

Development of a New Autosomal STR Multiplex with Additional Loci to Benefit Human Identity Testing

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Disclaimers

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Standards

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Our publications and presentations are made available at:
<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

Questions to Address

- **Which loci did I use and why?**
 - Why additional loci are important
 - Characterization of additional STR loci
- **How did I combine these new loci into one multiplex?**
 - The “Autoplex” 5-dye single amplification reaction
- **How can this be useful?**
 - SRM 2391b has been updated with certified and reference values for new loci

Which Loci Were Used and Why? Going Beyond the CODIS Loci

Aren't the Current STR Loci Good Enough?

- For general forensic matching of evidence to suspect, the 13 CODIS STR loci are sufficient
- For other human identity/relationship testing questions, more autosomal loci can be beneficial or even necessary

More Loci are Useful in Situations Involving Relatives

- **Missing Persons** and Disaster Victim Identification (kinship analysis)
- Immigration Testing (often limited references)
 - Recommendations for 25 STR loci
- Deficient Parentage Testing
 - often needed if only one parent and child are tested

Relationship testing labs are being pushed to answer more difficult genetic questions...and **we want to make sure the right tools are in place**

Characterizing New Loci

- New loci were chosen based on the following characteristics:
 - Genomic Position
 - Polymorphic Content
 - Span/Range of observed alleles
- Details about the characterization process have all been previously reported at length:

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Eighteenth International Symposium on Human Identification.
 See <http://www.promega.com/geneticidproc/>

- John Butler's talk at the 18th International Symposium on Human Identification (Promega 2007), "New Autosomal and Y-Chromosome STR Loci: Characterization and Potential Uses"

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Characterization of 26 MiniSTR Loci for Improved Analysis of Degraded DNA Samples

- Characterization of **26** new autosomal loci
- Primer sequences, GeneMapper bins and panels, genotypes on common samples, and allele frequency information **already available on STRBase**

<http://www.cstl.nist.gov/div831/strbase/miniSTR.htm>
<http://www.cstl.nist.gov/div831/strbase/newSTRs.htm>

Multiple Miniplexes

- 26 characterized loci** divided into 10 miniplexes
- One locus per dye color
- Allelic ladders created
- Amplicons <140 bp**
- miniSTRs
- Work with 100 pg DNA
- For degraded samples** (bones in missing persons cases)

NC = Non-CODIS or non-core

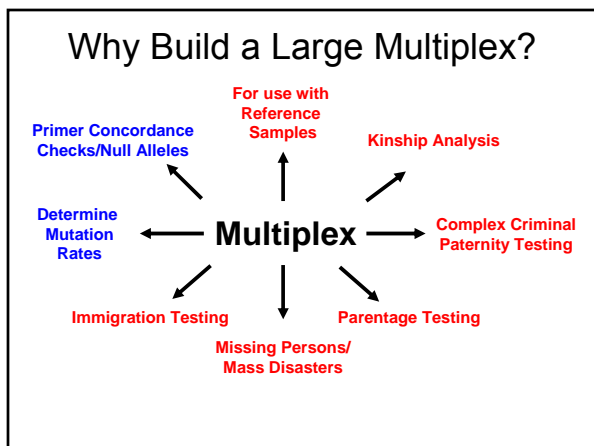
NC01 Loci

See Dixon et al. (2006) *Forensic Sci. Int.* 164: 33-44.

NC Miniplexes

NC01	NC02	NC03	
D10S1248	D1S1677	D3S3053	
D14S1434	D2S441	D6S474	
D22S1045			
	26 total new loci		
NC04	NC05	NC06	NC10
D1GATA113	D1S1627	D3S4529	D3S3053
D2S1776	D8S1115	D9S2157	D6S474
D4S2408	D9S324	D10S1430	D20S482
NC07	NC08	NC09	
D9S1112	D17S1301	D10S2327	
D12ATA63	D18S8534	D11S4463	
D14S1280	D20S1082	D17S974	

