

## 2. HEALTH EFFECTS

### 2.1 INTRODUCTION

The primary purpose of this chapter is to provide public health officials, physicians, toxicologists, and other interested individuals and groups with an overall perspective of the toxicology of sulfur dioxide. It contains descriptions and evaluations of toxicological studies and epidemiological investigations and provides conclusions, where possible, on the relevance of toxicity and toxicokinetic data to public health. A glossary and list of acronyms, abbreviations, and symbols can be found at the end of this profile.

### 2.2 DISCUSSION OF HEALTH EFFECTS BY ROUTE OF EXPOSURE

To help public health professionals and others address the needs of persons living or working near hazardous waste sites, the information in this section is organized first by route of exposure-inhalation, oral, and dermal; and then by health effect-death, systemic, immunological, neurological, reproductive, developmental, genotoxic, and carcinogenic effects. These data are discussed in terms of three exposure periods-acute (14 days or less), intermediate (15-364 days), and chronic (365 days or more).

Levels of significant exposure for each route and duration are presented in tables and illustrated in figures. The points in the figures showing no-observed-adverse-effect levels (NOAELs) or lowest-observed-adverse-effect levels (LOAELs) reflect the actual doses (levels of exposure) used in the studies. LOAELs have been classified into “less serious” or “serious” effects. “Serious” effects are those that evoke failure in a biological system and can lead to morbidity or mortality (e.g., acute respiratory distress or death). “Less serious” effects are those that are not expected to cause significant dysfunction or death, or those whose significance to the organism is not entirely clear. ATSDR acknowledges that a considerable amount of judgment may be required in establishing whether an end point should be classified as a NOAEL, “less serious” LOAEL, or “serious” LOAEL, and that in some cases, there will be insufficient data to decide whether the effect is indicative of significant dysfunction. However, the Agency has established guidelines and policies that are used to classify these end points. ATSDR believes that there is sufficient merit in this approach to warrant an attempt at distinguishing between “less serious” and “serious” effects. The distinction between “less serious” effects and “serious” effects is considered to be important because it helps the users of the profiles to identify levels of exposure at which major health effects start to appear. LOAELs or NOAELs should also help in

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determining whether or not the effects vary with dose and/or duration, and place into perspective the possible significance of these effects to human health.

The significance of the exposure levels shown in the Levels of Significant Exposure (LSE) tables and figures may differ depending on the user's perspective. Public health officials and others concerned with appropriate actions to take at hazardous waste sites may want information on levels of exposure associated with more subtle effects in humans or animals (LOAEL) or exposure levels below which no adverse effects (NOAELs) have been observed. Estimates of levels posing minimal risk to humans (Minimal Risk Levels or MRLs) may be of interest to health professionals and citizens alike.

Levels of exposure associated with carcinogenic effects (Cancer Effect Levels, CELs) of sulfur dioxide are indicated in Table 2- 1 and Figure 2- 1.

Estimates of exposure levels posing minimal risk to humans (Minimal Risk Levels or MRLs) have been made for sulfur dioxide. An MRL is defined as an estimate of daily human exposure to a substance that is likely to be without an appreciable risk of adverse effects (noncarcinogenic) over a specified duration of exposure. MRLs are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration within a given route of exposure. MRLs are based on noncancerous health effects only and do not consider carcinogenic effects. MRLs can be derived for acute, intermediate, and chronic duration exposures for inhalation and oral routes. Appropriate methodology does not exist to develop MRLs for dermal exposure.

Although methods have been established to derive these levels (Barnes and Dourson 1988; EPA 1990), uncertainties are associated with these techniques. Furthermore, ATSDR acknowledges additional uncertainties inherent in the application of the procedures to derive less than lifetime MRLs. As an example, acute inhalation MRLs may not be protective for health effects that are delayed in development or are acquired following repeated acute insults, such as hypersensitivity reactions, asthma, or chronic bronchitis. As these kinds of health effects data become available and methods to assess levels of significant human exposure improve, these MRLs will be revised.

A User's Guide has been provided at the end of this profile (see Appendix B). This guide should aid in the interpretation of the tables and figures for Levels of Significant Exposure and the MRLs.

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**2.2.1 Inhalation Exposure**

Table 2-1 and Figure 2-1 summarize the available quantitative information on the health effects that have been observed in humans and animals following inhalation exposure to sulfur dioxide. All exposure levels are expressed in parts per million (ppm).

**2.2.1.1. Death**

There have been several case reports of human deaths following acute exposure to high concentrations of sulfur dioxide (Atkinson et al. 1993; Charan et al. 1979; Harkonen et al. 1983; Huber and Loving 1991; Rabinovitch et al. 1989). In most studies, concentrations were not measured. In one study, analysis of gas samples at the time of rescue showed sulfur dioxide concentrations greater than 40 ppm (Rabinovitch et al. 1989). A sulfur dioxide level of 150 ppm was measured during the reenactment of an incident in which a 76-year-old asthmatic woman died of an asthma attack after inhaling vapors from a sulfite-based derusting agent used in her dishwasher (Huber and Loving 1991). Actual sulfur dioxide levels were probably higher since the quantity of derusting agent used in the investigation was approximately 7-10% of the amount originally used by the woman. A concentration of 100 ppm is considered immediately dangerous to life and health (HSDB 1998).

Excess mortality among humans occurred following exposure to high concentrations of sulfur dioxide and suspended particulate matter during acute smog episodes in London during the 1950s (Amdur et al. 1991; Mazumdar et al. 1982; WHO 1979). Peak daily concentrations as high as 4,000  $\mu\text{g}/\text{m}^3$  (1.5 ppm) sulfur dioxide and 6,000  $\mu\text{g}/\text{m}^3$  of smoke were recorded during some of the pollution episodes. The available evidence suggested that excess mortality may occur at sulfur dioxide levels at or above 500  $\mu\text{g}/\text{m}^3$  (0.2 ppm) (24-hour mean concentration) in combination with elevated particle levels. The increases in mortality were attributed to bronchitis and other causes of impairment of respiratory function (WHO 1979). Increased mortality from heart disease was also noted. The mortalities occurred mainly among the elderly and among those with pre-existing cardiac and/or respiratory disorders (WHO 1979).

A reanalysis of the mortality and London pollution data revealed that the mortalities were due almost entirely to smoke (Mazumdar et al. 1982). This study applied three types of analyses, including year-by-year multiple regression; stratification using nested quartiles of one pollutant with quantities of the other; and multiple

Table 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation

Key to <sup>a</sup> figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference	
					Less serious (ppm)	Serious (ppm)		
<b>ACUTE EXPOSURE</b>								
<b>Death</b>								
1	Rat (NS)	cont				590	(death in 8 out of 8 rats following an average of 31 hours of exposure)	Cohen et al. 1973
2	Rat (NS)	4hr		593		965	(death in 3 of 8 rats within 2 weeks following exposure)	Cohen et al. 1973
3	Mouse (Swiss Albino)	30 min				3000	(LC <sub>50</sub> )	Hilado and Machado 1977
<b>Systemic</b>								
4	Human-n	10 min	Resp		1-8		(decreased tidal volume; increased respiratory rate)	Amdur et al. 1953
			Cardio		1-8		(increased pulse)	
5	Human-n	1-6 hr	Resp		1M		(increased nasal airflow resistance; reduced FEV <sub>1</sub> and FEF <sub>25-75</sub> )	Andersen et al. 1974
6	Human-n	10-30 min	Resp	1	5		(70-75% increased flow resistance after 1 min; cough, sense of irritation)	Frank et al. 1962

Table 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (continued)

Key to figure <sup>a</sup>	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
7	Human-n	5 min	Resp		0.6-0.8	(100% or more increase in specific airway resistance in 13 of 16 subjects)	Islam et al. 1992
8	Human-n	NS	Resp		1	(increased specific airway resistance following deep inhalation of sulfur dioxide)	Lawther et al. 1975
9	Human-n	10min	Resp		5	(increased airway resistance during quiet mouth breathing)	Lawther et al. 1975
10	Human-n	20 min	Resp		8M	(erythema of trachea and main bronchi; increase in inflammatory cells in bronchoalveolar lavage fluid)	Sandstrom et al. 1989
11	Human-n	20min	Resp		4	(increased number of lysozyme positive macrophages in bronchoalveolar lavage fluid)	Sandstrom et al. 1989
12	Human-a	1, 3, 5 min	Resp			0.5 (3 min exposure: 173% increase in airway resistance; wheezing, chest tightness, dyspnea)	Balmes et al. 1987

Table 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (continued)

Key to figure <sup>a</sup>	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
13	Human-a	5 min	Resp		0.5	(238% increase in airway resistance during exercising)	Bethel et al. 1983b
14	Human-a	3 min	Resp		0.5	(bronchoconstriction and wheezing when also exposed to cold, dry air)	Bethel et al. 1984
15	Human-a	5 min	Resp		0.25	(134% increased exercise-induced bronchoconstriction)	Bethel et al. 1985
16	Human-a	20 min	Resp		1 M	(small increase in total respiratory resistance)	Heath et al. 1994
17	Human-a	10 min	Resp		0.75 M	(median concentration that resulted in a 100% increase in specific airway resistance)	Horstman et al. 1986
18	Human-a	30 min	Resp	0.5			Jorres and Magnussen 1990
19	Human-a	50 min	Resp		0.5	(32% increased nasal work of breathing; 60% increased respiratory resistance; decreased $V_{\max 50\%}$ and $V_{\max 75\%}$ ; 24% decreased $FEV_1$ )	Koenig et al. 1985

Table 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (continued)

Key to figure <sup>a</sup>	Species (strain)	Exposure/duration/frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
20	Human-a	15 min	Resp	0.1			Koenig et al. 1990
21	Human-a	5 min	Resp	0.2	0.4	(34% increased specific airway resistance)	Linn et al. 1983b
22	Human-a	1 hr	Resp	0.2	0.4	(29% increased specific airway resistance; 2% decreased FEV <sub>1</sub> )	Linn et al. 1987
23	Human-a	10-75 min	Resp	0.25 M	0.5 M	(2-3-fold increase in specific airway resistance during exercise compared to controls)	Roger et al. 1985
24	Human-a	40 min	Resp	0.5	0.75	(significant increase in airway resistance; decreases in FEV <sub>1</sub> , V <sub>max50%</sub> , MEF <sub>40%(P)</sub> , and clinical symptoms during moderate exercise)	Schachter et al. 1984
25	Human-a	10 min	Resp		0.1 <sup>b</sup>	(significant increases in airway resistance during moderate exercise)	Sheppard et al. 1981
26	Rat (Wistar)	8 hr	Resp			800 M (loss of cilia and cell necrosis in trachea and main bronchus)	Stratmann et al. 1991

Table 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (continued)

Key to figure <sup>a</sup>	Species (strain)	Exposure/duration/frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
27	Mouse (ICR)	30, 60, 90, 120 min	Resp		20F (degenerative changes to the olfactory epithelium after 16 min of exposure)		Min et al. 1994
28	Gn Pig (NS)	1-3 hr	Resp		2.6 (20% increased resistance and 10% decreased compliance)		Amdur 1959
29	Gn Pig (Hartley)	1 hr	Resp	1 M			Chen et al. 1992b
30	Gn Pig (Perlbright-white)	5d 8hr/d	Resp			5 F (severe destruction of ciliated epithelium and polymorphonuclear infiltrates)	Riedel et al. 1992
31	Dog (NS)	20-40 min	Resp		1.1-141 (5% decreased compliance; 121% increased resistance)		Balchum et al. 1959
32	Dog (NS)	30-40 min	Resp		1.8-148 (8.5% decreased compliance; 150-300% increased resistance)		Balchum et al. 1960b
33	Rabbit (NS)	10-20 min	Resp		200-300F (transient decrease in cough reflex and Hering-Breuer inflation reflex <sup>o</sup> )		Hanacek et al. 1991



Table 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (continued)

Key to figure <sup>a</sup>	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
<b>Immunological/Lymphoreticular</b>							
34	Gn Pig (Perbright-wh ite)	5d 8hr/d			5F (increased sensitization to inhalation exposure to ovalbumin)		Riedel et al. 1992
35	Hamster (Golden Syrian)	4 hr			50M (decreased endocytosis by pulmonary macrophage when exposure occurred during exercise)		Skornik and Brain 1990
<b>Reproductive</b>							
36	Mouse (CF-1)	7h/d; Gd6-15		25 F			Murray et al. 1979, 1977
37	Rabbit (New Zealand)	7h/d; Gd6-18		70 F			Murray et al. 1979, 1977
<b>Developmental</b>							
38	Mouse (CF-1)	7h/d; Gd6-15			25 (reduced fetal weight and increased skeletal variations)		Murray et al. 1979, 1977
39	Mouse (CD-1)	Gd7-18				32 (increased time for the righting reflex and negative geotaxis)	Singh 1989

Table 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (continued)

Key to <sup>a</sup> figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference	
					Less serious (ppm)	Serious (ppm)		
40	Rabbit (New Zealand)	7h/d; Gd6-18			70	(increased skeletal variations)	Murray et al. 1979, 1977	
<b>INTERMEDIATE EXPOSURE</b>								
<b>Systemic</b>								
41	Rat (Buffalo)	12 wk; 5d/wk; 1h/d	Resp		30M	(inflammation of bronchial mucosa)	Krasnowska et al. 1998	
42	Rat (NS)	3hr/d; 5d/wk; 2-42 d	Resp		400	(epithelial necrosis, loss of cilia, and increased numbers and activity of goblet cells)	Lamb and Reid 1968	
43	Hamster (Syrian)	19-74 days; 5 d/wk; 4 hr/d	Resp		650	(dilated bronchi and alveolar ducts; small scattered areas of focal emphysema)	Goldring et al. 1970	
44	Rabbit (New Zealand)	5 wk 6 d/wk 2 hr/d	Resp  Bd Wt				70-300 M (decreased respiratory rate; rhinitis; tracheitis; bronchopneumonia)	Miyata et al. 1990
					70-300M	(body weight gain 25% less than controls)		
<b>Immunological/Lymphoreticular</b>								
45	Gn Pig (Hartley)	6 wk 5 d/wk 4 hr/d					5 M (increased delayed-type dyspneic symptoms after challenge by <i>C. albicans</i> resulting in 3/12 guinea pigs dying)	Kitabatake et al. 1995

Table 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (continued)

Key to <sup>a</sup> figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
<b>Neurological</b>							
46	Mouse (CD-1)	24 d; continuous		12 F	30F (transient changes in the frequency of various activity-exploration behaviors)		Petruzzi et al. 1996
<b>Reproductive</b>							
47	Mouse (CD-1)	24 d; continuous		30			Petruzzi et al. 1996
<b>Developmental</b>							
48	Mouse (CD-1)	24 d, continuous		30			Petruzzi et al. 1996
<b>CHRONIC EXPOSURE</b>							
<b>Systemic</b>							
49	Monkey (Cynomol- gus)	78 wk 7 d/wk 23.3 hr/d	Resp	5.1			Alarie et al. 1975
			Cardio	5.1			
			Hemato	5.1			
			Hepatic	5.1			
			Renal	5.1			
			Bd Wt	5.1			

Table 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (continued)

