

Whole-Exome Sequencing: Technical Details

Jim Mullikin

Director, NIH Intramural Sequencing Center
Head, Comparative Genomics Unit

Whole Exome Sequencing, Why?

- Focuses on the part of the genome we understand best, the exons of genes
- Exomes are ideal to help us understand high-penetrance allelic variation and its relationship to phenotype.
- A whole exome is 1/6 the cost of whole genome and 1/15 the amount of data

Biesecker *et al.* *Genome Biology* 2011, **12**:128

Twinbrook Research Building

→
NISC occupies
entire 5th floor



5625 Fishers Lane, Rockville MD

NISC Sequence Production

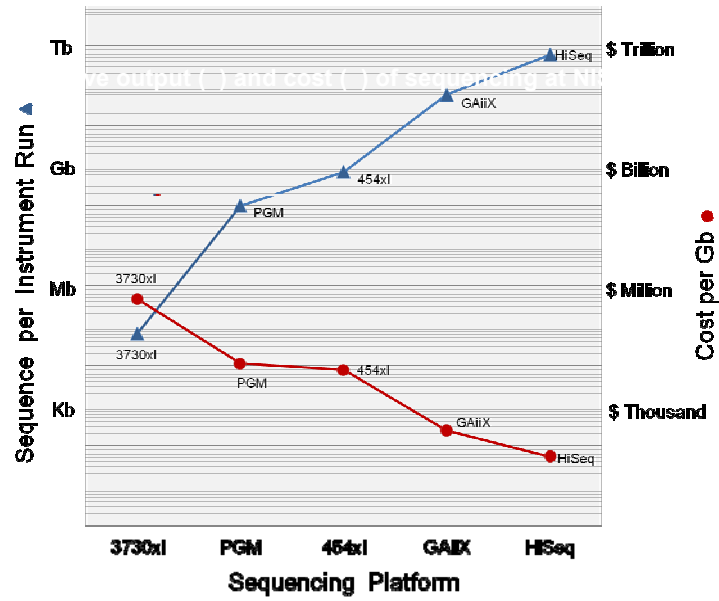


Feb. 2010

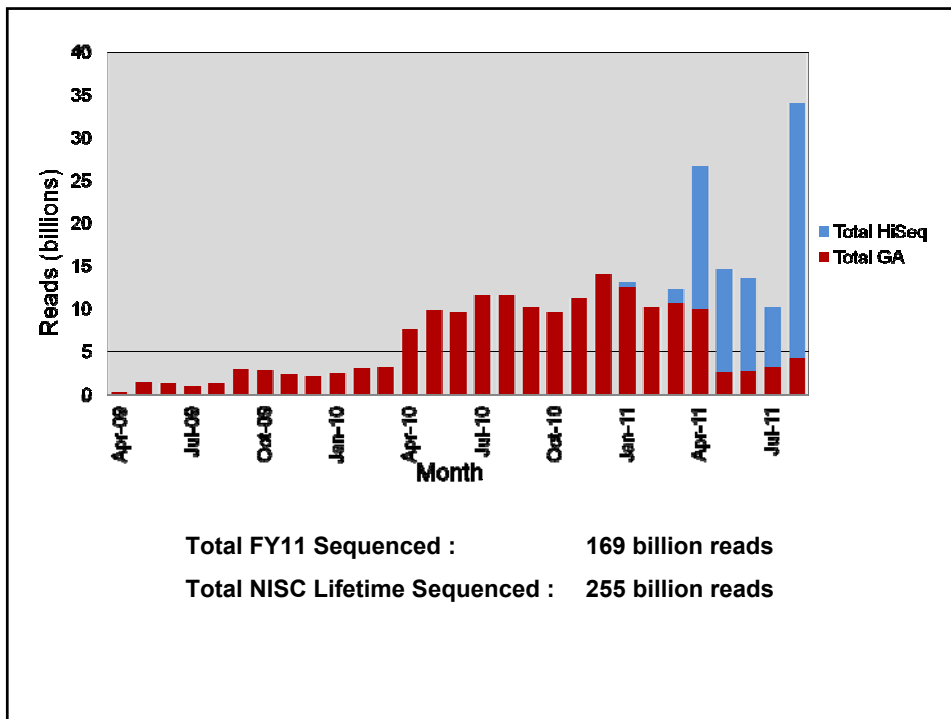
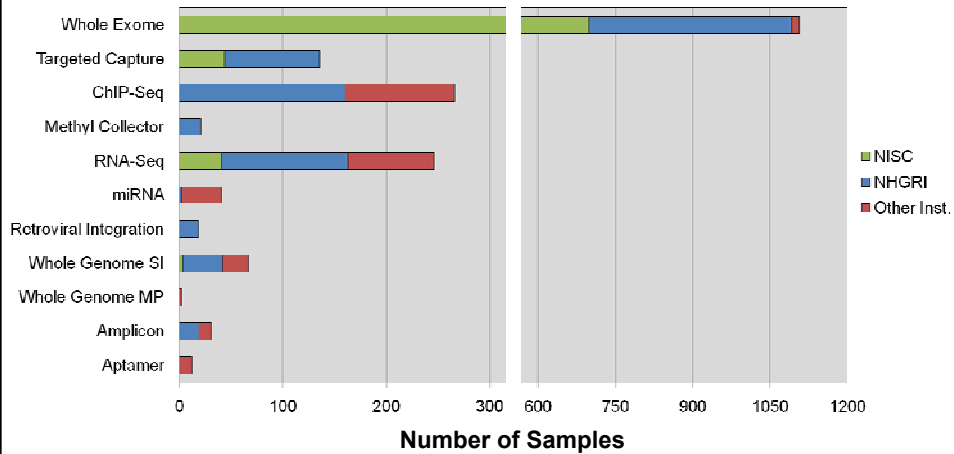
NISC Sequence Production



