

# FIELD SCALE EVALUATION OF BIOSTIMULATION FOR REMEDICATION OF URANIUM-CONTAMINATED GROUNDWATER AT FRC AREA 3

Stanford University



Craig Criddle  
Scott Fendorf

Anna Michalak  
Jennifer Nyman

Michael Fienen  
Margy Gentile  
Matt Ginder-Vogel

Julie Stevens  
Weimin Wu

Peter Kitanidis  
Jian Luo

ETH

Frank-Andreas Weber

RETEC



Robert Hickey  
Raj Rajan Luke Smith  
Dan Wagner

LBNL



Susan Hubbard  
Ken Williams

INEEL



Gill Geesey  
David Cummings

Miami University

Chiachi Hwang  
Matthew Fields

EAWAG

Olaf Cirpka

Oak Ridge National Laboratories



Craig Brandt  
Baohua Gu  
Philip Jardine

Ye-Kyoung Ku  
Stacy Rast  
Jeff Riggs Hui Yan

Scott Brooks  
Sue Caroll  
Tonia Mehlhorn

Jizhong Zhou  
Yul Roh

Jack Carley

Terry Gentry

Bobette Nourse

NABIR Field Research Center

Argonne

Ken Kemner

Ken Lowe



David Watson

George Houser  
Kirk Hyder

Michigan State University

Terry Marsh  
James Tiedje  
Mary Beth Leigh

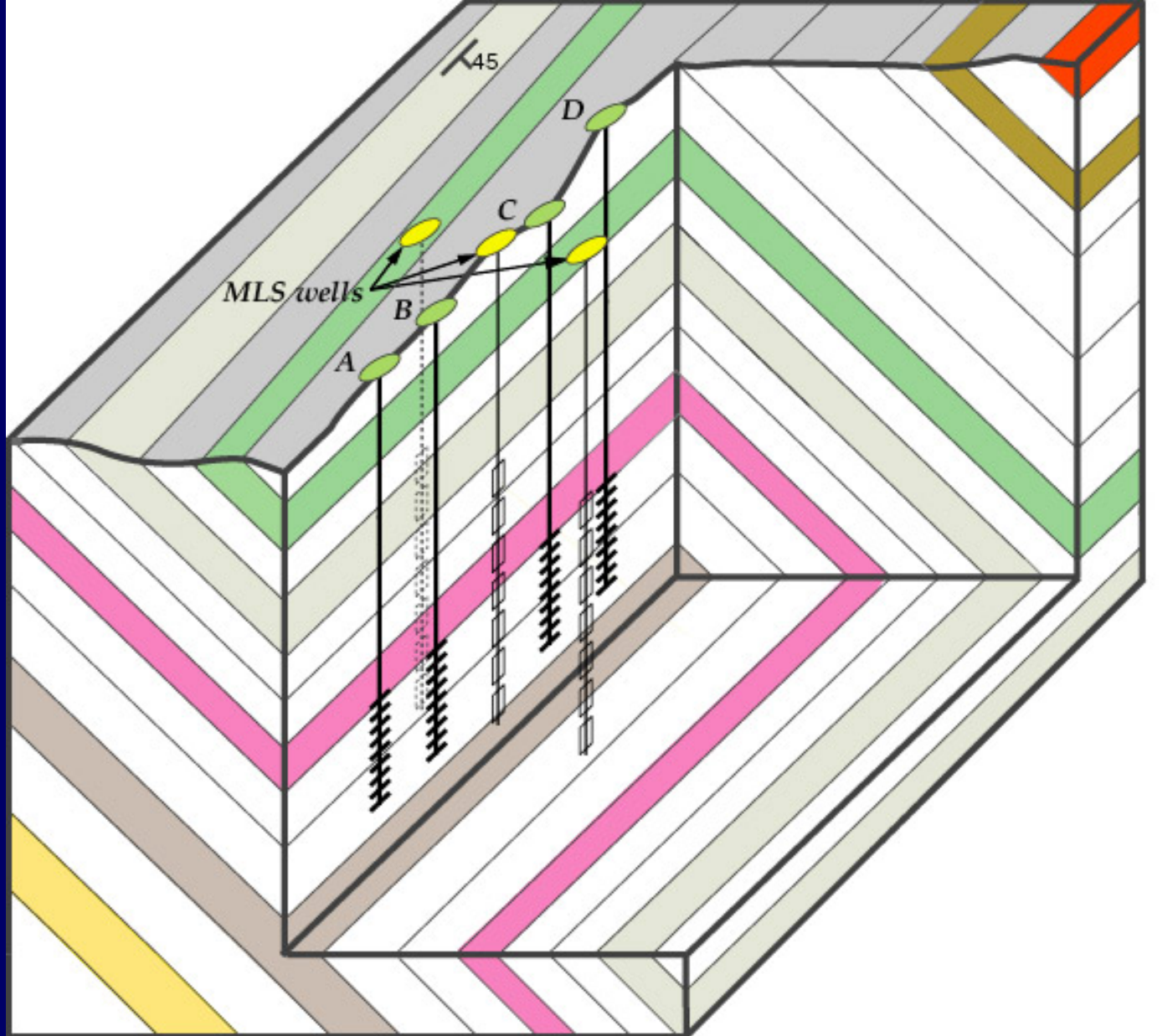


# Overview

- Selection of a treatment zone
- **Gaining hydraulic control**
- Conditioning
- Biostimulation

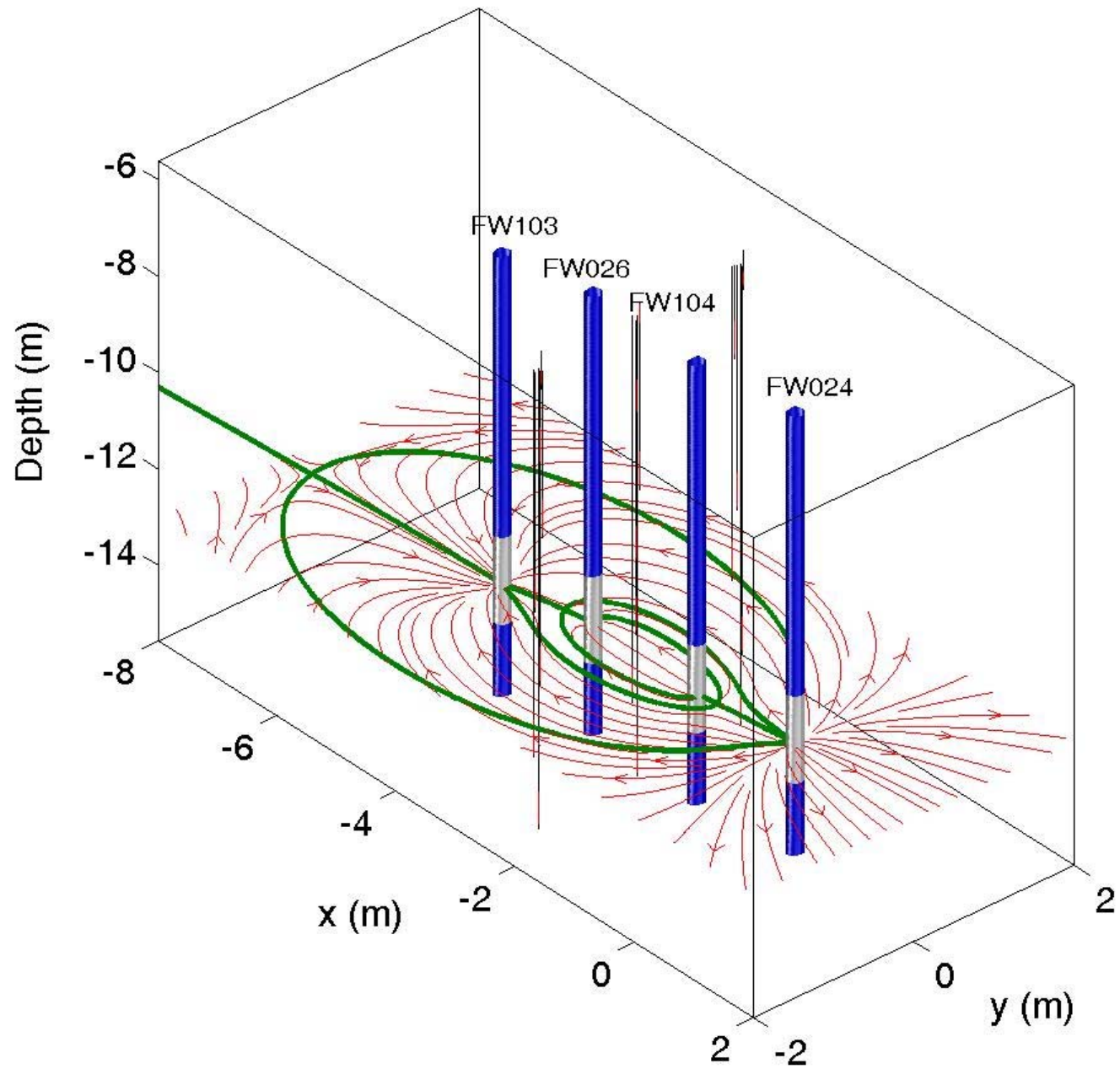
# Hydraulic control in a highly contaminated aquifer:

- Nested recirculation wells
- Aboveground removal of clogging agents and inhibitors
- Clean water tracer study
- Staged remediation



Screened  
Interval =  
38-45'

Cross-sectional view of the injection/extraction wells and the MLS wells.

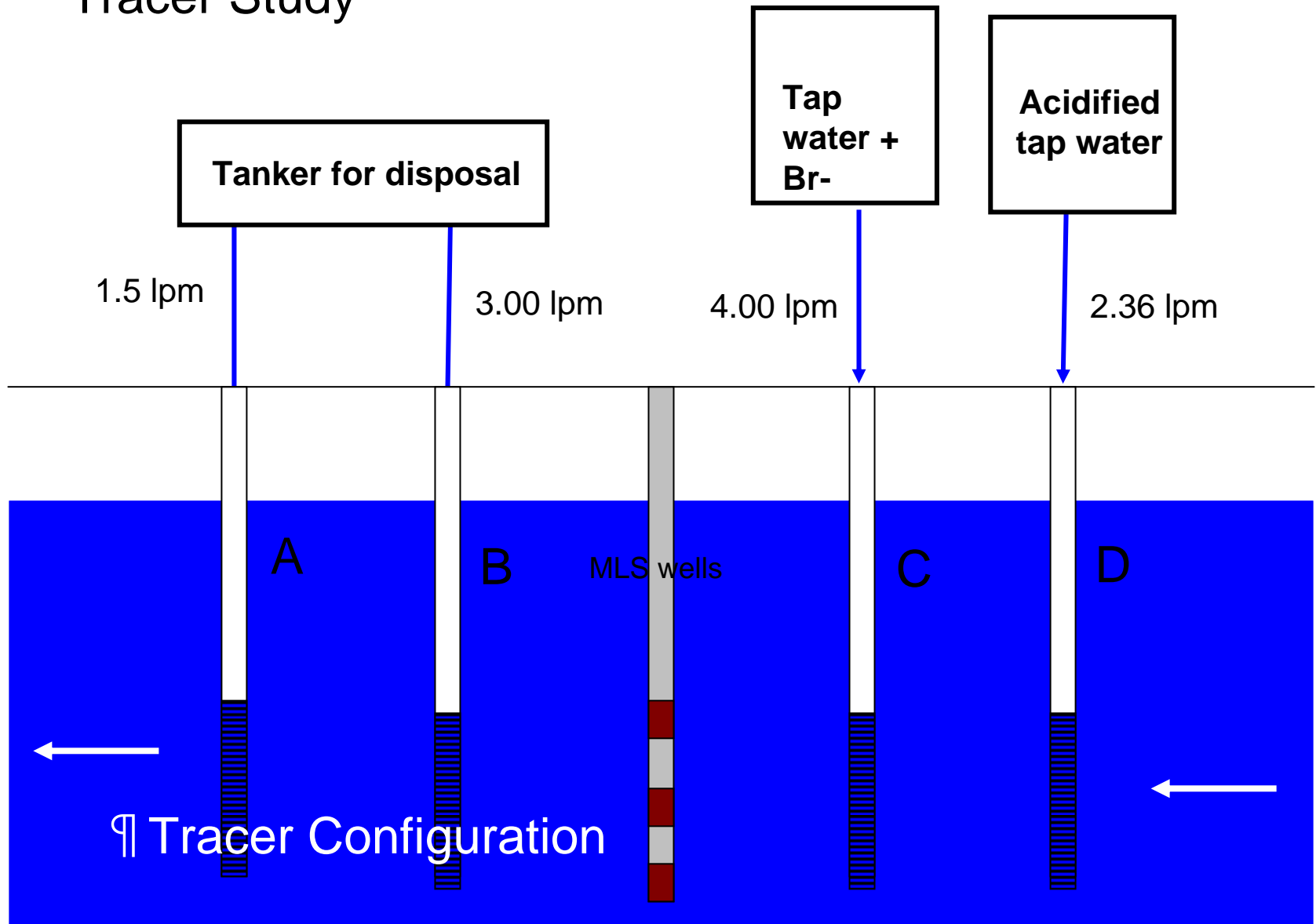




# 3D - nonuniform flow field

QuickTime™ and a  
Cinepak decompressor  
are needed to see this picture.

# Tracer Study





# MLS wells

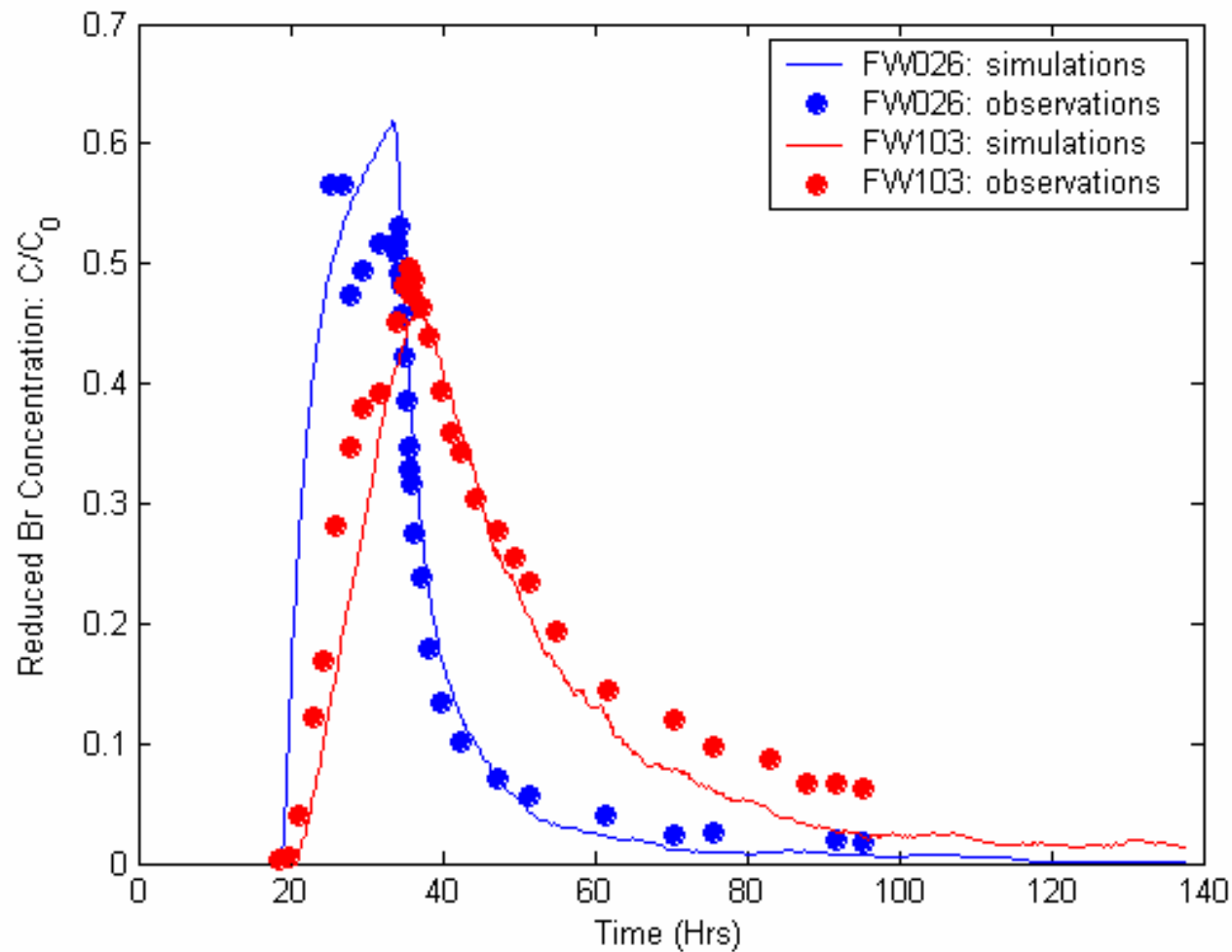


**Well B**

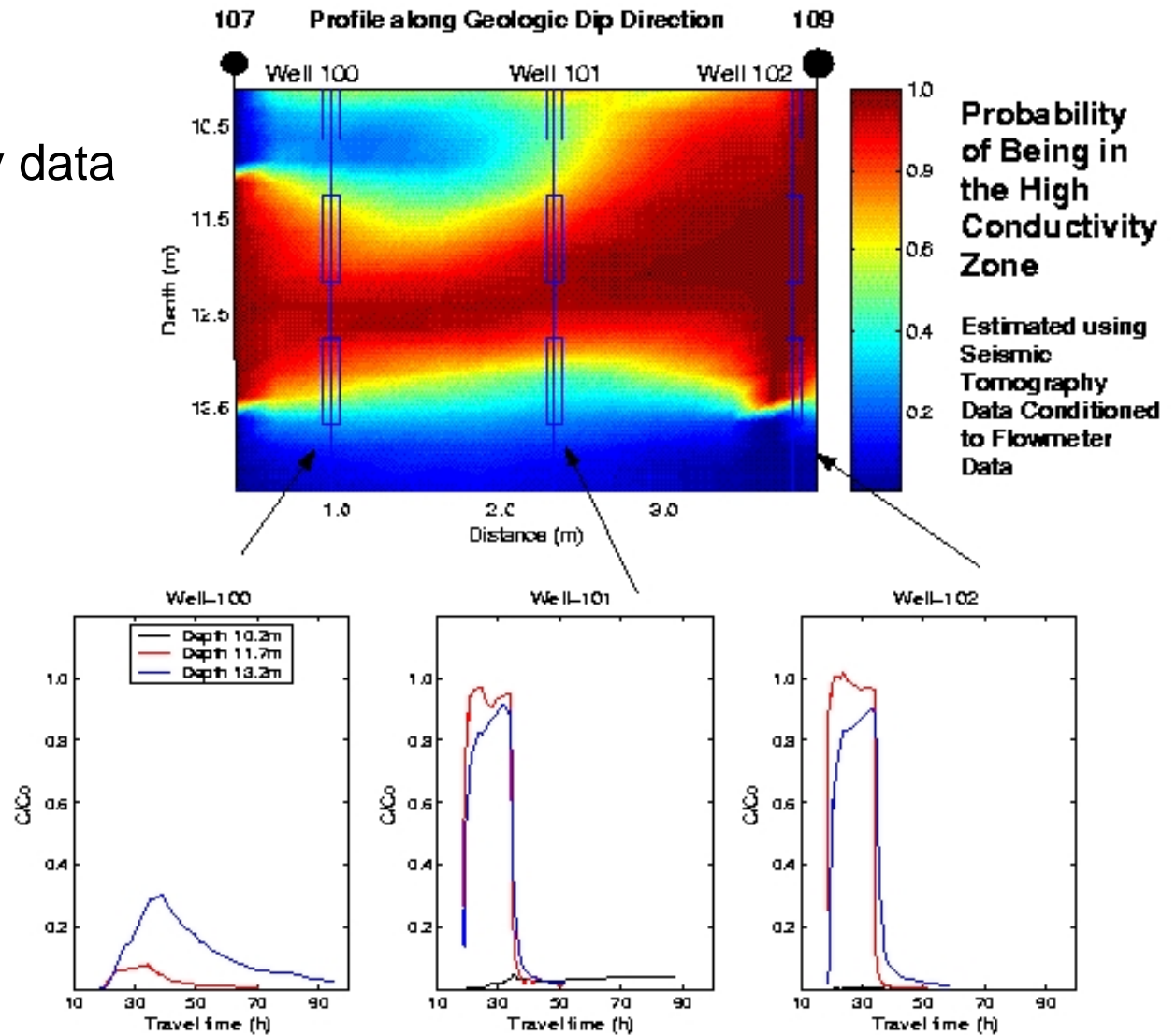
**Well C**

QuickTime™ and a  
TIFF (PackBits) decompressor  
are needed to see this picture.

