	.1
<i>Asteromphalus flabellatus</i> (Brebisson) Greville	2.4	---
<i>Biddulphia alternans</i> (Bailey) Van Heurck	---	3.6
<i>Biddulphia aurita</i> (Lyngbye) Brebisson	---	.2
<i>Biddulphia regia</i> (Schultze) Ostenfeld	1.6	---
<i>Campylodiscus limbatus</i> Brebisson	1.2	---
<i>Cerataulina pelagica</i> (Cleve) Hendey	5.2	12.2
<i>Chaetoceros</i> sp.	4.0	3.3
<i>Chaetoceros atlanticum</i> Cleve	1.2	.8.2
<i>Chaetoceros coarctatum</i> Lauder	24.4	15.6
<i>Chaetoceros concavicornis</i> Mangin	---	3.3
<i>Chaetoceros costatum</i> Pavillard	---	.8
<i>Chaetoceros danicum</i> Cleve	2.0	2.6
<i>Chaetoceros decipiens</i> Cleve	7.5	62.8
<i>Chaetoceros pendulum</i> Karsten	---	2.2
<i>Chaetoceros peruvianum</i> Brightwell	---	.8
<i>Chaetoceros sociale</i> Lauder	---	16.4
<i>Climacodium frauenfeldianum</i> Grunow	1.2	1.6
<i>Coccconeis scutellum</i> Ehrenberg	.4	---
<i>Corethron criophilum</i> Castracane	12.6	47.0
<i>Coscinodiscus</i> sp.	24.4	34.7
<i>Coscinodiscus asteromphalus</i> Ehrenberg	---	2.0
<i>Coscinodiscus centralis</i> Ehrenberg	3.6	---
<i>Coscinodiscus grani</i> Gough	13.2	---
<i>Coscinodiscus granulosus</i> Grunow	1.2	1.6
<i>Coscinodiscus lineatus</i> Ehrenberg	111.6	18.6
<i>Coscinodiscus marginatus</i> Ehrenberg	50.4	4.2
<i>Coscinodiscus nitidus</i> Gregory	12.4	7.3
<i>Coscinodiscus noblis</i> Grunow	1.6	---
<i>Coscinodiscus oculus iridis</i> Ehrenberg	---	5.6
<i>Coscinodiscus radiatus</i> Ehrenberg	6.0	25.6
<i>Coscinodiscus stellaris</i> var. <i>symbolophora</i> (Grunow) Jorgensen	---	18.0
<i>Coscinodiscus sub-bulliens</i> Jorgensen	3.2	---
<i>Coscinodiscus wailesii</i> Gran and Angst	20.0	25.8
<i>Cyclotella caspia</i> Grunow	13.6	---
<i>Cylindrotheca closterium</i> (Ehrenberg) Reimann and Lew	18.0	14.7
<i>Dimerogramma</i> sp.	---	1.0
<i>Diploneis crabro</i> Ehrenberg	---	1.2
<i>Ditylum brightwellii</i> (West) Grunow	3.6	14.2

Table 2. (continued)

	<u>Near</u> <u>Shore</u>	<u>Far</u> <u>Shore</u>
<i>Eucampia zodiacus</i> Ehrenberg	1.2	---
<i>Guinardia flaccida</i> (Castracane) Peragallo	130.0	236.2
<i>Gyrosigma balticum</i> (Ehrenberg) Cleve	---	20.0
<i>Gyrosigma hippocampus</i> (Ehrenberg) Hassall	---	.4
<i>Hemiaulus hauckii</i> Grunow	1.2	---
<i>Hemiaulus sinensis</i> Greville	.8	---
<i>Hemidiscus cuneiformis</i> Wallich	.4	---
<i>Lauderia borealis</i> Gran	.8	13.6
<i>Leptocylindrus danicus</i> Cleve	33.6	3161.1
<i>Leptocylindrus minimus</i> Gran	13.6	43.2
<i>Licmophora flabellata</i> (Carmichael) Agardh	.4	---
<i>Licmophora paradoxa</i> var. <i>tincya</i> (Agardh) Hustedt	.4	---
<i>Melosira moniliformis</i> (Muller) Agardh	---	1.6
<i>Navicula</i> sp.	6.8	.4
<i>Navicula directa</i> (Smith) Cleve	.4	---
<i>Navicula hennedyii</i> W. Smith	2.0	---
<i>Navicula palpebralis</i> (Brebisson) Smith	2.4	---
<i>Nitzschia</i> sp.	6.4	---
<i>Nitzschia lorenziana</i> Grunow	11.6	---
<i>Nitzschia pungens</i> Grunow	4.2	569.6
<i>Nitzschia recta</i> Grunow	.8	---
<i>Nitzschia seriata</i> Cleve	29.8	---
<i>Nitzschia spathulata</i> Brebisson	.4	---
<i>Paralia sulcata</i> (Ehrenberg) Cleve	91.2	77.5
<i>Plagiogramma vanheurckii</i> Grunow	0.4	---
<i>Plagiogramma staurophorum</i> (Gregory) Heilberg	15.6	10.8
<i>Pleurosigma</i> sp.	9.2	---
<i>Pleurosigma angulatum</i> (Quekett) W. Smith	.8	16.9
<i>Pleurosigma elongatum</i> W. Smith	6.4	3.2
<i>Pleurosigma hamuliferum</i> Brun	12.8	.8
<i>Pleurosigma normanii</i> Ralfs	50.4	1.2
<i>Rhaphoneis amphiceros</i> Ehrenberg	---	5.4
<i>Rhaphoneis surirella</i> (Ehrenberg) Grunow	---	.8
<i>Rhizosolenia alata</i> Brightwell	81.8	147.1
<i>Rhizosolenia alata</i> f. <i>gracillima</i> (Cleve) Grunow	---	3.8
<i>Rhizosolenia alata</i> f. <i>indica</i> (Peragallo) Gran	.4	1.8
<i>Rhizosolenia bergenii</i> Peragallo	---	.4
<i>Rhizosolenia calcar-avis</i> Schultze	.4	.4
<i>Rhizosolenia delicatula</i> Cleve	21.6	39.2
<i>Rhizosolenia fragilissima</i> Bergon	1.6	.6
<i>Rhizosolenia hebetata</i> f. <i>hiemalis</i> Gran	65.6	---
<i>Rhizosolenia hebetata</i> f. <i>semispina</i> (Hensen) Gran	---	12.3
<i>Rhizosolenia imbricata</i> Brightwell	130.0	32.6
<i>Rhizosolenia imbricata</i> var. <i>shrubsolei</i> (Cleve) Van Heurck	3.6	---
<i>Rhizosolenia setigera</i> Brightwell	20.4	1.8
<i>Rhizosolenia stolterfothii</i> Peragallo	106.4	181.1
<i>Rhizosolenia styliformis</i> Brightwell	1.2	20.1

Table 2. (continued)

	Near Shore	Far Shore
<i>Schroederella delicatula</i> (Peragallo) Pavillard	10.4	2.0
<i>Stauroneis amphioxys</i> Gregory	.4	.8
<i>Stephanopyxis palmeriana</i> (Greville) Grunow	16.4	---
<i>Striatella unipunctata</i> (Lyngbye) Agardh	3.6	---
<i>Synedra</i> sp.	---	1.0
<i>Synedra tabulata</i> var. <i>fasciculata</i> (Lyngbye) Hustedt	---	.6
<i>Tabellaria fenestrata</i> var. <i>asterionelloides</i> Grunow	---	3.2
<i>Thalassionema nitzschiooides</i> Hustedt	434.8	103.3
<i>Thalassiosira aestivalis</i> Gran and Angst	3.6	---
<i>Thalassiosira decipiens</i> (Grunow) Jorgensen	7.6	30.0
<i>Thalassiosira delicate</i> Ostenfeld	---	.2
<i>Thalassiosira gravida</i> Cleve	45.0	84.0
<i>Thalassiosira nordenskioldii</i> Cleve	442.0	9.0
<i>Thalassiosira rotula</i> Meunier	109.2	3.3
<i>Thalassiothrix frauenfeldii</i> Grunow	533.8	115.1
<i>Triceratium favus</i> Ehrenberg	1.2	---
Unidentified pennate diatoms >20 microns	---	5.5

