VI. REFERENCES

- 1. Greenburg L: Benzol poisoning as an industrial hazard--I. The chemistry and industrial uses of benzol--II. Acute benzol poisoning. Public Health Reports 41:1357-75, 1926
- 2. Gerarde HW: The aromatic hydrocarbons, in Patty FA (ed): Industrial Hygiene and Toxicology, rev ed 2. New York, Interscience Publishers, 1962, vol II, pp 1219-40
- 3. Stanford Research Institute: Chemical Economics Handbook. Menlo Park, California, pp 618.5021A-E, 618.5022A-B
- 4. Benzene (Benzol). Inhalation of Concentrated Vapors May Cause Acute, Chronic or Fatal Poisoning. Controlling Chemical Hazards, Ser No 6. US Dept of Labor, Div Labor Standards, 1946, 24 pp
- 5. Benzene, Hygienic Guide Series. American Industrial Hygiene Association, Hygienic Guides Committee, 1970
- 6. 1970 chemical production data. Chem Eng News 49:12A, 1971
- 7. Chemical Safety Data Sheet SD2-Benzene, rev 3. Washington, DC, Manufacturing Chemists Association, 1960, 15 pp
- 8. Weast RC (ed): Handbook of Chemistry and Physics-- A Ready Reference Book of Chemical and Physical Data, ed 52. Cleveland, The Chemical Rubber Publishing Co, 1971
- 9. Sherwood RJ: Benzene: The interpretation of monitoring results.
 Ann Occup Hyg 15: 409-21, 1972
- 10. Benzene in motor gasoline. International Study Group for Conservation of Clean Air and Water, Western Europe (Stichting Concawe), The Hague, 1973
- 11. Bowden JN: Status of unleaded and low-lead gasoline composition--Interim report FLRL No 16, AD 747421. Aberdeen Proving Ground, Md, US Army Coating and Chemical Laboratory, 1972
- 12. Pagnotto LD, Elkins HB, Brugsch HG, Walkley EJ: Industrial benzene exposure from petroleum naphtha--I. Rubber coating industry. Am Ind Hyg Assoc J 22:417-21, 1961
- 13. Legge TM: Chronic benzol poisoning. J Ind Hyg 1:539-41, 1920
- 14. Selling L: Benzol as a leucotoxin-- Studies on the degeneration and regeneration of the blood and haematopoietic organs. Johns Hopkins Hosp Rep 17:83-136, 1916
- 15. Hamilton A: Benzene (benzol) poisoning. Arch Pathol 11:434-54, 1931

- 16. Smith AR: Chronic benzol poisoning among women industrial workers—A study of the women exposed to benzol fumes in six factories. J Ind Hyg 10:73-93, 1928
- 17. Greenburg L, Mayers MR, Goldwater L, Smith AR: Benzene (benzol) poisoning in the rotogravure printing industry in New York City. J Ind Hyg Toxicol 21:395-420, 1939
- 18. Bowditch M, Elkins HB: Chronic exposure to benzene (benzo1)--I. The industrial aspects. J Ind Hyg Toxicol 21:321-30, 1939
- 19. Greenburg, L: Benzol poisoning as an industrial hazard--VII. Results of medical examination and clinical tests made to discover early signs of benzol poisoning in exposed workers. Public Health Reports 41:1526-39, 1926
- 20. Erf LA, Rhoads CP: The hematological effects of benzene (benzol) poisoning. J Ind Hyg Toxicol 21:421-35, 1939
- 21. Hunter FT: Chronic exposure to benzene (benzo1)--II. The clinical effects. J Ind Hyg Toxicol 21:331-54, 1939
- 22. Mallory TB, Gall EA, Brickley WJ: Chronic exposure to benzene (benzol)--III. The pathologic results. J Ind Hyg Toxicol 21:355-77, 1939
- 23. Browning E: Toxicity and Metabolism of Industrial Solvents. New York, Elsevier Publishing Company, 1965, pp 3-65
- 24. Gerarde HW: Toxicology and Biochemistry of Aromatic Hydrocarbons. New York, Elsevier Publishing Company, 1960, pp 97-108
- 25. Flury F: [II. Modern occupational intoxications. IIa. Modern occupational intoxications from the aspect of pharmacology and toxicology.] Arch Exp Path Pharmakol 138:65-82, 1928 (Ger)
- 26. A new domestic poison. Lancet 1:105, 1862

. . . .

- 27. Averill C: Benzole poisoning. Br Med J 1:709, 1889
- 28. Cesaro AN: [Is absorption of benzene through the skin possible?] Med Lavoro 37:151-56, 1946 (Ital)
- 29. Conca GL, Maltagliati A: [Transcutaneous absorption of benzene.]
 Med Lavoro 46:194-98, 1955 (Ital)
- 30. Hanke J, Dutkiewicz T, Piotrowski I: [The absorption of benzene through the skin in men.] Med Pracy 12: 413-26, 1961
- 31. Dutkiewicz T, Tyras H: A study of the skin absorption of ethylbenzene in man. Br J Indust Med 24:330-32, 1967

