

Commentary

Rising CO₂ – future ecosystems

'Every beginning biology student knows that photosynthesis will increase if you give a plant a 'squirt' of CO₂ – given enough light, nutrients, and water, and a suitable temperature. Logic tells us that if this is so, then more CO₂ in the atmosphere should mean more photosynthesis. This, in turn, should mean more yield or accumulated carbon in plants. This logic is fine for beginning biology; unfortunately, nature is not that simple' (Lemon, 1983).

This *Special Issue* of *New Phytologist* focuses on the responses of ecosystems to increased CO₂ concentration. The responses of plants are central to this focus, but the questions being asked have changed, and nature's complexities become paramount. Our concern is the human effect on the composition of the atmosphere and how it could have profound effects on our economic and social systems, options for energy production and use, and our capacity to grow food and fiber for an expanding population. The primary interaction between plants and atmospheric CO₂ is just the starting point for our analysis.

Research context

Lemon (1983), and the contributors to the international conference on which he was reporting, laid out a research agenda for investigating the responses of plants to future atmospheric CO₂ concentrations. The mostly short-term experiments that were appropriate for understanding the fundamental physiology of plants or the commercial aspects of CO₂ enrichment of glasshouse atmospheres (Witter & Robb, 1964) were seen as insufficient for understanding the more complex issues of plant productivity in a future, CO₂-

enriched atmosphere. The conference participants urged experimental work with CO₂ enrichment at all levels to elucidate biochemical, physiological and microbial responses, as well as community-scale responses and species interactions in complex environments.

Now, almost 20 years later, a great deal of that research agenda has been taken on. Not only do we know much more about the response of photosynthesis to a 'squirt' of CO₂ (Cousins *et al.* – see pp. 275–284 in this issue; Rodriguez *et al.* – pp. 337–346; Williams *et al.* – pp. 285–293), we have also studied everything from the effect of CO₂ concentration on the genetic control of stomatal density (Gray *et al.*, 2000) to the quality of bread and wine made from CO₂-enriched plants (Kimball *et al.* – pp. 295–303; Bindi *et al.*, 2001). Hundreds of plant species have been exposed to experimental manipulations of CO₂ concentration, and the unit of reference has progressed from small, potted plants in growth cabinets, to groups of plants in glasshouses or field chambers, to intact ecosystems and forest stands (Box 1). The CO₂ treatments have been combined with simultaneous manipulations of temperature, water, nitrogen, ozone, light, and competition. Research programs have increasingly been focused on describing how the primary responses to CO₂ concentration will be manifested in future ecosystems, understanding the feedbacks between those primary responses and the atmospheric and climatic systems, and developing plant and ecosystem models to make the predictions of plant responses to a future atmosphere.

These trends – larger-scale experiments, a focus on future ecosystems, and modeling – are reflected in the papers presented in this volume. A wide range of ecosystems is considered (Fig. 1): agricultural systems in Japan, Germany, and Arizona (USA); grasslands and pastures in Switzerland, Australia, New Zealand, and Minnesota (USA); bogs throughout Europe; a desert in Nevada (USA); and forests in Italy and Tennessee (USA).

Scale

In 1982, H. Z. Enoch spoke of the need for the scientific community to participate in a multinational effort to study the effects of elevated atmospheric CO₂ concentration on managed and unmanaged ecosystems, an admittedly expensive and difficult endeavor (Lemon, 1983). At the time, most of the information on CO₂ responses of plants came from short-term experiments (days or weeks) of potted plants in controlled-environment chambers (Kimball, 1983). It was recognized, however, that the short-term responses might not prevail over longer time periods, and that interactions between a plant and its environment (both biotic and abiotic)

Box 1 FACE technology

Exposure of ecosystems to controlled levels of elevated CO₂ under open-field conditions requires sophisticated free-air CO₂ enrichment (FACE) technology.