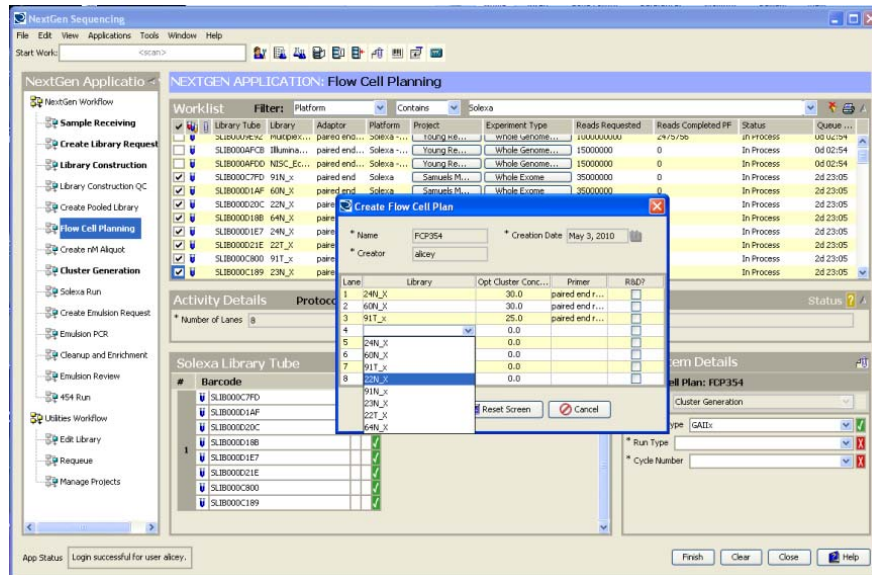
March 2011



Sequencing applications processed through NGS production pipeline from April 2009 to August 2011.



