

# **New Approaches for High-Throughput Identification and Characterization of Protein Complexes**

## **Center for Molecular and Cellular Systems**

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### **Collaborating Laboratories**

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## **Team Leaders**

### **Core:**

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### **High Throughput Complex Processing**

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### **Mass Spectrometry**

**Greg Hurst, Richard Smith**

### **Molecular and Cellular Imaging**

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### **Bioinformatics and Computing**

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**Carol Giometti (ANL) gel electrophoresis**

**Ray Gesteland (U. Utah) mass spectrometry**

**Malin Young (SNL) cross-linking**

**Mike Giddings (U. North Carolina) mass spectrometry**

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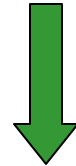
# Goal 1

## ***“Identify and Characterize the Molecular Machines of Life”***

*“...instead of a cell dominated by randomly colliding individual protein molecules, we now know that nearly every major process in a cell is carried out by assemblies ... of proteins...Indeed an entire cell can be viewed as a factory that contains an elaborate network of interlocking assembly lines, each of which is composed of a set of large protein machines.”*

*Bruce Alberts, “The Cell as a Collection of Protein Machines: Preparing the Next Generation of Molecular Biologists,” *Cell*, **92**, 291 (1998)*

**Protein complexes are key to biological function**



Understand the network of reactions that occur in sufficient detail to predict, test, and comprehend the responses of a biological system to changes

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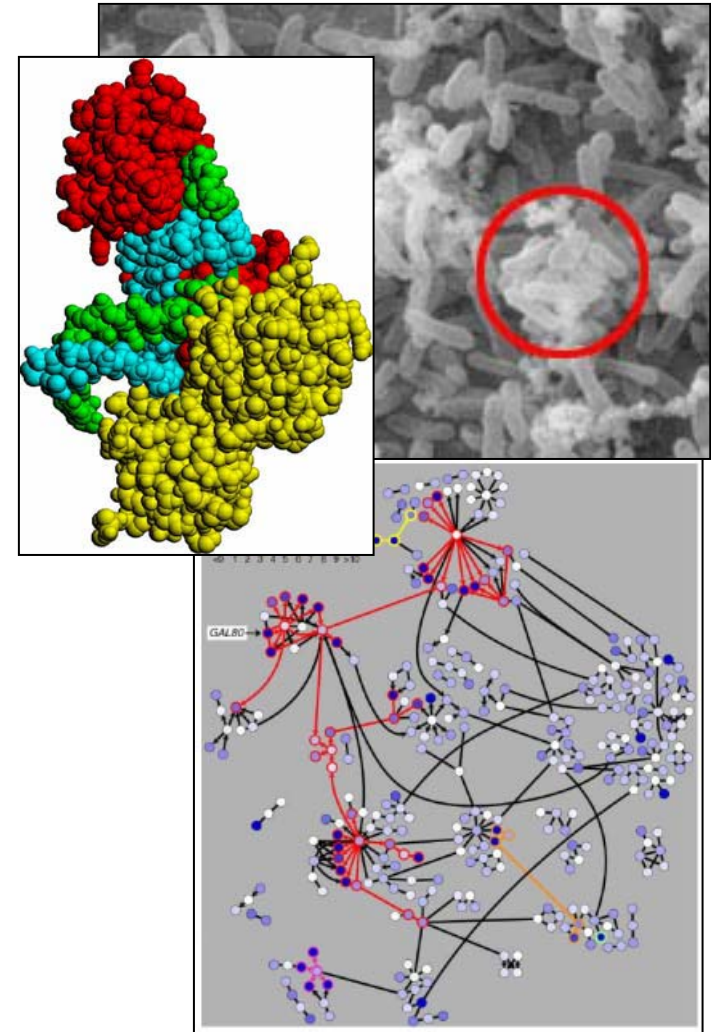
## **Goal 1 includes three main steps**

- Identify complement of protein complexes and their components
- Elucidate function and dynamics of complexes—intermediates, nature of interactions, cellular location, kinetics
- Establish how changes arising from environmental stress, development, etc., affect complex formation and function

**which lay the foundation for GTL**

# Impact of Goal 1

- ⊄ Molecular level understanding of protein complexes and, ultimately, networks
- ⊄ Predict/change behavior of organism and community
- ⊄ Predict function, biological pathways by homology
- ⊄ Discover new functions



# Identification and Characterization of Protein Machines

- ⊄ **New approaches needed for large-scale studies**
  - 4 **No single tool will provide all required information**
  - 4 **Computational tools must be integrated from beginning**
    - ⊄ **Analyze, compare, predict, share data**
    - ⊄ **Quality assessment**
    - ⊄ **Guide experimental design and data collection**



**Develop integrated approach to correlate identified complexes with data from gene expression, protein expression, imaging, and other methods**