- **Origins** L. Hartwell Allen (1992), who coined the acronym FACE, attributes the first attempts to H. Lundegårdh in the 1920s. Lundegårdh devised a ground-level tube distribution system using CO₂ from decomposing manure. In the 1960s, D. W. Kretzman studied the effects of CO₂ on many field-grown crops, attempting (unsuccessfully) to get a yield benefit economically – as was becoming standard in glasshouse horticultural production – using ground-level release systems in combination with windbreaks. Additional ground-level releases were made later to use CO₂ as a tracer of atmospheric dispersion, and by the early 1970s Allen and colleagues had FACE-type research as a goal.
- **Air pollution** Allen (1992) describes attempts to simulate the effects of CO₂ releases on plant responses. One conclusion from these and from the prior ground-level releases was that the CO₂ concentration at the upper canopy level, where the greatest effect of CO₂ on photosynthesis would be expected, was much lower than that at ground level. Fortunately, air pollution scientists had also been attempting to control the concentration of gases over field plots, and had begun to experiment with releasing the gases near the top of the canopy. The circular system designed by McLeod *et al.* (1985) probably came closest to providing the dispersal needed for large-area CO₂-enrichment plots.
- **Brookhaven National Laboratory** In 1986, Lance Evans, from Manhattan College, together with Keith Lewin and George Hendrey from Brookhaven National Laboratory (BNL) proposed a consortial attempt to develop a FACE system using concepts from McLeod and other air pollution scientists. Also, Jackson Mauney from the USDA-ARS Western Cotton Research Laboratory identified geological CO₂ wells and a fertilizer factory at Yazoo City, Mississippi, that might be potential sources of cheap CO₂ for research – and publicised these to US researchers. Enthusiasm was high, and the upshot was that the DOE funded BNL to develop FACE apparatus and, subsequently, to conduct a FACE experiment on cotton.
- **Apparatus for diverse ecosystems** The BNL team designed a FACE release system for 1-m-tall agricultural crops that featured a circular array of vertical vent pipes with each pipe having its own computer controlled valve (Hendrey, 1993). CO₂ was released only upwind of the plots, with the decision as to which pipes to release from, and the release rate, continually updated (in seconds). With firmer data on CO₂ requirements in hand – and with the realization that scientific labor dominates research budgets in spite of the high cost of commercially available CO₂ (Kimball, 1992) – the decision was made to move the system to Maricopa, Arizona, near where a team of CO₂ researchers led by Bruce Kimball and Jackson Mauney had been using open-top chambers to study the effects of elevated CO₂ on cotton. The first FACE experiment with publishable biological data was conducted there in 1989 (Hendrey, 1993). Following their success, the BNL apparatus were adapted to enable enrichment of forest ecosystems (e.g. DeLucia *et al.*, 1999) and to accommodate release of ozone in interactive experiments (e.g. Karnosky *et al.*, 2001) – opening up the technology for use on diverse ecosystems during the past decade. Most of the FACE projects listed in the following websites, and many of the papers in this volume, utilize BNL-designed apparatus:

BNL FACE Group	http://www.face.bnl.gov/
Carbon Dioxide Information and Analysis Center	http://cdiac.esd.ornl.gov/programs/FACE/face.html
GCTE Elevated CO ₂ Network	http://gcte-focus1.org/co2.html
- **Recent advances** The BNL system uses blowers to predilute CO₂ with air before release, an aspect of their design which enhances the uniformity and control of the concentration over the plots. However, in 1993, it was realized that the blowers introduce sufficient air turbulence to warm the plant canopy significantly on calm nights (Pinter *et al.*, 2000), emphasizing the need for control plots with identical air flow properties. It also led Okada *et al.* (pp. 251–260 in this issue) and Miglietta *et al.* (pp. 465–476) to devise alternative FACE designs that release pure un-prediluted CO₂ over the plots, thus eliminating the blowers. FACE technology continues to evolve.

could alter the system-level response to CO₂ (Lemon, 1983). Many of these problems were addressed by new experiments conducted in various field chambers. In short-statured systems such as a salt marsh and tundra, open-top chambers allowed the treatment of intact ecosystems (Mooney *et al.*, 1991). Field chambers also permitted multiyear exposures of tree species without the artifacts associated with confining root systems in pots (Norby *et al.*, 1999).

