

## Detection of Rabbit anti-VEGF in paraffin-embedded rat tissue

### Antibody Information:

Vector Goat Elite Kit  
Vector Laboratories, Inc.  
Burlingame, CA 94010  
[www.vectorlabs.com](http://www.vectorlabs.com)  
1-800-227-6666  
Catalog #

Contains everything needed to make blocking reagent, secondary antibody and label antibody

Avidin/Biotin Blocking Kit  
Vector Laboratories, Inc.  
Burlingame, CA 94010  
[www.vectorlabs.com](http://www.vectorlabs.com)  
1-800-227-6666  
Catalog #SP-2001

Normal Goat Serum  
Jackson ImmunoResearch Laboratories, Inc.  
West Grove, PA 19390  
[www.jacksonimmuno.com](http://www.jacksonimmuno.com)  
1-800-367-5296  
Catalog #PK-6105

Primary antibody: (VegF (A-20-G), goat polyclonal)  
Santa Cruz Biotechnology, Inc.  
Santa Cruz, CA 95060  
[www.scbt.com](http://www.scbt.com)  
1-800-457-3801  
Catalog #sc-152-G  
Concentration: 200ug/ml

Microwave Information  
Panasonic High Power 1200W  
Model number NN-S732WL  
Conditions for retrieval dependent upon this microwave.

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### Staining Procedure

-Positive Control Tissue: Islets of Langerhans (pancreas)  
-Stain localization: Cell Membrane

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Deparaffinize and hydrate slides through the following solutions.

Xylene	2 times	5 minutes
100% EtOH	2 times	3 minutes
95% EtOH	2 times	3 minutes
1X Automation Buffer	2 times	5 minutes

1. Quench endogenous peroxidase by placing slides in 3% hydrogen peroxide for 15 minutes.

2. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.

3. Perform Heat Induced Epitope Retrieval using MWO

Place a full rack of slides in Tissue Tek<sup>®</sup> container containing 200 mls 1X citrate buffer.

MWO for 5 minutes at level 3

Cool for 1 minute (Add 50 mls citrate buffer to container)

MWO for 5 minutes at level 3 Temp. \_\_\_\_\_

Cool 20 minutes at room temperature

Rinse in distilled water 3 X 2 minutes each

Place slides in buffer for 5 minutes

4. Apply block from Vector Goat Elite kit and incubate for 20 minutes.

Lot# \_\_\_\_\_ Exp. Date \_\_\_\_\_

5. Apply Avidin/Biotin block

Lot# \_\_\_\_\_ Exp \_\_\_\_\_ New Kit yes / no

Apply avidin block - 15 min @ RT.

Quick rinse in 1X AB.

Apply biotin block - 15 min @ RT.

No wash, wipe excess block and apply primary antibody

**DO NOT RINSE SLIDES WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY.**

6. Apply primary antibody VEGF at the following dilutions and incubate for 1 hour at RT.

Lot# \_\_\_\_\_ Exp Date \_\_\_\_\_

1:1000

For negative control slides, normalize the protein concentration of normal goat serum to the protein concentration of the primary antibody (VEGF) and make the following dilutions. Apply normal goat serum to the slides and incubate for one hour.

Lot# \_\_\_\_\_ Reconstituted Date \_\_\_\_\_

1:1000

7. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
8. Apply secondary from Vector Goat Elite kit and incubate for 30 minutes.
9. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
10. (Prepare Label antibody 30 minutes prior to use)  
Apply Label antibody from Vector Goat Elite Kit for 30 minutes. 2 drops Reagent A + 5 ml diluent -> Mix and then add 2 drops Reagent B
11. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
12. Apply liquid Dako DAB Chromagen for 6 minutes in the dark.  
(Add 1 drop of DAB per ml of substrate)  
Lot # \_\_\_\_\_ Exp. Date \_\_\_\_\_  
New Kit yes / no
13. Rinse in tap water 3 minutes.
14. Counterstain with Modified Harris Hematoxylin for 20 seconds.
15. Rinse in tap water until water is clear.
16. Place slides in 1X AB for 1 minute to blue.
17. Dehydrate through the following solutions.

95% Ethanol	1 change	3 minutes
100% EtOH	3 changes	3 minutes
Xylene	2 changes	5 minutes

18. Coverslip

updated 8/8/2003

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