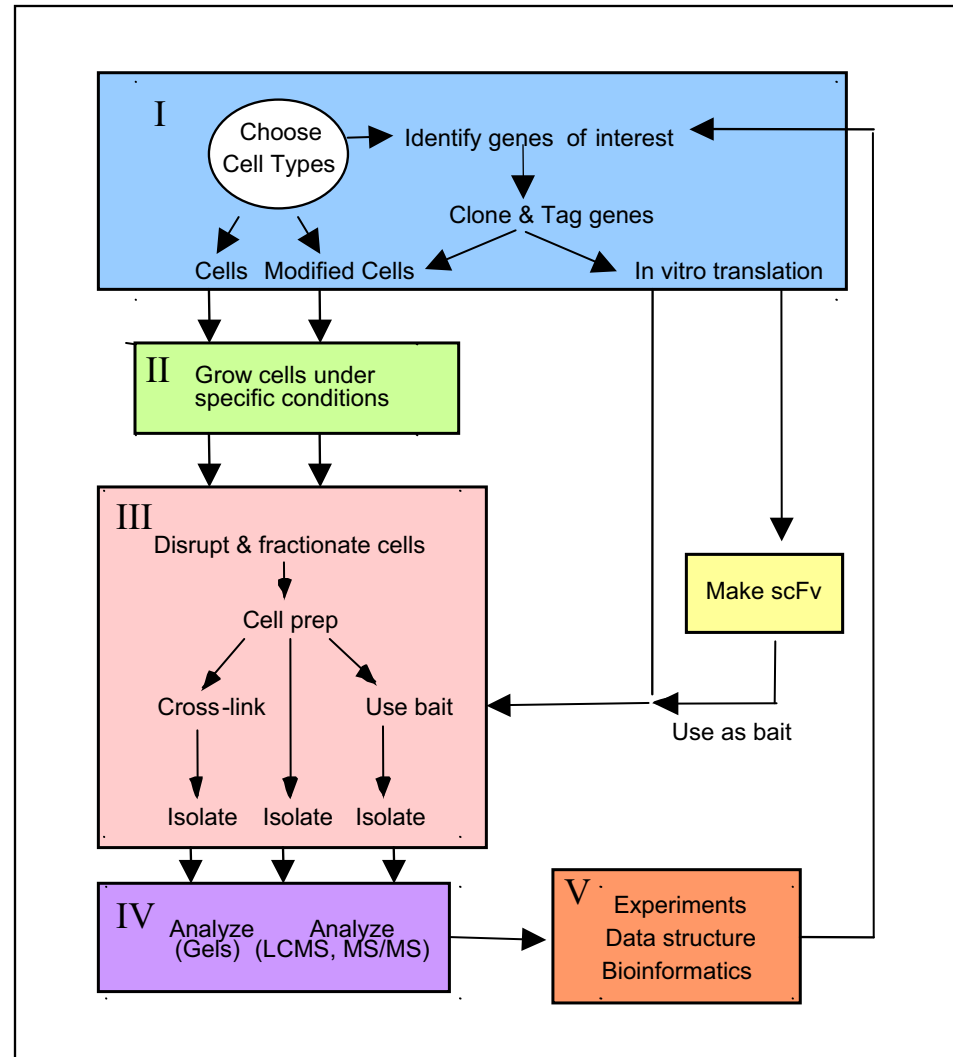
# Strategy to Achieve Goal 1

- € **Initiate protein complex identification using affinity separation combined with mass spectrometry and computational tools**
  - 4 **Use multiple approaches, non-optimized techniques**
  - 4 **Focus on targeted complexes**
  
- € **Evaluate new approaches for high-throughput identification**
  - 4 **Identify bottlenecks, opportunities for automation**
  - 4 **Establish dynamic R&D program to develop new, integrated analytical and computational tools**
  
- € **Incorporate additional tools, data to characterize complexes**
  - 4 **Imaging tools to characterize complexes in cells**
  - 4 **Tools to identify interaction interfaces**

# An Approach for High Throughput Identification of Protein Complexes

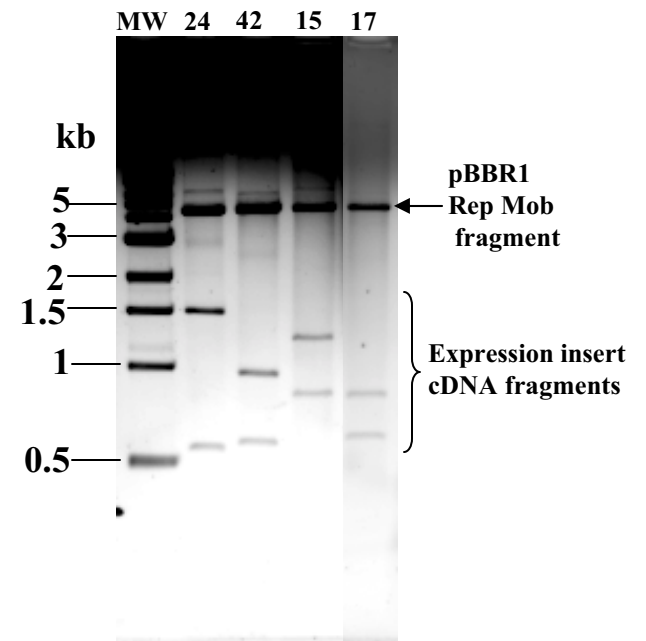
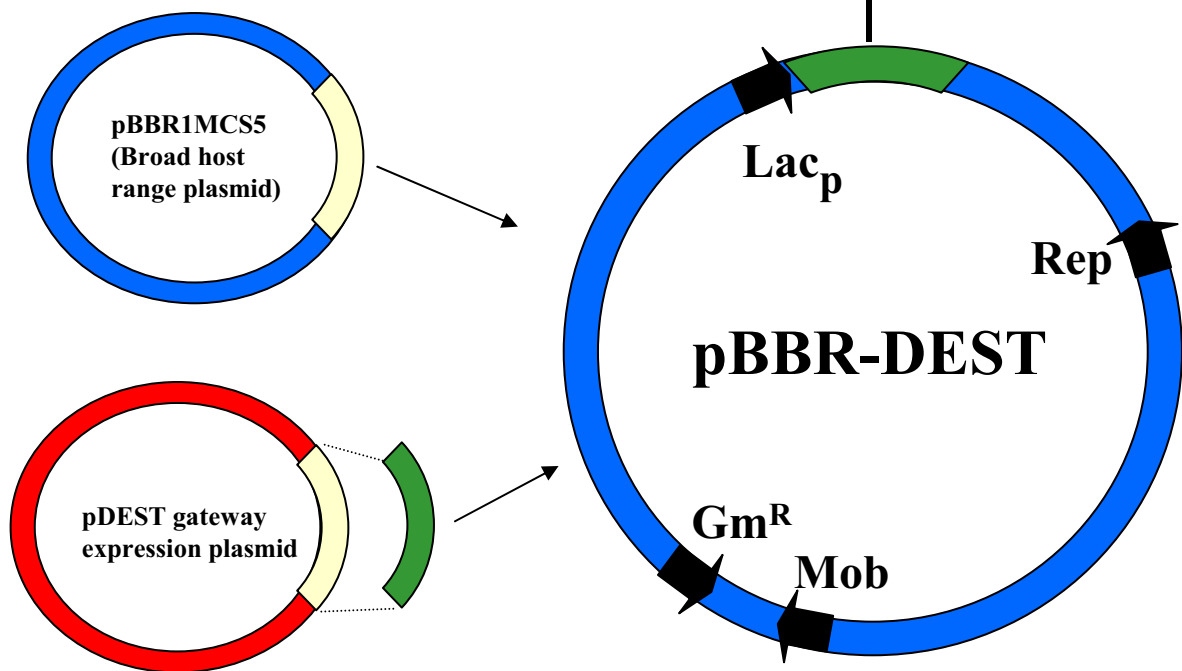
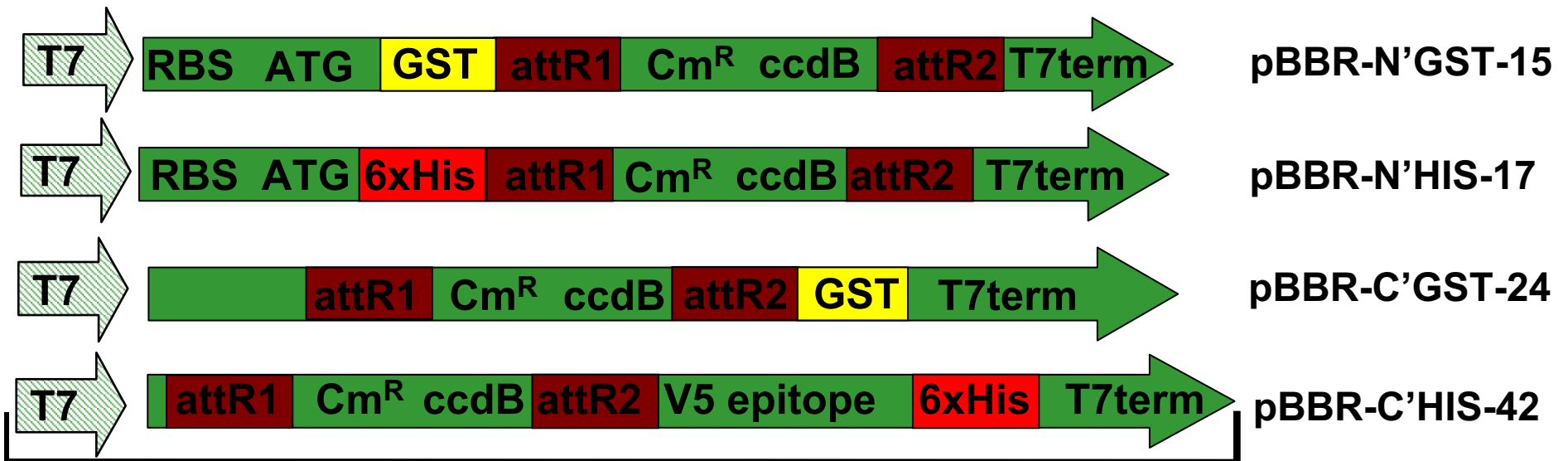
Combine complex isolation, mass spectrometry and data analysis