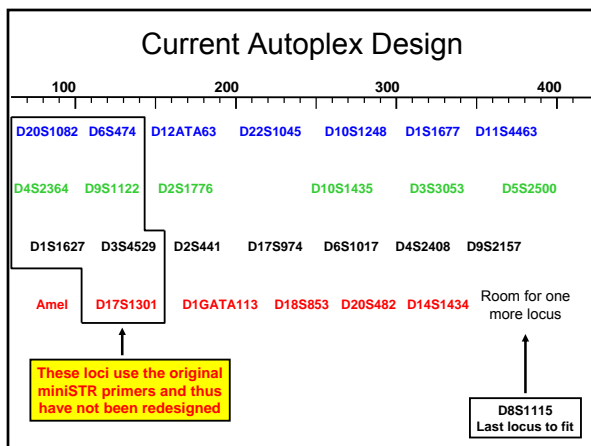
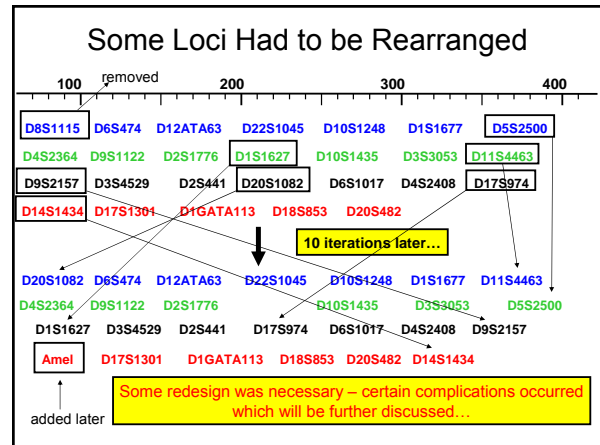
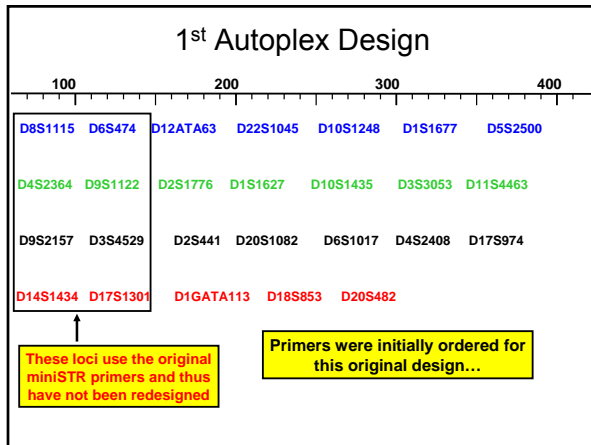
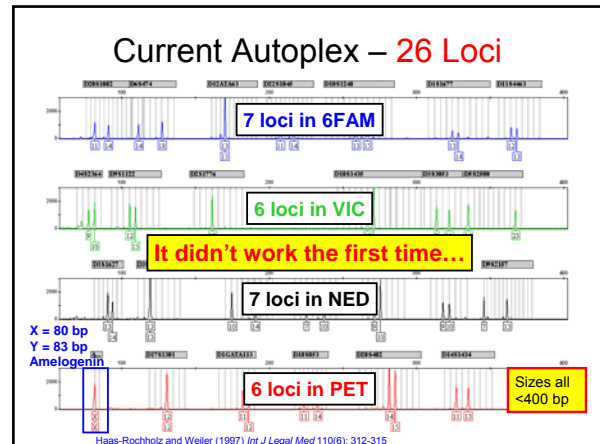
Removed because they were problematic



The Multiplex Design Process

The Design of the Multiplex

- **Goal:** A single amplification 5-dye multiplex to combine the 26 new autosomal loci + Amelogenin in one reaction (27plex)
- How can this be achieved?
 - Initial placement of all loci within 6FAM, VIC, NED, and PET dye channels (the size standard is in the LIZ channel)
 - Primer redesign for all but 7 of the original miniSTR loci
 - Trial and error of primer compatibility, as well as balancing for all working primers

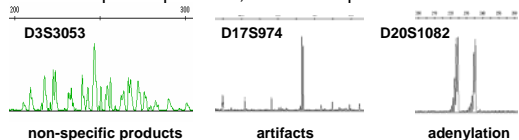


Where do we begin?

- Once the initial design has been laid out, primers were designed using Primer3
- Next, they were tested for multiplex-ability with AutoDimer developed by Pete Vallone
- Finally, the newly designed primers were mapped in LaserGene to confirm amplicon size and ensure primers flank the repeat
- Primers were then ordered
 - Forward fluorescent dye-labeled from ABI
 - Reverse non-dye labeled from Operon

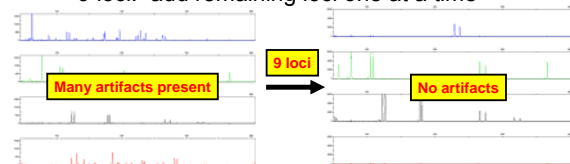
What Next?

