

**Using the Scientific Literature  
to Become a Lifelong Learner  
and NIST Research Update**

John M. Butler  
National Institute of Standards and Technology

AFDAA winter meeting (Austin, TX)  
January 28, 2010

*Becoming an Expert Witness – Literature Tracking*

**Presentation Outline for Literature Portion**

- Why bother with the literature?
- What is the literature you should be concerned with reading?
- What are some strategies for reading the literature?
- What resources exist for finding papers?
- What resources exist for storing and retrieving information related to the literature?
- How do I go about writing an article?

**Are You an Expert?**

- What kind of expert witness will you be?
- Do you know the field as well as you need to?
- **Reading the literature is critical to your ability to be an effective expert!**

**Why Discuss the Literature?**

- Hopefully you have a desire to continue to learn throughout your career
- The publications of others are key to learning
- How we manage information is critical to success in the information age we live in today


**Revised Quality Assurance Standard  
Requirement for Literature Review**  
Quality Assurance Standards for Forensic DNA Testing Laboratories  
(effective July 1, 2009)

**5.1.3.2.** The laboratory shall have a program approved by the technical leader for the **annual review of scientific literature** that documents **the analysts' ongoing reading of scientific literature. The laboratory shall maintain or have physical or electronic access to a collection of current books, reviewed journals, or other literature applicable to DNA analysis.**

[http://www.fbi.gov/hq/lab/fsc/backissu/oct2008/standards/2008\\_10\\_standards01b.htm](http://www.fbi.gov/hq/lab/fsc/backissu/oct2008/standards/2008_10_standards01b.htm)

I am trying to get you “hooked” on literature

- **I want to teach you to fish** rather than just giving you some fish...



Give a man a fish and you feed him for a day. Teach a man to fish and you feed him for a lifetime.  
Chinese Proverb

“Give a man a fish; you have fed him for today. Teach a man to use the Net and he won't bother you for weeks.”  
—Author unknown

### Benefits of Literature Scans

- Become familiar with authors and institutions
- Will improve you as a writer and a presenter
- Will improve your lab's performance
- Over time you will be building your knowledge
- Remember: **You don't have to master every paper...**

How many have read any scientific article in the past month?

### The Value of a Journal Club

- Some potential approaches
  - Have specific people looking at individual journals
  - Bring relevant articles to attention of everyone
- ***J Forensic Sci* and *FSI Genetics* will cover ~90% of relevant articles in forensic DNA**
  - Scan journal, distill information, distribute to group
- rQAS requires literature to be available

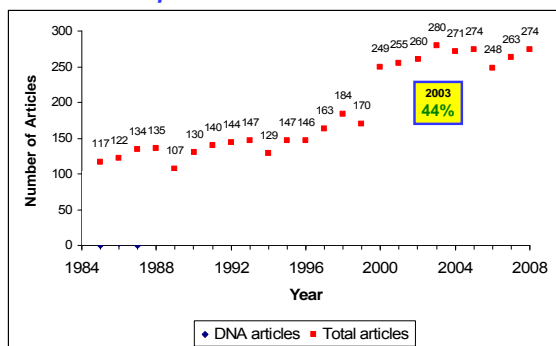
### How not to do it based on my experience

- Passing around individual journals with a reader list attached
  - Very inefficient process because journals get stuck on someone's desk
  - It becomes challenging to find a specific issue before it is returned to a central repository
  - Some information may not be as relevant (for research) many months later

### Forensic Science Publications



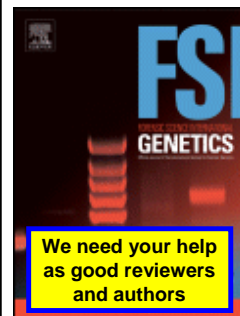
### Journal of Forensic Sciences DNA publications vs total articles



From PubMed searches performed Feb 2009 using <http://www.ncbi.nlm.nih.gov/sites/entrez>


### Forensic Science International: Genetics

<http://www.fsigenetics.com/>



**Editor-in-Chief:**  
 Angel Carracedo (Spain)  
**Associate Editors:**  
 Peter M. Schneider (Germany)  
 John M. Butler (USA)

**FSI: Genetics is a new journal dedicated exclusively to the field of forensic genetics.** It has been launched in 2007 by Elsevier Publishers in affiliation with the International Society of Forensic Genetics. **All members of the ISFG receive a free subscription of this journal** (print and online version) as part of their membership benefits.