- 4 Bioinformatics
- 4 Cloning, tagging
- 4 Controlled cell growth
- 4 Affinity isolation
- 4 scFv production
- 4 Cross-linking
- 4 Separation
- 4 MS analysis
- 4 Data analysis, archival

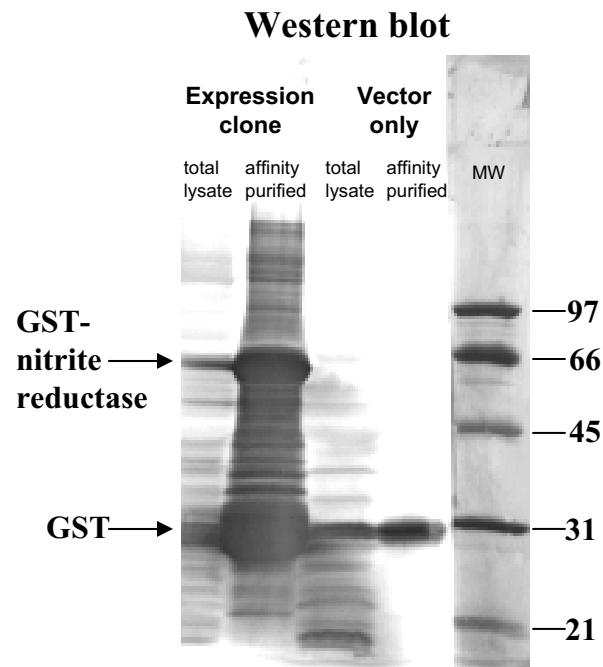
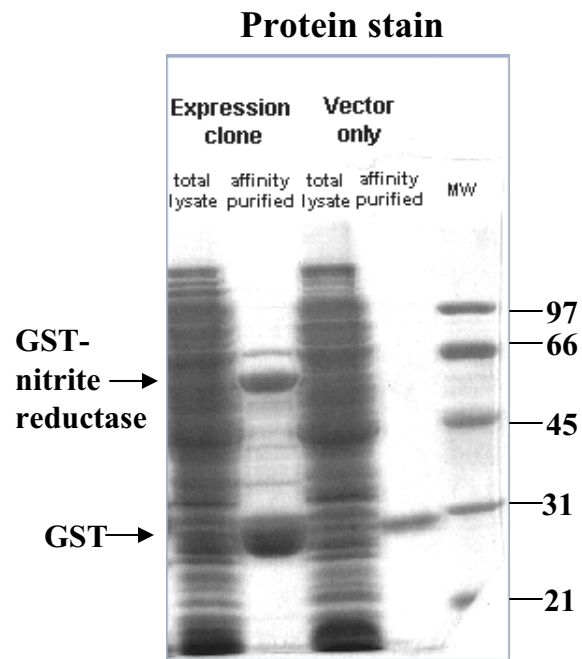
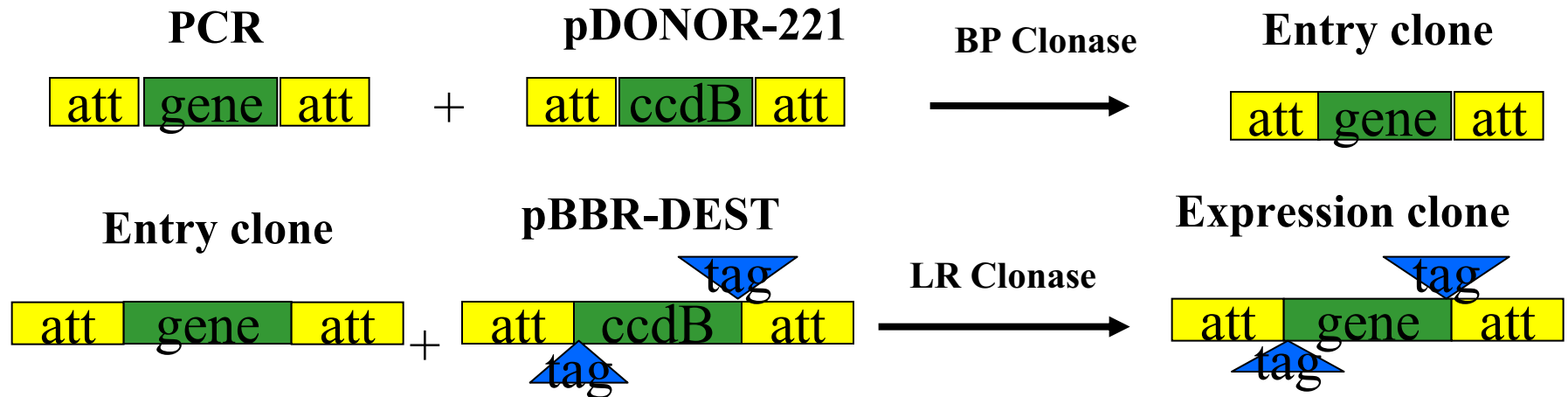




# Modified pDEST Vectors for Protein Expression in *R. palustris*



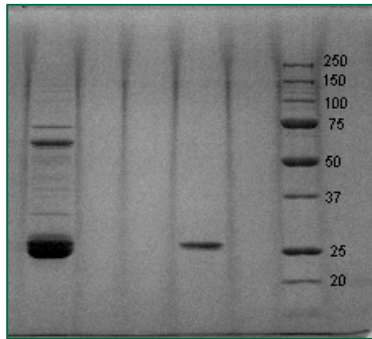
# Modified Gateway system for production of affinity tagged *R. palustris* proteins



**Mass Spectrometry**

	# of peptides identified	
	MALDI	nano LC
		MS/MS
<b>GST-nitrite reductase band</b>		
GST peptides	5	13
GST/linker peptides	2	1
nitrite reductase peptides	9	11
linker/nit. red. peptides	0	1
<b>GST band</b>		
GST peptides	10	22
GST/linker peptides	1	1
nitrite reductase peptides	0	0
linker/nit. red. peptides	0	0

