

Gene \times Disease Interaction on Orbitofrontal Gray Matter in Cocaine Addiction

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Context: Long-term cocaine use has been associated with structural deficits in brain regions having dopamine-receptive neurons. However, the concomitant use of other drugs and common genetic variability in monoamine regulation present additional structural variability.

Objective: To examine variations in gray matter volume (GMV) as a function of lifetime drug use and the genotype of the monoamine oxidase A gene, MAOA, in men with cocaine use disorders (CUD) and healthy male controls.

Design: Cross-sectional comparison.

Setting: Clinical Research Center at Brookhaven National Laboratory.

Patients: Forty individuals with CUD and 42 controls who underwent magnetic resonance imaging to assess GMV and were genotyped for the MAOA polymorphism (categorized as high- and low-repeat alleles).

Main Outcome Measures: The impact of cocaine addiction on GMV, tested by (1) comparing the CUD group with controls, (2) testing diagnosis \times MAOA interac-

tions, and (3) correlating GMV with lifetime cocaine, alcohol, and cigarette smoking, and testing their unique contribution to GMV beyond other factors.

Results: (1) Individuals with CUD had reductions in GMV in the orbitofrontal, dorsolateral prefrontal, and temporal cortex and the hippocampus compared with controls. (2) The orbitofrontal cortex reductions were uniquely driven by CUD with low-MAOA genotype and by lifetime cocaine use. (3) The GMV in the dorsolateral prefrontal cortex and hippocampus was driven by lifetime alcohol use beyond the genotype and other pertinent variables.

Conclusions: Long-term cocaine users with the low-repeat MAOA allele have enhanced sensitivity to gray matter loss, specifically in the orbitofrontal cortex, indicating that this genotype may exacerbate the deleterious effects of cocaine in the brain. In addition, long-term alcohol use is a major contributor to gray matter loss in the dorsolateral prefrontal cortex and hippocampus, and is likely to further impair executive function and learning in cocaine addiction.

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DRUG ADDICTION IS A chronic disease associated with deficits in brain dopamine¹ (DA) and brain function in regions underlying the impaired response inhibition and salience attribution syndrome (see Goldstein and Volkow² for review). These regions encompass the reward and the inhibitory circuitry that contain DA-receptive neurons, where ventral prefrontal regions such as the orbitofrontal cortex (OFC) have received much emphasis.^{2,3} Multiple neuroimaging studies in the past decade demonstrated a reliable pattern of functional deficits during cognitive/emotional challenges that involve reward contingencies (salience attribution) and inhibitory control (response inhibition) in cocaine use disorders (CUD).^{4,5} For example, positron emission tomography and functional magnetic reso-

nance (MR) imaging studies have demonstrated that DA-related functional deficits in the OFC may underlie disproportionate salience attribution to cocaine and compulsive drug intake.^{3,5-7}

Although relatively few, studies have tested structural alterations in the same circuitry where functional activations are compromised and have documented such deficits.⁸ Individuals with cocaine addiction have shown decreased gray matter volume (GMV) or thinner cortex in the dorsolateral prefrontal cortex (DLPFC), OFC, and anterior cingulate cortex⁹⁻¹²; other regions included the insula, temporal cortex, and amygdala compared with healthy controls (CON).¹¹⁻¹⁴ Because DA projections influence cerebral morphologic characteristics during development and throughout adulthood, it is expected that long-term exposure to substances that trigger supraphysi-

ologic DA levels in the synapse, such as cocaine, might cause persistent cellular changes resulting in reduced neural volume compared with nonexposed individuals.⁸ Moreover, positron emission tomography studies have shown that the reduction in brain metabolism in DLPFC, OFC, and anterior cingulate cortex in cocaine abusers is associated with loss of postsynaptic DA markers.¹⁵

Addiction to crack cocaine involves long-term concurrent use of other substances that are known to influence brain morphologic characteristics.¹⁶⁻¹⁹ More than 60% of individuals with CUD also had a comorbid alcohol use disorder and more than 80% smoked cigarettes, further compounding GM loss throughout the brain.¹⁶⁻²⁰ These high comorbidity rates make the assessment of long-term drug use other than cocaine imperative for the generalizability of the results to community samples of individuals with CUD. Therefore, the present study used MR imaging and whole-brain voxel-based morphometry (VBM) analysis to test changes in cerebral GMV as a function of CUD and in correlation with the chronicity of lifetime drug use. This analysis, however, does not indicate whether the predicted structural alterations result *uniquely* from years of chronic drug use. It is possible that individuals with CUD had reduced DA and reduced neural volume in the relevant brain circuits before disease onset, which could have predisposed them to drug use and addiction. The potential contribution of genetic differences to GMV may be present before disease onset and may interact with long-term drug use, rendering some individuals with CUD more sensitive to GM loss than others.

Genetic variations that interact with and affect brain development may contribute to behaviors that increase addiction liability.²¹ The product of the monoamine oxidase A gene, *MAOA*, is an enzyme that regulates the metabolism of monoamine neurotransmitters, thereby modulating brain function and structure.^{22,23} During prenatal development, the *MAOA* enzyme is crucial for catabolic degradation of DA and norepinephrine,²³ inducing changes with long-term consequences during childhood.²⁴ The *MAOA* genotype (defined as OMIM +309850), a variable number tandem repeat (uVNTR) region, is divergent in primates, suggesting that it plays a pivotal role in differential *MAOA* expression in both humans and monkeys.²⁵ The *MAOA* genotype is relevant to GMV in healthy CON.^{26,27} In a large VBM study, healthy carriers of the low-repeat allele of *MAOA* (*MAOA*^{*L}) had reduced GMV in the cingulate cortex and bilateral amygdala and increased GMV in the OFC compared with high-repeat allele (*MAOA*^{*H}) carriers.²⁸ Furthermore, in the presence of extreme environmental challenge (childhood abuse), *MAOA*^{*L} genotype increases the risk of antisocial behaviors in adulthood, pointing to a gene \times environment interaction.²⁹ Studies have also suggested association of *MAOA*^{*L} with the risk of alcohol addiction.^{30,31} We reasoned that, for individuals with CUD, the disease onset and its progression could be viewed as an environmental challenge,³² possibly influencing GMV in affected members of the *MAOA*^{*L} genotype (CUD-L group).

Therefore, in this study we predicted a main effect of addiction by which individuals with CUD would have reductions in GMV compared with CON. Next, we hypothesized a gene \times disease interaction driven mostly by GMV loss in the CUD-L group. We hypothesized that a

model containing both genetic and long-term drug use variables would better explain the predicted morphologic deficits in CUD.

METHODS

PARTICIPANTS

Eighty-two right-handed men (40 with CUD and 42 CON) were recruited by advertisement in local newspapers. All participants provided informed consent in accordance with the local institutional review board. Physical/neurologic, psychiatric, and neuropsychological examinations were conducted and included tests of intellectual functioning (Wide-Range Achievement Test 3 reading³³ and the Matrix Reasoning subset of the Wechsler Abbreviated Scale of Intelligence³⁴), Beck Depression Inventory (BDI)³⁵ to assess symptoms in the past 2 weeks, the Addiction Severity Index,³⁶ and the Structured Clinical Interview for DSM-IV Axis I Disorders (research version).³⁷ All participants were healthy, were not taking any medications, and were excluded if they had contraindications to the MR imaging environment (eg, metal in the body or claustrophobia), history of head trauma or loss of consciousness (>30 minutes), other neurologic disease, abnormal vital signs at time of screening, history of major medical conditions (cardiovascular, endocrinologic, oncologic, or autoimmune diseases), major psychiatric disorders (other than cocaine dependence and alcohol abuse for the CUD group and/or nicotine dependence for both groups), and urine positive (by means of a urinalysis kit [Biopsych; Biopsych Triage, San Diego, California] for psychoactive drugs or their metabolites (phencyclidine, benzodiazepines, amphetamines, cannabis, opiates, barbiturates, and inhalants) except for cocaine in CUD.

All participants in the CUD group were current users: urine was positive for cocaine in all but 6 of the 40 individuals, and they reported use a mean (SD) of 2.1 (1.5) days before the study. Current use or dependence on other drugs was denied and corroborated by preimaging urine tests in all participants (urine was negative for all other drugs in all participants). **Table 1** contains the demographic and clinical comparisons between the CUD and CON groups with nested genotype comparisons.

GENOTYPING

The DNA samples for *MAOA* genotyping were extracted from whole blood (PAXgene Blood DNA Kit; Qiagen Inc, Valencia, California) from each participant. The polymerase chain reactions were performed as previously described.²⁷ In humans and primates, categorization of common genetic variability is based on a functional polymorphism in the promoter region of the *MAOA* gene; uVNTR, 3.5 or 4 repeats (ie, "high") and 2, 3, or 5 repeats ("low") is common in the population in whom 3 and 4 occur in a ratio of approximately 60:40 in men. Compared with the high variant, the low variant has relatively lower transcriptional activity in human nonneural cell lines.^{27,38} In this sample, alleles were observed in expected ranges by means of GeneScan version 3.7 and Genotyper version 3.6 software (both Applied Biosystems, Carlsbad, California). Genetic analyses resulted in 42 participants classified as having the low-*MAOA*-repeat alleles (22 CUD-L and 20 CON-L) and 40 having the high-repeat alleles (18 CUD-H and 22 CON-H).

