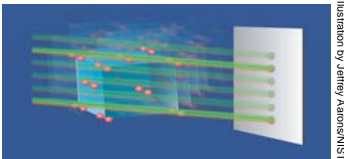


# Overview of the Human Identity Projects at NIST


Dr. Peter M. Vallone  
Applied Genetics Group  
U.S. National Institute of Standards and Technology (NIST)  
Presented to FSSA  
Adelaide, South Australia  
September 13, 2010

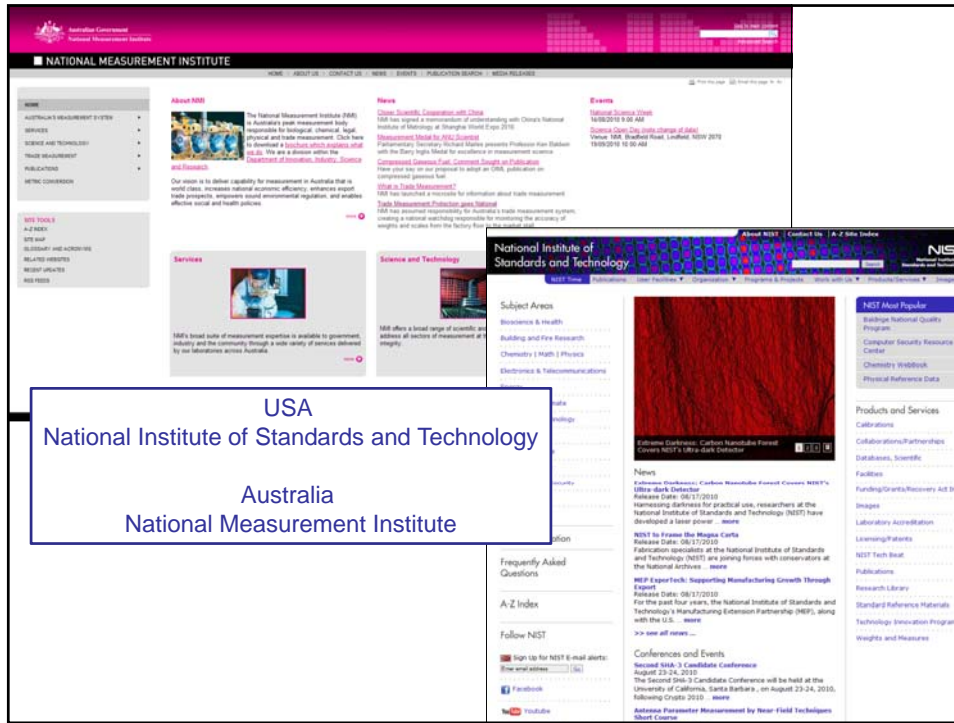
## What is NIST?

- U.S. National Institute of Standards and Technology
- Founded 1901
- Main campus in Gaithersburg, Maryland  
30 miles north of Washington D.C.



To promote U.S. innovation and industrial competitiveness by advancing **measurement science**, **standards**, and **technology** in ways that enhance economic security and improve quality of life.





## NIST Products and Services

### Measurement Research

- ~2,200 publications per year

### Standard Reference Data

- ~100 different types
- ~6,000 units sold per year
- ~130 million data downloads per year

Copyright Robert Rath

### Standard Reference Materials

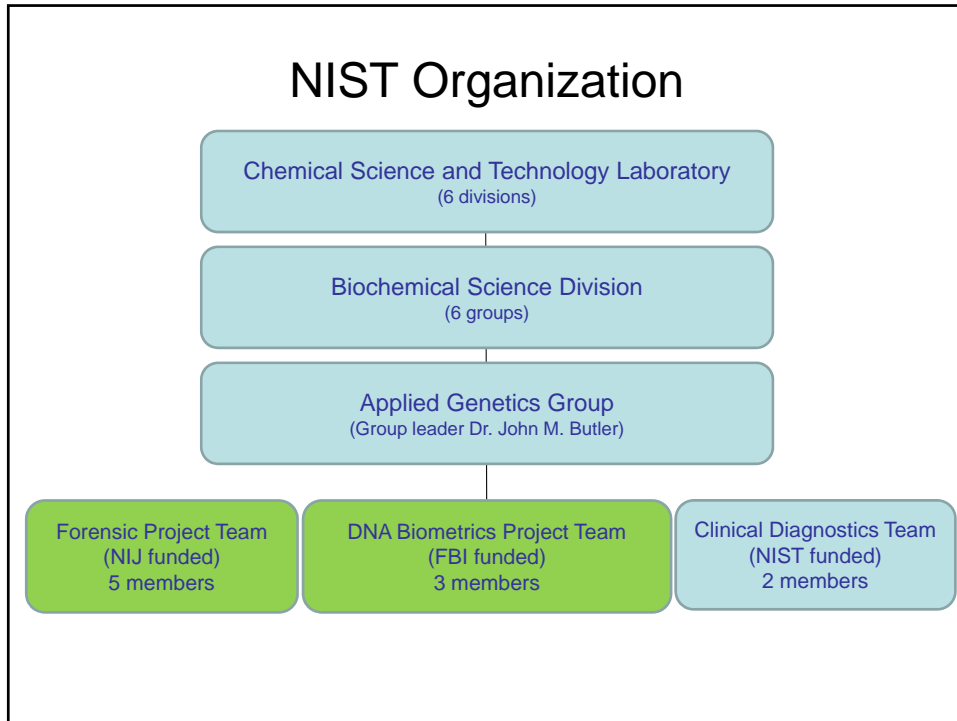
- ~1,300 products available
- ~33,000 units sold per year

### Calibration Tests

- ~16,000 test per year

### Laboratory Accreditation


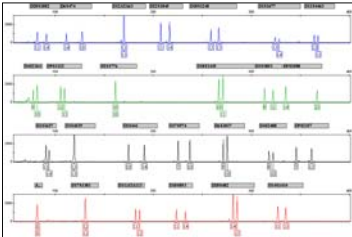

- ~800 accreditations of testing and calibration laboratories per year



### Our Group Goals

**Provide support for the forensic DNA typing community**

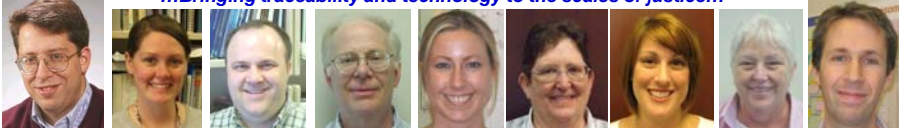
- Standards
- Technology
- Training and education



## The NIST Human Identity Project Team (Forensic DNA & DNA Biometrics)

Funding from the **National Institute of Justice (NIJ)** through the NIST Office of Law Enforcement Standards and the **FBI S&T Branch** through the NIST Information Access Division

...Bringing traceability and technology to the scales of justice...