Although much was learned from field chamber experiments, they fell short of the need expressed by Enoch. Field chambers create artificial environmental conditions, and plants often grow differently inside than outside (Kimball

et al., 1997). They can accommodate young trees, but not mature tree stands or forest ecosystems. Hence, the development of free-air CO₂ enrichment (FACE) technology for controlling an elevated CO₂ concentration in the open air was a critical advancement enabling the study of CO₂ effects on ecosystems. The history of FACE technology is described in Box 1.

The importance of the substantial increase in scale afforded by FACE systems is clear in many of the papers in this issue. Edwards *et al.* (pp. 359–369) report that elevated CO₂ concentration increased seedling growth of pasture species when grown individually in pots, but not when they were



Fig. 1 The effects of atmospheric CO₂ enrichment have been investigated in a wide range of ecosystem types, including crop systems in Arizona, USA; a bog in Finland; the Mojave desert in Nevada, USA; and a deciduous forest in Tennessee, USA. *Photos courtesy of Bruce Kimball, Topi Ylä-Mononen, Travis Huxman and Steve Eberhardt, respectively.*

grown in a native pasture within their FACE plots. Measurements of the exchange of CO₂ between the atmosphere and rice paddy flood water were made in a FACE experiment (Koizumi *et al.*, pp. 231–239), but they would not have been possible in a chamber system with blowers that alter micrometeorological conditions. Physiological responses to elevated CO₂ concentration often take on different meaning at a larger scale. Ottman *et al.* (pp. 261–273) suggest that the CO₂ effect on stomatal closure might be an advantage under limited water supply but a disadvantage when water supply is ample. Wullschlegel & Norby (pp. 489–495) found that the effect of elevated CO₂ concentration on stomatal closure, measured on upper canopy leaves under ideal conditions, did not scale to a reduction in season-long, whole-canopy transpiration in a tree stand.

The larger scale of FACE experiments makes possible measurements that otherwise would be unattainable. Norby *et al.* (pp. 477–487) were able to address questions about the growth responses of trees that had reached canopy closure, not heretofore possible in a deciduous forest system. They note that their study trees were in a linear growth phase, but had they been grown in open-top chambers, the experiment would have ended just at the critical transition from exponential growth. Even in those FACE experiments in which the experimental unit is relatively small, the larger exposure unit allowed for a wide range of simultaneous measurements and manipulations (Reich *et al.*, pp. 435–448).

Ecosystems of the future

The primary rationale for all of the studies reported in this issue concerns prediction of the future behaviour of ecosystems in an atmosphere with a higher concentration of CO₂. The most critical issues vary in the different ecosystems. In agricultural systems, the objective might be to predict productivity or quality of the marketable product in response to high CO₂ (Kim *et al.*, pp. 223–229; Kimball *et al.*, pp. 295–303; Lilley *et al.* (b), pp. 385–395); this must be done in relation to technological improvements and crop breeding (Amthor, 1998), as well as the overriding influences of environmental stress. In unmanaged systems such as the desert and prairie, effects of CO₂ concentration on diversity may be the predominant issue. Smith *et al.* (2000) showed that in a high rainfall year elevated CO₂ concentration stimulated the establishment and spread of an invasive annual grass in the Nevada desert FACE experiment; this has the potential to accelerate the fire cycle, reduce biodiversity, and alter ecosystem function in the deserts of western North America. The primary rationale for experiments in forests derives from their very large role in the global carbon budget and the importance of understanding exchanges and feedbacks between forests and a future atmosphere. Forest ecosystems are difficult to manipulate as intact systems because of their size and longevity; hence, forest experiments focus on testing specific

hypotheses about forest response (Norby *et al.*, 1999, also pp. 477–487).

Ecosystems provide essential services to humans, and there is increasing concern that those services might be jeopardized by the combined impacts of global change (Daily *et al.*, 1997). The provision of food, fiber, and water is of obvious importance. A less obvious ecosystem service is carbon sequestration, and this has been a particular focus of research because of the possibilities of feedbacks to the climate system. The effects of elevated CO₂ concentration on carbon fluxes have been considered at multiple scales: leaf (Tjoelker *et al.*, pp. 419–424), whole-plant (Sakai *et al.*, pp. 241–249), and whole system (Hoosbeek *et al.*, pp. 459–463; Craine *et al.*, pp. 425–434). Nutrient limitations apparently prevented any increases in C storage in bogs (Hoosbeek *et al.*). Stable isotope analysis provides a valuable tool for assessing the mechanisms of sequestration in soil in FACE experiments because the CO₂ that is added to the treatment plots is depleted in ¹³C (Leavitt *et al.*, pp. 305–314).

