









### **STR Typing Properties**

Fragment analysis

- Typically tetra nucleotide repeats
- PCR products range in size from ≈100-450 bp
- Non-coding region, no phenotype/ancestry information
- No linkage, allele frequencies can be multiplied for random match probabilities (<u>'1 in a trillion'</u>)
- 13 CODIS markers are typed in multiplex PCR kits from Promega, Qiagen, Life Technologies
  - 5plex to 23plex range (0.5 ng of genomic DNA)
  - Fluorescently labeled PCR primers

# Quality Assurance Standards for Forensic DNA Testing Laboratories

- The standards describe the quality assurance requirements that laboratories performing forensic DNA testing or utilizing the Combined DNA Index System (CODIS) shall follow to ensure the quality and integrity of the data generated by the laboratory.
- STANDARD 9.5.5 The laboratory shall check its DNA procedures annually or whenever substantial changes are made to a procedure against an appropriate and available NIST standard reference material or standard traceable to a NIST standard.
- STANDARD 9.5.1 Where quantitation is used, quantitation standards shall be used.

http://www.fbi.gov/about-us/lab/codis/qas-standards-for-forensic-dna-testing-laboratories-effective-9-1-201:

### **NIST Standard Reference Materials** http://www.cstl.nist.gov/biotech/strbase/srm\_tab.htm Traceable standards to ensure accurate measurements in our nation's crime laboratories 1 SRM 2391c Just released 2391c Certificate of Analysis with expanded CODIS core loci and Helps meet QAS Std. 9.5.5 Y-STRs and ISO 17025 SRM 2391c - CODIS core STRs SRM 2392 & 2392-I – mtDNA SRM 2395 - Y-STRs SRM 2372 – Human DNA quantitation Calibration with SRMs enables confidence in comparisons of Standard Reference Material results between laboratories

Description of Components in SRM 2391c					
Component	Description	2391c contains confirmed			
Α	50 μL of anonymous <b>female</b> genomic DNA	genotypes for:			
В	50 μL of anonymous <b>male</b> genomic DNA	51 autosomal STR loci (includes FBI core loci, European core loci, others)			
С	50 μL of anonymous <b>male</b> genomic DNA				
D	50 μL of <b>mixed-source</b> (Components A and C)	Allele calls (repeats) are			
E	Two 6 mm punches of CRL-1486 cells spotted on <b>903 paper</b>	confirmed by Sanger sequencing performed at NIST			
F	Two 6 mm punches of HTB-157 cells spotted on <b>FTA paper</b>				
	Liquid components ≈ 2 ng/μL Paper 75,00 cells per spot				

# • Repeat motif is confirmed by Sanger sequencing • Loci amplified (heterozygous samples separated on a gel) and sequenced Component C (21) = [TCTA]<sub>4</sub>[TGTA]<sub>2</sub>[TCTA]<sub>2</sub>[TGTA]<sub>2</sub>[TCTA]<sub>11</sub>

Table from SRM 2391c			Component					
Locus	A	В	c	D	E	0	F	
D1S1656	17.3.17.3	11.14	11.15	11,15,				
D2S1338	18,23	17,17	19,19	18,15	Genoty	ypes	are	provided for
D2S441	10,10	10,14	10,10	10		, ,		•
D3S1358	15,16	15,19	16,18	15,16	е	acn	com	ponent
D5S818	11,12	12,13	10,11	10,11				
D7S820	11,11	10.10	10,12	10,11				
D8S1179	13.14	10.13	10,17	10.13.1		1-1-	•	
D8S1115	15,16	15,17	9,9	9.15	Forensic labs using commercial			
D10S1248	15.16	13.13	12,16	12.15	CTD 4		1.:4	منمهمام اماريمما
D128391	18.3,22	19,24	19,23	18.3,19.	STRIYP	nng	KILS S	hould obtain
D13S317	8.8	9,12	11,11	8,1	+	ho c	ama	results
D16S539	10.11	10.13	10,10	10.1	L	ne s	anne	resuits
D18S51	12,15	13,16	16,19	12,15,1				
D198433	13,14	16,16.2	13.2,15.2	13,13.2,1				
D21S11	28,32.2	32,32.2	29,30	28,29,3	These m	ator	iale a	llow forensic
D2281045	15,15	15,17	16,16	15,1	THESE III	atti	1013 0	mow forchist
CSF1PO	10.10	10.11	10,12	10,1	labs to v	alid	ate/c	ertify:
FGA	21,23	20,23	24,26	21,23,2	1005 10 1	una	acc, c	citily.
Penta D	9,13	8,12	10,11	9,10,1				
Penta E	5,10	7,15	12,13	5,10,1				
SE33	16,18	17,18	28.2,31.2	16,18,28	<ul> <li>DNA ex</li> </ul>	xtra	ction	method
TH01	8,9.3	6,9.3	6,8	6,8,5	(1			
TPOX	8,8	8,11	11,11	8,1	<ul> <li>PCR (ki</li> </ul>	its a	nd cy	/cler)
vWA	18,19	17,18	16,18	16,18	•		,	•
Amelogenin	X.X	X.Y	X,Y	X.1	<ul> <li>Electro</li> </ul>	opho	retic	separation
DYS19	100	14	15	15				•
DYS385a		13	13	13	and de	eteci	ion	
DYS385b		17	15	15	• Allala	II:		£4
DYS389I		13	12	12	<ul> <li>Allele</li> </ul>	caiiii	ng so	itware
DYS389II		31	27	27			200	