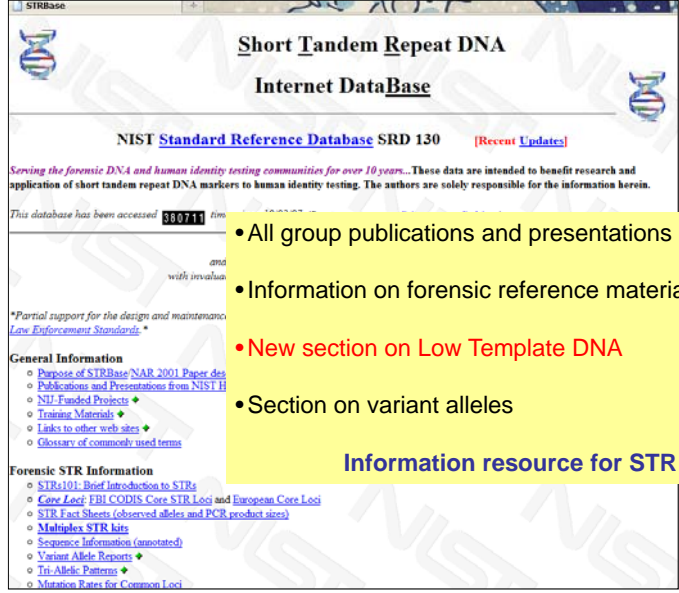
|   |                |               |                |               |                   |                     |               |   |
|---|----------------|---------------|----------------|---------------|-------------------|---------------------|---------------|---|
| John<br>Butler                                  | Erica<br>Butts | Mike<br>Coble | Dave<br>Duewer | Becky<br>Hill | Margaret<br>Kline | Kristen<br>O'Connor | Jan<br>Redman | Pete<br>Vallone                                   |
| <small>Project Leader,<br/>Forensic DNA</small> |                |               |                |               |                   |                     |               | <small>Project Leader,<br/>DNA Biometrics</small> |


|                              |                                   |                                   |                     |                           |
|------------------------------|-----------------------------------|-----------------------------------|---------------------|---------------------------|
| Workshops<br>& Textbooks     | Mixtures,<br>mtDNA & Y            | Concordance<br>& LT-DNA           | Kinship<br>Analysis | Rapid PCR<br>& Biometrics |
| DNA Extraction<br>Efficiency | Software Tools<br>& Data Analysis | Variant alleles<br>& Cell Line ID | STRBase<br>Support  |                           |

<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

## STRBase

<http://www.cstl.nist.gov/strbase/>





John  
Butler

- All group publications and presentations are available
- Information on forensic reference materials
- **New section on Low Template DNA**
- Section on variant alleles

**Information resource for STR typing**

## Projects within the Forensic and Biometric Project Teams

- Concordance testing with STR kits
- Additional STR loci
- DNA extraction efficiency
- Kinship analysis
- Rapid PCR protocols
- Mixture interpretation
- Low template PCR
- Workshops and training

## Concordance Testing



Becky Hill

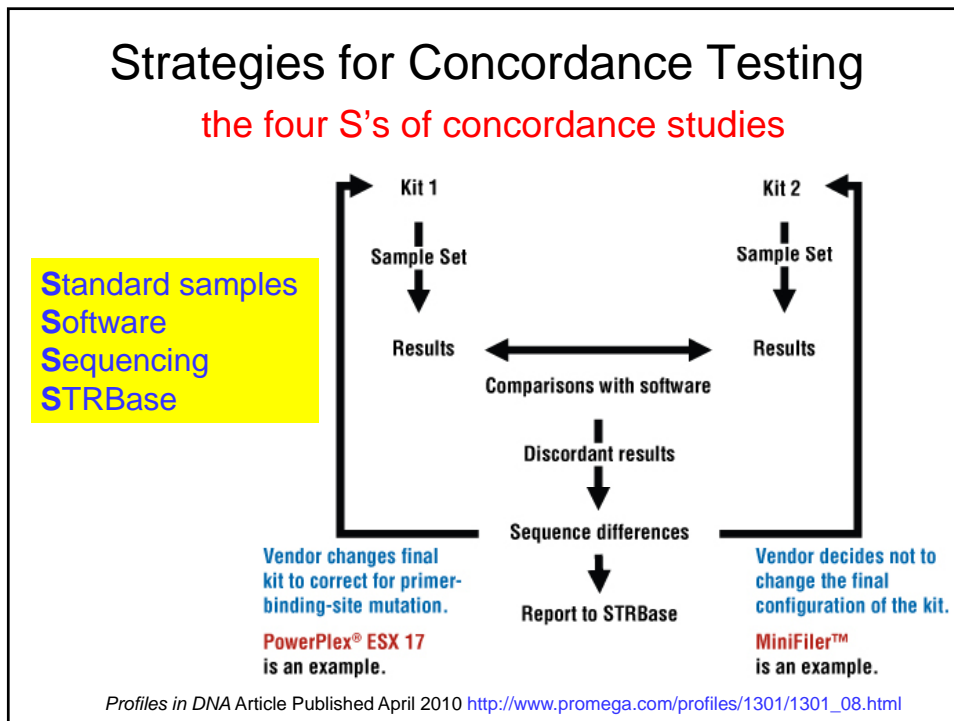
- When new STR typing kits are released, concordance testing is performed
  - Buffer and master mix improvements
  - PCR primer sequences may change
- Type a set of NIST U.S. population samples (n > 600) to evaluate concordance
- Provides a service to the forensic community and kit manufactures