Future ecosystems will be impacted not just by rising CO₂ concentration, but by a suite of atmospheric and climatic changes. FACE experiments are usually not as amenable to multifactor manipulations as smaller-scale experiments, but in this issue there are reports about interactions between CO₂ and N in rice and wheat (Kim *et al.*; Kimball *et al.*), prairie species (Craine & Reich, pp. 397–403; Lee *et al.*, pp. 405–418), and grasses (Daepf *et al.*, pp. 347–358). Interactions with water supply were studied in wheat (Kimball *et al.*; Williams *et al.*). Air temperature is very difficult to manipulate in open-air systems. Lilley *et al.* (a) (pp. 371–383) grew subterranean clover and phalaris grass in tunnels in which CO₂ concentration and temperature were controlled. Previous reports (Newton *et al.*, 1994) had reported that the abundance of clover in pastures increases with rising CO₂ concentration. Lilley *et al.* found that elevated temperature caused clover abundance to decrease, although this effect was counteracted by elevated CO₂ concentration.

Modeling

Despite our best efforts to control environmental conditions and avoid artifacts in FACE and other experimental systems, we cannot duplicate future ecosystems or the atmospheric and climatic conditions that will occur at a certain future date. Soils in our experimental systems developed under current conditions, and the plants are today's genotypes. To predict ecosystem responses to future conditions we must rely on models, and we want the response functions in those models (Rodriguez *et al.*) to be informed by the most realistic data possible. FACE experiments are particularly useful for this. In modeling plant responses to elevated CO₂ concentration in field chambers, it is necessary to account for the chamber effects, which are in fact plant responses to the altered microclimate due to the chamber enclosure and the plants themselves. Because of their composite nature, the chamber effects

are arguably harder to model than the plant responses to elevated CO₂ concentration *per se*. Assuming that the effects of elevated CO₂ concentration and the chambers on plants are multiplicative, one may compare the relative responses of the plants between the model and observation. The assumption can be disproved, however, by physiological considerations. In FACE experiments, by contrast, the results are almost free from artifacts, and, hence, the modeled plant responses to elevated CO₂ concentration can be compared with the observed ones without having to worry about the confounding effects of chambers (Kimball *et al.*, 1997). Grossman-Clarke *et al.* (pp. 315–335) compared model predictions of wheat productivity and water use with results from the Arizona FACE experiment. The model successfully described qualitative and quantitative behavior of the crop under elevated CO₂ concentration, making it possible to use the model for predictions about future behavior with greater confidence.

Testing models of unmanaged ecosystems in future CO₂ concentrations with experimental data is more problematic. In perennial systems in which the FACE experiment is imposed on existing vegetation (e.g. the desert FACE of Nowak *et al.*, pp. 449–458; the bog experiment of Hoosbeek *et al.*; or the deciduous forest FACE of Norby *et al.*), the CO₂ treatment is an abrupt increase in CO₂ concentration, to which some ecosystem processes could respond in quite a different way from those under gradually increasing CO₂ concentration (Cannell & Thornley, 1998). Luo & Reynolds (1999), nonetheless, pointed out that the ecosystem changes in FACE experiments can be analysed to elucidate responses of the individual ecosystem processes to the step change in CO₂ concentration, and these individual responses could be incorporated into a model to predict the whole-ecosystem responses to the increasing CO₂ concentration.

Synthesis

A synthesis of the effects of rising CO₂ concentration on ecosystems as reported in the papers in this issue and elsewhere in the literature cannot be undertaken lightly. We can safely conclude that in most systems photosynthesis is increased by CO₂ enrichment, and this generally results in increased plant growth. It is more difficult to make general statements about whole-system responses, such as carbon storage, water yield, and species composition, that apply across a wide range of ecosystems and the different spatial and temporal scales of their dominant processes. Predictions about the behavior of future ecosystems in an atmosphere with a higher concentration of CO₂ require an understanding of how the primary responses to CO₂ interact with the attributes of the different systems. As reports in this issue show, we should not expect a bog and a desert, nor a wheat field and a tree plantation, to respond identically to CO₂ enrichment – nature is not that simple.