Key to <sup>a</sup> figure	Species (strain)	Exposure/ duration/ frequency	System	NOAEL (ppm)	LOAEL		Reference
					Less serious (ppm)	Serious (ppm)	
50	Gn Pig (Hartley)	52 wk; 7 d/wk; 22 wk; 22 hr/d	Resp	5.7	5.7 (increase in the size of hepatocytes accompanied by cytoplasmic vacuolation)		Alarie et al. 1972
			Cardio	5.7			
			Hemato	5.7			
			Hepatic	1.0			
			Renal	5.7			
Bd Wt	5.7						
<b>Cancer</b>							
51	Mouse LX	2 yr 5 d/wk 5 min/d				500 (CEL: lung adenomas in 28/58 male and female mice; lung carcinomas in 4/30 females)	Peacock and Spence 1967

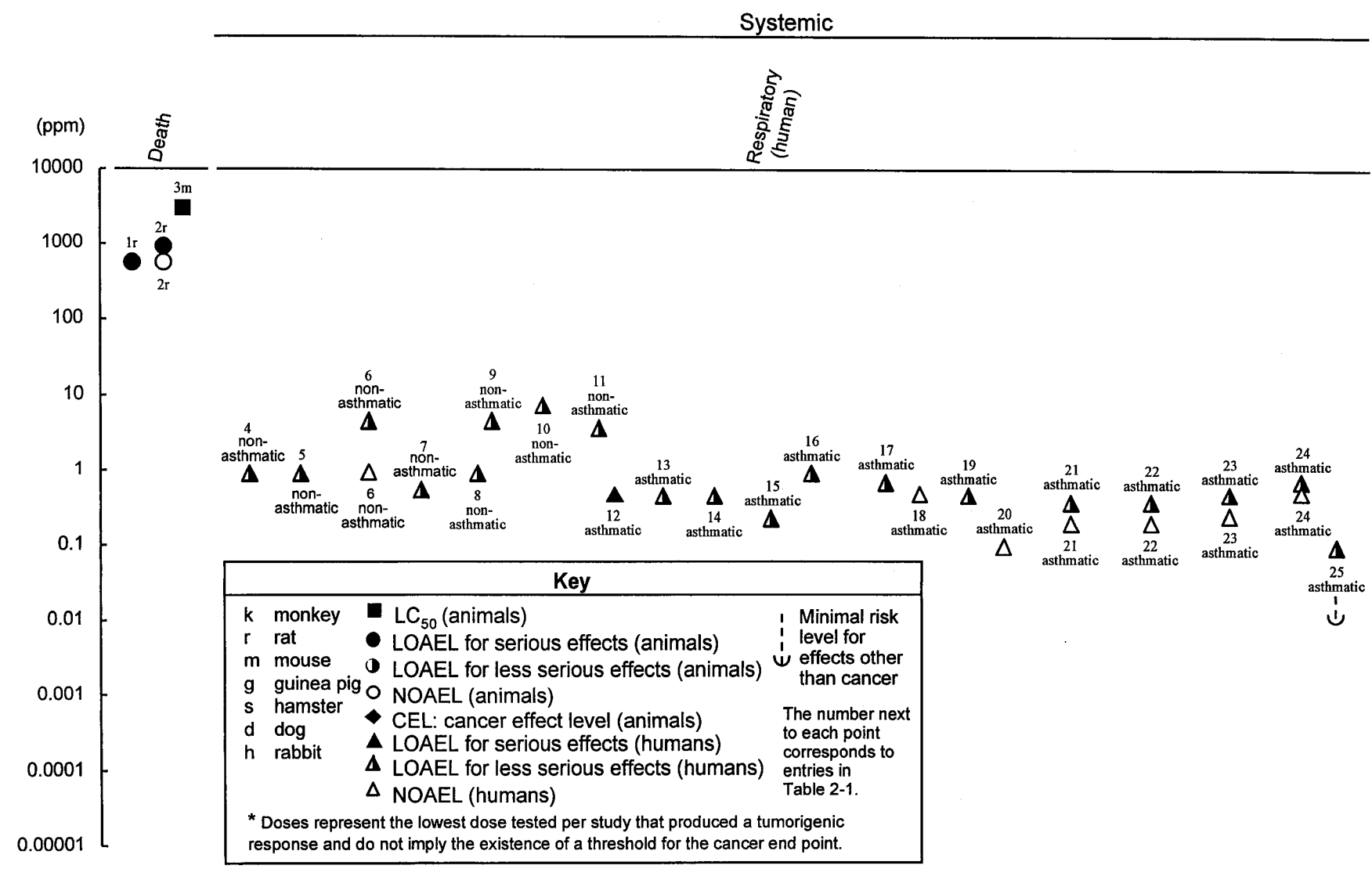
<sup>a</sup>The numbers correspond to entries in Figure 2-1.

<sup>b</sup>Used to derive an acute inhalation Minimal Risk Level (MRL) of 0.01 ppm; the minimal LOAEL of 0.1 ppm was divided by an uncertainty factor of 9 to account for human variability and use of a minimal LOAEL.

<sup>c</sup>The Hering-Breuer reflex involves reaction of the lung to inflation. Inflation of the lung inhibits inspiration and brings on expiration.

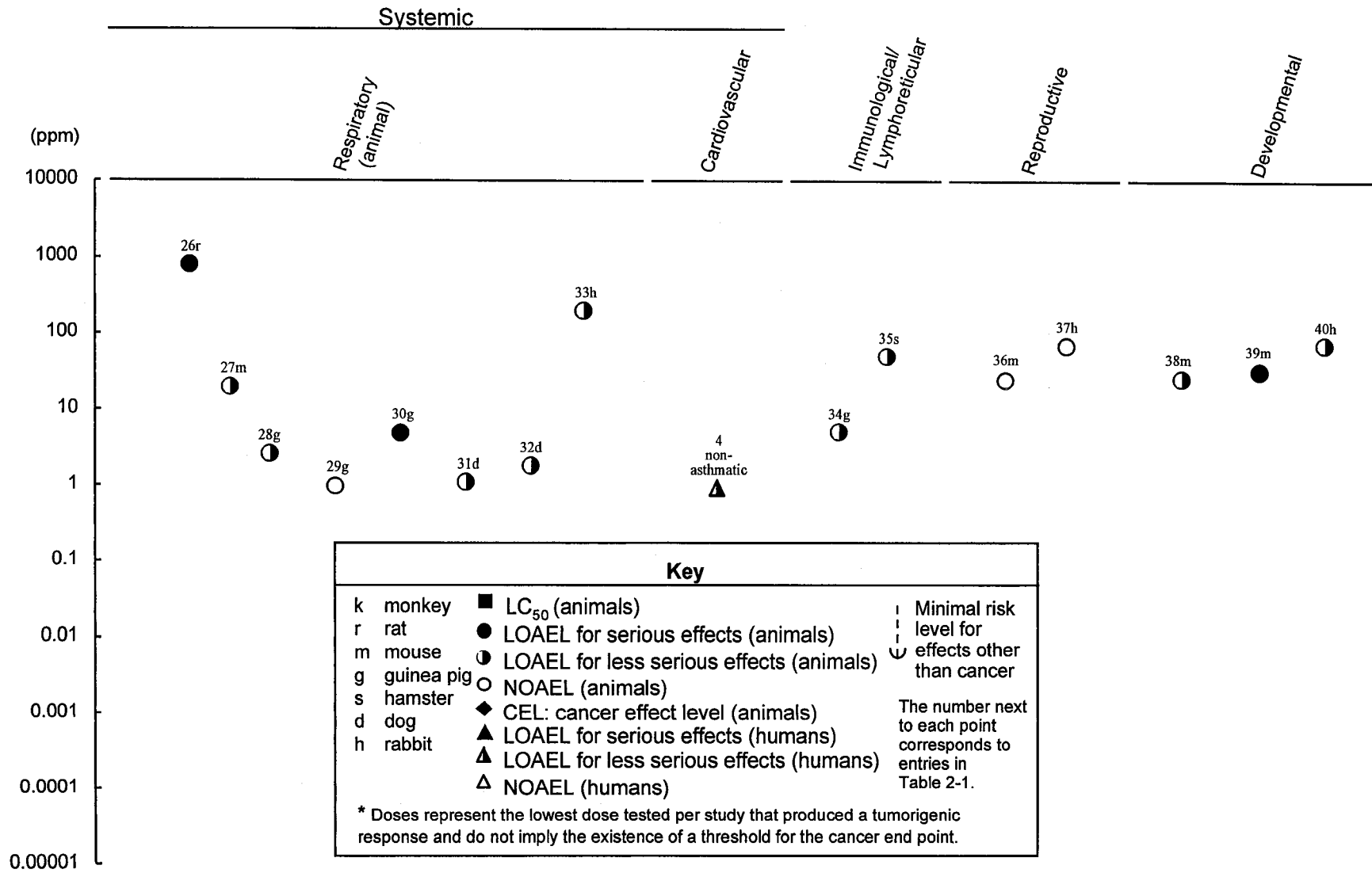
Bd Wt = body weight; Cardio = cardiovascular; CEL = cancer effect level; d = day(s); F = female; FEF = forced expiratory flow during the middle half of the expired volume; FEV<sub>1</sub> = forced expiratory volume in one second; Gd = gestation day; Gn Pig = guinea pig; Hemato = hematological; hr = hour(s); Human-a = asthmatic human; Human-n = nonasthmatic human; LOAEL = lowest-observed-adverse-effect level; M = male; MEF<sub>40% (P)</sub> = maximal expiratory flow at 60% of vital capacity below total lung capacity on the partial flow volume curve; min = minute(s); NOAEL = no-observed-adverse-effect level; NS = not specified; Resp = respiratory; V<sub>max50%</sub> = maximal flow at 50% of the vital capacity; V<sub>max75%</sub> = maximal flow at 75% of the vital capacity; wk = week(s)

**Figure 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation**  
**Acute (≤14 days)**



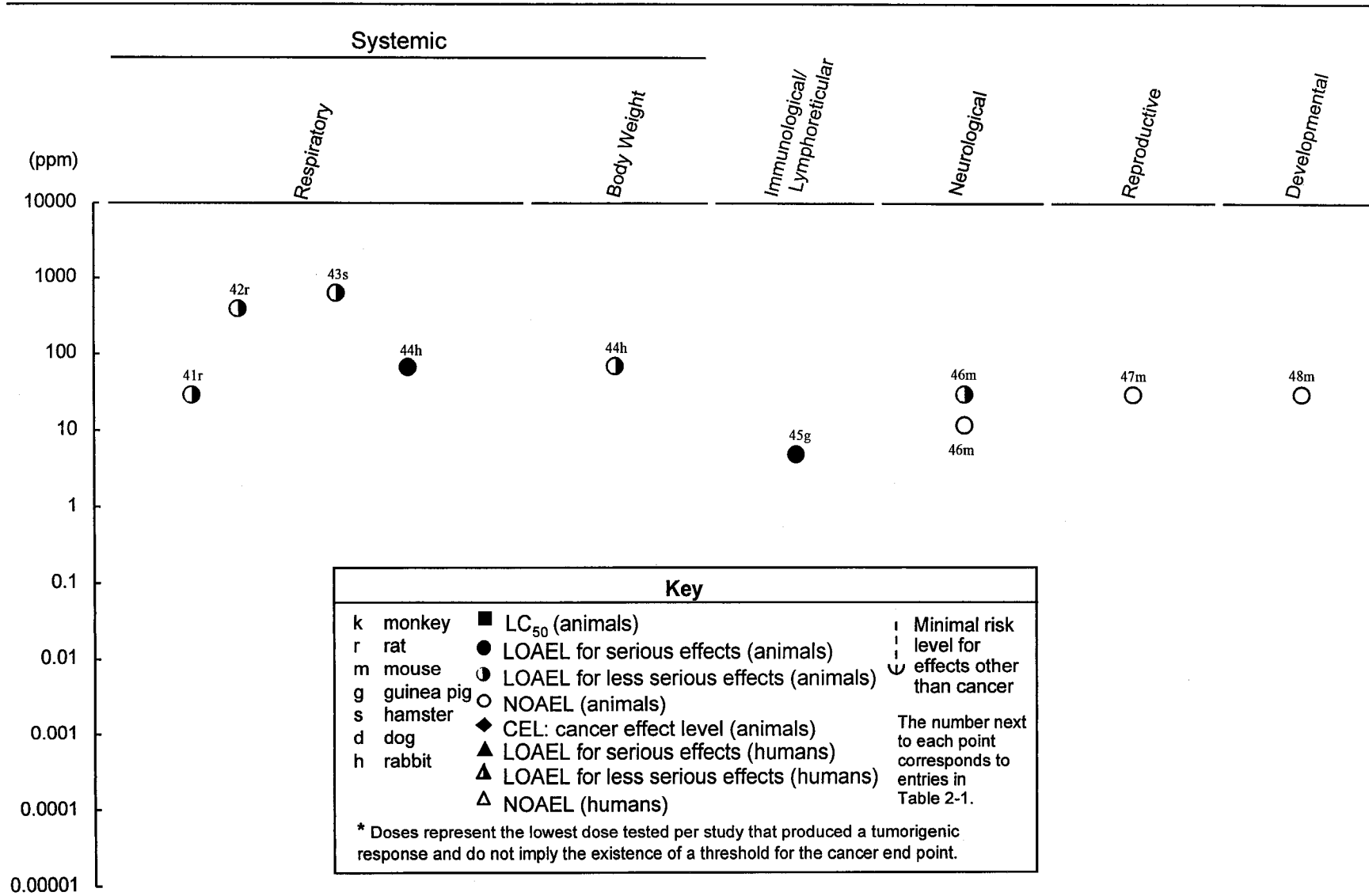
**Figure 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (cont.)**

**Acute ( $\leq 14$  days)**

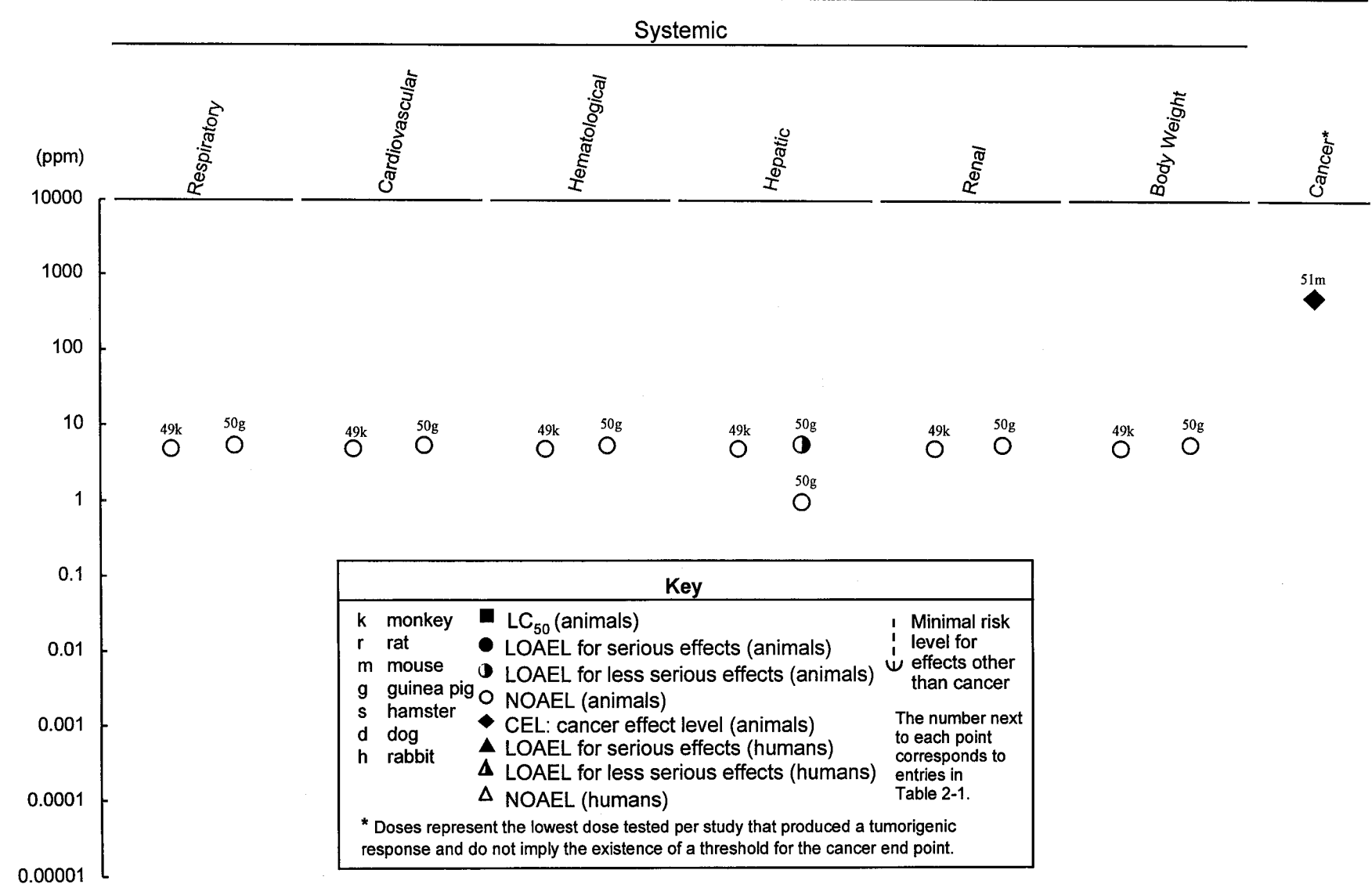


**Figure 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (cont.)**

**Intermediate (15-364 days)**



**Figure 2-1. Levels of Significant Exposure to Sulfur Dioxide - Inhalation (cont.)**  
**Chronic (≥365 days)**





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regression of a special subset of high-pollution days. However, a later reexamination of mortality and London pollution data revealed that the log of acid aerosol concentrations was more strongly associated with raw total mortality in bivariate analyses than with smoke or sulfur dioxide (Thurston et al. 1989).