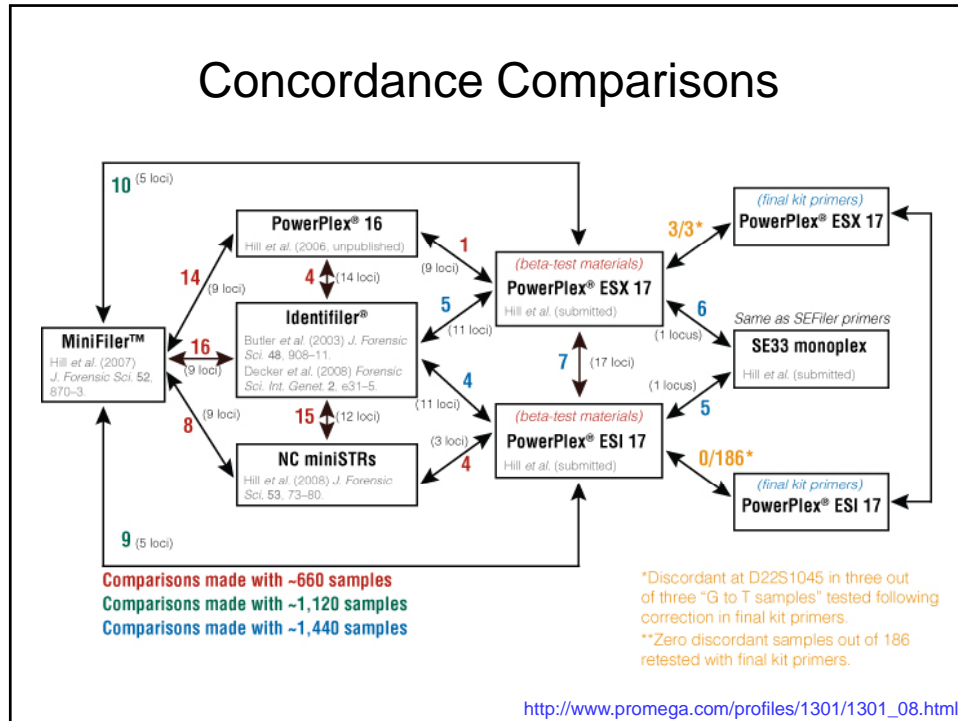
## STR Loci Present in Commercial Kits

| U.S.    |                |           | Europe    |          |         |          |
|---------|----------------|-----------|-----------|----------|---------|----------|
| PP16    | Identifiler    | MiniFiler | ESX/ESI17 | NGM      | SEfiler | SGM Plus |
| TPOX    | TPOX           |           |           |          |         |          |
| CSF1PO  | CSF1PO         | CSF1PO    |           |          |         |          |
| D5S818  | <u>D5S818</u>  |           |           |          |         |          |
| D7S820  | <u>D7S820</u>  | D7S820    |           |          |         |          |
| D13S317 | <u>D13S317</u> | D13S317   |           |          |         |          |
| FGA     | <u>FGA</u>     | FGA       | FGA       | FGA      | FGA     | FGA      |
| vWA     | <u>vWA</u>     |           | vWA       | vWA      | vWA     | vWA      |
| D3S1358 | <u>D3S1358</u> |           | D3S1358   | D3S1358  | D3S1358 | D3S1358  |
| D8S1179 | <u>D8S1179</u> |           | D8S1179   | D8S1179  | D8S1179 | D8S1179  |
| D18S51  | <u>D18S51</u>  | D18S51    | D18S51    | D18S51   | D18S51  | D18S51   |
| D21S11  | <u>D21S11</u>  | D21S11    | D21S11    | D21S11   | D21S11  | D21S11   |
| TH01    | TH01           |           | TH01      | TH01     | TH01    | TH01     |
| D16S539 | D16S539        | D16S539   | D16S539   | D16S539  | D16S539 | D16S539  |
|         | D2S1338        | D2S1338   | D2S1338   | D2S1338  | D2S1338 | D2S1338  |
|         | D19S433        |           | D19S433   | D19S433  | D19S433 | D19S433  |
|         |                |           | D12S391   | D12S391  |         |          |
|         |                |           | D1S1656   | D1S1656  |         |          |
|         |                |           | D2S441    | D2S441   |         |          |
|         |                |           | D10S1248  | D10S1248 |         |          |
|         |                |           | D22S1045  | D22S1045 |         |          |
|         |                |           | SE33      |          | SE33    |          |



U.S. is looking to expand the core loci (18-20 total) to provide more international overlap