Nevertheless, tremendous progress is being made in providing the data and understanding needed for making – and

having confidence in – predictions about the future. Papers in this volume have tackled some of the thorny problems of detecting changes in soil carbon, seeking functional group classifications of plant response, and scaling from leaf to stand. The end-point of experiments in agricultural systems is no longer simply yield, but includes consideration of nutritional quality for grazers or humans. Technological advances in CO₂ enrichment technology are allowing ecosystem-scale experiments in a greater diversity of ecosystems. The ongoing research described here is not the culmination of Enoch's call for a multinational effort on CO₂ responses of ecosystems, but part of a steady process of hypothesis formulation and testing at ever-increasing scales and levels of complexity. That process needs to continue.

Acknowledgements

Many of the papers in this volume were inspired by the FACE 2000 Conference held in Tsukuba, Japan, in June, 2000. The conference was sponsored by CREST (Core Research for Evolutional Science and Technology) of Japan Science and Technology Corporation. This summary contributes to the Global Change and Terrestrial Ecosystems (GCTE) project of the International Geosphere-Biosphere Programme. It was written at the Oak Ridge National Laboratory, which is managed by UT-Battelle, LLC, for the US Department of Energy under contract DE-AC05-00OR22725.

**Richard J. Norby^{1*}, Kazuhiko Kobayashi²
and Bruce A. Kimball³**

¹Environmental Sciences Division, Oak Ridge National Laboratory, Oak Ridge, Tennessee, 37831-6422, USA;

²National Institute of Agro-Environmental Sciences, Tsukuba, Ibaraki 305-8604, Japan; ³US Water Conservation Laboratory, USDA, Agricultural Research Service, Phoenix, Arizona 85040-8832, USA

*Author for correspondence
(tel +1 865 576 5261;
fax +1 865 576 939;
email rjn@ornl.gov)

References

- Allen Jr LH. 1992. Free-air CO₂ enrichment experiments: an historical overview. *Critical Reviews in Plant Sciences* 11: 121–134.
- Amthor JS. 1998. Perspective on the relative insignificance of increasing atmospheric CO₂ concentration to crop yield. *Field Crops Research* 58: 109–127.
- Bindi M, Fibbi L, Miglietta F. 2001. Free Air Enrichment (FACE) of grapevine (*Vitis vinifera* L.): II. Growth and quality of grape and wine in response to elevated CO₂ concentrations. *European Journal of Agronomy* 14: 145–155.

