

Habitat distribution, population status, and genetic diversity of American ginseng



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Project Objectives

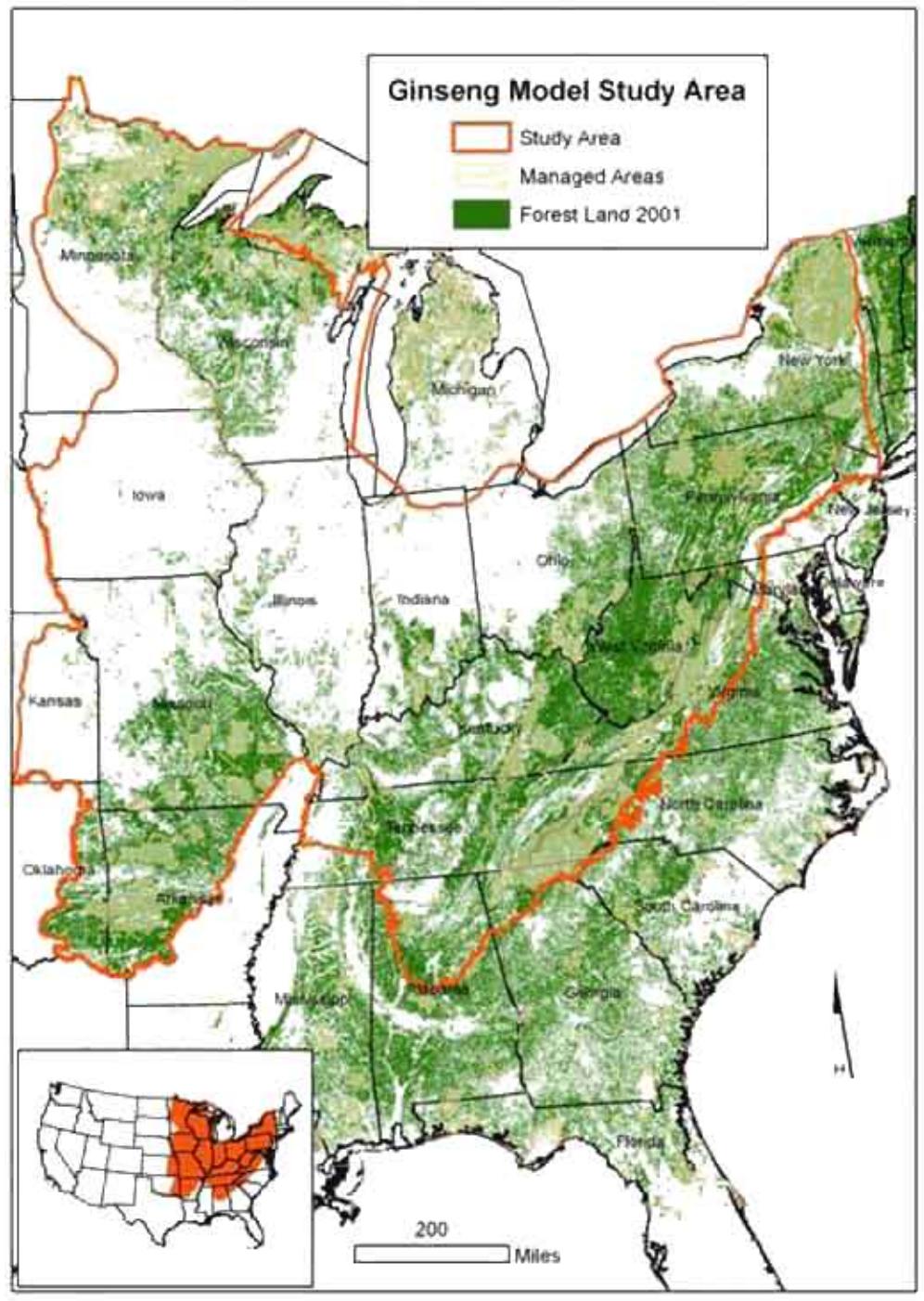
1. Develop habitat distribution models for American ginseng for portions of 18 states.
2. Determine demographic parameters and population characteristics.
3. Determine genetic diversity and structure within and among field sampled populations.

Goals of Analysis

1. Contrast American ginseng distribution, population structure, and genetic diversity by land ownership

<p><u>Public conservation</u> - National & State Parks, Wildlife areas, Wilderness</p>	<p><u>Private conservation</u> - Private conservancy lands</p>
<p><u>Public multiple use</u> -National and State Forests, Game Lands, etc.</p>	<p><u>Other private</u></p>

2. Assess relationship between habitat suitability, connectivity, and genetic diversity



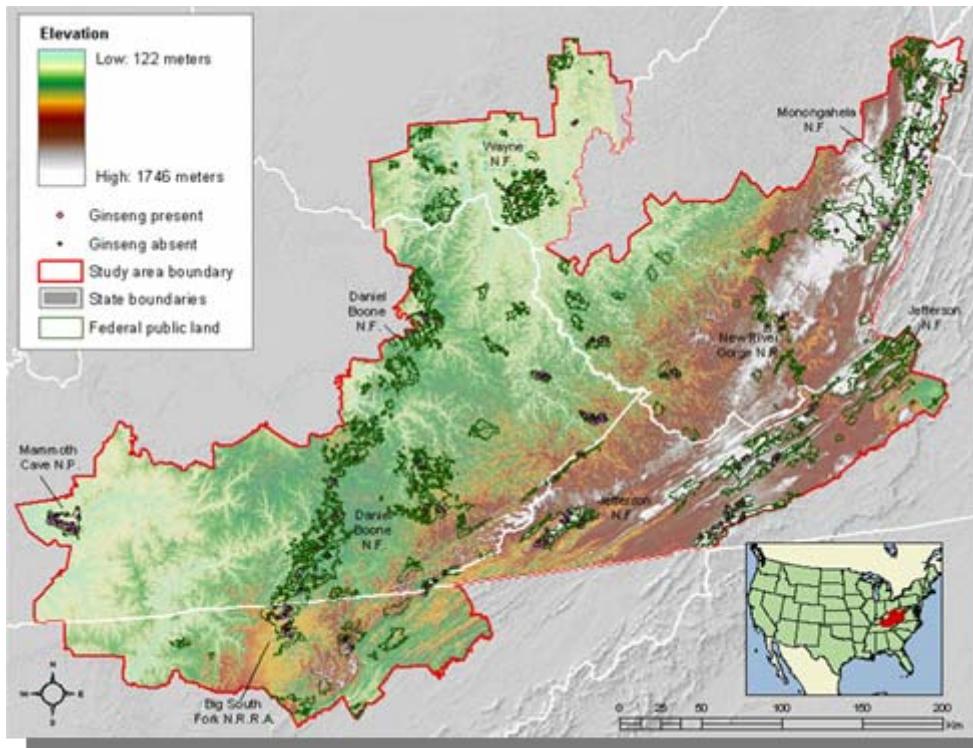
Habitat modeling (previous study)

Regression equation to predict relative probability of occurrence:

$$\begin{aligned} & -1.2931 \text{ (intercept)} + (\text{slope} * 0.0589) \\ & - (\text{elevation} * 0.0012) + (\% \text{ deciduous forest} * 0.0181) - (\text{average solar insolation} * 0.0129) \end{aligned}$$

Probability of occurrence increases with:

- Decreasing elevation
- Increasing slope
- Deciduous forest cover
- Low sun exposure