- 32. Dutkiewicz T, Tyras H: [The quantitative estimation of toluene skin absorption in man.] Arch Gewerbepathol Gewerbehyg 24: 253-57, 1968 (Ger)
- 33. Srbova J, Teisinger J, Skramovsky S: Absorption and elimination of inhaled benzene in man. Arch Ind Hyg Occup Med 2:1-8, 1950
- 34. Teisinger J, Bergerova-Fiserova V, Kudrna J: [The metabolism of benzene in man.] Procovni lekarstvi 4:175, 1952 (Pol)
- 35. Duvoir MR, Fabre A, Derobert L: [The significance of benzene in the bone marrow in the course of benzene blood diseases.] Arch Mal Prof 7:77, 1946 (Fr)
- 36. Hunter CG: Aromatic solvents. Ann Occup Hyg 9:191-97, 1966
- 37. Stewart RD, Dodd HC, Baretta ED, Schaeffer AW, Mutchler JE: Chronic overexposure to benzene vapor. Toxicol Pharmacol 10:381, 1967; abst
- 38. Helmer KJ: Accumulated cases of chronic benzene poisoning in the rubber industry. Acta Med Scand 118:354-75, 1944
- 39. Savilahti M: [More than 100 cases of benzene poisoning in a shoe factory.] Arch Gewerbepathol Gewerbehyg 15:147-57, 1956 (Ger)
- 40. Juzwiak I: [Studies on the state of health of shoe plant workers exposed to benzene and its homologues.] Med Przemyslowa 20:67-72, 1969 (Pol)
- 41. Vigliani EC, Saita G: Benzene and leukemia. N Engl J Med 271:872-76, 1964
- 42. Pollini G, Colombi R: [Medullary chromosome damage in aplastic anemia caused by benzol.] Med Lavoro 55: 241-55, 1964 (Ital)
- 43. Tough IM, Court Brown WM: Chromosome aberrations and exposure to ambient benzene. Lancet 1:684, 1965
- 44. Forni A, Moreo L: Cytogenetic studies in a case of benzene leukaemia. Eur J Cancer 3:251-55, 1967
- 45. Forni A, Moreo L: Chromosome studies in a case of benzene-induced erythroleukaemia. Eur J Cancer 5:459-63, 1969
- 46. Tough IM, Smith PG, Court Brown WM, Harnden DG: Chromosome studies on workers exposed to atmospheric benzene. Eur J Cancer 6:49-55, 1970
- 47. Forni A, Pacifico E, Limonta A: Chromosome studies in workers exposed to benzene or toluene or both. Arch Environ Health 22:373-78, 1971

- 48. Forni A, Cappellini A, Pacifico E, Vigliani EC: Chromosome changes and their evolution in subjects with past exposure to benzene. Arch Environ Health 23:385-91, 1971
- 49. Hartwich G, Schwanitz G: [Chromosome studies after chronic exposure to benzol.] Dtsch Med Wochenschr 97:45-49, 1972 (Ger)
- 50. Buckton KE, Jacobs PA, Court Brown WM: A study of the chromosome damage persisting after x-ray therapy for ankylosing spondylitis. Lancet 2:676-82, 1962
- 51. Bender MA, Gooch PC: Persistent chromosome aberrations in irradiated human subjects. Radiat Res 16:44-53, 1962
- 52. Bender MA, Gooch PC: Persistent chromosome aberrations in irradiated human subjects--II. Three and one half year investigation. Radiat Res 18:389-96, 1963
- 53. Goh K-O: Total-body irradiation and human chromosomes--Cytogenetic studies of the peripheral blood and bone marrow leukocytes seven years after total-body irradiation. Radiat Res 35:155-70, 1968
- 54. Thorpe JJ: Epidemiologic survey of leukemia in persons potentially exposed to benzene. J Occup Med 16:375-82, 1974
- 55. Cavignaux L: [Confirmed intoxications.] Cah Med Interprof 2: 28-31, 1962 (Fr)
- 56. Butarewicz L, Gosk S, Gluszczowa M: [Examination of the state of health of women workers in the leather industry, especially from the gynecological point of view.] Med Przemyslowa 20:137-48, 1969 (Pol)
- 57. Hardy HL, Elkins HB: Medical aspects of maximum allowable concentrations-Benzene. J Ind Hyg Toxicol 30:196-200, 1948
- 58. Kozlova TA, Volkova AP: [The blood picture and phagocytic activity of leucocytes in workers having contact with benzol.] Gig Sanit 25:29-34, 1960 (Rus)
- 59. Horiuchi K, Horiguchi S, Aratake K: Studies on the maximum allowable concentration of benzene in the air of workshops. Osaka City Med J 9:79-90, 1963
- 60. Cassan G, Baron J: [Usefulness of blood tests in workers exposed to benzene.] Arch Mal Prof 17:602-604, 1956 (Fr)
- 61. Lazarew NW, Brussilowskaja AJ, Lawrow JN, Lifschitz FB: [Cutaneous permeability for petroleum ether and benzene.] Arch Hyg 106:112-22, 1931 (Ger)
- 62. Carpenter CP, Shaffer CB, Weil CS, Smyth HF Jr: Studies on the inhalation of 1:3-butadiene with a comparison of its narcotic effect

- with benzol, toluol, and styrene, and a note on the elimination of styrene by the human. J Ind Hyg Toxicol 26:69-78, 1944
- 63. Jonek J, Olknowski Z, Zieleznik B: Histochemical studies on the spinal cord of mice poisoned with benzene. Acta Histochem 20:286-96, 1965
- 64. Schrenk H, Yant WP, Pearce SJ, Patty FA, Sayers RR: Absorption, distribution and elimination of benzene by body tissues and fluids of dogs exposed to benzene. J Ind Hyg Toxicol 23:20-34, 1941
- 65. Desoille H, Philbert M, Albahary C: [Hormonal influences in chronic benzene intoxication in guinea pigs. Influence of gestation on white and red blood cell counts in guinea pigs with and without moderate benzene intoxication during the entire gestation period.]