### A Brief Review of Contents (March 2007-Sept 2009)

- STR population data (40)
- Y-STRs (32)
- Mitochondrial DNA (28)
- X-STRs (13)
- Non-human DNA (11)
- SNPs (10)
- Mixtures (8)
- Low-level DNA (8)
- Degraded DNA/miniSTRs (6)
- Paternity testing (6)
- Rapid screening/portable devices (4)
- Phenotype information (4)
- DNA extraction (4)
- DNA quantitation (3)
- Disaster victim identification (3)
- DNA databases (3)
- Variant & tri-alleles (3)
- Mutation rates (4)
- Forensic pathology (3)
- Statistical issues (2)
- Expert systems (2)
- Review articles (2)
- Other articles not classified (14)

**213 articles published in 11 issues**

### AAFS 2009 Topics Regarding Forensic DNA

*From abstracts of presentations at AAFS meeting in Denver, CO (Feb 2009)*

- Improved DNA extraction
- Predicting hair color and ancestry with SNPs
- X-chromosome STRs
- **Familial searching**
- **Low level DNA samples**
- miniSTRs
- DNA screening assays
- Optimizing database labs
- Microfluidic biochip systems
- Use with property crimes
- Recovery from handguns
- DNA from IEDs
- Expert systems
- Automation with robotics
- DNA quantitation – qPCR
- PCR directly from blood
- mtDNA
- RNA
- Non-human DNA (dogs & cows)
- **Mixture interpretation**

### Approaches to Maintaining an Awareness of the Literature

- Foster environment where any employee can bring helpful information to their supervisor and team members
- Prepare reference lists on topics of interest to your lab (or team)
  - Examples:
    - **Listing of all articles on mixture interpretation**
    - Gathering 70 articles on low-copy number DNA (pdf files and reference list)

### How to Read a Scientific Article

- Skim the article first
  - Start with title and abstract (may consider authors as well)
  - Scan tables, figures and figure captions
- Examine results and conclusions
  - Do the data presented support the statements made?
- Do not worry about trying to comprehend the entire article at first
  - I very rarely read an article from start to finish in its entirety
- Highlight key points and make notes on the paper itself so you can go back to them later to refresh your memory

### Approaches for On-Going Information Searches on Topics of Interest

- Review entire journal listing of articles
  - Pick up journal or view table of contents on-line
- Directed searches on specific topics
  - PubMed
- Sign up for table of contents delivery via email
- **Examine publications cited in review article**

### Forensic Science Review Article

*June 15, 2009 issue of Analytical Chemistry*

Anal. Chem. 2009, 81, 4695-4711

### Forensic Science

**T. A. Brettell\***  
Department of Chemical and Physical Sciences, Cedar Crest College, 100 College Drive, Allentown, Pennsylvania 18104-6196

**J. M. Butler**  
Biochemical Science Division, National Institute of Standards and Technology, Gaithersburg, Maryland 20899-8312

**J. R. Almirall**  
Department of Chemistry and Biochemistry and International Forensic Research Institute, Florida International University, University Park, Miami, Florida 33199

Review Contents

Forensic DNA Analysis  
Collection, Characterization, Preservation, Extraction, and Quantitation of Biological Material

Short Tandem Repeats  
Single Nucleotide Polymorphisms

Y-Chromosome and X-Chromosome Analysis  
Mitochondrial DNA Typing

Nonhuman DNA Typing Systems  
DNA Databases, Missing Persons, and Disaster Victim Identification

Interpretation and Statistical Weight of DNA Typing Results  
General Reviews

**2009 review article covers 160 DNA articles published in 2007-2008**

### PubMed Searches

The screenshot shows a search for 'Forensic DNA typing and STRs' on the PubMed website. The results list several articles, including one by Takayama T, Takada R, Suzuki R, Nagata S, Watanabe Y, Kumaga R, Adachi Y, Butler JM, and Log H, published in 2008. The URL <http://www.ncbi.nlm.nih.gov/PubMed> is displayed at the bottom.

### The Public Library of Science (PLoS) offers free on-line access to scientific articles

The screenshot shows the PLoS ONE article page for 'An Ultra-High Discrimination Y Chromosome Short Tandem Repeat Multiplex DNA Typing System' by Eric K. Hanson and Jack Ballantyne. The URL <http://www.plos.org/> is displayed at the bottom.

The screenshot shows Google Scholar search results for 'Forensic DNA typing'. It lists several articles, including 'Population genetics in forensic DNA typing' and 'Forensic DNA typing: A review'. The URL <http://scholar.google.com> is displayed at the bottom.

### Reference Manager Software

The advertisement for Reference Manager software features a central image of the software interface. A yellow box at the bottom contains the text: 'Demonstrate Reference Manager searches and generation of bibliography'. The URL <http://www.refman.com/> is displayed at the bottom.