Penta D  
Penta E





### Additional STR Loci

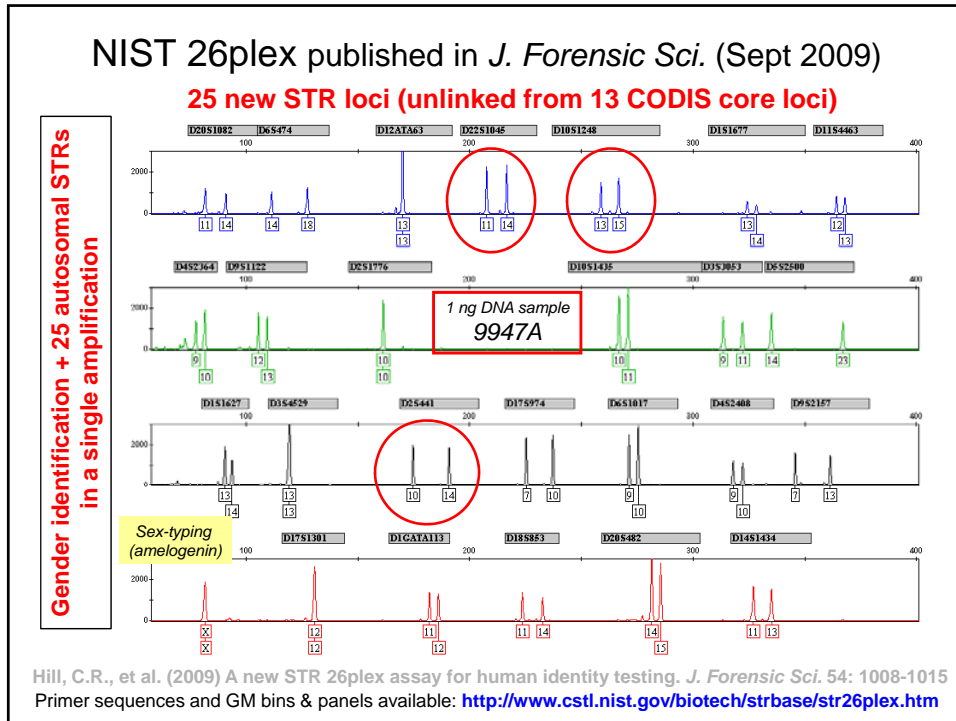
Pete Vallone

Becky Hill

- Characterizing additional STR loci for human identity testing (26 total)
  - Degraded DNA (miniSTRs)
  - Kinship analysis
  - Potential candidates for core loci
  - New core loci adopted by Europe
- In house assays developed for typing
- Population data collected on NIST samples

Europe adopts new loci **D10S1248**, **D14S1434** and **D22S1045** Gill *et al. Forensic Sci. Int.* 2006;156:242-244  
**D2S441** replaces **D14S1434** Gill *et al. Forensic Sci. Int.* 2006;163:155-157

Hill, C.R., Kline, M.C., Coble, M.D., Butler, J.M. (2008) Characterization of 26 miniSTR loci for improved analysis of degraded DNA samples. *J. Forensic Sci.* 53(1):73-80.



## DNA Extraction Efficiency



Erica Butts

- Using known amounts of DNA to estimate true extraction efficiency

### Sources of DNA:

- Highly characterized extracted DNA (52.44 ng/μL)
- Human cell lines (6 pg of DNA per cell)
- Whole blood (white blood cell count of 4.0 million WBC/mL)

The starting amount of DNA is known



## Qiagen EZ1 Advanced

EZ1 Advanced uses magnetic separation and multiple washes to purify DNA

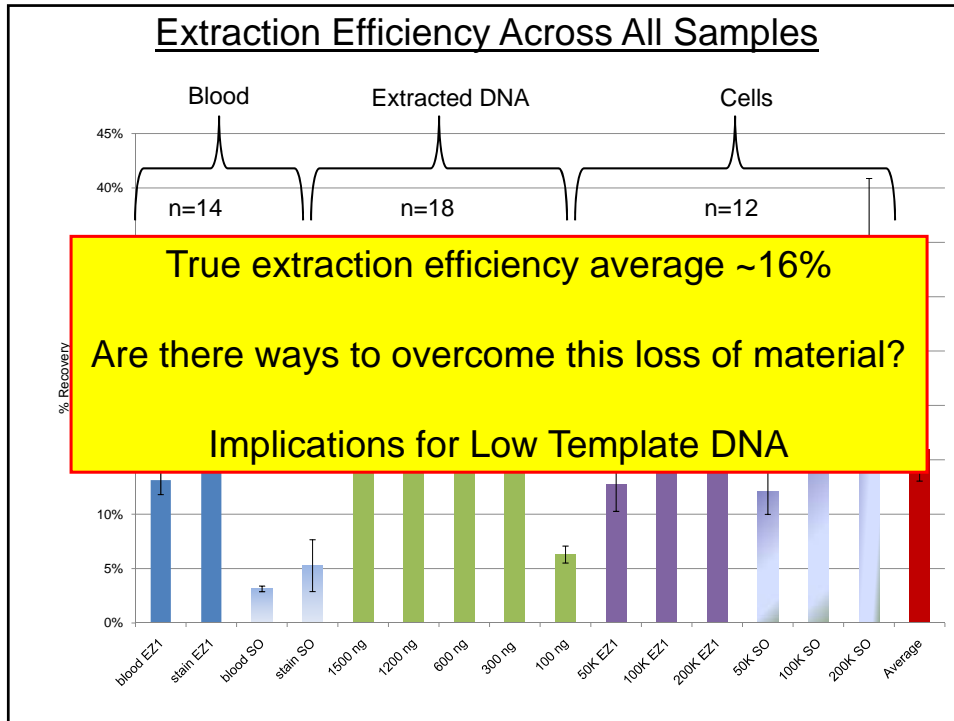
- Swabs & Stains: G2 Buffer and Proteinase K added to sample
- Blood: Total sample volume brought up to 200  $\mu\text{L}$  with G2 Buffer
- Incubated at 56°C for 15 minutes then 95°C for 5 minutes
  - Vortex periodically through incubation (~every 5 minutes)




## Modified Salt Out

- Manual extraction process
- Involves a Proteinase K digest
- Saturated Ammonium Acetate solution to separate DNA
- Absolute Ethanol wash to precipitate DNA
- Rehydrated with 100  $\mu\text{L}$  TE





## Kinship Analysis



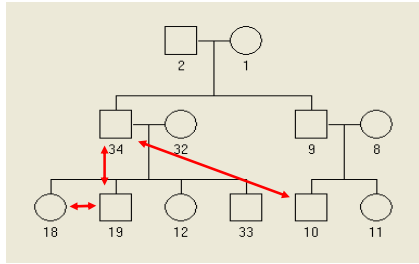
Kristen O'Connor

**Work by Kristen Lewis O'Connor, NIST NRC Postdoc**  
 (PhD research with Bruce Weir at University of Washington on familial search issues)