MR IMAGE ACQUISITION AND VOXEL-BASED MORPHOMETRY

All participants underwent T1-weighted anatomic MR imaging on a 4-T imager (Varian/Siemens, Malvern, Pennsylvania), with

Table 1. Demographic and Drug Exposure Factors

	CUD ^a (n=40)		CON ^a (n=42)		Tests ^b
	Low (n=22)	High (n=18)	Low (n=20)	High (n=22)	
Participant Characteristics					
Age, y	45 (1)	45 (1)	40 (1)	38 (1)	CUD > CON; $F_{1,81}=12.3, P<.001$
Race, No. black/white	18/4	17/1	14/6	12/10	CUD; $\chi^2_{3,82}=9.7, P=.03$
Education, y	13 (0.3)	13 (0.4)	13 (0.3)	14 (0.5)	$F_{1,74}=2.3, P=.08$
SES	32 (2)	26 (2)	34 (3)	34 (3)	$F_{1,74}=2.1, P=.10$
Verbal IQ ^c	93 (3)	86 (2)	92 (3)	101 (3)	CUD < CON; $F_{1,74}=5.2, P=.002$
Nonverbal IQ ^d	9 (1.0)	9 (1.0)	10 (0.5)	11 (1.0)	CUD × MAOA; $F_{1,74}=10.2, P=.002$
BDI symptoms score	9 (2)	9 (2)	2 (1)	5 (1)	$F_{1,74}=2.2, P=.09$ CUD > CON; $F_{1,74}=16.2, P=.03$
Drug Exposure Factors					
Age at CUD onset, y	24 (1.3)	28 (2.0)	NA	NA	CUD-L < CUD-H; $t_{36}=-2.07, P=.09$
Cocaine intake, g/occasion	1.9 (1.0)	1.6 (0.4)	NA	NA	$t_{36}=-0.3, P=.82$
Cocaine use, y	19 (1.4)	19 (1.3)	NA	NA	$t_{36}=-0.052, P=.97$
Cigarette smokers, No. (%)	17 (77)	13 (72)	5 (25)	4 (18)	CUD > CON; $\chi^2_{3,74}=23.2, P=.001$
Cigarettes, No./d	8 (1)	9 (1)	6 (1)	4 (2)	$F_{3,43}=.64, P=.32$
Age at smoking onset, y	15 (2)	17 (1)	17 (2)	19 (1)	$F_{3,43}=1.54, P=.16$
Years of smoking	21 (2)	22 (2)	4 (5)	2 (2)	CUD > CON; $F_{3,40}=4.0, P=.001$
Alcohol abuse, No. (%)	15 (68)	13 (72)	NA	NA	$t_{28}=0.1, P=.97$
Alcohol consumption, oz	62 (10)	59 (6)	34 (1)	30 (3)	CUD > CON; $F_{3,74}=2.2, P=.007$
Age at alcohol abuse onset, y	15 (1)	16 (1)	NA	NA	$t_{21}=-0.87, P=.21$
Years of drinking ^e	19 (2.0)	17 (3.0)	5 (1.0)	2 (0.5)	CUD > CON; $F_{1,74}=49.7, P<.001$

Abbreviations: BDI, Beck Depression Inventory; CON, controls; CUD, cocaine use disorders; CUD-H, CUD with high-repeat monoamine oxidase gene (MAOA) genotype; CUD-L, CUD with low-repeat MAOA genotype; NA, not applicable; SES, socioeconomic status.

^aValues are mean (SEM) unless otherwise noted. "Low" and "High" indicate low- and high-repeat MAOA genotype.

^bResults of general linear model with significant results labeled (eg, CUD).

^cFrom the Wide-Range Achievement Test, third edition.³³

^dFrom the Matrix Reasoning subset of the Wechsler Abbreviated Scale of Intelligence.³⁴

^eThe number of lifetime years of drinking (note that CON had years of drinking, although alcohol abuse was ruled out).

Sonata gradient set. The MR imaging variables of the 3-dimensional modified driven-equilibrium Fourier transform^{39,40} sequences were as follows: echo time/repetition time, 7/15 milliseconds; $0.94 \times 0.94 \times 1.00 \text{ mm}^3$ spatial resolution; axial orientation; 256 readout; and 192×96 phase-encoding steps, within a 16-minute imaging time. The modified driven-equilibrium Fourier transform sequence is particularly effective for white matter (WM)–GM tissue differentiation.⁴¹

All structural data were analyzed with MATLAB 7.0 (MathWorks, Inc, Natick, Massachusetts; <http://www.mathworks.com>) and statistical parametric mapping (SPM5; Wellcome Department of Cognitive Neurology, London, England; <http://www.fil.ion.ucl.ac.uk/spm>) with VBM5.1 toolbox (Christian Gaser, PhD, Department of Psychiatry, University of Jena, Jena, Germany; <http://dbm.neuro.uni-jena.de/vbm/>). Preprocessing (spatial normalization, tissue segmentation, and bias correction) was conducted by means of a unified model. Images were normalized to standard proportional stereotactic space (Montreal Neurological Institute). Tissue probability maps (International Consortium for Brain Mapping, European version) were subsequently applied, segmenting the images of all 82 participants into GM, WM, and cerebrospinal fluid (CSF) tissue classes for each individual following Bayesian rule.^{42,43} A hidden Markov random field⁴⁴ was applied to minimize the noise level by "removing" isolated voxels of one tissue class that are unlikely to be members of this tissue class, thus increasing the accuracy of the individual participant tissue probability maps. Finally, Jacobian modulation was applied to compensate for the expansion/contraction that occurs during nonlinear transformation and to restore the original absolute GMV in the segmented GM images. The voxel resolution after normalization was $1 \times 1 \times 1 \text{ mm}$. Statistical analysis of

the regional GMV was performed after smoothing the normalized and modulated segments by means of an isotropic 12-mm³ full-width at half-maximum gaussian kernel.

Total brain tissue was computed as a sum of the extracted GMV and WM volume (WMV) for each participant. We did not analyze WMV in this study because other methods, such as diffusion tensor imaging, are more sensitive for this purpose (VBM's WM T1 signal intensities are not correlated with the WM integrity).⁴⁵ As in other studies,⁴⁶⁻⁴⁸ CSF was not used in the calculation for total brain tissue because the value outputs by SPM5 are susceptible to artifacts (eg, if voxels are not fully differentiated as GM or WM, they can be mislabeled as CSF). In addition, GM and WM tend to vary together; however, CSF is variable from day to day and may increase as GM decreases, misleading the total brain calculation.⁴⁹

STATISTICAL ANALYSIS

Statistical analysis for the demographic and drug exposure factors was performed by means of a general linear model with a 2 (diagnosis: CUD vs CON) × 2 (genotype: low vs high) comparison or *t* tests or χ^2 , as needed, in SPSS (SPSS, Inc, Chicago, Illinois),⁵⁰ as documented in Table 1. In SPM5, general linear model 2×2 was used for the GM maps, controlling for total brain tissue and age, for the diagnosis main effect (CUD < or > CON) and the genotype main effect (MAOA*L < or > MAOA*H). Then, we conducted planned comparisons between CUD and CON of the same allele variation: CUD-L < CON-L and CUD-H < CON-H. Separate whole-brain regression analyses, controlling for total brain tissue and age, were

Table 2. Statistical Parametric Mapping Results of GM Differences^a

Region, BA	MNI Coordinates Peak Voxel			z Score	Cluster Size, mm ³	SPM5 P Value ^b
	x	y	z			
CUD < CON (n=82)						
OFC, 11	-22	24	-16	3.79	393	.04
DLPFC, 9	36	20	27	3.99	293	.02
Temporal, 37	50	-53	2	3.89	1116	.04
Hippocampus	35	-12	-16	4.30	5238	.04
Parahippocampus, 34	31	2	-18	3.55	5238	.02
CUD-L < CON-L (n=42)						
OFC, 11	-21	25	-16	4.01	1251	.04
Gyrus rectus	10	38	-21	3.74	1404	.04
Gyrus rectus	-8	37	-24	3.50	206	.04
DLPFC, 6	-51	-3	38	3.45	763	.04
DLPFC, 6	53	-1	33	4.08	16 313	.04
DLPFC, 9	16	48	13	4.35	16 313	.04
Temporal, 37	50	-60	0	3.91	7333	.04
Hippocampus	32	-13	-16	3.20	373	.04
CUD-H < CON-H (n=40)						
DLPFC, 9	35	19	29	3.43	105	.24
DLPFC, 6	50	1	28	3.18	637	.33
Hippocampus	39	-10	-14	3.84	2379	.04

Abbreviations: BA, Brodmann area; CON, controls; CON-H, CON with high-repeat monoamine oxidase gene (*MAOA*) genotype; CON-L, CON with low-repeat *MAOA* genotype; CUD, cocaine use disorders; CUD-H, CUD with high-repeat *MAOA* genotype; CUD-L, CUD with low-repeat *MAOA* genotype; DLPFC, dorsolateral prefrontal cortex; GM, gray matter; MNI, Montreal Neurological Institute; OFC, orbitofrontal cortex.

^aObtained with SPM5; corrected for age and total brain tissue.

^bThe *P* values are from the respective SPM5 analysis (Wellcome Department of Cognitive Neurology, London, England; <http://www.fil.ion.ucl.ac.uk/spm>), false discovery rate corrected.

then conducted to test associations between GMV and lifetime years of cocaine use (in the CUD sample [*n* = 40], small-volume correction was used⁵¹). Lifetime years of alcohol and cigarette use was evaluated in the whole sample (*n* = 82). All SPM5 analyses were performed controlling for age and total brain tissue, with extent threshold of 100 voxels and a threshold set at *P* < .05, corrected with a false discovery rate equivalent to a *T* threshold of 3.3. Labels for the resulting coordinates were inspected by means of software (Anatomy Toolbox; Institute of Neuroscience and Medicine, Jülich, Germany) and a coplanar stereotactic atlas of the human brain.⁵²

The voxels of interest were extracted with SPM5 EasyROI toolbox (http://www.sbirc.ed.ac.uk/cybil/cp_download.html) with an isotropic volume of the whole cluster around the significant peak voxel coordinates of the main effect results (CUD < CON from **Table 2**). This approach resulted in raw GMV values for each participant in each of these regions, allowing the measures to be used for figures and in SPSS⁵⁰ to conduct general linear model analysis, covarying for total brain tissue, age, race, verbal intelligence, and BDI symptoms (as documented in the "Results" section). These SPSS analyses were Bonferroni corrected for the 5 main effect regions, making the CUD × *MAOA* interaction results significant at *P* < .01. To understand the contribution to variability in GMV of all the variables studied and the potentially unique effects of long-term drug use, we used the voxels of interest in SPSS to conduct multiple regression analysis on each of the main effect coordinates. The model consisted of 2 hierarchical blocks: in the first block we entered total brain tissue, age, race, verbal intelli-

gence, BDI, and *MAOA*. In the second block, we entered the lifetime drug use variables.

RESULTS

CHARACTERISTICS OF COCAINE ADDICTION

Individuals with CUD were significantly older than the CON group (mean [SEM] age, 45 [1] vs 39 [1] years), with no genotype effects (*P* = .28-.67). Additional differences included race (fewer whites in the CUD group than the CON group) and higher depression symptom score in CUD (9 [2]) than CON (3 [1]), with no genotype effects (*P* = .53-.89) and lower verbal intelligence (CUD, 90 [2]; CON, 97 [2]) and an interaction with the CUD-H group having lower scores than the CON-H group (*P* < .002). There were no differences between the groups in years of education and socioeconomic status⁵³ (Table 1).