# Tracer study simulations



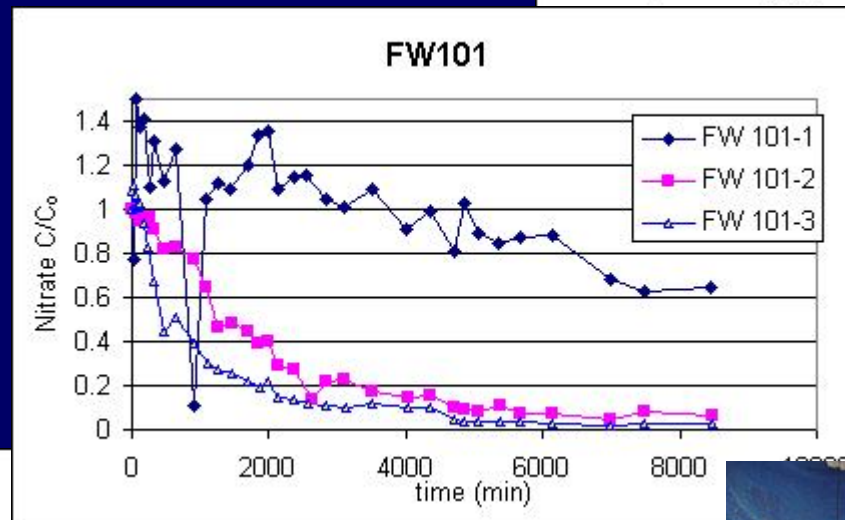
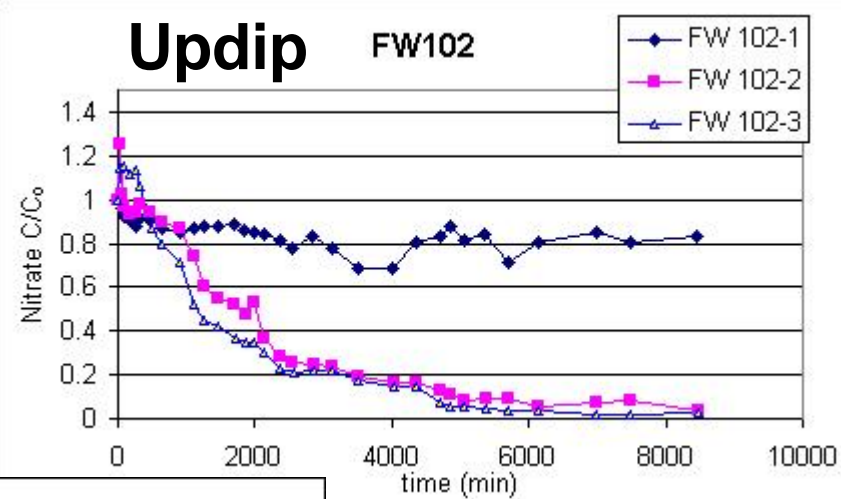
Seismic tomography data complements tracer measurements.



Hubbard et al.,  
2003  
Mehnhorn et al., 2003

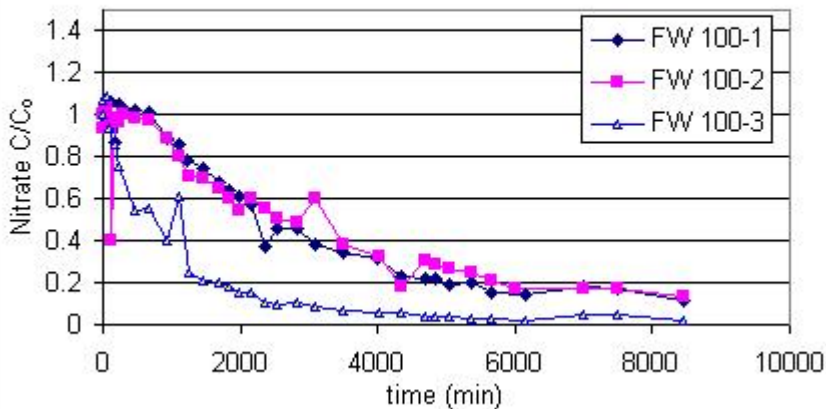
Tracer Breakthrough at 3 Multi-Level Samplers along Geologic Dip Direction

# Effect of tracer clean water flush on nitrate in MLS wells



Mid-depths were flushed well  
Bottom depth was poorly flushed

### Downdip FW100



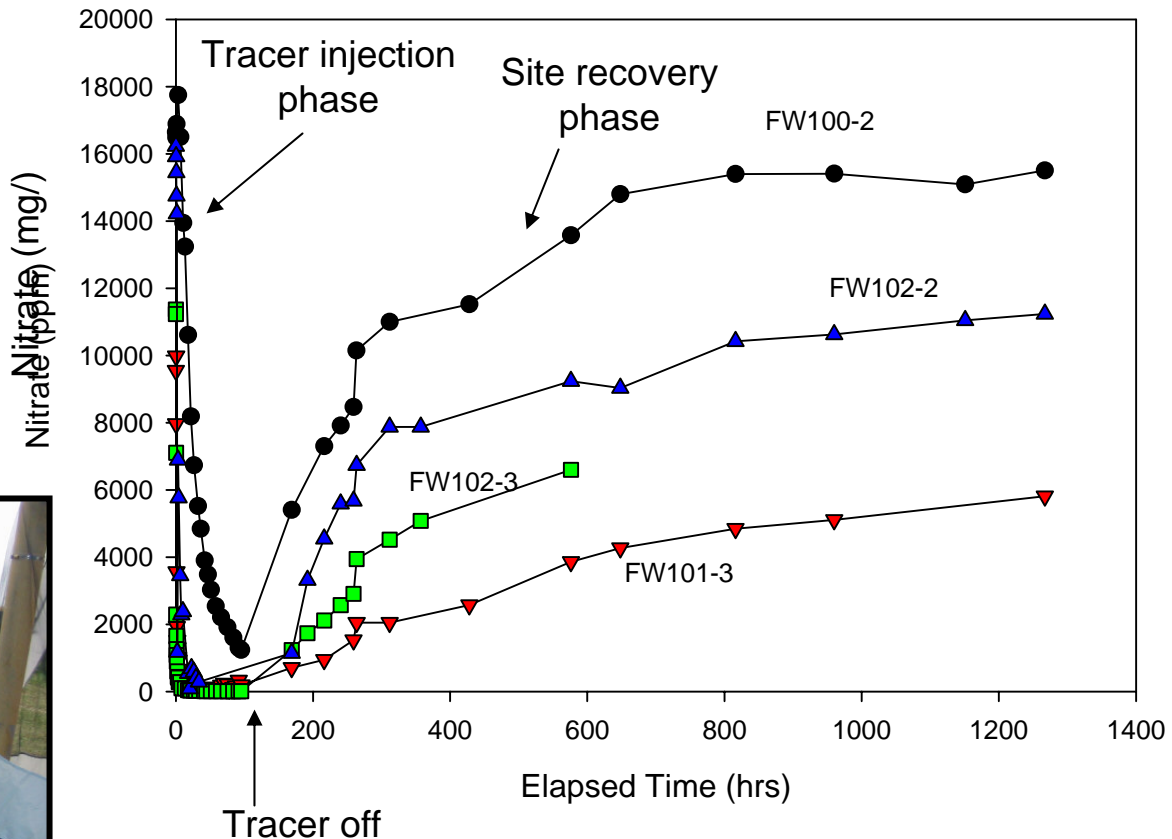
All depths were flushed

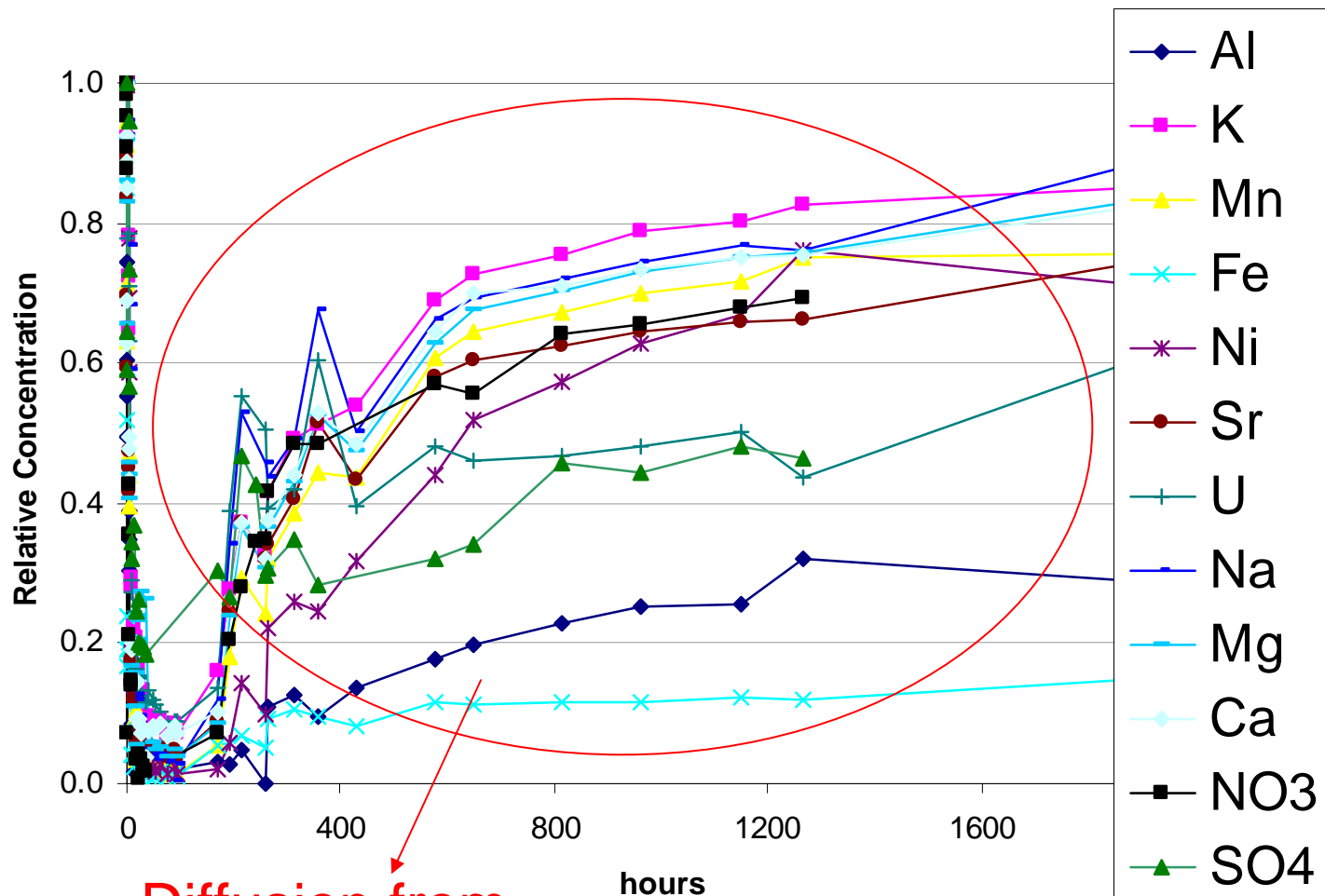


# Natural gradient site recovery solute breakthrough

Natural gradient  
contaminant transport  
monitored during site  
recovery.

Quantification of solute  
residence times, direction  
of groundwater flow, and  
strike vs. dip interactions.



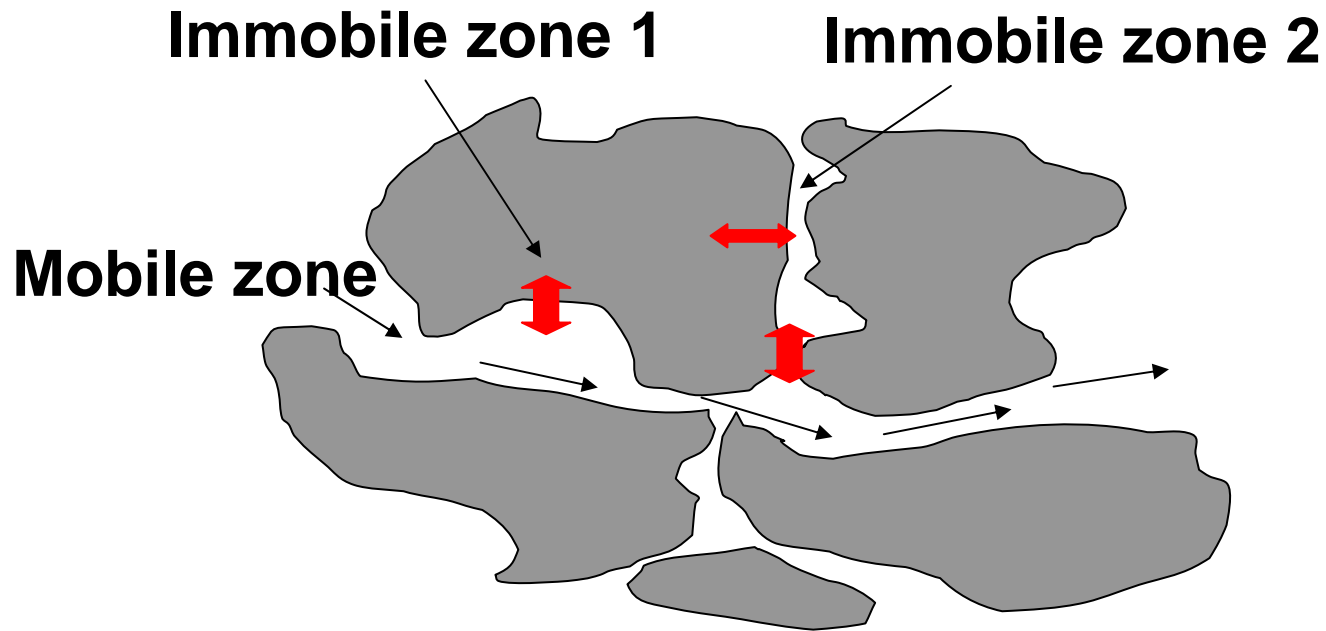


Diffusion from matrix

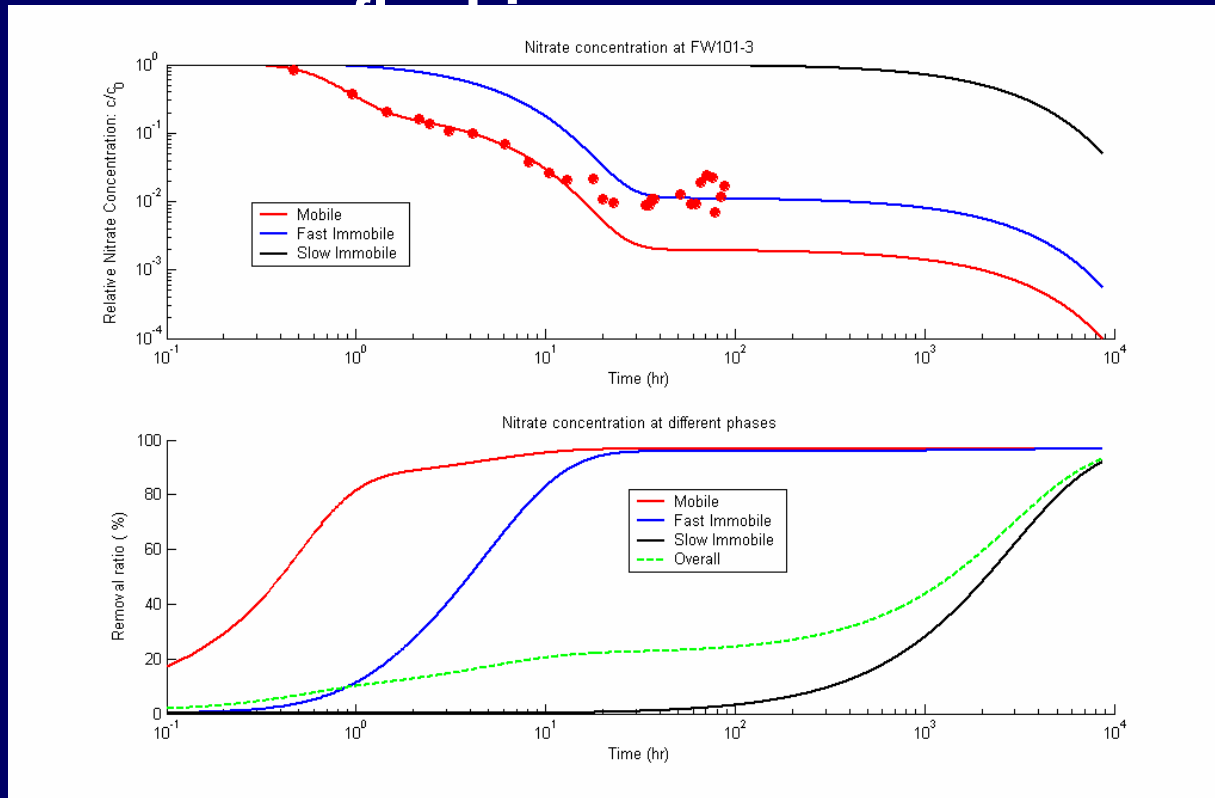
# Mass transfer during the flush

## Model assumptions:

- Kinetically controlled sorption/desorption
- Kinetic mass transfer between two regions



# Modeling of



- The half-life of nitrate in the second immobile region is about 3 months. To deplete the second immobile zone would take about one year.
- The mobile region definitely responds to flushing and a low average Nitrate concentration can be maintained while removing the Nitrate as it enters the mobile zone.