# **Next Generation Sequencing**

Forensic Applications

- Going in depth into STR loci and beyond
  - STRs are useful for legacy (databases)
  - SNPs within STRs identify 'sub-alleles'
  - Millions of bases of sequence variants (SNPs)
- Opens up new human identity applications: biogeographical ancestry, externally visible traits, complex kinship, degraded samples, mixtures, low template, and other applications

Applications are currently being addressed by the forensic genetics community (Kayser and deKnijff 2011)

### **Next Generation Sequencing**

- Challenges
  - Repeating sequences (STRs) and read lengths
  - Sample amount requirements (> 1 ng)
  - Cost and time per unit of information
  - Data analysis (storage, assembly, interpretation)
  - Policy, privacy, disease related markers

  - Standards/reference materials
    - Nomenclature
    - Accuracy of sequence information
    - Errors, platform and bioinformatics-based bias

### Requirements for a NGS forensic SRM?

Information gathering stage

- Materials:
  - Genomic DNAs?
  - Cell line DNAs?
- Is a full genome standard needed for forensic applications?
- · Is it enough to fully characterize
- 'core' loci and have good/high — Use current forensic SRMs? confidence in other non-core loci? - PCR amplicons of forensically relevant markers?
- How many components?
  - Family samples paternity trio?
  - Mixtures?
- How much material is needed?
- Certify additional markers beyond core STR loci

Ongoing discussion with Multiplex Biomolecular Science group 'Genome in a bottle' consortium – Justin Zook and Marc Salit

## Characterization of a NGS forensic SRM?

- Involve multiple platforms/technologies?
  - Short and long read technologies
  - Genomic versus targeted (PCR product) sequencing
  - Interlaboratory studies (pilot prior to SRM production)
- For the core forensic markers (STRs, mito)
  - Continue to include Sanger confirmation
  - Develop specific primer sets for core loci?
- Call SNPs based on consensus from multiple platforms and assembly algorithms

### Vision for the new SRM

### **Certified for**

- Core autosomal STRs
- X and Y STRs
- · Mitochondrial genome
- 'Forensic' SNPs (identification, ancestry, kinship, and phenotype)
- Other: InDels, additional SNPs
- Sufficient material for NGS experiments
- STRs characterized by Sanger sequencing and multiple NGS platforms
- 'Forensic' SNPs characterized with assays (if available) and consensus between multiple NGS platforms
- Uses: test algorithms, library preparation, validation of platforms, enable consistency between labs/platforms

# Current Focus - Characterization of SRMs 2392 and 2392-I

- Used for mitochondrial DNA sequencing
- The mtGenome was amplified using an overlapping primer pair strategy
- The amplicons were purified, quantitated and pooled in an equimolar mixture (100 ng)
- Platforms
  - Ion Torrent (Edge Biosystems)
  - 5500xl (in house, to be completed this month)
  - Illumina (Beckman, results soon)
  - Hope to have multiple systems in house (2013)

### Human HL-60

- Sole component of NIST SRM 2392-I
- NGS accurately called all sequence variants compared to the rCRS (33 variants)
- 315.1 C insertion (homopolymer stretch)
- Heteroplasmy at 12,071 (correctly called)
- Heteroplasmy at 16,069? (artifact of PCR)

Thank you for your attention!	
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