### Literature Management

- Used to spend 2+ hours per week in the library
- Now can access articles via NIST Virtual Library from my desk
- Consolidated Reference Manager database

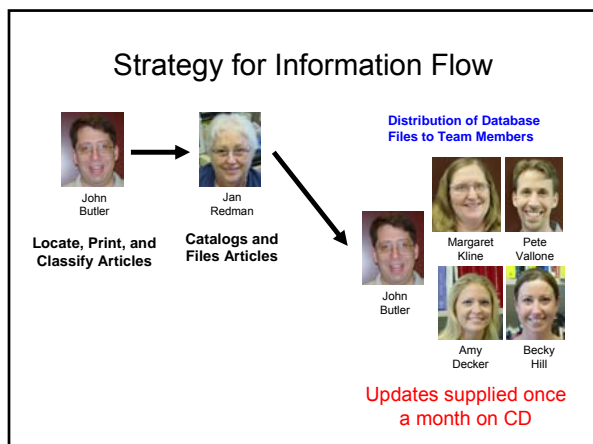
### Our Project Team Library

**>8,000 references gathered and cataloged in Reference Manager**

- Started by Christian Ruitberg
- Maintained now by **Jan Redman**
- **Updated Monthly** and provided to NIST Human Identity Project Team

**NIST Human Identity Project Team**

**Comprehensive set of forensic DNA articles**



### Reference Manager Database

As of Jan 2010: 4887 references in AllRef and 3369 references in STR\_Ref

Adding almost 1000 new articles each year

Ref ID	Authors	Title
2153	Butler, J.M.	Forensic DNA typing by capillary electrophoresis using the ABI Prism 310 and 3100 genetic analyzers for STR analysis
2201	Butler, J.M.	Duplication of DYS19 flanking regions in other parts of the Y chromosome
2461	Butler, J.M.	Locus-specific brackets for reliable typing of Y-chromosome short tandem repeat markers
2477	Butler, J.M.	Chromosomal duplications along the Y-chromosome and their potential impact on Y-STR interpretation
2492	Butler, J.M.	U.S. population data for the multi-copy Y-STR locus DYS464
2500	Butler, J.M.	Allele frequencies for 27 Y-STR loci with U.S. Caucasian, African American, and Hispanic samples
3015	Butler, J.M.	Genetics and Genomics of Core Short Tandem Repeat Loci Used in Human Identity Testing
3015	Butler, J.M.	Short tandem repeat typing technologies used in human identity testing
3035	Butler, J.M.	STRs vs. SNPs: thoughts on the future of forensic DNA testing

### Fruits of a Good Literature Collection

#### Review Articles

*John M. Butler, Ph.D.*

Genetics and Genomics of Core Short Tandem Repeat Loci Used in Human Identity Testing

*Analytical Chemistry (June 15, 2007 issue)*

#### Textbooks

**2nd Edition 688 pp.**  
**Feb 2005**

#### Textbooks

**Forensic Science**

**T. A. Brentell\***  
Department of Chemical and Physical Sciences, Cedar Crest College, 100 College Drive, Allentown, Pennsylvania 18104-6192

**J. M. Butler**  
Biological Science Division, National Institute of Standards and Technology, Gaithersburg, Maryland 20899-6311

**J. R. Albinell**  
Department of Chemistry and Biochemistry and International Forensic Research Institute, Florida International University, University Park, Miami, Florida 33199

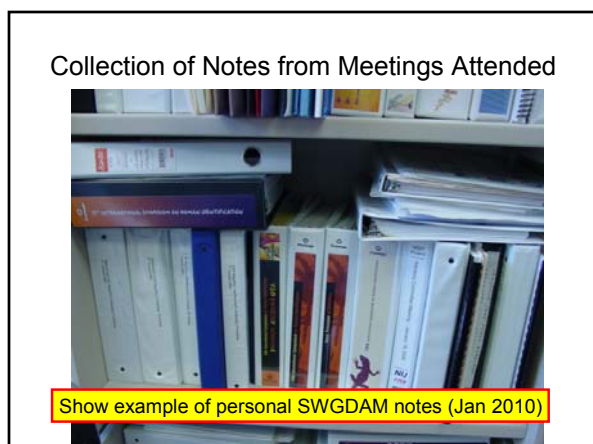
### And a Useful Reference Website...

**Short Tandem Repeat DNA Internet Database**

NIST Standard Reference Database SRD 130 [Recent Updates]

Reference Listing regarding STRs and DNA Typing

Complete Set of References (Numerical Order) 3816 references as of 1/12/07



- ### Why you need to write up your work
- Peer-review usually generates quality information
  - Talks are not held to the same standard as a written publication (that has been reviewed)
  - A written publication is also accessible to those who did not attend a presentation and is archived for future scientists to read


### The Peer-Review Process Based on My Perspective as an Editor

- Authors write article according to journal guidelines (each journal has an "Instructions for Authors")
- Steps during review
  - Article submitted to journal by corresponding author
  - Assigned to an editor
  - Editor asks 2 or more scientists to review the article in a specific timeframe (usually 2-3 weeks)
  - Editor takes reviews into consideration and responds to author with **Accept, Revise, or Reject**; "Revise" is most common
  - Author revises article and resubmits it for another review

Unfortunately, busy scientists often do not complete their review in a timely fashion (requiring the editor to remind them)

### How to Write a Scientific Article

- **Outline the ideas first** with a purpose and plan
  - Decide on scope, design experiments, & collect data
- Write Materials and Methods section first
- Prepare all figures & tables
  - captions should be stand-alone
- Write Results and Discussion based on data shown in figures & tables
- Write Introduction to provide context to your work
- Prepare reference list according to journal format
- **Write abstract last**
  - Most critical piece since it will be the most read!