 Arch Mal Prof 28:329-39, 1967 (Fr)
- 66. Deichmann WB, MacDonald WE, Bernal E: The hemopoietic tissue toxicity of benzene vapors. Toxicol Appl Pharmacol 5:201-24, 1963
- 67. Nau CA, Neal J, Thornton M: C9-C12 fractions obtained from petroleum distillates--An evaluation of their potential toxicity. Arch Environ Health 12:382-93, 1966
- 68. Wolf MA, Rowe VK, McCollister DD, Hollingsworth RL, Oyen F: Toxicological studies of certain alkylated benzenes and benzene--Experiments in laboratory animals. Arch Ind Health 14:387-98, 1956
- 69. Novikov YV: [Effect of small benzene concentrations on higher nervous activity of animals in chronic experiments.] Gig Sanit 21:20-25, 1956 (Rus); also in USSR Literature on Air Pollution and Related Occupational Diseases--A survey. BS Levine (trans1), USPHS, 1960, vol 2, pp 185-91
- 70. Horiuchi K, Horiguchi S, Morioka S: Maximum allowable concentration of benzene in an animal experiment. Osaka City Med J 13:1-8, 1967
- 71. Shils ME, Goldwater LJ: Nutritional factors affecting the toxicity of some aromatic hydrocarbons with special reference to benzene and nitrobenzene compounds—A review. J Ind Hyg Toxicol 31:175-89, 1949
- 72. Winternitz MC, Hirschfelder AD: Studies upon experimental pneumonia in rabbits--Parts I to III. J Exp Med 18:657-65, 1913
- 73. Kline BS, Winternitz MC: Studies upon experimental pneumonia in rabbits--V. The role of the leucocyte in experimental pneumonia. The relation of the number of organisms injected to the mortality. J Exp Med 18:50-60, 1913
- 74. Weiskotten HG, Steensland HS: Action of benzol--IV. Spontaneous infections with special reference to the diphasic leucopenia (rabbit). Exp Res 37:215-23, 1917

- 75. White WC, Gammon AM: The influence of benzol inhalations on experimental pulmonary tuberculosis in rabbits. Trans Assoc Amer Phys 29:332-337, 1914
- 76. Camp WE, Baumgartner EA: Inflammatory reactions in rabbits with a severe leucopenia. J Exp Med 22:174-92, 1915
- 77. Williams RT: Detoxification Mechanisms. New York, Wiley and Sons, 1959, pp 188-94
- 78. Cornish HH, Ryan RC: Metabolism of benzene in nonfasted, fasted, and aryl-hydroxylase inhibited rats. Toxicol Appl Pharmacol 7:767-71, 1965
- 79. Mitchell JR: Mechanism of benzene-induced aplastic anemia. Fed Am Soc Exp Biol 30:561, 1971; abst
- 80. Posner HS, Mitoma C, Udenfriend S: Enzymatic hydroxylation of aromatic compounds. Arch Biochem Biophys 94:269-79, 1961
- 81. Saito FU, Kocsis JJ, Snyder R: Effect of benzene on hepatic drug metabolism and ultrastructure. Toxicol Appl Pharmacol 26:209-17, 1973
- 82. Drew RT, Fouts JR: The lack of effects of pretreatment with phenobarbital and chlorpromazine on the acute toxicity of benzene in rats. Toxicol Appl Pharmacol 27:183-93, 1974
- 83. Lee EW, Kocsis JJ, Snyder R: Acute effect of benzene on 59Fe incorporation into circulating erythrocytes. Toxicol Appl Pharmacol 27:431-36, 1974
- 84. Parkinson GS: Benzene in motor gasoline—An investigation into possible health hazards in and around filling stations and in normal transport operations. Ann Occup Hyg 14:145-53, 1971
- 85. Sherwood RJ: Evaluation of exposure to benzene vapour during the loading of petrol. Br J Ind Med 29:65-9, 1972
- 86. Blaney L: Early detection of benzene toxicity. Ind Med Surg 19:227-28, 1950
- 87. Dolin BH: Determination of benzene: Detection and estimation of benzene in the presence of toluene, xylene, and other substances. Ind Eng Chem, Anal Ed 15:242-47, 1943
- 88. Levine BS (ed): Quantitative determination of benzene in the air. USSR Literature on Air Pollution and Related Occupational Diseases 8:47-51, 1963
- 89. Smith BS, Pierce JO: The use of plastic bags for industrial air sampling. Am Ind Hyg Assoc J 31:343-48, 1970