Associations have also been reported between daily acute mortality and pollution episodes involving sulfur dioxide and suspended particulate matter in Philadelphia, PA, (1973-1988) and Steubenville, OH, (1974-1984) (Moolgavkar et al. 1995a, 1995b). Although the mortality has been attributed to the particulate component of air pollution, recent reanalyses of data have indicated that it is premature to single out one specific component as being responsible for the observed association between air pollution and mortality (Moolgavkar et al. 1995a, 1995b, 1996).

In eastern and western Europe, associations between short-term increases in air pollutant levels and daily mortality rates have recently been evaluated by the Air Pollution and Health: a European Approach (APHEA) project (Katsouyanni et al. 1997). The purpose of the project was to provide a standardized method of evaluation between 15 European cities, and the primary pollutants examined were sulfur dioxide and particulates. Effects were evaluated for both current day increases in air pollution levels and for lag periods of 1-5 days past exposure. Statistical techniques were used to control for confounding factors such as various air pollutants, temperature, relative humidity, influenza epidemics, season, year, month, holidays, and day of the week. Despite efforts to ensure consistency in evaluation methods, the sulfur dioxide findings were variable. For instance, an association between short-term increases of sulfur dioxide levels in air and increased daily mortality was not observed in some cities (Bacharova 1996; Ballester et al. 1996; Verhoeff et al. 1996). In another city, analysis of sulfur dioxide alone indicated an association with daily mortality, but the effect was not observed when sulfur dioxide was analyzed in combination with other air pollutants (Anderson et al. 1996). Weak but significant associations between short term increases in sulfur dioxide levels and daily mortality were noted in other cities (Spix and Wichmann 1996; Sunyer et al. 1996; Touloumi et al. 1996).

Results from the pooling and metaanalysis of data from 12 cities suggested a weak but statistically significant association between increased sulfur dioxide levels and daily mortality in western European cities but not in central eastern Europe (Katsouyanni et al. 1997). In most studies sulfur dioxide concentrations were highest during the winter, but effects were noted primarily during the summer. A possible explanation may be an increased exposure to outdoor air during the summer months (Sunyer 1996).

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Inconsistencies in the various APHEA studies may have resulted from several limitations with epidemiological studies of air pollutant mixtures. For example, there may be confounding effects due to air pollutants that were not measured. Also, concentrations were obtained from monitoring sites and this provides no information about personal exposures within individuals of a population. Lastly, there is little understanding about the mechanism of toxicity associated with air pollutants. Therefore, separating the effects of individual air pollutants within a mixture is very unlikely with the technology available at this time (Ballester et al. 1996; Katsouyanni et al. 1997; Loomis et al. 1996; Moolgavkar et al. 1996).

The approximate LC<sub>50</sub> for a 30minute exposure in Swiss albino mice was 3,000 ppm (Hilado and Machado 1977). In this study, there were no deaths during or after 30 minutes of exposure to 1,190 ppm sulfur dioxide. However, the study is limited because it did not indicate the age, sex, or the number of animals used in the experiment. The LC<sub>50</sub> value associated with death in mice is shown in Table 2-1 and plotted in Figure 2- 1.

The time course of mortality in groups of 8 rats was examined following continuous exposure to 590-500,000 ppm sulfur dioxide (Cohen et al. 1973). Average mortality times were 1866,750, 176, <10 and <2 minutes for rats exposed to 590,925,2350,50,000, or 500,000 ppm sulfur dioxide respectively. Mortality in groups of 8 rats was monitored for two weeks following a 4-hour exposure to 224-1,319 ppm sulfur dioxide (Cohen et al. 1973). None of the rats died following exposure to 224 or 593 ppm sulfur dioxide. Three rats exposed to 965 ppm died. Five rats exposed to 1,168 ppm died 1 to 48 hours after exposure, and all rats exposed to 1,319 ppm died 1 to 24 hours after exposure. Results of these studies are illustrated in Table 2- 1 and plotted in Figure 2-1.

### 2.2.1.2 Systemic Effects

**Respiratory Effects.** In humans, and in particular asthmatics, respiratory changes are a primary response following acute exposure to sulfur dioxide. Numerous controlled clinical studies have examined pulmonary lung function, usually assessed by measurement of increases in specific airway resistance and/or decreases in forced expiratory volume or forced expiratory flow, in human subjects exposed to sulfur dioxide. The methods of exposure to sulfur dioxide usually involved oronasal, nose-only, or mouth breathing techniques. Several chamber studies have been performed in a number of investigations in various laboratories.

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In 10 healthy, nonasthmatic individuals, exposure to sulfur dioxide (via a chamber) at concentrations of up to 1.0 ppm for up to 40 minutes was associated with only a slight increase in subjective, mild, upper-respiratory symptoms such as sore throat and ability to taste and smell sulfur dioxide (Schachter et al. 1984). There was no effect on lung function parameters. However, reductions in forced expiratory volume and forced expiratory flow and an increase in nasal airflow resistance were observed in 15 healthy subjects exposed nose-only to  $\geq 1.0$  ppm sulfur dioxide for 1-6 hours (Andersen et al. 1974). Decreased tidal volume and increased respiratory rate were observed in 14 healthy subjects exposed to 1-8 ppm sulfur dioxide for 10 minutes (Amdur et al. 1953). Specific airway resistance was increased in 26 healthy individuals exposed to 0.6-0.8 ppm sulfur dioxide for 5 minutes (Islam et al. 1992). A significant decrease in nasal mucus flow was seen in 15 healthy subjects at 5 and 25 ppm sulfur dioxide (Anderson et al. 1974). Reduced bronchial clearance was observed at 5 ppm (Wolff et al. 1986). Exercise increased the rate of bronchial clearance (Wolff et al. 1986). Increased airway resistance during rest was observed in 7 healthy subjects exposed to 4-6 ppm sulfur dioxide in a body plethysmograph for 10 minutes (Nadel et al. 1965). When 11 healthy subjects were exposed mouth only to 5 ppm sulfur dioxide for 10-30 minutes, increased flow resistance was noted (Frank et al. 1962). Cough, a sense of irritation, and increased salivation were also seen at 5 ppm. No effects were observed at 1 ppm. Erythema of the trachea and main bronchi was seen in 22 healthy individuals exposed to 8 ppm sulfur dioxide for 20 minutes (Sandstrom et al. 1989a, 1989b). This effect was accompanied by an increase in inflammatory cells in bronchoalveolar lavage fluid. Increased numbers of inflammatory cells were also observed in the bronchoalveolar lavage fluid in groups of 4-10 subjects exposed to 4 ppm sulfur dioxide for 20 minutes (Sandstrom et al. 1989a, 1989b).

A series of inhalation exposure studies were conducted in healthy subjects with various concentrations of sulfur dioxide (Lawther et al. 1975). Pulmonary function changes were not observed in 13 subjects following quiet nasal breathing of 1 ppm sulfur dioxide for 1 hour. However, significant increases in specific airway resistance were noted in 12 subjects who inhaled 25 deep breaths of 1 ppm sulfur dioxide. Increases in specific airway resistance were also noted after the subjects took 25 deep breaths of filtered air, but the response was greater with sulfur dioxide exposure. In 17 subjects, significant increases in specific airway resistance were observed after inhaling 16 deep breaths of filtered air. Additional dose-related increases in resistance were observed after inhaling 8, 16, and 32 deep breaths of 3 ppm sulfur dioxide. Specific airway resistance increased significantly in 14 subjects who breathed 5 ppm sulfur dioxide quietly by mouth for 10 minutes. Resistance was highest immediately after exposure and, depending on the sensitivity of the subject, lasted from 5 to 65 minutes following exposure. Additional dose-related increases in resistance were not

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observed following exposure to 10-30 ppm sulfur dioxide. Subjects could detect higher levels of sulfur dioxide and it appears that they tried to protect themselves with shallow breathing.

The results of controlled laboratory studies in humans have established that asthmatics are particularly sensitive to the respiratory effects of sulfur dioxide. In a study of 21 asthmatics exposed for 10 minutes to sulfur dioxide by mouthpiece while at rest, increases in airway resistance were observed at 1,3, and 5 ppm (Sheppard et al. 1980).

Controlled studies have indicated that the pulmonary effects of sulfur dioxide can be significantly enhanced by exercise. Significant increases in airway resistance have been clearly demonstrated in moderately exercising asthmatics (ventilation range: 35-60 L/minute) exposed briefly (3-10 minutes) to 0.40-1.0 ppm sulfur dioxide (Bethel et al. 1985; Linn et al. 1983a, 1984a; Roger et al. 1985; Schachter et al. 1984). Significant changes in airway resistance were observed in young adult mild asthmatics exposed, while exercising, to a concentration of sulfur dioxide as low as 0.25 ppm through a mouthpiece (Sheppard et al. 1981). The two most sensitive asthmatics exhibited some degree of bronchoconstriction, as evidenced by a slight increase in specific airway resistance ( $SR_{aw}$ ) following inhalation of 0.1 ppm sulfur dioxide through a mouthpiece for 10 minutes (Sheppard et al. 1981). A dose-response relationship, as measured by airway resistance, was observed in the two sensitive subjects following exposures to 0.25 and 0.5 ppm sulfur dioxide. At 0.25 ppm, the difference between baseline specific airway resistance and specific airway resistance ( $\Delta SR_{aw}$ ) after inhalation of sulfur dioxide was approximately 5 Lxcm  $H_2O/L/s$  (units of  $SR_{aw}$ ). At 0.5 ppm, the  $\Delta SR_{aw}$  exceeded 15 Lxcm  $H_2O/L/s$ .

Lung function changes in asthmatics exposed by inhalation to 0.25 ppm sulfur dioxide have been reported by other investigators. In a chamber study of moderately exercising asthmatics, the concentration of sulfur dioxide required to produce an increase in airway resistance 100% greater than the response to clean air [designated as  $PC(SO_2)$ ] has been determined (Horstman et al. 1986). Analysis of the cumulative percentage of subjects plotted as a function of  $PC(SO_2)$  revealed that 25% of the subjects exhibited a  $PC(SO_2)$  of 0.25-0.5 ppm sulfur dioxide. The study authors suggested that the 25% of the mild asthmatics who were very sensitive to sulfur dioxide could possibly exhibit bronchoconstriction if they were to perform normal exercise routines in some highly industrialized areas of the United States. Increases in specific airway resistance were observed in 9-19 moderately exercising asthmatics exposed oronasally to 0.25 ppm sulfur dioxide for 5 minutes (Bethel et al. 1985). A dose-related increase in specific airway resistance was seen in

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asthmatics following a 3-minute exposure (via mouthpiece) to  $\geq 0.25$  ppm sulfur dioxide (Myers et al. 1986a, 1986b).

Some studies of asthmatics have reported a lack of significant lung function changes in asthmatics following exposures to 0.1-0.5 ppm (Jorres and Magnussen 1990; Koenig et al. 1990). Bronchoconstrictive responses to sulfur dioxide are highly variable among individual asthmatics (Horstman et al. 1986). In some studies, asthmatics were preselected for sensitivity to sulfur dioxide and this may explain the range of sulfur dioxide-induced responses obtained by different investigators.

In addition to exercise, sulfur dioxide-induced bronchoconstriction can be increased by cold or dry air (Sheppard et al. 1984). For instance, the concentration resulting in a 100% increase in specific airway resistance in 8 asthmatics exposed to sulfur dioxide in dry, cold air was  $0.51 \pm 0.2$  ppm, whereas it was  $0.6 \pm 0.41$  ppm for dry, warm air and  $0.87 \pm 0.41$  ppm humid, warm air.

In several controlled acute studies, the authors have reported that, because of the severity of the pulmonary response, exposures to sulfur dioxide had to be terminated for some of the asthmatic subjects while other subjects required medical attention. Two out of seven asthmatics required a bronchodilator after exposure to cold air and 0.5 ppm sulfur dioxide (Bethel et al. 1984). Two of 10 subjects exposed to 0.5 ppm sulfur dioxide were unable to complete the experiment (Koenig et al. 1985). Similar events were reported by other investigators (Balmes et al. 1987; Horstman et al. 1986, 1988; Linn et al. 1984a, 1984b, 1984c; Roger et al. 1985).

Controlled studies in asthmatic subjects have demonstrated that repeated exposures to sulfur dioxide reduce the responsiveness of asthmatics to the chemical. For instance, the pulmonary response of 10 exercising asthmatics exposed to 1.0 ppm sulfur dioxide in a chamber was attenuated after repeated exercise (Kehrl et al. 1987). Bronchoconstriction was less severe in 14 exercising asthmatics exposed to 0.6 ppm sulfur dioxide on the second day of a 2-day exposure period (Linn et al. 1984a). Specific airway resistance in 28 asthmatics was significantly less after second and third exercises when compared with the first exercise (Roger et al. 1985).