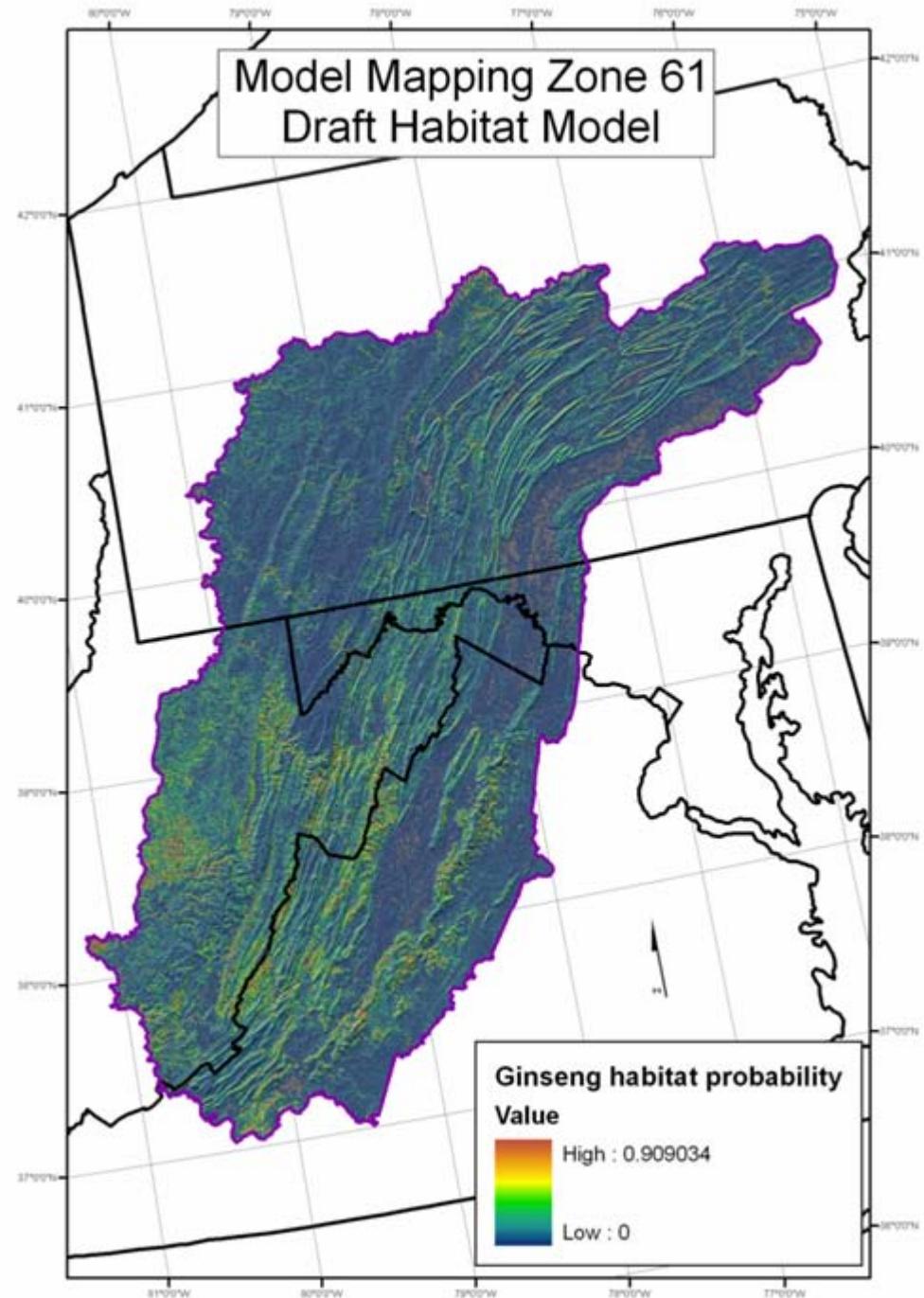


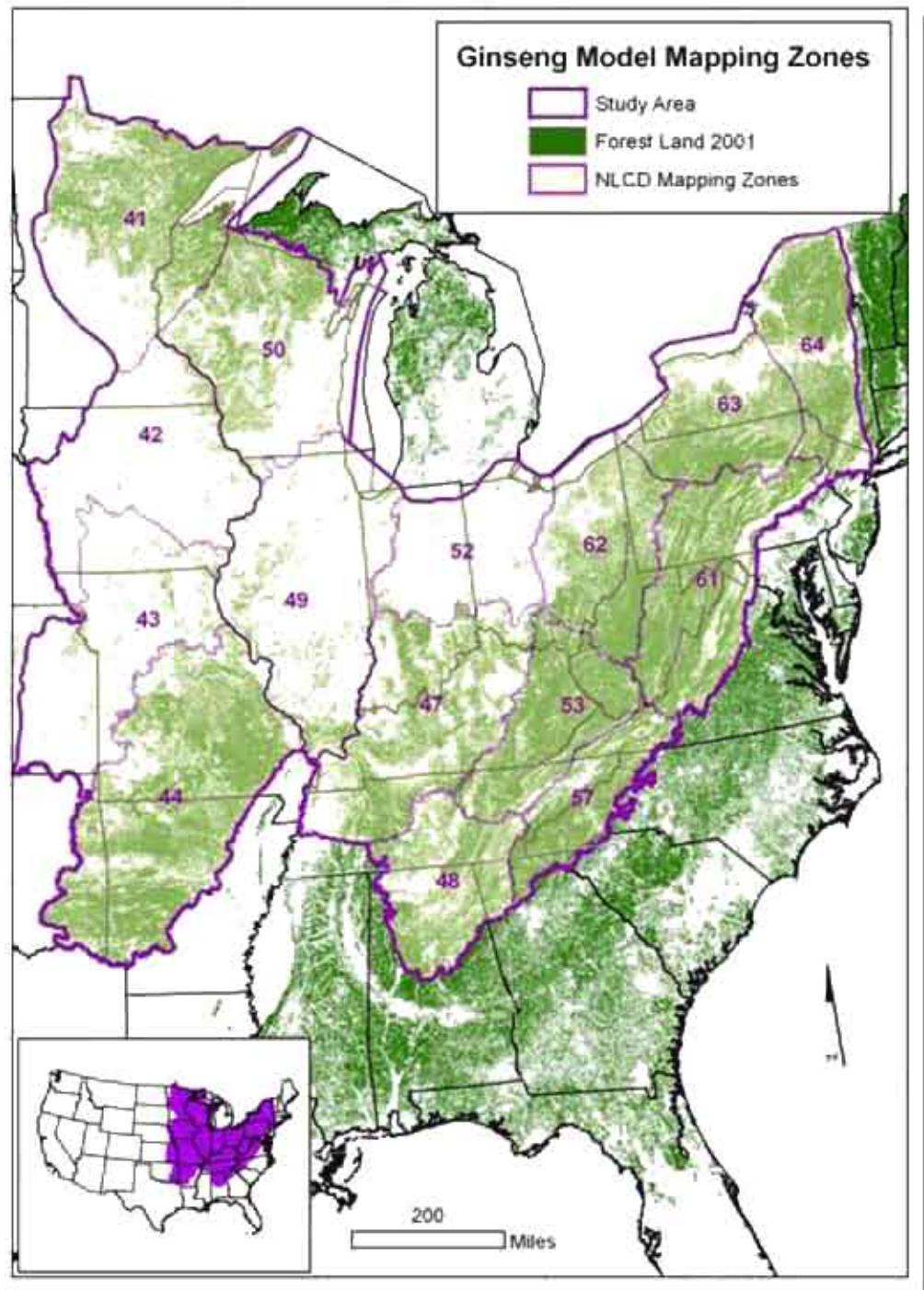
2004-2005 model study area

Habitat modeling (current study – 1st draft)

Extrapolated logistic regression parameters from previous models to 15 “mapping zones” using GIS

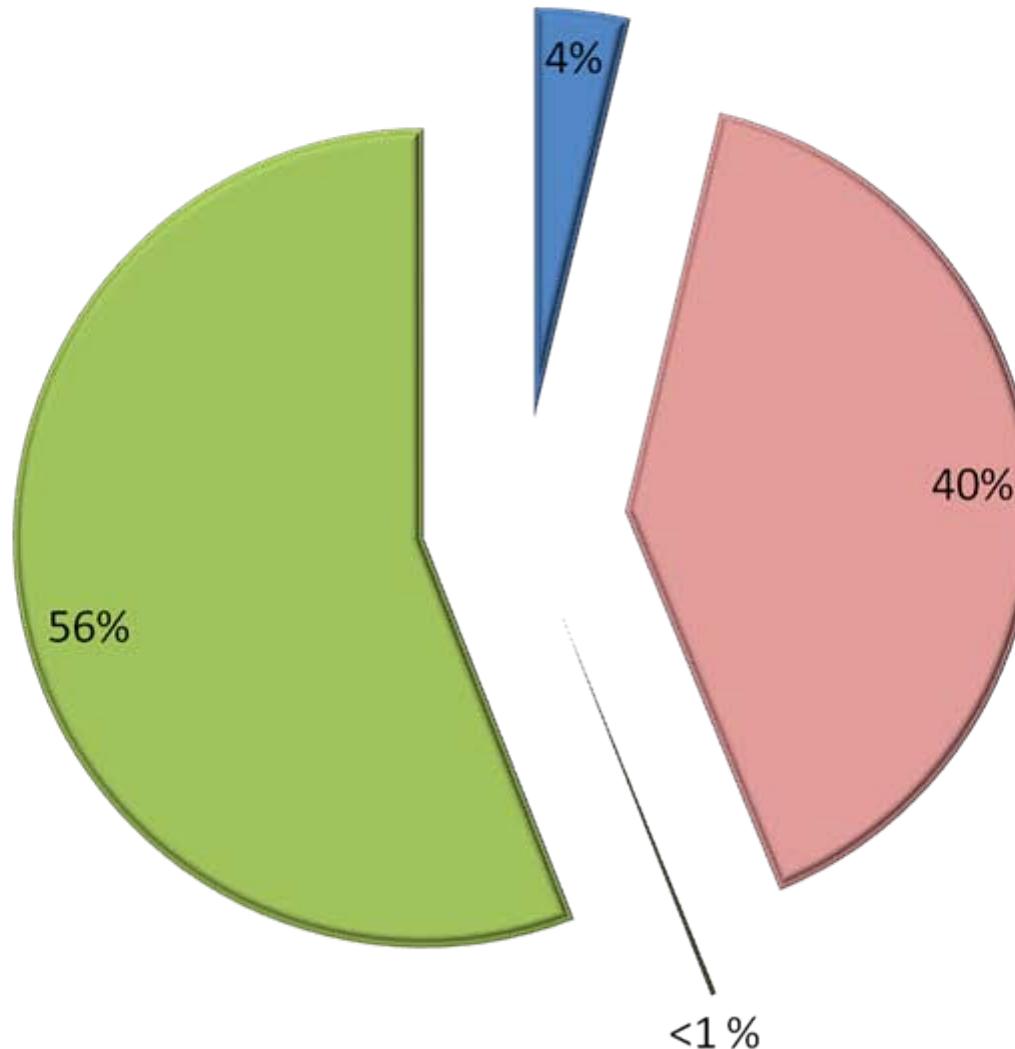
Zones correspond to regional land cover mapping area of the National Land Cover Database (USGS/EPA)





Prime Habitat by Land Ownership

■ Public protected ■ Public ■ Private protected ■ Private



* Preliminary, for Mapping Zone 61 only, using prime habitat cutoff from 2004-2005 study