- 90. Ovrum P: Determination of atmospheric benzene concentration by displacement following adsorption on silica gel. Br J Ind Med 13:210-13, 1956
- 91. Elkins HB, Pagnotto LD, Comproni EM: The ultraviolet spectrophotometric determination of benzene in air samples adsorbed on silica gel. Anal Chem 34:1797-1801, 1962
- 92. Van Mourik JHC: Experiences with silica gel as adsorbent. Am Ind Hyg Assoc J 26:498-509, 1965
- 93. Feldstein M, Balestrieri S, Levaggi DA: The use of silica gel in source testing. Am Ind Hyg Assoc J 28:381-85, 1967
- 94. Buchwald H: Activated silica gel as an adsorbent for atmospheric contaminants. Occup Health Rev 17:14-18, 1965
- 95. Fraust CL, Hermann ER: Charcoal sampling tubes for organic vapor analysis by gas chromatography. Am Ind Hyg Assoc J 27:68-74, 1966
- 96. Reid FH, Halpin WR: Determination of halogenated and aromatic hydrocarbons in air by charcoal tube and gas chromatography. Am Ind Hyg Assoc J 29:390-96, 1968
- 97. Baernstein HD: Photometric determination of benzene, toluene, and their nitro derivatives. Ind Eng Chem, Anal Ed 15:251-53, 1943
- 98. Maffett PA, Doherty TF, Monkman JL: A direct method for the collection and determination of micro amounts of benzene or toluene in air. Am Ind Hyg Assoc Quart 17:186-88, 1956
- 99. Analytical Abstracts Committee: Analytical Abstracts, Benzene.
 American Industrial Hygiene Association, 1965
- 100. Koljkowsky P: Indicator-tube method for the determination of benzene in air. Analyst 94:918-20, 1969
- 101. Ash RM, Lynch JR: The evaluation of gas detector tube systems--Benzene. Am Ind Hyg Assoc J 32:410-11, 1971
- 102. Sherwood RJ, Carter FWG: The measurement of occupational exposure to benzene vapour. Ann Occup Hyg 13:125-46, 1970
- 103. Levadie R, Harwood JF: An application of gas chromatography to analysis of solvent vapors in industrial air. Am Ind Hyg Assoc J 21:20-24, 1960
- 104. Whitman NE, Johnston AE: Sampling and analysis of aromatic hydrocarbon vapors in air: A gas-liquid chromatographic method. Am Ind Hyg Assoc J 25:464-69, 1964

- 105. Rules and Regulations on Sanitation of Factories and Mercantile Establishments, Rule 59. Maine Department of Health and Welfare, December 9, 1954.
- 106. Regulations for the Control and Prevention of Occupational Disease in Industry, Regulation 4. Florida Industrial Commission, December 16, 1957.
- 107. American Conference of Governmental Industrial Hygienists: Transactions of the Eighth Annual Meeting. Cincinnati, Ohio, ACGIH, 1946, p 40
- 108. American Conference of Governmental Industrial Hygienists: Transactions of the Ninth Annual Meeting. Cincinnati, Ohio, ACGIH, 1947, p 44
- 109. American Conference of Governmental Industrial Hygienists: Transactions of the Tenth Annual Meeting. Cincinnati, Ohio, ACGIH, 1948, p 31
- 110. American Conference of Governmental Industrial Hygienists: Transactions of the Nineteenth Annual Meeting. Cincinnati, Ohio, ACGIH, 1957, p 47
- 111. American Conference of Governmental Industrial Hygienists: Threshold Limit Values for Substances in Workroom Air Adopted by ACGIH for 1963. Cincinnati, Ohio, ACGIH, 1963.
- 112. American Conference of Governmental Industrial Hygienists: Documentation of the Threshold Limit Values for Substances in Workroom Air, ed 3. Cincinnati, Ohio, 1971, ACGIH, p 22
- 113. United States of America Standards Institute: Acceptable Concentrations of Benzene Z37.4-1969. New York, 1969, 8 pp
- 114. Convention 136 concerning protection against hazards of poisoning arising from benzene, adopted by the Conference at its 56th session, International Labour Conference, Geneva, June, 1971
- 115. Recommendation 144 concerning protection against hazards of poisoning arising from benzene, adopted by the Conference at its 56th session, International Labour Conference, Geneva, June, 1971
- 116. Benzene: Uses, Toxic Effects, Substitutes. Meeting of Experts on the Safe Use of Benzene and Solvents Containing Benzene, May 16-22, 1967. International Labour Office, Geneva, 1968
- 117. Smelyanskiy ZB, Ulanova IP: [New standards for permissible levels of toxic gases, fumes, and dust in the air of work areas.] Ind Hyg Occup Dis No 5:7-15, 1959 (Rus)

..