A series of studies was conducted to examine respiratory function in healthy, atopic, or asthmatic adolescents (n=8-9, aged 12-17 years) and 10 healthy male seniors (aged 55-73 years) following exposure by mouth only to a mixture of 1 ppm sulfur dioxide and  $1 \text{ mg/m}^3$  sodium chloride aerosol (Koenig et al. 1982a, 1982b,

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1983; Rondinelli et al. 1987). Exposures consisted of 30 minutes of rest and 10 minutes of exercise for adolescents and 20 minutes of rest and 10 minutes of exercise for seniors. Statistically significant decreases in forced expiratory volume in 1 second (FEV<sub>1</sub>) were observed in all groups of subjects following exposure to the sulfur dioxide and sodium chloride mixture during exercise. The magnitude of decreases were 23% in asthmatic adolescents, 18% in atopic adolescents, 8% in seniors, and 6% in healthy adolescents (Koenig et al. 1982a, 1982b, 1983; Rondinelli et al. 1987). Based on a comparison of FEV<sub>1</sub> values, it was concluded that respiratory health status and not age is the primary factor in determining susceptibility to sulfur dioxide (Rondinelli et al. 1987). Asthmatic adolescents were most sensitive to sulfur dioxide and normal adolescents were least sensitive. Responses in seniors and healthy adolescents were similar. However, the doses inhaled by healthy adolescents were about 20% greater because of a longer exposure period and a more strenuous exercise period. A further decrease in FEV<sub>1</sub> may have occurred in seniors had they received the same doses as adolescents. In two of the studies, it was verified that exposure to a mixture of sulfur dioxide and sodium chloride aerosol resulted in effects which did not differ significantly from those obtained from exposure to sulfur dioxide alone (Koenig et al. 1982b, 1982a). The authors stated that in the environment sulfur dioxide occurs together with one or more droplet aerosols and that the studies represented a realistic exposure scenario.

Adaptation to irritant concentrations of sulfur dioxide is a recognized occurrence in workers (Department of Labor 1975). The potential tolerance to the pulmonary effects of sulfur dioxide has been studied in asthmatics. Eight asthmatic subjects were exposed to 0.5 ppm sulfur dioxide with a mouthpiece for 3 minutes while performing voluntary eucapnic hyperpnea (rapid deep breathing to deplete arterial carbon dioxide) (Sheppard et al. 1983). The subjects received three 3minute exposures with a 30 minute rest period between exposures. Specific airway resistance was increased significantly more after the first exposure to sulfur dioxide than after the second or third exposures. Tolerance was not observed when the exposures were repeated 24 hours later and then 7 days later.

A case study describes a sulfide dust explosion in a copper mine that liberated large amounts of sulfur dioxide (Rabinovitch et al. 1989). Analysis of the gas sample obtained at the time of rescue showed sulfur dioxide concentrations >40 ppm, with no other toxic gases present. Effects resulting from the exposure consisted of burning of the nose and throat, dyspnea, and severe airway obstruction that was only partially reversed 2 years after the exposure. In another case report of an industrial accident at a paper mill involving exposure to high but unmeasured concentrations of sulfur dioxide, histological examination of the lungs of two subjects who died revealed extensive sloughing of the mucosa of large and small airways along with

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hemorrhagic alveolar edema. Acute symptoms observed in the survivors included irritation of the nose and throat, tightness in the chest, and intense dyspnea. Serial pulmonary function tests revealed severe airway obstruction in one subject on the first day of examination. Another survivor who used a protective mask developed mild airway obstruction by day 116. No pulmonary function abnormalities were noted in the third survivor, a fireman who arrived later on the scene.

Bronchial hypersensitivity can develop following a single exposure to very high concentrations of sulfur dioxide, a syndrome referred to as reactive airway dysfunction syndrome or RADS (Brooks et al. 1985; Brooks 1992; Brooks et al. 1985; Goldstein et al. 1979; Harkonen et al. 1983). In this syndrome, the bronchial epithelial damage results in increased sensitization and nonspecific hypersensitivity to a wide range of other irritant stimuli.

Bronchitis has been reported in pulp mill workers following brief accidental exposures to 100 ppm sulfur dioxide (Skalpe 1964).

Epidemiological studies on the relationship between sulfur dioxide exposure and respiratory effects have been conducted. However, these studies are limited because of the difficulties in separating potential effects of sulfur dioxide from those of particulates and other air pollutants. A longitudinal study of 343 children exposed to sulfur dioxide and particulate sulfate was conducted in three towns in Arizona, two of which have smelters (Dodge et al. 1985). The study period was from 1979 to 1982. The annual average sulfur dioxide concentrations at two monitoring sites were 0.005 ppm and 0.04 ppm. However, in the area of highest pollution, children were exposed intermittently to high levels of sulfur dioxide (peak 3-hour mean exceeded 1 ppm) and moderate levels of particulate sulfate. A significant correlation between the prevalence of cough and pollution levels was noted. There were no significant changes in lung function over a 3-year period.

Lung function has been studied in 200 school-age children following several acute air pollution episodes in Steubenville, OH (Dockery et al. 1982). One day in 1979, the children were exposed to mean daily concentrations of 0.17 ppm sulfur dioxide and 0.27 mg/m<sup>3</sup> total suspended particles, levels which exceeded the 24-hour standard in 1979. The children were examined in three weekly visits following each pollution alert. The children were measured again in five weekly examinations in the spring and fall of 1980. Slight (2-3%) decreases in forced vital capacity (FVC) and forced expiratory volume in 0.75 seconds (FEV<sub>0.75</sub>) were noted in the children the day following the acute pollution episode in 1979 and 1-2 weeks later. The

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decreases were statistically significant and reversible after 2-3 weeks. The effects of total suspended particles cannot be separated from those of sulfur dioxide.

In New York, the association between seasonal air pollution levels and forced expiratory volume in 0.75 seconds ( $FEV_{0.75}$ ) was examined in two age groups of children (5-8 and 9-13 years-old) (Shy et al. 1973). At the time of the study, sulfur dioxide levels in polluted areas ranged from 63-71  $\mu\text{g}/\text{m}^3$  (0.024-0.027 ppm) and levels of total suspended particulates (TSPs), suspended sulfates, and nitrogen dioxide were elevated in comparison to non-polluted areas. Levels of sulfur dioxide, TSPs, and suspended sulfates were greatly reduced three years prior to the initiation of the study. Previous sulfur dioxide levels ranged from 364-435  $\mu\text{g}/\text{m}^3$  (0.14-0.17 ppm). Associations between pollutant levels and reduced  $FEV_{0.75}$  were only observed in the older group of children. The authors concluded that persistent respiratory effects may result from 5-10 year exposures to high air pollution levels during early childhood. Effects associated with individual air pollutants were not separated in this study.

The association between peak expiratory flow rates (PEFR) and sulfur dioxide and particulate air pollution was examined in 60 asthmatic children (aged 9-14 years) living in Budapest during the period of September 13 through December 5, 1993 (Agocs et al. 1997). Sulfur dioxide concentrations ranged from 11-185  $\mu\text{g}/\text{m}^3$  (0.004-0.071), and levels exceeded World Health Organization (WHO) limits on several days. Confounding effects such as temperature, humidity, weekday, and time trends were controlled and each air pollutant was assessed individually. There were no consistent associations between air pollution levels and PEFR. Study limitations were addressed by the authors and included medication use by many of the children, incomplete control of seasonal factors, and a lack of personal exposure data.

In London, increases in sulfur dioxide levels by 14  $\mu\text{g}/\text{m}^3$  (0.005 ppm) were associated with a 12% increase in the number of emergency room visits for wheezing episodes in children (Buchdahl et al. 1996).

Confounding effects from season, temperature, and wind speed were controlled, but effects from the other air pollutants, including ozone and nitrogen dioxide, were not separated. In the evaluation of ozone, effects from sulfur dioxide and nitrogen dioxide were controlled, and ozone was found to have the strongest association with wheezing episodes in children.

Difficulty breathing and cough were reported by 91-94% of workers in a Yugoslavian broom manufacturing plant who were exposed to average sulfur dioxide concentrations of 0.29 and 57 ppm in the summer and winter, respectively (Savic et al. 1987). There appeared to be no control for confounding factors such as



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occupational exposure to dust and smoking. A study was also conducted of workers exposed to sulfur dioxide in a refrigerant manufacturing plant (Kehoe et al. 1932). Prior to 1927 concentrations of sulfur dioxide averaged between 80-100 ppm. After the installation of a ventilation system, sulfur dioxide levels typically ranged between 5-35 ppm with occasional peaks as high as 50-70 ppm. The incidence of respiratory irritation and shortness of breath during heavy activity were significantly increased in 100 exposed workers. Chest X-rays revealed no differences between exposed and unexposed workers at the same plant. Occupational exposure to other chemicals was not described and there was no control for smoking.

A significant decrease in forced expiratory volume in 1 second (FEV<sub>1</sub>) was observed in 113 workers exposed to  $\geq 1$  ppm sulfur dioxide for an unspecified time in a copper smelter in Salt Lake City (Smith et al. 1977). Smoking status was assessed in the study. However, workers were also exposed to respirable dust. A study conducted years later at the same copper smelter found no significant relationships between pulmonary function and exposure to  $\geq 5$  ppm sulfur dioxide in 430 workers (Lebowitz et al. 1979). Exposure durations ranged from 0 to >20 years and confounding factors such as age, smoking, and exposure to dust were corrected. The authors suggested that the previous findings of Smith et al. (1977) may have resulted from limitations of the study, such as a small sample size and inadequate correction for age.

In another study of 953 copper smelter workers in Garfield, Utah, a significant reduction in FVC and FEV<sub>1</sub> was associated with long-term (>20 years) exposure to 0.4-3.0 ppm sulfur dioxide (Archer and Gillan 1978). Smoking status was assessed in the study. The reduction was observed with increasing duration of exposure in both smokers and nonsmokers but was not observed in controls. However, workers were also exposed to arsenic, copper, manganese, iron, and other trace metals.

Workers (4,506 and 5,943) exposed to mean concentrations of approximately 0.84-1.2 ppm sulfur dioxide for an unspecified time period at two British steel plants did not experience an increase in respiratory symptoms or reduction in respiratory performance (Lowe et al. 1970). Confounding factors such as smoking and occupational exposure to dust were controlled.

In a study of 56 workers exposed to 2-36 ppm sulfur dioxide in pulp mills for 1 month to 40 years, cough was reported by 56% of workers, sputum production by 46%, and difficulty breathing by 22% (Skalpe 1964). Symptoms were reported in higher percentages by workers  $\leq 50$ -years-old. Maximal expiratory flow rate was significantly reduced in workers <50-years-old. The author speculated the most likely reason for increased

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effects in the younger workers was that minor respiratory effects are more likely detected due to decreased prevalence of respiratory symptoms at a younger age. Occupational exposure to other chemicals was not reported but smoking status was controlled.

Variability in the findings of occupational studies may have resulted from common limitations in each study. Personal exposure levels were not measured in any of the studies but were estimated from area samples. Levels of sulfur dioxide may vary widely within a small area of a plant and exposures could differ significantly between workers in the area. Also, workers are usually exposed to multiple toxic substances and it is difficult to separate the effects of individual compounds.

Though erratic results were obtained in the occupational studies, it is evident that effects occurred at higher concentrations than in controlled chamber studies of asthmatics. Workers represent a subgroup which is healthier than the general population. Due to breathing difficulties, individuals with respiratory disorders such as asthma would generally be excluded from working in areas with high sulfur dioxide concentrations. Therefore, it is expected that changes in lung function would be observed at lower concentrations in controlled studies of asthmatics than in occupational studies.

Studies in experimental animals have supported the findings of pulmonary effects of sulfur dioxide in humans following inhalation exposure. Sixteen guinea pigs exposed to 2.6 ppm sulfur dioxide in an exposure chamber for 1 hour showed a 20% increase in resistance accompanied by a 10% decrease in compliance (Amdur 1959). The response to sulfur dioxide increased when exposure was increased to 3 hours. Increased pulmonary flow resistance was noted in guinea pigs exposed to 0.16-835 ppm sulfur dioxide (Amdur 1966). In a nose-only inhalation exposure study, 6-9 guinea pigs exposed to 1.0 ppm sulfur dioxide for 1 hour did not display any effects on airway responsiveness (Chen et al. 1992b). In a study designed to assess the bronchial sensitization to ovalbumin in 6 guinea pigs following exposure to 5 ppm sulfur dioxide for 6 hours per day for 5 days, serious pulmonary effects, including severe destruction of ciliated epithelium of the bronchioles, partial collapse of alveoli, and polymorphonuclear infiltrates were observed (Riedel et al. 1992).

Decreased compliance and increased resistance were noted in 12 anesthetized dogs exposed to 1.1-141 ppm sulfur dioxide for 2040 minutes using a mask fitted so that the dogs breathed through the nose and the mouth (Balchum et al. 1959). The average decrease in compliance for 11 dogs was 5%. One dog exposed to 141 ppm for 40 minutes displayed a 121% decrease in compliance. Dogs exposed to 45 ppm for 16 minutes displayed increased resistance and decreased compliance of the lung (Balchum et al. 1960b). Decreased

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compliance and increased resistance were also seen in 10 anesthetized dogs exposed to 1.8-148 ppm sulfur dioxide through a tracheal cannula (Balchum et al. 1960b). Compliance was decreased from control levels at an average of 8.5% (statistically significant at  $p < 0.05$ ). Resistance was increased by 150-300%. The increase in resistance occurred within 9 seconds after the onset of sulfur dioxide exposure and disappeared quickly at the end of exposure.

Acute-duration exposure to high concentrations of sulfur dioxide can result in biochemical, clinical, and histological changes in the respiratory systems of the mouse and the rabbit. A transient decrease in cough reflex and the Hering-Breuer inflation reflex were observed in 18 rabbits exposed to 200-300 ppm sulfur dioxide for 10-20 minutes (Hanacek et al. 1991).

Four mice exposed to 20 ppm sulfur dioxide for up to 120 minutes exhibited degenerative changes in the olfactory epithelium (Min et al. 1994). A substantially increased number of polymorphonuclear lymphocytes in the trachea was noted in 3-5 rats exposed to 230 ppm sulfur dioxide for 5 hours (Farone et al. 1995). A loss of cilia and cell necrosis in the trachea and main bronchus were observed in 5 rats exposed to 800 ppm sulfur dioxide for 8 hours (Stratmann et al. 1991).

Rats exposed to 25 ppm sulfur dioxide for up to 5 days displayed nasal epithelial metaplasia and basal cell hyperplasia (Fowlie et al. 1990).

Respiratory effects from exposure to sulfur dioxide for intermediate time periods have also been studied in animals. Mild bronchitic lesions were seen in 72 hamsters exposed to 650 ppm sulfur dioxide for 4 hours/day, 5 days/week, for 19-74 days (Goldring et al. 1970). Decreased respiratory rate, rhinitis, tracheitis, and bronchopneumonia were observed in 6 rabbits exposed to 70-300 ppm sulfur dioxide for 6 weeks (Miyata et al. 1990). Nasopharyngitis and lipid peroxidation of lung tissue were observed in guinea pigs exposed to 10 ppm sulfur dioxide for 1 hour/day, for 30 days (Haider 1985).

Significantly increased activities of acid phosphatase and alkaline phosphatase, decreased numbers of epithelial cells, and increased numbers of leukocytes were observed in the bronchial lavage fluid of rats exposed to 30-40 ppm sulfur dioxide for 1 hour/day, 5 days/week, for 12 weeks (Krasnowska et al. 1998). A histopathological evaluation of the bronchial mucosa revealed damage to the epithelium accompanied by infiltration of leukocytes, destruction of cilia, and squamous cell metaplasia, which were more pronounced in rats examined three weeks past exposure compared to those examined immediately after exposure.