- Examine impact of additional STR loci (and other genetic markers) on addressing specific kinship questions
- Simulate likelihood ratio distributions with different sets of STR loci and different potential relationships
- Examine different software programs (and develop approaches for lab validation including investigating possible standard data sets for software testing)

## Likelihood Ratios with 15 Loci

$$LR = \frac{\text{Probability of the evidence (genotypes) given a particular hypothesis}}{\text{Probability of the evidence (genotypes) given an alternative hypothesis}}$$



Sets of multigenerational families

| Comparison             | LR for 34 & 19 | LR for 18 & 19 | LR for 34 & 10 |
|------------------------|----------------|----------------|----------------|
| Parent-Child           | 1.28E+06       |                |                |
| Full Siblings          |                | 2.76E+07       |                |
| Half Siblings          |                |                | 6.65E-01       |
| Cousins                |                |                |                |
| Uncle-Nephew           |                |                | 6.65E-01       |
| Grandparent-Grandchild |                |                | 6.65E-01       |

Parent/Child      Full Sibs      Uncle/Nephew

LR calculations were performed with GeneMarker® HIDv1.90

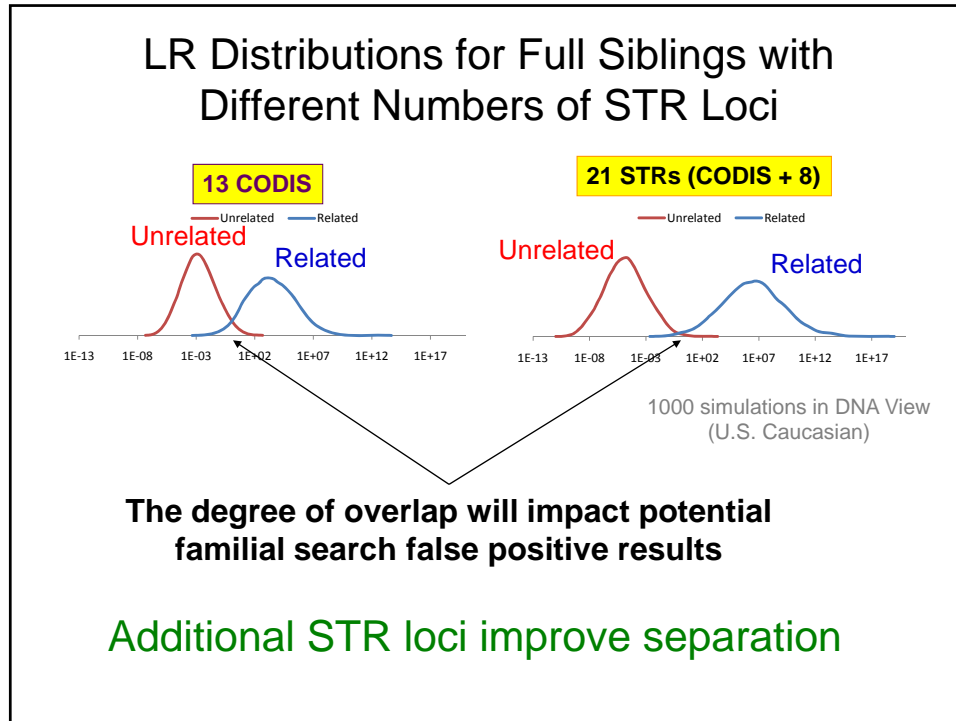
## Benefit of Additional Loci Likelihood Ratios with 40 Loci

| Comparison             | 15             | 40             | 15             | 40             | 15             | 40             |
|------------------------|----------------|----------------|----------------|----------------|----------------|----------------|
|                        | LR for 34 & 19 | LR for 34 & 19 | LR for 18 & 19 | LR for 18 & 19 | LR for 34 & 10 | LR for 34 & 10 |
| Parent-Child           | 1.28E+06       | 6.68E+16       | 9.08E+05       | 0.00E+00       | 0.00E+00       | 0.00E+00       |
| Full Siblings          | 3.22E+04       | 5.73E+12       | 2.76E+07       | 1.57E+19       | 6.07E-03       | 3.30E+03       |
| Half Siblings          | 7.38E+03       | 8.63E+11       | 4.89E+04       | 4.99E+12       | 6.65E-01       | 8.98E+05       |
| Cousins                | 1.95E+02       | 1.32E+08       | 8.96E+02       | 1.05E+09       | 1.52E+00       | 2.17E+04       |
| Uncle-Nephew           | 7.38E+03       | 8.63E+11       | 4.89E+04       | 4.99E+12       | 6.65E-01       | 8.98E+05       |
| Grandparent-Grandchild | 7.38E+03       | 8.63E+11       | 4.89E+04       | 4.99E+12       | 6.65E-01       | 8.98E+05       |



Parent/Child      Full Sibs      Uncle/Nephew

LR calculations were performed with GeneMarker® HIDv1.90

15 STR loci typed with commercial Identifier kit  
 25 STR loci typed with an in-house NIST assay  
 Hill, C.R., Butler, J.M., Vallone, P.M. (2009) A 26plex autosomal STR assay to aid human identity testing. *J. Forensic Sci.* 54(5): 1008-1015.