- Cannell MGR, Thornley JHM. 1998. N-poor ecosystems may respond more to elevated [CO₂] than N-rich ones in the long term. A model analysis of grassland. *Global Change Biology* 4: 431–442.
- Cousins AB, Adam NR, Wall GW, Kimball BA, Pinter Jr PJ, Leavitt SW, Lamorte RL, Matthias AD, Ottman MJ, Thompson TL, Webber AN. 2001. Reduced photorespiration and increased energy-use efficiency in young CO₂-enriched sorghum leaves. *New Phytologist* 150: 275–284.
- Craine JM, Reich PB. 2001. Elevated CO₂ and nitrogen supply alter leaf longevity of grassland species. *New Phytologist* 150: 397–403.
- Craine JM, Wedin DA, Reich PB. 2001. Grassland species effects on soil CO₂ flux track the effects of elevated CO₂ and nitrogen. *New Phytologist* 150.
- Daapp M, Nösberger J, Lüscher A. 2001. Nitrogen fertilization and developmental stage alter the response of *Lolium perenne* to elevated CO₂. *New Phytologist* 150: 425–434.
- Daily GC, Alexander A, Ehrlich PR, Goulder L, Lubchenco J, Matson PA, Mooney HA, Postel S, Schneider SH, Tilman D, Woodwell GM. 1997. *Ecosystem services: benefits supplied to human societies by natural ecosystems. Issues in Ecology, issue 2*. Washington, DC, USA: Ecological Society of America.
- DeLucia EH, Hamilton JG, Naidu SL, Thomas RB, Andrews JA, Finzi A, Lavine M, Matamala R, Mohan JE, Hendrey GR, Schlesinger WH. 1999. Net primary production of a forest ecosystem with experimental CO₂ enrichment. *Science* 284: 1177–1179.
- Edwards GR, Newton PCD, Tilbrook JC, Clark H. 2001. Seedling performance of pasture species under elevated CO₂. *New Phytologist* 150: 359–369.
- Gray JE, Holroyd GH, van der Lee FM, Bahrami AR, Sijmons PC, Woodward FI, Schuch W, Hetherington AM. 2000. The HIC signalling pathway links CO₂ perception to stomatal development. *Nature* 408: 713–716.
- Grossman-Clarke S, Pinter Jr PJ, Kartschall T, Kimball BA, Hunsaker DJ, Wall GW, Garcia RL, LaMorte RL. 2001. Modeling a spring wheat crop under elevated CO₂ and drought. *New Phytologist* 150: 315–335.
- Hendrey GR, ed. 1993. *Free-air carbon dioxide enrichment for plant research in the field*. Boca Raton, FL, USA: CK Smoley.
- Hoosbeek MR, van Breemen N, Berendse F, Grosvernier P, Vasander H, Wallén B. 2001. Limited effect of increased atmospheric CO₂ concentration on ombrotrophic bog vegetation. *New Phytologist* 150: 459–463.
- Karnosky DF, Oksanen E, Dickson DE, Isebrands JG. 2001. Impacts of interacting greenhouse gases on forest ecosystems. In: Karnosky DF, Scarascia-Mugnozza G, Ceulemans R, Innes J, eds. *The impact of carbon dioxide and other greenhouse gases on forest ecosystems*. New York, USA: CABI Press, (In press.)
- Kim HY, Lieffering M, Miura S, Kobayashi K, Okada M. 2001. Growth and nitrogen uptake of CO₂-enriched rice under field conditions. *New Phytologist* 150: 223–229.
- Kimball BA. 1992. Cost comparisons among free-air CO₂ enrichment, open-top chamber, and sunlit controlled-environment chamber methods of CO₂ exposure. *Critical Reviews in Plant Sciences* 11: 265–270.
- Kimball BA, Morris CF, Pinter Jr PJ, Wall GW, Hunsaker DJ, Adamsen FJ, Lamorte RL, Leavitt SW, Thompson TL, Matthias AD, Brooks TJ. 2001. Elevated CO₂, drought and soil nitrogen effects on wheat grain quality. *New Phytologist* 150: 295–303.
- Kimball BA, Pinter Jr PJ, Wall GW, Garcia RL, Lamorte RL, Jak PMC, Frumau KFA, Vugts HF. 1997. Comparisons of responses of vegetation to elevated carbon dioxide in free-air and open-top chamber facilities. In: Allen Jr LH, Kirkham MB, Olszyk DM, Whitman CE, eds. *Advances in carbon dioxide research*. Madison, WI, USA: American Society of Agronomy, Crop Science Society of America, Soil Science Society of America, 113–130.
- Kimball BA. 1983. Carbon dioxide and agricultural yield: an assemblage and analysis of 430 prior observations. *Agronomy Journal* 75: 779–788.
- Koizumi H, Kibe T, Mariko S, Ohtsuka T, Nakadai T, Mo W, Toda H, Seiichi N, Kobayashi K. 2001. Effect of free-air CO₂ enrichment (FACE) on CO₂ exchange at the flood-water surface in a rice paddy field. *New Phytologist* 150: 231–239.
- Leavitt SW, Pendall E, Paul EA, Brooks T, Kimball BA, Pinter Jr PJ, Johnson HB, Matthias AD, Wall GW, Lamorte RL. 2001. Stable-carbon isotopes and soil organic carbon in wheat under CO₂ enrichment. *New Phytologist* 150: 305–314.
- Lee TD, Tjoelker MG, Ellsworth DS, Reich PB. 2001. Leaf gas exchange responses of 13 prairie grassland species to elevated CO₂ and increased nitrogen supply. *New Phytologist* 150: 405–418.
- Lemon ER, ed. 1983. *CO₂ and plants*. Boulder, CO, USA: Westview Press.
- Lilley JM, Bolger TP, Gifford RM. 2001a. Productivity of *Trifolium subterraneum* and *Phalaris aquatica* under warmer, high CO₂ conditions. *New Phytologist* 150: 371–383.
- Lilley JM, Bolger TP, Peoples MB, Gifford RM. 2001b. Nutritive value and the nitrogen dynamics of *Trifolium subterraneum* and *Phalaris aquatica* under warmer, high CO₂ conditions. *New Phytologist* 150: 385–395.
- Luo Y, Reynolds JF. 1999. Validity of extrapolating field CO₂ experiments to predict carbon sequestration in natural ecosystems. *Ecology* 80: 1568–1583.
- McLeod AR, Alexander K, Hatcher P. 1985. Open-air fumigation of field crops: criteria and design for a new experimental system. *Atmospheric Environment* 19: 1639–1649.
- Miglietta F, Peressotti A, Vaccari FP, Zaldei A, deangelis P, Scarascia-Mugnozza G. 2001. Free air CO₂ enrichment (FACE) of a poplar plantation: the POPFACE fumigation system. *New Phytologist* 150: 465–476.
- Mooney HA, Drake BG, Luxmoore RJ, Oechel WC, Pitelka LF. 1991. Predicting ecosystem responses to elevated CO₂ concentrations. *Bioscience* 41: 96–104.
- Newton PCD, Clark H, Bell CC, Glasgow EM, Campbell BD. 1994. Effects of elevated CO₂ and simulated seasonal-changes in temperature on the species composition and growth-rates of pasture turves. *Annals of Botany* 73: 53–59.
- Norby RJ, Todd DE, Fuels J, Johnson DW. 2001. Allometric determination of tree growth in a CO₂-enriched sweetgum stand. *New Phytologist* 150: 477–487.
- Norby RJ, Wullschlegel SD, Gunderson CA, Johnson DW, Ceulemans R. 1999. Tree responses to rising CO₂: implications for the future forest. *Plant, Cell & Environment* 22: 683–714.
- Nowak RS, DeFalco LA, Wilcox CS, Jordan DN, Coleman JS, Seemann JR, Smith SD. 2001. Leaf conductance decreased under free-air CO₂ enrichment (FACE) for three perennials in the Nevada desert. *New Phytologist* 150: 449–458.