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Exposure of 12 rats to 400 ppm sulfur dioxide for 3 hours/day, 5 days/week, for up to 42 days resulted in airway effects similar to those observed in humans with chronic bronchitis (Lamb and Reid 1968). A loss of cilia, epithelial necrosis, and a reduction in goblet cell numbers were observed during the first 2-4 days of exposure. By 3-6 weeks of exposure, signs of healing were evident and included a thickened epithelium and the reappearance of shortened cilia. During the same time period, goblet cells increased in size and number and appeared in distal airways, areas in which they are normally lacking. The sialidase-sensitive mucous in distal airways was replaced by sialidase-resistant mucous within three weeks of exposure. Tracheal gland size continually increased throughout the exposure period. Similar effects were observed in another study in which rats were exposed to 400 ppm sulfur dioxide for 3 hours/day, 5 days/week, for up to 3 weeks (Basbaum et al. 1990). An 8-9 fold increase in mucin mRNA transcripts was also observed.

The respiratory toxicity of sulfur dioxide following chronic-duration inhalation exposure has been studied in animals. No effects on lung function were observed in 50 guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972). Likewise, no lung function changes or histopathological alterations in the lung were observed in 9 monkeys exposed to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975).

In summary, the available data indicate that sensitive asthmatics may respond to concentrations of sulfur dioxides as low as 0.1 ppm. Healthy nonasthmatics respond to higher concentrations of sulfur dioxide ( $\geq 1.0$  ppm). Factors that can exacerbate the respiratory effects of sulfur dioxide include exercise and breathing of dry or cold air. Animal data support the human data on respiratory effects of sulfur dioxide.

**Cardiovascular Effects.** Human, nonasthmatic subjects ( $n \leq 14$ ) exposed to 1-8 ppm sulfur dioxide showed increased pulse rate (Amdur et al. 1953).

No evidence of histological lesions in the heart was found in 9 monkeys following exposure to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). Likewise, no microscopic lesions were detected in the hearts of 50 guinea pigs that were exposed by inhalation to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972). Increased lipid peroxidation was observed in hearts of 6 guinea pigs exposed to 10 ppm sulfur dioxide for 1 hour/day, for 30 days (Haider 1985). The biological significance of the result is unknown.

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**Gastrointestinal Effects.** Nausea and vomiting were observed in 3 humans exposed to >40 ppm sulfur dioxide during an accident at a copper mine (Rabinovitch et al. 1989).

No studies were located regarding gastrointestinal effects in animals after inhalation exposure to sulfur dioxide.

**Hematological Effects.** Blood samples were taken from PO0 workers exposed to sulfur dioxide for a minimum of 2 years in a refrigerant manufacturing plant (Kehoe et al. 1932). Prior to 1927, the sulfur dioxide concentration averaged between 80-100 ppm. After the installation of a ventilation system, sulfur dioxide levels typically ranged between 5-35 ppm with occasional peaks as high as 50-70 ppm. Small but significant differences in the numbers of polymorphonuclear leukocytes and lymphocytes were observed. The authors questioned the biological significance of the hematological findings and concluded that it was probably due to the high rate of respiratory infection in both exposure and control groups.

An increase in blood levels of methemoglobin was observed in 45-59 workers in a broom manufacturing factory who were exposed to average sulfur dioxide concentrations of 0.29 and 57.0 ppm in the summer and winter, respectively (Savic et al. 1987). The mean percentage of methemoglobin was elevated during the winter when windows were closed and sulfur dioxide levels were highest. With the exception of dust, exposure to other workplace chemicals was not described.

Numerous studies in animals have demonstrated oxidative effects on erythrocytes. An increase in erythrocyte deformability was observed in 14 rats exposed to 1 ppm sulfur dioxide for 24 hours (Baskurt et al. 1990). Osmotic hemolysis and sulfhemoglobin levels were increased in erythrocytes of 50 rats exposed to 0.9 ppm sulfur dioxide for 24 hours (Baskurt et al. 1988). An increase in the erythrocyte deformability index and lipid peroxidation were noted in the erythrocytes of 12 guinea pigs exposed to 10 ppm sulfur dioxide for 30 days (Dikmenoglu et al. 1991). Significantly increased ratios of methemoglobin and sulfhemoglobin, lipid peroxidation, and increased fragility of erythrocytes, were observed in 7 rats exposed to 10 ppm sulfur dioxide for 1 hour/day, for 45 days (Etlik et al. 1997). Significant changes in antioxidant enzyme activities and increased lipid peroxidation were observed in the erythrocytes of 15 rats exposed to 10 ppm sulfur dioxide for 1 hour/day, 7 days/week, for 8 weeks (Gumusht et al. 1998).

Mixed results have been obtained for sulfur dioxide-induced effects on blood cell numbers in animals. No hematological effects were observed in a chronic toxicity study of 9 monkeys exposed to 5.12 ppm sulfur

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dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). Likewise, no hematological effects were noted in guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972). However, statistically significant increases in white and red blood cell counts, as well as hematocrit and hemoglobin levels were observed in 7 rats exposed to 10 ppm sulfur dioxide for 1 hour/day for 45 days (Etlik et al. 1997). Increased hematocrit levels were also observed in 50 rats exposed to 0.9 ppm sulfur dioxide for 24 hours (Baskurt et al. 1988).

Animal studies support the finding in humans that sulfur dioxide induces oxidative effects in erythrocytes. However, both human and animal studies are limited. In the occupational studies, levels of sulfur dioxide varied greatly and individual exposure levels were not measured. In addition, confounding factors such as smoking or exposure to other chemicals were not controlled. Because multiple doses were not tested in any of the animal studies, the dose response relationship is unknown. Data from well designed occupational or animal studies are required before the impact of sulfur dioxide exposure on blood can be assessed.

**Musculoskeletal Effects.** No studies were located regarding musculoskeletal effects in humans or animals after inhalation exposure to sulfur dioxide.

**Hepatic Effects.** No studies were located regarding hepatic effects in humans after inhalation exposure to sulfur dioxide.

Information on hepatic effects in animals from inhalation exposure is very limited. In a chronic toxicity study in monkeys, no histological lesions were detected in 9 monkeys exposed to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). In contrast, microscopic examination of 50 guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks, showed an increase in the size of hepatocytes accompanied by cytoplasmic vacuolation (Alarie et al. 1972).

Liver triglyceride and esterified cholesterol levels were increased in 9 rats fed either a standard diet or a high cholesterol diet and exposed to 10 ppm sulfur dioxide for 24 hours/day, 7 days/week, for 15 days (Lovati et al. 1996). However, exposure of 9 diabetic rats to 5 or 10 ppm sulfur dioxide under the same conditions resulted in a significant reduction in liver triglyceride levels. Additional experiments demonstrated that triglyceride levels increased in nondiabetic rats due to a reduction in lipid catabolism. Because insulin regulates the activity of lipases, it appears that a concurrent drop in insulin levels was responsible for the decreased catabolism in the nondiabetic rats. However, in diabetic rats it was postulated that triglyceride

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catabolism may have increased to provide an alternate energy source for glucose. Contrary findings were observed in 6 guinea pigs whose liver cholesterol level decreased following exposure to 10 ppm sulfur dioxide for 1 hour/day for 30 days (Haider 1985). Liver triglyceride levels were not measured, but total lipid concentrations in livers of exposed animals were unaffected. Additional studies are needed to define the role of sulfur dioxide in the hepatic metabolism of lipids.

**Renal Effects.** No studies were located regarding renal effects in humans after inhalation exposure to sulfur dioxide.

Information on renal effects in animals from inhalation exposure is very limited. No renal effects were observed in a chronic toxicity study of 9 monkeys exposed to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). Likewise, no renal effects were noted in 50 guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972).

**Endocrine Effects.** No studies were located regarding endocrine effects in humans after inhalation exposure to sulfur dioxide.

Significant dose-related decreases in plasma insulin levels were observed in 9 rats exposed to 5 or 10 ppm sulfur dioxide for 24 hours/day, 7 days/week, for 15 days, but significant effects on insulin levels were not noted in diabetic rats exposed to sulfur dioxide under the same conditions (Lovati et al 1996).

**Dermal Effects.** No studies were located regarding dermal effects in humans after inhalation exposure to sulfur dioxide. Skin irritation was observed in 6 rats and 12 guinea pigs exposed to 10 ppm sulfur dioxide for 1 hour/day for 30 days (Haider 1982, 1985).

**Ocular Effects.** In a case report of a paper mill accident in which five persons were exposed to high concentrations (not specified) of sulfur dioxide for less than 5 minutes, reversible conjunctivitis and superficial corneal burns were noted (Charan et al. 1979). In another case report dealing with a pyrite dust explosion that resulted in nine persons being exposed to high levels of sulfur dioxide, conjunctival irritation and corneal erosion were observed (Harkonen et al. 1983). The concentration of sulfur dioxide was not measured, but based on experimental explosions, the concentration was estimated to be 30-1,600 ppm. Burning of the eyes was reported in a case study of an accident at a copper mine in which 3 miners were exposed to >40 ppm sulfur dioxide (Rabinovitch et al. 1989). Irritation and tearing were reported by 65% of

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workers in a Yugoslavian broom manufacturing plant who were exposed to sulfur dioxide levels averaging between 0.29 ppm in summer and 57 ppm in winter (Savic et al. 1987).

Eye irritation was observed in 6 rats and 12 guinea pigs exposed to 10 ppm sulfur dioxide for 1 hour/day for 30 days (Haider 1982,1985).

**Body Weight Effects.** No studies were located regarding body weight effects in humans after inhalation exposure to sulfur dioxide.

Following 5 weeks of exposure to 70-300 ppm sulfur dioxide, body weight gains of the 6 treated rabbits were 25% lower than those of controls (Miyata et al. 1990). No body weight effects were observed in a chronic toxicity study of 9 monkeys exposed to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). Likewise, no body weight effects were noted in 50 guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972) or in 15 rats exposed to 10 ppm sulfur dioxide for 1 hour/day, 7 days/week, for 8 weeks (Gumuslu et al. 1998).

### 2.2.1.3 Immunological and Lymphoreticular Effects

In a study of 10 mild asthmatic subjects, prior inhalation of 0.2 ppm sulfur dioxide for 6 hours did not significantly affect the provocation dose of *Dermatophagoides pteronyssinus* allergen required to produce a 20% decrease in forced expiratory volume in 1 second (Devalia et al. 1994). However, prior inhalation exposure to a combination of 200 ppb sulfur dioxide and 400 ppb nitrogen dioxide significantly reduced the provocation dose of allergen required to cause a 20% decrease in forced expiratory volume in 1 second.

Increased prevalence of allergies was observed in 556 children (aged 7-13 years) living near an aluminum smelter in Norway for seven years or more (Soyseth et al. 1996). Prevalence was highest in children who lived in the area between the ages of 19-36 months and were exposed to sulfur dioxide levels of 20-24  $\mu\text{g}/\text{m}^3$  (0.008-0.009 ppm). However, there was no evidence of a dose response relationship. Fluoride levels were also increased in the area, but other air pollutants were not discussed. The data was controlled for age, sex, and in utero exposure to cigarette smoke. However, there was no control for possible confounders such as fluoride or other air pollutants.



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The incidence of acute respiratory disease was compared in groups of 2,705-8,991 children (aged 1-1,2 years) living in high and low pollution areas (based on 1973 national primary standards) within four regions of the United States (French et al. 1973). All polluted areas had high concentrations of sulfur dioxide ranging from 63-275  $\mu\text{g}/\text{m}^3$  (0.024-0.1 ppm). Contaminants which were also present in high concentrations in polluted areas included sulfates in the Salt Lake Basin, metals in the Rocky Mountain communities, and total suspended particulates (TSPs) in New York and Chicago. Generally, increases in incidence of respiratory infection were observed in children who lived in polluted areas for a minimum of 3 years. In the Salt Lake Basin and Rocky Mountain region, effects were greatest in children 4 years-old and younger. However, in Chicago, there was no association between respiratory illness and air pollution in children who were too young to attend nursery school. The discrepancy in results may have resulted from differences in the pollution mixtures or from inadequate control of confounding factors.

Increased sensitization to antigen was reported in a study of guinea pigs exposed by inhalation to sulfur dioxide (Riedel et al. 1992). In this experiment, groups of six guinea pigs were exposed to either clean air or 5 ppm sulfur dioxide for 8 hours/day for 5 days, with intermittent inhalation of ovalbumin (8 mg aerosolized) in order to sensitize the animals. Control animals were also sensitized with ovalbumin. Antibodies against ovalbumin were measured before and after sham and sulfur dioxide exposures. Animals were exposed to the ovalbumin for 45 minutes following sham or sulfur dioxide exposure on days 3,4, and 5. Seven days after the last exposure, the guinea pigs were tested for bronchial sensitization to ovalbumin. Sulfur dioxide-treated animals were found to be sensitized to ovalbumin, while pretreatment with anti-inflammatory drugs (methylprednisolone, indomethacin, or nedocromilsodium) for 6 days beginning 12 hours before the first sulfur dioxide exposure prevented ovalbumin sensitization.

In an acute-duration inhalation study with hamsters ( $\leq 12/\text{group}$ ), there was a significant reduction in endocytosis by pulmonary macrophage (process used in defending lung against pathogens and foreign bodies) following exposure to 50 ppm sulfur dioxide for 4 hours while exercising (Skornik and Brain 1990). However, exposure of rats and mice to 0.32-0.43 ppm sulfur dioxide together with 87-113  $\mu\text{g}/\text{m}^3$  sulfate for 4 hours prior to or 17 hours following infection with *Staphylococcus aureus* or Group C *Streptococci* had no effect on clearance or phagocytosis of *S. aureus* or rates of *Streptococcal* killing and proliferation (Goldstein et al. 1979).

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**2.2.1.4 Neurological Effects**

Reflexes were examined in 100 workers exposed to sulfur dioxide in a refrigerant manufacturing facility for a minimum of 2 years (Kehoe et al. 1932). Prior to 1927, the sulfur dioxide concentration averaged between 80-100 ppm. After the installation of a ventilation system, sulfur dioxide levels typically ranged between 5-35 ppm with occasional peaks as high as 50-70 ppm. A significant number of workers had reflex response times that were slower or faster than normal. The authors stated that the effect did not indicate neurological injury but reflected differences in irritability. Neurological effects of sulfur dioxide exposure have been described as psychological responses to general toxicity (Parmeggiani 1983). The Kehoe et al. (1932) study is limited because occupational exposure to other chemicals was not well characterized.

Seizures and prostration were observed in groups of 8 rats prior to death, which occurred following exposure to 2,350,50,000 and 500,000 ppm sulfur dioxide for an average of 176 minutes, <10 minutes, and <2 minutes respectively (Cohen 1973). Transient changes in the frequency of certain behavioral patterns, such as grooming and digging, were noted in 10 male and 10 female mice continuously exposed to 30 ppm sulfur dioxide and observed during the first 9 days prior to mating in a reproductive study (Petruzzi et al. 1996). There was no evidence of neurological effects in the offspring of the mice.

Increased lipid peroxidation was observed in the brains of 6 guinea pigs exposed to 10 ppm sulfur dioxide for 1 hour/day for 30 days (Haider et al. 1982). However, the study is limited due to small sample size and administration of only one dose. In addition similar studies are not available and the reproducibility of results cannot be verified. Therefore, the biological significance of the results is not known.