In terms of drug exposure factors, all participants with CUD used cocaine (smoked crack) in the past 0 to 7 days before imaging and met DSM-IV⁵⁴ criteria for current cocaine dependence. The participants with CUD reported use of a mean (SEM) of 1.7 (0.8) g of cocaine per occasion with no genotype effects (*P* = .82). The years of lifetime cocaine use was 19 (1), with no genotype effects (*P* = .97). The age at CUD onset was 26 (1) years, and participants with CUD-L tended to be younger at onset (by approximately 4 years; *P* = .09, 2-tailed) than those with CUD-H. In addition to long-term cocaine use, the CUD sample also had a substantial lifetime use of cigarettes and alcohol. A larger proportion of individuals with CUD (30 [75%]) than CON (9 [21%]) reported cigarette smoking, with no difference in the number of cigarettes smoked per day (CUD, 8 [1]; CON, 5 [2]) and with no genotype effects (*P* = .32). In addition, 70% of the CUD group were also diagnosed as having alcohol abuse; their age at onset was 16 (1) years and they consumed 60 (8) ounces per occasion, with no genotype effects (*P* = .21).

GM EFFECTS OF COCAINE ADDICTION

Total GMV was reduced with greater age across all participants (*r* = -0.30, *P* = .007) with no diagnosis or genotype effects (*P* = .85), and there were no main effects and no interactions in total WMV (*P* = .21). Controlling for age and total brain tissue, individuals with CUD had GMV reductions in the left OFC (Brodmann area [BA] 11) ($F_{1,72} = 6.5$; *P* = .002), right DLPFC (BA 9) ($F_{1,72} = 27.5$; *P* = .001), temporal cortex (BA 37) ($F_{1,72} = 5.3$; *P* = .02), and hippocampus and parahippocampal gyrus ($F_{1,72} = 8.6$; *P* = .002) compared with CON (CUD < CON; Table 2, **Figure 1**). The *F* values in parentheses throughout the "Results" section represent the main effects of addiction after controlling for the potential influences of total brain tissue, age, race, verbal intelligence, and BDI symptoms.

At this SPM threshold (*P* < .05, false discovery rate corrected), there were no regions of increased GMV in CUD compared with CON and no main effects of genotype as assessed with *MAOA***L* > or < *MAOA***H* contrasts. However, there was a significant CUD × *MAOA* interaction effect exclusively in the OFC ($F_{1,68} = 5.2$; *P* = .003).

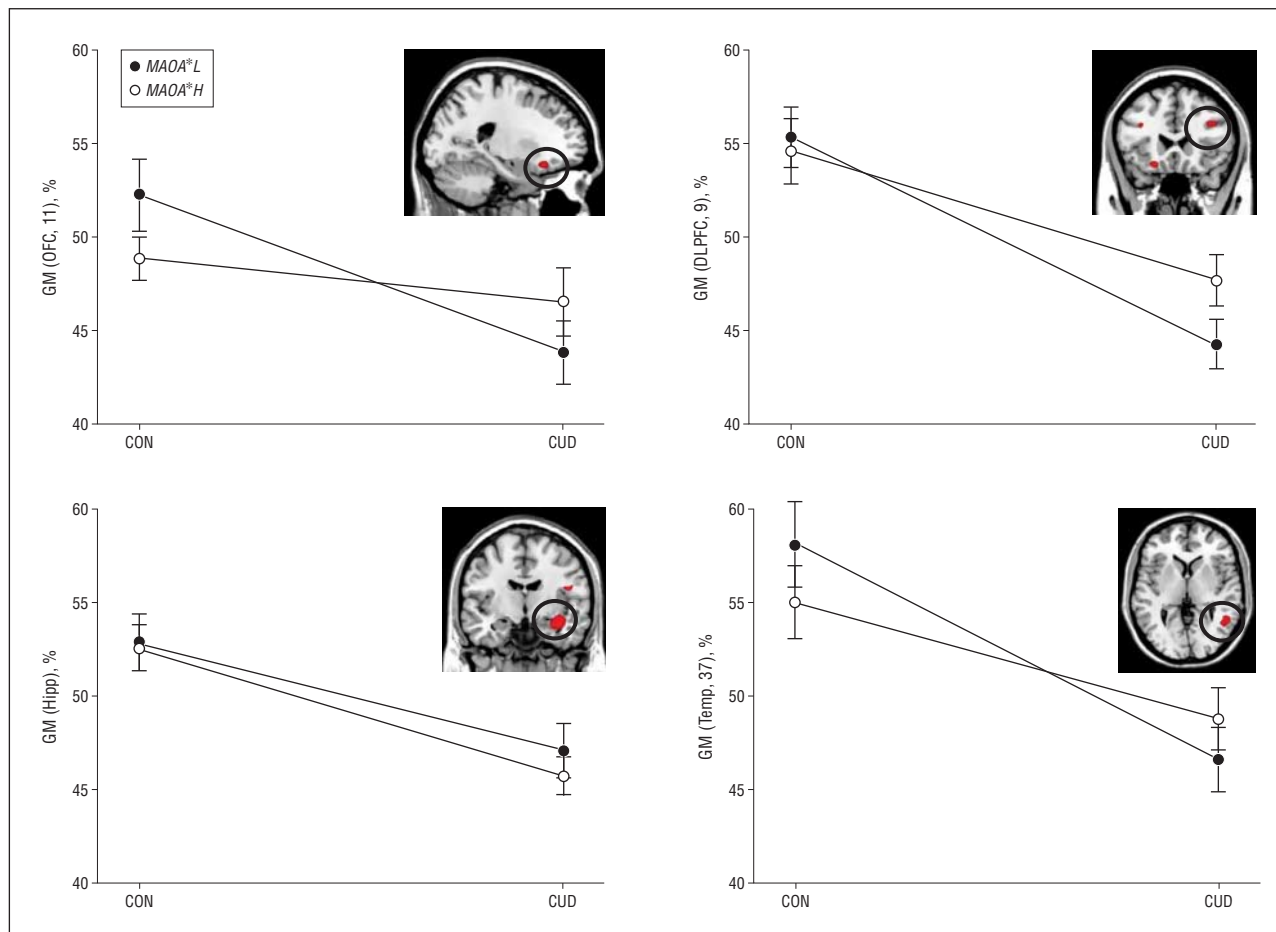


Figure 1. Gray matter (GM) volume reductions as a function of cocaine addiction (CUD < CON, 82 individuals; CUD indicates cocaine use disorders, and CON, controls). Each brain region (DLPFC, dorsolateral prefrontal cortex; Hipp, hippocampus; OFC, orbitofrontal cortex; Temp, temporal; numbers in parentheses are Brodmann areas) is presented with a graph using the voxels of interest to show that the main effects of addiction are contributed by both genotype groups (except for the OFC). The y-axis units display the percentage of GM volume in the cluster around the peak coordinates listed in Table 2. Error bars represent standard error of the mean. The GM volume map in each of the graphs shows the clusters of significance between the diagnostic groups ($P < .05$, false discovery rate corrected, 100 voxels minimum). The parahippocampus is not shown because its values were identical to the hippocampus volumes of interest, as they came from the same cluster. MAOA*H and MAOA*L indicate high- and low-repeat monoamine oxidase A genotype, respectively.

GENE \times DISEASE INTERACTION

After examining our planned contrasts and to investigate the source of the gene \times disease interaction effect in the OFC, we matched the CUD and CON participants on allele variation (Table 2). Comparing CUD-H with CON-H (Figure 2, blue) demonstrated a diagnosis effect of GMV reductions in the hippocampus; however, this contrast did not produce significant results in any of the other main effect regions, including the OFC, even at a reduced threshold. Comparing CUD-L with CON-L (Figure 2, red) showed robust GMV reductions in the OFC, DLPFC, temporal cortex, and hippocampus, similar to the main effects of addiction. Here, however, the results not only included the OFC between the anterior branches of the medial and lateral orbital sulci (BA 11) but also encompassed the medial edge of the orbital surface, ie, gyrus rectus (Table 2). The general linear model SPSS analyses using our voxels of interest in these OFC coordinates in all participants, and controlling for the covariates as listed earlier, showed that the CUD-L group had significantly less GMV than the CUD-H group and both CON groups in the left OFC (MAOA \times CUD; $F_{1,68} = 4.2$; $P = .007$) and bilateral gyrus rec-

tus (MAOA \times CUD; left, $F_{1,68} = 10.6$, $P = .002$; right, $F_{1,68} = 14.8$, $P = .001$) (Figure 2). This interaction was unique to the OFC (all other voxels of interest in Table 2, MAOA \times CUD, $P = .10-.76$).

LIFETIME DRUG USE AND OTHER VARIABLES

To understand the contribution of drug use duration in this sample, we conducted multiple regressions in SPM5 of GMV with years of drug use, controlling for age and total brain volume. In the CUD group ($n = 40$), with increasing years of cocaine exposure, there were more volume reductions in the OFC ($r = -0.44$, $P = .003$), DLPFC ($r = -0.41$, $P = .008$), and hippocampus ($r = -0.46$, $P = .003$); a similar pattern of results was obtained in the CUD group with lifetime alcohol (all $r = -0.34$ to -0.65 , $P = .008-.001$) and with cigarette smoking (all $r = -0.31$ to -0.52 , $P = .008-.001$) (Table 3, SPM results). In Figure 3, the whole-brain correlation results of all 3 drugs were overlaid, showing a visible overlap of the detrimental effects of all drugs on the hippocampus.

