

Detection of HDAC2 in Formalin-Fixed, Paraffin-Embedded Human Tissue

Reagents:

[1X Automation Buffer](#)
[3% Hydrogen Peroxide](#)
[Antibody Diluent](#)
[Citrate Buffer](#)
[DAB Chromagen](#)
[Hematoxylin](#)

Antibody Information:

Kit: Mouse IgG Elite Kit
Vector Laboratories, Inc.
Burlingame, CA 94010
www.vectorlabs.com
1-800-227-6666
Catalog # PK-6102

Note: The Vector Mouse Elite Kit contains solutions needed to make the block, secondary, and label reagents.

Avidin Biotin Blocking Kit
Vector Laboratories, Inc.
Burlingame, CA 94010
www.vectorlabs.com
1-800-227-6666
Catalog # SP-2001

Primary antibody: Mouse anti-HDAC2 (C-8) Monoclonal Antibody
Santa Cruz Biotechnology, Inc.
Santa Cruz, CA 95060
www.scbt.com
1-800-457-3801
Catalog # sc-9959

Negative control serum: Normal Mouse Serum
Jackson ImmunoResearch Laboratories, Inc.
West Grove, PA 19390
www.jacksonimmuno.com
1-800-367-5296
Catalog # 015-000-001

Staining Procedure

Positive Control Tissue: Breast carcinoma and teratoma

Stain Localization: Nuclear

Deparaffinize and hydrate slides through the following solutions.

Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Automation Buffer	2 times	5 minutes

1. Quench endogenous peroxidase by placing slides in 3% hydrogen peroxide for 15 minutes.
2. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
3. Unmasking technique using the decloaker.
Add 500 ml distilled water to the pan of the decloaker.
Place full rack of slides in 200 ml of 1X citrate buffer and place in the decloaker.
Decloak for 5 minutes. Pressure _____
Depressurize for 10 minutes.
Remove pan top and cool for 10 minutes. Temperature before cooling _____
Rinse in distilled water twice for 3 minutes each.
4. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
5. Apply blocking solution from the Vector Mouse Elite Kit and incubate for 20 minutes at room temperature.
Exp. Date _____ New Kit: yes / no

DO NOT RINSE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.

6. Apply the Avidin Biotin Blocking Kit
Lot# _____ Exp Date _____ New Kit: yes / no
Apply avidin block - 15 minutes at room temperature.
Quick rinse in 1X Automation Buffer.
Apply biotin block - 15 minutes at room temperature.
No wash, wipe excess block and apply primary antibody

DO NOT RINSE SLIDES WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY.

7. Apply primary antibody (HDAC2) at a 1:10 dilution and incubate for one hour at room temperature
Lot# _____ Exp Date _____

For negative control slides, normalize the protein concentration of the normal mouse serum to the protein concentration of the primary antibody (HDAC2). Use this to make a 1:10 dilution. Apply to slides and incubate for one hour at room temperature.
Lot# _____ Reconstituted Date _____

- 8. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
- 9. Apply secondary antibody from Vector Mouse Elite Kit and incubate for 30 minutes at room temperature.
- 10. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
- 11. Apply label antibody from Vector Mouse Elite Kit and incubate for 30 minutes at room temperature.
- 12. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
- 13. Apply liquid Dako DAB Chromagen for 6 minutes in the dark.
(Add 1 drop of DAB per ml of substrate)
Lot# _____ Exp Date _____ New Kit yes / no
- 14. Rinse in tap water 3 minutes.
- 15. Counterstain with Modified Harris Hematoxylin for 20 seconds.
- 16. Rinse in tap water until water is clear.
- 17. Gently agitate slides in 1X Automation buffer until they turn blue.
- 18. Dehydrate through the following solutions.

95% Ethanol	1 change	3 minutes
100% Ethanol	3 changes	3 minutes
Xylene	2 changes	5 minutes

19. Coverslip

Updated 12/15/06