**2.2.1.5 Reproductive Effects**

In a cross-sectional study of spontaneous abortions in an industrial community in Finland, no evidence was found that exposure to sulfur dioxide was associated with a risk of spontaneous abortions (Hemminki and Niemi 1982). Types of industries in the area included pulp paper, metal, viscose rayon, and chemical. Hydrogen sulfide and carbon disulfide were industry-related pollutants which were also present in the area. High sulfur dioxide levels in air were associated with abnormal sperm morphology and decreased motility in 325 18-year-old males in the Czech Republic (Selevan et al. 1995). Because sulfur dioxide was used as a surrogate for all air pollutants, it cannot be determined which chemical or mixture of chemicals was responsible for the effect on sperm.

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Reproductive effects, as measured by completed pregnancies, litter size, sex ratio, and neonatal mortality, were not observed in 10 male and 10 female mice continuously exposed to 5, 12, or 30 ppm sulfur dioxide for 9 days prior to mating until days 12-14 of pregnancy (Petruzzi et al. 1996). Pregnancy rate and the number of implants, resorptions, and live fetuses were not affected in 32 mice exposed to 25 ppm sulfur dioxide for 7 hours/day on gestation days 6-16 or in 20 rabbits exposed to 70 ppm sulfur dioxide for 7 hours/day on gestation days 6-18 (Murray et al. 1979). The offspring were not assessed for reproductive function in these studies.

**2.2.1.6 Developmental Effects**

An association between exposure to increased levels of air pollution during pregnancy and a reduction in birth weight has been reported in humans (Wang et al. 1997). Decreased birth weights were noted for exposure to both sulfur dioxide and TSPs. However, evaluation of these results is complicated by limitations such as confounding effects of various air pollutants and a lack of personal exposure data. Therefore, differentiation of potential developmental effects associated with individual pollutants of a mixture is not possible.

No developmental effects were observed in a study in which 13 pregnant mice were exposed to 32-250 ppm sulfur dioxide on gestation days 7-17 (Singh 1982). Neurological effects, measured by reflexes and learning ability, were not observed in the offspring of 10 mice continuously exposed to 5-30 ppm sulfur dioxide 9 days prior to mating through gestation days 12-14 (Petruzzi et al. 1996). Although there were some transient behavioral effects in dams, there were no signs of systemic maternal toxicity. In another developmental study in mice in which pregnant females were exposed to 32 ppm or 65 ppm on gestation days 7-18, increased time for righting reflex on postnatal day 1 and increased negative geotaxis on postnatal day 10 were reported in offspring (Singh 1989). The duration of exposure for each day was not stated. No visible signs of maternal toxicity and no effect on the number of live births were observed. Sulfur dioxide at a concentration of 65 ppm significantly decreased the birth weight (about 89% of controls) of the pups. Reduced body weight and delayed ossification in sternbrae and occipital bones were observed in the offspring of 32 mice exposed to 25 ppm sulfur dioxide for 7 hours/day on gestation days 6-15 (Murray et al. 1979). Increased numbers of skeletal variations, such as non-ossified sections of frontal bones, fusion of occipital and parietal bones, and extra ribs, were also observed in the offspring of 20 rabbits exposed to 70 ppm sulfur dioxide for 7 hours/day on gestation days 6-18 (Murray et al. 1979). Decreased food intake was the only sign of maternal toxicity in mice and rabbits.

### 2.2.1.7 Genotoxic Effects

Assays of clastogenic effects in humans following occupational exposure to sulfur dioxide via inhalation show mostly positive results. Increases in chromosome aberrations and sister chromatid exchanges were detected in lymphocytes from 42 workers at an Indian fertilizer plant who were exposed to an average concentration of 41.7 mg/m<sup>3</sup> (15.92 ppm) of sulfur dioxide (Yadav and Kaushik 1996). Similar findings were observed in another study of 40 workers exposed to sulfur dioxide in a Chinese sulfuric acid factory (Meng and Zhang 1990a). Exposure concentrations ranged from 0.34 to 11.97 mg/m<sup>3</sup> (0.13 to 4.57 ppm). In addition, increases in the frequencies of lymphocytes with micronuclei were noted (Meng and Zhang 1990b). Confounding exposures were not discussed in these studies. A significant increase in the frequency of chromosomal aberrations was found among 19 workers at a sulfite pulp factory (Nordenson et al. 1980). Exposure concentrations were not reported and workers were also exposed to chlorine and dust. One study of potential chromosomal abnormalities in workers exposed to sulfur dioxide while working in the aluminum industry revealed that sulfur dioxide did not cause an effect (Sorsa et al. 1982).

No studies were located regarding genotoxic effects in animals after inhalation exposure to sulfur dioxide.

Other genotoxicity studies are discussed in Section 2.5.

### 2.2.1.8 Cancer

There is no definitive evidence for an increased cancer potential from sulfur dioxide in humans. Several epidemiological studies have been conducted on copper smelter workers and pulp and paper workers who can be exposed to sulfur dioxide (IARC 1992). However, the studies conducted in copper smelters focused primarily on the association between arsenic exposure and cancer. It has also been difficult to separate the potential effects of sulfur dioxide and arsenic exposures in the studies of copper smelter workers. Studies of the potential carcinogenicity of sulfur dioxide are discussed in the following paragraphs.

A cohort study of 5,403 male copper smelter workers showed that respiratory cancer risks in workers exposed to sulfur dioxide for 212 months were not significantly increased when controls for exposure to arsenic were in place (Lubin et al. 1981). The concentration of sulfur dioxide was not specified in the study. In a follow-up mortality study in the same cohort of smelter workers, a clear dose-response relationship between arsenic exposure and respiratory cancer was demonstrated (Welch et al. 1982). Although an

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apparent relationship of mortality to sulfur dioxide exposure was observed, workers in the medium and high sulfur dioxide exposure categories had higher exposures to arsenic than those in the low sulfur dioxide exposure groups. The concentrations of sulfur dioxide associated with each exposure category were not specified. The studies by Lubin et al. (1981) and Welch et al. (1982) are limited because exposures to sulfur dioxide and arsenic could not be completely separated.

The results of a case-cohort study of mortality in workers from eight copper smelters in the United States revealed that arsenic was the primary cause of lung cancer in copper smelter workers after adjusting for cigarette smoking and sulfur dioxide exposure (Enterline et al. 1987).

A nested case-control study of lung cancer among 308 workers in a large chemical facility revealed significantly elevated risks for workers with moderate and high potential exposure ( $\geq 1$  year) to sulfur dioxide (Bond et al. 1986). For workers who had been exposed to sulfur dioxide, the odds ratio for lung cancer was 1.40. Application of multivariate analyses showed a significant trend ( $p=0.003$ ) of increasing lung cancer risk associated with increasing intensity of sulfur dioxide exposure for the comparison with the decedent controls. The odds ratios for lung cancer were 0.48 for low exposure, 1.69 for moderate exposure, and 1.45 for high exposure. No trend was apparent for the comparison with the living controls. Also, the risk of lung cancer did not increase with duration of exposure to sulfur dioxide. Confounding effects from exposure to numerous chemicals was also possible. Products manufactured at the facility included chlorinated solvents, plastics, chlorine, caustic soda, ethylene, styrene, epoxy, latex, magnesium metal, chlornitrogen products, and glycols. Employees were exposed to an average of 7.5 chemicals during their career, and the most common exposures included chlorine, sulfur dioxide, hydrogen chloride, and carbon tetrachloride. The study authors indicated that the findings must be interpreted with caution because such findings have not been reported among other workers with similar exposures.

The relationship between ambient air pollution (including sulfur dioxide) and lung cancer has been examined. Lung cancer cases (2,439 males, 765 females) were identified in Helsinki, Finland, and standardized incidence ratios were calculated for 33 subareas of Helsinki for 1975-1978, 1979-1982, and 1983-1986 (Ponka et al. 1993). Mean annual concentrations of sulfur dioxide were 0.005-0.008 ppm. After adjustment for age, sex, and level of education, the lung cancer risk increased slightly, but nonsignificantly, with increasing sulfur dioxide concentration. Lung cancer was 1.3% higher in the subareas with the highest sulfur dioxide concentrations ( $\geq 0.008$  ppm) compared with the subareas with the lowest concentrations ( $< 0.005$  ppm). There was no consistent relationship between the concentration of nitrogen dioxide and the

## 2. HEALTH EFFECTS

incidence of lung cancer. The study authors concluded that sulfur dioxide had little, if any, effect on the risk of lung cancer.

A retrospective cohort study employing a Poisson regression model for time trends of mortality to detect subtle effects of air pollution on lung cancer mortality in Japan was conducted (Tango 1994). The trend of mortality in females, aged 40-79, was examined in 23 wards of the Tokyo metropolitan area. The size of the cohort was not specified. The ward-specific time trend of nitrogen dioxide and sulfur dioxide concentrations for the years 1972 through 1988 was estimated. The result of this study showed a statistically significant ( $p=0.0055$ ) association between exposure to nitrogen dioxide and an increased trend of lung cancer mortality. The association for sulfur dioxide exposure was nonsignificant ( $p=0.0655$ ).

One chronic-duration animal study investigated the potential carcinogenicity of inhaled sulfur dioxide in mice (Peacock and Spence 1967). An experimental group of 35 male and 30 female mice was exposed to 500 ppm sulfur dioxide for 5 minutes/day, 5 days/week, for 2 years. The control group consisted of 41 males and 39 females. Female mice exposed to sulfur dioxide exhibited a significant increase in the incidence of lung tumors (13/30 adenomas and carcinomas versus 5/30 in controls; 4/30 primary carcinomas versus none in the controls). The incidence of lung adenomas and carcinomas was also higher in the treated males (15/28 versus 11/35 in controls), but the increase was not significant. The incidence of primary carcinomas in treated males was similar to that of the controls. These data provide limited evidence for the carcinogenicity of sulfur dioxide in the mice. However, the determination of carcinogenic potential is complicated by study limitations. A dose-response relationship could not be assessed because multiple doses were not tested. Due to the small group sizes, it cannot be concluded that increased tumor incidences did not result from chance alone. Quality studies in additional species are required before the carcinogenicity status of the compound can be determined.

## 2.2.2 Oral Exposure

### 2.2.2.1 Death

No studies were located regarding death in humans or animals after oral exposure to sulfur dioxide.

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**2.2.2.2 Systemic Effects**

No studies were located regarding respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, endocrine, dermal, ocular, body weight, or other systemic effects after oral exposure to sulfur dioxide.

No studies were located regarding the following health effects in humans or animals after oral exposure to sulfur dioxide:

**2.2.2.3 Immunological and Lymphoreticular Effects****2.2.2.4 Neurological Effects****2.2.2.5 Reproductive Effects****2.2.2.6 Developmental Effects****2.2.2.7 Genotoxic Effects**

Genotoxicity studies are discussed in Section 2.5.

**2.2.2.8 Cancer**

No studies were located regarding cancer in humans or animals after oral exposure to sulfur dioxide.

**2.2.3 Dermal Exposure****2.2.3.1 Death**

No studies were located regarding death in humans or animals after dermal exposure to sulfur dioxide.

**2.2.3.2 Systemic Effects**

No studies were located regarding respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, endocrine, body weight, or other systemic effects in humans or animals after dermal exposure to sulfur dioxide.

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**Dermal Effects.** Sulfur dioxide is a severe irritant to the skin (Department of Labor 1975). Sulfur dioxide is a liquid under pressure at 0°C. On the skin, the liquid produces burns from the freezing effect of rapid evaporation (Department of Labor 1975).

No studies were located regarding dermal effects in animals after dermal exposure to sulfur dioxide.

**Ocular Effects.** Sulfur dioxide is a severe irritant to the eyes (Department of Labor 1975). The eye irritation level is 10 ppm. Exposure of the eyes to liquid sulfur dioxide from pressurized containers can cause corneal burns and opacification, resulting in loss of vision. It has been reported that ocular damage from exposure to liquid sulfur dioxide is not due to freezing but is characteristic of chemical burns (Grant 1974). Sulfurous acid, which is formed when sulfur dioxide contacts moist surfaces, appears to be the primary cause of eye injury.

Application of pure sulfur dioxide gas (room temperature and atmospheric pressure) directly to the eyes of rabbits for 5 seconds has produced severe damage to the cornea and conjunctiva, similar to effects observed with acid burns (Grant 1974). Opacity of corneas was noted immediately after and 6 months following exposure.

No studies were located regarding the following health effects in humans or animals after dermal exposure to sulfur dioxide.

#### 2.2.3.3 Immunological and Lymphoreticular Effects

#### 2.2.3.4 Neurological Effects

#### 2.2.3.5 Reproductive Effects

#### 2.2.3.6 Developmental Effects

#### 2.2.3.7 Genotoxic Effects

Genotoxicity studies are discussed in Section 2.5.



### 2.2.3.8 Cancer

No studies were located regarding cancer in humans or animals after dermal exposure to sulfur dioxide.

## 2.3 TOXICOKINETICS

Sulfur dioxide, a highly water-soluble gas, is rapidly absorbed by the mucosa of the nose and upper respiratory tract. Absorption in the lower respiratory tract is increased with enhanced ventilation associated with a transition from nasal to oronasal breathing. Upon contact with moist mucous membranes, sulfur dioxide is hydrolyzed to sulfites which are taken up by the blood and readily distributed throughout the body. Sulfites can either be oxidized to sulfates by sulfite oxidase, primarily in the liver, or they can react with proteins to form S-sulfonate. Sulfates are excreted in the urine.

### 2.3.1 Absorption

#### 2.3.1.1 Inhalation Exposure

Studies in humans have shown that sulfur dioxide, a highly water-soluble gas, is rapidly absorbed by the mucosa of the nose and upper respiratory tract (Kleinman 1984; Speizer and Frank 1966). Absorption of sulfur dioxide in the upper respiratory tract is more efficient during nose breathing than during mouth breathing (WHO 1979). Absorption of sulfur dioxide by the mucosa of the lower respiratory tract is minimal because of minimal delivery to this region, but is increased with increased ventilation associated with a transition from nasal to oronasal breathing at a mean minute ventilation of 30 L/min (EPA 1986d).

Studies in animals indicate that sulfur dioxide is rapidly absorbed by mucosa following inhalation exposure. In rabbits exposed to 100, 200, or 300 ppm sulfur dioxide, 90-95% absorption of sulfur dioxide by tissues in the upper respiratory tract was observed (Dalhamn and Strandberg 1961). The rate of absorption of sulfur dioxide by tissues in the nasal cavity was found to be higher than that of tissues of the mouth or pharynx (Dalhamn and Strandberg 1961). A subsequent study in rabbits revealed that absorption of sulfur dioxide is dependent on concentration. At high concentrations ( $\geq 100$  ppm), sulfur dioxide absorption in tissues of the respiratory tract was  $\geq 90\%$ , while at low concentrations ( $\leq 0.1$  ppm), absorption was approximately 40% (Strandberg 1964). Studies in dogs have supported the findings that sulfur dioxide is readily absorbed by

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mucosa in the upper respiratory tract and that the nose is more efficient than the mouth in removing sulfur dioxide (Balchum et al. 1959, 1960a; Frank et al. 1969).