To understand the contribution of all the variables studied and the unique effects of long-term drug use, we conducted hierarchical regression analyses in SPSS. As

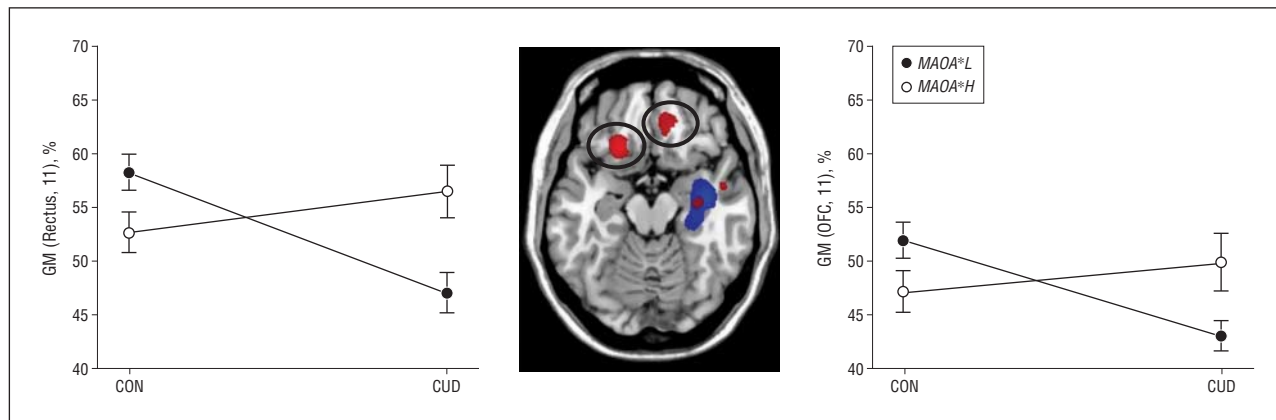


Figure 2. Gene \times disease interaction in the orbitofrontal cortex (OFC; numbers in parentheses are Brodmann areas). The gray matter (GM) volume measures in CUD-L $<$ CON-L (red) and CUD-H $<$ CON-H (blue) are overlaid on the SPM5 canonical template (Wellcome Department of Cognitive Neurology, London, England; <http://www.fil.ion.ucl.ac.uk/spm>) (CUD indicates cocaine use disorders, and CON, controls; H and L refer to high- and low-repeat monoamine oxidase A gene, *MAOA*, genotype, respectively). The respective interaction graphs show regional GM volume differences between the groups, in which the individuals in the CUD-L group have less GM than those in the CUD-H group and both CON groups. Error bars represent standard error of the mean. The y-axis units display the percentage of GM volume cluster around the peak coordinates in Table 2 ($P < .05$, false discovery rate corrected, 100 voxels minimum).

Table 3. Multiple Regression Analyses With GMV and Lifetime Drug Use^a

Region, BA	MNI Coordinates Peak Voxel			z Score	Cluster Size, mm ³	SPM5 P Value ^b
	x	y	z			
Cocaine Use, Lifetime y						
OFC, 11	24	38	-19	3.35	845	.04 ^c
DLPFC, 46	51	35	-8	2.95	430	.13
Hippocampus	25	-4	-18	3.53	1088	.01 ^c
Alcohol Use, Lifetime y						
DLPFC, 9	-36	-21	30	3.58	919	.02
Temporal, 20	44	-8	-17	3.75	4878	.02
Hippocampus	29	-18	-16	4.25	15 481	.01
Cigarettes, Lifetime y						
Hippocampus	33	-6	-22	4.62	603	.03

Abbreviations: BA, Brodmann area; DLPFC, dorsolateral prefrontal cortex; GMV, gray matter volume; MNI, Montreal Neurological Institute; OFC, orbitofrontal cortex.

^aCorrected for age and total brain tissue.

^bThe *P* values are from the respective SPM5 analysis (Wellcome Department of Cognitive Neurology, London, England; <http://www.fil.ion.ucl.ac.uk/spm>), false discovery rate corrected.

^cSmall volume correction was used.

documented in **Table 4**, total GM tissue was not significantly affected by any of the variables except for the known effect of reduced total GMV with greater age. As for the OFC, the block 1 variables contributed 21% to the GMV variance (driven by the *MAOA* genotype, age, and race). The drug use variables accounted significantly for an additional 19% of unique variance to the OFC GMV. This effect was driven by lifetime cocaine use. In the DLPFC, lifetime alcohol and cocaine use contributed the most unique variability to GMV, adding 24% to the 17% that was explained by the block 1 variables). In the temporal cortex, race and alcohol use were most predictive of GM differences between the groups. Results for the hippocampus were the most striking, showing that lifetime alcohol use contributed 30% of unique variance. Notably, in the hippocampus

and DLPFC, alcohol and cocaine use contributed more variability than the block 1 variables.

COMMENT

These findings demonstrate a distributed pattern of GMV loss in participants with CUD compared with CON in the OFC, DLPFC, temporal, and hippocampal regions. Exclusively in the OFC, GMV reductions were driven by increasing years of cocaine use and by individuals in the CUD-L group having smaller GMV, showing a gene \times disease interaction. The pattern of GMV in other regions was not affected by the genotype; rather, GMV loss in the temporal region and especially the DLPFC and hippocampus was driven primarily by drug use, especially by alcohol use.

REDUCED GMV IN COCAINE ADDICTION

Participants with CUD had reduced GMV in the right dorsolateral region of the prefrontal cortex, in BA 9, a region critical for monitoring information in working memory and in the controlled retrieval of information.⁵⁵ Specifically in CUD, these regions showed a deficit in functional activation during a go/no-go task, and deficits in these regions were associated with poor inhibitory control.⁵⁶ With the use of measures of cortical thickness, this precise DLPFC region was found to be thinner in participants with CUD than well-matched CON participants.⁸ Additional GMV reductions were found in this study in the inferior posterior temporal cortex, BA 37, associated with object naming and recognition memory, and found to have reduced GMV in opiate-dependent individuals.⁵⁷ This temporal region is particularly sensitive to age-dependent damage in Alzheimer disease.⁵⁸ This region is located immediately adjacent to the posterior parahippocampal gyrus and the hippocampus, also found to have reduced GMV in those with CUD compared with CON in this study. The hippocampus plays a role in extinction of currently nonrelevant but previously rewarding stimuli and in retrieval of information

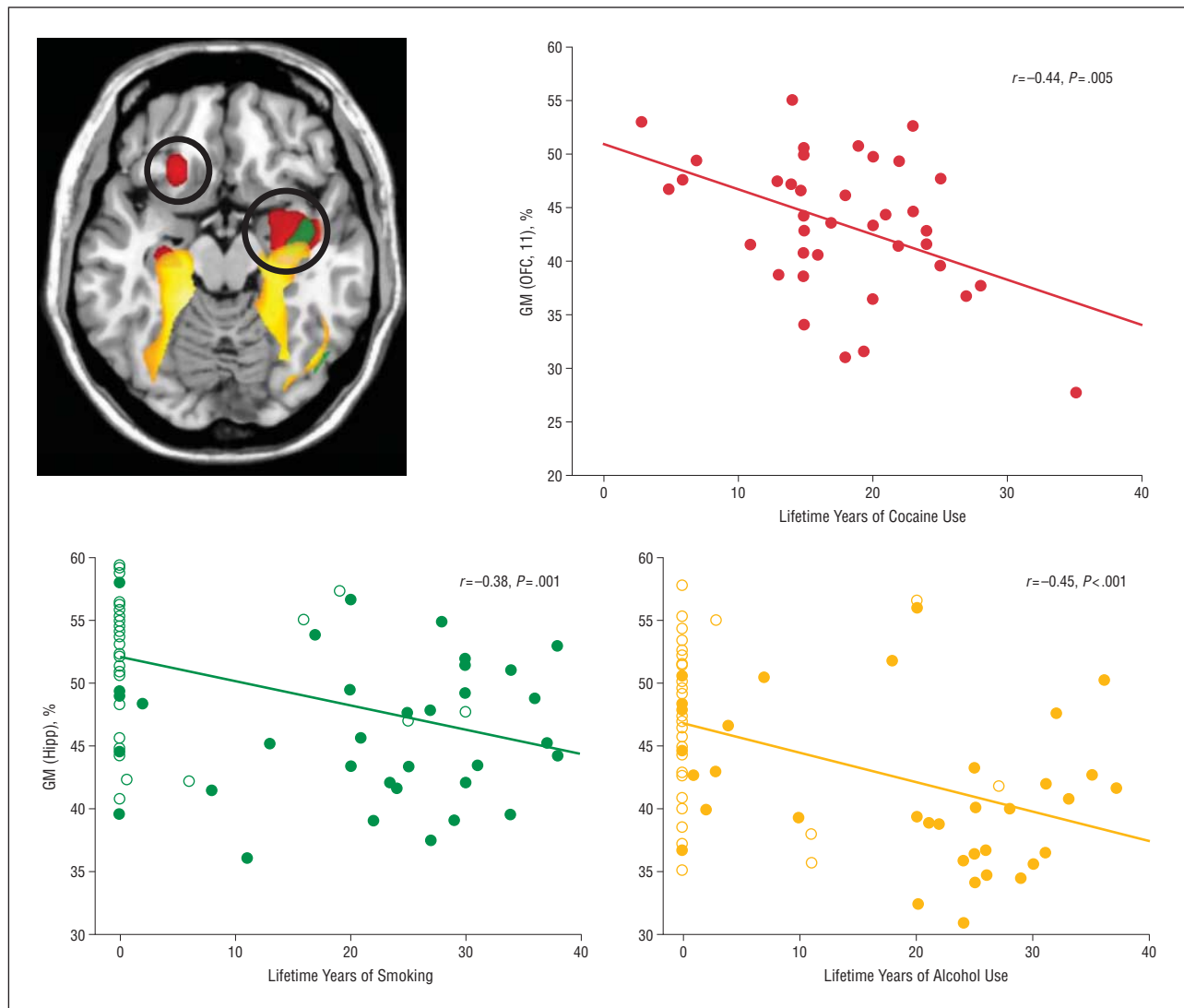


Figure 3. Lifetime effects of drug use on gray matter (GM) volume. The image shows correlation of GM volume with lifetime use of each drug (cocaine, red; alcohol, yellow; smoking, green) overlaid on the SPM5 canonical template (Wellcome Department of Cognitive Neurology, London, England; <http://www.fil.ion.ucl.ac.uk/spm>). The respective scatterplots are also overlaid with the correlations of GM volume (y-axis) and lifetime years of cocaine use in the cocaine use disorders group (red) and lifetime years of alcohol use (yellow) and smoking (green) in all participants (open circles represent controls), with the respective slope ($P < .001$, uncorrected, 100 voxels minimum). Hipp indicates hippocampus, and OFC, orbitofrontal cortex; number in parentheses is Brodmann area.