Dinophyceae

<i>Amphidinium</i> sp.	.4	.2
<i>Amphidinium acutissimum</i> Schiller	---	13.0
<i>Amphidinium acutum</i> Lahmann	---	.6
<i>Amphidinium carterae</i> Hulbert	.4	---
<i>Amphidinium crassum</i> Lohmann	3.6	---
<i>Amphidinium sphenoides</i> Wulff	1.6	.8
<i>Amphidinium wislouchi</i> Hulbert	.4	---
<i>Amphidoma</i> sp.	1.6	---
<i>Amphisolenia globifera</i> Stein	.4	---
<i>Ceratium bucephalum</i> Cleve	.8	---
<i>Ceratium contrarium</i> (Gourret) Pavillard	---	.4
<i>Ceratium extensum</i> (Gourret) Cleve	---	1.4
<i>Ceratium furca</i> (Ehrenberg) Claparede and Lachmann	---	2.8
<i>Ceratium fusus</i> (Ehrenberg) DuJardin	3.6	3.7
<i>Ceratium horridum</i> (Cleve) Gran	.4	---
<i>Ceratium lineatum</i> (Ehrenberg) Cleve	12.8	14.8
<i>Ceratium macroceros</i> (Ehrenberg) VanHoffen	1.2	.4
<i>Ceratium massiliense</i> (Gourret) Jorgensen	.8	---
<i>Ceratium minutum</i> Jorgensen	.8	4.8
<i>Ceratium pentagonum</i> Gourret	.4	.4
<i>Ceratium teres</i> Kofoed	.4	---
<i>Ceratium trichoceros</i> (Ehrenberg) Kofoed	---	.8
<i>Ceratium tripos</i> (Muller) Nitzsch	4.8	20.0
<i>Ceratium tripos</i> var. <i>atlanticum</i> (Ostenfeld) Paulsen	8.0	22.0
<i>Cochlodinium constrictum</i> (Schutt) Lemmerman	1.2	---
<i>Dinophysis</i> sp.	.4	---
<i>Dinophysis acuminata</i> Claparee and Lachmann	2.4	---
<i>Dinophysis acuta</i> Ehrenberg	.8	---
<i>Dinophysis fortii</i> Pavillard	---	2.8
<i>Dinophysis lachmannii</i> Paulsen	.4	---

Table 2. (continued)

	<u>Near</u> <u>Shore</u>	<u>Far</u> <u>Shore</u>
<i>Dinophysis microterygia</i> Dang	---	1.6
<i>Dinophysis norvegica</i> Claparede and Lachmann	.4	---
<i>Dinophysis ovum</i> Schutt	---	.7
<i>Dinophysis sphaerica</i> Stein	---	.5
<i>Diplopsalis lenticula</i> Bergh	.8	---
<i>Glenodinium</i> sp.	---	.1
<i>Glenodinium lenticula</i> (Bergh) Schiller	.8	---
<i>Goniaulax</i> sp.	1.2	---
<i>Goniaulax birostris</i> Stein	.4	---
<i>Goniaulax diacantha</i> (Meunier) Schiller	4.0	---
<i>Goniaulax diegensis</i> Kofoid	---	5.2
<i>Goniaulax excavata</i> (Braarud) Balech	1.6	6.4
<i>Goniaulax polyedra</i> Stein	1.2	---
<i>Goniaulax polygramma</i> Stein	.8	---
<i>Goniaulax spinifera</i> (Claparede and Lachmann) Diesing	.4	---
<i>Goniaulax unicornis</i> Lebour	.4	---
<i>Gymnodinium</i> sp.	.4	10.5
<i>Gymnodinium arcticum</i> Wulff	---	1.4
<i>Gymnodinium danicans</i> Campbell	.4	---
<i>Gymnodinium dissimile</i> Kofoid and Swezy	4.8	2.7
<i>Gymnodinium minutum</i> Hulbert	.4	---
<i>Gymnodinium nelsoni</i> Martin	7.2	---
<i>Gymnodinium simplex</i> (Lohmann) Kofoid and Swezy	1.2	---
<i>Gymnodinium splendens</i> Lebour	2.4	---
<i>Gymnodinium stellatum</i> Hulbert	4.0	---
<i>Gyrodinium</i> sp.	7.6	1.4
<i>Gyrodinium dominans</i> Hulbert	.8	---
<i>Gyrodinium estuariale</i> Hulbert	.4	---
<i>Gyrodinium fusiforme</i> Kofoid and Swezy	17.6	---
<i>Gyrodinium gloicum</i> Hulbert	1.2	---
<i>Gyrodinium metum</i> Hulbert	4.0	---
<i>Gyrodinium pellucidum</i> Wulff	4.0	---
<i>Gyrodinium spirale</i> (Bergh) Kofoid and Swezy	.8	---
<i>Gyrodinium undulans</i> Hulbert	.8	---
<i>Gyrodinium uncatenum</i> Hulbert	8.0	---
<i>Hemidinium</i> sp.	.4	---
<i>Heterocapsa triquetra</i> (Ehrenberg) Stein	17.6	.4
<i>Katodinium rotundatum</i> (Lohmann) Loeblich	21.2	---
<i>Noctiluca miliaris</i> Suriray	1.2	---
<i>Orithocercus thurni</i> (Schmidt) Kofoid and Skogsberg	---	.2
<i>Oxytoxum</i> sp.	.4	---
<i>Oxytoxum sceptrum</i> (Stein) Schroder	---	.2
<i>Oxytoxum scolopax</i> Stein	.4	1.2
<i>Oxytoxum sphaeroideum</i> Stein	1.2	---
<i>Podolampas elegans</i> Schutt	---	.1
<i>Prorocentrum</i> sp.	.8	1.3
<i>Prorocentrum aporum</i> (Schiller) Dodge	9.2	---
<i>Prorocentrum dentatum</i> Stein	---	3.8
<i>Prorocentrum maximum</i> (Gourret) Schiller	.8	---

Table 2. (continued)

	Near Shore	Far Shore
<i>Prorocentrum micans</i> Ehrenberg	187.4	459.9
<i>Prorocentrum minimum</i> (Pavillard) Schiller	8.0	---
<i>Prorocentrum rostratum</i> Stein	2.8	---
<i>Protoperidinium</i> sp.	6.4	---
<i>Protoperidinium abei</i> (Paulsen) Balech	.4	---
<i>Protoperidinium brevipes</i> (Paulsen) Balech	3.2	---
<i>Protoperidinium cerasus</i> (Paulsen) Balech	6.4	6.4
<i>Protoperidinium conicum</i> (Gran) Balech	3.2	---
<i>Protoperidinium depressione</i> (Bailey) Balech	---	.4
<i>Protoperidinium leonis</i> (Pavillard) Balech	1.6	---
<i>Protoperidinium nipponicum</i> (Abe) Balech	---	.4
<i>Protoperidinium oblongum</i> (Aurivallis) Parke and Dodge	.8	---
<i>Protoperidinium oceanicum</i> (VanHoffen) Balech	1.2	---
<i>Protoperidinium ovatum</i> Pouchet	.8	---
<i>Protoperidinium pallidum</i> (Ostenfeld) Balech	1.6	---
<i>Protoperidinium pellucidum</i> Bergh	.8	---
<i>Protoperidinium pentagonum</i> (Gran) Balech	4.8	---
<i>Protoperidinium steinii</i> (Jorgensen) Balech	---	.2
<i>Protoperidinium subinerme</i> (Paulsen) Balech	.4	---
<i>Scrippsiella trochoidea</i> (Stein) Loeblich	11.6	---
Unidentified dinoflagellate cysts	11.2	---
Unidentified dinoflagellates	---	7.8