- 118. Volkova ZA: [The relationship between human health and environmental conditions (at work and in everyday life) as a method for verifying the safety criteria for human exposure to chemical substances.] Consultation on methods used in the USSR for establishing biologically safe levels of toxic substances, World Health Organization, OH/WP/72.5, December 1972 (Rus)
- 119. Parke DV, Williams RT: Studies in detoxication—The metabolism of benzene—(a) The determination of benzene; (b) The elimination of unchanged benzene in rabbits. Biochem J 46:236-42, 1953
- 120. Dutton GJ: Uridine diphosphate glucuronic acid as glucuronyl donor in the synthesis of "ester," aliphatic and steroid glucuronides. Biochem J 64:693-701, 1956
- 121. Guertin DL, Gerarde HW: Toxicological studies on hydrocarbons-- IV.
 A method for the quantitative determination of benzene and certain alkylbenzenes in blood. Arch Ind Health 20:262-65, 1959
- 122. Hunter CG: Solvents with reference to studies on the pharmaco-dynamics of benzene. Proc Roy Soc Med 61:913-15, 1968
- 123. Sherwood RJ: One man's elimination of benzene (C6H6). Proceedings of the 3rd Annual Conference on Environmental Toxicology, AMRL TR-72-130, Dayton, 1972
- 124. Walkley JE, Pagnotto LD, Elkins HB: The measurement of phenol in urine as an index of benzene exposure. Am Ind Hyg Assoc J 22:362-67, 1961
- 125. Elkins HB: The Chemistry of Industrial Toxicology, ed 2. New York, John Wiley & Sons, 1959
- 126. Truhaut R: [Determination of a tolerable limit of benzene in work environment.] Arch Mal Prof 29:5-22, 1968 (Fr)
- 127. Teisinger J, Fiserova-Bergerova V: [Comparative value of the determination of urinary sulfates and phenol for the evaluation of the atmospheric benzene concentration.] Arch Mal Prof 16:221-32, 1955 (Fr)
- 128. Docter JH, Zielhuis RL: Phenol excretion as a measure of benzene exposure. Ann Occup Hyg 10:317-26, 1967
- 129. Deichmann W, Schafer LJ: Phenol studies. Am J Clin Pathol 12:129-43, 1942
- 130. Buchwald H: The expression of urine analysis results--Observations on the use of a specific gravity correction. Ann Occup Hyg 7:125-36, 1964

- 131. Theis RC, Benedict SR: The determination of phenols in the blood.

 J Biol Chem 61:67-71, 1924
- 132. Buchwald H: The colorimetric determination of phenol in air and urine with a stabilized diazonium salt. Ann Occup Hyg 9:7-14, 1966
- 133. Gibbs HD: Phenol tests--III. The indophenol test. J Biol Chem 72:649-64, 1927
- 134. Rainsford SG, Lloyd Davies TA: Urinary excretion of phenol by men exposed to vapour of benzene--A screening test. Br J Ind Med 22:21-26, 1965
- 135. Van Haaften AB, Sie ST: The measurement of phenol in urine by gas chromatography as a check on benzene exposure. Am Ind Hyg Assoc J 26:52-58, 1965
- 136. White LD, Taylor DG, Mauer PA, Kupel RE: A convenient optimized method for the analysis of selected solvent vapors in the industrial atmosphere. Am Ind Hyg Assoc J 31:225-32, 1970
- 137. Kupel RE, White LD: Report on a modified charcoal tube. Am Ind Hyg Assoc J 32:456, 1971
- 138. Standard recommended practice for use of the terms precision and accuracy as applied to the measurement of a property of a material, ASTM E 177-71. Philadelphia, American Society of Testing and Materials, 1971, 18 pp
- 139. Levine L, Fahy JP: Evaluation of urinary lead determinations. I. The significance of the specific gravity. J Ind Hyg Toxicol 27:217-23, 1945
- 140. Wintrobe MW: Clinical Hematology, ed 6. Philadelphia, Lea and Febiger, 1967
- 141. Berlin NI, Waldmann TA, Weissman SM: Life span of the red blood cell. Physiol Rev 39: 577-616, 1959
- 142. Conn RB: Normal laboratory values of clinical importance -- normal hematologic values, in Beeson PB, McDermott W (eds): Cecil-Loeb Textbook of Medicine, ed 13. Philadelphia, WB Saunders Co, 1971

VII. APPENDIX I

METHOD FOR

SAMPLING AND ANALYTICAL PROCEDURES

FOR DETERMINATION OF BENZENE

The following sampling and analytical method for analysis of benzene in air employs adsorption on charcoal, followed by desorption, and gas chromatographic measurement. This is a modified method derived from White et al [136] and Kupel and White. [137] Additional data are contained in Part IV under Sorbability of Benzene on Charcoal and Accuracy and Precision Data.

Atmospheric Sampling

(a) Equipment Used

The sampling train is composed of a charcoal tube, a vacuum pump, and a flowmeter. A personal sampler pump or a dependable hand pump, eg, a detector tube pump may be calibrated to produce the desired volume of air.

(b) Calibration of Sampling Instruments

Air sampling instruments may be calibrated with a wet test meter or other suitable reference over a normal range of flowrates and pressure drops. The calibration is conducted at least annually and at any time following repairs or modifications to the sampling system. Similarly, wet test meters should be calibrated upon procurement, at least annually, and after each repair. Calibration curves shall be established for each sampling pump and shall be used in adjusting the pumps prior to field use. The volumetric flowrate through the sampling system shall be spot checked

and the proper adjustments made before and during each study to assure obtaining accurate airflow data.

(1) Flowmeter Calibration Test Method

- (A) With the wet test meter in a level position, check to ascertain that the water level just touches the calibration point on the meter. If the water level is low, add water 1 to 2 F warmer than room temperature to the fill point and run the meter for 30 minutes before calibration.
- (B) Check the voltage of the pump battery with a voltmeter to assure adequate voltage for calibration. Charge the pump battery if needed.
- (C) Break the tips of a charcoal tube to produce openings of a least 2 mm in diameter.
- (D) Assemble the calibration train in series, with the test meter, then the charcoal tube, and finally the pump.
- (E) Turn the pump on, adjusting the rotameter float to a selected reading on the rotameter scale. Wait until the float indicates a steady reading.
- (F) The pointer on the meter should turn clockwise and indicate a pressure drop of not more than 1.0 inch of water. Operate the system for 10 minutes before starting the calibration. If the pressure is greater, recheck the system.
- (G) Data for the calibration include the serial number; meter reading, start and finish; starting time, finish time, and elapsed time; air temperature; barometric pressure; serial number of the

pump and rotameter; the name of the person performing the calibration; and the date.