# Verification of *R. palustris* Fusion Proteins Expressed in *E. coli*—Two Approaches



Affinity capture of tagged proteins from lysed cells

1D PAGE

whole eluate digestion

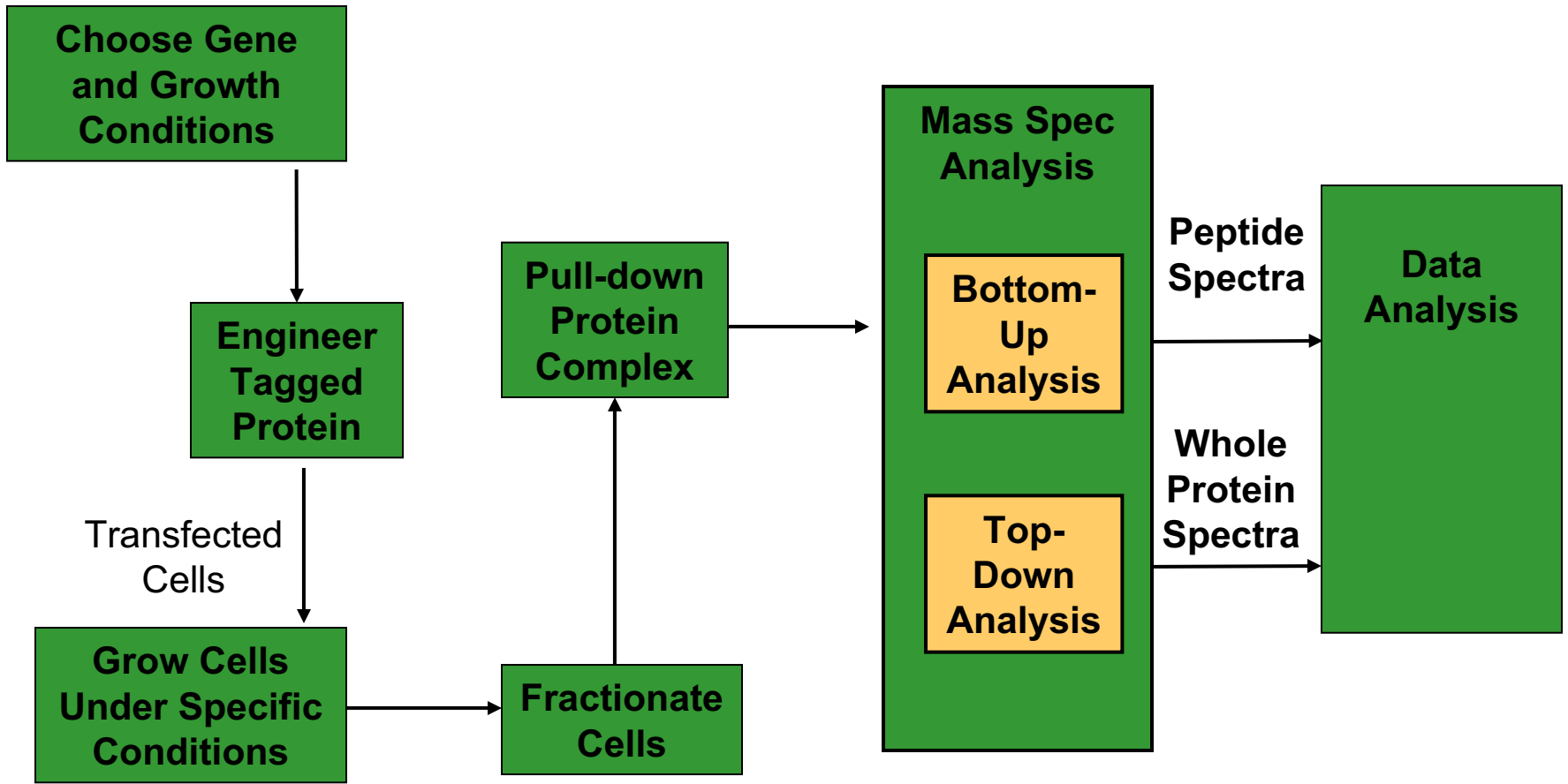
In-gel digestion and mass spectrometric identification of individual gel bands

LC-MS-MS of digest peptides; identification of proteins via SEQUEST

<u>Fusion Protein</u>	<u>No. of peptides identified from:</u>		<u>Others</u>
	<u>target protein</u>	<u>affinity tag</u>	<u>identified</u>
Rpal 4709 + N-terminal GST	45	8	2
Rpal 4709 + C-terminal 6-His & V5 epitope	31	3	19
Rpal 5426 + C-terminal 6-His & V5 epitope	35	3	8

*These are candidate methods for analysis of **protein complexes** isolated via affinity purification*