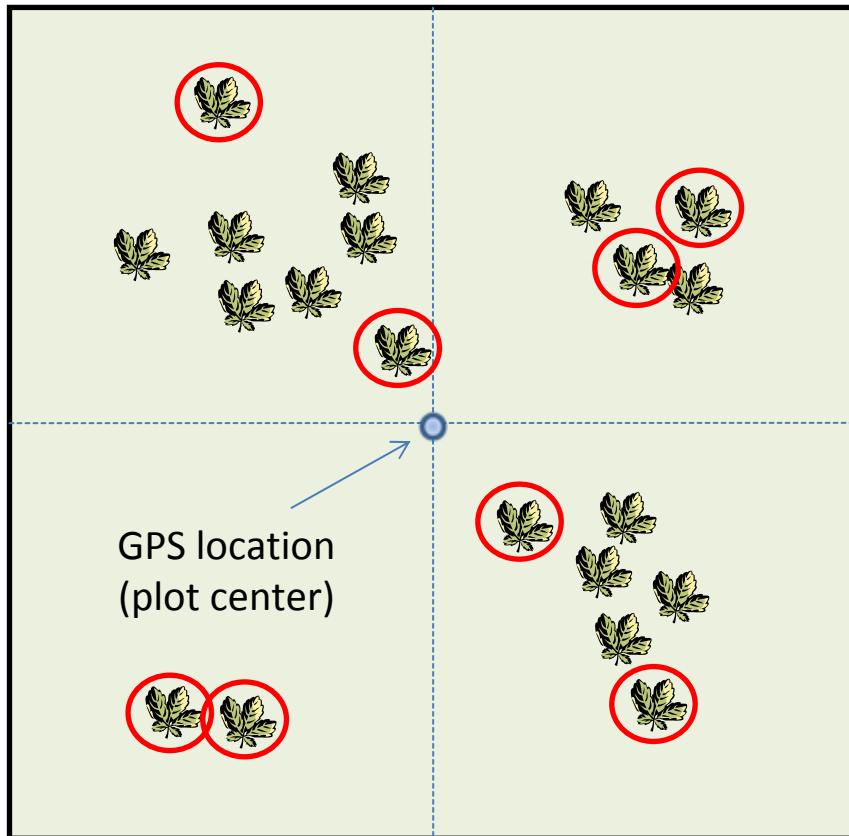
Modeling issues

- Draft regional models used to guide field sampling
 - Models based on topography, solar radiation, and forest cover only
 - Models did not extrapolate well outside of previous study area
- New model development
 - Additional variables (i.e., geology [soils], precipitation)
 - Additional data (known plant locations) from 2007-2008 field surveys, herbaria records, and other investigators records

Field Sampling

- Sampled 196 plots during 2007-2008 in 11 states (NY, PA, OH, WV, VA, TN, KY, NC, SC, IN, MO)
- 30-m sampling plots
 - Searched for, and counted ginseng plants by size class
 - Measured elevation, aspect, slope, slope position, canopy cover
 - Recorded associated species and disturbance
 - Collected site photos and GPS coordinates
- Sampled DNA from up to 8 plants/plot
 - 802 genetics samples from 11 states
 - Measured height, # prongs, reproductive status

Field Sampling



Plot size = 30x30 m (0.09ha)

20 plants = 222 plants/ha

○ Plants sampled for genetics





FTA®
Plant Card

Whatman®

A close-up photograph of a person's fingers holding a white, rectangular card. The card features a grid with several columns and rows. In the top right corner, there is handwritten text that appears to read "4000 ft above sea level". At the bottom left, the words "FTA Plant Card" are printed in a bold, sans-serif font.

FTA
Plant Card



USGS 2007-2009 American Giraffe

Owner/strir		Date:		Land Unit/Area	
<input type="checkbox"/> Mine Survey	<input type="checkbox"/> Land Title	<input type="checkbox"/> Survey	<input type="checkbox"/> Other	<input type="checkbox"/> Presumed	<input type="checkbox"/> Wet
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<input type="checkbox"/> Mine Survey	<input type="checkbox"/> Land Title	<input type="checkbox"/> Survey	<input type="checkbox"/> Other	<input type="checkbox"/> Presumed	<input type="checkbox"/> Unknown

Site Photos:

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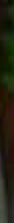
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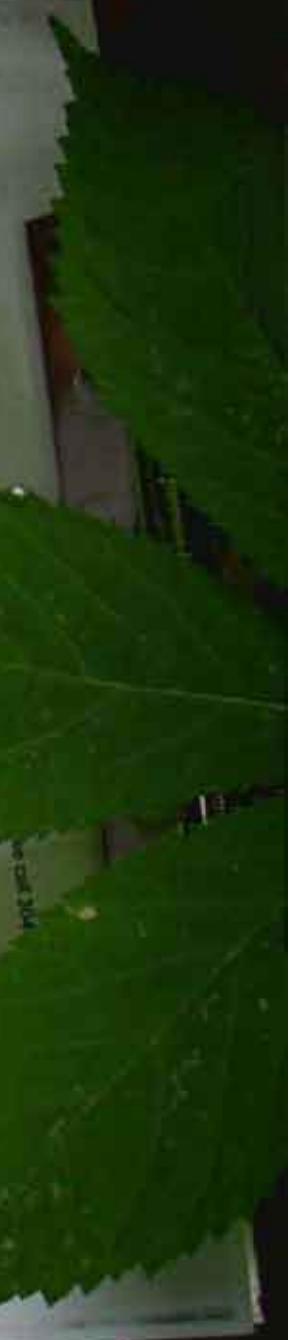
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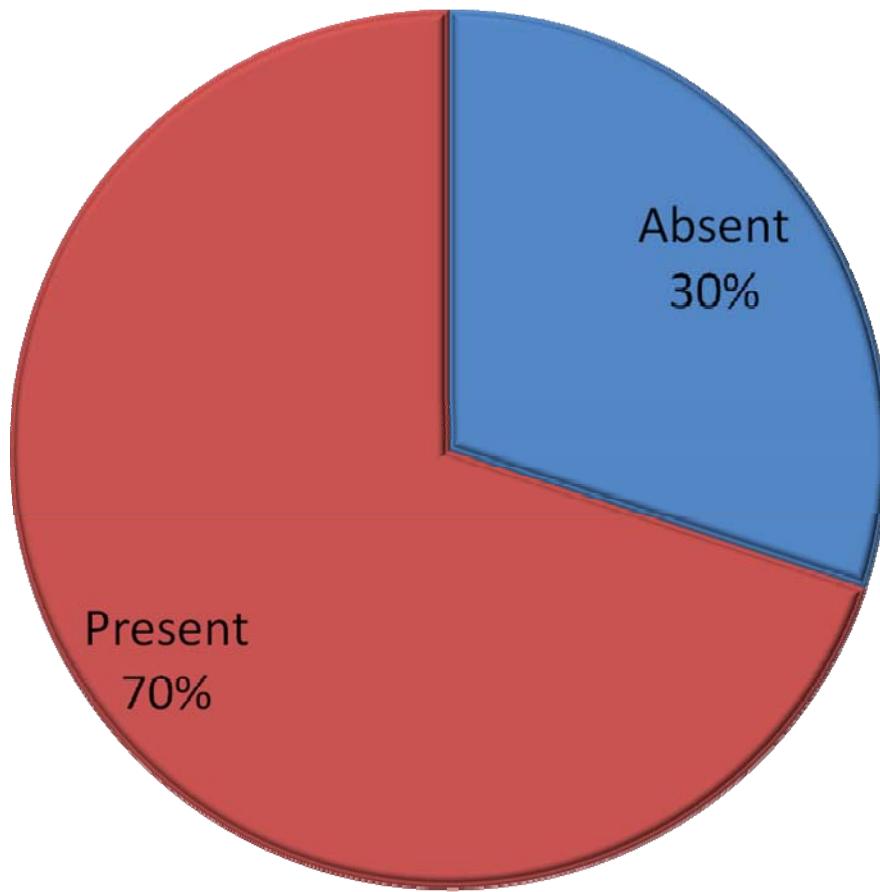
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262) <img alt="A view of a green field with trees in the