- (H) Adjust the rotameter float to at least 3 other readings and record the pertinent data in step G at each reading.
- (I) Correct the readings to standard conditions of pressure and temperature by means of the gas law equation.
- (J) Use graph paper to plot the actual airflow and the rotameter readings. Determine the rotameter reading which will result in a l liter/minute flowrate for the pump being calibrated.

(c) Sampling Procedure

The equipment should be set up in a proper locale. The tips of the charcoal tube are broken off producing openings of at least 2 mm in diameter; the filled end of the tube is inserted toward the pump. The tube should always be in a vertical position during sampling. The pump is started and a 10-liter sample is taken at a flowrate of 1 liter/minute. Slower flowrates may be used to lengthen the sampling period but the 1 liter/minute rate should not be exceeded. After the sample is taken, each end of the tube should be capped (plastic caps are provided with commercial tubes). The samples will remain stable for at least 2 weeks which permits shipment for analysis; however, samples should be analyzed as soon as possible in keeping with good laboratory practices.

Analytical

(a) Principle of the Method

A known volume of air is drawn through a charcoal tube to trap the organic vapors present. The charcoal in the tube is transferred to a small

test tube and desorbed with carbon disulfide and an aliquot of the desorbed sample is injected into a gas chromatograph. The area of the resulting peak is determined and compared with areas obtained from the injection of standards.

(b) Range and Sensitivity

The lower limit for benzene with instrument attenuation and splitter techniques is 0.01 mg for each sample. This value can be lowered by reducing the attenuation or by eliminating the splitter. The upper limit value for benzene is 6.0 mg/sample. This value is the number of milligrams of benzene which the front section will collect before a significant amount passes to the backup section. The charcoal tube consists of 2 sections of activated charcoal separated by a section of urethane foam [see description in (f)(2)]. If a particular atmosphere is suspected of containing a large amount of contaminant, it is recommended that a smaller than normal sampling volume be taken.

(c) Interferences

- (1) When the amount of water in the air is so great that condensation actually occurs in the tube, organic vapors will not be trapped. Only water present as a mist is a problem, not water vapor.
- (2) Any compound with the same retention time in the gas chromatograph as benzene at the operating conditions described in this method could be considered an interference. This type of interference can be overcome by changing the operating conditions of the instrument.

(d) Accuracy and Precision

The accuracy and precision determined by a representative laboratory test with benzene (see also <u>Accuracy and Precision Data</u> in Part IV) was found to be:

	Accuracy	Precision
Motor driven laboratory pump	7.6%	4.2%
Approved coal mine personal		
sampling pump (calibrated		
with no in-line resistance)	13.6%	10.1%
•		•
Approved coal mine personal		
sampling pump (calibrated		
with charcoal tube in line)	8.8%	11.6%

The accuracy includes single-day systematic error by 1 operator. Precision represents the single-day accuracy on several different tubes and includes tube-to-tube deviation under controlled laboratory conditions. [138]

(e) Advantages and Disadvantages of the Method

The sampling device is small, portable, and involves no liquids: one basic method is provided for determining many different organic solvents. Interferences are minimal and most can be eliminated by altering chromatographic conditions. In addition, the analysis is accomplished using a rapid instrumental method.

One disadvantage of the method is that the amount of sample which can be obtained is limited by the amount of benzene which the tube will hold before overloading as indicated by benzene recovery at the outlet end of the tube. Also, the precision is limited by the reproducibility of the pressure drop across the tubes, which affects the flowrate, thus causing the volume to be imprecisely measured.

(f) Apparatus consists of:

- (1) An approved coal mine dust personal sampling pump or any vacuum pump whose flow can accurately be determined at 1 liter/minute or less for an area sample.
- (2) Charcoal tubes: Glass tubes with both ends flame-sealed, 7 cm long with a 6-mm O.D. and a 4-mm I.D., containing two sections of 20/40 mesh activated charcoal separated by a 2-mm portion of urethane foam. The absorbing section contains 100 mg of charcoal, the backup section, 50 mg. A 3-mm portion of urethane foam is placed between the outlet end of the tube and the backup section. A plug of glass wool is placed in front of the absorbing section. The pressure drop across the tube must be less than 1 inch of mercury at a flowrate of 1 liter/minute. Tubes with the above specifications are commercially available.
- (3) Gas chromatograph equipped with a flame ionization detector.
- (4) Column (20 ft x 1/8 in) with 10% FFAP stationary phase on 80/100 mesh acid washed DMCS Chromosorb W solid support.
- (5) A mechanical or electronic integrator or a recorder and some method for determining peak area.
 - (6) Small glass-stoppered test tubes or equivalent tubes.

(7) Syringes: 10 μ liter syringe, and other convenient sizes for preparation of standards.

(g) Reagents

- (1) Spectroquality carbon disulfide
- (2) Benzene, preferably chromatoquality grade.
- (3) Bureau of Mines Grade A helium.
- (4) Prepurified hydrogen.
- (5) Filtered compressed air.

(h) Procedure

(1) Cleaning of Equipment

All equipment used for the laboratory analysis should be washed in detergent followed by tap and distilled water rinses.