pertinent to these learning mechanisms; as such, the hippocampus is implicated in drug-context memory and in relapse to drug-seeking behaviors.^{59,60} Together with the hippocampus, the regions found to have reduced GMV in CUD in the current study are associated with drug-craving⁶¹ and drug-seeking⁶⁰ behaviors. Because the hippocampus, in concert with DLPFC regions, has an important executive role in inhibiting previously acquired drug reward mechanisms,⁶² these GMV decrements may perpetuate the impaired response inhibition and salience attribution syndrome in drug addiction.²

The neurochemistry of these affected brain regions is modulated by tonic and phasic DA action.^{1,63,64} In humans, the *in vivo* concentration of DA receptors is related to neural volume, as demonstrated by a recent imaging study showing a voxel-wise relationship between DA D₂ receptor availability (positron emission tomography with fallypride labeled with fluorine 18) and GMV in the DLPFC (BA 6 and 9) and temporal and parahippocampal gyrus,⁶⁵

regions that were found to have reduced GMV in CUD in the present study. Medium spiny neurons are the principal targets of DA terminals, and DA depletion in animal studies results in neurons with shorter and fewer spines compared with nonexposed neurons.⁶⁶ Because long-term drug use and addiction are associated with reduced DA D₂ receptor availability,^{67,68} neuronal volume is predicted to be similarly reduced, as evident especially in prefrontal cortical DA projections from the ventral tegmental area.^{69,70} Studies in humans found a reduction of *N*-acetylaspartate (suggested as a putative marker for neuronal cell loss or damage) concentrations in CUD and increased levels of myoinositol (a marker of glial activation) in frontal cortical regions.⁷¹

The present results demonstrate reduced volume of the OFC in the left hemisphere, whereas the rest of the main effect regions were right lateralized. These results may support the notion of a disrupted regional lateralization in drug addiction,⁷² which is posited to be inherited⁸;

Table 4. Contribution of Demographic, Genetic, and Drug Use Variables to GMV^a

Multiple Regression	Total GMV	OFC	DLPFC, BA 9	Temporal, BA 37	Hippocampus
Block 1 ^b	0.08 (5,63), .44	0.21 (6,62), .02	0.17 (6,62), .02	0.24 (6,62), .001	0.25 (6,62), .008
Block 2 ^c	0.02 (3,59), .79	0.19 (3,59), .004	0.24 (3,59), .001	0.12 (3,59), .05	0.30 (3,59), <.001
Age	-0.20	0.20 ^d	0.02	0.05	-0.20
Race	0.07	0.25 ^d	0.19	0.31 ^e	0.14
Volume	NA	-0.08	0.07	0.07	-0.17
WRAT-3	0.08	-0.16	-0.11	0.00	-0.05
BDI	0.00	0.02	0.04	-0.02	-0.16
<i>MAOA</i> genotype	0.06	0.43 ^e	0.06	-0.02	-0.06
Lifetime cocaine use	-0.05	-0.40 ^e	-0.26 ^d	-0.26	-0.20
Lifetime alcohol use	-0.06	-0.04	-0.55 ^e	-0.44 ^e	-0.59 ^e
Lifetime smoking	-0.12	0.08	-0.13	-0.17	-0.20

Abbreviations: BA, Brodmann area; BDI, Beck Depression Inventory; DLPFC, dorsolateral prefrontal cortex; GMV, gray matter volume; *MAOA*, monoamine oxidase A gene; NA, not applicable; OFC, orbitofrontal cortex; WRAT-3, Wide-Range Achievement Test, third edition.³³

^aUnless otherwise indicated, values are standardized beta coefficients.

^bValues are adjusted R^2 , df , and P value (contains the variables age, race, total brain tissue, WRAT-3, BDI, and *MAOA* genotype).

^cValues are adjusted R^2 , Δ (change), df , and P value (contains the block 1 variables and block 2 variables plus years of cocaine use, alcohol abuse, and cigarette smoking with the corresponding standardized β coefficients).

^d $P < .05$.

^e $P < .001$.

it may start developing before disease onset and may indeed contribute to its onset and progression together with the influence of particular traits, such as impulsivity.⁷³

GENE \times DISEASE INTERACTION IN THE OFC

In this study, the CUD-L group had significantly smaller volume than the CUD-H group and both CON groups in the OFC and gyrus rectus (BA 11). The OFC has been implicated in a wide variety of externalizing behavior disorders, and patients with specific damage to the OFC demonstrate more impulsive behavior than patients with other prefrontal damage.^{74,75} The anterior part of the OFC consists of eulaminate (6-layer) cortex, including granular layer IV.⁷⁶ Neurons in the OFC BA 11 of the macaque monkey code novelty, with rapid habituation,⁷⁷ and BA 11 is strongly linked with DLPFC areas (also found in this study to have reduced GMV in CUD), which together may guide goal-directed motivation.⁷⁸ The projections from the OFC to the entorhinal cortex, which innervates the pyramidal cells of the hippocampus, may underlie the process through which information about the emotional significance of stimuli is remembered.⁷⁹ Limited GMV in the OFC may undermine its functional connections with dorsolateral and entorhinal regions, thereby impairing the ability to make advantageous decisions.^{69,80} Supporting poor connectivity is a study finding disruption in WM fiber tracts to the OFC in CUD, which may further impair the OFC connectivity to the DLPFC and hippocampus regions.⁸¹ The regional GM loss we documented herein may correspond to WM loss, which is more reliably documented in manual segmentation or diffusion tensor imaging studies than VBM.⁴⁵

The selectivity of *MAOA* on DA degradation is not entirely known because *MAOA* influences other neurotransmitters that may affect GM.⁸² Although there is pharmacologic evidence that serotonin levels are enhanced after *MAOA* inhibition, immunohistochemical and autoradiographic studies have established that *MAOA* is predominantly localized in catecholaminergic neurons.⁸³ The se-

lectivity of *MAOA* specifically on DA degradation may also be relevant during prenatal development, when *MAOA* is crucial for catabolic degradation of DA, norepinephrine, and perhaps also serotonin.⁸⁴ Indeed, recent studies have shown that *MAO* (A and B) regulates neural progenitor cells during brain development, an effect mediated through serotonin.⁸⁵ Dopamine depletion in adults, as reliably documented in CUD,³ can trigger large-scale gene expression changes through multiple regulatory subunit changes in messenger RNA expression levels.⁸⁶ Although the *MAOA* uVNTR polymorphism analyzed in this study is not directly indicative of brain *MAOA* activity,⁸⁷ this genetic variant was linked to the differences in levels of the DA metabolite homovanillic acid in CSF.⁸⁸

It remains unknown whether the mechanisms by which decreased transcriptional activity of *MAOA* might increase GM in the OFC in healthy controls²⁸ but interact with cocaine use to selectively diminish OFC in the present study. The modulating effect of the *MAOA* genotype on structural variability may have started during early brain development, clearly before disease onset, and possibly continued its effect at adolescence at onset of the disease process. Interestingly, the CUD-L group in this study had a slightly younger age at onset of cocaine use. It is possible that these individuals who later developed CUD had reduced GMV in the OFC before disease onset because developmental factors, such as maternal smoking, are associated with increased likelihood of drug experimentation and decreased thickness of the OFC in adolescence.⁸⁹ In this context, it is noteworthy that the *MAOA**L genotype was associated with risk of alcoholism and antisocial alcoholism.³¹ It is also noteworthy that other factors in addition to the *MAOA* polymorphism affect the enzyme's expression. In a recent article, our group demonstrated that the *MAOA* gene is subjected to epigenetic modifications.⁹⁰ This finding, together with the well-established evidence that the drugs of abuse cause epigenetic aberrations,⁹¹ led us to propose that the *MAOA* methylation pattern in CUD might be influenced by drug use, causing dysregulation of its expression.

Gray matter in the OFC, showing deficit in CUD-L, was uniquely driven by increasing years of cocaine exposure. In fact, the OFC was the only region affected specifically by cocaine and not years of alcohol use. It is possible that the OFC of individuals with CUD-L is more sensitive to the neurotoxic effects of cocaine than that of individuals with CUD-H exposed to similar amounts of the drug. Supporting this specificity is evidence from studies in rats showing that long-term stimulants limit spine density in the OFC (while long-term opiate use may increase spine density),^{92,93} perhaps making the OFC sensitive to morphologic changes depending on the drug of abuse.⁹⁴ Additional morphometric damage can be caused by smoking exposure because long-term smoking inhibits MAO_A⁹⁵ and high-affinity nicotinic receptors in the human OFC increase after smoking.⁹⁶ A recent VBM study showed that GMV in DLPFC and inferior frontal regions is reduced in cigarette smokers.⁹⁷ However, consistent with the current results, nicotine administration to adolescent rats elicited less severe region-dependent effects than alcohol.⁹⁸

REGION-SPECIFIC EFFECTS OF LIFETIME ALCOHOL USE

Lifetime alcohol use was the major contributor of GMV deficit in the DLPFC, temporal cortex, and hippocampus of participants with CUD, contributing unique variability to GMV above and beyond the MAOA polymorphism and any of the other factors tested, more so than cocaine and cigarette smoking. In this study, we measured severity as the number of lifetime years of use. Animal models of binge alcohol administration, controlling for severity in a dose-dependent manner, support a direct link between high levels of alcohol consumption and neurotoxic effects in the hippocampus and surrounding dentate gyrus and associated entorhinal-perirhinal cortex during adolescence.⁹⁸ Similarly, reduced hippocampal volume was found among adolescents with alcohol use disorders.^{99,100} Gray matter loss in the hippocampus may lead to more drug seeking, as demonstrated by animal studies showing that blocking neurogenesis in the adult rat hippocampus caused increased cocaine seeking and more self-administration,¹⁰¹ further facilitating a vicious cycle of cocaine use.¹⁰¹ The observed GMV reductions in the hippocampus, perhaps due to chronic alcohol use, may increase cocaine use through strong resistance to extinction of drug-seeking behavior.¹⁰¹

CAVEATS

Our groups differed in age, ethnicity, verbal intelligence, and symptoms of depression. Demographic effects of difference in the lifetime trajectory of drug addiction are a source of variability and a contributor to the overall impact of the disease.¹⁰² Lower verbal intelligence could indicate compromised education due to drug use during adolescence (note that the differences due to genotype are partly supported by another study¹⁰³). The BDI measure (reflecting symptoms in the past 2 weeks) cannot be separated from drug effects (such as acute withdrawal).¹⁰⁴ Rather than excluding these effects, we studied their impact in explaining GMV differences between the

groups, enhancing the generalizability of the current results.²⁰ On the subject of enhancing generalizability, it is important to remember that our findings come from a male sample; women are largely understudied in drug addiction, a limitation of generalizability that needs to be addressed in future studies. Our sample of individuals with CUD also underrepresented whites compared with African Americans, and the latter show GM effects in the OFC and temporal cortex. This represents a confounding factor in this study, but it also highlights the evidence of racial differences in GMV that need to be accounted for beyond the clinical variable of interest. Indeed, in this study we demonstrated through hierarchical regression analysis that the MAOA and cocaine use effects contribute unique variability to GMV beyond other effects.