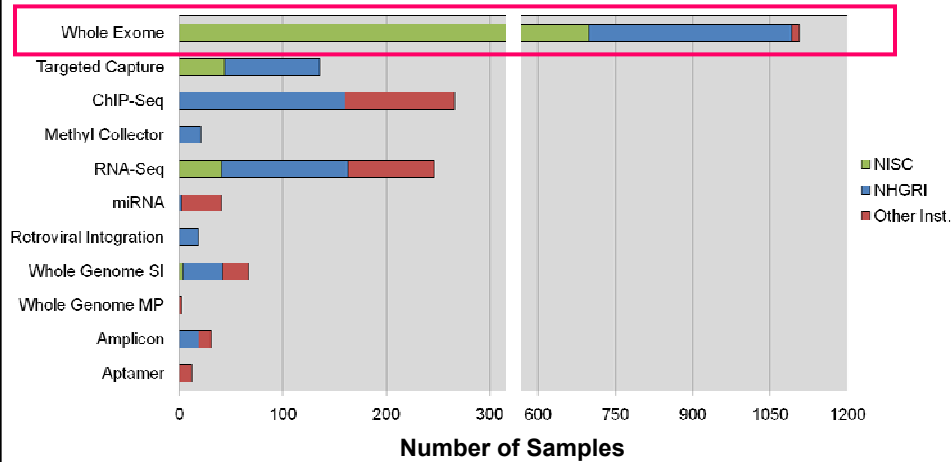
Cimarron Software based NextGen LIMS



Computational Resources for 6 GAiiX and 3 HiSeq2000

- **Linux cluster**
 - 1000 cores
 - 250 for production
 - 900TB disk
 - 250TB for production with 75TB available
 - 15TB/month long term storage
 - **Network**
 - 1 and 10 Gigabit-Ethernet

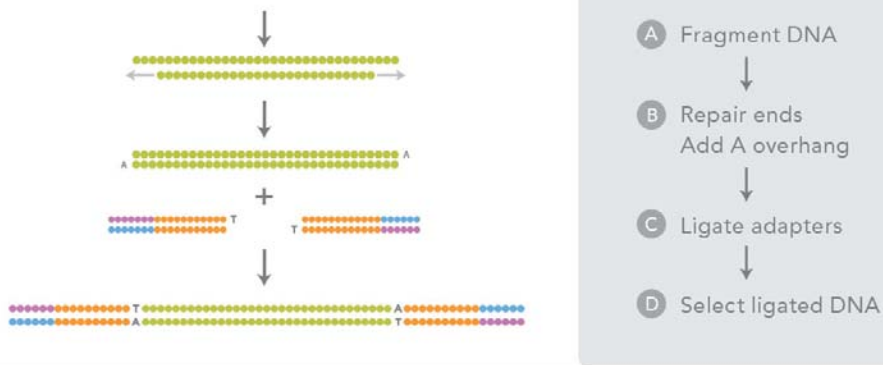
Sequencing applications processed through NGS production pipeline from April 2009 to August 2011.



Exome Sequencing Pipeline

- Sample DNA Fragmentation
- Illumina Library Preparation
- Exome Enrichment
- Cluster Generation
- Sequencing and Basecalling
- Sequence Read Alignment
- Variation Detection

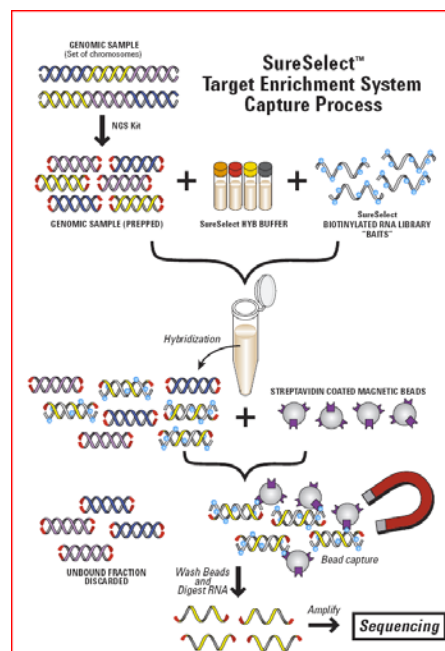
Library Preparation



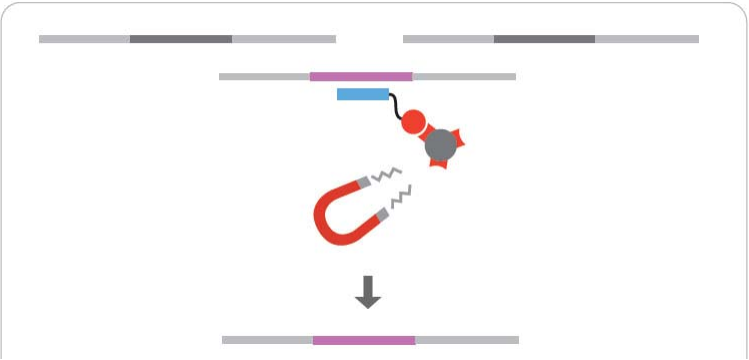
<http://www.illumina.com/support/literature.ilmn>

Agilent Technologies SureSelect method

Whole-exome kit
38Mb and 50Mb



<http://cp.literature.agilent.com/litweb/pdf/5990-3532EN.pdf>



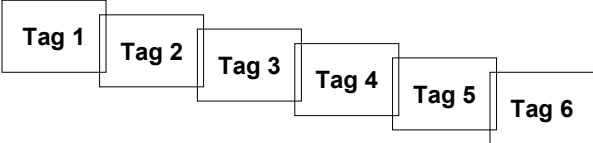
The diagram illustrates the Illumina TruSeq Exome Enrichment process. It shows a DNA template with a specific region highlighted in pink. A probe with a red hook and a grey bead is shown binding to this region. An arrow points to the final state where the probe has captured the target DNA, forming a loop.