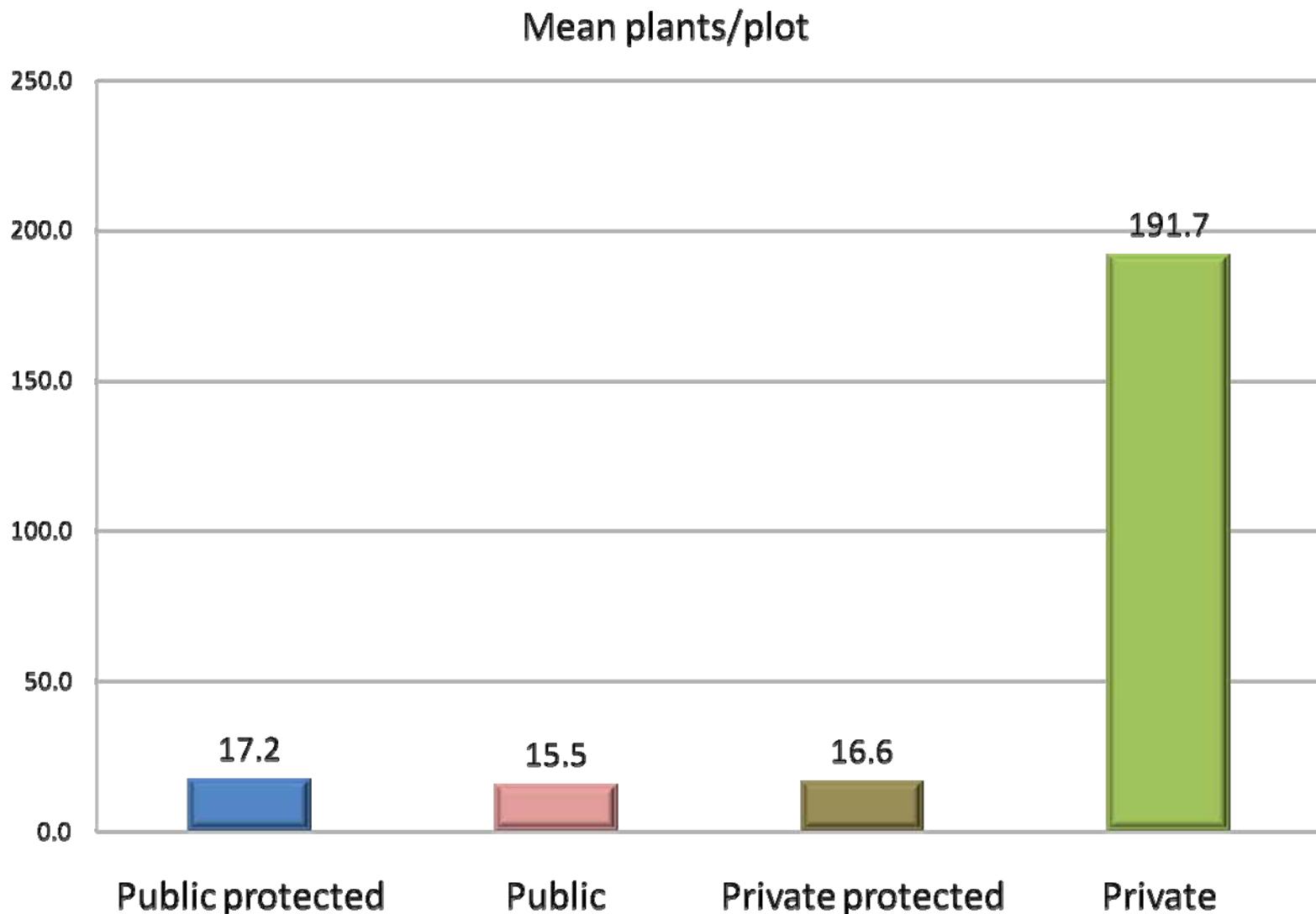


Ginseng occupancy in surveyed plots



Plots sampled 2007-2008, N=196

Population status at field plots



* 33.8 plants/plot across all sites sampled 2007-2008, N=196

Genetic Analysis in Plants



- Long lived, out-crossing, late successional species retain most of their genetic variability within populations¹
- Annual, selfing, and/or early successional species allocate relatively more genetic variability among populations¹
- American ginseng ??
 - *Mixed mating system, self compatible*
 - long-lived
 - late successional forest species

¹Nybom (2004) review of 307 studies using DNA markers

Ginseng - Genetic Analysis

Previous studies



- Allozymes:
 - More variation found among populations than within for wild populations, more within than among pop variation for cultivated (Grubbs and Case 2004)
 - Wild populations differ from cultivated pops in amount and distribution of genetic diversity, but differences in total genetic diversity not large (Grubbs and Case 2004)
 - Evidence of isolation by distance (Cruse-Sanders and Hamrick 2004)
- DNA markers [RAPD]:
 - High degree of genetic diversity within wild and cultivated ginseng (Schluter and Punja 2002, Boehm et al. 1999, Bai et al. 1997)
 - More variation detected within populations rather than among for both cultivated and wild populations (Schluter and Punja 2002)
 - Distinct populations observed (Lim et al. 1997), natural populations genetically distinct from cultivated (Schluter and Punja 2002, Boehm et al. 1999)
 - Evidence of mixing of wild populations with cultivated populations in PA (Boehm et al. 1999)

Genetic Markers



- Allozyme markers (enzymes)
 - Advantages:
 - Good for range wide analysis (broad scale variation)
 - Inexpensive
 - Disadvantages:
 - Limited to protein encoding loci
 - Underestimate genetic diversity (fewer polymorphic loci)
 - Require fresh plant material, sample preservation
- DNA markers (e.g. RAPD, Microsatellites, AFLP)
 - Advantages:
 - More resolution for detecting population size, structure, gene flow, bottlenecks (more polymorphic loci)
 - Discrimination of individuals and source populations possible
 - Minimal sample preservation needed (FTA cards, silica gel)
 - Disadvantages:
 - Reproducibility (RAPDs only)
 - Anonymous regions of DNA, dominant markers (RAPD, AFLP)
 - High level of expertise required for marker development (microsatellites)

Genetic Markers - Microsatellites



- Short sections of repeated sequences of DNA
- Mutations in non-coding (neutral) DNA region
 - Some mutations common to genus, species, populations
 - Some mutations unique to individual
- Variation in length of repeats = marker
- Uses sequence information

Flanking region

CTAGCTAG **GATAGATAGATA**

Flanking region

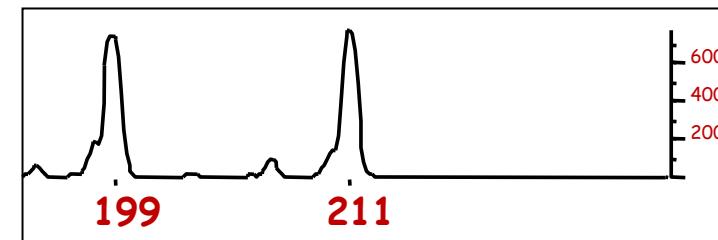
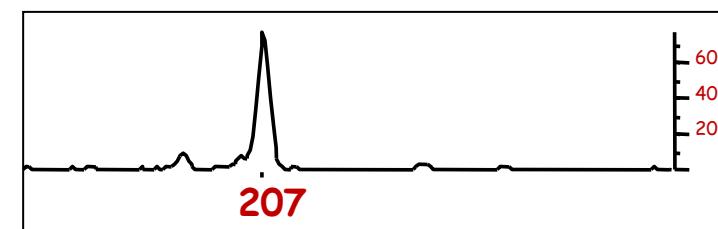
TTGTCA