- Okada M, Lieffering M, Nakamura H, Yoshimoto M, Kim HY, Kobayashi K. 2001. Free-air CO₂ enrichment (FACE) using pure CO₂ injection: system description. *New Phytologist* 150: 251–260.
- Ottman MJ, Kimball BA, Pinter Jr PJ, Wall GW, Vanderlip RL, Leavitt SW, Lamorte RL, Matthias AD, Brooks TJ. 2001. Elevated CO₂ increases sorghum biomass under drought conditions. *New Phytologist* 150: 261–273.
- Pinter Jr PJ, Kimball BA, Wall GW, Lamorte RL, Hunsaker DJ, Adamsen FJ, Frumau KFA, Vugts HF, Hendrey GR, Lewin KF, Nagy J, Johnson HB, Wechsung F, Leavitt SW, Thompson TL, Matthias AD, Brooks TJ. 2000. Free-air CO₂ enrichment (FACE): blower effects on wheat canopy microclimate and plant development. *Agric. For. Meteorol.* 103: 319–333.
- Reich PB, Tilman D, Craine J, Ellsworth D, Tjoelker MG, Knops J, Wedin D, Naeem S, Bahaeddin D, Goth J, Bengston W, Lee TD. 2001. Do species and functional groups differ in acquisition and use of C, N and water under varying atmospheric CO₂ and N availability regimes? A field test with 16 grassland species. *New Phytologist* 150: 435–448.
- Rodríguez D, Ewert F, Goudriaan J, Manderscheid R, Burkart S, Weigel HJ. 2001. Modelling the response of wheat canopy assimilation to atmospheric CO₂ concentrations. *New Phytologist* 150: 337–346.
- Sakai H, Yagi K, Kobayashi K, Kawashima S. 2001. Rice carbon balance under elevated CO₂. *New Phytologist* 150: 241–249.
- Smith SD, Huxman TE, Zitzer SE, Charlet TN, Housman DC, Coleman JS, Fenstermaker LK, Seemann JR, Nowak RS. 2000. Elevated CO₂ increases productivity and invasive species success in an arid ecosystem. *Nature* 408: 79–82.
- Tjoelker MG, Oleksyn J, Lee TD, Reich PB. 2001. Direct inhibition of leaf dark respiration by elevated CO₂ is minor in 12 grassland species. *New Phytologist* 150: 419–424.
- Williams DG, Gempko V, Fravolini A, Leavitt SW, Wall GW, Kimball BA, Pinter Jr PJ, Lamorte R, Ottman M. 2001. Carbon isotope discrimination by *Sorghum bicolor* under CO₂ enrichment and drought. *New Phytologist* 150: 285–293.
- Wittwer SH, Robb W. 1964. Carbon dioxide enrichment of greenhouse atmospheres for food crop production. *Economic Botany* 18: 34–56.
- Wullschlegel SD, Norby RJ. 2001. Sap velocity and canopy transpiration in a sweetgum stand exposed to free-air CO₂ enrichment (FACE). *New Phytologist* 150: 489–498.