## Rapid and Direct PCR





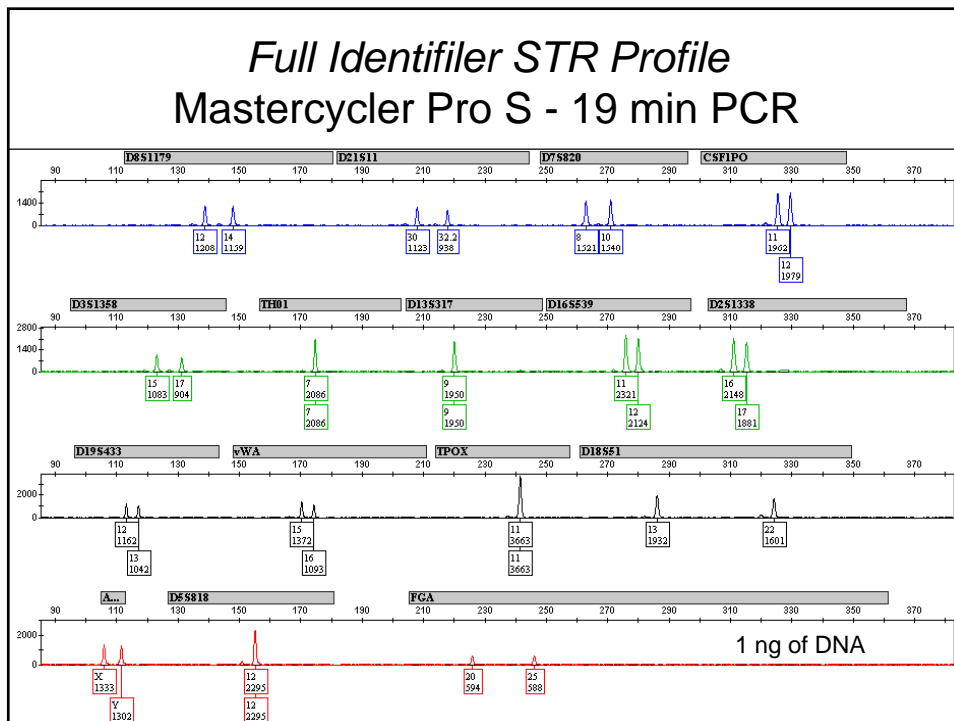
Pete Vallone

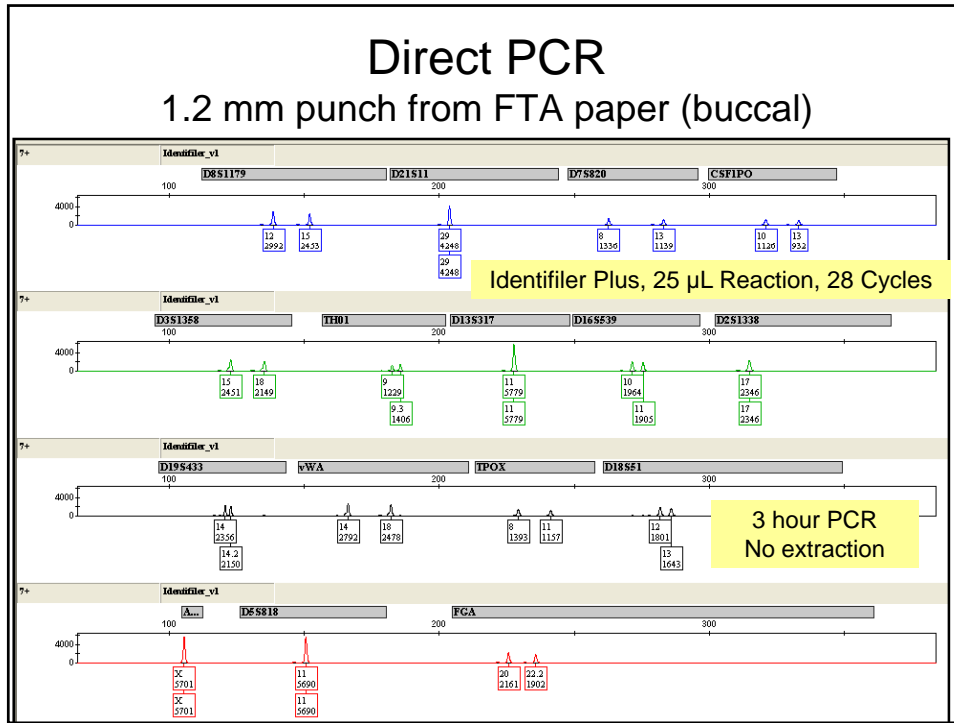
Erica Butts

- Developing methods for faster and robust amplification of STR typing kits
  - Reduce cycling time from 3 h to 30 min
- Faster thermal cyclers
- Faster DNA polymerases
- For reference samples (non-casework)
  - 1ng of template, single source
- Apply to rapid integrated systems for DNA typing


### DNA Analysis Approaches (non-integrated)

| Steps Involved           | Traditional Protocols  | Rapid Improvements (Buccal)  | Rapid Improvements (Direct PCR)  |
|--------------------------|--|--|--|
| Collection               |  |  |  |
| Extraction               | 1.5 hours<br><small>Manual</small>   | 30 min.<br><small>Automated Extraction<br/>Qiagen EZ1 Advanced</small> | <br>Blood Stain |
| Quantitation             | 1.5 hours<br><small>qPCR</small>   | 1.5 hours  |  |
| Amplification            | 3.5 hours  | <36 min.<br><small>Rapid PCR conditions</small>                        | 3 hour<br><small>Using PowerPlex 16 HS or Identifier</small>                                       |
| Separation/<br>Detection | 1 hour<br><small>Capillary Electrophoresis<br/>ABI 3130xl</small>          | 1 hour   | 1 hour   |
| Data Interpretation      | Time may vary depending on software, sample quality, and analyst expertise |  |  |
| Total Time               | Minimum ~7.5 hours   | ~4 hours   | ~4 hours   |