CTAGCTAG **GATAGATAGATA**

TTGTCA

CTAGCTAG **GATA** TTGTCA

CTAGCTAG **GATAGATAGATA** TTGTCA



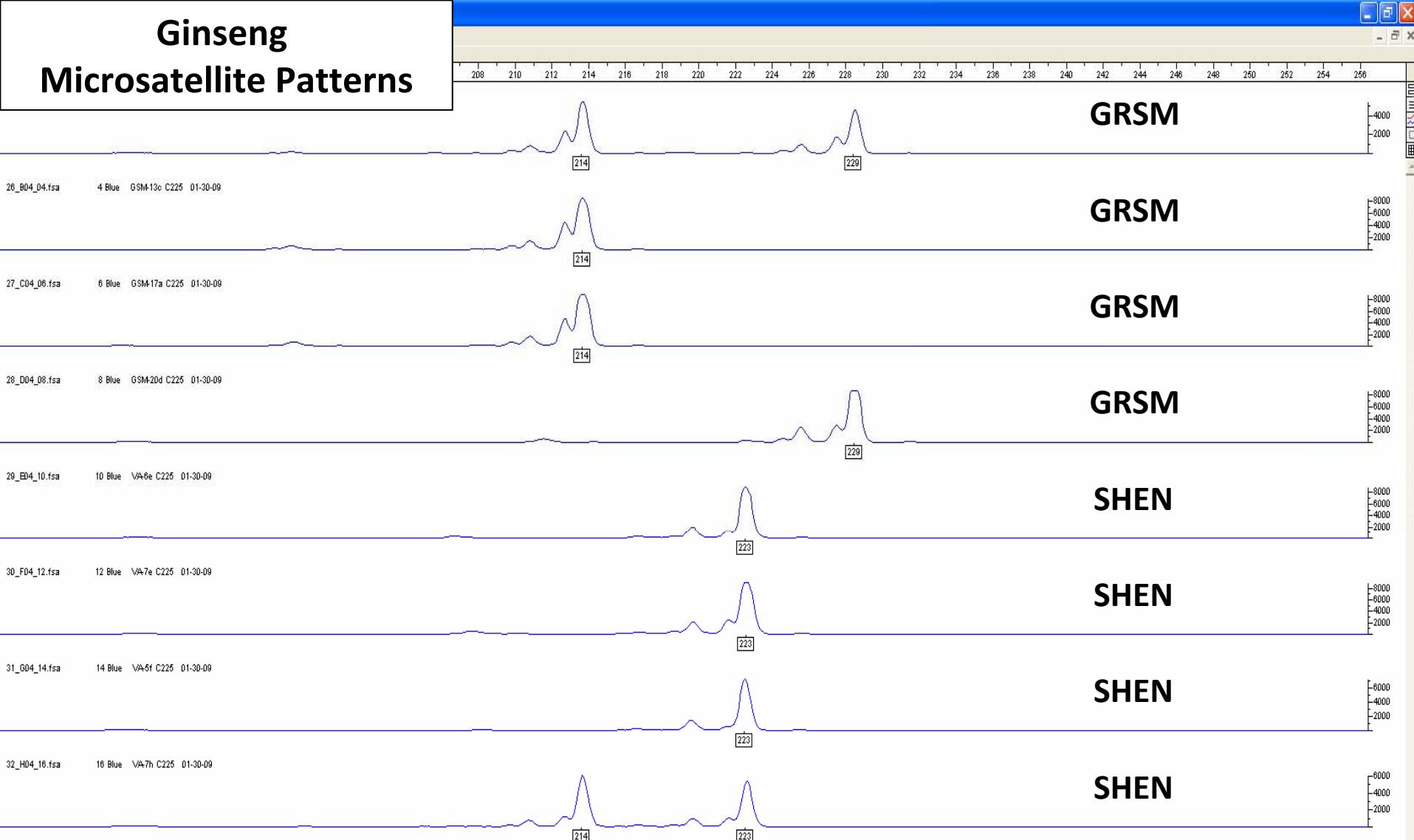
Genetic Markers – Microsatellites



King Lab

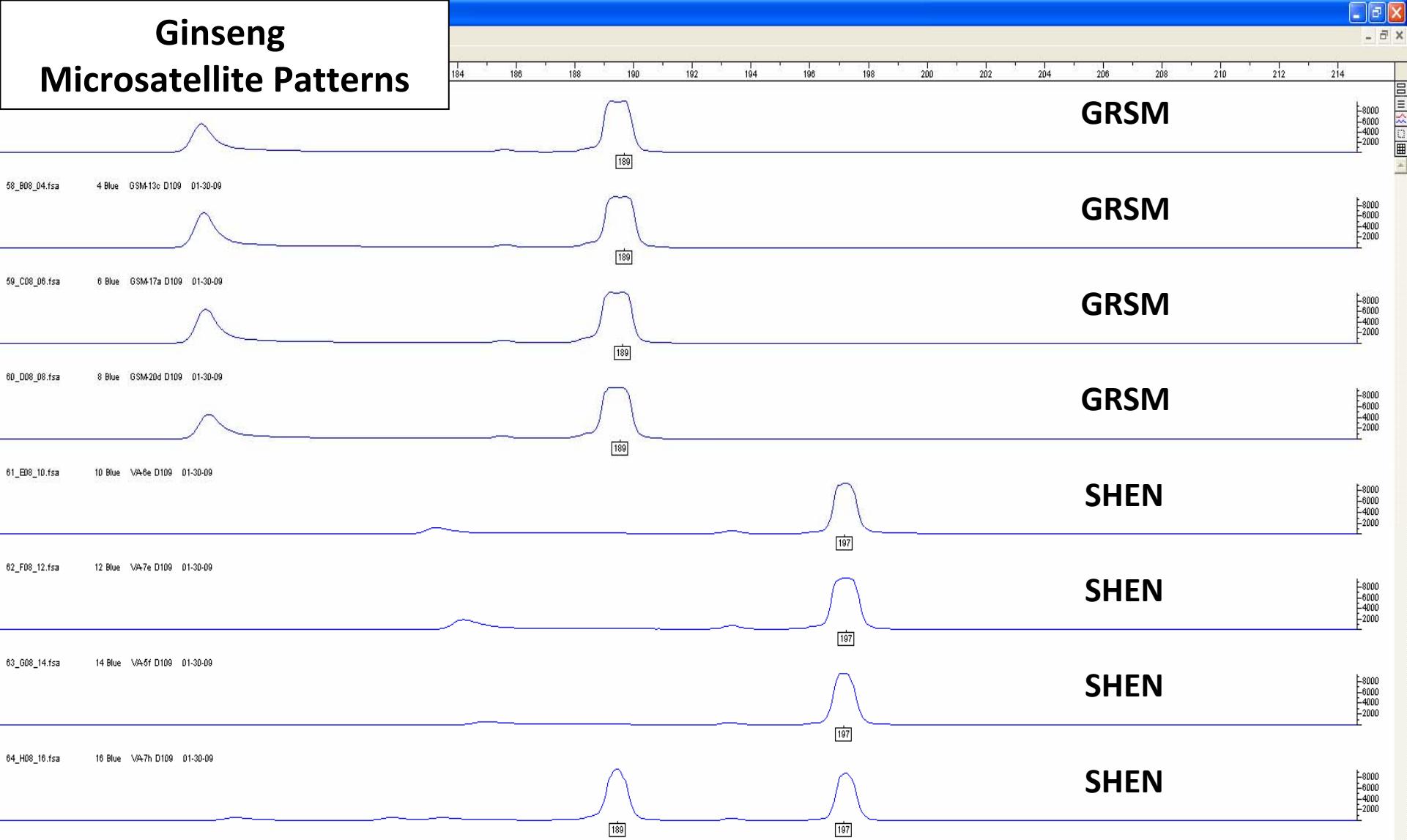
- Need 12-15 markers for population and individual discrimination
- Tested published markers for Korean ginseng (*Panax ginseng*)
 - Kim et al. 2007 [10/14], Ma et al. 2007 [14/22]
 - Amplification problems in *Panax quinquefolius*
 - Lack of polymorphism
- Developed our own set of 25 microsatellite markers for American ginseng, 14 look most promising
 - Enriched microsatellite DNA libraries created
 - Markers screened and tested in our laboratory