(2) Collection and Shipping of Samples

Both ends of the charcoal tube are broken to provide openings of at least 2 mm (one-half the I.D. of the tube). The smaller section of charcoal in the tube is used as a backup section and is, therefore, placed nearest the sampling pump. Tubing may be used to connect the back of the tube to the pump, but no tubing must ever be placed on the front of the charcoal tube. Because of the high resistance of the charcoal tube, the sampling method places a heavy load on the personal sampling pump; therefore, it should not be assumed that the pump will run a full 8 hours without a recharging of the battery.

One or more charcoal tubes serving as blanks are treated in the same manner as the sample tubes (break, seal, ship) except that no air is drawn through them.

If bulk samples are submitted in addition to charcoal tubes, they are to be shipped in a separate container.

(3) Analysis of Samples

(A) Preparation

Each charcoal tube is scored with a file and broken open in front of the first section of charcoal. The glass wool is removed and discarded, the charcoal in the first (larger) section is transferred to a small stoppered test tube, the foam separating section is removed and discarded, and the second section is transferred to another test tube. The two charcoal sections are then analyzed separately.

(B) Desorption

Prior to analysis, 0.5 ml of carbon disulfide is pipetted into each test tube to desorb the benzene from the charcoal. Desorption is complete in 30 minutes if the sample is stirred occasionally.

EXTREME CAUTION MUST BE EXERCISED AT ALL TIMES WHEN USING CARBON DISULFIDE BECAUSE OF ITS HIGH TOXICITY AND FIRE AND EXPLOSION HAZARDS. IT CAN BE IGNITED BY HOT STEAM PIPES. ALL WORK WITH CARBON DISULFIDE MUST BE PERFORMED UNDER AN EXHAUST HOOD.

- (C) Gas chromatographic conditions

 Typical operating conditions for a gas chromatograph

 are:
 - (i) 85 cc/min (70 psig) helium carrier gas flow.
 - (ii) 65 cc/min (24 psig) hydrogen gas flow to detector.
 - (iii) 500 cc/min (50 psig) airflow to detector.
 - (iv) 200 C injector temperature.
 - (v) 200 C manifold temperature (detector).

- (vi) 90 C oven temperature isothermal.
- (vii) Use either dual column differential operation or uncompensated mode.

(D) Injection

To eliminate difficulties arising from blowback or distillation within the syringe needle, the solvent flush injection technique is employed to inject the sample into the gas chromatograph. The $10-\mu$ 1 syringe is first flushed with solvent several times to wet the barrel and plunger, then 3 μ 1 of solvent is drawn into the syringe to increase the accuracy and reproducibility of the injected sample volume. Next, the needle is removed from the solvent and the plunger is pulled back about 0.2 μ l to separate the solvent flush from the sample with an air pocket to be The needle is then immersed in the sample and a $5-\mu 1$ used as a marker. aliquot is withdrawn. Prior to injection in the gas chromatograph, the plunger is pulled back a short distance to minimize sample evaporation from the needle tip. Duplicate injections should be made of each sample and the No more than a 3% difference should result in the peak areas standard. that are recorded.

(E) Measurement of area

The area of the sample peak is measured by an electronic integrator or some other suitable form of area measurement and preliminary sample results are read from a standard curve prepared as outlined below.

(i) Standards Preparation and Desorption Efficiency

(1) Preparation of Standards

It is convenient to prepare standards in terms of mg/ 0.5 ml of carbon disulfide because this is the quantity used for benzene desorption from the charcoal. To prepare a 0.3 mg/ 0.5 ml standard, 6.0 mg of benzene (converted to microliters for easy measurement) is injected into exactly 10 ml of carbon disulfide in a glass-stoppered flask. The excess quantity of benzene is used to minimize error due to carbon disulfide volatility. A series of standards is then prepared, varying in concentration over the desired range, and analyzed under the same gas chromatographic conditions and during the same time period as the unknown samples. Curves are established by plotting concentration vs average peak area.

(2) Determination of Desorption Efficiency

The desorption efficiency, ie, the percentage of benzene desorbed from the charcoal, is determined only once, provided the same batch of charcoal is always used.

Activated charcoal, equivalent to the amount in the first section of the sampling tube (100 mg), is measured into a 2-in, 4-mm I.D. glass tube, flame-sealed at one end, and capped with a paraffin film or equivalent at the open end. A known volume of benzene, usually equivalent to that present in a 10-liter sample at a concentration equal to the federal standard, is injected directly into the activated charcoal with a microliter syringe and the tube again capped with more paraffin film. A minimum of 5 tubes are prepared in this manner and allowed to stand for at least 1 day to assure complete adsorption of the benzene onto the charcoal.

These tubes are desorbed and analyzed in exactly the same manner as the sampling tubes.

The results of each analysis are compared to the standards to determine the average percentage (desorption efficiency) that is desorbed. The desorption efficiency is then used as a factor in all sample analyses. The desorption efficiency, determined in this manner, has been shown to be essentially the same as that obtained by analysis of a known amount of benzene vapor trapped on the charcoal and the determined value, therefore, is used because of its simplicity. Each laboratory should determine its own desorption efficiency. For comparison purposes, NIOSH determined a value of 96% for benzene on one batch of charcoal.