# ORNL GTL Process Flowchart

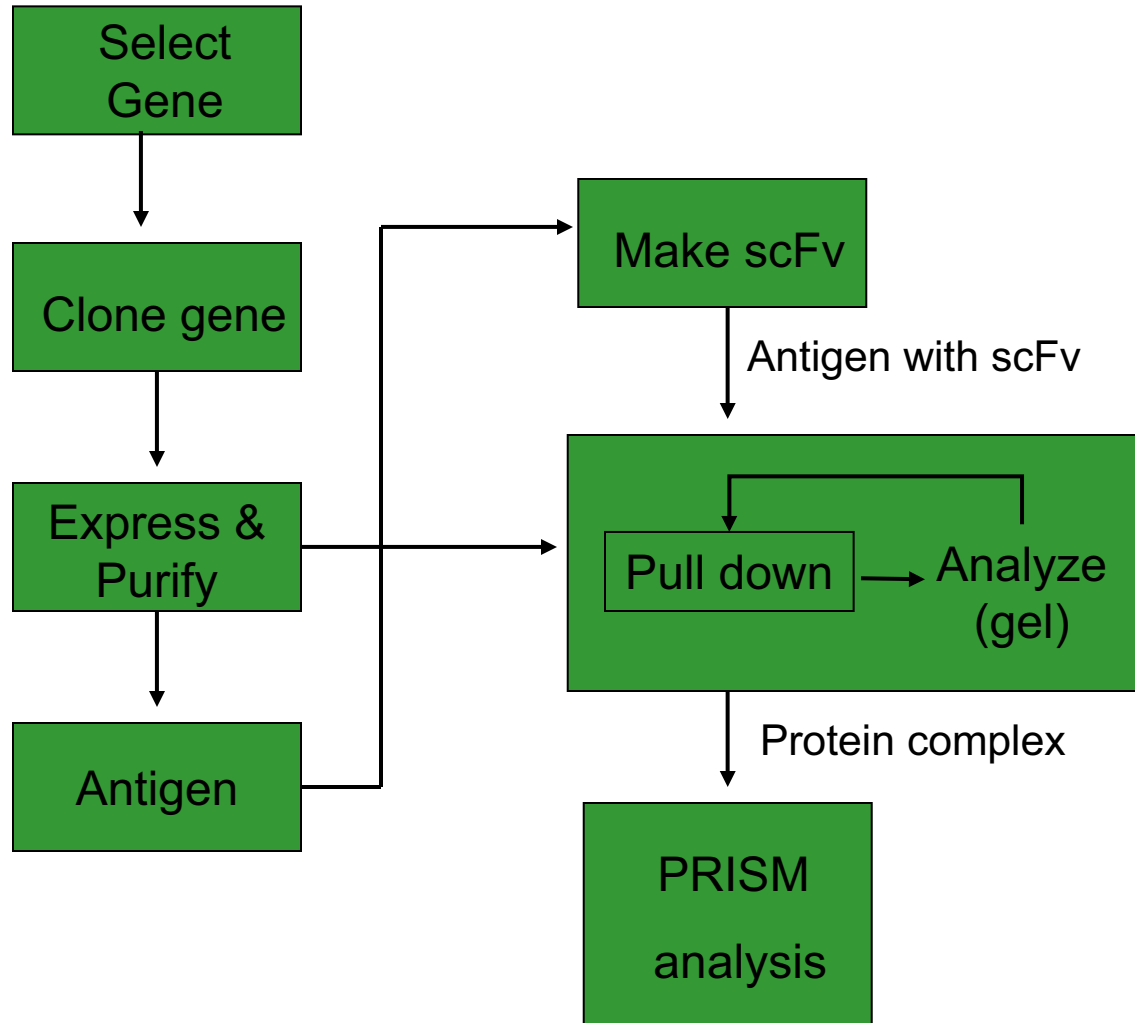


# Analysis of expression of affinity tagged *R. palustris* genes

Gene	Function	Affinity Tag	Expression
<i>nirK</i>	Nitrite reductase	N-His	++
		C-His	+
		N-GST	++
		C-GST	0
<i>groEL-2</i>	chaperonin	N-His	++
		C-His	+++
		N-GST	+
		C-GST	+
<i>groEL-1</i>	chaperonin	N-His	+++
		C-His	+++
		N-GST	++
		C-GST	+
<i>soxB</i>	thiosulfate oxidation	N-His	++
		C-His	++
		N-GST	0
		C-GST	0
<i>soxC</i>	thiosulfate oxidation	N-His	++
		C-His	++
		N-GST	+
		C-GST	+
<i>hupS</i>	uptake hydrogenase small subunit	N-His	0
		C-His	++
		N-GST	0
		C-GST	++
<i>hupL</i>	uptake hydrogenase large subunit	N-His	++
		C-His	++
		N-GST	0
		C-GST	++