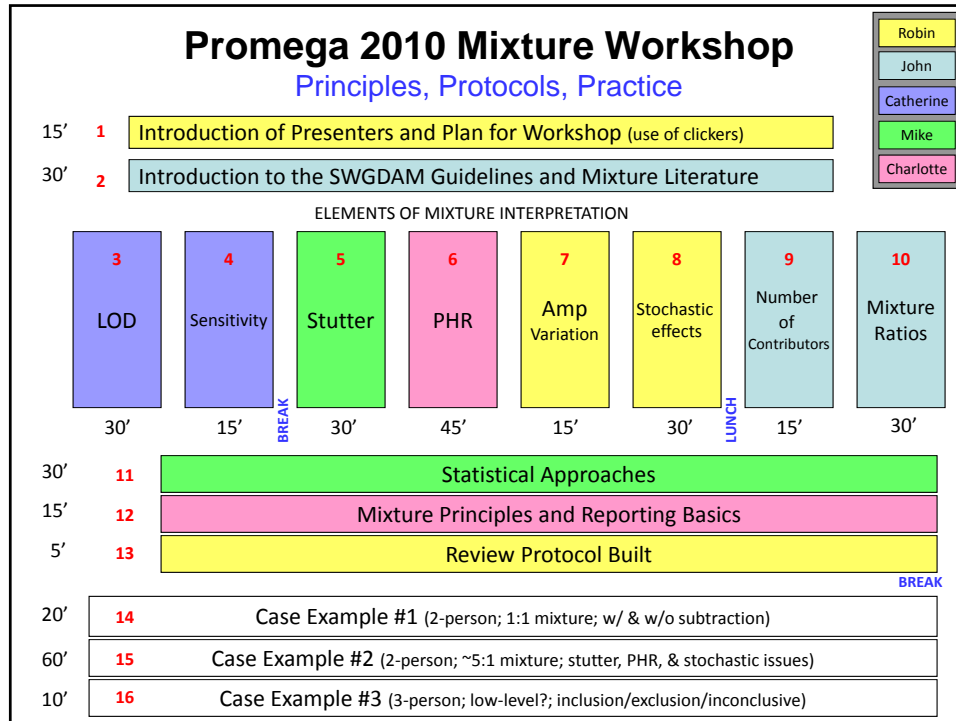
## DNA Mixture Interpretation




Mike  
Coble

- Evaluate software for DNA mixture interpretation
  - True Allele
  - FSS i<sup>3</sup> (update)
  - USACIL (custom software)
- Developing training materials and data sets based upon SWGDAM guidelines
  - ‘Mixture of the month’
  - Mixture section on STRBase





## Low Template DNA



Becky Hill

- Amplifying pristine (non-casework) DNA samples
- Varying
  - Amount of DNA (100 pg, 30 pg, 10 pg)
  - Number of cycles
  - STR kit (Identifiler, Identifiler Plus, Powerplex 16, HS)
- Evaluating
  - Allele drop out
  - Allele drop in
  - High stutter
  - Heterozygote peak height balance

Performing control experiments provides a baseline of PCR performance with low amounts of DNA template

## New STRBase Website on LT-DNA (LCN)

<http://www.cstl.nist.gov/biotech/strbase/LTDNA.htm>

**Information on Low Template / Low Copy Number DNA Testing**

**General Information**

- o [Purpose of STRBase/](#)
- o [Publications and Prese](#)
- o [NIJ-Funded Projects](#)
- o [Training Materials](#) ◆
- o [Links to other web site](#)
- o [Glossary of commonl](#)

**Forensic STR Informa**

- o [STRs101: Brief Introd](#)
- o [Core Loci: FBI COD](#)
- o [STR Fact Sheets \(obs](#)
- o [Multiplex STR kits](#)
- o [Sequence Information](#)
- o [Variant Allele Reports](#)
- o [Tri-Allelic Patterns](#) ◆
- o [Mutation Rates for Co](#)
- o [Published PCR primer](#)
- o [Y-chromosome STRs](#)

Sessions were held at several recent is widely referred to as low copy num readers better understand this topic.

\*At the International Society for Forensic Genetics (ISFG) Statistical Committee meeting (the Caddy report), Bruce Budowle (Science Service), At the International Symposium on Human Identification (formerly of Orchid Cellmark), Bruce Budowle (consultant, formerly of Orchid Cellmark), Bruce Budowle (NYC Office of Chief Medical Examiner), B...

**Presentations on LTDNA**

- o [John Butler - ISHI \(Promega\)](#)
- o [Becky Hill - ISHI \(Promega\)](#)
- o [Theresa Caragine - ISHI \(Pro](#)

**LTDNA Validation Data**  
Labs having validation data on this: [john.butler@nist.gov](mailto:john.butler@nist.gov)

**NIST Sensitivity Data with low level DNA templates**  
10 replicate amplifications for each condition with two fully heterozygous, single-source samples

Low Copy Number (LCN) DNA Panel Discussion

### Scientific Issues with Analysis of Low Amounts of DNA

**John M. Butler and Carolyn R. "Becky" Hill**  
National Institute of Standards and Technology

20<sup>th</sup> International Symposium on Human Identification  
Las Vegas, NV  
October 15, 2009

- o [Low-template DNA Information](#) NEW ←
- o [miniSTRs \(short amplicons\)](#) ◆
- o [Null Alleles](#) - discordance observed between STR kits ◆
- o [STR Reference List](#) - now 3303 references ◆

## Complete Set of NIST Sensitivity Data Available on New LT-DNA Website

<http://www.cstl.nist.gov/biotech/strbase/LTDNA.htm>

**NIST Sensitivity Data with low level DNA templates**  
10 replicate amplifications for each condition with two fully heterozygous, single-source samples

*Click on links to see summaries and DNA profiles observed*

| STR kit - PCR conditions                    | Sample 1               | Sample 2               |
|---|------------------------|------------------------|
| <a href="#">Identifiler - 28 cycles</a>     | <a href="#">100 pg</a> | <a href="#">100 pg</a> |
|   | <a href="#">30 pg</a>  | <a href="#">30 pg</a>  |
|   | <a href="#">10 pg</a>  | <a href="#">10 pg</a>  |
| <a href="#">Identifiler - 31 cycles</a>     | <a href="#">100 pg</a> | <a href="#">100 pg</a> |
|   | <a href="#">30 pg</a>  | <a href="#">30 pg</a>  |
|   | <a href="#">10 pg</a>  | <a href="#">10 pg</a>  |
| <a href="#">PowerPlex 16 HS - 31 cycles</a> | <a href="#">100 pg</a> | <a href="#">100 pg</a> |
|   | <a href="#">30 pg</a>  | <a href="#">30 pg</a>  |
|   | <a href="#">10 pg</a>  | <a href="#">10 pg</a>  |
| <a href="#">PowerPlex 16 HS - 34 cycles</a> | <a href="#">100 pg</a> | <a href="#">100 pg</a> |
|   | <a href="#">30 pg</a>  | <a href="#">30 pg</a>  |
|   | <a href="#">10 pg</a>  | <a href="#">10 pg</a>  |