George Whitesides  
on how to write a scientific article

*Adv. Mater.* (2004) 16(15): 1375-1377

**Whitesides' Group: Writing a Paper**<sup>®</sup>\*

By George M. Whitesides<sup>\*</sup>

**1. What is a Scientific Paper?**

A paper is an organized description of hypotheses, data and conclusions, intended to instruct the reader. Papers are a central part of research. If your research does not generate papers, it might just as well not have been done. "Interesting and unpublished" is equivalent to "non-existent".

Realize that your objective in research is to formulate and test hypotheses, to draw conclusions from these tests, and to teach these conclusions to others. Your objective is not to "collect data".

A paper is not just an archival device for storing a completed research program; it is also a structure for planning

do not agree on the outline, any text is useless. Much of the time in writing a paper goes into the text; most of the thought goes into the organization of the data and into the analysis. It can be relatively efficient in time to go through several (even many) cycles of an outline before beginning to write text; writing many versions of the full text of a paper is slow.

All writing that I do—papers, reports, proposals (and, of course, slides for seminars)—I do from outlines. I urge you to learn how to use them as well.

**2.2. How Should You Construct an Outline?**

The paper is now a good plan for the research program. You should write and rewrite these plans/outlines throughout the

process. Why not? In this work - ... What does it mean? "What hypotheses did I mean to test?"; "What ones did I

**author of more than 950 scientific articles and 50 patents**

### "Source Attribution" and Literature Categories

- **Always cite your sources**
  - Important to know where something came from because you might need to go back to it
  - Not all information is of equal value or importance

**Literature can be subdivided into several categories:**

- **Peer-reviewed literature** (containing data)
- Reports (evaluating a methodology)
- Review articles (commenting on other's data)
- Non-peer reviewed literature (representing the authors' opinions only) — e.g., conference proceedings

### Additional Thoughts

- Make time each week to continue your education
  - read an article once a week during lunch
  - read during your commute (unless you are driving!)
- Take detailed notes with each meeting you attend and then **share what you learned with others** (this will help you understand the information better)




### APPLIED GENETICS Group Major Programs Currently Underway

- **Forensic DNA**
  - New loci and assays (26plex)
  - **STR kit testing**
  - Ancestry SNP assays
  - **Low-template DNA studies**
  - Mixture interpretation
  - STR nomenclature
  - **Variant allele cataloging** and sequencing
  - Expert systems review
  - Training workshops to forensic DNA laboratories
  - Validation information and **software tools**
  - **Textbook** – 3<sup>rd</sup> ed. (2 vol.)
- **Clinical Genetics**
  - CMV SRM
  - Huntington's SRM
- **Ag Biotech**
  - "universal" GMO detection/quantitation (35S promoter)
- **DNA Biometrics**
  - **Rapid PCR** methods
  - Efforts to standardize testing of future portable DNA systems
  - Kinship analysis
- **Cell Line Authentication**



**The NIST Human Identity Project Team**  
 (Forensic DNA & DNA Biometrics)

Funding from the National Institute of Justice (NIJ) through the NIST Office of Law Enforcement Standards and the FBI S&T Branch through the NIST Information Access Division  
 ...Bringing traceability and technology to the scales of justice...



**The Topics & Tasks**

**Technology**

- Research programs in STRs, SNPs, miniSTRs, Y-STRs, mtDNA, qPCR, LCN, mixtures, rapid PCR
- Assay and software development, expert system and kinship software review

**Standards**

- Standard Reference Materials (SRMs 2391b, 2392, 2395, 2372)
- Standard Information Resources (STRBase website = SRD 130)
- Interlaboratory Studies (DNA quantitation, mixture interpretation)

**Training Materials & Workshops**

- Textbooks on Forensic DNA Typing and review articles written
- PowerPoint and pdf files made available for download
- Training workshops conducted to scientists, lawyers, and students on validation and other topics

**The Triumphs**

**Achievements since 2000:**

- >110 publications
- >300 presentations
- >40 workshops
- 3 textbooks

**National Institute of Justice** <http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

Rapid PCR work published in *FSI Genetics* (Dec 2008)

**Full STR profiles in 36 minutes (instead of 3 hour PCR)**

Contents lists available at ScienceDirect  
 Forensic Science International: Genetics  
 journal homepage: www.elsevier.com/locate/fgi

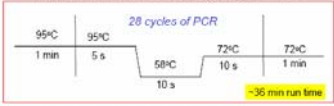
Short communication  
 Demonstration of rapid multiplex PCR amplification involving 16 genetic loci<sup>®</sup>  
 Peter M. Vallone<sup>\*</sup>, Carolyn R. Hill, John M. Butler

National Institute of Standards and Technology, Biomedical Science Division, 100 Bureau Drive, Mail Stop 8371, Gaithersburg, MD 20899-8371, United States

**Complete concordance of STR allele calls (for 60 samples) between the rapid and standard thermal cycling protocols** were observed although there was incomplete adenylation at several of the loci examined and some PCR artifacts were detected. Using less than **750 pg of template DNA and 28 cycles, STR peaks for all loci were above a 150 relative fluorescent unit (RFU) detection threshold** with fully adequate inter-locus balance and heterozygote peak height ratios of greater than 0.84.

