KIS™ Test District Correlation Meeting

August 20, 2009









- The Charm KISTM test is an antibiotic detection test for fresh or thawed kidney tissue.
- Its principle of detection is microbial inhibition. Bacteria, cultured in agar with purple pH indicator media and kidney extract, generate acid that produces a yellow color. In the presence of antibiotic, the bacterial growth is inhibited and the test remains blue/purple.





Essentially, there are only a few steps to the test

- Out tissue and absorb sample Note-Work in batches. It's helpful to loosen all of the swabs from the KIS™ tests prior to swabbing the kidneys.
- Activate swab for two minutes
 Note-Be sure EACH swab is kept in the top clear liquid for two minutes.
- Incubate tests and read results
 Note-Incubate at 64 ± 2 C for appropriate time. Time depends on when results are to be read and may be lot dependant. Be sure to check times prescribed on the packaging.





- MUST have a Negative Control Sample with each batch of samples and for each lot used
 - Note-Different lot numbers may prescribe different incubation times.
 Use a negative control for each lot used.
- Negative Control can be used up to 5 days (refrigerated)
 - Note-Video and Written material state 24 hours! Label appropriately to ensure proper storage.
- Negative Control Sample MUST remain yellow for sample batch results to be valid



Additional Notes Heating Block Use

- Allow ~ 30-60 minute for temperature equilibration
- Avoid setting unit in drafty areas. This will cause longer equilibration times.
- Determine when results will be read
 - □ ~3 hours
 - Set timer using prescribed time listed on kit packaging
 - Remove tubes when prescribed time has elapsed
 - □ Up to 16 hours
 - Subtract 15 minutes from prescribed time on kit packaging
 - Leave and allow tubes to cool in heating block



Additional Notes Batching and Handling Samples

- To avoid mess.
 - Recommend placing Kidney in plastic bag
 - ☐ Slice bag (ex. Use scalpel)
 - □ Using the cookie cutter end, score kidney through hole in bag
- Handle Samples in Batches
 - This helps because of the time determinative 2 minute activation step.
 - One possible solution is shown in the video.
 - Score and swab samples
 - Place swabs in tubes BUT DO NOT engage.
 - Engage tube and start two minutes. Engage remaining tubes down the line.



Additional Notes KIS™ Results Interpretation



 Yellow or yellow/green colors are negative.



 Blue/purple colors are positive.

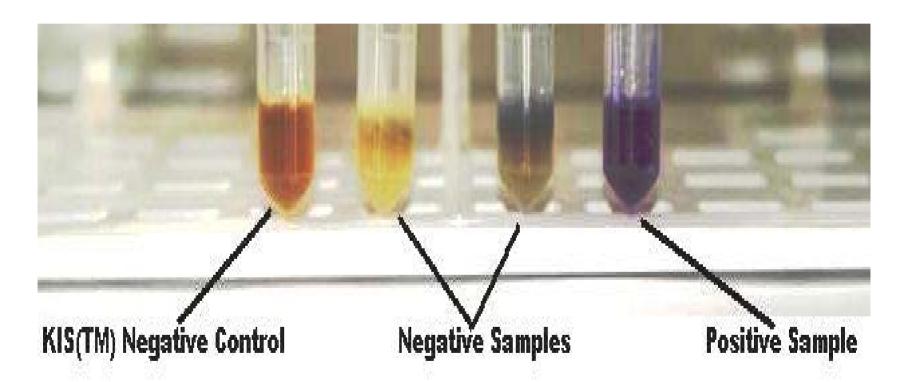


Yellow or yellow/green in lower half of vial with blue/purple or brown in upper half of vial are CAUTION. These samples should be interpreted as negative.



Additional Notes KIS™ Results Interpretation

Note-Read lower half of vial



NOTE: Kit Negative Control Note Brownish Color

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Question:

■ How should the inspector interpret the color changes associated with the KIS™ test?



- Answer:
- "P"ervasive (spread through-out) "P"urple color means "P"ositive – remember keep the "P"s together! A Yellow or Yellow/green color in the lower half of the tube means Negative. See below for pictures of positive and negative results ALWAYS READ THE BOTTOM HALF OF THE VIAL.



- Answer:
- 2. The KIS tube comes inoculated with bacteria. The inspector adds kidney extract from the animal being tested to the tube. If the kidney extract that was added to the tube does not contain antibiotics, the bacteria will grow and multiply. These growing bacteria produce acid. The acid build-up in the tube changes the color from purple to yellow. The bacteria grew, there was acid production, and the color changed from purple to yellow. This is a negative test.



- Answer:
- 3. When microbial inhibitors (antibiotics) are present in the kidney extract from the carcass being tested, the bacteria will not grow, acid will not be produced, and the color will not change (it will remain purple)



KIS™ Questions?

- Contact RIMD via AskFSIS Sampling
- Call 1-800-233-3935 and select #4 for RIMD