Ginseng Microsatellite Patterns



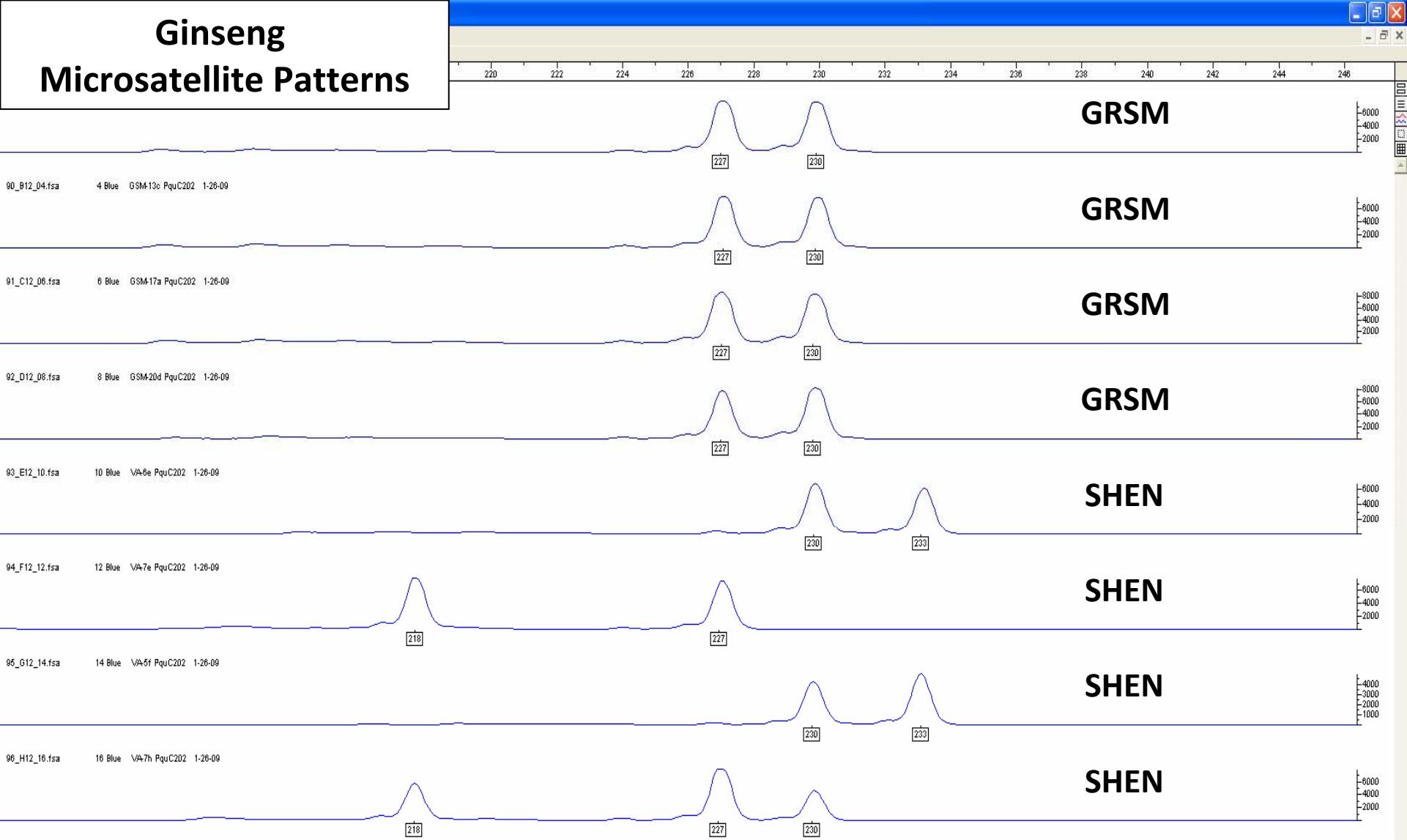
PquC225

Ginseng Microsatellite Patterns



PquD109

Ginseng Microsatellite Patterns



PquC202

Test of microsatellite markers

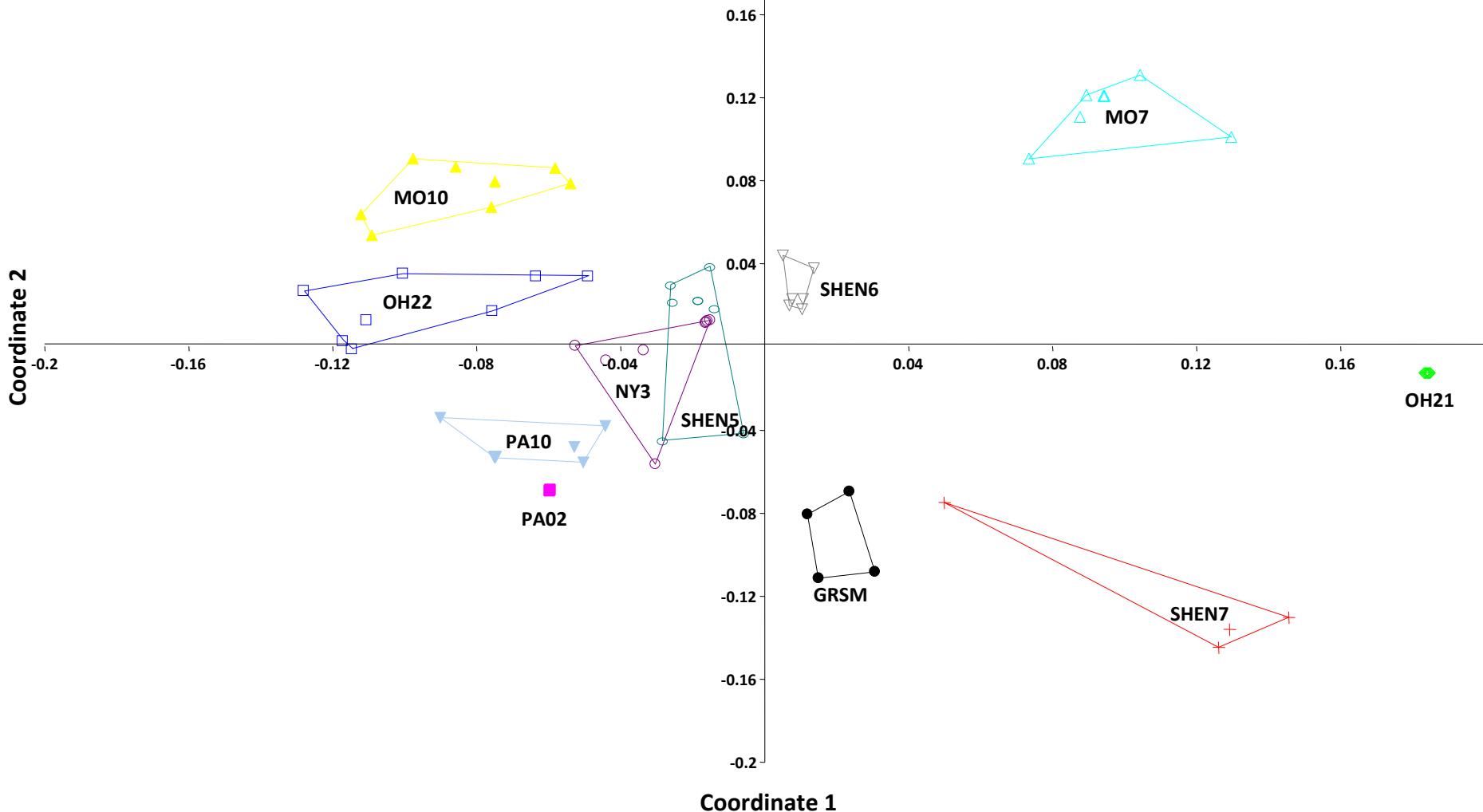


- Subset of sites across geographic range, and land ownerships (78 individuals)
- Test of allele frequency differences across 12 of 14 loci, 3-9 alleles each, (66 markers)
- Sites:
 - GRSM = Great Smoky National Park (4 plants)
 - SHEN5, SHEN6, SHEN7 = Shenandoah National Park, VA (3 sites, 18 plants)
 - MO10 = Roaring River State Park, MO (8 plants)
 - MO7 = Caney Mountain Conservation Area, MO (8 plants)
 - PA02 = Allegheny National Forest, PA (8 plants)
 - PA10 = Private land, western PA (8 plants)
 - NY3 = Private land, Cairo, NY (8 plants)
 - OH22 = Cultivated ginseng farm, OH (8 plants)
 - OH21 = Edge of Appalachia Preserve, OH (8 plants)



Microsatellite DNA Collection Differentiation

Non-metric Multidimensional Scaling (MDS)
Plot of individual Jaccard's distance values

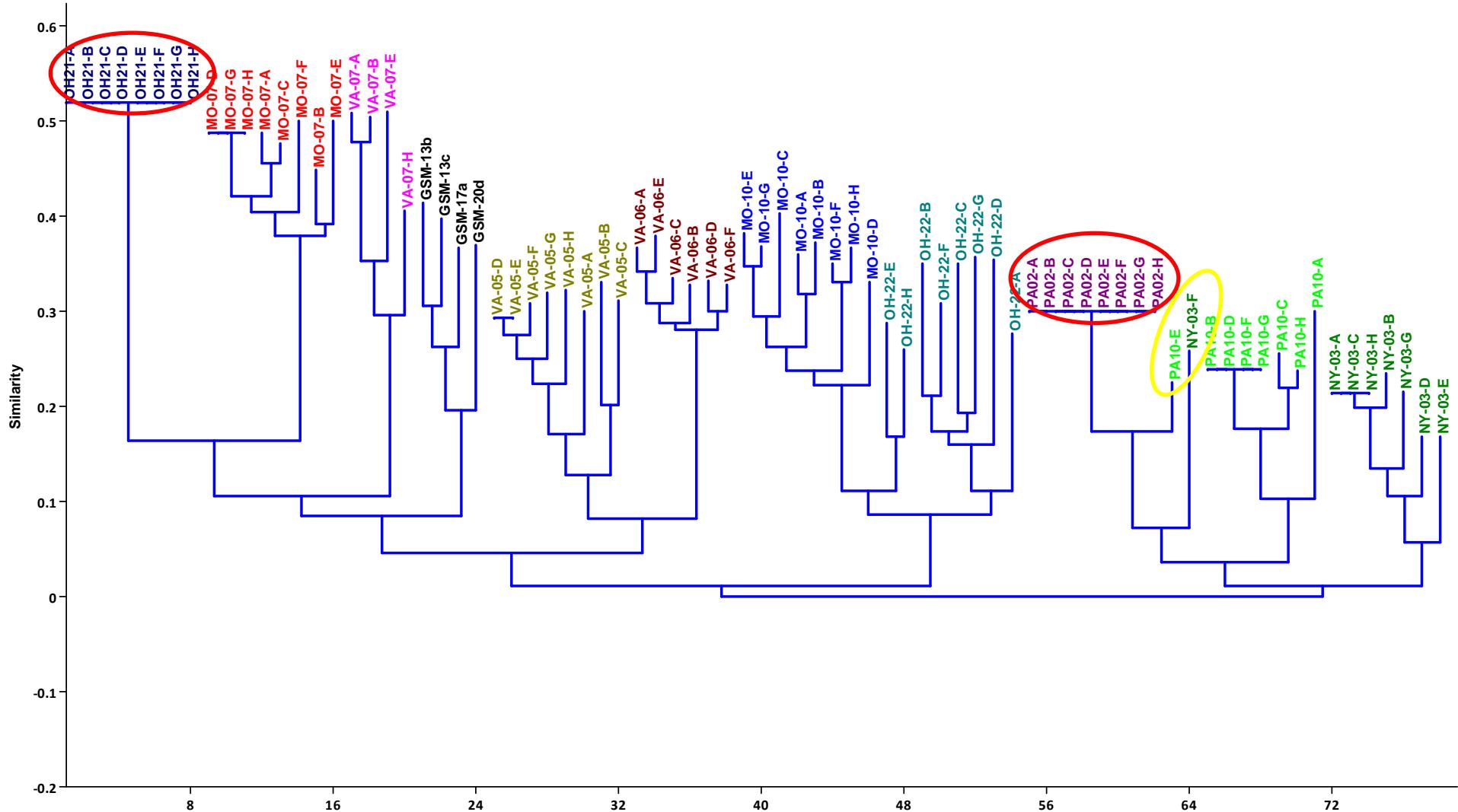


Polygons depict inter- and intra-collection variation in Jaccard's Distance

Microsatellite DNA Collection Differentiation



Neighbor Joining Tree on Jaccard's distance values





Φ_{PT} - Test of Genetic Differentiation

(F_{ST} analog for binomial data)



Collection	GRSM	M07	M010	NY	OH22	OH21	PA02	PA10	SHEN5	SHEN6	SHEN7	Prob.
GRSM	0.000	0.002	0.002	0.002	0.003	0.007	0.004	0.001	0.005	0.007	0.023	GRSM
M07	0.806	0.000	0.001	0.001	0.003	0.001	0.002	0.001	0.001	0.002	0.003	M07
M010	0.747	0.804	0.000	0.001	0.001	0.001	0.001	0.002	0.001	0.001	0.002	M010
NY	0.651	0.760	0.680	0.000	0.001	0.001	0.001	0.001	0.002	0.001	0.006	NY
OH22	0.639	0.714	0.483	0.455	0.000	0.001	0.001	0.001	0.001	0.001	0.004	OH22
OH21	0.889	0.909	0.889	0.859	0.788	0.000	0.122	0.002	0.001	0.002	0.004	OH21
PA02	0.848	0.923	0.855	0.771	0.691	0.000	0.000	0.001	0.001	0.001	0.003	PA02
PA10	0.681	0.838	0.752	0.531	0.542	0.899	0.771	0.000	0.001	0.002	0.004	PA10
SHEN5	0.667	0.776	0.675	0.427	0.529	0.854	0.783	0.667	0.000	0.002	0.001	SHEN5
SHEN6	0.768	0.851	0.765	0.637	0.648	0.958	0.944	0.812	0.576	0.000	0.002	SHEN6
SHEN7	0.657	0.830	0.777	0.734	0.672	0.890	0.898	0.769	0.679	0.771	0.000	SHEN7
Φ_{PT}	GRSM	M07	M010	NY	OH22	OH21	PA02	PA10	SHEN5	SHEN6	SHEN7	

Pair-wise Φ_{PT} (below diagonal) and probability Φ_{PT} value is > 0 (above).
 Comparison in yellow is not statistically significant.