# Overview

- Selection of the treatment zone
- Gaining hydraulic control
- **Conditioning**
- Biostimulation

# Conditioning - removal of clogging agents, inhibitors, adjustment of pH

## 1. Recirculate and flush at pH 4-4.5

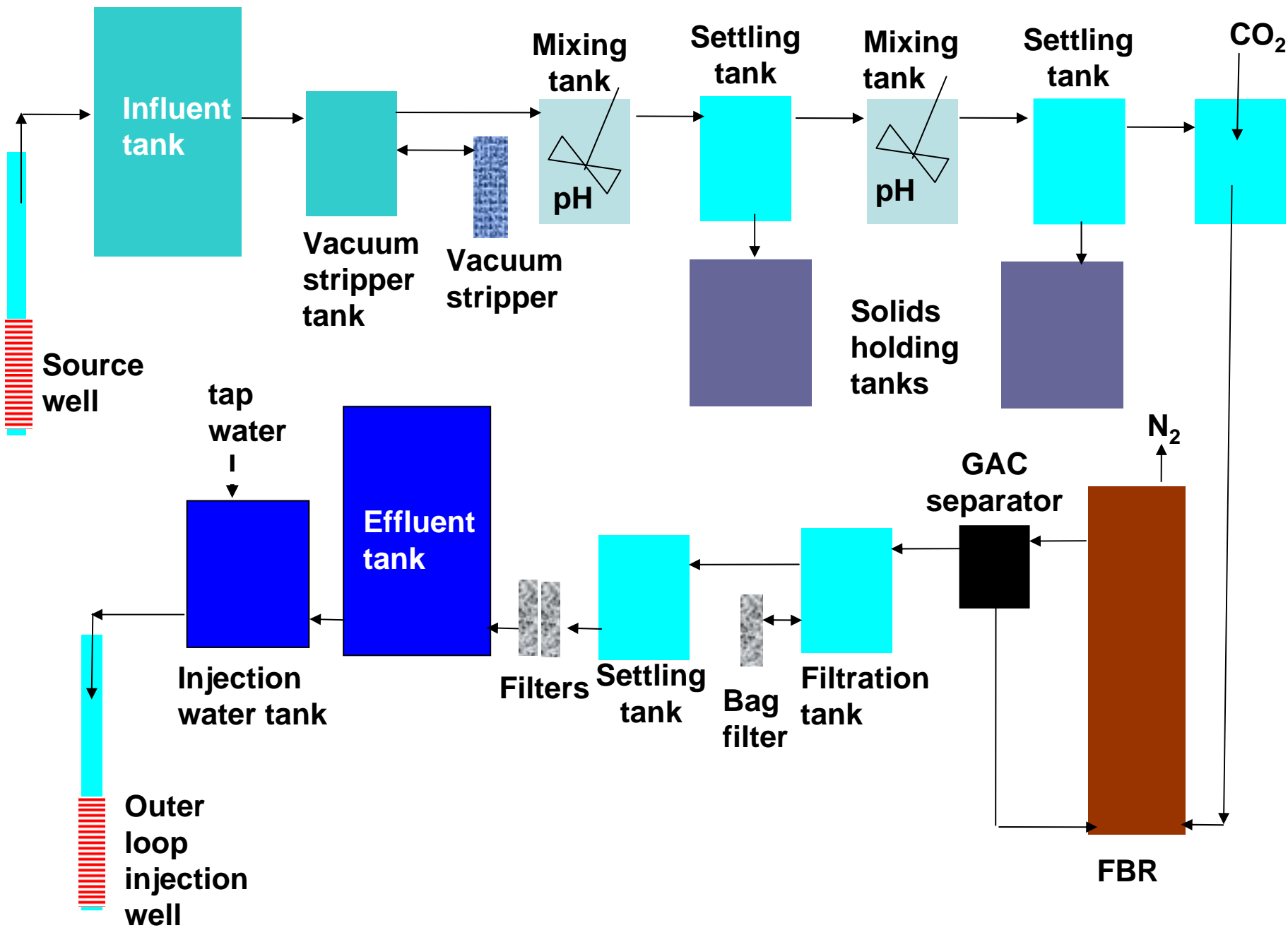
Precipitate Al and Ca *ex-situ*

Remove  $\text{NO}_3^-$  by denitrification in FBR

Vacuum strip to remove VOCs and  $\text{N}_2$

## 2. Recirculate and flush at pH 6-6.3

# ABOVEGROUND PROCESS TRAIN



# Clogging agents

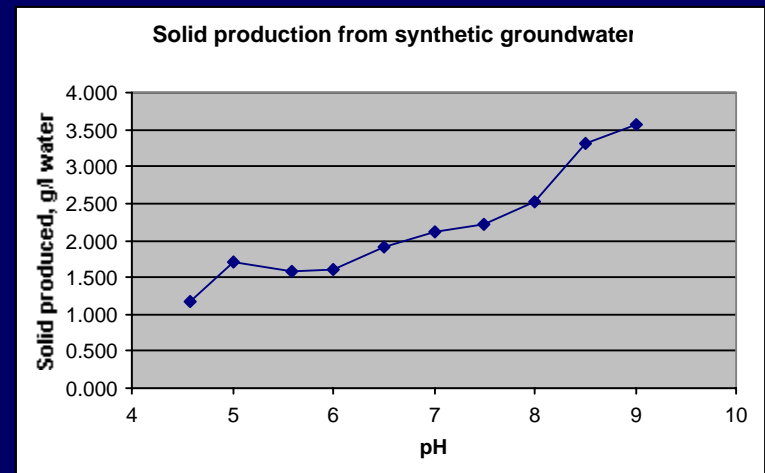
- Aluminum hydroxide form at pH 5.
- Calcium and magnesium carbonates form at pH 7-9.
- N<sub>2</sub> gas forms during denitrification.
- High levels of biomass are produced during denitrification.



pH adjusted to 7 with 50% liquid from denitrifying batch cultures

pH adjusted to 7 with Na<sub>2</sub>CO<sub>3</sub>

pH adjusted to 7 with KOH



2 g/L solids produced

## Tanker for chemical sludge disposal



## Bag filters for disposal of biomass

The “Big Top” where extracted groundwater is treated to enable metal reduction *in-situ*



# Inside the Big Top



# The aboveground treatment train



Vacuum stripper



Two-step  
chemical precipitation



Fluidized bed reactor  
(FBR)

**Well TPB16  
enrichment** → **innoculum**



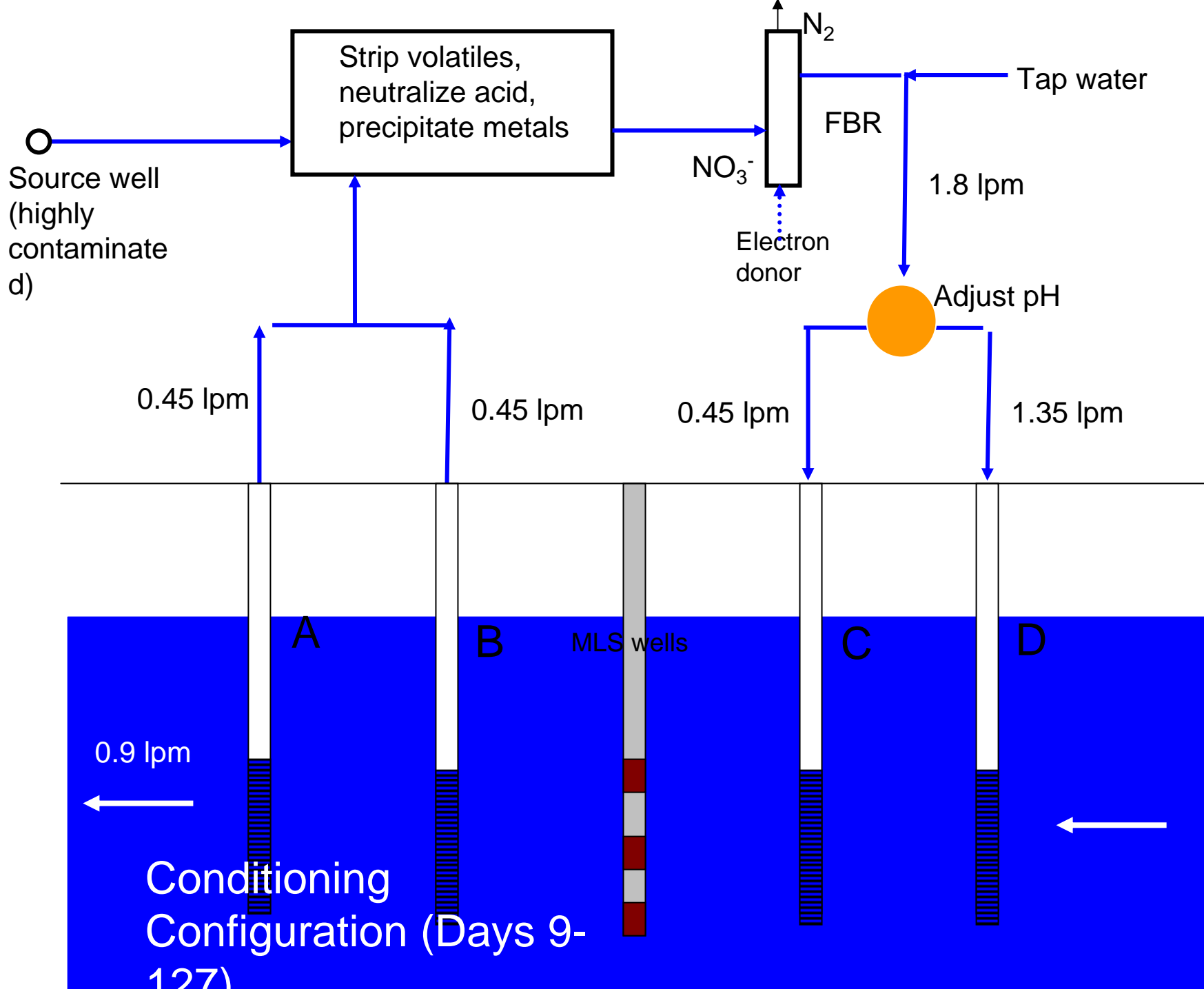
**Two pilot scale  
FBRs**

**innoculum** →

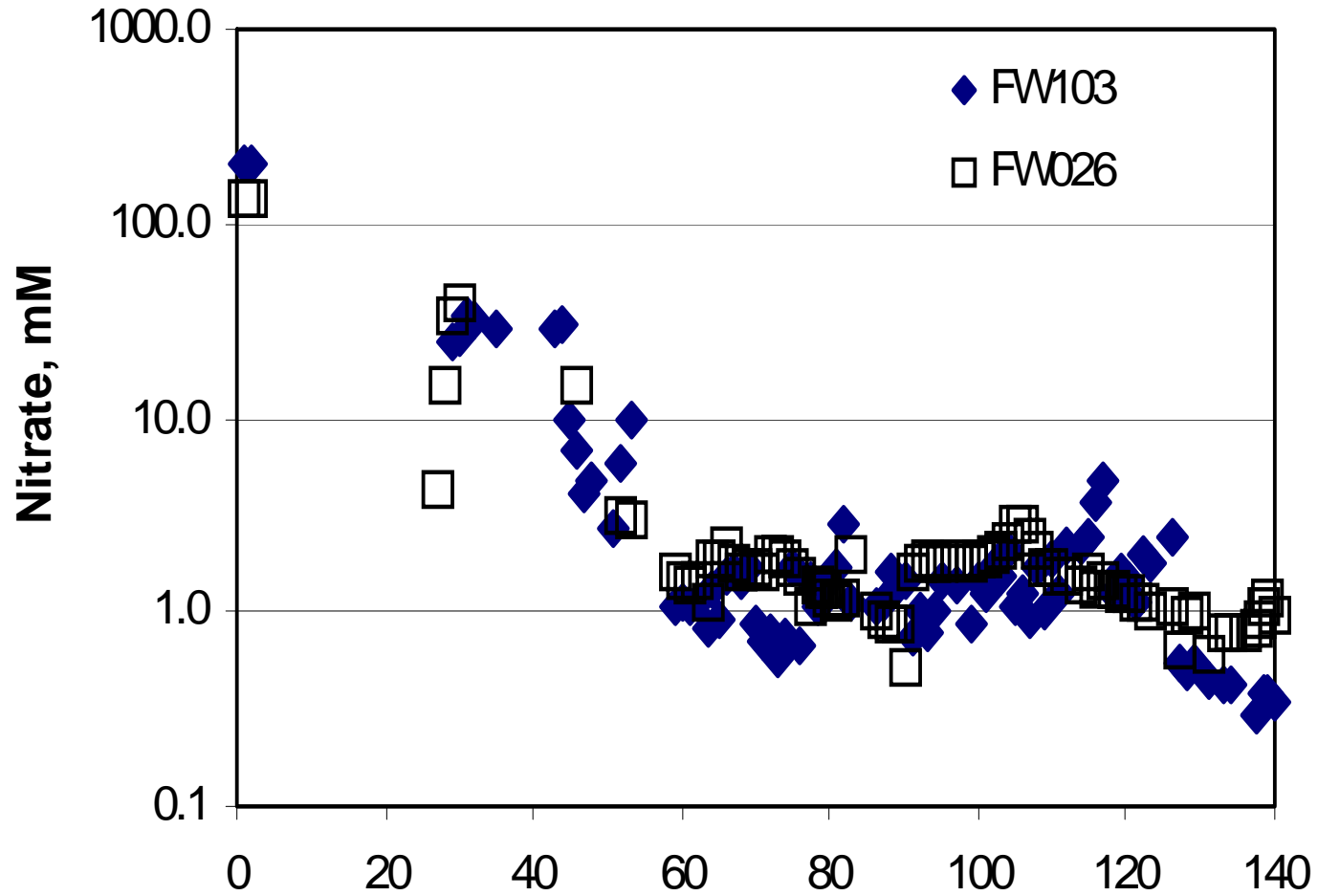


**Full scale  
FBR**

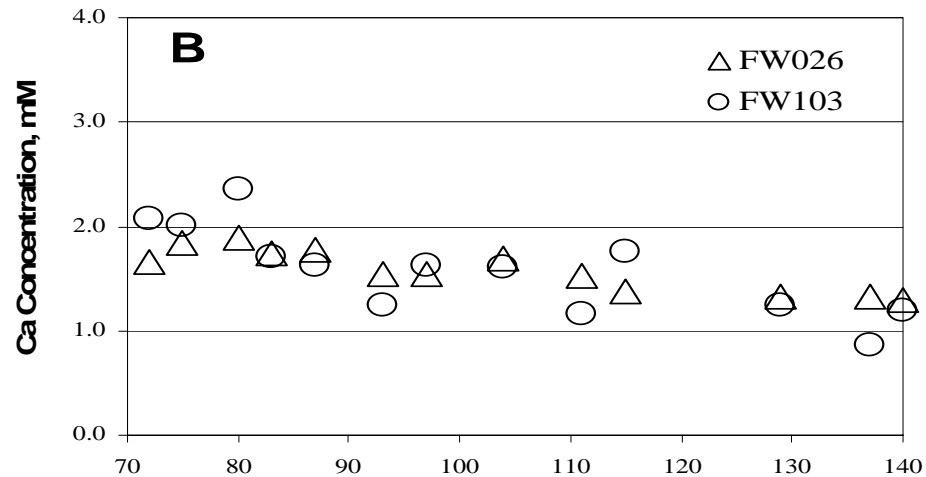
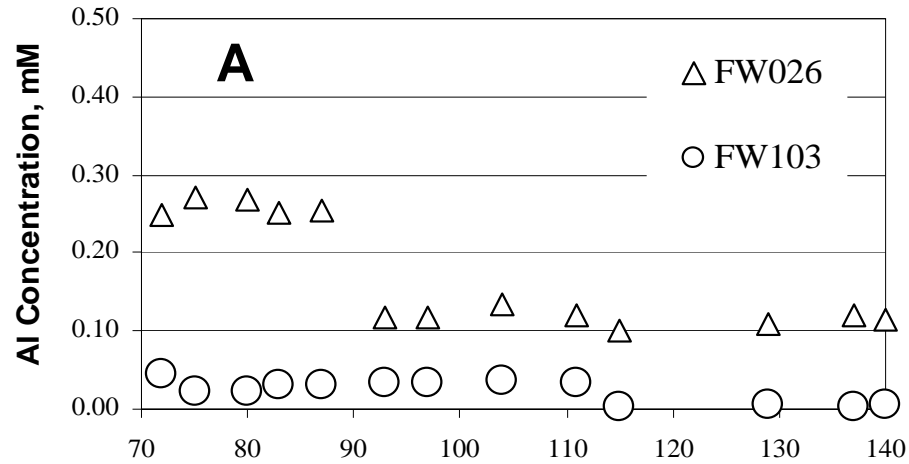