**PowerPlex 16 HS – 34 cycles**

Legend: Green = full (correct) type, Red = allele dropout, Black = locus dropout, Black = allele drop-in

Sample #1 (MT97150)

Sample #2 (PT84411)

MT97150 - 10 pg, amp #1

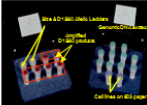




## Forensic DNA Standard Reference Materials

| SRM    | Name  | FY06 | FY07 | FY08 | FY09 | Avg   | Remaining | Current \$* |
|--------|---|------|------|------|------|-------|-----------|-------------|
| 2372   | Human DNA Quantitation Std                        | 0    | 0    | 160  | 147  | 153.5 | 1,078     | \$372       |
| 2390   | DNA Profiling                                     | 2    | 0    | 1    | 0    | 0.8   | 3         | \$833       |
| 2391B  | PCR-Based DNA Profiling                           | 86   | 81   | 125  | 140  | 108   | 107       | \$811       |
| 2392   | Mitochondrial DNA Sequencing                      | 8    | 6    | 0    | 12   | 6.8   | 165       | \$883       |
| 2392-1 | Mitochondrial DNA Sequencing<br>(Human HL-60 DNA) | 6    | 32   | 20   | 19   | 19.3  | 176       | \$365       |
| 2395   | Human Y-Chromosome DNA<br>Profiling               | 34   | 39   | 72   | 88   | 58.3  | 136       | \$383       |

\*As of Oct 7, 2009

### PCR-based DNA Profiling Standard

|   |  |   |  |  |
|---|--|---|--|--|
| <p><b>SRM 2391</b><br/>(1995)</p>  | <p><b>SRM 2391a</b><br/>(2000)</p>  | <p><b>SRM 2391b</b><br/>(2003, r2008)</p>  | <p><b>SRM 2391c</b><br/>(planned 2010)</p> <div style="border: 1px solid black; padding: 2px; font-size: small;"> <p>Would like your input into the desired number of components &amp; loci certified</p> </div> | <p><b>Will likely run out of SRM 2391b in mid-2011</b></p> |
|---|--|---|--|--|

\*coverage for all commercially available kit STR loci at the time of release

[http://www.cstl.nist.gov/biotech/strbase/pub\\_pres/Promega2009poster\\_SRM2391c.pdf](http://www.cstl.nist.gov/biotech/strbase/pub_pres/Promega2009poster_SRM2391c.pdf)

## SRM 2391c Plans

- 6 components (different DNA samples)
- Inclusion of a mixture sample
- Genomic DNA + DNA spotted on paper
- Will certify both autosomal STRs and Y-STRs
- Will replace both SRM 2391b and SRM 2395
  - Goal to have lower cost and single STR SRM
- Have typed >25 DNA candidate samples to assess allele ranges and combinations
- Have studied different formats
  - buccal swab DNA, etc.

## Workshops and Training

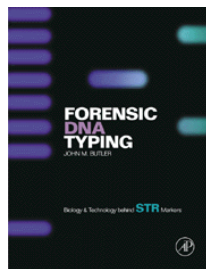
So far in 2010...

- 10 articles published
- Over 20 presentations given
- DNA as a Biometric Workshop (AAFS 2010)
  - Pete Vallone
- Mixture workshops (at Promega 2010 and AAFS 2011)
  - Mike Coble and John Butler

<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm#Presentations>

## The Expansion of *Forensic DNA Typing*

1<sup>st</sup> Edition

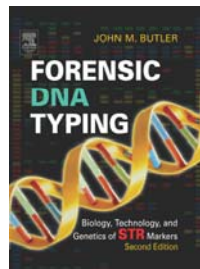


Jan 2001  
335 pp.  
17 chapters



John Butler

2<sup>nd</sup> Edition

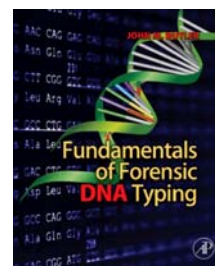


Feb 2005  
688 pp.  
24 chapters

**Chinese Translation (2007)** Y. Hou, translator

**Japanese Translation (2009)** Y. Fukuma, translator

3<sup>rd</sup> Edition



Sept 2009

**Fundamentals**  
18 chapters (504 pp.)










**Advanced Topics**  
25 chapters (~600 pp.)

**Planned for 2011**

**The NIST Human Identity Project Team**  
(Forensic DNA & DNA Biometrics)

Funding from the **National Institute of Justice (NIJ)** through the NIST Office of Law Enforcement Standards and the **FBI S&T Branch** through the NIST Information Access Division

*...Bringing traceability and technology to the scales of justice...*

|   |   |   |   |   |  |   |   |   |
|---|---|---|---|---|--|---|---|---|
|  |  |  |  |  |  |  |  |  |
| John Butler   | Erica Butts   | Mike Coble  | Dave Duewer   | Becky Hill  | Margaret Kline   | Kristen O'Connor  | Jan Redman  | Pete Vallone  |
| <small>Project Leader,<br/>Forensic DNA</small>                                   |   |   |   |   |  |   |   | <small>Project Leader,<br/>DNA Biometrics</small>                                   |
| Workshops<br>& Textbooks  | Mixtures,<br>mtDNA & Y  | Concordance<br>& LT-DNA   | Kinship<br>Analysis   | Rapid PCR<br>& Biometrics   |  |   |   |   |
| DNA Extraction<br>Efficiency  | Software Tools<br>& Data Analysis   | Variant alleles<br>& Cell Line ID   | STRBase<br>Support  |   |  |   |   |   |

<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>  
**peter.vallone@nist.gov**  
**301-975-4872**