Similarly, additional factors affect GM reductions, including, for example, long-term lack of sleep (affecting the OFC)¹⁰⁵ and acute depression affecting hippocampal volume.¹⁰⁴ Both are common problems in CUD and should be further investigated in future studies. The present study had active, currently using participants with CUD (90% had urine positive for cocaine), and a case could be made for OFC reductions during acute use that may recover with abstinence. However, even after prolonged abstinence of 2 to 4 years, GM reductions were still found in comparison of substance-dependent individuals and controls, pointing to persistent and enduring GM deficits in the OFC.^{46,106}

In a VBM study in healthy control participants, MAOA**L* has had increased lateral OFC volume (BA 47) compared with MAOA**H*.²⁸ Conversely, in this study, the CUD-L group had significantly less GMV than CUD-H and both CON groups in the *medial* OFC and gyrus rectus. In the same previous study, healthy individuals with MAOA**L* had reduced GM encompassing the entire cingulate gyrus and particularly in the anterior cingulate, a region not evident in the current results. While inspecting CON-L vs CON-H in our data, we could find a similar pattern including the anterior cingulate, using $P < .05$, uncorrected (results not shown). Differences in findings may stem from the use of varied methods with varied populations of controls and individuals with CUD. Other morphology studies found deficit in regions in which we did not find reduced GMV (eg, amygdala,¹³ anterior cingulate,¹¹ and insula¹¹); conversely, none of the studies found the hippocampus GMV deficits that we found in this sample, although we studied CUD with comorbid alcohol abuse, which has been associated with hippocampal volume loss. Future studies should continue to assess genotype differences within CUD because this study suggests CUD-L to be associated with potentially more extensive deficits than CUD-H (eg, earlier age at onset is a major risk factor for a more severe course of illness).

CONCLUSIONS

The extensive GMV loss in the OFC, DLPFC, temporal, and hippocampal regions in individuals with CUD underlies demographic, genetic, and drug use factors. Exclusively in the OFC, GMV reductions were driven by increasing years of cocaine use and by individuals with

CUD-L having smaller GMV, showing a gene \times disease interaction. The population we studied had already started using drugs, which constrains the ability to track causes and effects of the substance abuse.^{8,21} Addiction liability can be characterized dimensionally among already affected individuals insofar as indexes of severity.³² These results suggest that loss of GMV among individuals with CUD is multidetermined and can be assessed with a model that includes genetic, behavioral, and drug use factors that we speculate have interacted continuously throughout the lifespan. Studies are emerging in support of this notion, that gene \times environment interactions take different forms at different ontogenic stages of development during the lifespan.^{32,107} Therefore, the next generation of neurogenetic studies will have to document complex interactions over protracted developmental trajectories to explain the effects contributing to multifaceted psychopathology as drug addiction.

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REFERENCES

- Volkow ND, Wang GJ, Fowler JS, Logan J, Gattley SJ, Hitzemann R, Chen AD, Dewey SL, Pappas N. Decreased striatal dopaminergic responsiveness in detoxified cocaine-dependent subjects. *Nature*. 1997;386(6627):830-833.
- Goldstein RZ, Volkow ND. Drug addiction and its underlying neurobiological basis: neuroimaging evidence for the involvement of the frontal cortex. *Am J Psychiatry*. 2002;159(10):1642-1652.
- Volkow ND, Fowler JS. Addiction, a disease of compulsion and drive: involvement of the orbitofrontal cortex. *Cereb Cortex*. 2000;10(3):318-325.
- Garavan H, Kaufman JN, Hester R. Acute effects of cocaine on the neurobiology of cognitive control. *Philos Trans R Soc Lond B Biol Sci*. 2008;363(1507):3267-3276.
- Goldstein RZ, Alia-Klein N, Tomasi D, Zhang L, Cottone LA, Maloney T, Telang F, Caparelli EC, Chang L, Ernst T, Samaras D, Squires NK, Volkow ND. Is decreased prefrontal cortical sensitivity to monetary reward associated with impaired motivation and self-control in cocaine addiction? *Am J Psychiatry*. 2007;164(1):43-51.
- London ED, Ernst M, Grant S, Bonson K, Weinstein A. Orbitofrontal cortex and human drug abuse: functional imaging. *Cereb Cortex*. 2000;10(3):334-342.
- Kaufman JN, Ross TJ, Stein EA, Garavan H. Cingulate hypoactivity in cocaine users during a GO-NOGO task as revealed by event-related functional magnetic resonance imaging. *J Neurosci*. 2003;23(21):7839-7843.
- Makris N, Gasic GP, Kennedy DN, Hodge SM, Kaiser JR, Lee MJ, Kim BW, Blood AJ, Evans AE, Seidman LJ, Iosifescu DV, Lee S, Baxter C, Perlis RH, Smoller JW, Fava M, Breiter HC. Cortical thickness abnormalities in cocaine addiction—a reflection of both drug use and a pre-existing disposition to drug abuse? *Neuron*. 2008;60(1):174-188.
- Liu X, Matochik JA, Cadet JL, London ED. Smaller volume of prefrontal lobe in poly-substance abusers: a magnetic resonance imaging study. *Neuropsychopharmacology*. 1998;18(4):243-252.
- Fein G, Di Sclafani V, Meyerhoff DJ. Prefrontal cortical volume reduction associated with frontal cortex function deficit in 6-week abstinent crack-cocaine dependent men. *Drug Alcohol Depend*. 2002;68(1):87-93.
- Franklin TR, Acton PD, Maldjian JA, Gray JD, Croft JR, Dackis CA, O'Brien CP, Childress AR. Decreased gray matter concentration in the insular, orbitofrontal, cingulate, and temporal cortices of cocaine patients. *Biol Psychiatry*. 2002;51(2):134-142.
- Matochik JA, London ED, Eldreth DA, Cadet JL, Bolla KI. Frontal cortical tissue composition in abstinent cocaine abusers: a magnetic resonance imaging study. *Neuroimage*. 2003;19(3):1095-1102.
- Makris N, Gasic GP, Seidman LJ, Goldstein JM, Gastfriend DR, Elman I, Albaugh MD, Hodge SM, Ziegler DA, Sheahan FS, Caviness VS Jr, Tsuang MT, Kennedy DN, Hyman SE, Rosen BR, Breiter HC. Decreased absolute amygdala volume in cocaine addicts. *Neuron*. 2004;44(4):729-740.
- Bartzokis G, Beckson M, Lu PH, Edwards N, Rapoport R, Wiseman E, Bridge P. Age-related brain volume reductions in amphetamine and cocaine addicts and normal controls: implications for addiction research. *Psychiatry Res*. 2000;98(2):93-102.
- Volkow ND, Fowler JS, Wang GJ, Hitzemann R, Logan J, Schlyer DJ, Dewey SL, Wolf AP. Decreased dopamine D2 receptor availability is associated with reduced frontal metabolism in cocaine abusers. *Synapse*. 1993;14(2):169-177.
- Taki Y, Kinomura S, Sato K, Goto R, Inoue K, Okada K, Ono S, Kawashima R, Fukuda H. Both global gray matter volume and regional gray matter volume negatively correlate with lifetime alcohol intake in non-alcohol-dependent Japanese men: a volumetric analysis and a voxel-based morphometry. *Alcohol Clin Exp Res*. 2006;30(6):1045-1050.
- Fein G, Landman B, Tran H, McGillivray S, Finn P, Barakos J, Moon K. Brain atrophy in long-term abstinent alcoholics who demonstrate impairment on a simulated gambling task. *Neuroimage*. 2006;32(3):1465-1471.
- Almeida OP, Garrido GJ, Lautenschlager NT, Hulse GK, Jamrozik K, Flicker L. Smoking is associated with reduced cortical regional gray matter density in brain regions associated with incipient Alzheimer disease. *Am J Geriatr Psychiatry*. 2008;16(1):92-98.
- Durazzo TC, Rothlind JC, Cardenas VA, Studholme C, Weiner MW, Meyerhoff DJ. Chronic cigarette smoking and heavy drinking in human immunodeficiency virus: consequences for neurocognition and brain morphology. *Alcohol*. 2007;41(7):489-501.
- Pennings EJ, Leccese AP, Wolff FA. Effects of concurrent use of alcohol and cocaine. *Addiction*. 2002;97(7):773-783.
- Vanyukov MM, Maher BS, Devlin B, Tarter RE, Kirillova GP, Yu LM, Ferrell RE. Haplotypes of the monoamine oxidase genes and the risk for substance use disorders. *Am J Med Genet B Neuropsychiatr Genet*. 2004;125(1):120-125.
- Fowler JS, MacGregor RR, Wolf AP, Arnett CD, Dewey SL, Schlyer D, Christman D, Logan J, Smith M, Sachs H, Aquilonius SM, Bjurling P, Halldin C, Hartvig P, Leenders KL, Lundqvist H, Orelund L, Staltnacke CG, Langstrom B. Mapping human brain monoamine oxidase A and B with 11C-labeled suicide inactivators and PET. *Science*. 1987;235(4787):481-485.
- Shih JC, Thompson RF. Monoamine oxidase in neuropsychiatry and behavior. *Am J Hum Genet*. 1999;65(3):593-598.
- Wakschlag LS, Kistner EO, Pine DS, Biesecker G, Pickett KE, Skol AD, Dukic V, Blair RJ, Leventhal BL, Cox NJ, Burns JL, Kasza KE, Wright RJ, Cook EH Jr. Interaction of prenatal exposure to cigarettes and MAOA genotype in pathways to youth antisocial behavior. *Mol Psychiatry*. 2010;15(9):928-937.
- Inoue-Murayama M, Mishima N, Hayasaka I, Ito S, Murayama Y. Divergence of ape and human monoamine oxidase A gene promoters: comparative analysis of polymorphisms, tandem repeat structures and transcriptional activities on reporter gene expression. *Neurosci Lett*. 2006;405(3):207-211.
- Vanyukov MM, Maher BS, Devlin B, Kirillova GP, Kirisci L, Yu LM, Ferrell RE. The MAOA promoter polymorphism, disruptive behavior disorders, and early onset substance use disorder: gene-environment interaction. *Psychiatr Genet*. 2007;17(6):323-332.
- Sabol SZ, Hu S, Hamer D. A functional polymorphism in the monoamine oxidase A gene promoter. *Hum Genet*. 1998;103(3):273-279.
- Meyer-Lindenberg A, Buckholz JW, Kolachana B, Hariri AR, Pezawas L, Blasi G, Wabnitz A, Honea R, Verchinski B, Callicott JH, Egan M, Mattay V, Weinberger DR. Neural mechanisms of genetic risk for impulsivity and violence in humans. *Proc Natl Acad Sci U S A*. 2006;103(16):6269-6274.
- Caspi A, McClay J, Moffitt TE, Mill J, Martin J, Craig IW, Taylor A, Poulton R. Role of genotype in the cycle of violence in maltreated children. *Science*. 2002;297(5582):851-854.
- Saito T, Lachman HM, Diaz L, Hallikainen T, Kauhanen J, Salonen JT, Ryyänänen OP, Karvonen MK, Syvälahti E, Pohjalainen T, Hietala J, Tiihonen J. Analysis of monoamine oxidase A (MAOA) promoter polymorphism in Finnish male alcoholics. *Psychiatry Res*. 2002;109(2):113-119.
- Samochowiec J, Lesch KP, Rottmann M, Smolka M, Sygailo YV, Okladnova O, Rommelspacher H, Winterer G, Schmidt LG, Sander T. Association of a regula-