- Illumina TruSeq Exome Enrichment
- 62Mb of exome targeted

<http://www.illumina.com/support/literature.ilmn>

NISC Exome Process


Indexed Libraries



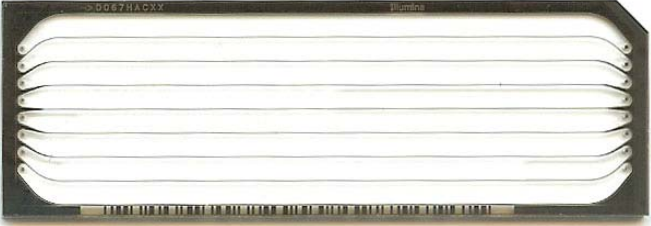
Tag 1 Tag 2 Tag 3 Tag 4 Tag 5 Tag 6

Balance and Pool

Illumina TruSeq Exome Enrichment

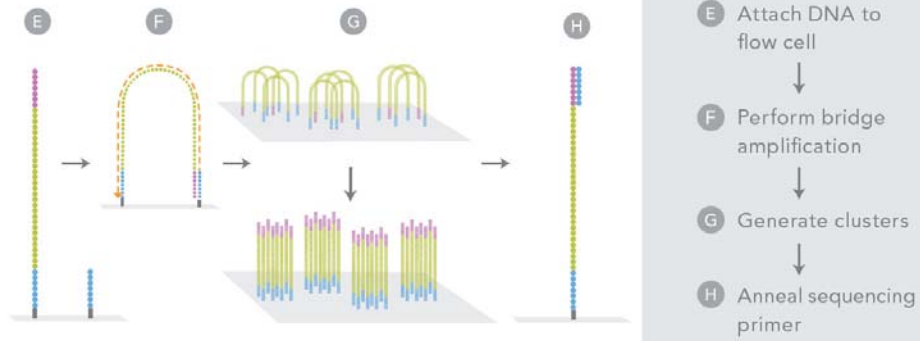


Sequence on two lanes



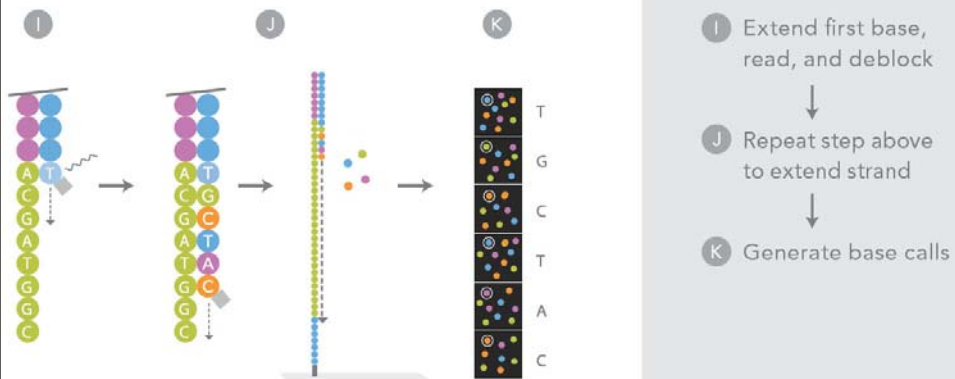
The diagram shows the NISC Exome Process. It starts with six indexed libraries (Tag 1 to Tag 6) which are balanced and pooled. This pool then undergoes Illumina TruSeq Exome Enrichment. The final step is sequencing on two lanes, as shown by the image of a sequencing lane.