Studies in humans have indicated that 12-15 % of sulfur dioxide absorbed on nasal mucosa is desorbed and exhaled (Speizer and Frank 1966). The remaining sulfur dioxide metabolites are absorbed into the systemic circulation or are delivered to the lower respiratory system by repeated absorption and desorption from mucosa (Frank et al. 1969). Systemic absorption of sulfur dioxide metabolites from tissues of the upper respiratory tract has been demonstrated in animals. In dogs a small segment of trachea was isolated and perfused with radiolabeled sulfur dioxide ( $^{35}\text{SO}_2$ ) while the lungs were ventilated with air to prevent entry of the  $^{35}\text{SO}_2$  (Balchum et al. 1960a). Detection of  $^{35}\text{S}$  in lungs, liver, spleen, and kidneys indicated systemic absorption from the tracheal mucosa. It appears that systemic absorption is more efficient from the lower respiratory tract. Blood and organ levels of  $^{35}\text{S}$  were compared in dogs following oronasal or tracheal exposure to  $^{35}\text{SO}_2$  (Balchum et al. 1959). Following oronasal exposure, levels of  $^{35}\text{S}$  were lower in the blood and several organs including the liver, spleen, and lymph nodes. In most dogs the level of  $^{35}\text{S}$  remained steady, thus suggesting a slow continued absorption.

### 2.3.1.2 Oral Exposure

No studies were located regarding absorption of sulfur dioxide after oral exposure in humans or animals.

### 2.3.1.3 Dermal Exposure

No studies were located regarding absorption of sulfur dioxide after dermal exposure in humans or animals.

## 2.3.2 Distribution

### 2.3.2.1 Inhalation Exposure

*S*-sulfonate ( $\text{R-S-SO}_3^-$ ), which forms from the reaction of the sulfur dioxide metabolite, sulfite, and proteins, was measured in the plasma of human subjects following continuous exposure to 0.3, 1.0, 3.0, 4.2, or 6.0 ppm sulfur dioxide in a chamber for periods of up to 120 hours (Gunnison and Palmes 1974). Plasma levels of *S*-sulfonate showed a positive correlation with air concentrations of sulfur dioxide.

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The results of studies in dogs indicate that absorbed sulfur dioxide metabolites are taken up by the blood and are readily distributed throughout the body (Balchum et al. 1960a; Frank et al. 1967; Yokoyama et al. 1971). The results of a study in dogs exposed to  $22 \pm 2$  ppm radiolabelled sulfur dioxide ( $^{35}\text{SO}_2$ ) for 30-60 minutes showed that the radioactivity levels appeared in blood within 5 minutes of the onset of exposure. It was estimated that from 5% to 18% of the radioactivity administered to dogs was contained in the blood by the end of exposure (Frank et al. 1969). In a subsequent study in dogs designed to further examine the distribution of radioactivity in the blood, blood-radioactivity levels increased progressively when the isolated upper airways were exposed to  $^{35}\text{SO}_2$  for 30-60 minutes (Yokoyama et al. 1971). Blood-radioactivity levels decreased slightly during a postexposure period that lasted up to 3 hours. The radiolabel was more concentrated in the plasma than in the red blood cells. Also, approximately one-third of the radioactivity in plasma was associated with proteins, especially  $\gamma$ -globulins. About two-thirds of the radioactivity associated with the red blood cells was intracellular.

The tissue distribution of radioactivity has been studied in dogs administered  $^{35}\text{SO}_2$  as a gas to either the intact upper airways or to an isolated segment of the trachea (Balchum et al. 1959, 1960a, 1960b). In dogs that breathed 0.47-148 ppm  $^{35}\text{SO}_2$ , via a tracheal cannula, the radioactivity levels were highest in the trachea and lungs, followed by the hilar lymph nodes, kidneys, and esophagus (Balchum 1960b). A lower concentration of radiolabel was noted in other tissues such as the heart muscle, liver, spleen, striated muscle, brain, ovaries, stomach, pancreas, eye, skin, and submaxillary gland. Experiments in dogs in which  $^{35}\text{SO}_2$  was inhaled through the nose and mouth at concentrations of 1-141 ppm, showed that a large proportion was deposited in the upper airways (Balchum et al. 1959, 1960a). Also, dogs breathing  $^{35}\text{SO}_2$  through the nose and mouth retained a smaller portion of the radiolabel in the trachea, lungs, hilar lymph nodes, liver, and spleen than those breathing similar concentrations via a tracheostomy (Balchum et al. 1959).

Following oronasal inhalation of radiolabeled sulfur dioxide by dogs, 0.01-0.18% of the  $^{35}\text{S}$  dose was measured in ovaries and 0.006-0.1% was measured in testicles (Balchum et al. 1959).

### 2.3.2.2 Oral Exposure

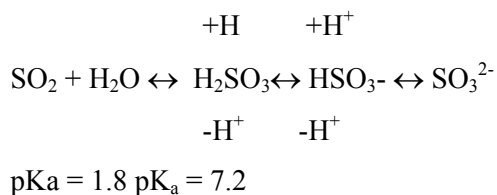
No studies were located regarding distribution of sulfur dioxide after oral exposure.

### 2.3.2.3 Dermal Exposure

No studies were located regarding distribution of sulfur dioxide after dermal exposure.

### 2.3.3 Metabolism

Following absorption, inhaled sulfur dioxide dissolves on the walls of the moist airways producing a mixture of sulfite, bisulfite, and hydrogen ions (Gunnison et al. 1987). The formation of sulfite ( $\text{SO}_3^{2-}$ ), bisulfite ( $\text{HSO}_3^-$ ), and hydrogen ions occurs in accordance with the equilibria depicted below:



Once formed, sulfite can be oxidized to sulfate, a reaction catalyzed by sulfite oxidase which occurs primarily in the liver (Ellenhorn and Barceloux 1988; Gunnison et al. 1987). Sulfite oxidase is present in mitochondria and has been detected in most tissues (Cabre et al. 1990). High sulfite oxidase activity has been measured in the liver, kidney, and heart, but the brain, spleen, lungs, and testis have been found to have low sulfite oxidase activity. Low pulmonary sulfite oxidase activity may explain the findings of a study in which the distribution of sulfite from inhaled sulfur dioxide was restricted largely to the major airways of the lung (Gunnison et al. 1987).

Experiments utilizing sulfite oxidase-competent rats have demonstrated an absence of sulfite in the plasma of rats following inhalation exposure to sulfur dioxide (Gunnison et al. 1987). Decreased activity of sulfite oxidase in sulfite oxidase-deficient rats results in higher *in vivo* concentrations of sulfite. S-sulfonate is the primary chemical form of absorbed sulfur dioxide in the bloodstream (Sheppard 1988; Yokohama et al. 1971).

Sulfite oxidase activity was found to be lower in the livers of young versus mature rats (Cohen 1974). In one day-old rats, sulfite oxidase activity was approximately 1/10 the level of adult rats. Activity increased as the rats matured and at 32 days of age, the liver sulfite oxidase activity was approximately one half the level of

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adult rats. The efficiency of the sulfite oxidation reaction depends primarily on the activity of sulfite oxidase (Gunnison and Palmes 1976). However, detoxification of sulfites is not solely dependent on sulfite oxidase activity. It has been demonstrated that rabbits clear injected sulfites at a faster rate than rhesus monkeys, but that the *in vitro* activity of sulfite oxidase in the primary metabolizing organs is greater for monkeys than rabbits (Gunnison et al. 1977). The study indicates that metabolism of sulfites is dependent on factors in addition to sulfite oxidase activity.

Sulfites can react and become associated with disulfide bonds of plasma proteins, resulting in the formation of S-sulfonates (Gunnison et al. 1987). Further metabolism and subsequent elimination of S-sulfonates is not known. However, it has been postulated that disulfide bonds may reassociate resulting in the release of sulfite (Gunnison and Palmes 1973). Another possibility is that glutathione may react with S-sulfonates to either separate S-sulfonates from proteins or form mixed disulfides and sulfites. It is expected that sulfites produced from such reactions would then be metabolized by sulfite oxidase. Reaction of sulfites with plasma proteins may protect tissues against injury, but may also prolong exposure to very low levels of sulfite (Gunnison and Benton 1971).

There is evidence of age related differences in the metabolism of sulfite to sulfate and in the formation of a sulfur trioxide radical intermediate. Levels of sulfur trioxide radicals and sulfite oxidase activity were measured in polymorphonuclear leukocytes (PMN's) obtained from healthy young adults (average age 25), healthy older adults (average age 65), 3 centenarians ( $\geq 100$  years), and 3 Down's syndrome patients (Constantin et al. 1996). Significantly increased amounts of sulfur trioxide radicals were observed in PMN's from healthy adults who had low sulfite oxidase activity. In centenarians and Down's syndrome patients, generation of the sulfur trioxide radical appeared to be the primary mechanism for detoxification of sulfite, and the sulfur trioxide radical formation was not correlated with the sulfite oxidase activity level.

Sulfur dioxide is excreted primarily in the urine as sulfate (Yokoyama et al. 1971). Increased levels of sulfates have been detected in the urine of dogs (Yokoyama et al. 1971) and humans (Savic et al. 1987) exposed to sulfur dioxide.

Decreased glutathione levels in the lungs of rats exposed to sulfur dioxide suggest that glutathione may be involved in the detoxification process (Langley-Evans et al. 1996). *In vitro* experiments have demonstrated that sulfites, metabolites of sulfur dioxide, react with reduced glutathione to form S-sulfogluthathione in a reaction which is catalyzed by thiol transferase (Kagadel et al. 1986). Conversion of S-sulfogluthathione by

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$\gamma$ -glutamyltranspeptidase yields S-sulfocysteinylglycine which is hydrolyzed to S-sulfocysteine by renal peptides. S-sulfogluthathione has been detected in lenses and intestinal mucosa of animals and S-sulfocysteine has been observed in body fluids.

**2.3.4 Elimination and Excretion****2.3.4.1 Inhalation Exposure**

The excretion of inhaled sulfur dioxide has been studied in dogs. In dogs exposed by inhalation to  $^{35}\text{SO}_2$ ,  $^{35}\text{S}$  was excreted primarily in the urine as sulfate (Yokoyama et al. 1971). An average of 84.4% of the urinary radioactivity was in the form of inorganic sulfate, while 92.4% was present as total sulfate (Yokoyama et al. 1971). In dogs exposed to  $^{35}\text{SO}_2$ , the rate of excretion of the radiolabel by the kidney has been shown to be approximately proportional to the level of radiolabel in plasma and whole blood (Frank et al. 1967; Yokoyama et al. 1971).

In humans it is estimated that 12-15% of sulfur dioxide absorbed to mucous membranes is desorbed and exhaled (Speizer and Frank 1966). In an experimental study in which the airways were surgically isolated in anesthetized dogs,  $^{35}\text{S}$  was detected in expired gas samples following exposure to 22 ppm  $^{35}\text{SO}_2$  (Frank et al. 1967). The concentration of  $^{35}\text{SO}_2$  in the expired gas samples was low, representing 1% of the exposure level. The study authors stated that the lungs were releasing the gas during expiration, presumably from pulmonary capillaries.

**2.3.4.2 Oral Exposure**

No studies were located regarding the elimination and excretion of sulfur dioxide after oral exposure.

**2.3.4.3 Dermal Exposure**

No studies were located regarding the elimination and excretion of sulfur dioxide after dermal exposure.

### 2.3.5 Physiologically Based Pharmacokinetic (PBPK)/Pharmacodynamic (PD) Models

Physiologically based pharmacokinetic (PBPK) models use mathematical descriptions of the uptake and disposition of chemical substances to quantitatively describe the relationships among critical biological processes (Krishnan et al. 1994). PBPK models are also called biologically based tissue dosimetry models. PBPK models are increasingly used in risk assessments, primarily to predict the concentration of potentially toxic moieties of a chemical that will be delivered to any given target tissue following various combinations of route, dose level, and test species (Clewell and Andersen 1985). Physiologically based pharmacodynamic (PBPD) models use mathematical descriptions of the dose-response function to quantitatively describe the relationship between target tissue dose and toxic end points

PBPK/PD models refine our understanding of complex quantitative dose behaviors by helping to delineate and characterize the relationships between: (1) the external/exposure concentration and target tissue dose of the toxic moiety, and (2) the target tissue dose and observed responses (Andersen et al. 1987; Andersen and Krishnan 1994). These models are biologically and mechanistically based and can be used to extrapolate the pharmacokinetic behavior of chemical substances from high to low dose, from route to route, between species, and between subpopulations within a species. The biological basis of PBPK models results in more meaningful extrapolations than those generated with the more conventional use of uncertainty factors.

The PBPK model for a chemical substance is developed in four interconnected steps: (1) model representation, (2) model parametrization, (3) model simulation, and (4) model validation (Krishnan and Andersen 1994). In the early 1990s validated PBPK models were developed for a number of toxicologically important chemical substances, both volatile and nonvolatile (Krishnan and Andersen 1994; Leung 1993). PBPK models for a particular substance require estimates of the chemical substance-specific physicochemical parameters, and species-specific physiological and biological parameters. The numerical estimates of these model parameters are incorporated within a set of differential and algebraic equations that describe the pharmacokinetic processes. Solving these differential and algebraic equations provides the predictions of tissue dose. Computers then provide process simulations based on these solutions.

The structure and mathematical expressions used in PBPK models significantly simplify the true complexities of biological systems. If the uptake and disposition of the chemical substance(s) is adequately described, however, this simplification is desirable because data are often unavailable for many biological processes. A

simplified scheme reduces the magnitude of cumulative uncertainty. The adequacy of the model is, therefore, of great importance, and model validation is essential to the use of PBPK models in risk assessment.

PBPK models improve the pharmacokinetic extrapolations used in risk assessments that identify the maximal (i.e., the safe) levels for human exposure to chemical substances (Andersen and Krishnan 1994). PBPK models provide a scientifically sound means to predict the target tissue dose of chemicals in humans who are exposed to environmental levels (for example, levels that might occur at hazardous waste sites) based on the results of studies where doses were higher or were administered in different species. Figure 2-2 shows a conceptualized representation of a PBPK model

At the current time, PBPK models for sulfur dioxide have not been developed.

## **2.4 MECHANISMS OF ACTION**

### **2.4.1 Pharmacokinetic Mechanisms**

Sulfur dioxide, a water-soluble gas, is readily absorbed through the upper respiratory tract of both animal species and humans. Once absorbed, sulfur dioxide is mostly metabolized in the liver by sulfite oxidase (Ellenhorn and Barceloux 1988). Sulfites are readily distributed throughout the body. Although significant species-specific differences in sulfite oxidase activity in the liver and kidney have been reported (Tejnorova 1978), potential species-specific differences in the activity of this enzyme in the respiratory tract (the target of sulfur dioxide toxicity in both humans and animals) have not been reported. Sulfur dioxide is excreted in the urine as sulfate.