Allocation to Collection



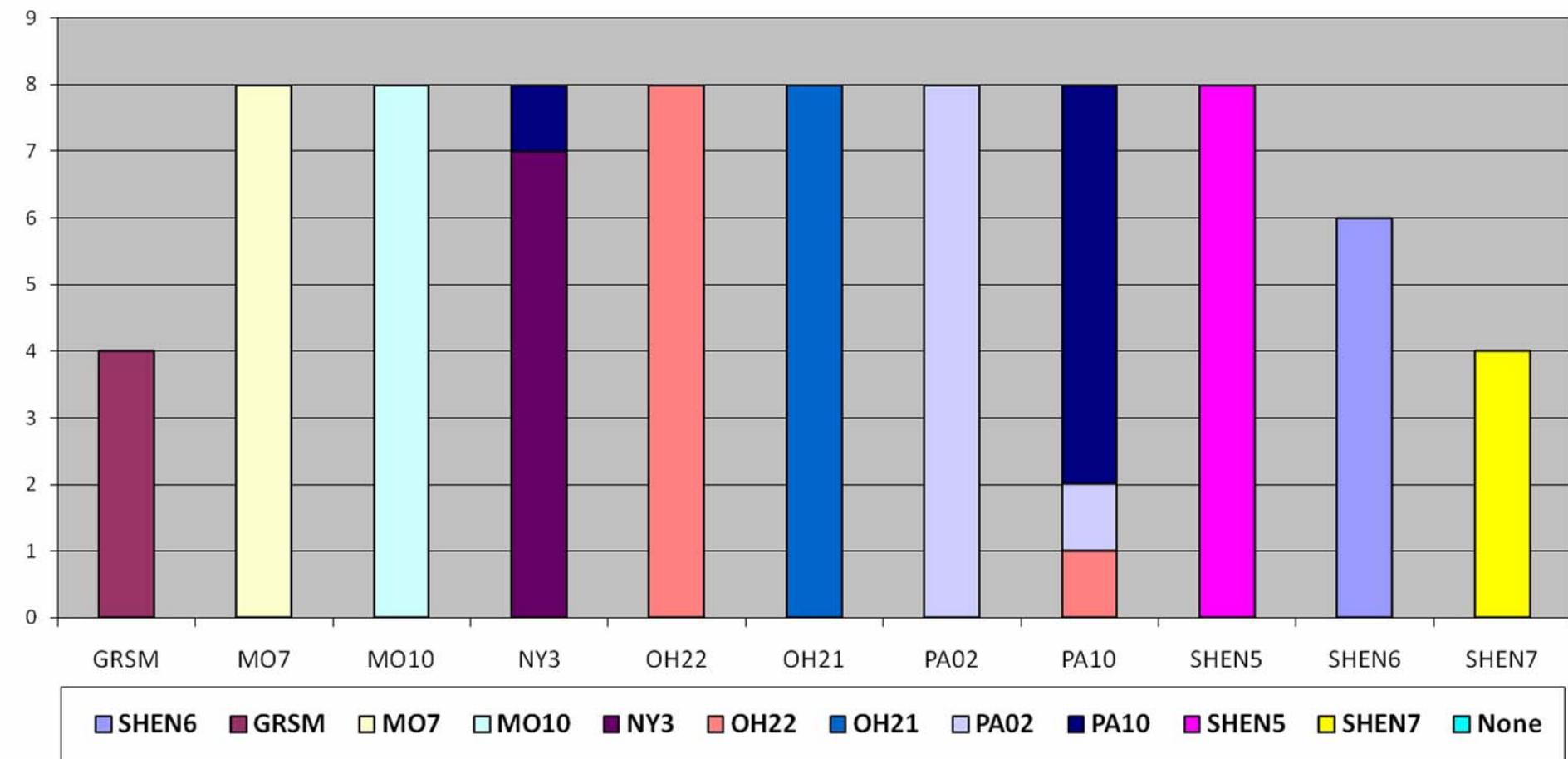
allocated to	GRSM	MO7	MO10	NY3	OH22	OH21	PA02	PA10	SHEN5	SHEN6	SHEN7
GRSM	4	0	0	0	0	0	0	0	0	0	0
MO7	0	8	0	0	0	0	0	0	0	0	0
MO10	0	0	8	0	0	0	0	0	0	0	0
NY3	0	0	0	7	0	0	0	0	0	0	0
OH22	0	0	0	0	8	0	0	1	0	0	0
OH21	0	0	0	0	0	8	0	0	0	0	0
PA02	0	0	0	0	0	0	8	1	0	0	0
PA10	0	0	0	1	0	0	0	6	0	0	0
SHEN5	0	0	0	0	0	0	0	0	8	0	0
SHEN6	0	0	0	0	0	0	0	0	0	6	0
SHEN7	0	0	0	0	0	0	0	0	0	0	4
Correct Assignment (%)	100.0%	100.0%	100.0%	87.5%	100.0%	100.0%	100.0%	75.0%	100.0%	100.0%	100.0%

Overall Assignment Success: 96.2%

Incorrect assignments are distributed vertically and are highlighted in yellow.



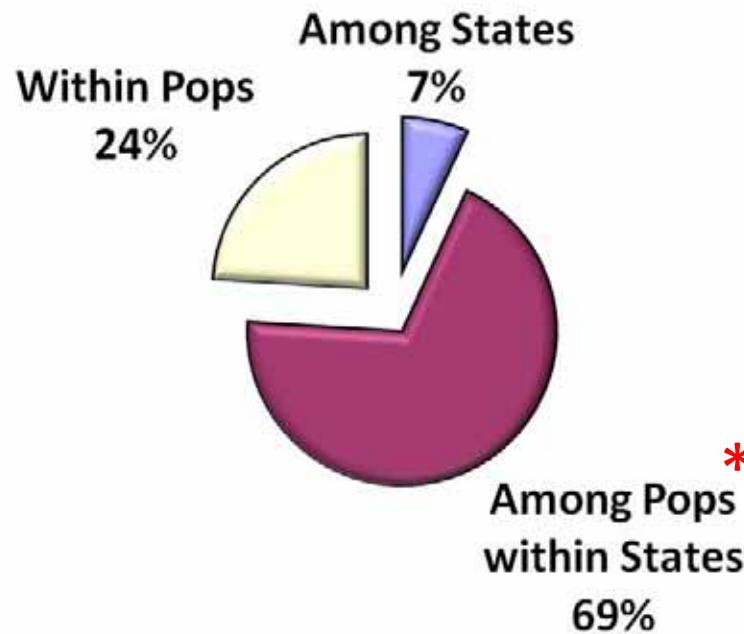
Microsatellite DNA Allocation Graphic





AMOVA (Analysis of MOlecular VAriance)

Partitioning of Genetic Variation



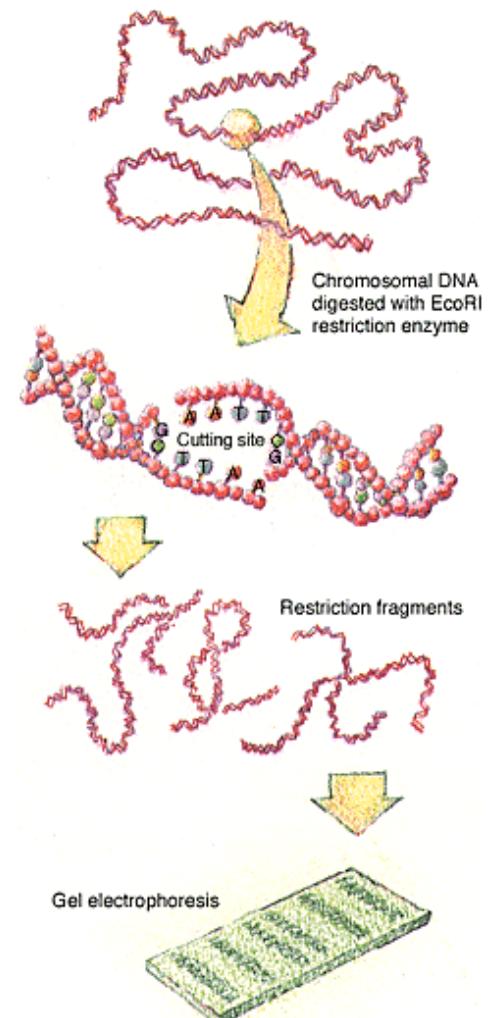
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Preliminary results (AMOVA, pair-wise Φ_{PT} , and assignment tests),
needs validation with larger dataset



Genetic Markers - AFLP

- AFLP = “Amplified Fragment Length Polymorphism”
 - No sequence information required
 - Highly repeatable
- Two step process:
 1. Cut DNA with restriction enzymes (primers), bind adapters to end of fragments
 2. Use PCR to amplify fragments
- Polymorphisms from:
 - Sequence variation in and around restriction sites
 - Insertions or deletions in amplified fragments
- High numbers of dominant (presence/absence) allelic markers produced