+++ Excellent ++ Good + Poor 0 None

# Heterologous Expression

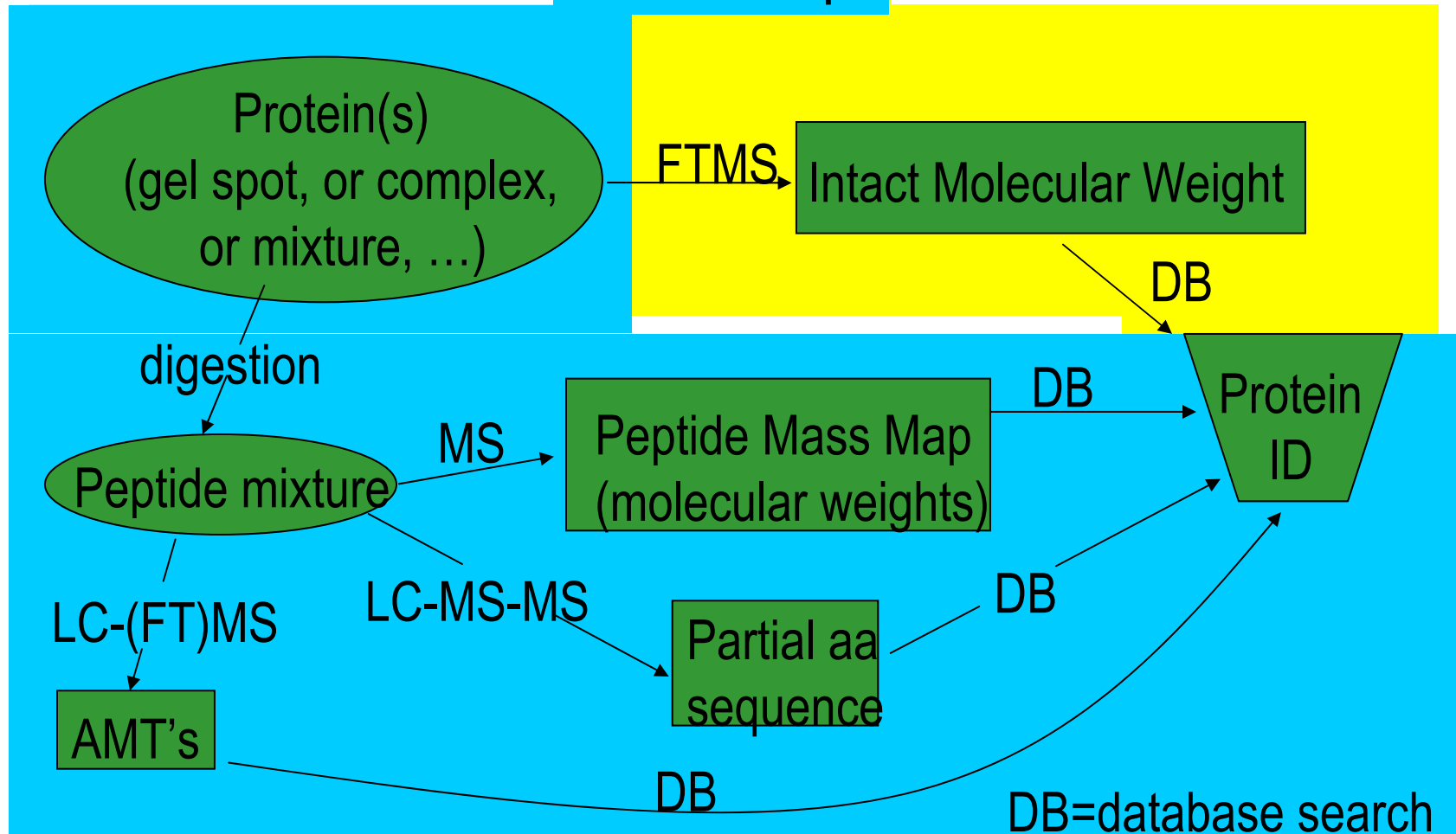


## Tagged proteins generated to date for pull-down studies of *S. Oneidensis*

Gene	Description	Annotation
hydB	periplasmic Fe hydrogenase small subunit	SO3921
hydA	periplasmic Fe hydrogenase large subunit	SO3920
napA	periplasmic nitrate reductase	SO0848
omcA	decaheme cytochrome C	SO1779
omcB	decaheme cytochrome C	SO1778
hoxK	Quinone-reactive Ni/Fe hydrogenase small subunit precursor	SO2099
petA	ubiquinol-cytochrome C reductase iron-sulfur subunit	SO0608
	flavocytochrome C flavin subunit	SO3301
	Gfo/ldh/MocA family oxidoreductase	SO3120
	oxidoreductase molybdopterin-binding	SO0715
nrfC	formate-dependent nitrite reductase	SO0483
ptpA	phosphotyrosine protein phosphatase	SO2208
ptpB	Tyrosine-specific protein phosphatase	SO3124
cpxP	Spheroplast protein y precursor	SO4476
msrA	methionine sulfoxide reductase (isoform A)	SO2337
msrB	methionine sulfoxide reductase (isoform B)	SO2588
eno	Enolase	SO3440
rnIB	ATP-dependent RNA helicase	SO0407
rpoD	RNA polymerase sigma-70 factor	SO1284
	Cytochrome c3	SO2727
rpoA	DNA-directed RNA polymerase alpha subunit	SO0256
rpoZ	DNA-directed RNA omega subunit	SO0360
hepA	RNA polymerase-associated protein	SO0575

# MS for Protein Identification

“Bottom-Up”



*Integrating “Top-Down” and “Bottom-Up” Mass Spectrometric Approaches for Proteomic Analysis of Shewanella oneidensis*, N.C. VerBerkmoes, J.L. Bundy, L. Hauser, K.G. Asano, J. Razumovskaya, F. Larimer, R.L. Hettich, and J.L. Stephenson, Jr., *J. Proteome Research*, in press for Vol 1, issue 3 (estimated June 2002).

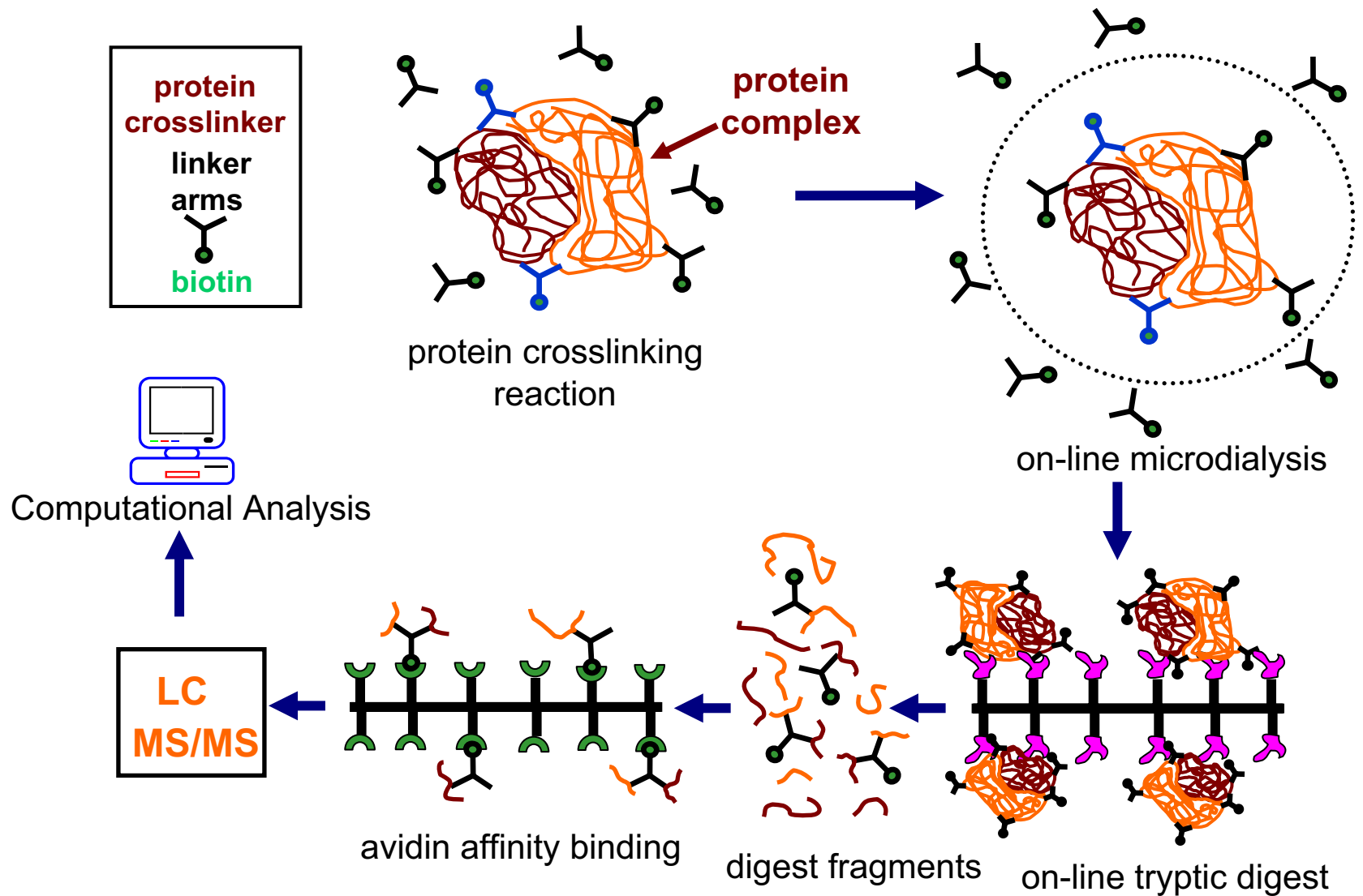


# **Crosslinking and Mass Spectrometry for Protein Complex Analysis**

**Oak Ridge National Laboratory,  
Sandia National Laboratories**

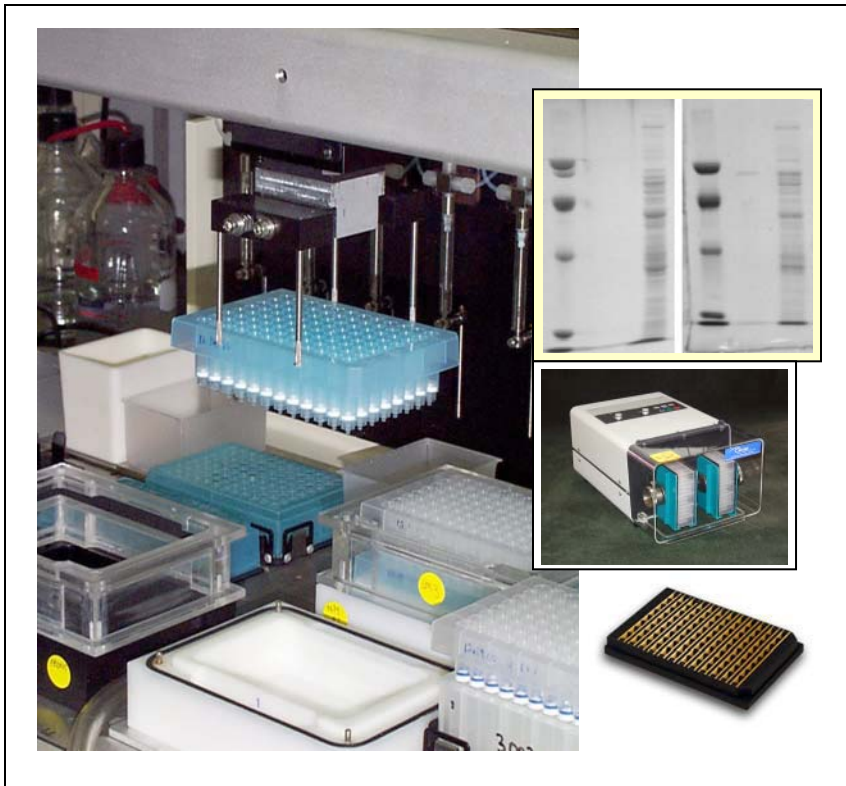
- ⊄ **Chemical crosslinking has potential for:**
  - 4 **Stabilizing “fragile” complexes**
  - 4 **Providing information on distances between particular residues in proteins or complexes**
  - 4 **Improving throughput for MS analysis of complexes**
  