Cluster Generation



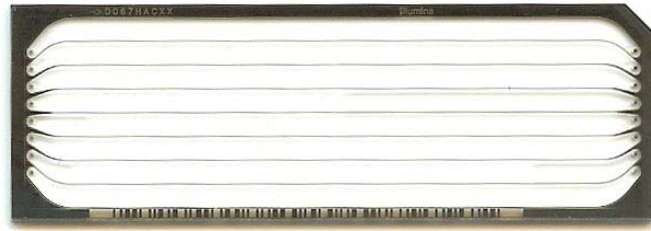
<http://www.illumina.com/support/literature.ilmn>

Sequencing



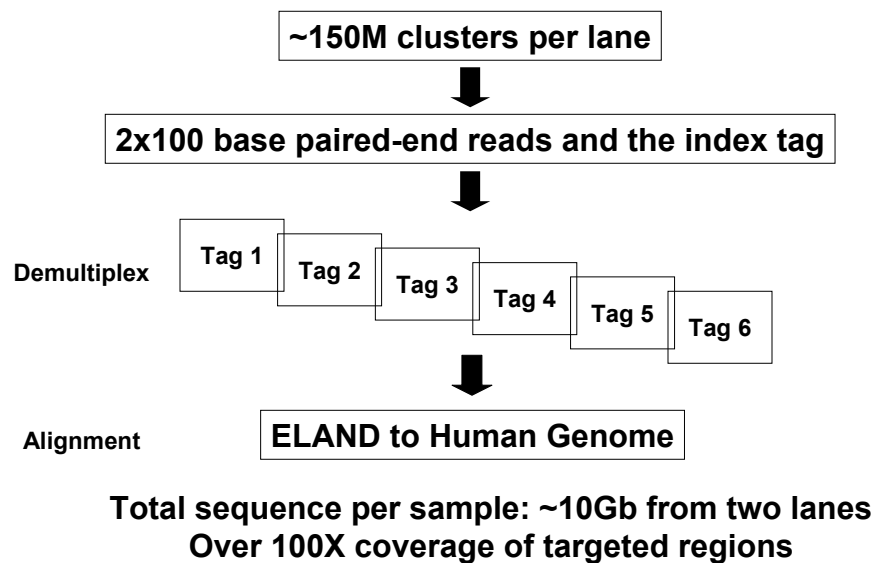
<http://www.illumina.com/support/literature.ilmn>