Haptophyceae

<i>Acanthecca aculeata</i> Kamptner	.4	---
<i>Coccolithus pelagicus</i> (Wallich) Schiller	3.6	---
<i>Cyclococcolithus leptoporus</i> (Murray and Blackman) Kamptner	.4	426.5
<i>Discosphaera tubifer</i> (Murray and Blackman) Ostenfeld	.4	.4
<i>Emiliania huxleyi</i> (Lohmann) Hay and Mohler	22.8	21.2
<i>Helicosphaera carteri</i> (Wallich) Kamptner	1.6	1.6
<i>Hymenomonas carterae</i> (Braarud and Fagerland) Braarud	4.8	---
<i>Hymenomonas roseola</i> Stein	3.2	12.0
<i>Pontosphaera syracusana</i> Lohmann	.4	---
<i>Rhabdosphaera claviger</i> Murray and Blackman	.4	---
<i>Scyphosphaera apsteinii</i> Lohmann	.8	---
<i>Syracosphaera pulchra</i> Lohmann	3.2	---
Unidentified coccolithophores	16.4	3.2

Table 2. (continued)

	<u>Near</u> <u>Shore</u>	<u>Far</u> <u>Shore</u>
<u>Chrysophyceae</u>		
<i>Calycomonas ovalis</i> Wulff	2.0	---
<i>Dictyocha fibula</i> Ehrenberg	19.8	91.0
<i>Distephanus speculum</i> (Ehrenberg) Haeckel	29.2	93.9
<i>Ebria tripartita</i> (Schumann) Lemmermann	2.4	---
<i>Mallomonas</i> sp.	2.4	---
<i>Olisthodiscus luteus</i> Carter	5.2	1.6
<u>Cyanophyceae</u>		
<i>Anacyclis marina</i> (Hansg) Drouet and Daily	---	180.3
<i>Nostoc commune</i> Vaucher	---	.2
<i>Oscillatoria erythraea</i> (Ehrenberg) Kutz	---	.8
<i>Oscillatoria submembranacea</i> Ardisson and Strafforel	---	2.3
<u>Euglenophyceae</u>		
<i>Euglena</i> sp.	4.8	.8
<i>Euglena acus</i> Ehrenberg	.4	---
<i>Eutreptia marina</i> Cunha	.4	---
<i>Eutreptia viridis</i> Perty	7.6	---
<u>Chlorophyceae</u>		
<i>Nannochloris atomus</i> Butcher	11673.2	1776.0
<i>Staurastrum leptocladum</i> var. <i>insidiae</i> West and West	---	.6
<u>Cryptophyceae</u>		
<i>Chilomonas marina</i> Ehrenberg	8.4	1.6
<i>Chroomonas amphioxidea</i> (Conrad) Butcher	.8	---
<i>Chroomonas salina</i> (Wislouch) Butcher	.8	---
<i>Chroomonas vectensis</i> Carter	9.2	.4
<i>Cryptomonas stigmatica</i> Wislouch	8.4	---
<i>Rhodomonas amphioxidea</i>	2.0	---
<u>Xanthophyceae</u>		
<i>Monodus guttula</i> Pascher	---	426.4
<u>Prasinophyceae</u>		
<i>Bipedinomonis pyriformis</i> Carter	.4	---
<i>Pyramimonas grossii</i> Parke	---	1047.4
<i>Pyramimonas micron</i> Conrad and Kufferath	.8	2.0
<i>Pyramimonas obovata</i> Carter	.4	---

Appendix I. Concentrations of phytoplankton observed at stations from the Belogorsk 78-04 cruise.

<u>Station 76</u>	<u>Cells/ Liter</u>	<u>Cells/ Liter</u>
<i>Nannochloris atomus</i>	54000	<i>Ditylum brightwellii</i> 40
<i>Dictyocha fibula</i>	144	<i>Asterionella glacialis</i> 768
<i>Distephanus speculum</i>	172	<i>Thalassiothrix frauenfeldii</i> 152
<i>Ebria tripartita</i>	24	<i>Thalassionema nitzschioides</i> 3048
<i>Olisthodiscus luteus</i>	24	<i>Licmophora flabellata</i> 8
<i>Acanthoica aculeata</i>	8	<i>Licmophora paradoxa v. tincta</i> 8
<i>Syracospaera pulchra</i>	64	<i>Striatella unipunctata</i> 40
Unidentified coccolithophorids	128	<i>Plagiogramma staurophorum</i> 88
<i>Emiliania huxleyi</i>	80	<i>Cocconeis scutellum</i> 8
<i>Discosphaera tubifera</i>	8	<i>Navicula</i> sp. 56
<i>Helicosphaera carteri</i>	16	<i>Navicula palpebralis</i> 40
<i>Paralia sulcata</i>	964	<i>Stauroneis amphioxis</i> 8
<i>Stephanopyxis palmeriana</i>	328	<i>Pleurosigma</i> sp. 184
<i>Corethron criophilum</i>	72	<i>Pleurosigma angulatum</i> 16
<i>Leptocylindrus danicus</i>	672	<i>Pleurosigma normani</i> 600
<i>Leptocylindrus minimus</i>	272	<i>Amphora crassa</i> 192
<i>Cyclotella caspia</i>	152	<i>Nitzschia</i> sp. 80
<i>Thalassiosira decipiens</i>	56	<i>Nitzschia pungens</i> 84
<i>Thalassiosira gravida</i>	636	<i>Nitzschia seriata</i> 400
<i>Thalassiosira nordenskioldii</i>	1760	<i>Nitzschia spathulata</i> 8
<i>Coscinodiscus</i> sp.	104	<i>Cylindrotheca closterium</i> 304
<i>Coscinodiscus centralis</i>	48	<i>Campylodiscus</i> sp. 24
<i>Coscinodiscus grani</i>	240	<i>Prorocentrum micans</i> 136
<i>Coscinodiscus marginatus</i>	800	<i>Prorocentrum apora</i> 64
<i>Coscinodiscus nitidus</i>	40	<i>Dinophysitis acuminata</i> 16
<i>Coscinodiscus radiatus</i>	20	Unidentified dinoflagellates 8
<i>Coscinodiscus wailesii</i>	32	<i>Amphidinium crassum</i> 56
<i>Coscinodiscus granulosus</i>	24	<i>Gymnodinium splendens</i> 48
<i>Coscinodiscus sub-bulliens</i>	40	<i>Gyrodinium uncatenum</i> 24
<i>Actinptychus senarius</i>	464	<i>Katodinium rotundatum</i> 24
<i>Asteromphalus flabellatus</i>	32	<i>Noctiluca miniaris</i> 8
<i>Hemidiscus cuneiformis</i>	8	<i>Diplopsalis lenticula</i> 16
<i>Triceratium favus</i>	24	<i>Glenodinium lenticula</i> 8
<i>Eucampia zodiacus</i>	24	<i>Heterocapsa triquetra</i> 56
<i>Hemiaulus sinensis</i>	16	<i>Protoperidinium</i> sp. 80
<i>Chaetoceros</i> sp.	8	<i>Protoperidinium pellucidum</i> 16
<i>Chaetoceros atlanticum</i>	24	<i>Scripsiella trochoidea</i> 16
<i>Chaetoceros coarctatum</i>	200	<i>Protoperidinium oceanicum</i> 24
<i>Rhizosolenia alata</i>	1344	<i>Protoperidinium oblongum</i> 16
<i>Rhizosolenia alata f. indica</i>	8	<i>Gonyaulax</i> sp. 24
<i>Rhizosolenia calcar-avis</i>	8	<i>Gonyaulax diacantha</i> 8
<i>Rhizosolenia delicatula</i>	248	<i>Gonyaulax polygramma</i> 16
<i>Rhizosolenia fragilissima</i>	24	<i>Gonyaulax birostris</i> 8
<i>Rhizosolenia hebetata f. hemiaulus</i>	120	<i>Ceratium fusus</i> 8
<i>Rhizosolenia imbricata</i>	360	<i>Ceratium lineatum</i> 28
<i>Rhizosolenia imbricata v. shrubsolei</i>	72	<i>Ceratium macroceros v. gallum</i> 24
<i>Rhizosolenia setigera</i>	184	<i>Ceratium pentagonum</i> 8
<i>Rhizosolenia stolterfothii</i>	1152	<i>Ceratium tripos</i> 8
<i>Rhizosolenia styliformis</i>	24	<i>Ceratium massiliense</i> 8
<i>Guinardia flaccida</i>	700	<i>Ceratium minutum</i> 16