(j) Calculations

- (1) Read the weight in milligrams corresponding to each peak area from the standard curve. No correction is necessary for the volume injected, since it is the same for both the sample determination and the standard curve.
- (2) The weight of benzene on the front section of the blank is subtracted from the weight determined for the front section of each sample; a similar procedure is followed for the backup sections. Amounts present on the front and backup sections of the same tube are then added together to determine the total amount detected in the sample. This total weight is then divided by the desorption efficiency to determine the corrected total number of milligrams in the sample. Milligrams are converted into ppm by volume in the air sampled by the following equation at 25 C and 760 mm Hg:

ppm = $\frac{24,450 \text{ ml/mole x mg/liter}}{\text{molecular wt}}$

For a 10-liter air sample of benzene:

ppm =
$$\frac{24,450 \text{ ml/mole x mg in sample/10 liters}}{78.11 \text{ g/mole}}$$

 $ppm = 31.30 \times mg \text{ in sample}$

VIII. APPENDIX II

METHODS FOR DETERMINATION OF

EXPOSURE AREAS TO BENZENE

Estimation of Concentration with Detector Tubes

(a) Atmospheric Sampling

(1) Equipment Used

A typical sampling train consists of a detector tube with a corresponding sampling pump. A specific manufacturer's pump may only be used with his detector tubes.

(2) Sampling Procedures

A specific procedure depends on the manufacturer's instructions but normally consists of breaking both tips off a detector tube, inserting the tube into the pump, and taking a specific number of strokes with the pump.

(3) Handling and Shipping of Samples

Detector tubes are not stable with time; the stain in some tubes fades in a few minutes. The tubes should be read immediately in accordance with the manufacturer's instructions and charts; no attempt should be made to save the used tubes.

(b) General Principles

Gas detector tubes contain a chemically impregnated packing which indicates the concentration of a contaminant in the air by means of a chemically produced color change. The color changes are not permanent or stable, so the stained tubes must be read immediately after the samples are taken. The length of stain or the color intensity is read according to the

manufacturer's instructions. This may involve comparing the stain with a chart, a color comparator, or a direct concentration reading from calibration marks on the tube. Detailed descriptions are provided by individual manufacturer's instructions.

Tubes obtained from commercial sources which bear the certified seal of NIOSH are considered to adhere to the requirements as specified for Approval of Gas Detector Tube Units in 42 CFR Part 84 (37 F.R. 19643). A user may perform his own calibration on commercially acquired tubes by generating accurately known concentrations of benzene in air and correlating concentration with stain length or color intensity.

(c) Range and Sensitivity

Certification standards require that certified tubes have a range from 1/2-5 times the time-weighted average concentration. The sensitivity varies with tube brands.

(d) Interferences

Interferences vary with tube brands. The manufacturer's instructions must be consulted.

(e) Accuracy

Certification standards by NIOSH under the provisions of 42 CFR Part 84 (37 F.R. 19643) specify reliability to within ±25% of the actual concentration in the range 0.75-5 times the standard and ±35% in the range from 0.5 up to, but not including, 0.75 times the standard.

(f) Advantages and Disadvantages

Unlike the charcoal tube method, the use of detector tubes (and portable instruments) is relatively inexpensive and rapid; there is far less time lag than that experienced with laboratory analytical results.

Rapid detecting units are valuable for determining whether a hazardous condition exists at a given location so that workers may be evacuated or suitable protective devices provided. In addition, industrial operators and process engineers need inexpensive and rapid tools for day-to-day evaluation of the atmospheric levels in a work area.

The accuracy of detector tubes is limited; at best they give only an indication of the contaminant concentration. In evaluating measurements performed with detector tubes, interferences, difficulty of end-point readings, and possible calibration inaccuracies must all be considered.

Measurement with Portable Instruments

(a) Atmospheric Sampling

(1) Equipment Used

Two classifications of portable meters that are applicable to atmospheric sampling are direct reading instruments and analytical instruments. Combustible gas meters and flame ionization meters are portable, direct reading instruments; portable variable-path infrared analyzers and gas chromatographs are both field analytical instruments. Any of the 4 meters mentioned are acceptable for benzene determinations if they are properly calibrated before use.

(2) Sampling Procedures

The most important sampling step is the meter calibration. Careful calibration must be performed either in the laboratory prior to onsite use or in the field using a container of specific benzene concentration. If calibration charts are inaccurate, erroneous readings will be made.

The actual field sampling is conducted according to the manufacturer's instructions. Readings should be corrected if necessary for variables such as temperature, humidity, atmospheric pressure, etc, and recorded along with time, place, temperature, etc.

(b) General Principles

Analysis is dependent on the type of meter used. The portable direct reading meters require no analysis because they usually provide usable concentration readings directly. Results obtained from the variable-path infrared analyzer and the gas chromatograph must be recorded, further analyzed, and compared with standards to obtain concentration values.

(c) Range and Sensitivity

The range and sensitivity vary with the instrument used; in general, the portable analysis meters are more sensitive than direct reading units.

(d) Interferences

Again, these vary with the instrument used. Water vapor or combustible gases interfere with benzene identification using combustible gas meters. Mixtures of any carbon containing compounds, other than benzene, will interfere in flame ionization determinations.

(e) Advantages and Disadvantages

The benefits and drawbacks of portable instruments are essentially the same as for detector tubes discussed previously. Where recording capability is possible, direct reading instruments have the advantage of continuous record availability.