Detection of MAdCAM-1 in Formalin Fixed Paraffin-Embedded Mouse Tissue

Reagents:

1X Automation Buffer
3% Hydrogen Peroxide
Antibody Diluent
Citrate Buffer
DAB Chromagen
Hematoxylin

Antibody Information

Blocking Serum: Normal Horse Serum
Jackson Immunoresearch Laboratories, Inc.
West Grove, PA 19390
www.jacksonimmuno.com
1-800-367-5296
Catalog # 008-000-001

Avidin/Biotin Blocking Kit Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # SP-2001

Primary antibody: Goat anti-mouse mMAdCAM-1 Antibody R&D Systems Minneapolis, MN 55413 www.rndsystems.com 1-800-343-7475 Catalog # AF993

Negative Serum Control: Normal Goat Serum Jackson Immunoresearch Laboratories, Inc. West Grove, PA 19390 www.jacksonimmuno.com 1-800-367-5296 Catalog #005-000-121

Secondary antibody: Biotinylated horse anti-goat IgG Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # BA-9500

<u>Label antibody: Peroxidase-conjugated Streptavidin SS Label</u>

Biogenex

San Ramon, CA 94583

www.biogenex.com

1-800-421-4149

Catalog # HK330-9K

Staining Procedure

Positive Control Tissue: Spleen

Stain Localization: Cytoplasmic (primarily around the germinal centers)

Deparaffinize and hydrate slides through the following solutions.

| Xylene | 2 times | 5 minutes |
|----------------------|---------|-----------|
| 100% Ethanol | 2 times | 3 minutes |
| 95% Ethanol | 2 times | 3 minutes |
| 1X Automation Buffer | 2 times | 5 minutes |

- 1. Quench endogenous peroxidase by placing slides in 3% hydrogen peroxide for 15 minutes.
- 2. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.

| 3 | . Unmasking technique using the decloaker. |
|---|--|
| | Add 500 ml distilled water to the pan of the decloaker. |
| | Place full rack of slides in 200 ml of 1X citrate buffer and place in the decloaker. |
| | Decloak for 5 minutes. Pressure |
| | Depressurize for 10 minutes. |
| | Remove pan top and cool for 10 minutes. Temperature before cooling |
| | Rinse in distilled water twice for 3 minutes each. |

- 4. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
- 5. Block with 10% Normal Horse Serum for 20 minutes at room temperature. Lot#______ Reconstituted Date______

DO NOT RINSE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.

| Apply avidin b | Exp Date New Kit: yes / no lock - 15 minutes at room temperature. |
|--|--|
| | 1X Automation Buffer. |
| | lock - 15 minutes at room temperature. |
| No wash, wipe | excess block and apply primary antibody |
| DO NOT RINS ANTIBODY. | SE SLIDES WITH BUFFER BEFORE ADDING PRIMARY |
| 7. Apply primary room temperate | antibody (MAdCAM-1) at a 1:500 dilution and incubate for one hourure. |
| _ | Exp Date |
| serum to match use this to make temperature. | ntrol slides, normalize the protein concentration of the normal goat the protein concentration of the primary antibody (MAdCAM-1), and e a 1:500 dilution. Apply to slides and incubate for one hour at room |
| Lot# | Reconstituted Date |
| for 30 minutes | nti-goat secondary antibody at a 1:500 dilution and incubate at room temperature. Reconstituted Date |
| 10. Rinse slides in | n 2 changes of 1X Automation Buffer for 5 minutes each. |
| | nix SS Label and incubate for 30 minutes at room temperature. Exp. Date |
| 12. Rinse slides in | n 2 changes of 1X Automation Buffer for 5 minutes each. |
| (Add 1 drop o | Dako DAB Chromagen for 6 minutes in the dark. of DAB per ml of substrate) Exp. Date New Kit: yes / no |
| 14. Rinse in tap w | vater 3 minutes. |
| 15 0 | with Modified Harris Hamatovylin for 20 seconds |
| 15. Counterstain | with Modified Harris Hematoxylin for 20 seconds. |
| | vater until water is clear. |

18. Dehydrate through the following solutions.

| 95% Ethanol | 1 change | 3 minutes |
|--------------|-----------|-----------|
| 100% Ethanol | 3 changes | 3 minutes |
| Xylene | 2 changes | 5 minutes |

19. Coverslip

Updated 10/17/06