HiSeq Flow Cell

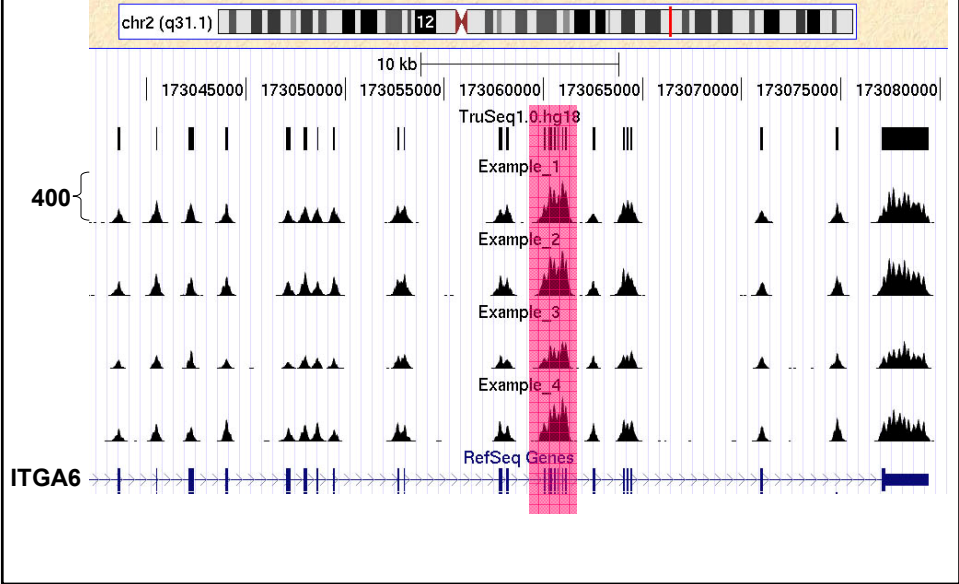


- 8 lanes
- 1.5 billion clusters
- Up to 300Gb per flow cell

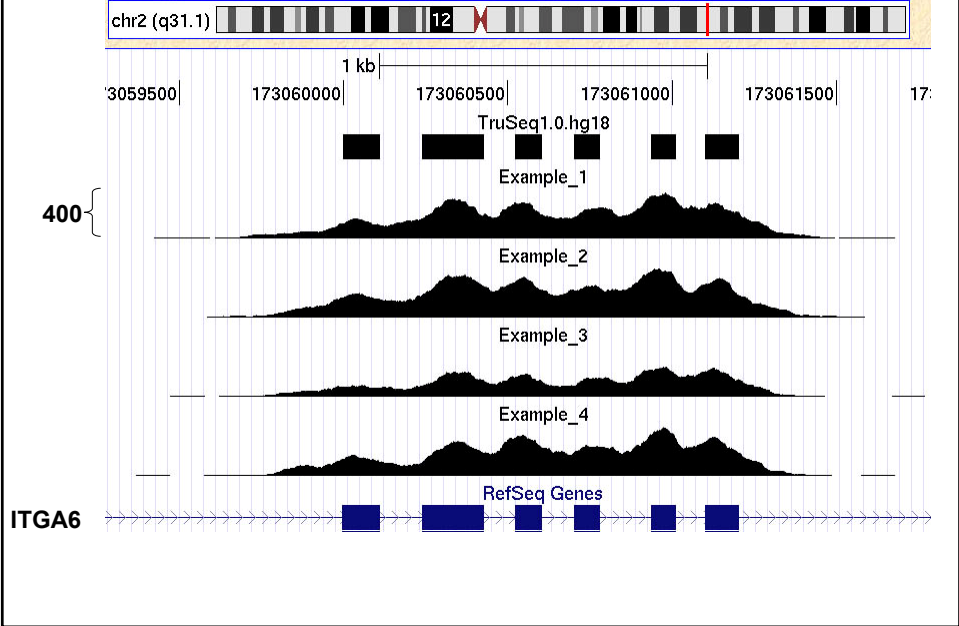
Sequencing and Data Processing

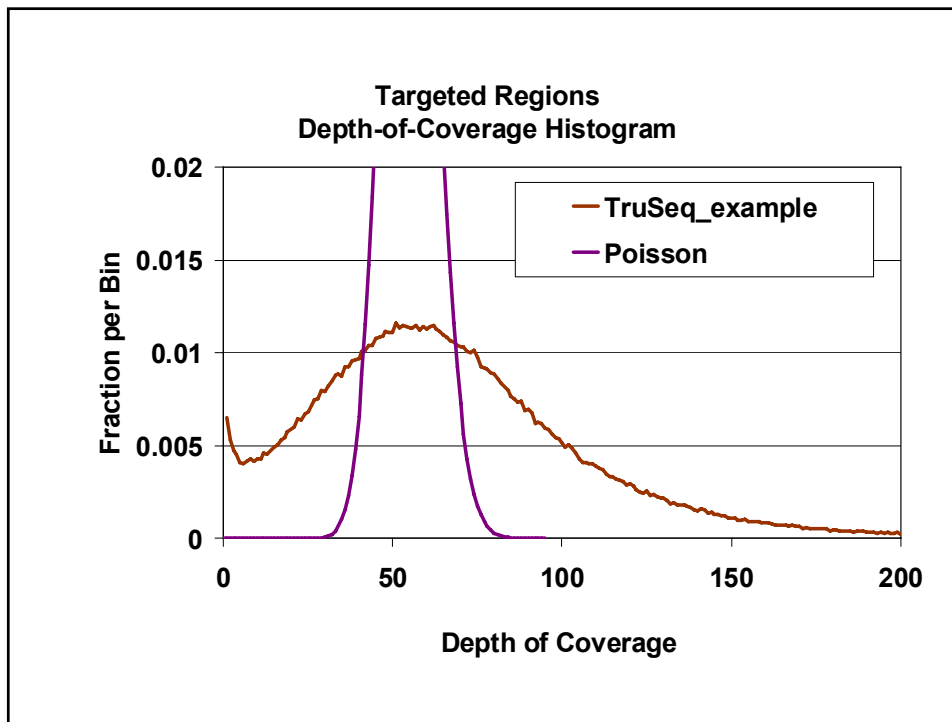


Read Depth-of-Coverage



Read Depth-of-Coverage





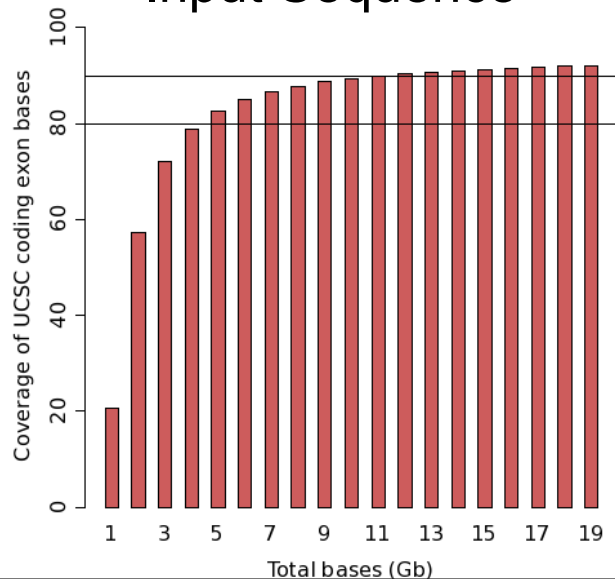
Refining the Alignment (diagCM)

- **ELAND is part of the standard pipeline**
- **ELAND accurately places reads in the correct genomic location**