# Nitrate removal at injection extraction wells during condi

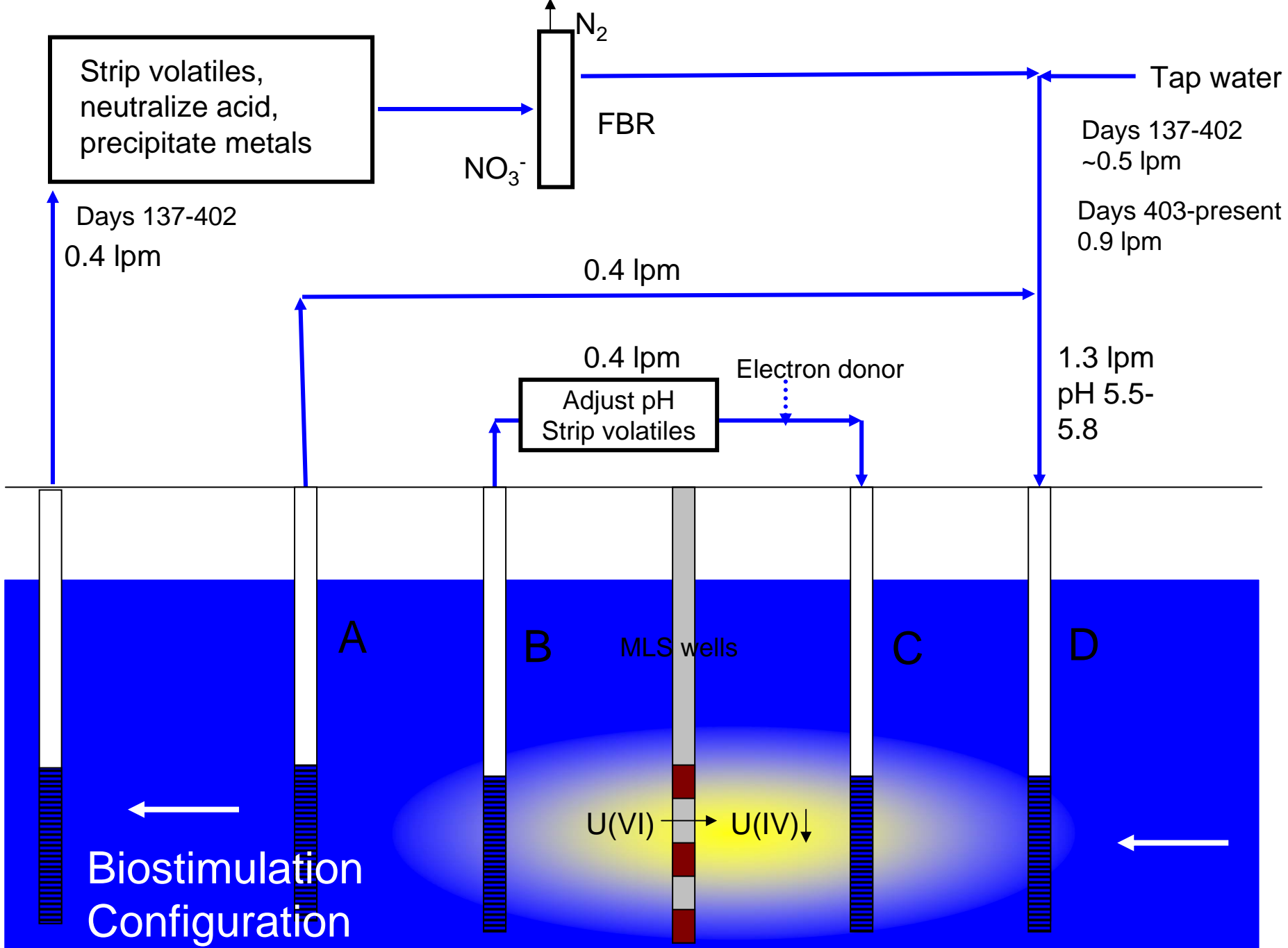


# Al and Ca removal at injection extraction wells during cond

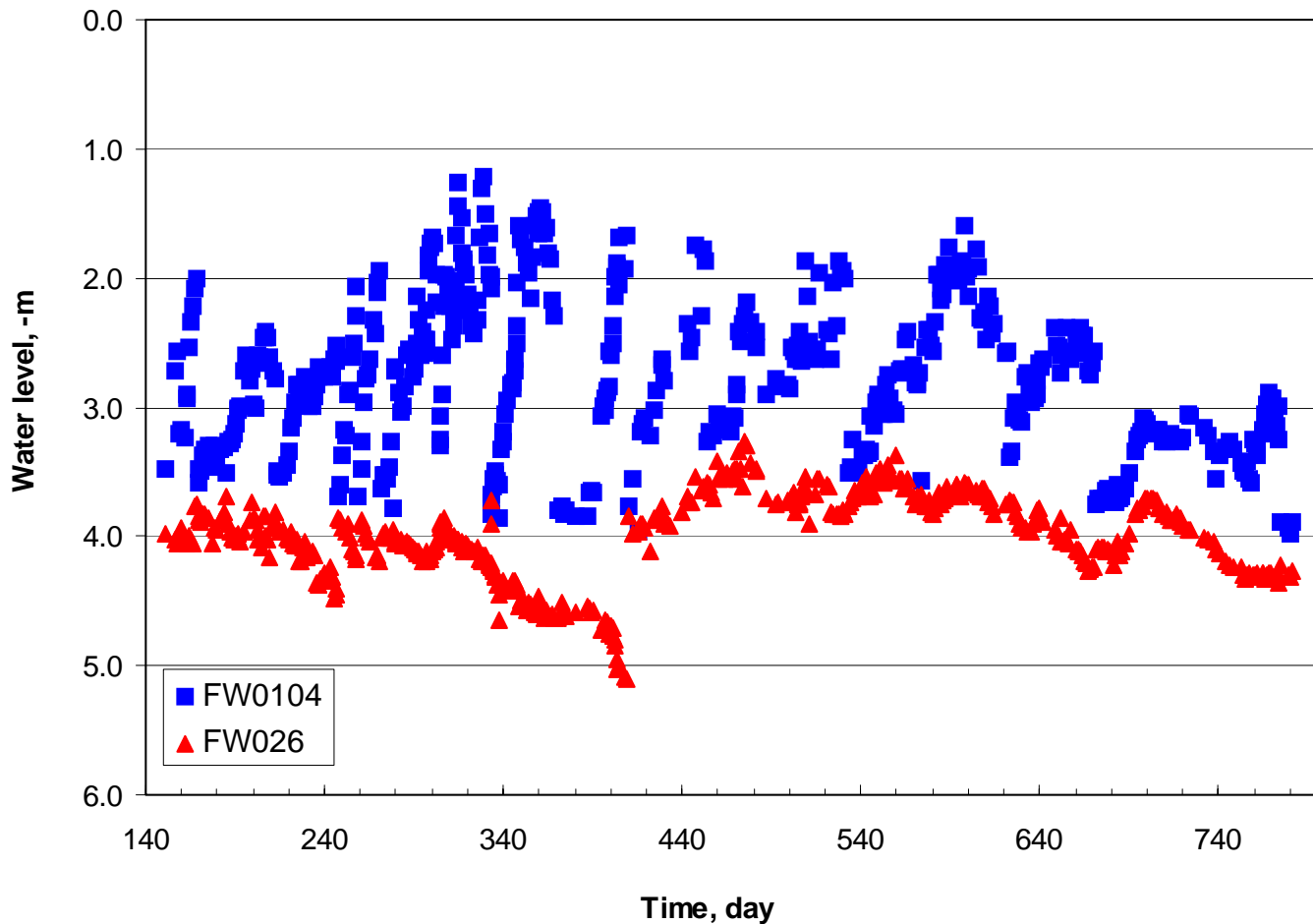


# Overview

- Selection of the treatment zone
- Gaining hydraulic control
- Conditioning
- **Biostimulation**



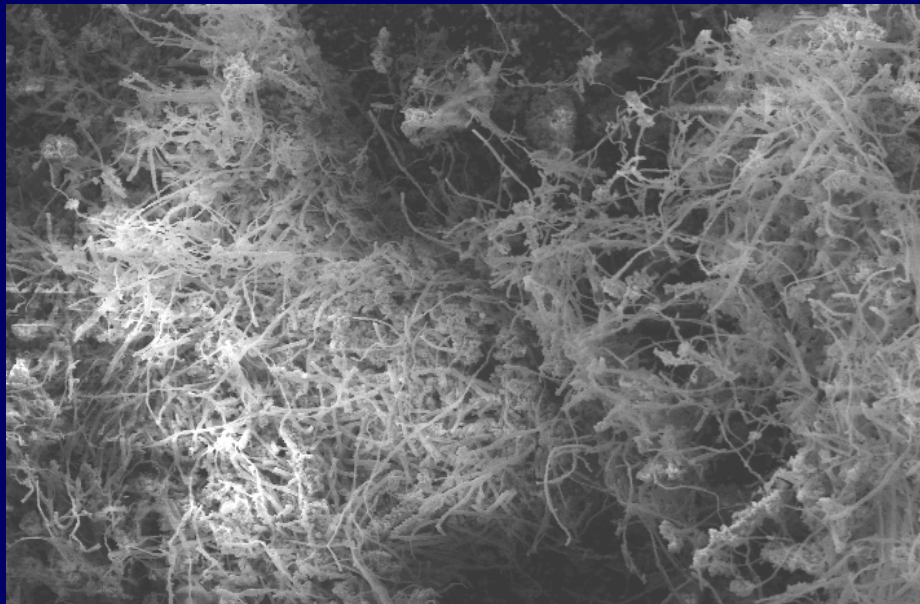
# Water level in inner loop injection and extraction wells during biostimulation



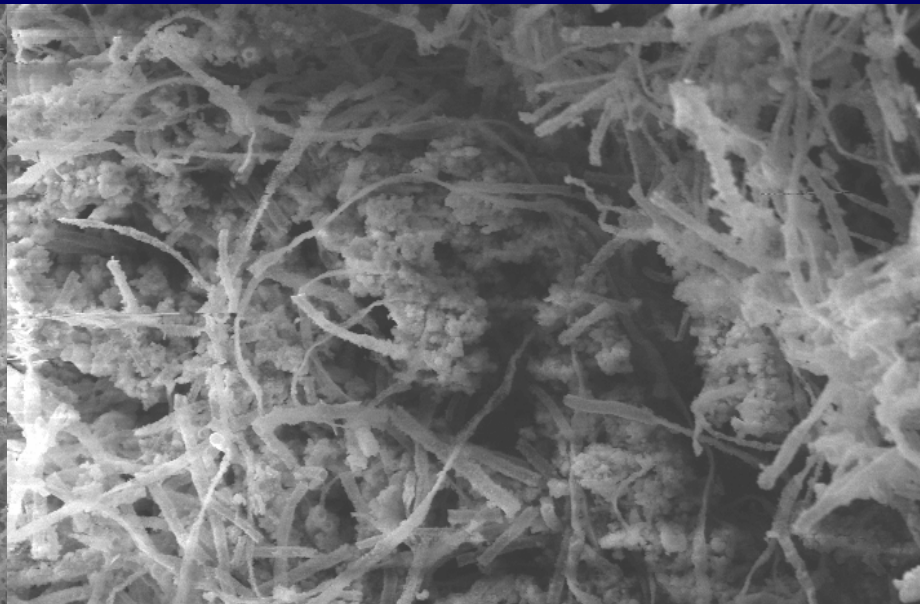
Surge block for cleaning



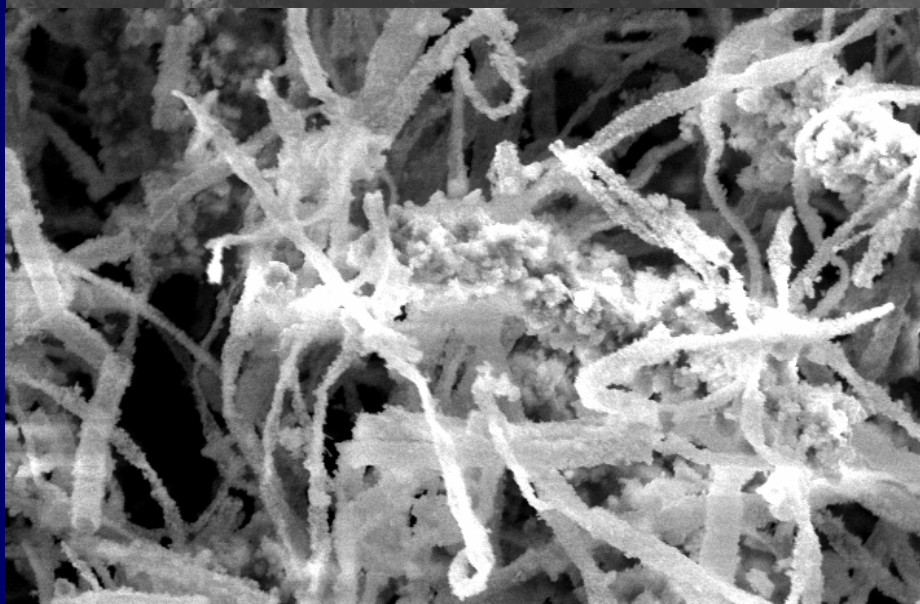
Clogged pump head screen.  
The white precipitate dissolved in  
a 2% HCl solution after 1.5 hour.



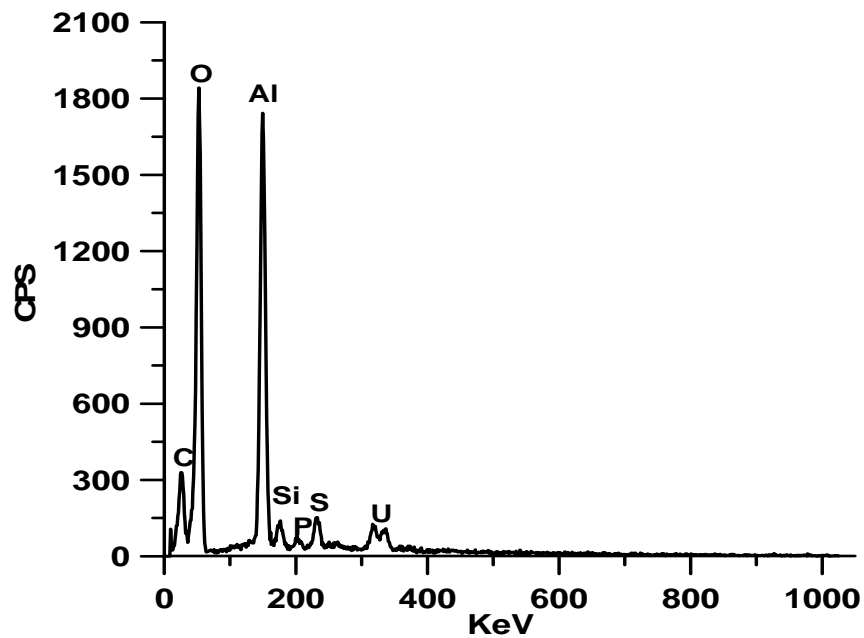
Acc.V Spot Magn Det WD Exp |-----| 100  $\mu$ m  
15.0 kV 4.0 150x SE 9.7 39845 Roh



Acc.V Spot Magn Det WD Exp |-----| 50  $\mu$ m  
15.0 kV 4.0 500x SE 9.7 39846 Roh

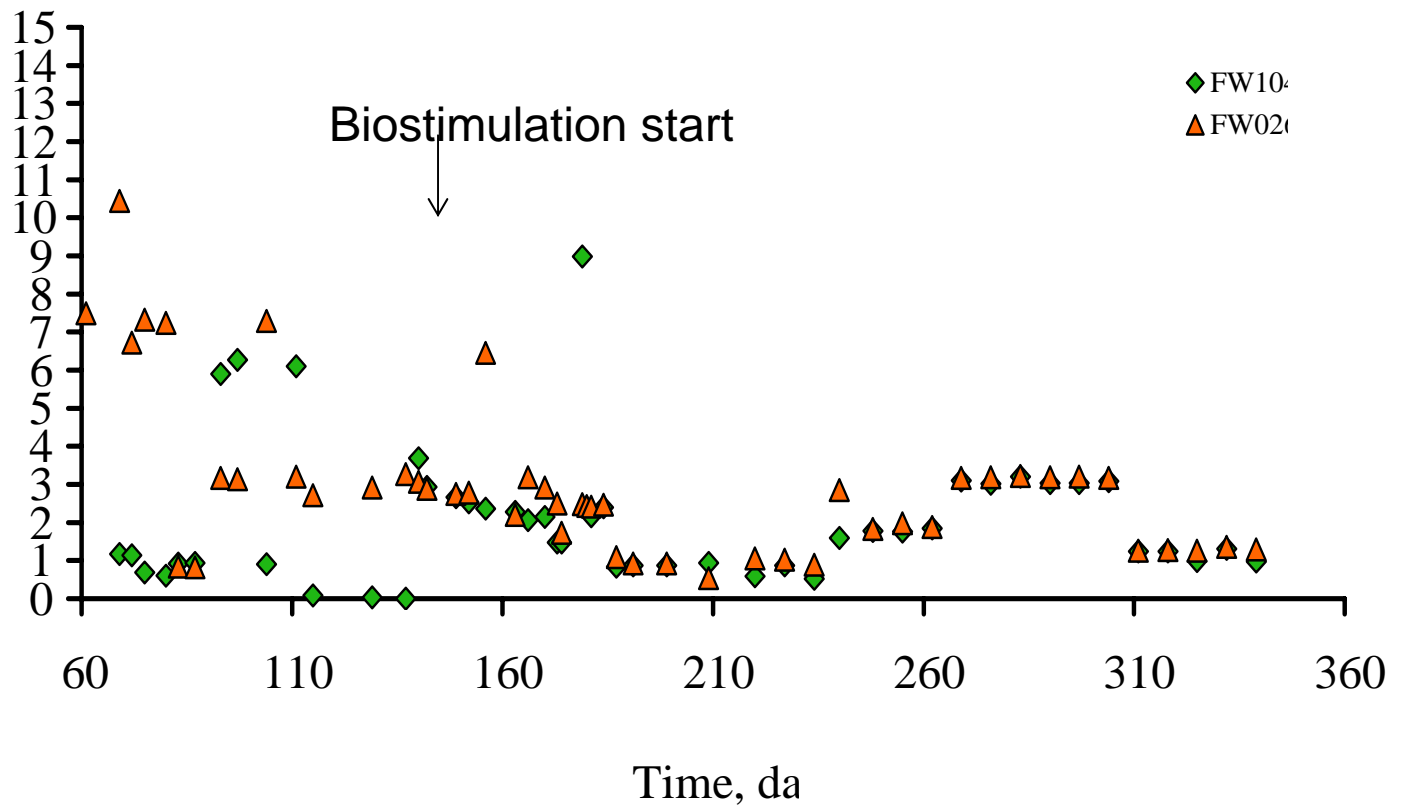


Acc.V Spot Magn Det WD Exp |-----| 20  $\mu$ m  
15.0 kV 4.0 1000x SE 9.5 39849 Roh





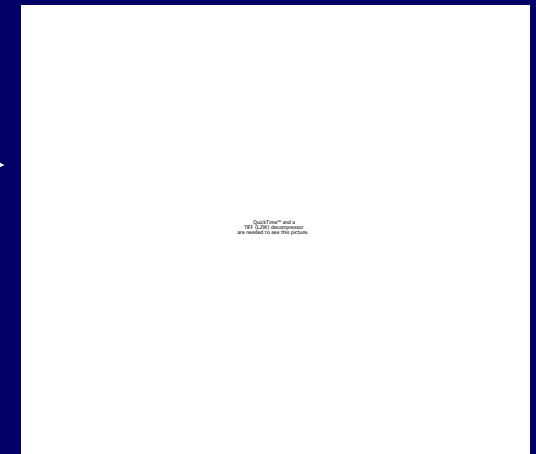
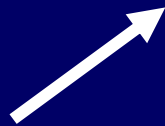
# Aluminum in inner loop injection and extraction wells



Biofouling of pump intake on inner loop extraction well - Day 245



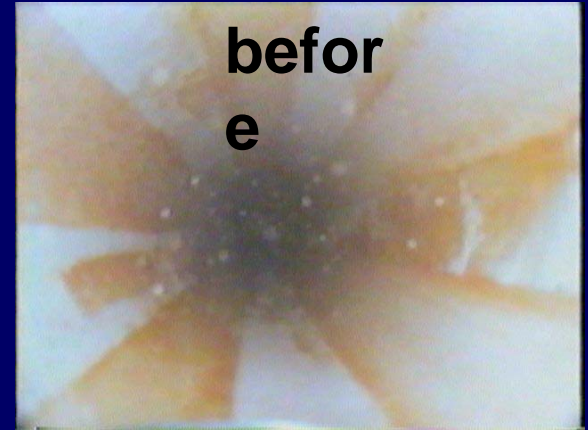
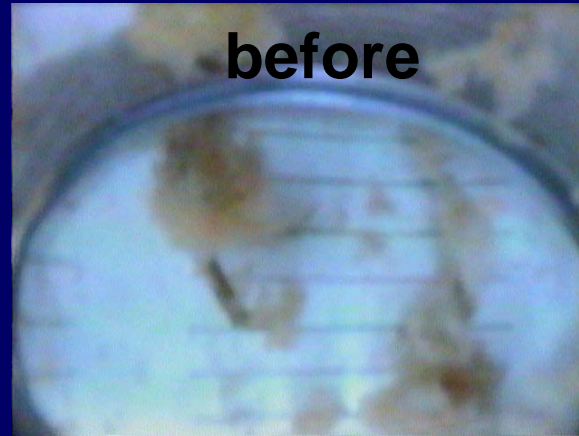
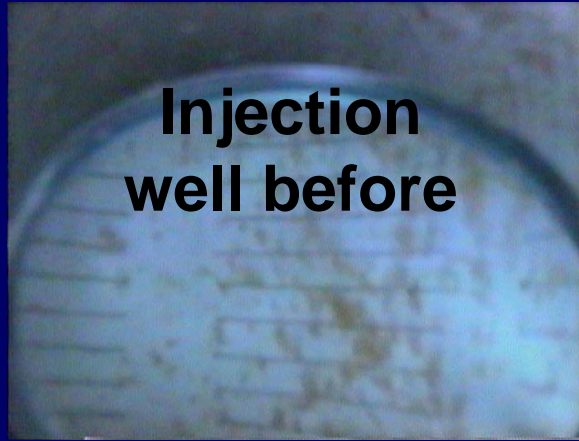
Surge block  
allowed for  
sampling of  
sediment/biomas  
s in wells



# Effects of surging: what a borehole camera shows

## reference well

QuickTime™ and a TIFF (LZW) decompressor are needed to see this picture.



