

Concordance Testing Comparing STR Multiplex Kits with a Standard Data Set

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Email: becky.hill@nist.gov

Phone: 301-975-4275

Carolyn R. (Becky) Hill, Margaret C. Kline, David L. Duewer, and John M. Butler

National Institute of Standards and Technology (NIST), 100 Bureau Drive, Gaithersburg, MD 20899-8314

Poster available for download from STRBase:
http://www.cstl.nist.gov/strbase/pub_pres/Hill-ISHI2011-Concordance.pdf

Concordance evaluations are important to detect allelic dropout or "null alleles" present in a data set. These studies are performed because there are a variety of commercial STR multiplex kits with different configurations of STR markers available to the forensic community [1]. The electrophoretic mobility of the markers can vary between kits because the primer sequences were designed to amplify different polymerase chain reaction (PCR) product sizes. When multiple primer sets are used, there is concern that allele dropout may occur due to primer binding site mutations that affect one set of primers but not another [2]. These null alleles become evident only when data sets are compared. Null alleles are a concern because they can result in a false-negative or incorrect exclusion of samples that come from a common source (if the samples have been typed with kits using different PCR primers). A base pair change in the DNA template at the PCR primer binding region can disrupt primer hybridization and result in a failure to amplify and detect an existing allele [2].

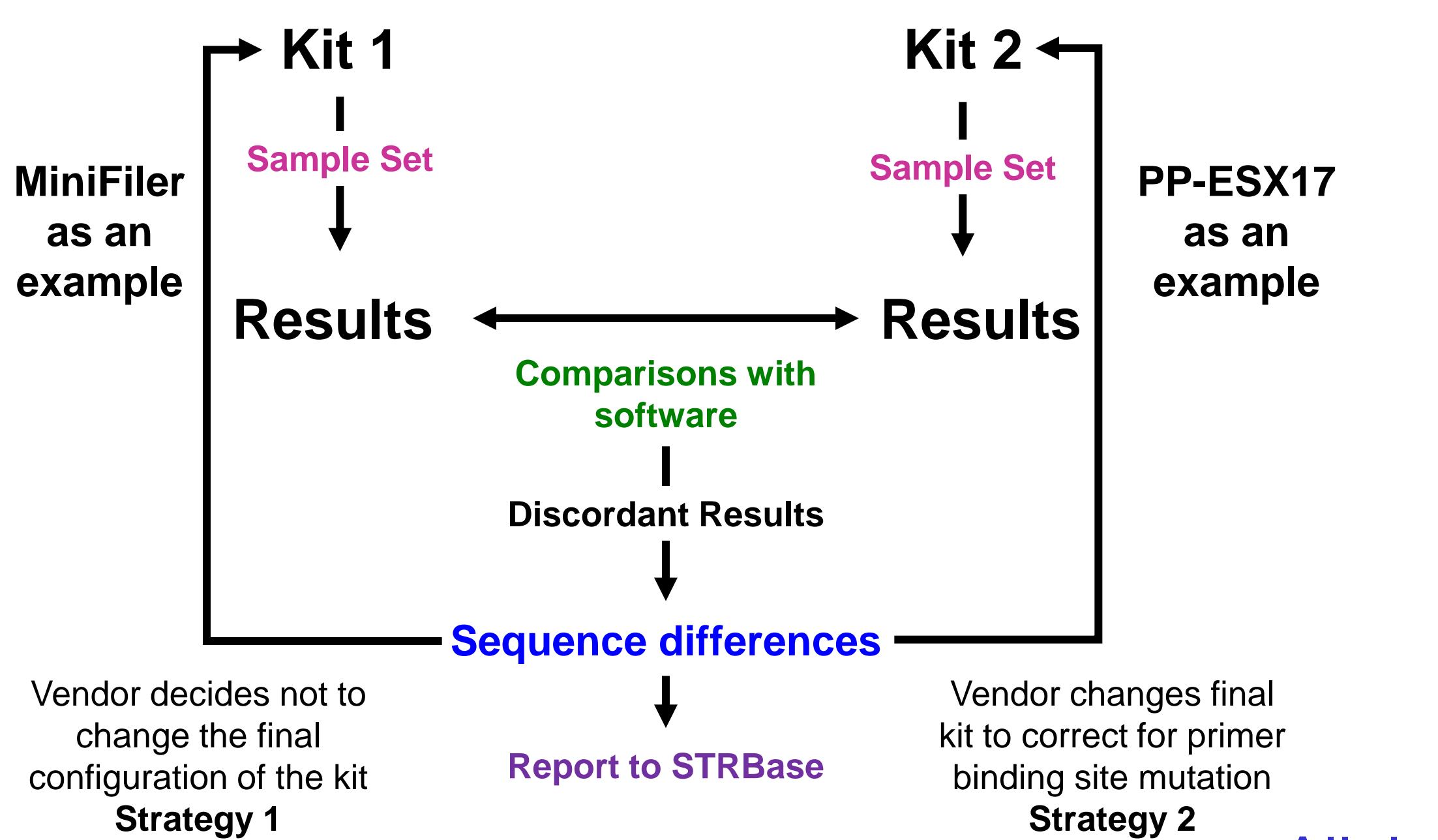
Multiple concordance studies have been performed at NIST with a standard sample set (~1450 in-house U.S. population samples) using various STR multiplex kits including Applied Biosystems Identifiler, MiniFiler, NGM, NGM SElect, SGM Plus, and Profiler Plus kits, Promega PowerPlex 16, ESX 17 and ESI 17 Systems, and Qiagen ESSplex, ESSplex SE, IDplex, and Hexplex kits [3-8]. Various discordant results have been identified using concordance software developed at NIST, confirmed by DNA sequencing and reported to the forensic community on the null allele web page of STRBase [9]. A summary of the results of our concordance studies will be shown in order to help assess the benefits of performing concordance testing using a standard data set with STR multiplex kits that have different primer sequences for the same markers.