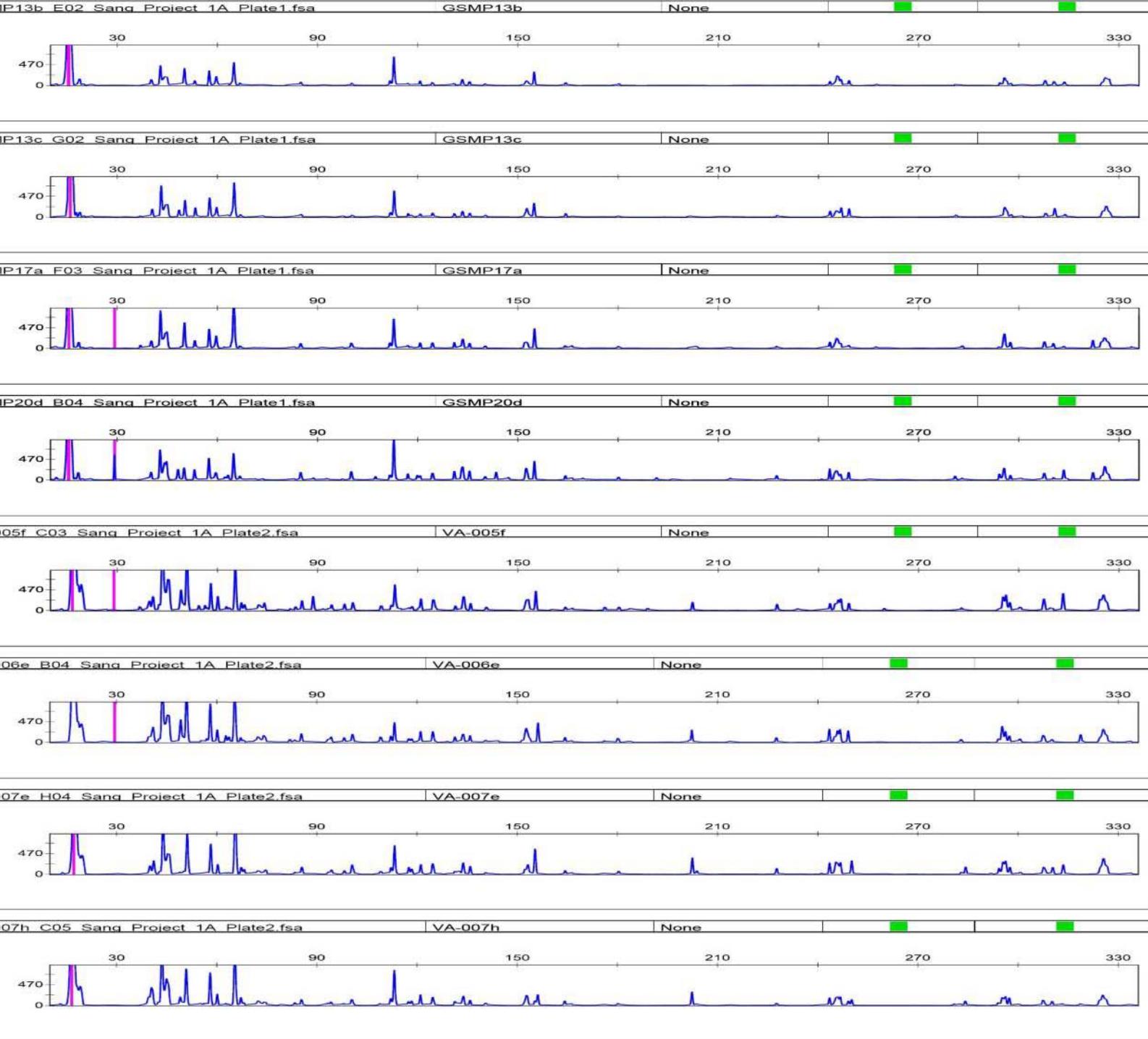


Genetic Markers – AFLP

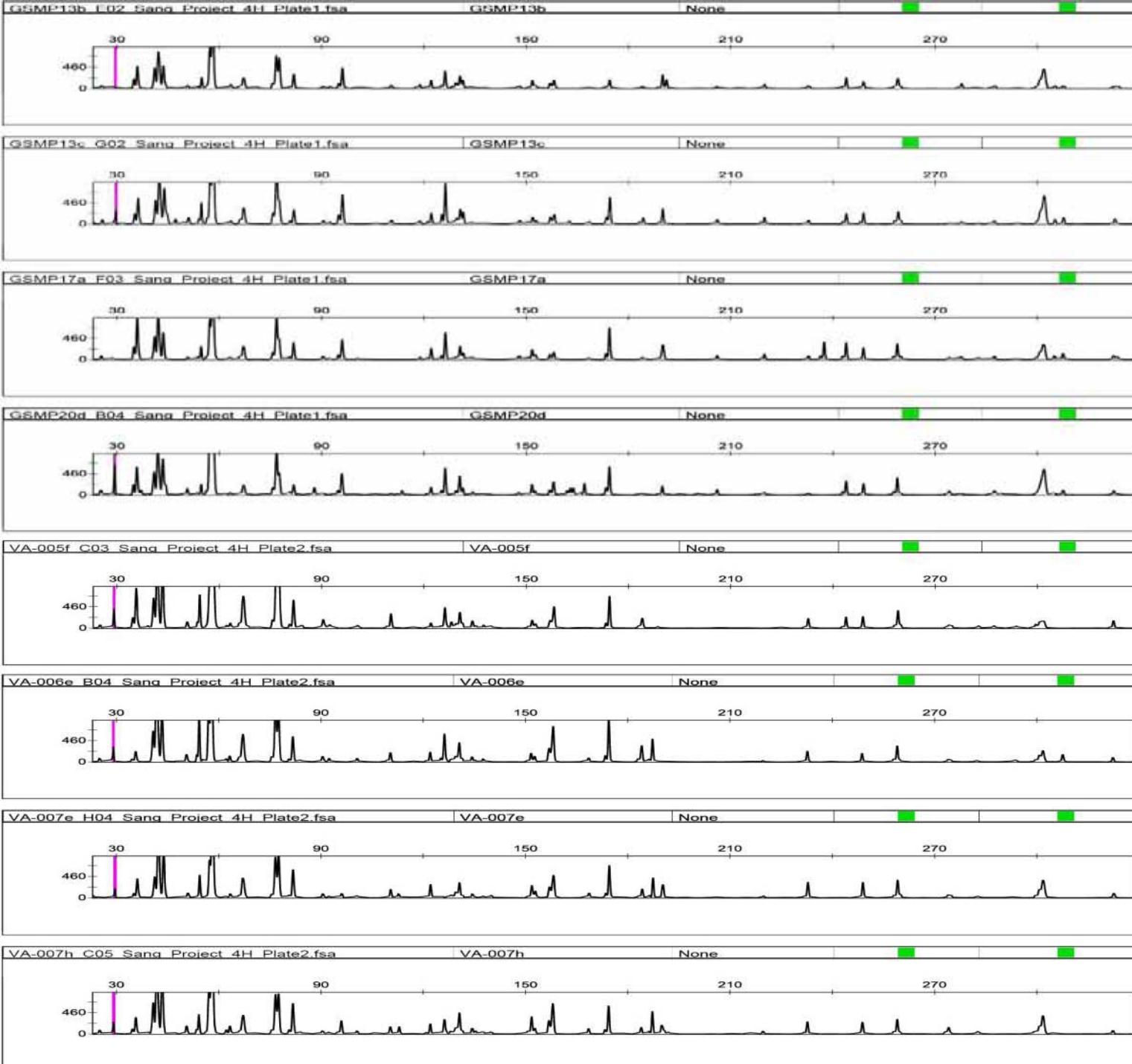
David DeViney, M.S. Student, ASU



- We tested 3 primer pairs for American ginseng:
 - 1) “Pqu1-A”: EcoRI-ACT, MseI-CAA
 - 2) “Pqu4-H”: EcoRI-ACC, MseI-CTT
 - 3) “Pqu7-A”: EcoRI-AGG, MseI-CAA
- Number of allelic markers generated for American ginseng:
 - 378 with 3 primers (1+2+3)
 - 240 with 2 primers (1+2)

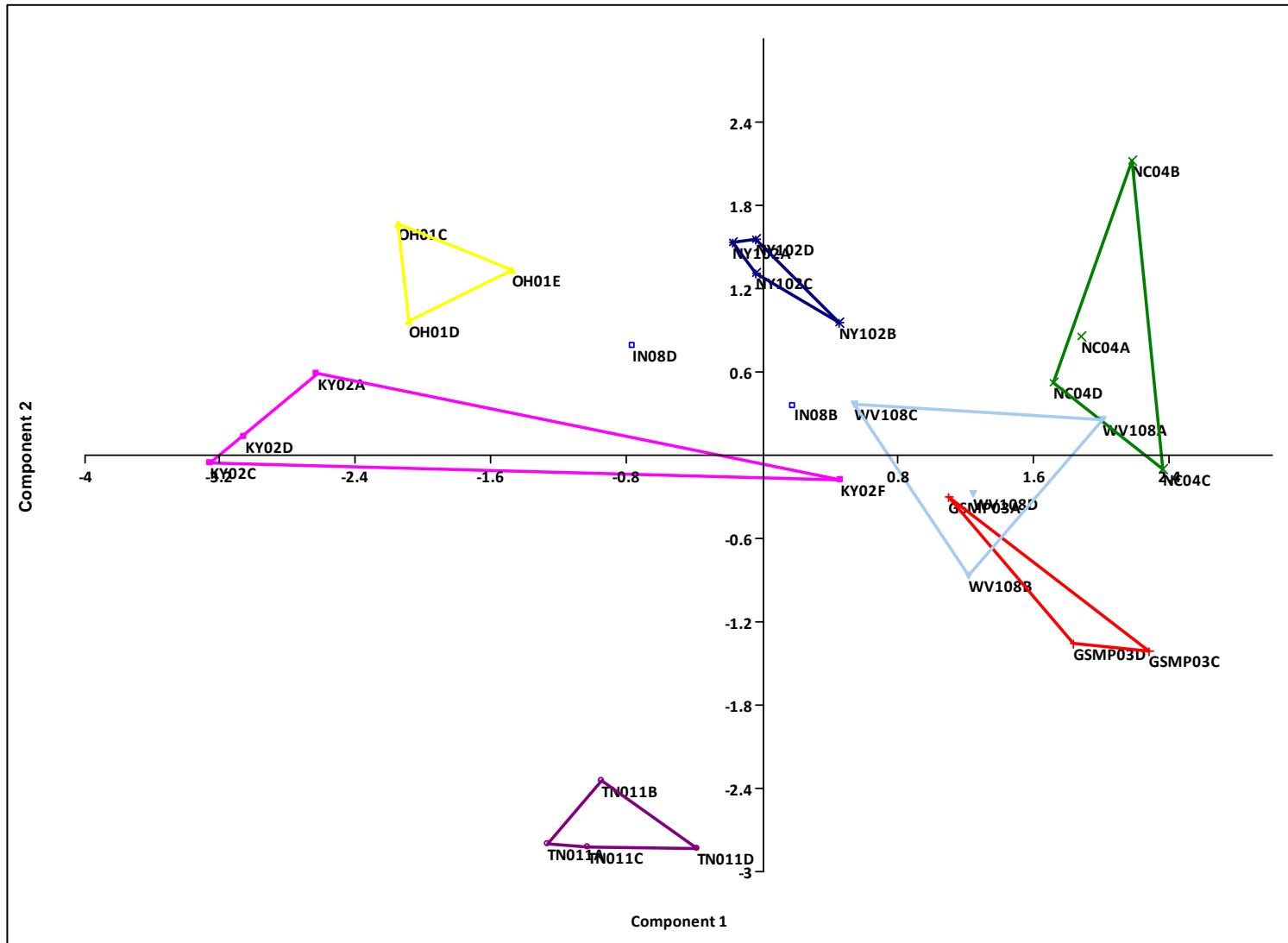


Pau4-H



Population discrimination – AFLP

Principal Components Analysis



Remaining tasks:

- Winter-Spring 2009
 - Develop new ginseng distribution models
 - Survey all genetic samples for AFLP and microsatellite variation
- Early summer 2009
 - Collect additional genetic samples
- Late summer-early fall 2009
 - Data analysis
 - Assess phylogeographic and population structure
 - Diversity within/among sites by land ownership
 - Genetic diversity compared to habitat distribution/quality

Acknowledgements

- Marcus Springmann (USGS) – field data collection, genetic laboratory assistance
- Chris Walter (USGS) – field data collection, assistance with GIS data compilation
- Pat Ford (FWS), state ginseng coordinators – contacts with land owners
- Bob Beyfuss, Eric Burkhart – contacts with private and state landowners
- Numerous landowners and land managers who provided access to their property for sampling