**Further Rapid PCR Work**

Rapid PCR Thermal Cycling Profile



- Much shorter hold times at each temperature
- Faster ramp rates between temperatures

Maximum heating rate of ~4°C/s on a GeneAmp 9700 (Applied Biosystems)

- **Examination of different enzyme mixes**
  - 0.5 x master mix PyroStart (Fermentas) (\$0.14/rxn)
  - 0.5 x master mix Premix Ex Taq (Takara) (\$0.22/rxn)
  - 0.25 µL = 1.25 units of SpeedStar (Takara) (\$1.09/rxn)
- **Evaluation of additional kits**
  - Identifiler, PP16, Yfiler, MiniFiler and Promega S5
- **Testing thermal cyclers with faster ramp rates**

**Four Thermal Cyclers Being Evaluated**

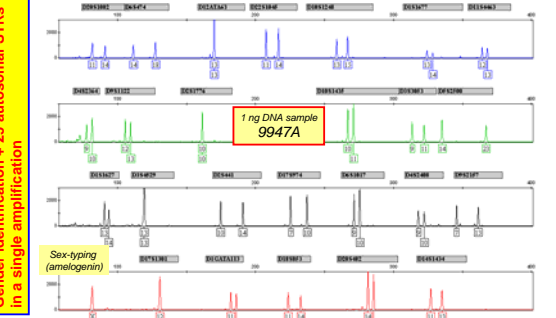
**How fast can we run 28 cycles?**

<b>GeneAmp 9700 (Applied Biosystems)</b> • Heating rate: 4°C/s • Heating mechanism: Peltier block (A0) • Tube format: 0.2 mL – 96 well plate • 28 cycles = 36 min (by the general rule) (Normal cycling cycle shown on the left) <b>36 minutes</b>	<b>SmartCycler (Cepheid)</b> • Heating rate: 10°C/s • Heating mechanism: heating plates and air circulating fan • Tube format: proprietary 25 µL tubes • 16 reactions per instrument, using to run 16 independent thermal cycling profiles • Can also be used for real time PCR • 28 cycles = 20 min <b>20 minutes</b>
<b>Mastercycler pro (Eppendorf)</b> • Heating rate: 6°C/s • Heating mechanism: Peltier block (A0) • Tube format: 0.2 mL – 96 well plate • 28 cycles = 19 min <b>19 minutes</b>	<b>Rotor-Gene Q (Qiagen)</b> • Heating rate: 15°C/s • Heating mechanism: Air chamber (spinning rotor) • Tube format: 0.1 mL – 72 tube/rotor • 28 cycles = 36 min <b>36 minutes</b>

[http://www.cstl.nist.gov/biotech/strbase/pub\\_pres/VallonePromega2009poster.pdf](http://www.cstl.nist.gov/biotech/strbase/pub_pres/VallonePromega2009poster.pdf)

NIST 26plex published in *J. Forensic Sci.* (Sept 2009)

**25 new STR loci (unlinked from 13 CODIS core loci)**

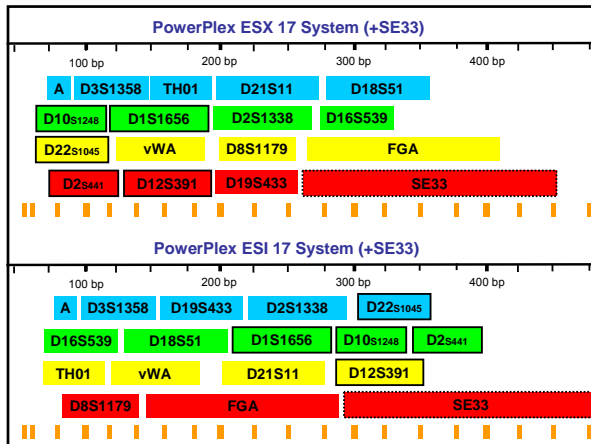


Gender identification + 25 autosomal STRs in a single amplification

Hill, C.R., et al. (2009) A new STR 26plex assay for human identity testing. *J. Forensic Sci.* 54: 1008-1015  
 Primer sequences and GM bins & panels available: <http://www.cstl.nist.gov/biotech/strbase/str26plex.htm>