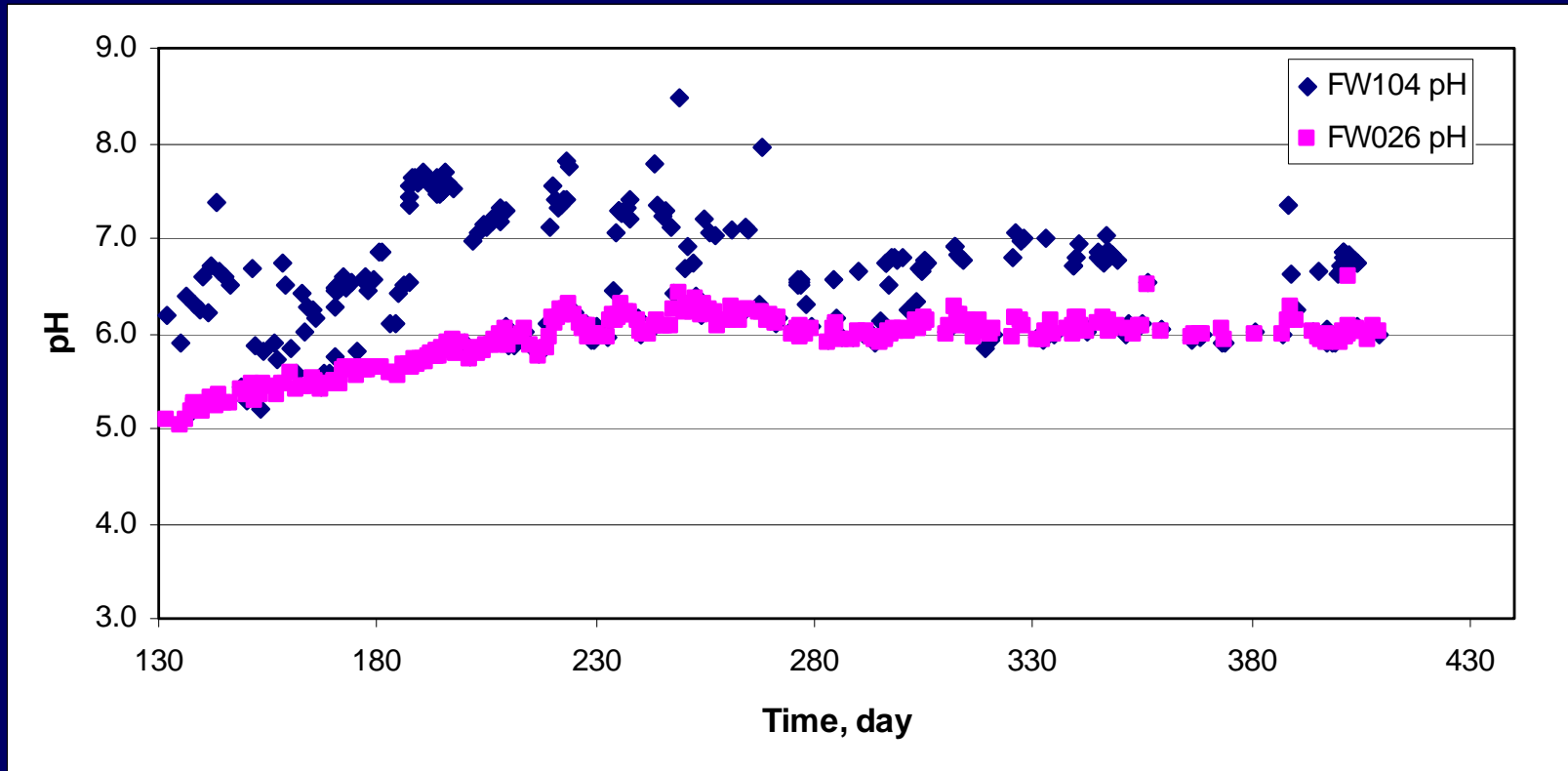
after

QuickTime™ and a TIFF (LZW) decompressor are needed to see this picture.

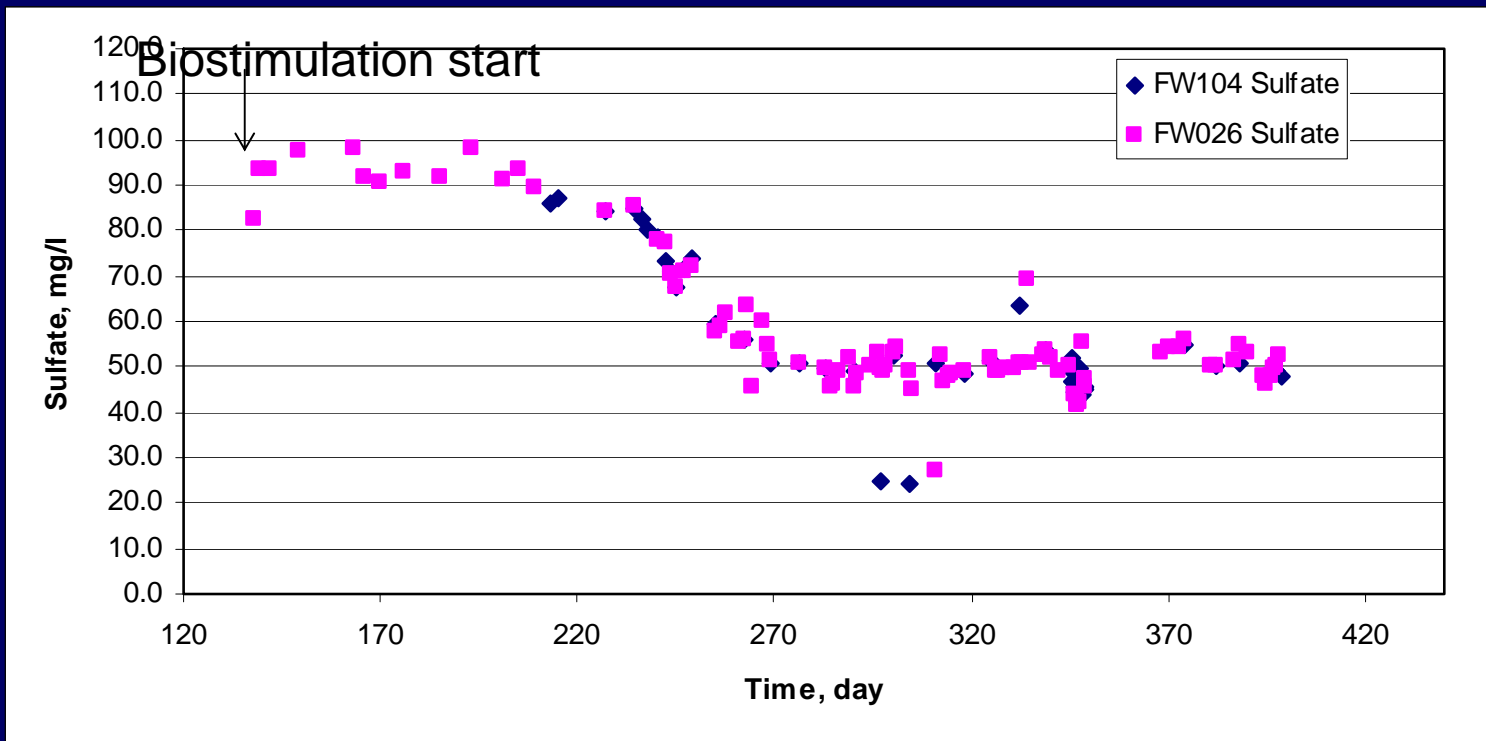
after

QuickTime™ and a TIFF (LZW) decompressor are needed to see this picture.

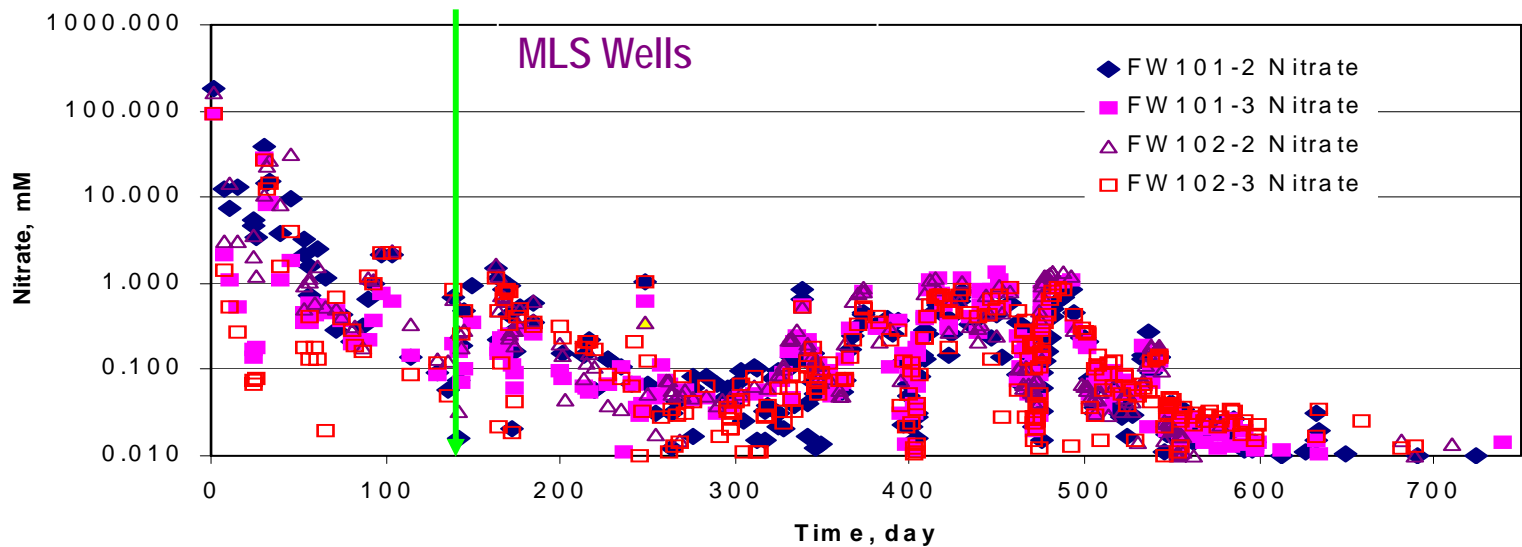
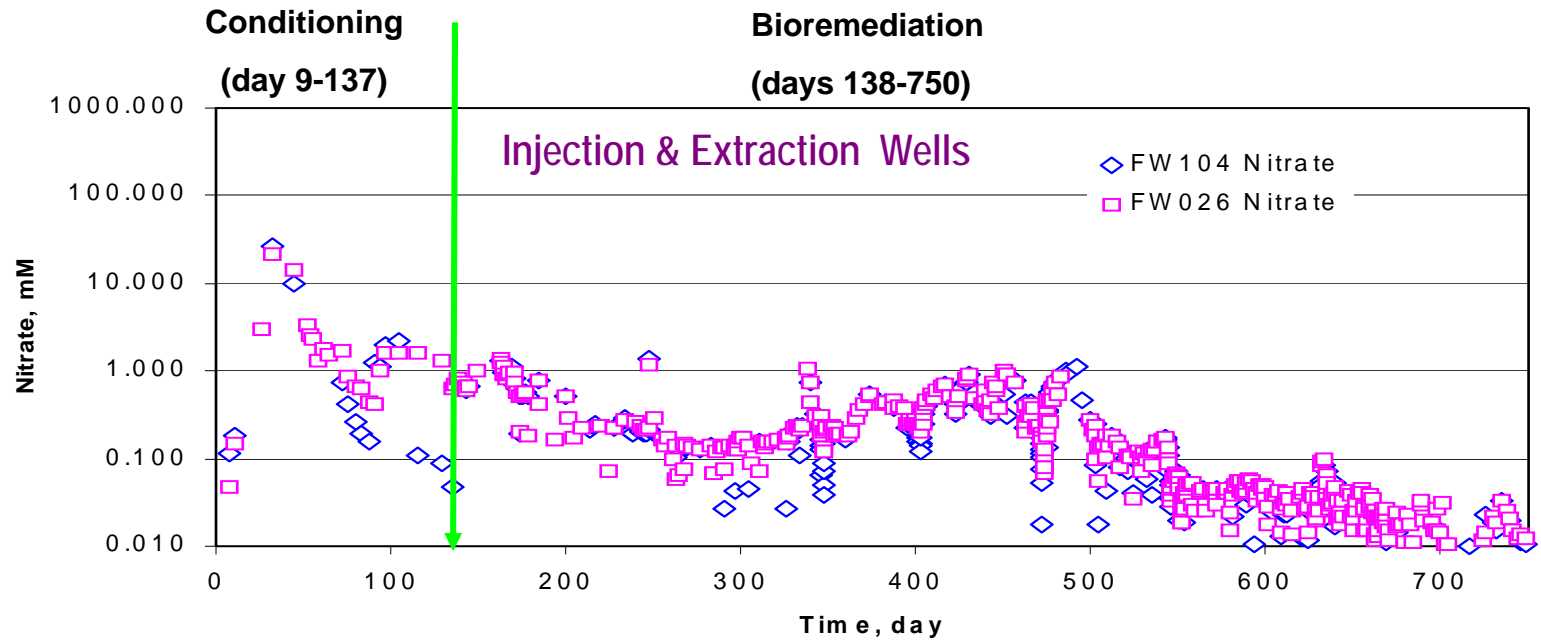
# pH in inner loop injection and extraction wells during biostimulation



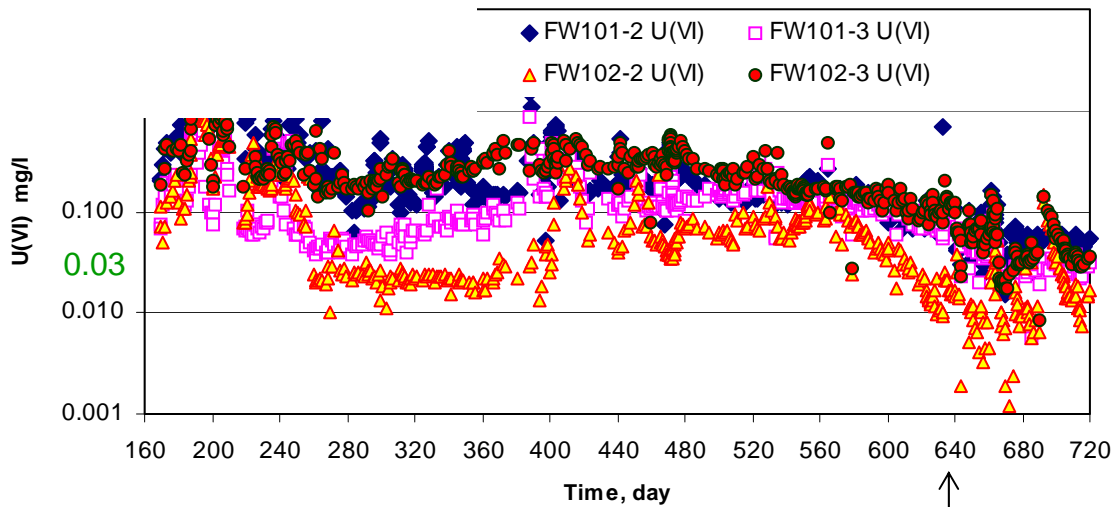
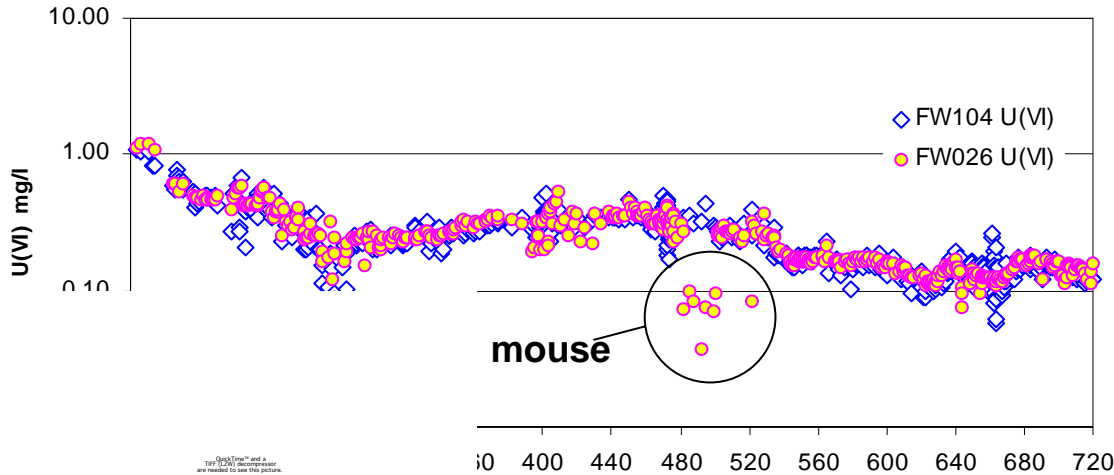
# Sulfate in inner loop injection and extraction wells



# Nitrate removal during biostimulation



# Dissolved U(VI) concentrations during biostimulation (Day 160-preset)



## Key Findings

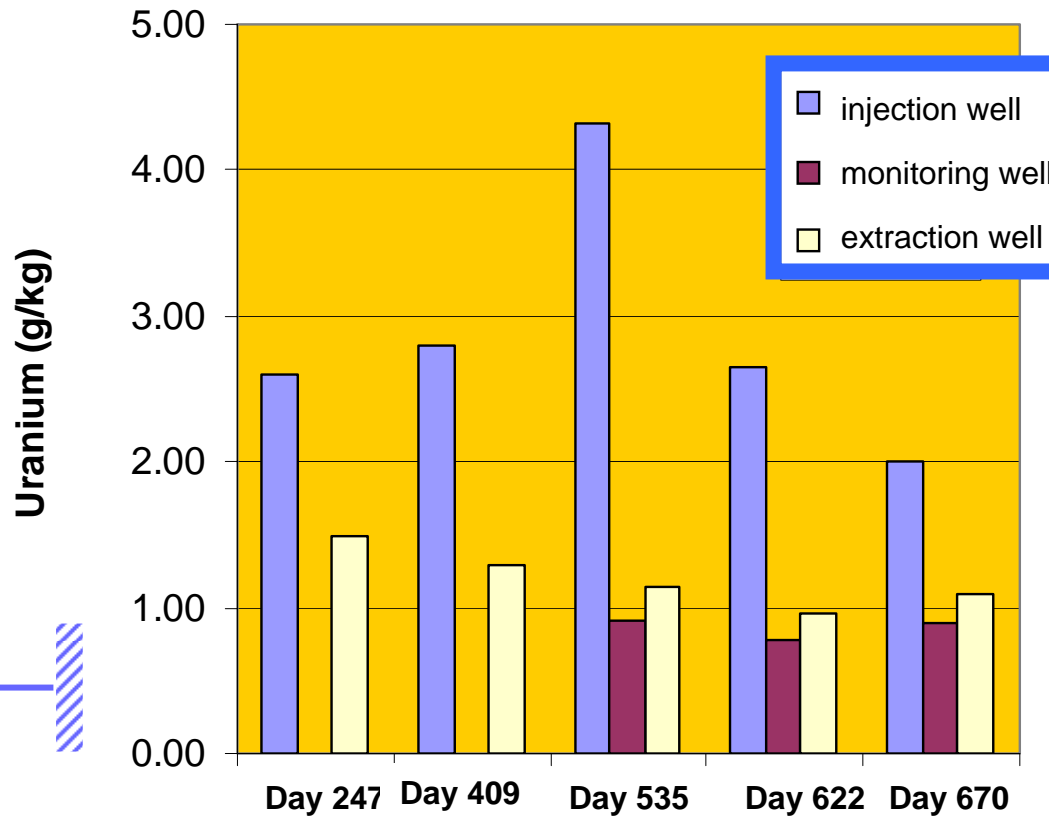
1. Ethanol addition stimulated *In situ* bioreduction of U(VI).
2. U(VI) concentration dropped below EPA MCL.
3. Sulfate reduction and Fe(III) Reduction were concomitant with U(VI) reduction.
4. U(IV) was stable under controlled anaerobic conditions

**Maximum concentration of uranium in drinking water of 0.03 mg/l (US EPA) is achievable.**

**O<sub>2</sub> removed from outer loop**



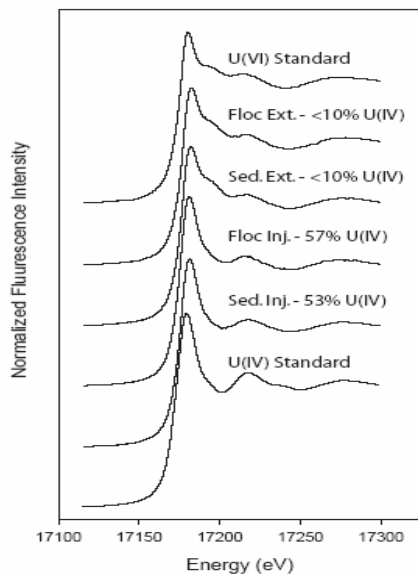
# Uranium Content in Sediments from Injection, Monitoring and Extraction Wells during Bioremediation



Uranium content in core samples from FW104 was in the range of 0.03-0.8 g/kg before bioremediation

Variability in the content of U in the injection well sediment is likely due to variability in the removal of U during the surging operation.