- 1st step: Once received, all new primers were amplified in singleplex
 - If the primers fail in singleplex, they will most likely fail in multiplex – **Candidates for redesign**
 - Initially, 7 loci out of 26 failed – 19 worked in singleplex
- How do we define failure?
 - Incomplete adenylation, presence of artifacts, low signal, non-specific products, or no PCR product



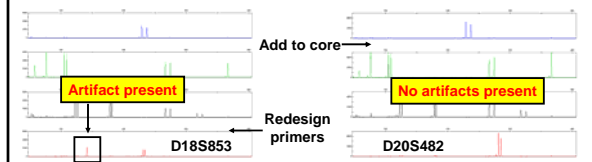
Begin Building the Multiplex

- Next, successful primers were tested in multiplex – 19 loci were combined together
- Then, the minimum amount of loci that work well together were determined
 - Can isolate by dye color – amplify loci in one dye channel at a time to hone in on failures
 - 9 loci: add remaining loci one at a time



Trial and Error

- When a locus was added to the “core-9” multiplex, decide whether it works or not
 - Check for artifacts and adenylation
- Then, build the multiplex from there
 - Keep adding to the core multiplex and redesign all primers that fail



Finalizing the Multiplex

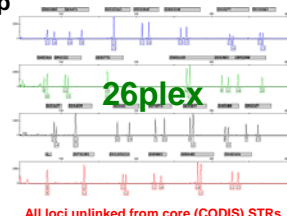
- Eventually the multiplex gained more loci with clean profiles: 18, 19, and 23plex
- Locus to locus primer balancing was performed with each multiplex
- Concordance and Mutation Rate studies were performed with the 23plex
- Currently, the multiplex has all but one locus (D8S1115) and we are working on redesigning further out (possibly > 400 bp)

Lessons Learned from Primer Redesign

- Some loci had to be redesigned in a completely different location
 - If artifacts are present or if it is a noisy baseline
- The fluorescent dye label can be switched to the reverse primer to mask an artifact
 - Can check forward and reverse primers separately
- With adenylation issues, a PIGTAIL (GTTTCTT) can be added to the 5' end of reverse primers
 - D1S1677, D3S3053, D11S4463, and D12ATA63
- Dye blobs can be filtered out with Edge Columns
 - Especially in the PET dye channel

Autoplex

- So far **25 STRs and amelogenin** in single multiplex (Eventual goal to have all 26 loci)
- Multiple loci in four dye channels
- **Amplicons 70 to 400 bp** (No longer 'miniSTRs')
- Typically use 1 ng DNA, 30 cycles
- **For reference samples** (a missing person's relatives)
 - “Autoplex” or “miniMegaplex”
 - All loci unlinked from core (CODIS) STRs

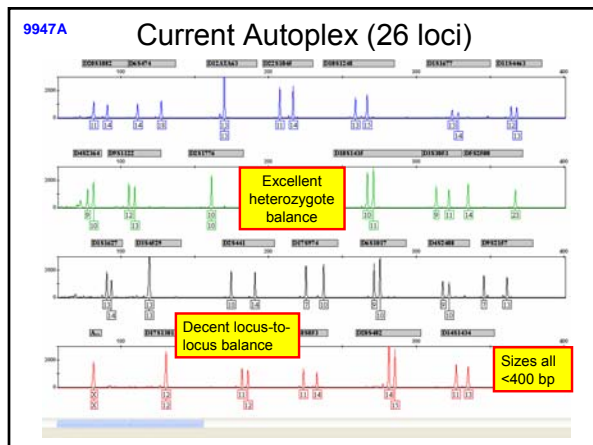


PCR Parameters

- Master Mix (MM)
 - 2 mM MgCl₂
 - 1x PCR Buffer
 - 1 Unit TaqGold
 - 0.2 μM Primer mix
 - 250 mM dNTPs
 - 0.16 mg/mL BSA
- 20μL reaction volume = 19μL MM + 1μL DNA sample (~1ng)