	<u>Cells/ Liter</u>		<u>Cells/ Liter</u>
Chilomonas mariana	40	Pleurosigma elongatum	128
Chroomonas vectensis	128	Pleurosigma hamuliferum	256
Cryptomonas stigmatica	56	Pleurosigma normani	392
Rhodomonas amphioxieia	40	Amphora crassa	72
 <u>Station 77</u>		Nitzschia sp.	40
Nannochloris atomus	101024	Nitzschia seriata	140
Mallomonas sp.	48	Euglenophyceae euglenales Euglena sp.	96
Dictyocha fibula	252	Unidentified dinoflagellate cysts	224
Distephanus speculum	388	Prorocentrum micans	3580
Ebria tripartita	16	Prorocentrum maximum	16
Pontosphaera huxleyi	8	Dinophysis norvegica	8
Unidentified coccolithophorids	112	Dinophysis lachmanii	8
Coccolithus pelagicus	8	Amphidinium crassum	16
Emiliania huxleyii	40	Cochlodinium constrictum	16
Cyclococcolithus leptoporus	8	Gymnodinium dissimile	96
Rhabdosphaera claviger	8	Gyrodinium sp.	16
Paralia sulcata	860	Gyrodinium fusiforme	352
Corethron criophilum	132	Heterocapsa triquetra	16
Cyclotella caspia	104	Protoperidinium sp.	24
Thalassiosira aestivalis	72	Scripsiella trochoidea	16
Thalassiosira decipiens	96	Gonyaulax diacantha	8
Thalassiosira gravida	264	Ceratium fusus	24
Thalassiosira nordenskioldii	7048	Ceratium lineatum	212
Thalassiosira rotula	2184	Ceratium tripos var. atlanticum	160
Lauderia annulata	16	Ceratium teres	8
Coscinodiscus sp.	384	Chilomonas marina	64
Coscinodiscus grani	24	Chroomonas vectensis	8
Coscinodiscus lineatus	2208	Cryptomonas stigmatica	32
Coscinodiscus marginatus	112		
Coscinodiscus nitidus	160	Dictyocha fibula	232
Coscinodiscus radiatus	100	Distephanus speculum	392
Coscinodiscus wailesii	152	Corethron criophilum	32
Coscinodiscus sub-bulliens	8	Thalassiosira gravida	1400
Actinptychus splendens	48	Coscinodiscus sp.	64
Actinptychus senarius	768	Coscinodiscus nitidus	16
Asteromphalus flabellatus	16	Coscinodiscus radiatus	176
Asterolampra marylandica	16	Coscinodiscus granulosus	32
Hemiaulus hauckii	16	Actinptychus senarius	160
Chaetoceros coarctatum	288	Rhizosolenia alata	8
Chaetoceros danicum	40	Rhizosolenia alata f. gracillima	32
Rhizosolenia alata	260	Rhizosolenia delicatula	8
Rhizosolenia hebetata f. hemiaulus	1192	Rhizosolenia setigera	24
Rhizosolenia imbricata	2240	Rhizosolenia stolterfothii	1288
Rhizosolenia setigera	176	Guinardia flaccida	56
Rhizosolenia stolterfothii	352	Synedra sp.	8
Guinardia flaccida	1892	Thalassiothrix frauenfeldii	1456
Thalassiothrix frauenfeldii	10496	Plagiogramma staurophorum	112
Thalassionema nitzschiooides	5624	Gyrosigma balticum	400
Plagiogramma staurophorum	224	Nitzschia pungens	24
Navicula sp.	72	Nitzschia lorenziana	232

	<u>Cells/ Liter</u>		<u>Cells/ Liter</u>
<i>Prorocentrum micans</i>	4760	<i>Guinardia flaccida</i>	121
<i>Dinophysis fortii</i>	16	<i>Thalassiothrix frauenfeldii</i>	5
<i>Ceratium lineatum</i>	24	<i>Thalassionema nitzschiooides</i>	36
<i>Ceratium tripos var. atlanticum</i>	344	<i>Pleurosigma angulatum</i>	23
<i>Ceratium trichoceros</i>	8	<i>Pyramimonas grossi</i>	98
<i>Ceratium contortum</i>	8	<i>Prorocentrum micans</i>	26
 <u>Station 79</u>		<i>Amphidinium sphenoides</i>	16
<i>Oscillatoria submembranacea</i>	4	<i>Protoperidinium cerasus</i>	32
<i>Dictyocha fibula</i>	68	<i>Protoperidinium depressum</i>	1
<i>Distephanus speculum</i>	34	<i>Gonyaulax excavata</i>	128
<i>Cyclococcilithus leptoporus</i>	389	<i>Ceratium lineatum</i>	2
<i>Paralia sulcata</i>	10	<i>Ceratium tripos</i>	3
<i>Corethron criophilum</i>	53	 <u>Station 81</u>	
<i>Leptocylindrus danicus</i>	66	<i>Dictyocha fibula</i>	256
<i>Leptocylindrus minimus</i>	78	<i>Distephanus speculum</i>	376
<i>Coscinodiscus sp.</i>	22	<i>Paralia sulcata</i>	96
<i>Coscinodiscus lineatus</i>	42	<i>Corethron criophilum</i>	84
<i>Coscinodiscus radiatus</i>	2	<i>Leptocylindrus danicus</i>	20
<i>Chaetoceros sp.</i>	4	<i>Thalassiosira gravida</i>	164
<i>Chaetoceros decipiens</i>	42	<i>Coscinodiscus lineatus</i>	160
<i>Rhizosolenia alata</i>	110	<i>Coscinodiscus nitidus</i>	4
<i>Rhizosolenia stolterfothii</i>	56	<i>Actinoptychus senarius</i>	156
<i>Rhizosolenia styliformis</i>	14	<i>Chaetoceros atlanticum</i>	72
<i>Guinardia flaccida</i>	426	<i>Chaetoceros coarctatum</i>	240
<i>Ditylum brightwellii</i>	2	<i>Rhizosolenia alata</i>	136
Unidentified pennate diatom >20 microns	2	<i>Rhizosolenia delicatula</i>	44
<i>Thalassiothrix frauenfeldii</i>	11	<i>Rhizosolenia fragilissima</i>	8
<i>Thalassionema nitzschiooides</i>	188	<i>Rhizosolenia stolterfothii</i>	100
<i>Pleurosigma angulatum</i>	38	<i>Guinardia flaccida</i>	916
<i>Cylindrotheca closterium</i>	2	<i>Synedra sp.</i>	8
<i>Pyramimonas grossi</i>	59	<i>Thalassiothrix frauenfeldii</i>	84
<i>Prorocentrum micans</i>	73	<i>Thalassionema nitzschiooides</i>	340
<i>Ceratium fusus</i>	2	<i>Raphoneis amphiceros</i>	12
<i>Ceratium tripos</i>	4	<i>Pleurosigma elongatum</i>	60
<i>Podolampas elegans</i>	1	<i>Nitzschia pungens</i>	636
 <u>Station 80</u>		<i>Cylindrotheca closterium</i>	44
<i>Hymenomonas roseola</i>	240	<i>Prorocentrum micans</i>	524
<i>Emiliania huxleyii</i>	400	<i>Dinophysis fortii</i>	16
<i>Paralia sulcata</i>	5	<i>Amphidinium sp.</i>	4
<i>Corethron criophilum</i>	6	<i>Amphidinium actutum</i>	8
<i>Leptocylindrus danicus</i>	11	<i>Amphidinium acutissimum</i>	32
<i>Thalassiosira delicatula</i>	4	<i>Gymnodinium dissimile</i>	4
<i>Coscinodiscus lineatus</i>	41	<i>Protoperidinium nipponicum</i>	8
<i>Coscinodiscus wailesii</i>	4	<i>Gonyaulax diegensis</i>	8
<i>Chaetoceros concavicornis</i>	3	<i>Ceratium fusus</i>	8
<i>Chaetoceros decipiens</i>	6	<i>Ceratium lineatum</i>	20
<i>Rhizosolenia alata</i>	115	<i>Ceratium pentagonum</i>	8
<i>Rhizosolenia stolterfothii</i>	40	<i>Ceratium tripos var. atlanticum</i>	28
		<i>Ceratium extensum</i>	8
		<i>Oxytoxum scolpax</i>	8