## XANES analysis of sediment samples taken from inner loop injection well confirms U(VI) reduction to U(IV)

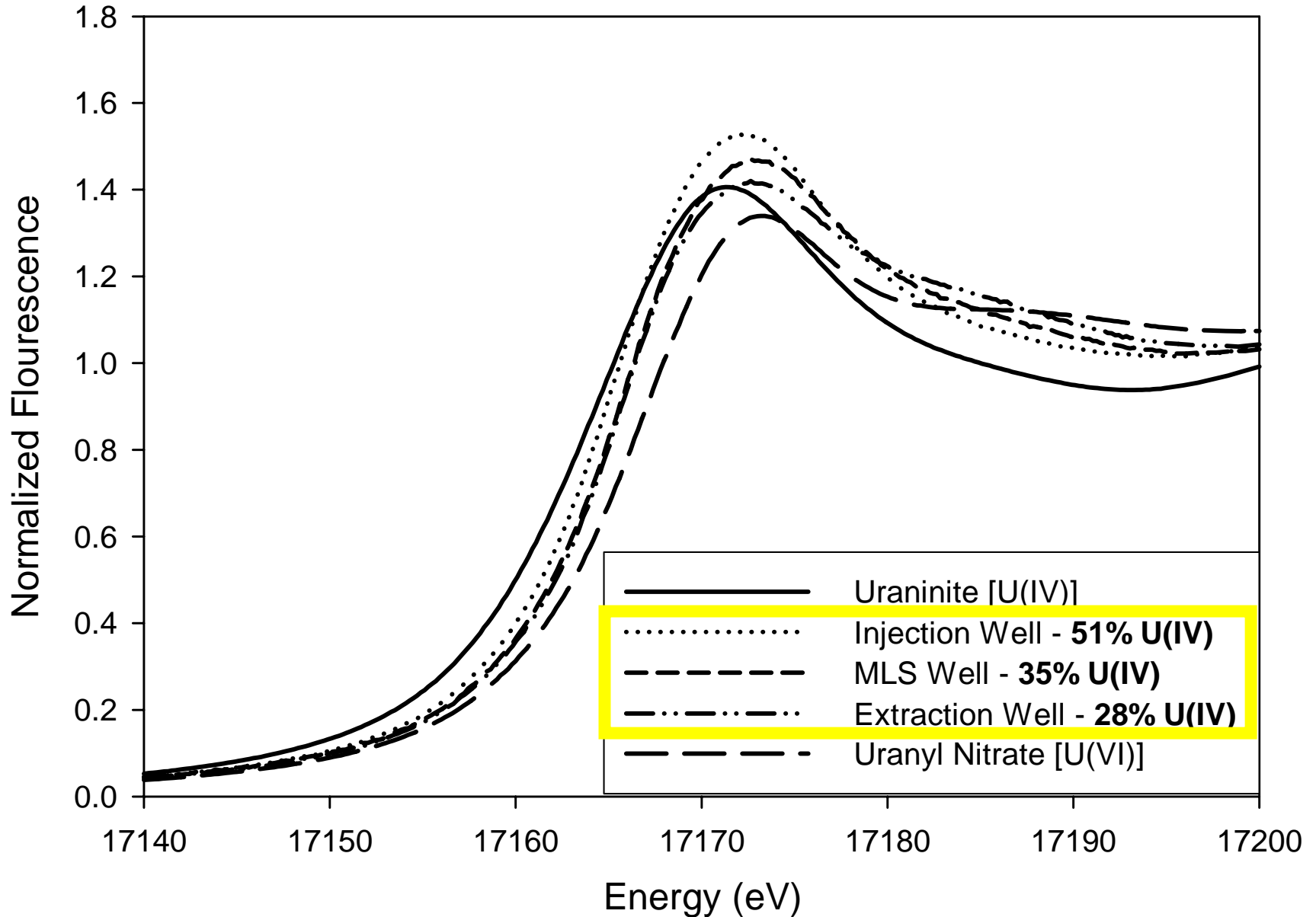


QuickTime™ and a  
TIFF (LZW) decompressor  
are needed to see this picture.

**Samples for day 409.**

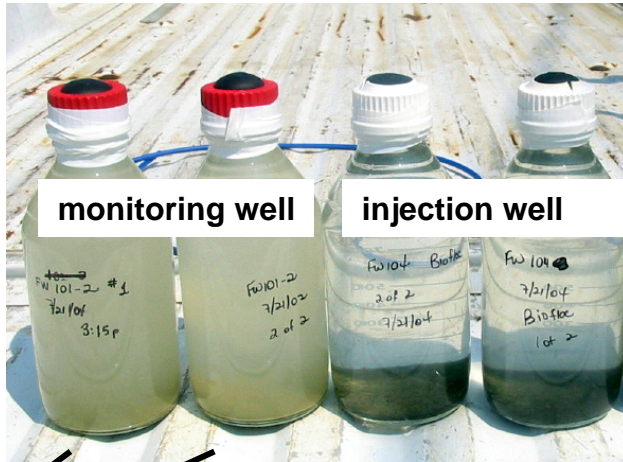
**U(IV) at the injection well was more than 50% of the total U. At the extraction well, it was less than 10%.**

## Uranium Redox State – Day 535



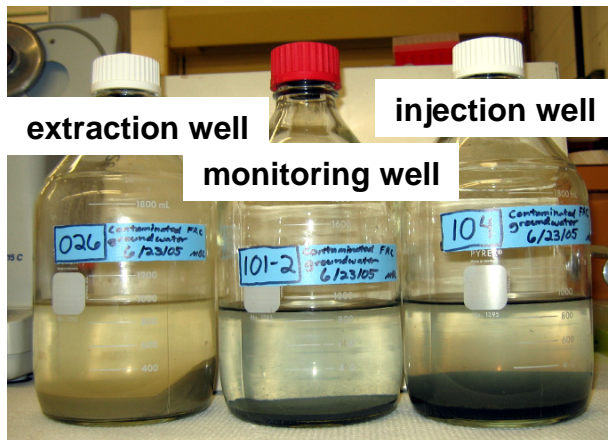
# The sediment changes color as microbial reduction progresses

Day 333



brown

Day 670



Now black



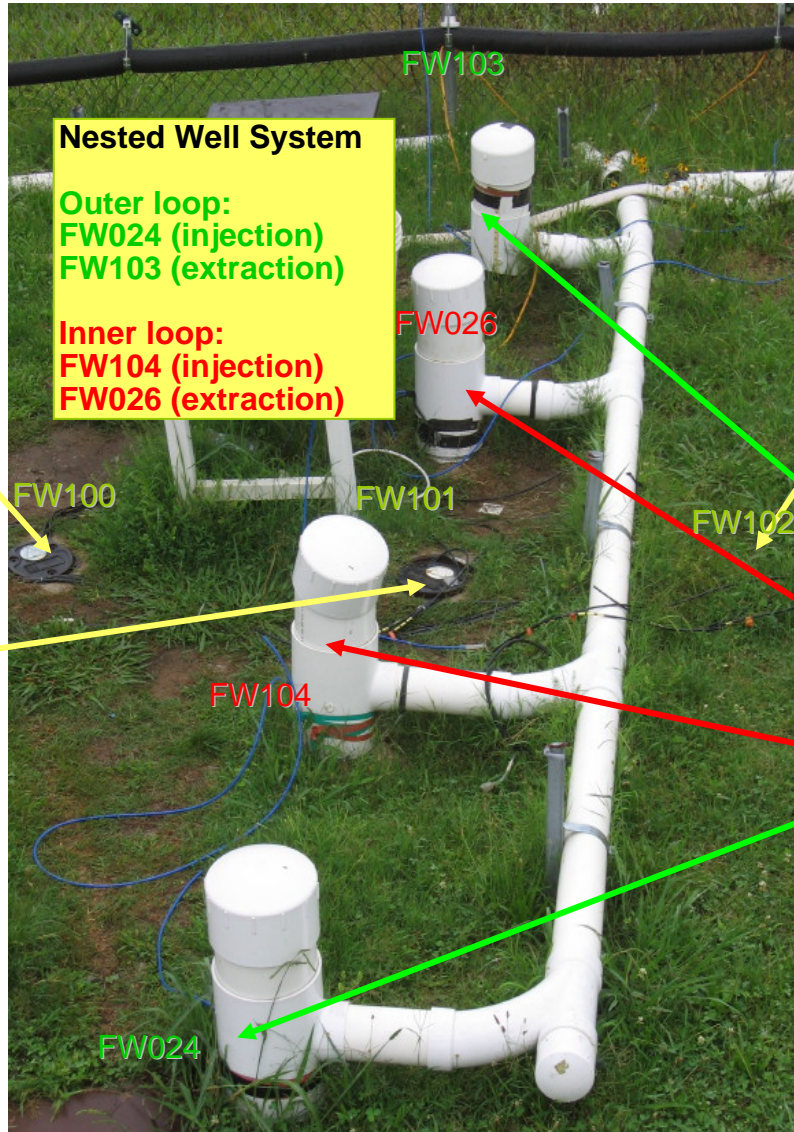
extraction well  
sample from day  
670 incubated 3  
days with no  
added ethanol

extraction well  
sample from day  
670 incubated 3  
days after adding  
100 mg/L ethanol

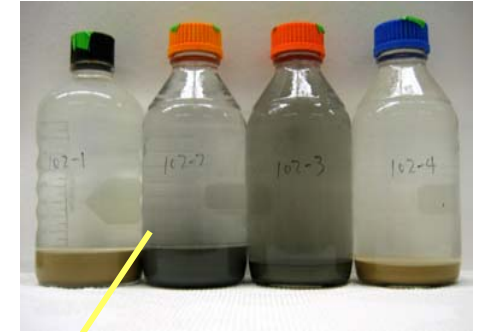
# Sediment from the treatment zone give visual evidence of reduction and expansion of the zone of reduction



Samples from FW100 at 45ft, 40 ft, 35ft and 30ft.



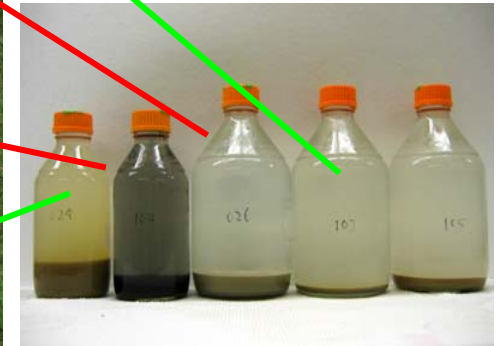
**Nested Well System**  
**Outer loop:**  
FW024 (injection)  
FW103 (extraction)  
**Inner loop:**  
FW104 (injection)  
FW026 (extraction)



Samples from FW102 at 45ft, 40 ft, 35ft and 30ft.



Samples from FW101 at 45ft, 40 ft, 35ft and 30ft.



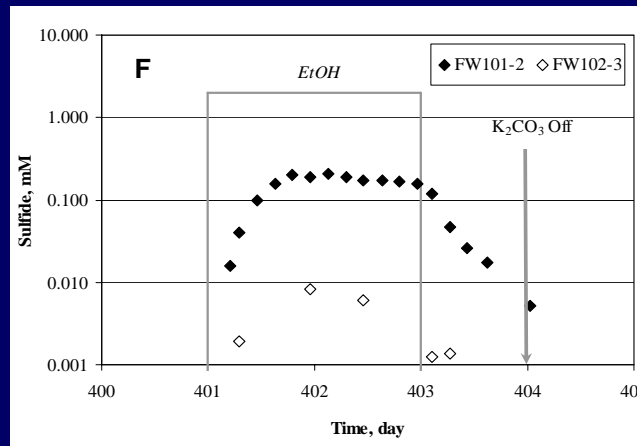
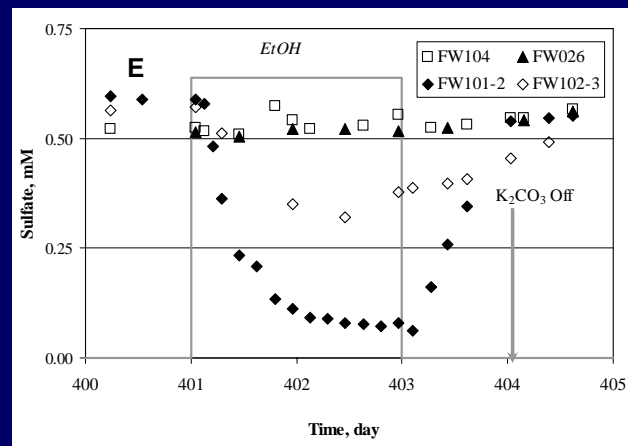
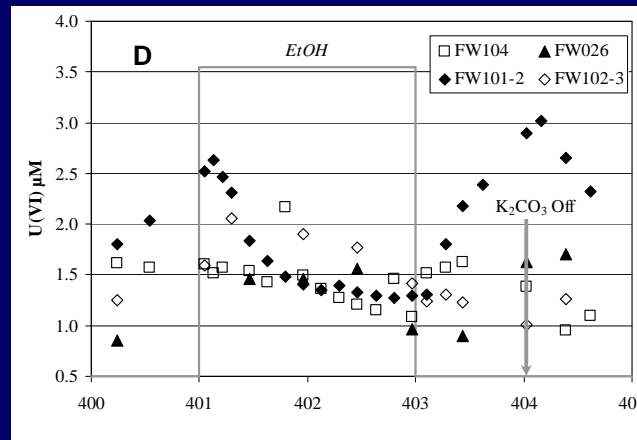
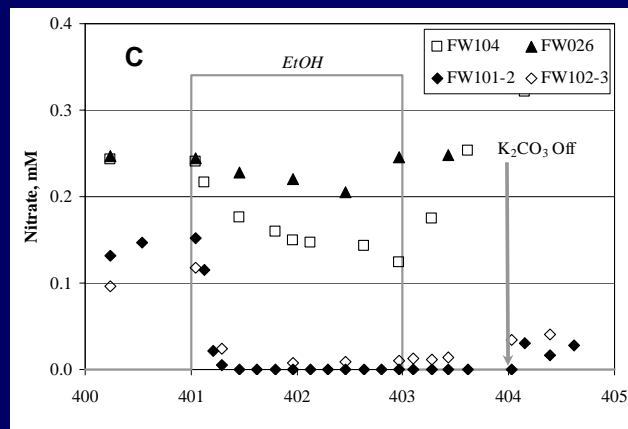
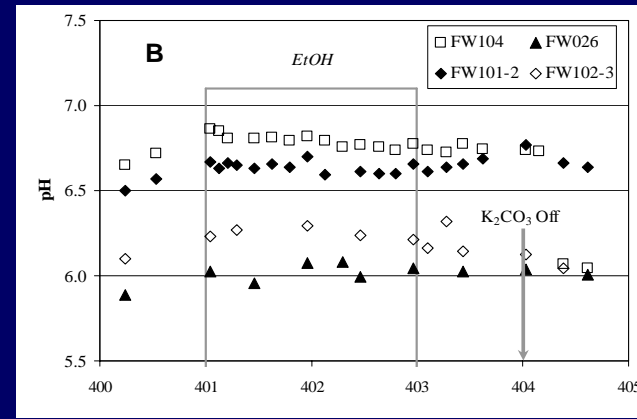
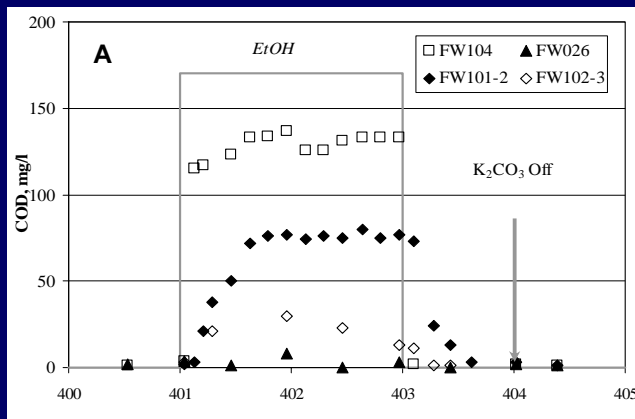
Samples from FW024, FW104, FW026, FW103 and FW105 (down gradient well).