Why Concordance Testing is Important

Concordance studies are necessary when comparing genotypes of samples that have been tested with STR multiplex kits that have varying configurations of the same loci. These markers are able to be in different positions because they have alternative primer sequences from one another. There is the potential for allelic dropout or a "null allele" if a primer binding site mutation impacts one of the primer pairs. The use of non-overlapping primers permits the detection of allele dropout. A base pair change in the DNA template in the PCR primer binding region can disrupt the hybridization of the primer and result in a failure to amplify and detect an existing allele. Null alleles are a concern because discordant results can impact DNA databases. Therefore, when discordant results are discovered, it is important to confirm the result with DNA sequencing to fully understand and characterize the direct cause of the mutation. At NIST we also establish concordance with standard reference materials (SRM 2391c: PCR-based Profiling Standard) and all next-generation STR typing kits.

NIST Strategies for Concordance Testing

NIST Four "S's" of Concordance: (a) NIST in-house samples are run with multiple kits/in-house assays to compare genotyping results; (b) concordance software is used to detect the differences between the results; (c) DNA sequencing is performed to confirm the results and determine the cause of the null allele; and (d) the final discordant results and verified null alleles are reported to the forensic community on STRBase: <http://www.cstl.nist.gov/strbase/NullAlleles.htm> [1]



NIST Samples Tested (~1450 Samples)

NIST U.S. population samples

- 260 African American, 262 Caucasian, 140 Hispanic, 3 Asian

U.S. father/son paired samples

- 200 African American, 200 Caucasian, 200 Hispanic, 202 Asian

50 anonymous blood samples

- 46 African American, 4 Caucasian

NIST SRM 2391c, PCR-based DNA Profiling Standard

- 4 genomic samples (with one mixture)
- 2 cell line samples (FTA and 903 paper)