	<u>Cells/ Liter</u>	<u>Cells/ Liter</u>	
<u>Station 82</u>		<u>Station 84</u>	
<i>Dictyocha fibula</i>	212	<i>Dictyocha fibula</i>	292
<i>Distephanus speculum</i>	4	<i>Paralia sulcata</i>	8
<i>Monodus</i> sp.	8528	<i>Corethron criophilum</i>	8
<i>Corethron criophilum</i>	8	<i>Coscinodiscus lineatus</i>	28
<i>Leptocylindrus danicus</i>	4	<i>Actinoptychus senarius</i>	316
<i>Coscinodiscus nitidus</i>	16	<i>Rhizosolenia alata</i>	8
<i>Coscinodiscus radiatus</i>	4	<i>Rhizosolenia delicatula</i>	164
<i>Actinoptychus senarius</i>	40	<i>Guinardia flaccida</i>	16
<i>Chaetoceros peruvianum</i>	8	<i>Ditylum brightwellii</i>	44
<i>Rhizosolenia fragilissima</i>	4	<i>Thalassiothrix frauenfeldii</i>	132
<i>Guinardia flaccida</i>	4	<i>Pleurosigma angulatum</i>	12
<i>Ditylum brightwellii</i>	8	<i>Pleurosigma elongatum</i>	4
Unidentified pennate diatom >20 microns	8	<i>Nitzschia pungens</i>	688
<i>Synedra</i> sp.	4	<i>Cylindrotheca closterium</i>	36
<i>Synedra tabulata</i> v. <i>fasiculata</i>	8	<i>Euglenophyceae euglenales Euglena</i> sp.	8
<i>Thalassionema nitzschioides</i>	8	Unidentified dinoflagellates	68
<i>Nitzschia pungens</i>	336	<i>Prorocentrum micans</i>	32
<i>Cylindrotheca closterium</i>	16	<i>Prorocentrum dentatum</i>	60
<i>Euglenophyceae euglenales Euglena</i> sp.	4	<i>Amphidinium acutissimum</i>	44
<i>Prorocentrum micans</i>	20	<i>Gymnodinium dissimile</i>	8
<i>Gyrodinium</i> sp.	4	<i>Protoperidinium cerasus</i>	32
<i>Ceratium fusus</i>	12	<i>Gonyaulax diegensis</i>	4
<i>Ceratium minutum</i>	8	<i>Ceratium fusus</i>	36
<i>Ceratium extensum</i>	4	<i>Ceratium lineatum</i>	60
<i>Oxytoxum scolpax</i>	4	<i>Ceratium tripos</i> var. <i>atlanticum</i>	8
		<i>Ceratium minutum</i>	12
		<i>Oxytoxum scolpax</i>	4
<u>Station 83</u>		<u>Station 88</u>	
<i>Dictyocha fibula</i>	3	<i>Nannochloris atomus</i>	35520
<i>Corethron criophilum</i>	4	<i>Olisthodiscus luteus</i>	32
<i>Leptocylindrus danicus</i>	390	<i>Hymenomonas carteri</i>	32
<i>Leptocylindrus minimus</i>	780	Unidentified coccolithophorids	64
<i>Coscinodiscus lineatus</i>	4	<i>Emiliania huxleyii</i>	24
<i>Coscinodiscus radiatus</i>	1	<i>Discosphaera tubifera</i>	8
<i>Chaetoceros concavicornue</i>	3	<i>Thalassiosira rotula</i>	24
<i>Chætoceros decipiens</i>	4	<i>Coscinodiscus marginatus</i>	32
<i>Ditylum brightwellii</i>	2	<i>Coscinodiscus nitidus</i>	32
Unidentified pennate diatom >20 microns	1	<i>Climacodium frauenfeldianum</i>	32
<i>Thalassiothrix frauenfeldii</i>	2	<i>Guinardia flaccida</i>	16
<i>Thalassionema nitzschioides</i>	8	<i>Thalassionema nitzschioides</i>	128
<i>Pleurosigma angulatum</i>	4	<i>Navicula</i> sp.	8
<i>Pyramimonas grossi</i>	140	<i>Stauroneis amphioxis</i>	16
<i>Prorocentrum micans</i>	6	<i>Pleurosigma normani</i>	24
<i>Ceratium fusus</i>	4	<i>Amphora arenaria</i>	8
<i>Ceratium tripos</i>	5	<i>Pyramimonas micron</i>	40
<i>Podolampas elegans</i>	1	<i>Prorocentrum micans</i>	40
		<i>Gyrodinium</i> sp.	8
		<i>Heterocapsa triquetra</i>	8
		<i>Protoperidinium cerasus</i>	24
		<i>Chilomonas marina</i>	32
		<i>Chroomonas vectensis</i>	8