**NIST Studies with New STR Kits**

- Over the past year, our NIST team has worked with Promega to evaluate their new STR kits
  - PowerPlex ESX 17 & ESI 17 - with new European loci
  - PowerPlex 16 HS – with new enzyme & buffer system
- **Concordance studies**, population data analysis, and additional validation information
- Plan to do similar studies with Applied Biosystems on their NGM and other new kits



### New NIST Software Tools

Developed by Dave Duewer (NIST)

**From NIST STRBase Website:**

- Lab Resources and Tools
  - Addresses for scientists working with STRs
  - Training Materials
  - STR Allele Sequencing
  - Population data
  - Data from NIST's S. Proulx's seminar
  - NIST-Developed Software: Index, AutoDimer, minSTR, and Multiplex\_QA**
  - NIST Standard Reference Material for PCR-Based Testing
  - New STR Markers under Development at NIST
  - Chromosomal Locations
  - DNA Advisory Board Quality Assurance Standards
  - Interlaboratory Studies
  - NIST Meeting 2005 Interlab Study: MINIST Data
  - Validation Information
  - DNA Quantitation - SRM 2172 (available as of October 5, 2007)
  - Technology for resolving STR alleles

**STR\_MatchSamples**

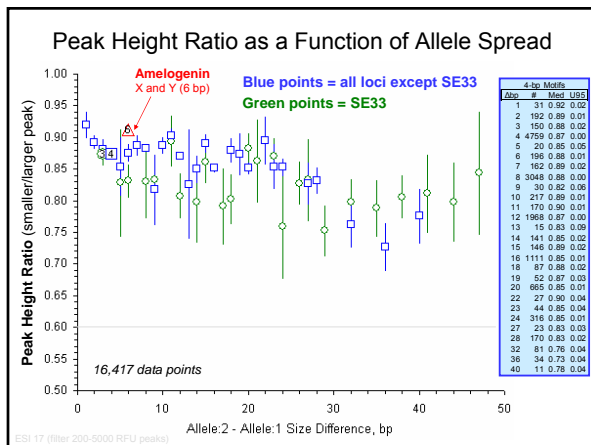
- An Excel-based tool developed to aid comparison of STR genotypes from two or more data sets.

**Tools under development (to aid validation studies)**

- Peak height ratio
- Inter-locus balance
- Stutter percentages
- Allele frequency

[http://www.cstl.nist.gov/biotech/strbase/tools/STR\\_MatchSamples.xls](http://www.cstl.nist.gov/biotech/strbase/tools/STR_MatchSamples.xls)

<http://www.cstl.nist.gov/biotech/strbase/software.htm>



### Trinucleotide D22S1045

#### Allele-Specific Stutter Percentages

ESX 17					ESI 17				
Allele	Size	#	Median	Stutter	Allele	Size	#	Median	Stutter
10	84.5	21	1.8		10	308.7	22	1.9	
11	87.4	134	3.0		11	311.8	98	2.8	
12	90.4	37	4.2		12	314.8	32	4.5	
14	96.4	51	7.2		14	321.0	36	6.1	
15	99.4	165	8.9		15	324.0	150	9.9	
16	102.4	120	10.5		16	327.1	94	9.8	
17	105.5	105	14.7		17	330.1	95	14.2	
Avg		633	7.2		Avg		527	7.0	
SD			4.6		SD			4.4	

633 data points      Avg + 3SD 21.0%

527 data points      Avg + 3SD 20.2%

### 532 Variant Alleles on STRBase

[http://www.cstl.nist.gov/biotech/strbase/var\\_tab.htm](http://www.cstl.nist.gov/biotech/strbase/var_tab.htm)

532 total variants reported as of 01/25/2010

**New Addition**

Core STR Loci (364)	Other Common STR Loci (122)	Y-STR Loci (46)
<ul style="list-style-type: none"> <li>CSF1PO (20)</li> <li>FGA (103)</li> <li>TH01 (19)</li> <li>TP02 (17)</li> <li>VWA (13)</li> <li>D3S1358 (21)</li> <li>D5S818 (16)</li> <li>D7S820 (26)</li> <li>D8S1179 (21)</li> <li>D13S317 (17)</li> <li>D16S539 (21)</li> <li>D18S51 (41)</li> <li>D21S11 (32)</li> </ul>	<ul style="list-style-type: none"> <li>D2S1338 (23)</li> <li>D19S433 (27)</li> <li>Penh4_D (37)</li> <li>Penh4_F (30)</li> <li>FL301 (1)</li> <li>FES.FPS (1)</li> <li>FL3B</li> <li>LPL</li> <li>SE33 (1)</li> <li>D13S1677 (1)</li> <li>D14S1434 (1)</li> </ul>	<ul style="list-style-type: none"> <li>DYS19 (2)</li> <li>DYS389I (2)</li> <li>DYS389II (1)</li> <li>DYS390 (2)</li> <li>DYS391</li> <li>DYS392 (4)</li> <li>DYS393 (1)</li> <li>DYS385 a,b (16)</li> <li>DYS438 (3)</li> <li>DYS439 (4)</li> <li>DYS437 (3)</li> <li>DYS448</li> <li>DYS456</li> <li>DYS458 (4)</li> <li>DYS633 (3)</li> <li>Y-GATA-H4 (1)</li> </ul>

### Many New Y-STR Variants Being Reported by Missouri State Highway Patrol

#### DYS439 Variants

At time of submission had run over 41,000 samples!