Commercial and In-house STR Multiplex Kits Used in NIST Concordance Studies

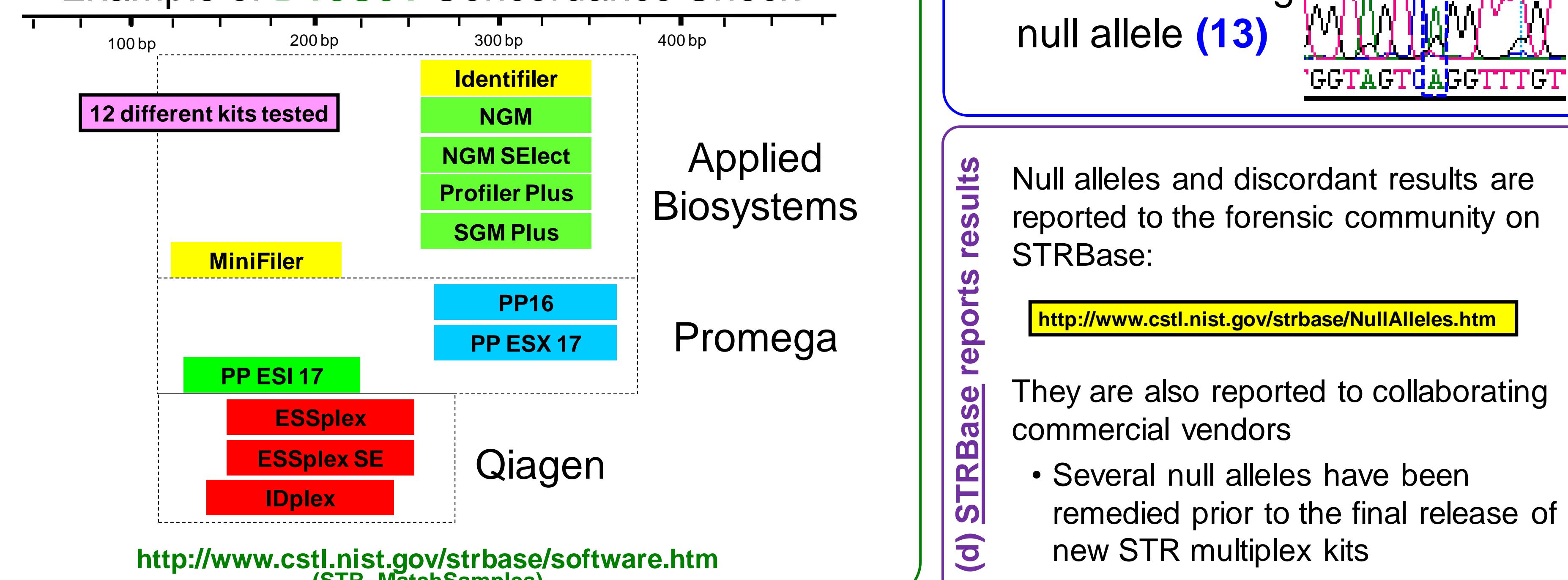
Applied Biosystems AmpF [®] STR Kits	Promega PowerPlex Systems	Qiagen Investigator HID Kits	In-House NIST Assays
Identifiler [3]	PowerPlex 16	ESSplex	miniSTRs [6,7]
Minifiler [4]	PowerPlex ESX 17 [5]	ESSplex SE	26plex [8]
Profiler Plus	PowerPlex ESI 17 [5]	Hexplex ESS	
SGM Plus	*PowerPlex 18D	IDplex	
NGM			
NGM SElect			

*PowerPlex 18D studies were performed by Erica Butts and Peter Vallone at NIST

Numbers of Primer Sets Compared for Each Marker

Marker	# of Sets	Marker	# of Sets
Amelogenin	13	D2S441	9
D18S51	12	D19S433	9
D21S11	12	D1S1656	7
FGA	12	D12S391	7
D3S1358	11	SE33	5
TH01	11	D5S818	4
D16S539	11	D7S820	4
vWA	11	D13S317	4
D8S1179	11	TPOX	3
D2S1338	10	CSF1PO	4
D10S1248	9	Penta D	1
D22S1045	9	Penta E	1

Example of D18S51 Concordance Check



Null alleles and discordant results are reported to the forensic community on STRBase:

<http://www.cstl.nist.gov/strbase/NullAlleles.htm>

They are also reported to collaborating commercial vendors

- Several null alleles have been remedied prior to the final release of new STR multiplex kits

<http://www.cstl.nist.gov/strbase/software.htm> (STR_MatchSamples)