	<u>Cells/ Liter</u>	<u>Cells/ Liter</u>
<u>Station 95</u>		
<i>Nannochloris atomus</i>		8
<i>Olisthodiscus luteus</i>	10464	104
<i>Hymenomonas carteri</i>	40	8
Unidentified coccolithophorids	40	32
<i>Emiliana huxleyii</i>	8	8
<i>Helicosphaera carteri</i>	80	8
<i>Corethron criophilum</i>	8	24
<i>Coscinodiscus marginatus</i>	16	8
<i>Rhizosolenia stolterfothii</i>	8	16
<i>Thalassiothrix frauenfeldii</i>	8	8
<i>Thalassionema nitzschiooides</i>	16	16
<i>Striatella unipunctata</i>	16	16
<i>Achnanthes</i> sp.	8	32
<i>Pyramimonas obovata</i>	16	24
<i>Eutreptia viridis</i>	40	40
<i>Prorocentrum rostratum</i>	24	16
<i>Gymnodinium nelsoni</i>	40	40
<i>Gymnodinium danicans</i>	8	16
<i>Gyrodinium dominans</i>	16	16
<i>Gyrodinium gloicum</i>	24	8
<i>Heterocapsa triquetra</i>	8	24
<i>Scripsiella trochoidea</i>	8	40
<i>Gonyaulax diacantha</i>	16	16
<i>Oxytoxum sphaeroideum</i>	24	16
<i>Chilomonas marina</i>	8	8
<i>Chroomonas vectensis</i>	16	
<u>Station 96</u>		
<i>Nannochloris atomus</i>		3704
<i>Distephanus speculum</i>		72
<i>Ebria tripartita</i>	1080	16
<i>Syracosphaera</i> sp.	8	24
<i>Hymenomonas carteri</i>	8	64
<i>Pontosphaera syracusana</i>	16	8
Unidentified coccolithophorids	8	104
<i>Coccolithus pelagicus</i>	24	16
<i>Emiliana huxleyii</i>	8	456
<i>Corethron criophilum</i>	80	8
<i>Thalassiosira nordenskioldii</i>	32	8
<i>Coscinodiscus marginatus</i>	32	8
<i>Coscinodiscus nitidus</i>	8	80
<i>Coscinodiscus wailesii</i>	24	80
<i>Climacodium frauenfeldianum</i>	16	80
<i>Chaetoceros decipiens</i>	24	104
<i>Rhizosolenia alata</i>	56	80
<i>Navicula directa</i>	32	80
<i>Navicula hennedyii</i>	8	360
<i>Pleurosigma normani</i>	24	8
<i>Phaeodactylum tricornutum</i>	8	88
<i>Nitzschia</i> sp.	16	88
<i>Nitzschia seriata</i>	8	80
	56	24
		72
		8

	<u>Cells/ Liter</u>		<u>Cells/ Liter</u>
Ceratium bucephalum	16	Katodinium rotundatum	16
Chilomonas marina	24	Heterocapsa triquetra	24
Cryptomonas stigmatica	80	Protoperidinium brevipes	64
 <u>Station 98</u>		Protoperidinium pallidum	32
Nannochloris atomus	56	Scripsiella trochoidea	16
Hymenomonas carteri	24	Gonyaulax diacantha	16
Unidentified coccolithophorids	8	Gonyaulax unicornis	8
Coccolithus pelagicus	8	Ceratium tripos	16
Emiliania huxleyii	48	Oxytoxum sp.	8
Coscinodiscus wailesii	8	 <u>Station 102</u>	
Coscinodiscus nobilis	16	Nannochloris atomus	1160
Biddulphia regia	24	Hymenomonas carteri	8
Hemialulus hauckii	8	Coscinodiscus wailesii	16
Rhizosolenia setigera	8	Guinardia flaccida	8
Dinophysis acuminata	8	Thalassionema nitzschiooides	8
Protoperidinium cerasus	48	Pyramimonas micron	8
Amphidoma sp.	32	Heterocapsa triquetra	24
Gonyaulax excavata	16	Protoperidinium cerasus	8
Chilomonas marina	16	Protoperidinium subinerme	8
Chroomonas amphioxea	8	 <u>Station 104</u>	
Cryptomonas salina	8	Nannochloris atomus	984
 <u>Station 99</u>		Dictyocha fibula	1
Nannochloris atomus	54912	Coscinodiscus radiatus	1
Calycomonas ovalis	32	Coscinodiscus wailesii	24
Olisthodiscus luteus	40	Chaetoceros decipiens	6
Hymenomonas roseola	40	Thalassiothrix frauenfeldii	5
Hymenomonas carteri	8	Pleurosigma angulatum	1
Syracospheara apsteini	16	Amphidinium sphenoides	8
Unidentified coccolithophorids	32	Heterocapsa triquetra	56
Coccolithus pelagicus	48	Protoperidinium cerasus	16
Emiliania huxleyii	48	Gonyaulax excavata	16
Halopappus sp.	8	Ceratium tripos	1
Helicosphaera carteri	8	 <u>Station 105</u>	
Coscinodiscus wailesii	24	Cyclococcolithus leptoporus	1072
Coscinodiscus nobilis	16	Corethron criophilum	8
Biddulphia regia	8	Coscinodiscus stellaris	192
Chaetoceros sp.	72	Coscinodiscus wailesii	264
Rhizosolenia delicatula	184	Coscinodiscus oculus iridis	8
Rhizosolenia stolterfothii	160	Rhizosolenia alata	728
Thalassiothrix frauenfeldii	16	Rhizosolenia alata f. indica	32
Navicula sp.	8	Rhizosolenia hebetata f. semispina	24
Pyramimonas micron	8	Guinardia flaccida	32
Eutreptia marina	8	Prorocentrum micans	8
Euglena acus	8	Dinophysis sphaerica	8
Prorocentrum sp.	16	Ceratium tripos	64
Prorocentrum rostratum	32	Ceratium minutum	8
Gymnodinium simplex	24		
Gyrodinium sp.	136		

	<u>Cells/ Liter</u>	<u>Cells/ Liter</u>	
<u>Station 106</u>			
<i>Distephanus speculum</i>	8	Dictyocha fibula	632
<i>Corethron criophilum</i>	16	<i>Distephanus speculum</i>	912
<i>Leptocylindrus danicus</i>	12	<i>Paralia sulcata</i>	160
<i>Coscinodiscus marginatus</i>	12	<i>Corethron criophilum</i>	8
<i>Coscinodiscus nitidus</i>	4	<i>Thalassiosira decipiens</i>	600
<i>Coscinodiscus radiatus</i>	4	<i>Coscinodiscus</i> sp.	40
<i>Coscinodiscus wailesii</i>	36	<i>Coscinodiscus wailesii</i>	48
<i>Actinptychus senarius</i>	48	<i>Chaetoceros decipiens</i>	512
<i>Chaetoceros coarctatum</i>	8	<i>Rhizosolenia alata</i>	800
<i>Rhizosolenia alata</i>	4	<i>Rhizosolenia stolterfothii</i>	72
<i>Rhizosolenia alata f. gracillima</i>	4	<i>Rhizosolenia styliformis</i>	48
<i>Rhizosolenia styliformis</i>	16	<i>Guinardia flaccida</i>	1440
<i>Ditylum brightwellii</i>	4	<i>Thalassionema nitzschiooides</i>	176
Unidentified pennate diatom >20 microns	72	<i>Pleurosigma angulatum</i>	120
<i>Plagiogramma staurophorum</i>	4	<i>Pyramimonas grossi</i>	136
<i>Tabellaria fenestriata</i> v. <i>asterionelloides</i>	44	<i>Prorocentrum micans</i>	2664
<i>Pleurosigma angulatum</i>	12	<i>Ceratium furca</i>	16
<i>Nitzschia pungens</i>	168	<i>Ceratium lineatum</i>	16
<i>Cylindrotheca closterium</i>	12	<i>Ceratium tripos</i>	136
<i>Prorocentrum</i> sp.	20		
<i>Amphidinium acutissimum</i>	20	<u>Station 114</u>	
<i>Gymnodinium dissimile</i>	20	<i>Dictyocha fibula</i>	36
<i>Gymnodinium articum</i>	4	<i>Distephanus speculum</i>	24
<i>Gonyaulax diegensis</i>	4	<i>Cyclococcolithus leptoporus</i>	72
<i>Ceratium lineatum</i>	16	<i>Melosira moniliformis</i>	32
<i>Ceratium tripos</i> var. <i>atlanticum</i>	8	<i>Corethron criophilum</i>	6
		<i>Lauderia borealis</i>	216
<u>Station 108</u>		<i>Coscinodiscus lineatus</i>	16
<i>Oscillatoria submembranacea</i>	2	<i>Coscinodiscus radiatus</i>	4
<i>Dictyocha fibula</i>	4	<i>Asterolampra marylandica</i>	2
<i>Distephanus speculum</i>	1	<i>Chaetoceros</i> sp.	62
<i>Corethron criophilum</i>	2	<i>Chaetoceros pendulum</i>	44
<i>Coscinodiscus radiatus</i>	42	<i>Rhizosolenia alata</i>	48
<i>Coscinodiscus wailesii</i>	28	<i>Rhizosolenia alata f. gracillima</i>	24
<i>Chaetoceros concavicornue</i>	7	<i>Rhizosolenia hebetata f. semispina</i>	24
<i>Rhizosolenia alata</i>	7	<i>Rhizosolenia stolterfothii</i>	44
<i>Guinardia flaccida</i>	1	<i>Guinardia flaccida</i>	440
<i>Thalassionema nitzschiooides</i>	8	<i>Thalassiothrix frauenfeldii</i>	112
<i>Pleurosigma angulatum</i>	4	<i>Pleurosigma angulatum</i>	40
<i>Pyramimonas grossi</i>	18	<i>Pyramimonas grossi</i>	8688
<i>Prorocentrum micans</i>	72	<i>Prorocentrum</i> sp.	4
<i>Dinophysis sphaerica</i>	1	<i>Prorocentrum micans</i>	204
<i>Gymnodinium</i> sp.	2	<i>Ornithocercus thurni</i>	4
<i>Ceratium tripos</i>	1	<i>Ceratium furca</i>	16
		<i>Ceratium tripos</i>	28