Allele Designation	Allele Size (bp)	Instrument	Amp Kit	Contributor	Verification/Confirmation Method(s)	Notes	Frequency
7 (<8)	194.26	ABI 3130d	PPY	Jackie Johnson, MO State Hwy Patrol	redo sample		1 in 41635
10.1	207.20	ABI 3130d	PPY	Jackie Johnson, MO State Hwy Patrol	redo sample		2 in 41635
11.1	211.22	ABI 3130d	PPY	Jackie Johnson, MO State Hwy Patrol	redo sample		1 in 41635
16 (>15)	230.44	ABI 3130d	PPY	Jackie Johnson, MO State Hwy Patrol	redo sample		1 in 41635



### 194 Tri-Allelic Patterns on STRBase

[http://www.cstl.nist.gov/biotech/strbase/tri\\_tab.htm](http://www.cstl.nist.gov/biotech/strbase/tri_tab.htm)

*194 total patterns reported as of 01/04/2010*

(click on loci listed below for details)

Core STR Loci (171)	Other Common STR Loci (23)	Y-STR Loci (1) <i>duplications or triplications</i>
CSE1PO (7)	D2S1348 (3)	DYS19
FGA (26)	D19S433 (3)	DYS389I
TH01 (3)	Penta D (6)	DYS389II
TP05 (15)	Penta E (11)	DYS390
VWA (20)	FlA01	DYS391
D18S1358 (7)	FES, FES (1)	DYS392
D5S818 (7)	FlB	DYS393
D7S820 (8)	LPL	DYS385 a/b
D8S1179 (11)	SE33	DYS438
D13S317 (9)		DYS439
D16S539 (8)		DYS437
D18S51 (26)		DYS448
D21S11 (18)		DYS456 (1)
		DYS458
		DYS613/GATA-C4
		Y-GATA-H4

### New STRBase Website on LT-DNA (LCN)

<http://www.cstl.nist.gov/biotech/strbase/LTDNA.htm>

Information on Low Template / Low Copy Number DNA Testing

**General Information**

- o Purpose of STRBase
- o Publications and Presentations
- o NII Funded Projects
- o Training Materials
- o Links to other web sites
- o Glossary of common terms

**Forensic STR Information**

- o STRs101: Brief Intro
- o Core Loci FBI CODIS
- o STR Fact Sheets: Loci
- o Multiplex STR kits
- o Sequence Information
- o Variant Allele Reports
- o Tri-Allelic Patterns
- o Mutation Rates for Core Loci
- o Published PCR primers
- o Y-chromosome STRs

**Presentations on LTDNA**

- o John Butler - NIST (Prommer)
- o Beckie Hill - NIST (Prommer)
- o Theresa Caspary - NIST (PhD)

**Low Copy Number (LCN) DNA Panel Discussion**

#### Scientific Issues with Analysis of Low Amounts of DNA

John Butler, NIST (Prommer)  
Beckie Hill, NIST (Prommer)  
Theresa Caspary, NIST (PhD)

**Low Copy Number (LCN) DNA Panel Discussion**

Theresa Caspary, Ph.D.  
Deputy Director  
October 15, 2009

### Complete Set of NIST Sensitivity Data Available on New LT-DNA Website

<http://www.cstl.nist.gov/biotech/strbase/LTDNA.htm>

NIST Sensitivity Data with low level DNA templates

10 replicate amplifications for each condition with two fully heterozygous, single-source samples

Click on links to see summaries and DNA profiles observed

STR kit - PCR conditions	Sample 1	Sample 2
Identifier - 28 cycles	100 pg	100 pg
	30 pg	30 pg
	10 pg	10 pg
Identifier - 31 cycles	100 pg	100 pg
	30 pg	30 pg
	10 pg	10 pg
PowerPlex 16 HS - 31 cycles	100 pg	100 pg
	30 pg	30 pg
	10 pg	10 pg
PowerPlex 16 HS - 34 cycles	100 pg	100 pg
	30 pg	30 pg
	10 pg	10 pg

PowerPlex 16 HS - 34 cycles

Sample #1 (MT97150)

Sample #2 (PT6411)

MT97150 - 10 pg, amp #1

### Comparison of Approaches

**Replicate Amplification with Consensus Profile**

Low amount of DNA examined

Stochastic effects

Amplification #1  
Amplification #2  
Amplification #3

Consensus Profile Developed (from repeated alleles observed)

Interpretation Rules Applied (based on validation experience) e.g., specific loci may dropout more

