

# **Identification of Neurofilament in Formalin-Fixed, Paraffin-Embedded Rodent Tissue**

## **Reagents:**

[1X Automation Buffer](#)

[3% Hydrogen Peroxide](#)

[Antibody Diluent](#)

[Citrate Buffer](#)

[DAB Chromagen](#)

[Hematoxylin](#)

## **Antibody Information**

Dako LSAB2 System HRP

Dako Corporation

Carpinteria CA 93013

[www.dakousa.com](http://www.dakousa.com)

1-800-235-5763

Code No. K0609

\*This kit includes reagents needed for link and label antibodies.

Primary antibody: Mouse anti-Neurofilament Protein

Dako Corporation

Carpinteria CA 93013

[www.dakousa.com](http://www.dakousa.com)

1-800-235-5763

Code No. M0762

Negative control serum: Normal Mouse Serum

Jackson ImmunoResearch Laboratories, Inc.

West Grove, PA 19390

[www.jacksonimmuno.com](http://www.jacksonimmuno.com)

1-800-367-5296

Catalog #015-000-001

## Staining Procedure

- Positive Control Tissue: brain
- Stain Localization: cytoplasmic

Deparaffinize and hydrate slides through the following solutions.

Xylene	2 times	5 minutes
100% EtOH	2 times	3 minutes
95% EtOH	2 times	3 minutes
1X Automation Buffer	2 times	5 minutes

1. Quench endogenous peroxidase by placing slides in 3% hydrogen peroxide for 15 minutes.
2. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
3. Unmasking Techniques using the decloaker.  
Add 500 ml D/W to the pan of the decloaker.  
Place full rack of slides in 200 ml of 1X citrate buffer and place in the decloaker.  
Decloak for 5 minutes. Pressure \_\_\_\_\_  
Depressurize for 10 minutes.  
Remove pan top and cool for 10 min. Temp \_\_\_\_\_  
Rinse in D/W, 2x for 3 min each

NO BLOCK REQUIRED

4. Apply primary antibody (Neurofilament) at 1:100 dilution and incubate for 10 mins.  
Lot# \_\_\_\_\_ Exp Date \_\_\_\_\_

For negative control slides, normalize the protein concentration of normal mouse serum to the protein concentration of the primary antibody (Neurofilament) and use this to make the 1:100 dilution for 10 min.

Lot# \_\_\_\_\_ Reconstituted Date \_\_\_\_\_

5. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.

LSAB2 Kit Lot# \_\_\_\_\_ Exp Date \_\_\_\_\_

6. Apply Link antibody (Yellow bottle) from LSAB2 kit for 10 minutes.
7. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
8. Apply label antibody (Red bottle) from LSAB2 kit for 10 minutes
9. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
10. Apply liquid Dako DAB Chromagen for 6 minutes in the dark.  
(Add 1 drop of DAB per ml of substrate)  
Lot# \_\_\_\_\_ Exp Date \_\_\_\_\_ New Kit: yes / no
11. Rinse in tap water 3 minutes.
12. Counterstain with Modified Harris Hematoxylin for 30 seconds.
13. Rinse in tap water until water is clear.
14. Place slides in 1X Automation Buffer for 1 minute with gentle agitation to blue slides.
15. Dehydrate through the following solutions.

95% Ethanol	1 change	3 minutes
100% EtOH	3 changes	3 minutes
Xylene	2 changes	5 minutes

16. Coverslip.  
updated 07/29/04