- ⊄ **Technical issues currently being addressed:**
  - 4 **Low abundance of crosslinked products**
  - 4 **Interpretation of mass spectrometry data**

# Protein Complex Analysis: Proposed Affinity Crosslinker Approach

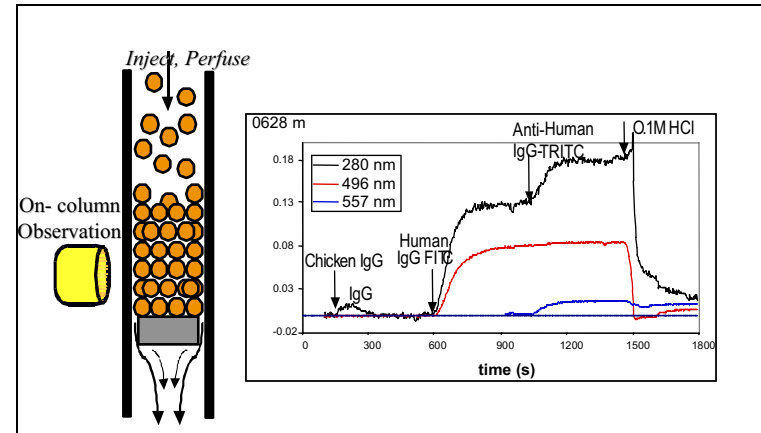


# AUTOMATION OF PROTEIN PRODUCTION & ANALYSES

A



B



**A. Macroscale HT Cloning and Sample Preparation**

C



**B. Microscale Sample Production for Mass Spec**

**C. Lab-On-A-Chip**

# Emerging approaches for characterizing protein complexes

## *Molecular and Cellular Imaging Subproject*



- ⊄ Characterize protein complexes in isolation, within cells, and on cell surfaces/interfaces
- ⊄ Employ multimodality approaches to molecular imaging—optical probes, molecular recognition force microscopy, afm/optical, (optical)<sup>n</sup>
- ⊄ Validate the composition of protein complexes
- ⊄ Determine the location of specific complexes at cellular and subcellular locations
- ⊄ Characterize dynamics, binding forces

# Bioinformatics and Computing

## ∄ Short-term goals

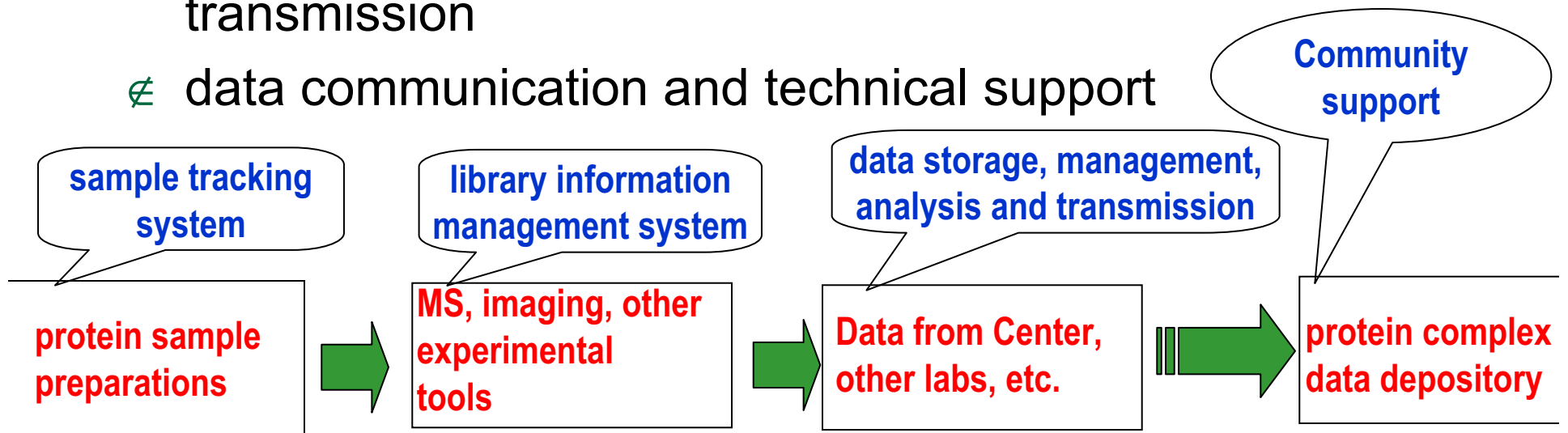
- 4 Create infrastructure for sample tracking, data collection and analysis
- 4 Improve tools for predicting and validating members of protein complexes
- 4 Build tools for interpreting MS data from cross-linked and modified proteins