	<u>Cells/ Liter</u>		<u>Cells/ Liter</u>
<u>Station 116</u>			
<i>Oscillatoria erythraea</i>	16	<i>Plagiogramma vanheurckii</i>	8
<i>Nostoc commune</i>	4	<i>Navicula hennedyii</i>	16
<i>Leptocylindrus danicus</i>	48	<i>Pleurosigma normani</i>	8
<i>Lauderia borealis</i>	20	<i>Phaeodactylum tricornutum</i>	40
<i>Coscinodiscus wailesii</i>	8	<i>Nitzschia recta</i>	8
<i>Actinioptychus senarius</i>	28	<i>Eutreptia viridis</i>	8
<i>Biddulphia aurita</i>	4	<i>Prorocentrum micans</i>	16
<i>Chaetoceros atlanticum</i>	8	<i>Prorocentrum apora</i>	8
<i>Chaetoceros costatum</i>	16	<i>Amphisolenia globifera</i>	8
<i>Rhizosolenia alata</i>	84	<i>Dinophysis acuminata</i>	16
<i>Rhizosolenia alata f. gracillima</i>	16	<i>Gyrodinium spirale</i>	8
<i>Rhizosolenia alata f. indica</i>	4	<i>Gyrodinium undulans</i>	16
<i>Rhizosolenia bergonii</i>	8	<i>Katodinium rotundatum</i>	8
<i>Rhizosolenia calcar-avis</i>	8	<i>Heterocapsa triquetra</i>	40
<i>Rhizosolenia delicatula</i>	8	<i>Protoperidinium cerasus</i>	16
<i>Rhizosolenia imbricata</i>	20	<i>Protoperidinium conicum</i>	64
<i>Ditylum brightwellii</i>	104	<i>Scripsiella trochoidea</i>	48
Unidentified pennate diatom >20 microns	12	<i>Protoperidinium pentagonum</i>	80
<i>Dimerogramma</i> sp.	20	<i>Protoperidinium abei</i>	8
<i>Plagiogramma staurophorum</i>	4	<i>Gonyaulax spinifera</i>	8
<i>Tabellaria fenestriata</i> v. <i>asterionelloides</i>	20	<i>Gonyaulax diacantha</i>	8
<i>Nitzschia pungens</i>	148	<i>Gonyaulax polyedra</i>	24
<i>Cylindrotheca closterium</i>	160	<i>Ceratium fusus</i>	16
<i>Staurastrum leptocladum</i>	12	<i>Ceratium lineatum</i>	16
<i>Euglenophyceae euglenales Euglena</i> sp.	4	<i>Oxytoxum scolpax</i>	8
<i>Prorocentrum micans</i>	20	<u>Station 141</u>	
<i>Prorocentrum dentatum</i>	16	<i>Nannochloris atomus</i>	6032
<i>Amphidinium actutum</i>	4	<i>Calymonas gracilis</i>	8
<i>Amphidinium acutissimum</i>	60	<i>Hymenomonas roseola</i>	24
<i>Protoperidinium steinii</i>	4	<i>Striatella unipunctata</i>	16
<i>Ceratium fusus</i>	4	<i>Amphidinium</i> sp.	8
<i>Ceratium tripos</i> var. <i>atlanticum</i>	4	<i>Gymnodinium minutum</i>	8
<i>Ceratium trichoceros</i>	8	<i>Gyrodinium spirale</i>	8
<i>Ceratium extensem</i>	16	<i>Hemidinium</i> sp.	8
<i>Oxytoxum sceptrum</i>	4	<i>Heterocapsa triquetra</i>	8
<i>Oxytoxum scolpax</i>	8	<i>Protoperidinium excentricum</i>	8
<u>Station 138</u>		<i>Protoperidinium leonis</i>	32
<i>Nannochloris atomus</i>	48	<i>Chroomonas vectensis</i>	32
<i>Distephanus speculum</i>	16	<i>Chroomonas amphioxaea</i>	8
<i>Schroederella delicatula</i>	208	<u>Station 143</u>	
<i>Coscinodiscus marginatus</i>	16	<i>Dictyocha fibula</i>	21
<i>Coscinodiscus nitidus</i>	16	<i>Distephanus speculum</i>	45
<i>Coscinodiscus wailesii</i>	24	<i>Corethron criophilum</i>	8
<i>Cerataulina pelagica</i>	104	<i>Leptocylindrus danicus</i>	18
<i>Chaetoceros decipiens</i>	88	<i>Leptocylindrus minimus</i>	6
<i>Rhizosolenia fragilissima</i>	8	<i>Coscinodiscus lineatus</i>	6
<i>Rhizosolenia setigera</i>	40	<i>Coscinodiscus radiatus</i>	6
<i>Ditylum brightwellii</i>	32	<i>Coscinodiscus wailesii</i>	1
		<i>Chaetoceros decipiens</i>	24