Result can be and usually is **Reliable & Reproducible**

**Single Amplification**

Low amount of DNA examined

Stochastic effects

Amplification #1 (only a single test)

Result can be **Unreliable**

Individual results may vary but a **consensus profile is reproducible** (based on our experience with sensitivity studies and replicate amplifications)

What "LCN Labs" Are Doing

### NIST SRM 2391b will need to be replaced within the next year

SRM	Name	FY06	FY07	FY08	FY09	Avg	Remaining	Current \$*
2372	Human DNA Quantitation Std	0	0	160	147	153.5	1,078	\$372
2390	DNA Profiling	2	0	1	0	0.8	3	\$833
2391B	PCR-Based DNA Profiling	86	81	125	140	108	107	\$811
2392	Mitochondrial DNA Sequencing	8	6	0	12	6.8	165	\$883
2392i	Mitochondrial DNA Sequencing (Human HL-60 DNA)	6	32	20	19	19.3	176	\$365
2395	Human Y-Chromosome DNA Profiling	34	39	72	88	58.3	136	\$383

\*As of Oct 7, 2009

PCR-based DNA Profiling Standard

SRM 2391 (1995)	SRM 2391a (2009)	SRM 2391b (2003, r2008)	SRM 2391c (planned 2010)
3 liquid samples	3 liquid samples	2 swabs	1 paper punch

Considering preparing DNA in different forms:

Plan to certify both autosomal and Y-STRs on SRM 2391c components

Would like your input into the desired number of components & loci certified

\*coverage for all commercially available kit STR loci at the time of release

[http://www.cstl.nist.gov/biotech/strbase/pub\\_pres/Promega2009poster\\_SRM2391c.pdf](http://www.cstl.nist.gov/biotech/strbase/pub_pres/Promega2009poster_SRM2391c.pdf)

### SRM 2372: Human DNA Quantitation Standard

Released in Oct 2007

- >300 units in use as of October 2009
- Used by more than 110 forensic laboratories worldwide
- Manuscript describing production published in *Anal. Bioanal. Chem.*
- Serves to adjust qPCR calibrants supplied by manufacturers and adjust for assay-specific bias

PCR RT

Kline, et al. (2005) *J. Forensic Sci.* 50(3):571-578

Anal Bioanal Chem (2009) 394:1183–1192  
DOI 10.1007/s00216-009-2782-0

## SRM 2372 Certification

ORIGINAL PAPER

### Production and certification of NIST Standard Reference Material 2372 Human DNA Quantitation Standard

Margaret C. Kline · David L. Duesel · Melody V. Smith · Janette W. Redn · Peter M. Vallone · Amy E. Decker

Code	# Labs	# Data sets			Assay
		A	B	C	
Quantifiler	20	75	75	78	qPCR, gender-neutral
Sybr_ALU	5	11	11	13	qPCR, gender-neutral
CES	3	16	16	16	qPCR, gender-neutral
monoT101	1	4	4	4	qPCR, gender-neutral
Probz_ALU	1	4	4	4	qPCR, gender-neutral
T101	1	3	3	3	qPCR, gender-neutral
XNF	1	3	3	3	qPCR, gender-neutral
Probz_mit	1	2	2	2	qPCR, gender-neutral
Gender	1	4	0	4	qPCR, male specific
Probz_Y	1	2	1	2	qPCR, male specific
	3	11	11	0	Direct, gender-neutral
	2	7	7	0	Direct, gender-neutral
	1	4	4	4	Direct, gender-neutral

**Interlab Assessment Study with SRM 2372 (performed prior to Quantifiler Duo and Plexor HY availability)**

**Do NOT use Component C with Quantifiler Duo**

## The Expansion of Forensic DNA Typing

1<sup>st</sup> Edition

Jan 2001  
335 pp.  
17 chapters

2<sup>nd</sup> Edition

Feb 2005  
688 pp.  
24 chapters

Chinese Translation (2007) Y. Hou, translator  
Japanese Translation (2009) Y. Fukuma, translator

3<sup>rd</sup> Edition

Sept 2009  
18 chapters (504 pp.)  
25 chapters (~600 pp.)  
Planned for Spring 2011

## Improved Reference Format

**Forensic DNA Typing**  
(2<sup>nd</sup> Edition)

*Full list of authors but no article title*

**Fundamentals**  
(3<sup>rd</sup> Edition)

*Subdivided by subject with article title provided*

**>1500 references total (>600 new since 2<sup>nd</sup> edition)**

## Chapters Re-ordered to Reflect DNA Testing Process

**Steps Involved**

- Collection
- Sample Storage
- Extraction
- Quantitation
- Amplification
- STR Markers
- Separation/Detection
- Data Interpretation
- Statistical Interpretation

**Fundamentals of Forensic DNA Typing (2009)**

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## Written as Part of My Job at NIST (no royalties to be received)

### Fundamentals of Forensic DNA Typing

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