- tory polymorphism in the promoter region of the monoamine oxidase A gene with antisocial alcoholism. *Psychiatry Res.* 1999;86(1):67-72.
32. Vanyukov MM, Kirisci L, Moss L, Tarter RE, Reynolds MD, Maher BS, Kirillova GP, Ridenour T, Clark DB. Measurement of the risk for substance use disorders: phenotypic and genetic analysis of an index of common liability. *Behav Genet.* 2009;39(3):233-244.
 33. Wilkinson G. *The Wide-Range Achievement Test 3: Administration Manual.* Wilmington, DE: Wide Range Inc; 1993.
 34. Wechsler D. *Wechsler Memory Scale Manual.* San Antonio, TX: Psychological Corp; 1987.
 35. Beck AT. *The Beck Depression Inventory (BD-II).* San Antonio, TX: Psychological Corp; 1996.
 36. McLellan AT, Kushner H, Metzger D, Peters R, Smith I, Grissom G, Pettinati H, Argeriou M. The fifth edition of the Addiction Severity Index. *J Subst Abuse Treat.* 1992;9(3):199-213.
 37. Ventura J, Liberman RP, Green MF, Shaner A, Mintz J. Training and quality assurance with the Structured Clinical Interview for DSM-IV (SCID-I/P). *Psychiatry Res.* 1998;79(2):163-173.
 38. Guo G, Ou XM, Roettger M, Shih JC. The VNTR 2 repeat in MAOA and delinquent behavior in adolescence and young adulthood: associations and MAOA promoter activity. *Eur J Hum Genet.* 2008;16(5):626-634.
 39. Deichmann R, Schwarzbauer C, Turner R. Optimisation of the 3D MDEFT sequence for anatomical brain imaging: technical implications at 1.5 and 3 T. *Neuroimage.* 2004;21(2):757-767.
 40. Lee JH, Garwood M, Menon R, Adriany G, Andersen P, Truwit CL, Uğurbil K. High contrast and fast three-dimensional magnetic resonance imaging at high fields. *Magn Reson Med.* 1995;34(3):308-312.
 41. Tardif CL, Collins DL, Pike GB. Sensitivity of voxel-based morphometry analysis to choice of imaging protocol at 3 T. *Neuroimage.* 2009;44(3):827-838.
 42. Ashburner J, Friston KJ. Voxel-based morphometry—the methods. *Neuroimage.* 2000;11(6, pt 1):805-821.
 43. Ashburner J, Friston KJ. Unified segmentation. *Neuroimage.* 2005;26(3):839-851.
 44. Cuadra MB, Cammoun L, Butz T, Cuisenaire O, Thiran JP. Comparison and validation of tissue modelization and statistical classification methods in T1-weighted MR brain images. *IEEE Trans Med Imaging.* 2005;24(12):1548-1565.
 45. Padovani A, Borroni B, Brambati SM, Agosti C, Broli M, Alonso R, Scifo P, Bellelli G, Alberici A, Gasparotti R, Perani D. Diffusion tensor imaging and voxel based morphometry study in early progressive supranuclear palsy. *J Neurol Neurosurg Psychiatry.* 2006;77(4):457-463.
 46. Tanabe J, Tregellas JR, Dalwani M, Thompson L, Owens E, Crowley T, Banich M. Medial orbitofrontal cortex gray matter is reduced in abstinent substance-dependent individuals. *Biol Psychiatry.* 2009;65(2):160-164.
 47. Szeszko PR, Christian C, MacMaster F, Lencz T, Mirza Y, Taormina SP, Easter P, Rose M, Michalopolou GA, Rosenberg DR. Gray matter structural alterations in psychotropic drug-naïve pediatric obsessive-compulsive disorder: an optimized voxel-based morphometry study. *Am J Psychiatry.* 2008;165(10):1299-1307.
 48. Pannacciulli N, Del Parigi A, Chen K, Le DS, Reiman EM, Tataranni PA. Brain abnormalities in human obesity: a voxel-based morphometric study. *Neuroimage.* 2006;31(4):1419-1425.
 49. O'Brien LM, Ziegler DA, Deutsch CK, Kennedy DN, Goldstein JM, Seidman LJ, Hodge S, Makris N, Caviness V, Frazier JA, Herbert MR. Adjustment for whole brain and cranial size in volumetric brain studies: a review of common adjustment factors and statistical methods. *Harv Rev Psychiatry.* 2006;14(3):141-151.
 50. Stevens J. *Applied Multivariate Statistics for the Social Sciences.* 2nd ed. Mahwah, NJ: Lawrence Erlbaum Assoc; 1992.
 51. Worsley KJ, Marrett S, Neelin P, Vandal AC, Friston KJ, Evans AC. A unified statistical approach for determining significant signals in images of cerebral activation. *Hum Brain Mapp.* 1996;4(1):58-73.
 52. Talairach J, Tournoux P. *Co-Planar Stereotaxic Atlas of the Human Brain.* New York, NY: Thieme Medical Publishers, Inc; 1988.
 53. Hollingshead AB. *Four-Factor Index of Social Status.* New Haven, CT: Yale University; 1975.
 54. First MB, Spitzer RL, Gibbon M, Williams J. *Structured Clinical Interview for DSM-IV Axis I Disorders—Patient Edition (SCID-I/P, Version 2.0).* New York: Biometrics Research Dept, New York State Psychiatric Institute; 1996.
 55. MacLeod AK, Buckner RL, Miezin FM, Petersen SE, Raichle ME. Right anterior prefrontal cortex activation during semantic monitoring and working memory. *Neuroimage.* 1998;7(1):41-48.
 56. Hester R, Garavan H. Executive dysfunction in cocaine addiction: evidence for discordant frontal, cingulate, and cerebellar activity. *J Neurosci.* 2004;24(49):11017-11022.
 57. Lyoo IK, Pollack MH, Silveri MM, Ahn KH, Diaz CI, Hwang J, Kim SJ, Yurgelun-Todd DA, Kaufman MJ, Renshaw PF. Prefrontal and temporal gray matter density decreases in opiate dependence. *Psychopharmacology (Berl).* 2006;184(2):139-144.
 58. Thangavel R, Sahu SK, Van Hoesen GW, Zaheer A. Modular and laminar pathology of Brodmann's area 37 in Alzheimer's disease. *Neuroscience.* 2008;152(1):50-55.
 59. Fuchs RA, Evans KA, Ledford CC, Parker MP, Case JM, Mehta RH, See RE. The role of the dorsomedial prefrontal cortex, basolateral amygdala, and dorsal hippocampus in contextual reinstatement of cocaine seeking in rats. *Neuropsychopharmacology.* 2005;30(2):296-309.
 60. Vorel SR, Liu X, Hayes RJ, Spector JA, Gardner EL. Relapse to cocaine-seeking after hippocampal theta burst stimulation. *Science.* 2001;292(5519):1175-1178.
 61. Kilts CD, Schweitzer JB, Quinn CK, Gross RE, Faber TL, Muhammad F, Ely TD, Hoffman JM, Drexler KP. Neural activity related to drug craving in cocaine addiction. *Arch Gen Psychiatry.* 2001;58(4):334-341.
 62. Levy D, Shabat-Simon M, Shalev U, Barnea-Ygael N, Cooper A, Zangen A. Repeated electrical stimulation of reward-related brain regions affects cocaine but not "natural" reinforcement. *J Neurosci.* 2007;27(51):14179-14189.
 63. Volkow ND, Wang GJ, Fowler JS, Thanos PP, Logan J, Gatley SJ, Gifford A, Ding YS, Wong C, Pappas N. Brain DA D2 receptors predict reinforcing effects of stimulants in humans: replication study. *Synapse.* 2002;46(2):79-82.
 64. Volkow ND, Wang GJ, Fowler JS, Logan J, Gatley SJ, Wong C, Hitzemann R, Pappas NR. Reinforcing effects of psychostimulants in humans are associated with increases in brain dopamine and occupancy of D₂ receptors. *J Pharmacol Exp Ther.* 1999;291(1):409-415.
 65. Woodward ND, Zald DH, Ding Z, Riccardi P, Ansari MS, Baldwin RM, Cowan RL, Li R, Kessler RM. Cerebral morphology and dopamine D₂/D₃ receptor distribution in humans: a combined [¹⁸F]fallypride and voxel-based morphometry study. *Neuroimage.* 2009;46(1):31-38.
 66. Meredith GE, Ypma P, Zahm DS. Effects of dopamine depletion on the morphology of medium spiny neurons in the shell and core of the rat nucleus accumbens. *J Neurosci.* 1995;15(5, pt 2):3808-3820.
 67. Volkow ND, Fowler JS, Wang GJ. The addicted human brain viewed in the light of imaging studies: brain circuits and treatment strategies. *Neuropharmacology.* 2004;47(suppl 1):3-13.
 68. Volkow ND, Fowler JS, Logan J, Gatley SJ, Dewey SL, MacGregor RR, Schlyer DJ, Pappas N, King P, Wang G-J, Wolf AP. Carbon-11-cocaine binding compared at subpharmacological and pharmacological doses: a PET study. *J Nucl Med.* 1995;36(7):1289-1297.
 69. Haber SN, Knutson B. The reward circuit: linking primate anatomy and human imaging. *Neuropsychopharmacology.* 2010;35(1):4-26.
 70. Du C, Yu M, Volkow ND, Koretsky AP, Fowler JS, Benveniste H. Cocaine increases the intracellular calcium concentration in brain independently of its cerebrovascular effects. *J Neurosci.* 2006;26(45):11522-11531.
 71. Chang L, Ernst T, Strickland T, Mehlinger CM. Gender effects on persistent cerebral metabolite changes in the frontal lobes of abstinent cocaine users. *Am J Psychiatry.* 1999;156(5):716-722.
 72. Hanlon CA, Wesley MJ, Roth AJ, Miller MD, Porrino LJ. Loss of laterality in chronic cocaine users: an fMRI investigation of sensorimotor control. *Psychiatry Res.* 2010;181(1):15-23.
 73. Hill SY, Wang S, Kostelnik B, Carter H, Holmes B, McDermott M, Zezza N, Stiffler S, Keshavan MS. Disruption of orbitofrontal cortex laterality in offspring from multiplex alcohol dependence families. *Biol Psychiatry.* 2009;65(2):129-136.
 74. Bechara A. The role of emotion in decision-making: evidence from neurological patients with orbitofrontal damage. *Brain Cogn.* 2004;55(1):30-40.
 75. Matsuo K, Nicoletti M, Nemoto K, Hatch JP, Peluso MA, Nery FG, Soares JC. A voxel-based morphometry study of frontal gray matter correlates of impulsivity. *Hum Brain Mapp.* 2009;30(4):1188-1195.
 76. Hof PR, Mufson EJ, Morrison JH. Human orbitofrontal cortex: cytoarchitecture and quantitative immunohistochemical parcellation. *J Comp Neurol.* 1995;359(1):48-68.
 77. de Araujo IE, Rolls ET, Velazco MI, Margot C, Cayeux I. Cognitive modulation of olfactory processing. *Neuron.* 2005;46(4):671-679.
 78. Djordjevic J, Zatorre RJ, Petrides M, Boyle JA, Jones-Gotman M. Functional neuroimaging of odor imagery. *Neuroimage.* 2005;24(3):791-801.
 79. Kringelbach ML. The human orbitofrontal cortex: linking reward to hedonic experience. *Nat Rev Neurosci.* 2005;6(9):691-702.
 80. Bechara A, Damasio H. Decision-making and addiction (part I): impaired activation of somatic states in substance dependent individuals when pondering decisions with negative future consequences. *Neuropsychologia.* 2002;40(10):1675-1689.
 81. Lim KO, Choi SJ, Pomara N, Wolkin A, Rotrosen JP. Reduced frontal white matter integrity in cocaine dependence: a controlled diffusion tensor imaging study. *Biol Psychiatry.* 2002;51(11):890-895.