- **Use cross-match, a Smith-Waterman aligner, to improve local alignment**

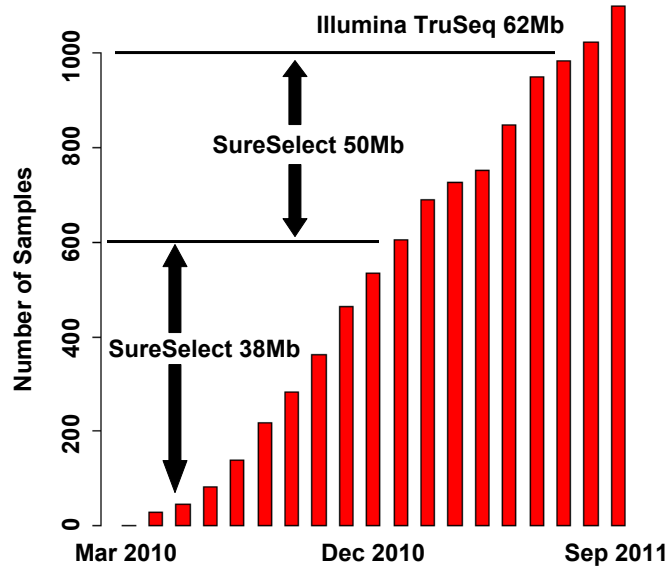
MPG of Haploid Regions

- Human autosomes are normally diploid
- MPG is designed to call two alleles for the autosomes, and X chromosome if the sample is from a female
- For samples from males, MPG is run in haploid mode on the non-sudoautosomal regions of X and Y
- Thus only testing for the four nucleotides