	<u>Cells/ Liter</u>		<u>Cells/ Liter</u>		
Rhizosolenia alata	4	Rhizosolenia alata	40		
Rhizosolenia hebetata f. semispina	3	Rhizosolenia delicatula	560		
Thalassionema nitzschiooides	22	Rhizosolenia hebetata f. semispina	40		
Pleurosigma angulatum	42	Rhizosolenia stolterfothii	1240		
Gyrosigma hippocampus	1	Rhizosolenia styliformis	320		
Pyramimonas grossi	86	Guinardia flaccida	600		
Prorocentrum micans	5	Ditylum brightwellii	80		
Dinophysis sphaerica	1	Thalassiothrix frauenfeldii	320		
Gymnodinium sp.	1	Thalassionema nitzschiooides	960		
Ceratium lineatum	5	Rhaphoneis amphiceros	40		
Ceratium tripos	14	Pleurosigma angulatum	40		
<u>Station 145</u>					
Cyclococcolithus leptoporus	189	Anacyctis marina	3600		
Corethron criophilum	3	Pyramimonas grossi	7000		
Leptocylindrus danicus	182	Prorocentrum micans	240		
Thalassiosira rotula	1	Ceratium lineatum	40		
Lauderia borealis	5	Ceratium tripos	40		
Coscinodiscus lineatus	4	<u>Station 147</u>			
Coscinodiscus radiatus	4	Dictyocha fibula	24		
Chaetoceros concavicornue	54	Distephanus speculum	16		
Rhizosolenia alata	2	Paralia sulcata	632		
Rhizosolenia hebetata f. semispina	3	Corethron criophilum	48		
Rhizosolenia stolterfothii	14	Thalassiosira gravida	112		
Synedra sp.	1	Lauderia borealis	32		
Pleurosigma angulatum	4	Coscinodiscus sp.	400		
Anacyctis marina	6	Coscinodiscus lineatus	48		
Pyramimonas grossi	131	Coscinodiscus nitidus	24		
Prorocentrum sp.	1	Biddulphia alternans	32		
Prorocentrum micans	8	Cerataulina pelagica	48		
Gymnodinium sp.	8	Chaetoceros atlanticum	80		
Glenodinium sp.	2	Chaetoceros coarctatum	40		
Ceratium tripos	1	Chaetoceros danicum	48		
<u>Station 146</u>		Chaetoceros sociale	328		
Oscillatoria submembranacea	40	Rhizosolenia alata	64		
Dictyocha fibula	400	Rhizosolenia imbricata	632		
Cyclococcolithus leptoporus	200	Rhizosolenia stolterfothii	768		
Paralia sulcata	640	Guinardia flaccida	656		
Corethron criophilum	240	Ditylum brightwellii	32		
Schroederella delicatula	40	Thalassiothrix frauenfeldii	152		
Leptocylindrus danicus	62400	Rhaphoneis surirella	16		
Thalassiosira nordenskioldii	180	Rhaphoneis amphiceros	56		
Thalassiosira rotula	40	Plagiogramma staurophorum	96		
Coscinodiscus sp.	160	Diploneis crabro	24		
Coscinodiscus asteromphalus	40	Pleurosigma hamuliferum	16		
Coscinodiscus marginatus	40	Nitzschia pungens	7752		
Coscinodiscus nitidus	40	Cylindrotheca closterium	24		
Coscinodiscus radiatus	160	Prorocentrum micans	360		
Coscinodiscus wailesii	40	Dinophysis micropterygia	32		
Biddulphia alternans	40	Dinophysis fortii	24		
		Gymnodinium articum	24		
		Gyrodinium sp.	16		
		Protoperidinium cerasus	40		
		Ceratium tripos var. atlanticum	32		
		Ceratium minutum	32		

	<u>Cells/ Liter</u>		<u>Cells/ Liter</u>
<u>Station 148</u>			
<i>Distephanus speculum</i>	24	<i>Thalassionema nitzschioides</i>	32
<i>Cyclococcolithus leptoporus</i>	2696	<i>Pyramimonas grossi</i>	1616
<i>Coscinodiscus lineatus</i>	24	<i>Gymnodinium</i> sp.	120
<i>Coscinodiscus wailesii</i>	40	<i>Ceratium furca</i>	8
<i>Chaetoceros decipiens</i>	588	<i>Ceratium tripos</i>	8
<i>Chaetoceros peruvianum</i>	8		
<i>Rhizosolenia alata</i>	56	<u>Station 183</u>	
<i>Rhizosolenia hebetata f. semispina</i>	48	<i>Distephanus speculum</i>	34
<i>Thalassionema nitzschioides</i>	160	<i>Corethron criophilum</i>	30
<i>Pyramimonas grossi</i>	2976	<i>Coscinodiscus</i> sp.	8
<i>Prorocentrum micans</i>	16	<i>Coscinodiscus nitidus</i>	2
<i>Ceratium furca</i>	16	<i>Coscinodiscus radiatus</i>	62
<i>Ceratium fusus</i>	8	<i>Actinoptychus senarius</i>	4
<i>Ceratium macroceros</i>	8	<i>Chaetoceros atlanticum</i>	4
<i>Ceratium tripos</i>	80	<i>Chaetoceros coarctatum</i>	24
		<i>Chaetoceros danicum</i>	4
<u>Station 150</u>		<i>Rhizosolenia setigera</i>	12
<i>Leptocylindrus danicus</i>	16	<i>Rhizosolenia styliformis</i>	4
<i>Thalassiosira gravida</i>	4	Unidentified pennate diatoms	
<i>Coscinodiscus nitidus</i>	8	>20 microns	16
<i>Actinoptychus senarius</i>	124	<i>Nitzschia pungens</i>	12
<i>Cerataulina pelagica</i>	196	<i>Dinophysis ovum</i>	2
<i>Ditylum brightwellii</i>	8	<i>Amphidinium acutissimum</i>	4
<i>Synedra tabulata v. fasciculata</i>	4	<i>Gymnodinium dissimile</i>	22
<i>Thalassiothrix frauenfeldii</i>	28	<i>Ceratium lineatum</i>	4
<i>Nitzschia pungens</i>	1628	<i>Ceratium tripos var. atlanticum</i>	4
Unidentified dinoflagellates	88		
<i>Prorocentrum micans</i>	120		
<i>Dinophysis ovum</i>	12		
<i>Amphidinium acutissimum</i>	100		
<i>Gymnodinium</i> sp.	80		
<i>Conyaulax diegensis</i>	88		
<i>Ceratium lineatum</i>	108		
<i>Ceratium tripos</i>	16		
<i>Ceratium tripos var. atlanticum</i>	12		
<i>Ceratium minutum</i>	36		
<u>Station 182</u>			
<i>Distephanus speculum</i>	8		
<i>Cyclococcolithus leptoporus</i>	3912		
<i>Corethron criophilum</i>	376		
<i>Leptocylindrus danicus</i>	56		
<i>Coscinodiscus radiatus</i>	48		
<i>Coscinodiscus stellaris</i>	168		
<i>Coscinodiscus wailesii</i>	48		
<i>Coscinodiscus occulus</i>	104		
<i>Chaetoceros decipiens</i>	80		
<i>Rhizosolenia alata</i>	728		
<i>Rhizosolenia hebetata f. semispina</i>	104		

(continued from inside front cover)

5. *The Status of the Marine Fishery Resources of the Northeastern United States.* By Margaret M. McBride and Bradford E. Brown. December 1980. viii + 13 p., 4 figs., 3 tables.
6. *Economic and Biological Data Needs for Fisheries Management, With Particular Reference to the New England and Mid-Atlantic Areas.* By Guy D. Marchesseault, Joseph J. Mueller, and Ivar E. Strand, Jr. December 1980. vi + 10 p., 1 fig., 3 tables.
7. *Methodology for Identification and Analysis of Fishery Management Options.* By Brian J. Rothschild, Richard C. Hennemuth, Jacob J. Dykstra, Leo C. Murphy, Jr., John C. Bryson, and James D. Ackert. December 1980. vi + 10 p., 5 figs., 1 app.
8. *Phytoplankton Community Structure in Northeastern Coastal Waters of the United States. I. October 1978.* By Harold G. Marshall and Myra S. Cohn. August 1981. v + 14 p., 4 figs., 1 app.

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74

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