82. Bortolato M, Chen K, Shih JC. Monoamine oxidase inactivation: from pathophysiology to therapeutics. *Adv Drug Deliv Rev.* 2008;60(13-14):1527-1533.
83. Westlund KN, Denney RM, Rose RM, Abell CW. Localization of distinct monoamine oxidase A and monoamine oxidase B cell populations in human brainstem. *Neuroscience.* 1988;25(2):439-456.
84. Shih JC, Chen K, Ridd MJ. Role of MAO A and B in neurotransmitter metabolism and behavior. *Pol J Pharmacol.* 1999;51(1):25-29.
85. Cheng A, Scott AL, Ladenheim B, Chen K, Ouyang X, Lathia JD, Mughal M, Cadet JL, Mattson MP, Shih JC. Monoamine oxidases regulate telencephalic neural progenitors in late embryonic and early postnatal development. *J Neurosci.* 2010;30(32):10752-10762.
86. Meurers BH, Dziewczapolski G, Shi T, Bittner A, Kamme F, Shults CW. Dopamine depletion induces distinct compensatory gene expression changes in DARPP-32 signal transduction cascades of striatonigral and striatopallidal neurons. *J Neurosci.* 2009;29(21):6828-6839.
87. Fowler JS, Alia-Klein N, Kriplani A, Logan J, Williams B, Zhu W, Craig IW, Telang F, Goldstein R, Volkow ND, Vaska P, Wang GJ. Evidence that brain MAO A activity does not correspond to MAO A genotype in healthy male subjects. *Biol Psychiatry.* 2007;62(4):355-358.
88. Zalsman G, Huang YY, Harkavy-Friedman JM, Oquendo MA, Ellis SP, Mann JJ. Relationship of MAO-A promoter (u-VNTR) and COMT (V158M) gene polymorphisms to CSF monoamine metabolites levels in a psychiatric sample of Caucasians: a preliminary report. *Am J Med Genet B Neuropsychiatr Genet.* 2005;132(1):100-103.
89. Lotfipour S, Ferguson E, Leonard G, Perron M, Pike B, Richer L, Séguin JR, Toro R, Veillette S, Pausova Z, Paus T. Orbitofrontal cortex and drug use during adolescence: role of prenatal exposure to maternal smoking and *BDNF* genotype. *Arch Gen Psychiatry.* 2009;66(11):1244-1252.
90. Shumay E, Fowler JS. Identification and characterization of putative methylation targets in the MAOA locus using bioinformatic approaches. *Epigenetics.* 2010;5(4):325-342.
91. LaPlant Q, Nestler EJ. CRACKing the histone code: cocaine's effects on chromatin structure and function [published online June 4, 2010]. *Horm Behav.* doi: 10.1016/j.yhbeh.2010.05.015.
92. Jentsch JD, Taylor JR. Impulsivity resulting from frontostriatal dysfunction in drug abuse: implications for the control of behavior by reward-related stimuli. *Psychopharmacology (Berl).* 1999;146(4):373-390.
93. Crombag HS, Gorny G, Li Y, Kolb B, Robinson TE. Opposite effects of amphetamine self-administration experience on dendritic spines in the medial and orbital prefrontal cortex. *Cereb Cortex.* 2005;15(3):341-348.
94. Everitt BJ, Hutcheson DM, Ersche KD, Pelloux Y, Dalley JW, Robbins TW. The orbital prefrontal cortex and drug addiction in laboratory animals and humans. *Ann N Y Acad Sci.* 2007;1121:576-597.
95. Fowler JS, Volkow ND, Wang GJ, Pappas N, Logan J, Shea C, Alexoff D, MacGregor RR, Schlyer DJ, Zezulkova I, Wolf AP. Brain monoamine oxidase A inhibition in cigarette smokers. *Proc Natl Acad Sci U S A.* 1996;93(24):14065-14069.
96. Perry DC, Dávila-García MI, Stockmeier CA, Kellar KJ. Increased nicotinic receptors in brains from smokers: membrane binding and autoradiography studies. *J Pharmacol Exp Ther.* 1999;289(3):1545-1552.
97. Brody AL, Mandelkern MA, Jarvik ME, Lee GS, Smith EC, Huang JC, Bota RG, Bartzokis G, London ED. Differences between smokers and nonsmokers in regional gray matter volumes and densities. *Biol Psychiatry.* 2004;55(1):77-84.
98. Oliveira-da-Silva A, Vieira FB, Cristina-Rodrigues F, Filgueiras CC, Manhães AC, Abreu-Villaça Y. Increased apoptosis and reduced neuronal and glial densities in the hippocampus due to nicotine and ethanol exposure in adolescent mice. *Int J Dev Neurosci.* 2009;27(6):539-548.
99. Wobrock T, Falkai P, Schneider-Axmann T, Frommann N, Wölter W, Gaebel W. Effects of abstinence on brain morphology in alcoholism: a MRI study. *Eur Arch Psychiatry Clin Neurosci.* 2009;259(3):143-150.
100. Nagel BJ, Schweinsburg AD, Phan V, Tapert SF. Reduced hippocampal volume among adolescents with alcohol use disorders without psychiatric comorbidity. *Psychiatry Res.* 2005;139(3):181-190.
101. Noonan MA, Bulin SE, Fuller DC, Eisch AJ. Reduction of adult hippocampal neurogenesis confers vulnerability in an animal model of cocaine addiction. *J Neurosci.* 2010;30(1):304-315.
102. Weston BW, Krishnaswami S, Maraty GT, Coly G, Kotchen JM, Grim CE, Kotchen TA. Cocaine use in inner city African American research volunteers. *J Addict Med.* 2009;3(2):83-88.
103. Qian QJ, Yang L, Wang YF, Zhang HB, Guan LL, Chen Y, Ji N, Liu L, Faraone SV. Gene-gene interaction between COMT and MAOA potentially predicts the intelligence of attention-deficit hyperactivity disorder boys in China. *Behav Genet.* 2010;40(3):357-365.
104. Bergouignan L, Chupin M, Czechowska Y, Kinkingnéhun S, Lemogne C, Le Bastard G, Lepage M, Garnero L, Colliot O, Fossati P. Can voxel based morphometry, manual segmentation and automated segmentation equally detect hippocampal volume differences in acute depression? *Neuroimage.* 2009;45(1):29-37.
105. Altena EV, Vrenken H, Van Der Werf YD, van den Heuvel OA, Van Someren EJ. Reduced orbitofrontal and parietal gray matter in chronic insomnia: a voxel-based morphometric study. *Biol Psychiatry.* 2010;67(2):182-185.
106. Ersche KD, Fletcher PC, Lewis SJ, Clark L, Stocks-Gee G, London M, Deakin JB, Robbins TW, Sahakian BJ. Abnormal frontal activations related to decision-making in current and former amphetamine and opiate dependent individuals. *Psychopharmacology (Berl).* 2005;180(4):612-623.
107. Caspi A, Langley K, Milne B, Moffitt TE, O'Donovan M, Owen MJ, Polo Tomas M, Poulton R, Rutter M, Taylor A, Williams B, Thapar A. A replicated molecular genetic basis for subtyping antisocial behavior in children with attention-deficit/hyperactivity disorder. *Arch Gen Psychiatry.* 2008;65(2):203-210.