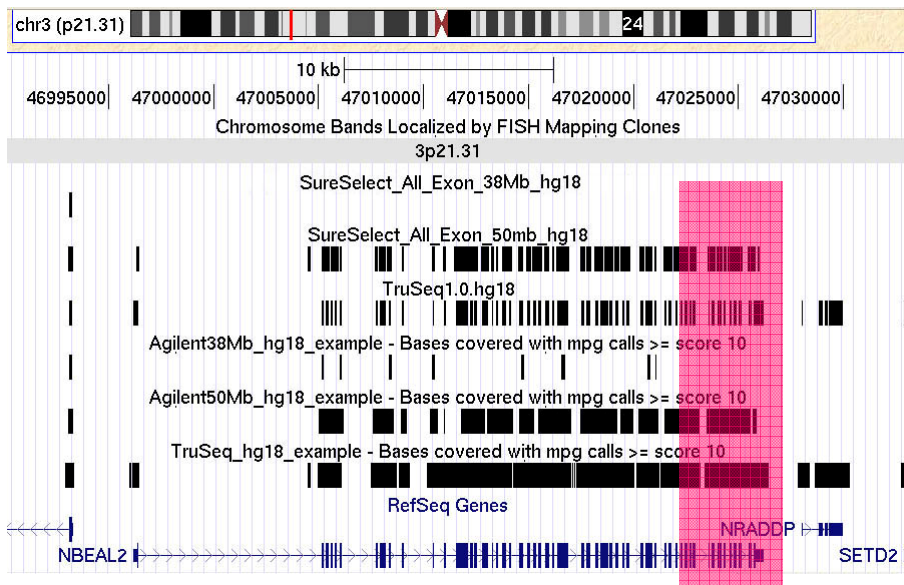
Exome Coverage versus Input Sequence



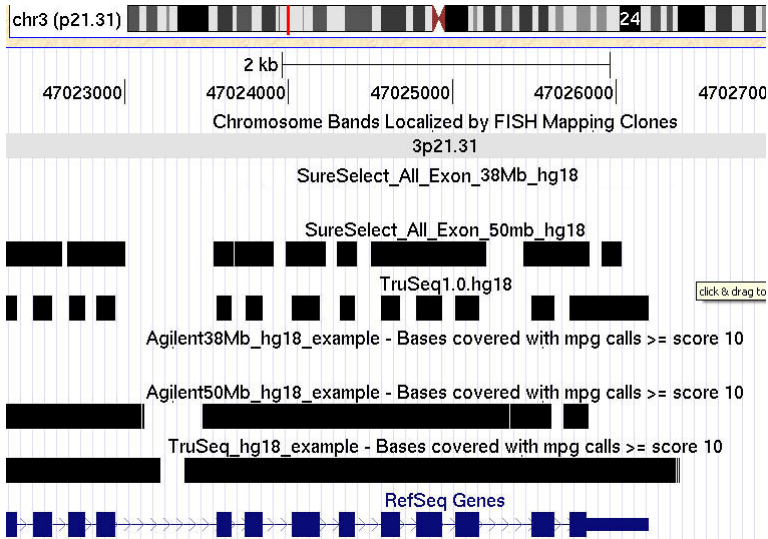
Whole Exomes Processed at NISC



NBEAL2



NBEAL2 5'



Exome Variation Statistics TruSeq 62Mb, Male Sample

Type	Total Genotype Calls	SNVs	Within-sample Heterozygosity
Total	133,047,403	142,361	0.00072
Auto	125,491,045	139,295	0.00076
chrX	6,842,299	2,600	NA
chrY	710,243	1,435	NA

Exome Variation Statistics TruSeq 62Mb, Female Sample

Type	Total Genotype Calls	SNVs	Within-sample Heterozygosity
Total	125,681,915	136,993	0.00075
Auto	120,559,746	132,616	0.00076
chrX	5,096,376	2,701	0.00034

Example Heterozygous SNV

145590901 145590911 145590921 145590931 145590941 145590951 145590961 145590971
TATGTACAAGGATGTGTCACAGCTCTCTGTTCGATCATCCTTGCAAAATCTTTCAGCATGGGGCAGCTTGGGAGGTTGTGTGCCTGAAAAGG
.....K.....
.....
.....
.....a.....
.....G.....
.....G.....
.....g.....
.....g.....
.....G.....
.....g.....
.....G.....
.....G.....
.....G.....
.....G.....
.....G.....
.....G.....
.....
.....
.....
.....
.....
.....g.....
.....c.....g.....g.....

Genotype Concordance

	Total Agreement with Genotype Chip (CCDS)
Whole Genome Shotgun	99.908%
SureSelect 38Mb	99.910%
SureSelect 50Mb	99.857%
TruSeq 62Mb	99.865%

Whole Exome Sequencing

- **Being applied to**
 - Undiagnosed Diseases Program (100's of samples)
 - ClinSeq (>1000 samples)
 - Variety of other PI driven projects (e.g. cancer)
- **Data generation rate per year**
 - 200 exomes per GAiiX
 - 1200 exomes per HiSeq 2000
- **Analysis results**
 - Genotype data for 90% of consensus coding exon bases (CCDS)
 - Accuracy of genotype calls over 99.5%

Exome Sequencing Pipeline

Sample DNA Fragmentation
Illumina Library Preparation
Exome Enrichment
Cluster Generation
Sequencing and Basecalling
Sequence Read Alignment
Variation Detection

Variant Annotation and Working With Whole-Exome Data

- One sample produces > 100k variants
- One hundred samples gives rise to 600k or more
- How does one work with such large datasets?
- The next speaker, Dr. Jamie Teer, will address these next steps

Acknowledgements

NIH Intramural Sequencing Center

- **Sequencing Operations**

- Bob Blakesley
- Alice Young
- Lab Staff

- **Bioinformatics**

- Gerry Bouffard
- Baishali Maskeri
- Jenny McDowell
- Meg Vemulapalli

- **IT Linux Support**

- Jesse Becker
- Matt Lesko

- **Mullikin Lab**

- Nancy Hansen

- Pedro Cruz

- Praveen Cherukuri

- **Biesecker Lab**

- Jamie Teer

<http://research.nhgri.nih.gov/>