# U(VI) bioavailability experiments

See Wu poster

# Days 399-409

Sequence:  
base on,  
ethanol on,  
ethanol off  
base off.  
(Summer)

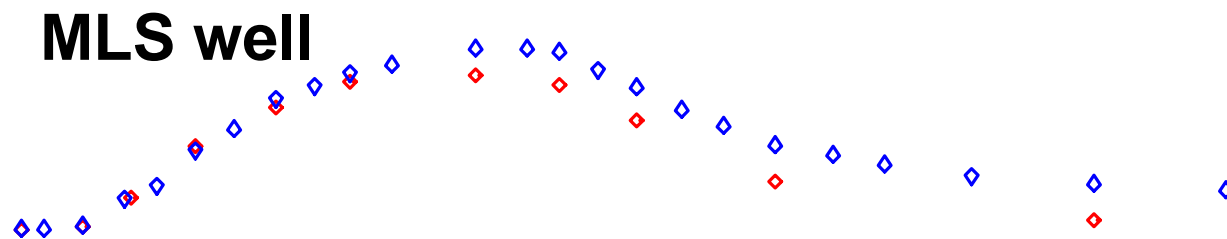
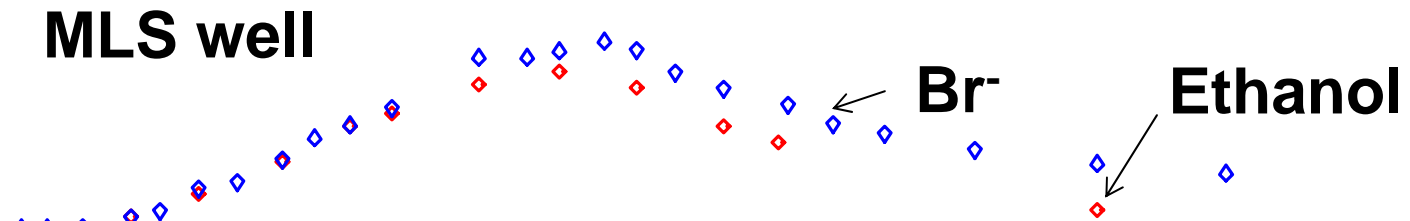
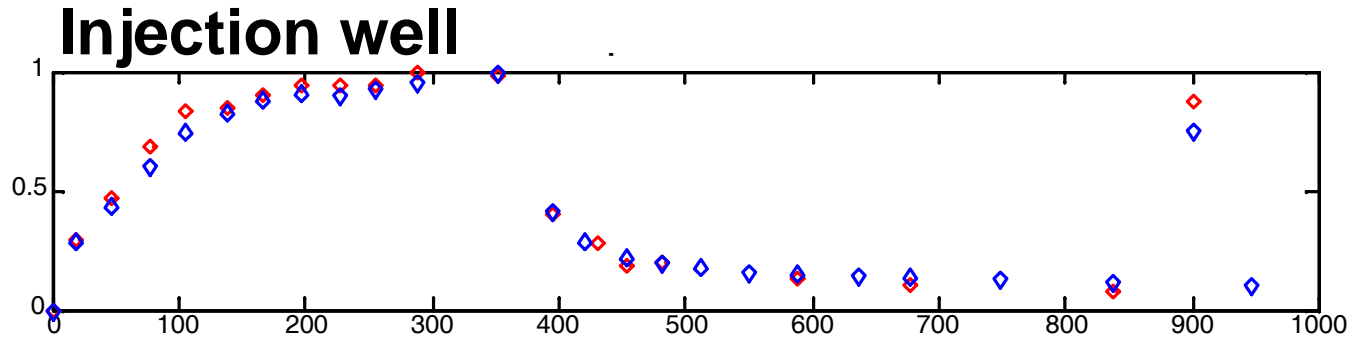


# Modeling

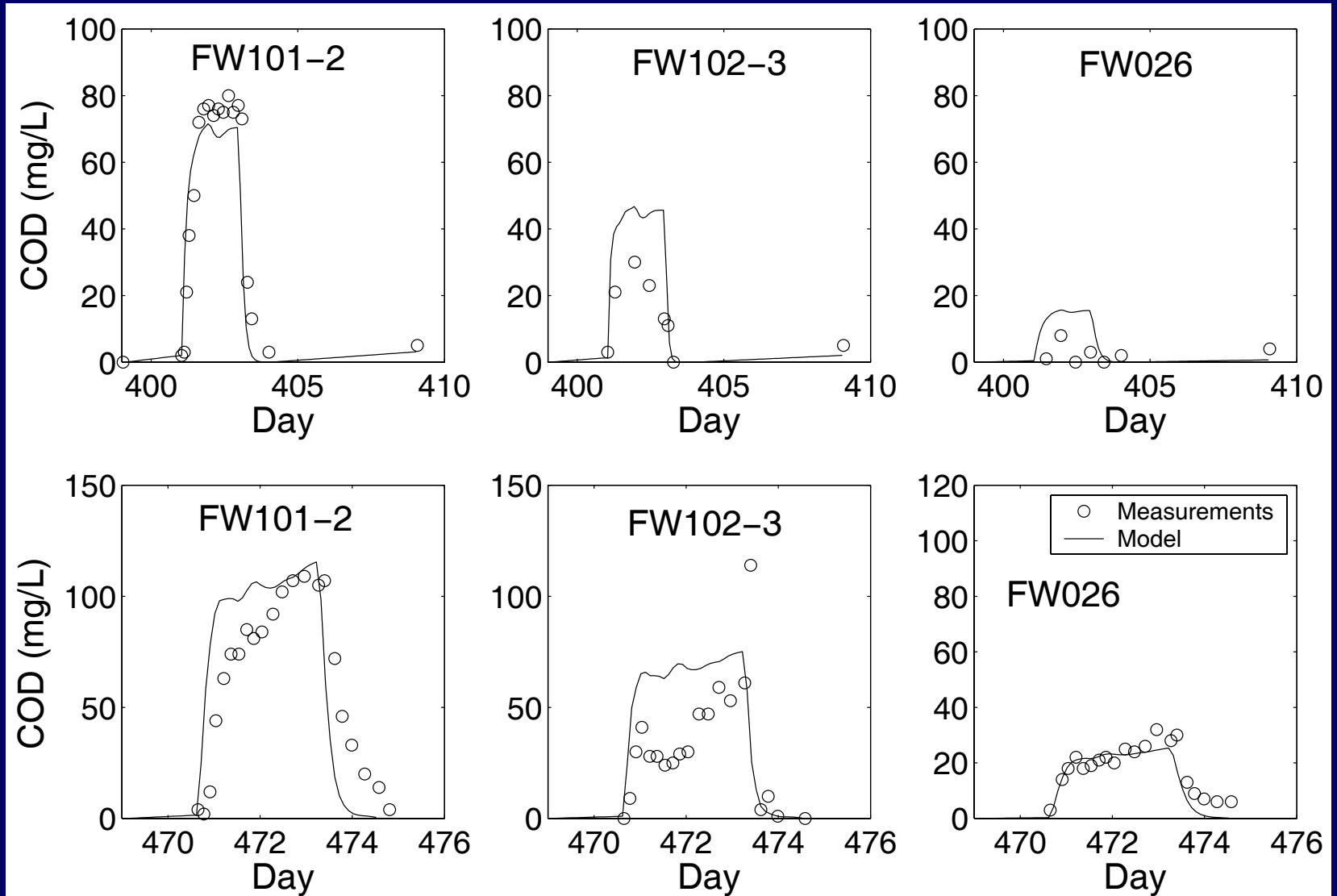
See Jian Luo poster



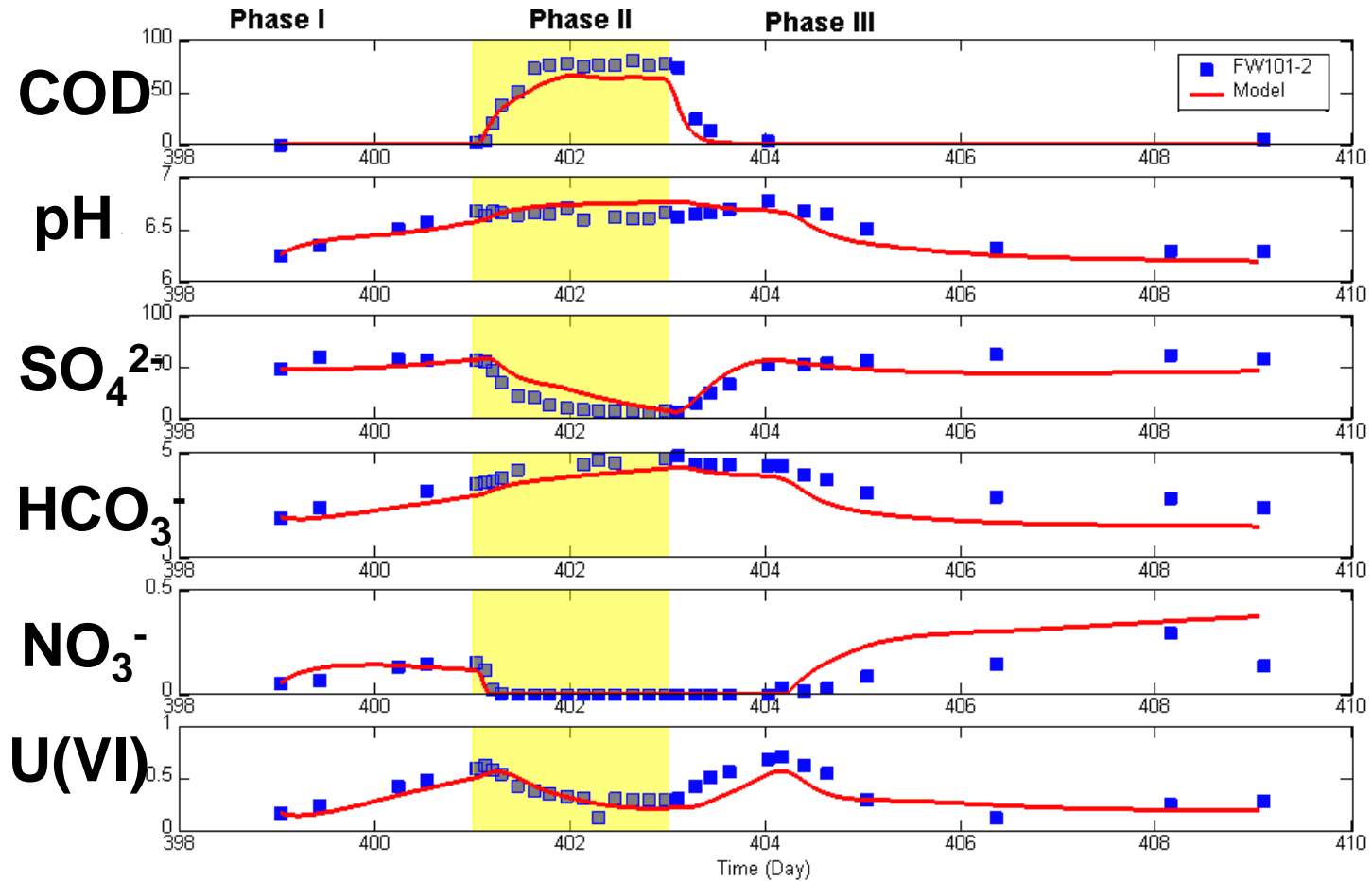
# Model calibration: ethanol and bromide tracer study



# Predictions for ethanol consumption



# Reactive transport simulation (Days 399-409)



# Microbiology

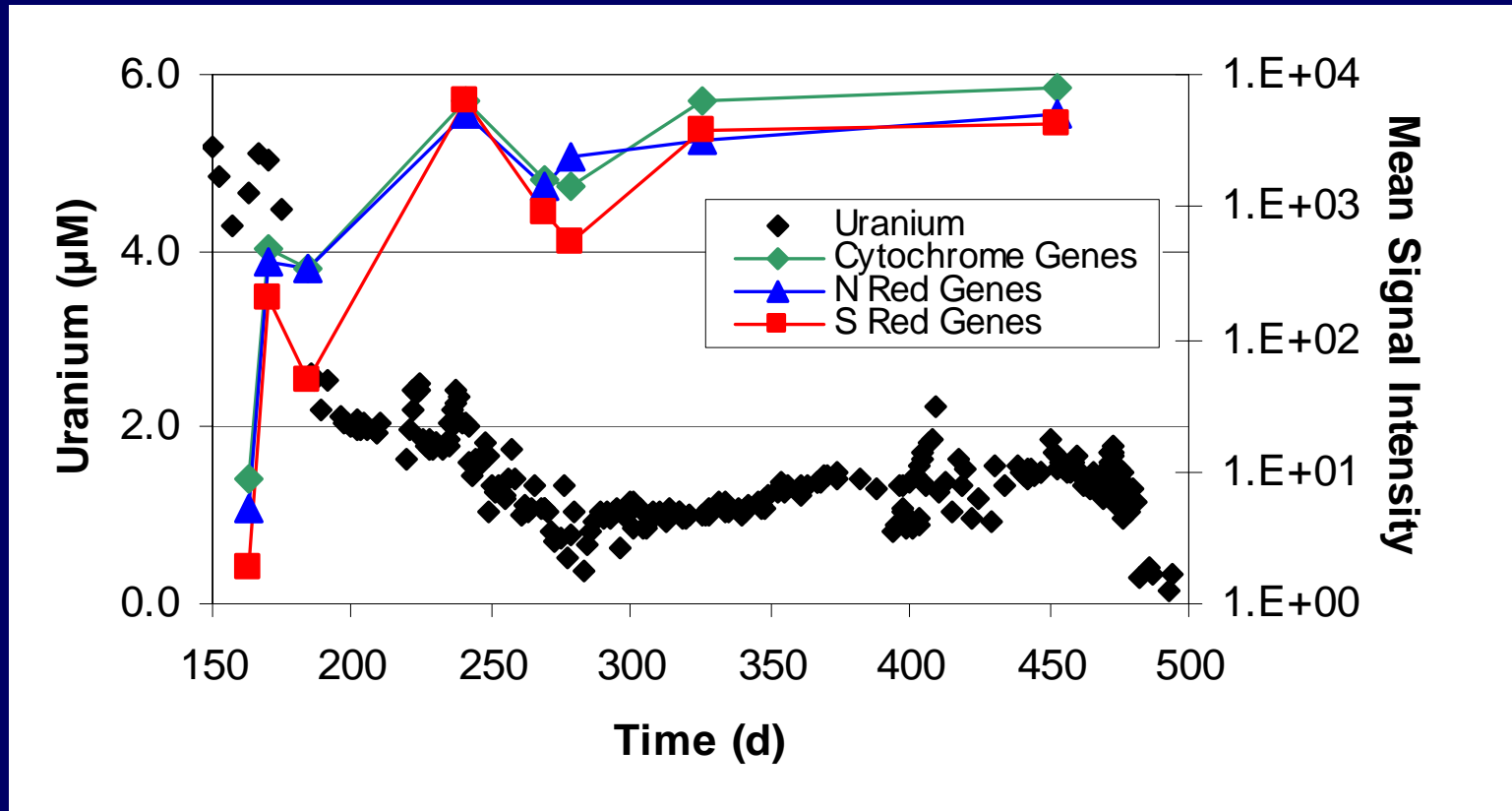
See Terry Gentry poster

## MPN values for different trophic groups (number/mL)

Well	Denitrifiers	Sulfate Reducers	Iron Reducers
Inner loop extraction	$3.5 \times 10^5$	$1.6 \times 10^5$	$2.0 \times 10^3$
MLS 101-2	$5.6 \times 10^2$	$1.4 \times 10^5$	$2.4 \times 10^3$
MLS102-2	$5.4 \times 10^5$	$0.92 \times 10^4$	$2.8 \times 10^2$
MLS102-3	$2.1 \times 10^6$	$2.4 \times 10^5$	$3.2 \times 10^3$
106 Control well	$5.4 \times 10$	0	0

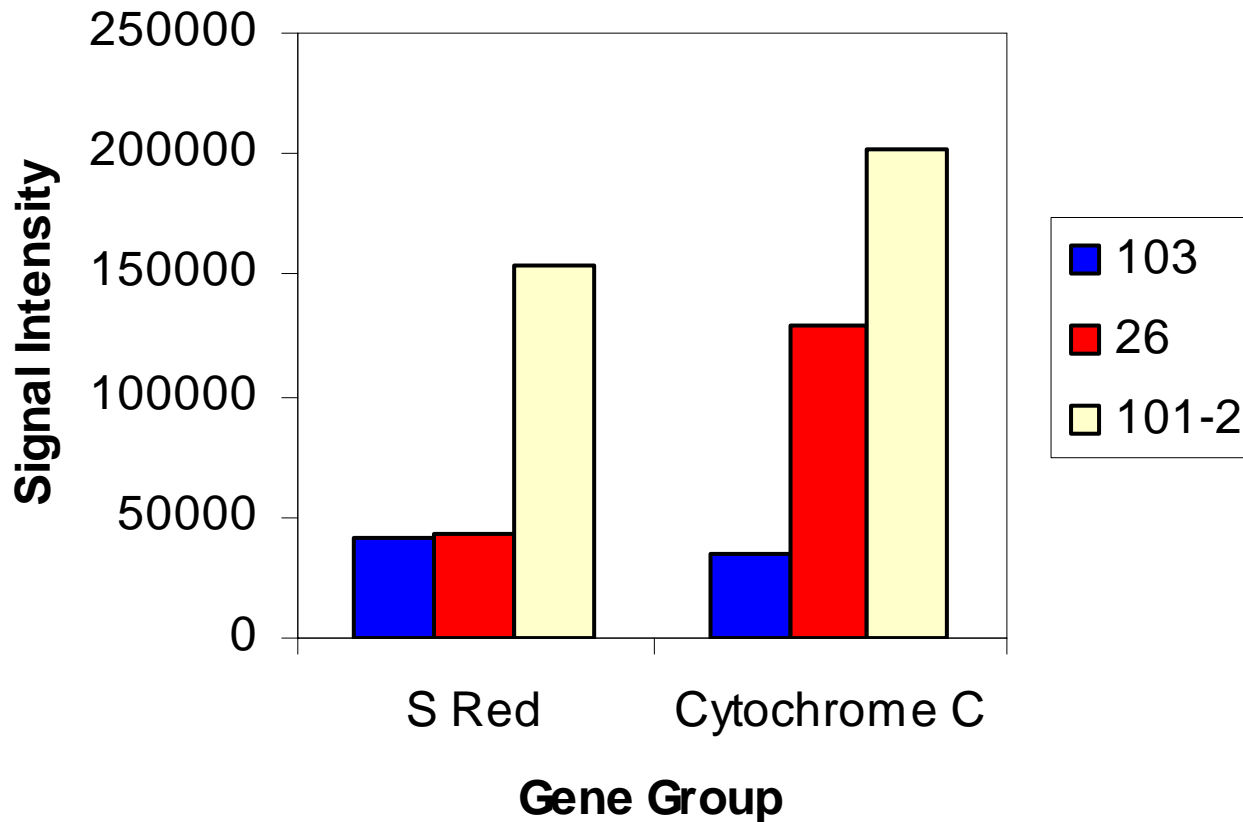
Note: MPN values for five replicates. Test wells sampled 8/20/04. Control well sampled 5/28/04.