Thermal Cycling Conditions

- Conditions for ABI 9700 in 9600 emulation mode
- 95°C Hot Start for 11 min
- 30 cycles
 - 94°C for 45 sec **Denaturation**
 - 59°C for 2 min **Annealing**
 - 72°C for 1 min **Elongation**
- 60°C soak for 60 min
- 25°C hold (∞)



Further Work with the Autoplex Studies were Performed with the 23plex

Evaluation of Autoplex (23plex)

- 660 U.S. population samples
 - U.S. Caucasian, African American, Hispanic
 - **Concordance testing** compared to miniSTR results
- 790 father/son samples
 - U.S. Caucasian, African American, Hispanic, Asian
 - **Mutation rate determination**
- 12 samples for **extended family testing**

>1450 samples examined so far
(multiple primer batches prepared)

Concordance Study to Check for Null Alleles

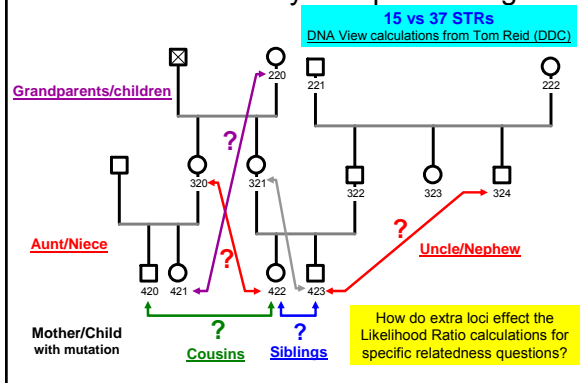
<http://www.cstl.nist.gov/biotech/strbase/NullAlleles.htm>

- 639 samples compared
- 14,058 total types (639 x 22 loci)
- 28 types discordant (0.20%)
- 99.80% concordance
- **Discordance has not yet been confirmed by sequencing**

Mutation Rates Measured for New STRs

- **395 father/son pairs** (790 samples total)
- 22 STR loci examined
- 8690 allelic transfers
- Only **6 mutations** were observed in total
- **0.069% mutation rate**
- 2-3 times less than typical 0.2% for common STRs

Extended Family Sample Testing



Comparison of Likelihood Ratios

Relationship Examined	15 STRs (Identifiler, ID15)	ID15 + Autoplex 22 STRs = 37 loci (A37)
Mother/Child* (*with single mutation)	0.214	5,200,000 Extra loci help...
Siblings	477	113,000 Extra loci help...
Uncle/Nephew	824	247,000 Extra loci help...
Cousins	0.45	2.25
Grandparents/ Grandchildren	0.53	1.42

Conclusions: Longer distance multi-generational questions cannot usually be solved with additional autosomal STRs...

SRM 2391b: DNA Profiling Standard Certificate of Analysis Update

- Genotyping and sequencing have been performed with SRM 2391b components (#1-12) for all 26 additional loci
- Cert... **More information on this to come in Margaret Kline's presentation on Saturday morning...** ...ed to all re...
- The Certificate of Analysis is in the process of being updated for all 26 loci (coming up in near future...)
- Using these values, bins and panels have been written in GeneMapper/ID
- **Purpose:** No commercial allelic ladders are available, but all genotypes are certified for the components of SRM 2391b

http://www.cstl.nist.gov/biotech/strbase/miniSTR/miniSTR_NC_loci_types.htm

In the Future...

- The D8S1115 will be redesigned to fit into the Autoplex – 27plex goal
- Allelic ladders for the final Autoplex will be prepared (made available to interested companies)
- All information will be available on STRBase
- A comprehensive publication is currently being written for submission to a forensic journal
- Use in new applications such as Rapid PCR (**initial findings to be presented by Pete Vallone on Saturday morning**)







Summary/Conclusions

- **26 unlinked loci** have been characterized and we have developed multiple miniplexes and an Autoplex (26plex)
- The Autoplex is a robust single amplification 5-dye multiplex reaction that can benefit the forensic community for reference purposes and relationship testing
- **NIST SRM 2391b** will include certified and reference values on these 26 additional autosomal STR loci

Thank you for your attention...

Our team publications and presentations are available at:
<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

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Collaborators			
 Jan Redman	 Amy Decker	 Dave Duewer	Mike Coble (now AFDIL) – early miniSTR work Tom Reid (DDC) - father/son samples