Key words: free-air CO₂ enrichment (FACE), ecosystems, atmospheric CO₂, modeling, scale.

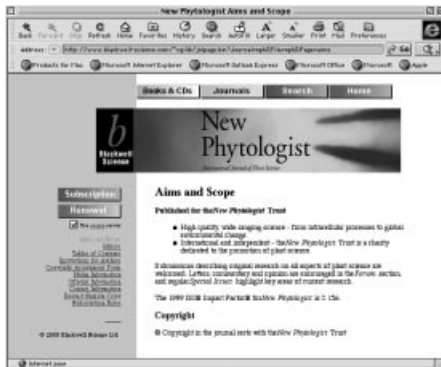


Have you seen the latest *New Phytologist Special Issue?*

Root Dynamics and Global Change: An Ecosystem Perspective
Editors Norby RJ, Fitter AH, Jackson RB
New Phytologist (2000) 147

As the reality of human-mediated global change becomes increasingly accepted by a sceptical public, so the scientific research that has identified this situation has become increasingly high profile. However, while we can almost see the leaves before us changing, the same is not true of plant roots – we ignore this hidden half at our peril. This *Special Issue* addresses root dynamics in the face of a globally changing environment and asks the key questions: Do atmospheric and climatic changes alter root production and root longevity? How do the changes impact on the whole plant and its microbial symbiotic partners? And, ultimately, how do these changes alter the ecosystem itself? The ecosystem perspective is especially important – root turnover is a key component of ecosystem metabolism and the capacity of ecosystems to store carbon. It is clear that the prime challenges still concern how to reach and analyze the roots themselves, but where there are gaps in our knowledge, many researchers are finding that the visible half – the leaves – often does provide a good analogy for roots. The reviews and original research reported here provide a comprehensive overview of the subject, and point the way ahead for systematic scientific exploration of this compelling topic of our times.

If you are interested in obtaining a copy, then let us know at Central Office (newphytol@lancaster.ac.uk) or the USA Office (newphytol@ornl.gov).



www.newphytologist.com

Did you know that you can read summaries of *New Phytologist* articles online, free of charge? The most up-to-date information about the journal is also available.

All online subscribers receive access to the fully navigable electronic version.

Supplementary material viewable online only at www.newphytologist.com, free of charge, can be submitted with articles. Alternatively, if the size or format of the supplementary material is such that it cannot be accommodated, authors can agree to make the material available on a permanent Website, to which links will be set up from www.newphytologist.com.

Visit the Website for further information about the journal, or get in touch with us at Central Office (newphytol@lancaster.ac.uk) or the USA Office (newphytol@ornl.gov).