## ∄ Long-term goals

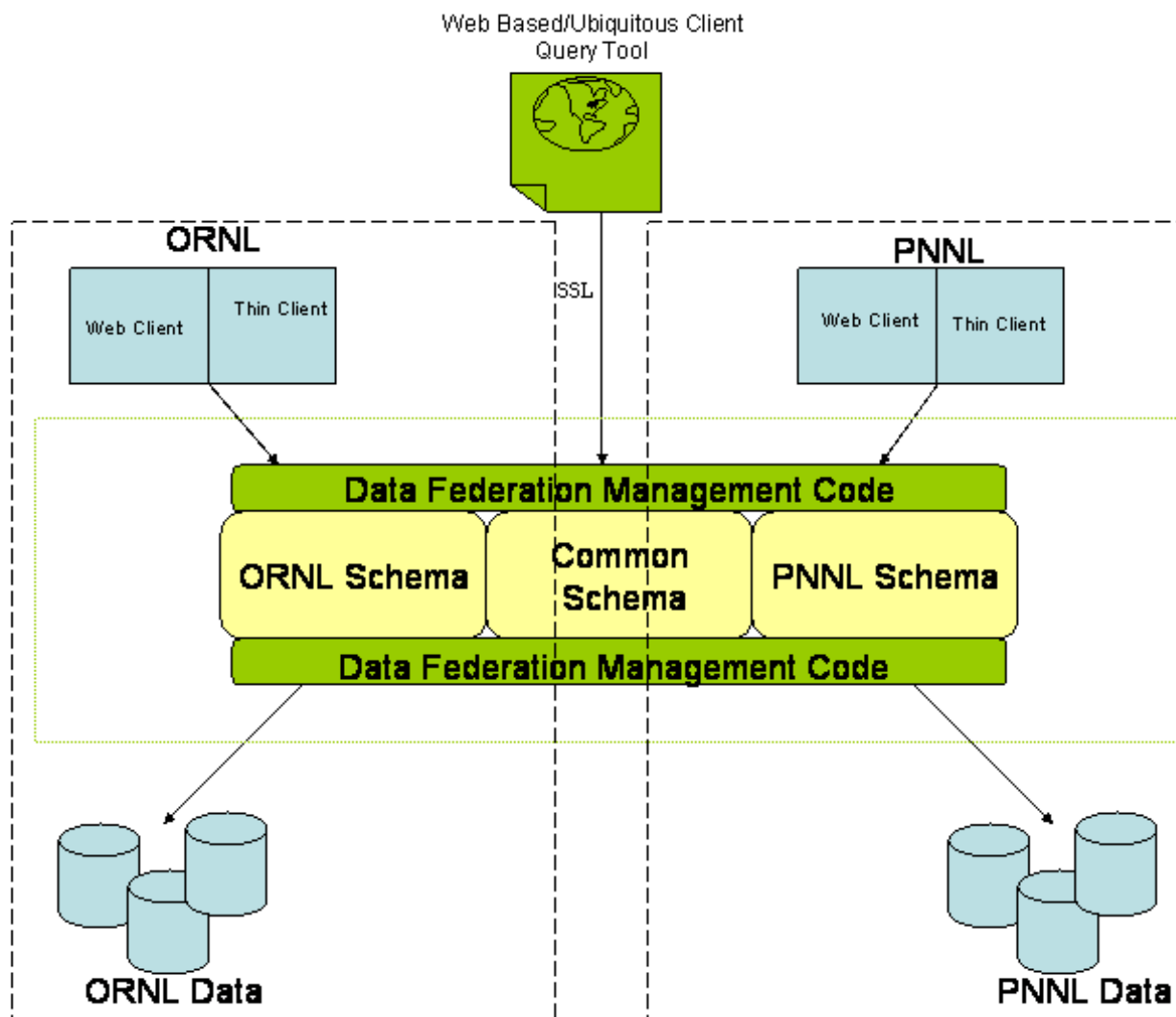
- 4 Predict protein structures involved in forming complexes
- 4 Predict function of protein complexes
- 4 Help build global architecture for integrating data necessary for successful systems biology

# Computational Tools Support All Aspects of Center

- ∉ sample tracking
- ∉ work flow monitoring
- ∉ library information management
- ∉ data processing, storage, management and transmission
- ∉ data communication and technical support

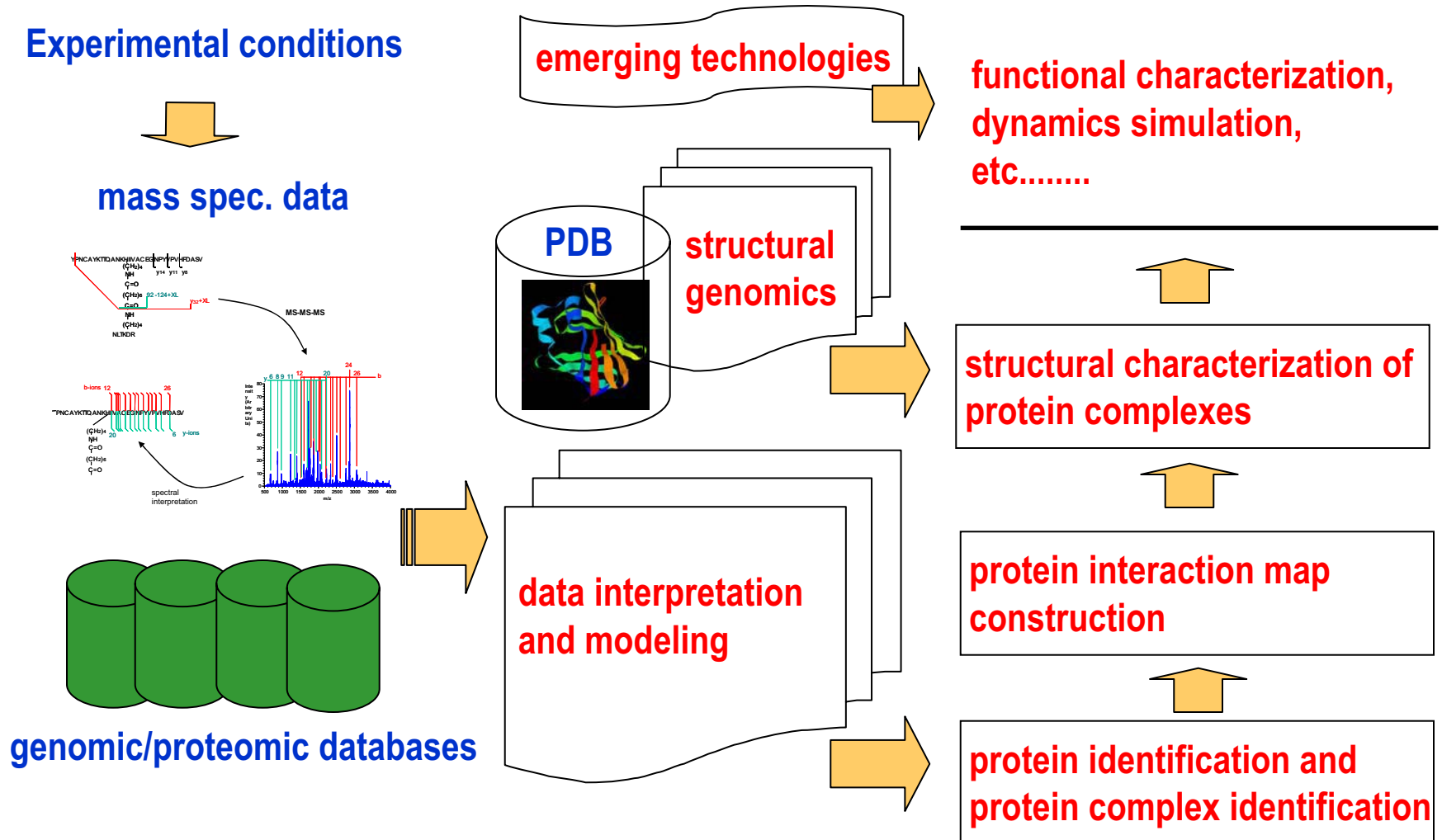


# GTL LIMS System Architecture



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# Computational Characterization of Protein Complexes





# Acknowledgements

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