Alleles Impacted by Insertions, Deletions, or Primer Binding Site Mutations

Marker	Kits with Correct Genotype	# Kits Compared	Correct Type	Kits with Null Allele/Discordant Genotype	Incorrect Type	Total Samples	Sequence Issue
Amel	ID/ESX17/ES17/PP16/MiniFiler/Pro+SGM-/ESS/ESSplexSE/IDplex/NGMs/Hexplex	13	X,Y		NGM*	657	yet to be determined
Amel	ID/ESX17/ES17/PP16/MiniFiler/Pro+SGM-/ESS/ESSplexSE/IDplex/NGMs/Hexplex	8	X,Y		NGM*	780	yet to be determined
Amel	ID/ESX17/ES17/PP16/MiniFiler/Pro+SGM-/ESS/ESSplexSE/IDplex/NGMs/Hexplex	13	X,Y		NGM*	653	yet to be determined
CSF1PO	PP16/MiniFiler/IDplex/PP18D	5	11,11	PP16 call	11,11,1	656	1 bp ins in ID amplicon outside of PP16 and MiniFiler primers [2]
CSF1PO	ID/MiniFiler/PP16/PP18D	5	9,12		CFT SNP 16 bp ds from repeat	12,12	662
D10S1248	ESX17/ES17/NGM/NGMs/ESS/ESSplexSE	7	14,16		G/FT SNP 24 bp ds from repeat	14,14	653
D10S1248	ESX17/ES17/NGM/NGMs/ESS/ESSplexSE	7	14,16		G/FT SNP 24 bp ds from repeat	14,14	653
D13S317	Pro+PP16/IDplex	5	9,11	Minifiler	11,11	656	4 bp del in the rev Minifiler primer binding site [2]
D13S317	Pro+PP16/IDplex	5	9,11	Minifiler	11,11	656	4 bp del in the rev Minifiler primer binding site [2]
D13S317	Pro+PP16/IDplex	5	9,12	Minifiler	12,12	656	4 bp del in the rev Minifiler primer binding site [2]
D13S317	Pro+PP16/IDplex	5	8,12	Minifiler	12,12	656	4 bp del in the rev Minifiler primer binding site [2]
D13S317	Pro+PP16/IDplex	5	10,13	Minifiler	13,13	656	4 bp del in the rev Minifiler primer binding site [2]
D13S317	Pro+PP16/IDplex	5	9,11	Minifiler	14,14	656	4 bp del in the rev Minifiler primer binding site [2]
D13S317	Pro+PP16/IDplex	5	9,12	Minifiler	14,14	656	4 bp del in the rev Minifiler primer binding site [2]
D13S317	Pro+PP16/IDplex	4	9,10	Minifiler	10,10	481	4 bp del in the rev Minifiler primer binding site [2]
D13S317	Pro+PP16/IDplex	4	10,11	Minifiler	11,11	481	4 bp del in the rev Minifiler primer binding site [2]
D18S51	Pro+PP16/IDplex	4	8,10	Minifiler	8,8	481	4 bp del in the rev Minifiler primer binding site [2]
D18S51	Pro+PP16/IDplex	4	10,12	Minifiler	12,12	481	4 bp del in the rev Minifiler primer binding site [2]
D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	11,15		CFT SNP 10 bp ds from repeat	11,11	653
D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	9,11		A/G SNP in Minifiler primer binding site [2]	9,9	656
D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	9,11		A/G SNP in Minifiler primer binding site [2]	12,12	656
D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	11,12		A/G SNP in Minifiler primer binding site [2]	11,11	656
D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	9,11		A/G SNP in Minifiler primer binding site [2]	14,14	481
D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	11,12		A/G SNP in Minifiler primer binding site [2]	14,14	481
D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	9,11		A/G SNP in Minifiler primer binding site [2]	14,14	481
D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	11,12		A/G SNP in Minifiler primer binding site [2]	14,14	481
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D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	11,12		A/G SNP in Minifiler primer binding site [2]	14,14	481
D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	9,11		A/G SNP in Minifiler primer binding site [2]	14,14	481
D18S51	ESX17/ES17/PP16/NGM/NGMs/ESS/ESSplexSE/IDplex	11	11,12		A/G SNP in Minifiler primer binding site [2]</td		