# FW101-2 Denitrification, Sulfate Reduction & Cytochrome C Genes



- Increased levels of cytochrome C genes correlated with lower uranium levels

# Sediment Sulfate Reduction and Cytochrome C Genes – 535 d

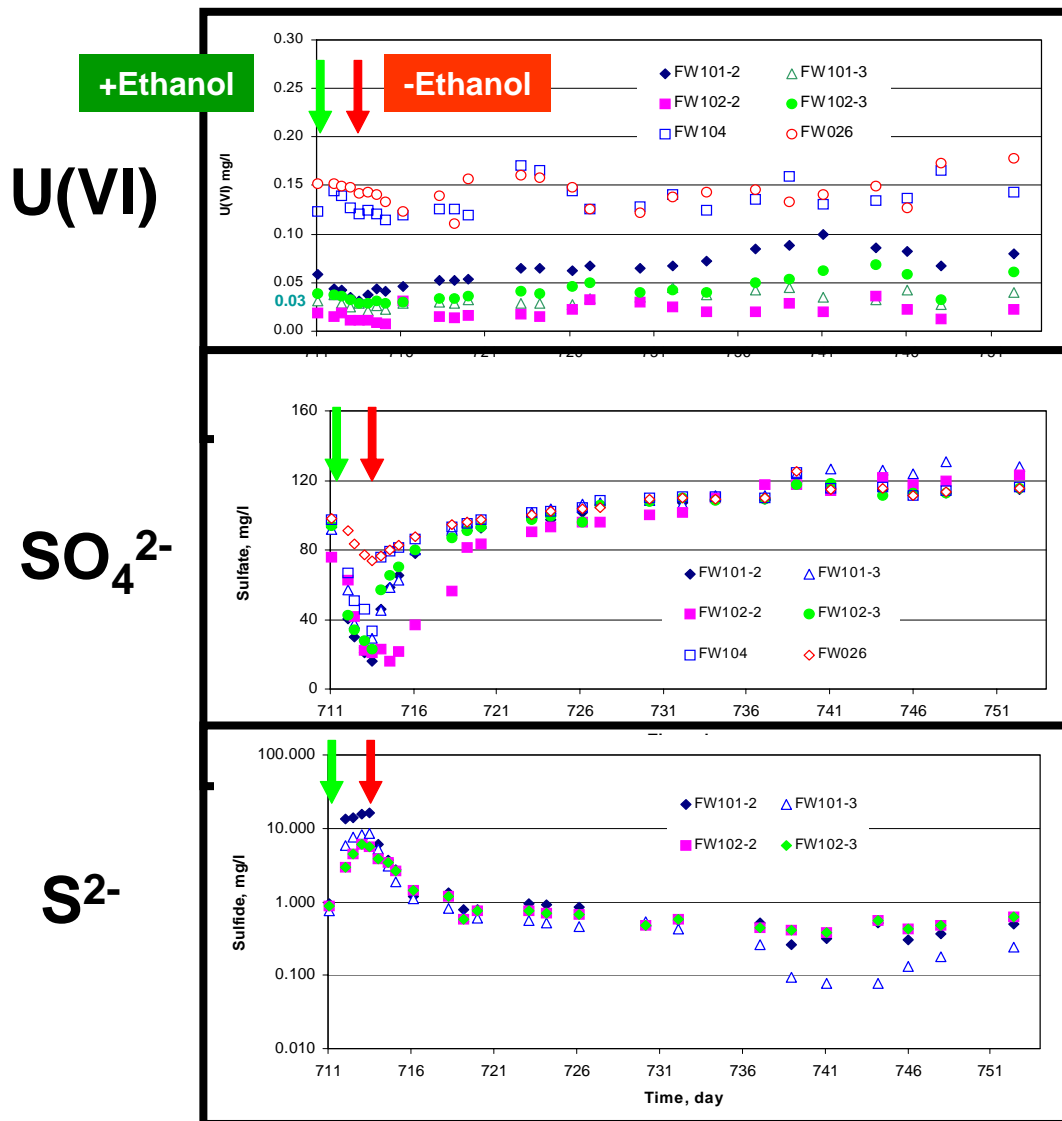


- Both sulfate reduction and cytochrome C genes were elevated in biostimulated sediment

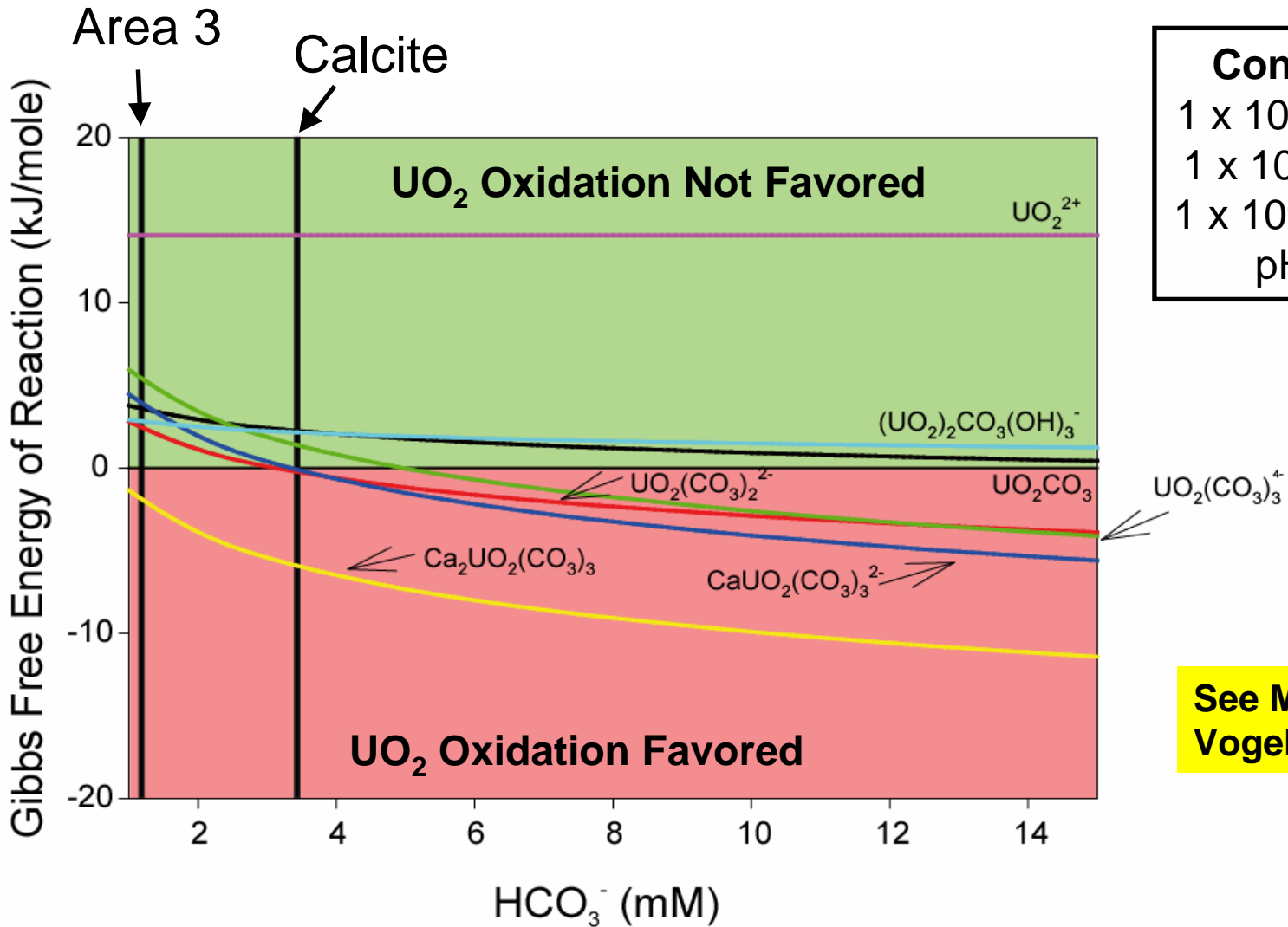
**Stability experiments  $\pm$  O<sub>2</sub>**



# System stability during a 41-day starvation period (days 713-754) - No O<sub>2</sub> in system



# UO<sub>2</sub>(Biogenic) Oxidation by Ferrihydrite



**Conditions**  
 $1 \times 10^{-6}$  M U(VI)  
 $1 \times 10^{-3}$  M  $\text{Ca}^{2+}$   
 $1 \times 10^{-5}$  M Fe(II)  
 pH = 7

See Matt Ginder-Vogel poster

High bicarbonate concentration results in UO<sub>2</sub> oxidation becoming more energetically favorable

# Conclusions: strategies for highly contaminated sites

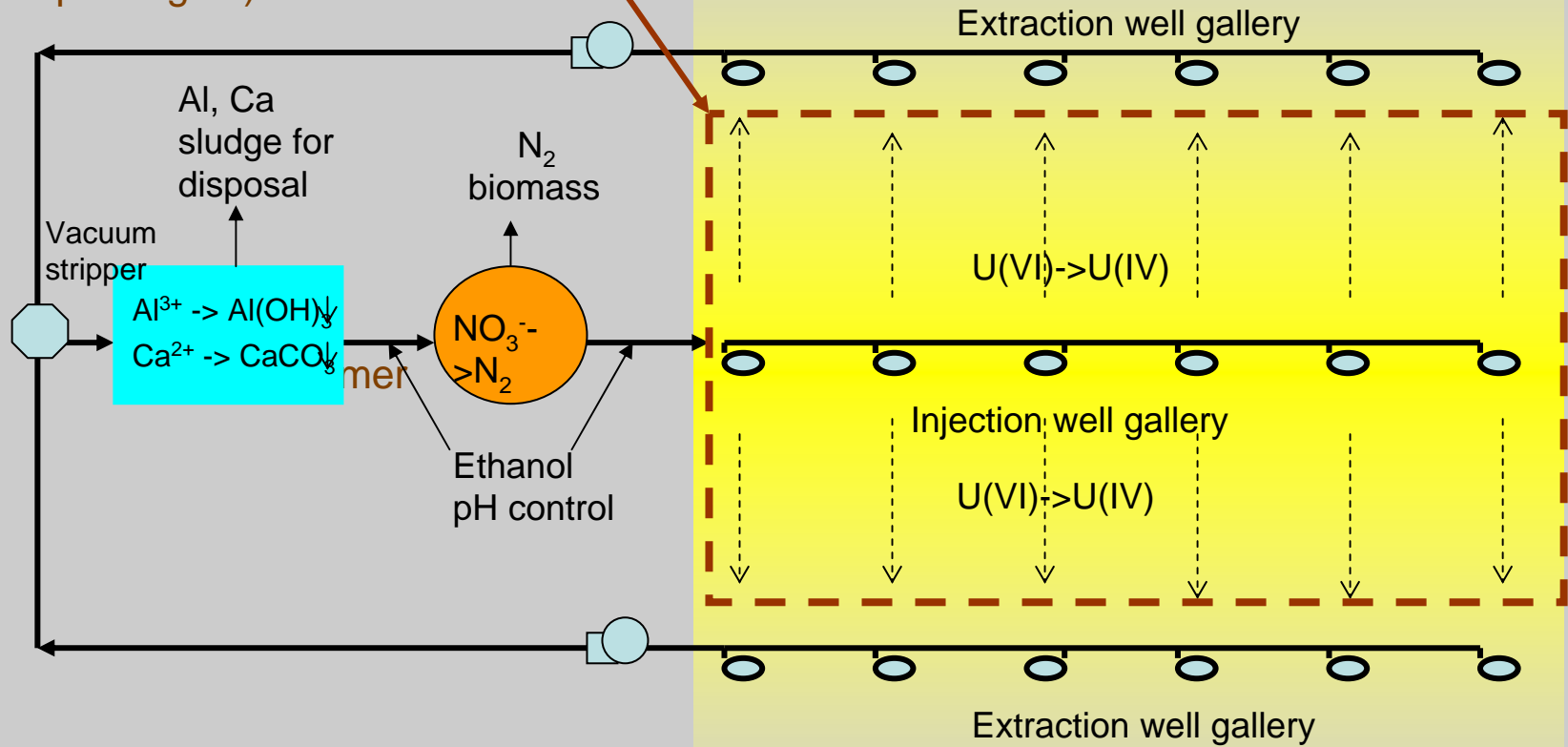
- Aboveground removal of inhibitors and clogging agents.
- Use of clean water flush to determine mass transfer rates and to condition a treatment zone.
- Staged treatment with soil conditioning before biostimulation.
- A nested recirculation scheme can protect a treatment zone from clogging agents and inhibitors.
- Mass transfer considerations may enable manipulation of contaminant bioavailability.

# Future Work

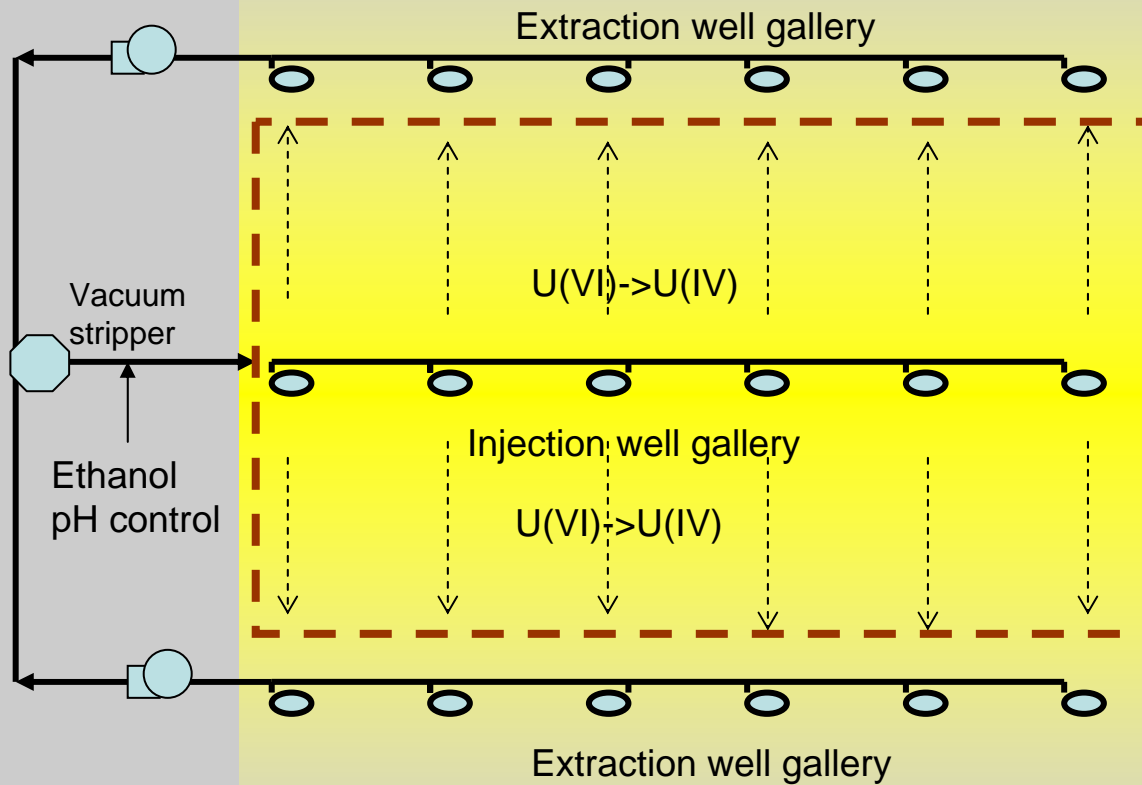
- Effects of oxygen on dissolved U(VI) levels.
- Evaluation of the extent of reduction required for stability.
- Microbial succession during stable and unstable operation
- Modeling of dissolved U flux from a reduced zone under different operational scenarios.

# Stage 1 -removal of aluminum, calcium, nitrate

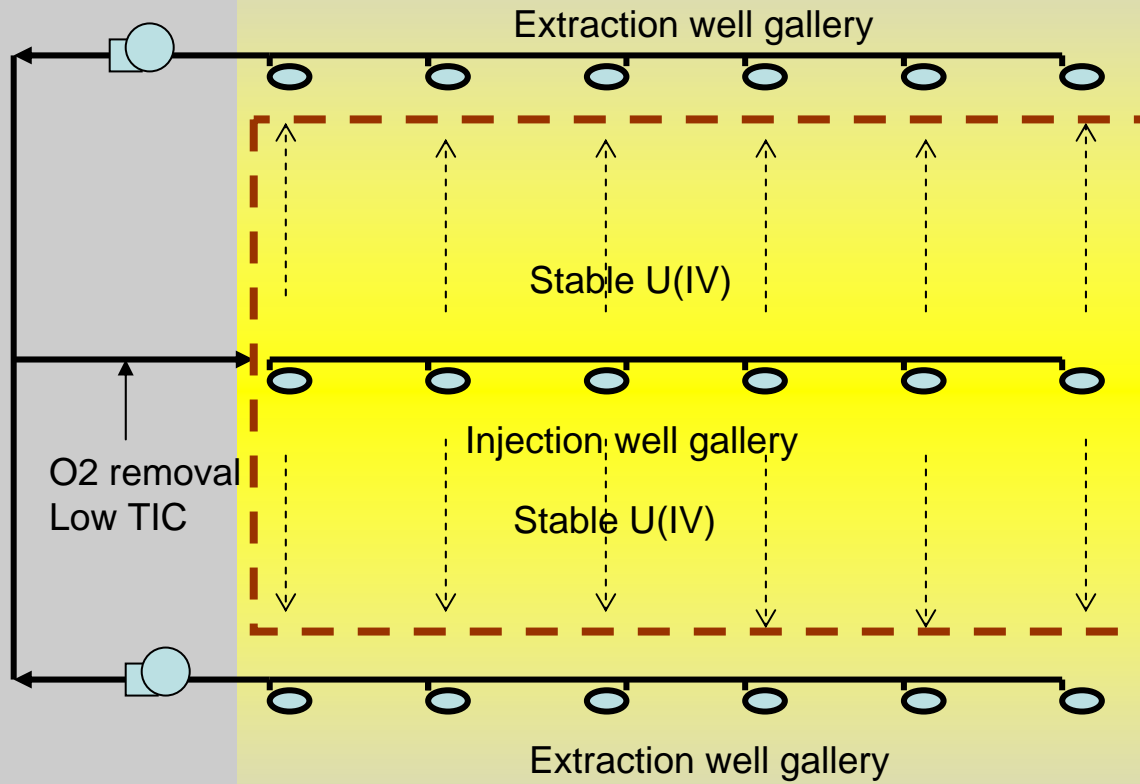
Former S-3 Ponds (now covered with parking lot)



## Stage 2 - conversion of U(VI) to U(IV)



### Stage 3 - Long-term maintenance of stable U(IV)



## Journal articles from the project

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Fields, M. W., T. Yan, S.-K. Rhee, S. L. Carroll, P. M. Jardine, D. B. Watson, C. S. Criddle, and J. Zhou. 2005. Impacts on microbial communities and cultivable isolates from groundwater contaminated with high levels of nitric acid-bearing uranium waste. Accepted for publication in *FEMS Microbiol. Ecol.*

Fienen, M.N., J. Luo, P. K. Kitanidis (2005). Semi-Analytical, Homogeneous, Anisotropic Capture Zone Delineation. *Journal of Hydrology*. In press.

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Gentile, M., Yan, T., Tiquia, S.M, Fields, M.W., Nyman, J., Zhou, J., and C.S. Criddle. Stability in a Denitrifying Fluidized Bed Reactor. Submitted/in review.

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Luo, J., Cirpka, O.A., Wu, W-M., Fienen, M.N., Jardine, P.M., Mehlhorn, T.L., Watson, D.B., Criddle, C.S., and Kitanidis, P.K. 2005. Mass-Transfer Limitations for Nitrate Removal in a Uranium-Contaminated Aquifer. Submitted.

Nyman, J.L.; Marsh, T.; Ginder-Vogel, M.; Gentile, M.; Fendorf, S. & Criddle, C.S. (2005). Heterogeneous Response to Biostimulation for U(VI) Reduction in Replicated Sediment Microcosms. Submitted.

### Conference Proceedings and Book Chapters:

Chen, J., S. Hubbard, M. Fienen, T. Mehlhorn, and D. Watson. (2003) Estimating Hydrogeological Zonation Using High-resolution Geophysical Data and Markov Chain Monte Carlo Methods. *Eos Trans. AGU*, 84(46), Fall Meet. Suppl., Abstract H21F-04.



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Criddle, C., J. Carley, O. Cirpka, M. Fields, M. Fienen, M. Gentile, T. Gentry, B. Gu, R. Hickey, P. Jardine, P. Kitanidis, K. Lowe, J. Luo, T. Mehlhorn, B. Nourse, J. Nyman, R. Rajou, D. Watson, W. Wu, J. Zhou. (2004) Biostimulation of in-situ uranium reduction at the NABIR Field Research Center using a nested recirculation scheme and aboveground ground water conditioning. DOE-NABIR workshop, Mar. 2004, Warrenton, VA.

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Fienen, M.N. and P. K. Kitanidis (2002). An Application of Bayesian Inverse Methods to Borehole Flowmeter Interpretation at Oak Ridge National Laboratory, USA. Presented at the NATO Advanced Study Institute, Trest Castle, Czech Republic, July 17-27, 2002.

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Nyman, J., S. Middleton, and C. S. Criddle, 2005. Bioengineering for the In-situ Remediation of Metals. Chapter 19 in Environmental Catalysis, Ed: Vicki Grassian. Marcel and Dekker Publishers, pp. 493-520.

Wu, W-M. J. Carley, M. Ginder-Vogel, T. Gentry, M. Fienen, J. Nyman, E. Tsai, S. Carroll, H. Yan, J. Luo, M. Gentile, T. Mehlhorn, M. W. Fields, B. Gu, D. Watson, J. Zhou, P. Jardine, S. Fendorf, P. Kitanidis, C. Criddle (2005). Field-Scale Demonstration of in situ Uranium (VI) Reduction at the NABIR Field Research Center, Oak Ridge, TN. Abstract submitted to the 105th American Society for Microbiology General Meeting, June 5-9, 2005.

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