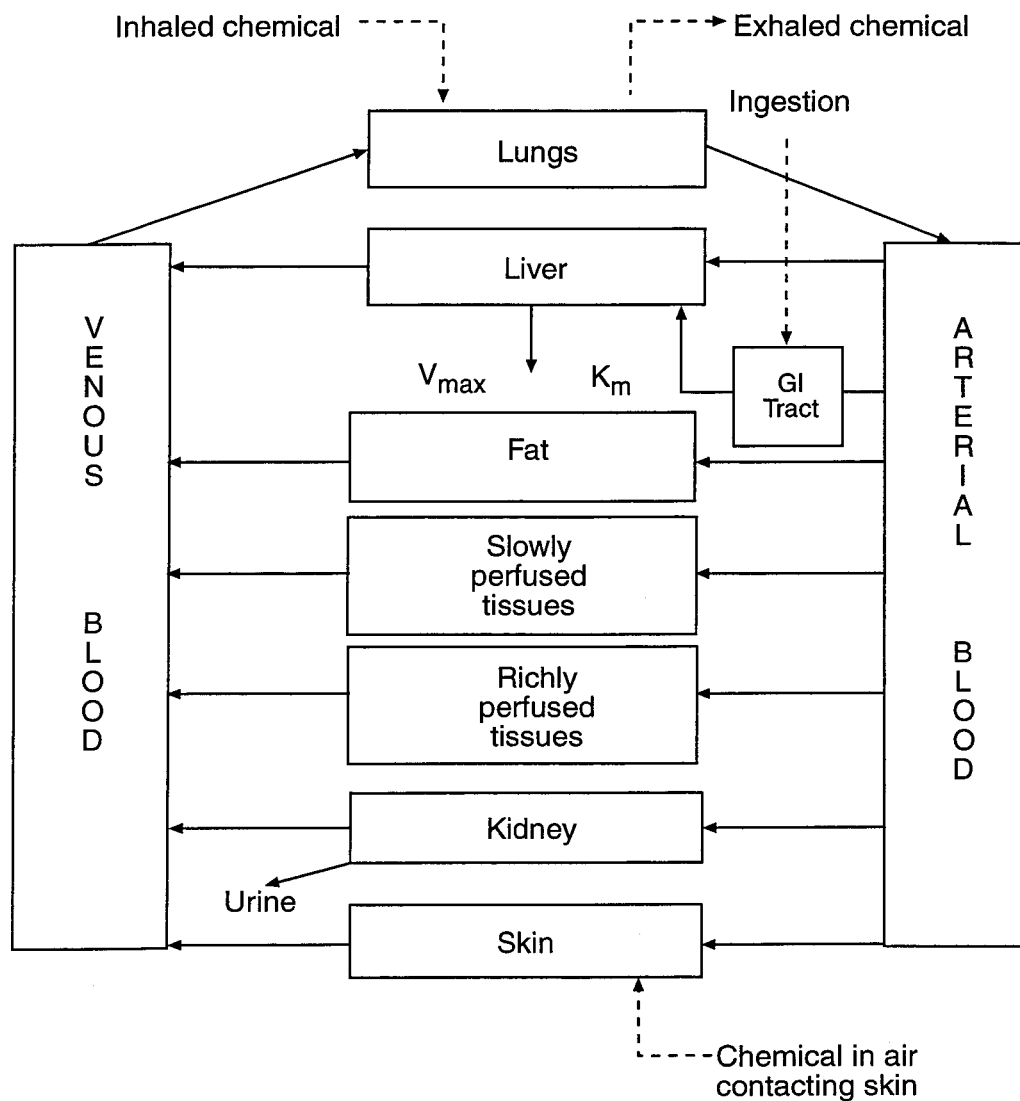
### **2.4.2 Mechanisms of Toxicity**

Sulfur dioxide-induced increase in airway resistance is due to reflex bronchoconstriction (Frank et al. 1962; Nadel et al. 1965). The reflex nature of bronchoconstriction during inhalation of sulfur dioxide has been demonstrated in humans and in cats (Nadel et al. 1965). Injection of atropine prevented the increase in airway resistance in healthy subjects exposed by inhalation to 4-6 ppm sulfur dioxide for 10 minutes. In another study, atropine increased airway conductance in 11 healthy subjects exposed to 8 ppm sulfur dioxide for 3 minutes but had no effect on 4 asthmatics (Snashall and Baldwin 1982). The finding suggested minimal contribution of cholinergic mechanisms in sulfur dioxide-induced bronchoconstriction in asthmatics. In



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**Figure 2-2. Conceptual Representation of a Physiologically-Based Pharmacokinetic (PBPK) Model for a Hypothetical Chemical Substance**



Note: This is a conceptual representation of a physiologically-based pharmacokinetic (PBPK) model for a hypothetical chemical substance. The chemical substance is shown to be absorbed via the skin, by inhalation, or by ingestion, metabolized in the liver, and excreted in the urine or by exhalation.

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anesthetized, paralyzed, artificially ventilated cats, sulfur dioxide increased pulmonary resistance, an effect prevented by complete cold block of the cervical vagosympathetic nerves or by the injection of atropine.

Studies indicate that sulfur dioxide-induced bronchoconstriction in cats arises from the activation of a muscarinic (cholinergic) reflex via the vagus nerves (Nadel et al. 1965; Sheppard 1988). However, there is some evidence from studies of human asthmatics for the existence of both muscarinic and nonmuscarinic components (Sheppard 1988), with the nonmuscarinic component possibly involving an effect of sulfur dioxide on airway mast cells.

Induction of sulfur dioxide-induced bronchoconstriction by non-cholinergic mechanisms has been demonstrated in humans. The role of prostaglandins in sulfur dioxide-induced bronchoconstriction was investigated by measuring airway responses in asthmatic subjects administered indomethacin, a prostaglandin synthetase inhibitor (Field et al. 1996). A small but significant reduction in airway responsiveness to a sulfur dioxide challenge was noted following administration of indomethacin, thus suggesting a minor role of prostaglandins.

Leukotrienes, which are released by mast cells, may also contribute to sulfur dioxide-induced bronchoconstriction. Administration of zafirlukast, a leukotriene receptor antagonist, to asthmatics resulted in a significant decrease in sulfur-dioxide induced airway responsiveness in 10 out of 12 subjects (Lazarus et al. 1997).

Evidence of noncholinergic mechanisms of sulfur dioxide-induced bronchoconstriction has also been obtained in various animal studies. In guinea pigs partial decreases in response to a sulfur dioxide challenge were noted in 5 out of 7 animals following atropine administration, but results were not significant (Hajj et al. 1996). Results suggested that cholinergic mechanisms may play a minor role in sulfur dioxide responses. However, tachykinins, neuropeptides which are present in the C-fibers of the vagus nerve, were demonstrated to play a larger role in sulfur dioxide-induced bronchoconstriction. Blocking of tachykinin receptors with antagonists, primarily the neurokinin-2 receptor, resulted in a significant attenuation of sulfur dioxide-induced bronchoconstriction in guinea pigs.

Afferent impulses in vagal fibers of the lower respiratory tract were measured in dogs exposed to 200-500 ppm sulfur dioxide for 2 minutes (Roberts et al. 1982). Fifteen out of 34 vagal C-fibers were

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stimulated, and the impulses were consistent with sulfur dioxide-induced contraction of tracheal smooth muscle.

It has been postulated that treatment of neonatal rats with capsaicin to eliminate tachykinins through the destruction of C-fibers would result in an attenuation of sulfur dioxide-induced respiratory effects. Responses in isolated lungs from capsaicin-treated and control guinea pigs were compared following exposure to 250 ppm sulfur dioxide for 15 minutes (Bannenberg et al. 1994). Capsaicin treatment eliminated the sulfur dioxide-induced release of tachykinins and significantly reduced bronchoconstriction. Minimal bronchoconstriction was present in the lungs of the capsaicin-treated guinea pigs, thus suggesting effects from additional, unknown mediators. However, different results were obtained in intact rats treated with capsaicin shortly after birth (Long et al. 1997). Exposure to sulfur dioxide at 3 months of age resulted in increased bronchoconstriction and airway responsiveness in the capsaicin treated rats. Enhanced *in vitro* responses of tracheal rings to cholinergic substances in the capsaicin group supported the theory that sulfur dioxide effects are primarily mediated through cholinergic components. Another possible explanation is that destruction of the C-fibers resulted in the loss of a protective mechanism in which tachykinins alter breathing patterns to prevent the entry of irritants into the lungs.

The ability of tachykinins to alter breathing patterns in response to a respiratory irritant has been demonstrated in rats. Acute exposure of rats to 5% sulfur dioxide through tracheal cannulas was characterized by an immediate reduction in breathing rate (Wang et al. 1996). However, the effect was no longer noted following treatment of the vagal nerves with capsaicin to destroy C-fibers or following a vagotomy.

The involvement of a cholinergic component in the alteration of breathing patterns following exposure to sulfur dioxide has also been demonstrated. Mice (4/group) who breathed various concentrations of sulfur dioxide between 17-298 ppm experienced a rapid reduction in breathing rate at each concentration (Alarie et al. 1973). When the mice were exposed to the same concentrations via a tracheal cannula, the respiratory rate was unaffected. It was therefore concluded that respiratory suppression occurs through the stimulation of cholinergic trigeminal nerve endings in the nasal mucosa.

In conclusion, sulfur dioxide-induced bronchoconstriction and respiratory inhibition appear to be mediated through vagal reflexes by both cholinergic and non-cholinergic mechanisms. Non-cholinergic components include but are not limited to tachykinins, leukotrienes, and prostaglandins. The extent to which cholinergic

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or non-cholinergic mechanisms contribute to sulfur dioxide-induced effects is not known and may vary between asthmatic and healthy individuals and between animal species.

The chemical mechanism underlying the bronchoconstrictive effect of sulfur dioxide has been examined (Sheppard 1988). Sulfur dioxide can dissolve in water to form bisulfite ion, sulfite ion, and hydrogen ion. Studies on the bronchoconstrictor effects of inhaled sulfur dioxide and acidic and basic sulfite solution suggest that sulfite ion is not likely to mediate sulfur dioxide-induced bronchoconstriction. However, the bisulfite ion, which is present at a ratio of 5:1 (bisulfite:sulfite) at physiologic pH and more chemically reactive than sulfite, might mediate the bronchoconstrictor effect. Bisulfite is a nucleophile that can react with many biomolecules by substitution at electrophilic sites (Sheppard 1988). Such reactions can cause disruption of disulfide bonds. It has been postulated that bisulfite formed at the airway surface during inhalation of sulfur dioxide may initiate bronchoconstriction by disrupting disulfide bonds present in tissue proteins (Sheppard 1988). Hydrogen ion production is not considered a likely factor since the concentration of hydrogen ions generated by inhaled sulfur dioxide is regarded to be insufficient to cause bronchoconstriction (Sheppard 1988).

Possible mechanisms for the augmentation of sulfur-dioxide bronchoconstriction by cold, dry air have been suggested (Sheppard et al. 1984). Dry air may contribute to bronchoconstriction through heat loss and subsequent airway cooling. Increased osmolarity from water evaporation in airways is another possible mechanism. Induction of bronchoconstriction through the inhalation of hypertonic aerosols has been demonstrated. It is also likely that drying of the upper airways decreases sulfur dioxide absorption in those areas, thus allowing greater concentrations to reach the lower airways.

Bronchial hyperactivity can develop after a single exposure to a very high concentration of sulfur dioxide (Brooks et al. 1985; Harkonen et al. 1983). Respiratory injury includes epithelial cell injury and destruction, airway mucosal edema and inflammation, and airway smooth muscle bronchospasm. This hyperactivity has been termed reactive airway dysfunction syndrome (RADS) (Brooks 1992; Brooks et al. 1985; Goldstein et al. 1979). Subjects with RADS react positively to methacholine challenge. The bronchial epithelial damage results in nonspecific hyperresponsiveness to a wide range of other irritant stimuli.

Inhalation exposure to sulfur dioxide has been shown to alter mucus secretion in animals (Sheppard 1988). There is some evidence that this effect may be mediated by a vagal reflex (Sheppard 1988). Support for this mechanism has been obtained from a study in dogs which showed that a sulfur dioxide-induced increase in

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submucosal gland secretion can be abolished by vagal cooling (Sheppard 1988). The mechanism differs from that of acidic aerosols in which toxicity is dependent on hydrogen ion content (Holma 1985). *In vitro* studies have shown that mucus viscosity increases when its pH is lowered below approximately 7.4. The available literature indicates that mucus of asthmatics has a lower pH and that toxicity of acid aerosols is related to a reduction in the buffering and hydrogen ion absorption capacity of the mucus. There is no evidence which indicates that sulfur dioxide-induced changes in mucous secretion are dependent upon hydrogen ion concentration.

Mechanisms which could contribute to clastogenic effects in human lymphocytes (Meng and Zhang 1990a, 1990b; Nordenson et al. 1980; Yadav and Kaushik 1996) have been discussed (Shapiro et al. 1977). Sulfites, metabolites of sulfur dioxide, can inhibit DNA synthesis and induce chromosomal aberrations in human lymphocytes (Yadav and Kaushik 1996). Bisulfite has been shown to deaminate cytosine in bacteria, a reaction which can lead to mutations by the replacement of Guanine-Cytosine (GC) sites with Adenine-Thymine (AT). Bisulfite can also catalyze the crosslinking of proteins and nucleic acids, such as DNA and histones (proteins to which the DNA is bound). Lastly, numerous free radicals are generated during the oxidation of bisulfite to sulfate, and interaction of the free radicals with nucleic acids is possible. Though many of these mechanisms have been demonstrated *in vitro*, it has yet to be verified that the mechanisms are plausible in humans.

### 2.4.3 Animal-to-Human Extrapolations

The acute pulmonary effect of sulfur dioxide following inhalation exposure has been consistent in both humans and animals. Dose-related increases in airway resistance have been demonstrated in guinea pigs, dogs, and human subjects. There appears to be a good correlation between the dose-response in guinea pigs and asthmatic subjects; therefore, the guinea pig is considered a good model that is predictive of the response of asthmatics (Amdur et al. 1991).

## 2.5 RELEVANCE TO PUBLIC HEALTH

### Overview

The inhalation route is the primary route of exposure to sulfur dioxide. In the atmosphere, sulfur dioxide occurs with other air pollutants including sulfuric acid, sulfur trioxide, ozone, nitrogen dioxide, and

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particulates. Sulfur dioxide can undergo transformation to sulfur trioxide and sulfuric acid. Sulfur dioxide tends to be a problem in urban industrial areas, particularly those where industrial activities utilize the combustion of fuels.

Table 2-2 summarizes the health effects of sulfur dioxide. The results of controlled exposure studies in humans have established that exposure to sulfur dioxide can result in lung function changes indicative of bronchoconstriction. Stimulation of sensory receptors in the airways by irritants and chemicals such as sulfur dioxide produces reflex bronchoconstriction that is mediated primarily via cholinergic pathways. Cool, dry air and exercise also cause bronchoconstriction possibly through increased respiration associated with cooling and drying of the airways (Sheppard et al. 1984). Exercising asthmatics, in particular, are sensitive to the pulmonary effects of sulfur dioxide at concentrations as low as 0.1 ppm (Sheppard et al. 1981). It has been estimated that asthmatics represent a subset of approximately 10 million people, or 4% of the population (EPA 1994a, 1994b). The true prevalence of asthmatics though, may be as high as 7-10% of the population. Responses to sulfur dioxide are variable among asthmatics. It is estimated that exposure to 0.2-0.5 ppm sulfur dioxide during moderate exercise would lead to substantial respiratory effects, in only about 10-20% of mild to moderate asthmatics, but the most sensitive individuals could likely experience incapacitating effects (EPA 1994b). The susceptibility of asthmatics to sulfur dioxide-induced respiratory effects is not clear. Severe asthmatics may actually be less prone to adverse respiratory effects because they are less likely to exercise outdoors and more likely to take medication (EPA 1994b). Healthy nonasthmatic individuals can show lung function changes following inhalation exposure to sulfur dioxide concentrations of  $\geq 1.0$  ppm. Sulfur dioxide levels in the most polluted urban industrial areas can exceed 1 ppm. Clinical studies have demonstrated that in adolescents and senior citizens, susceptibility to sulfur dioxide is determined primarily by respiratory health status and not age (Koenig et al. 1982a, 1982b, 1983; Rondinelli et al. 1987).

Most clinical studies in humans have demonstrated statistically significant but subtle effects (i.e., in the normal range) which are not considered pathological. Therefore the effects were classified as minimal according to the ATSDR definition which describes minimal effects as “those that reduce the capacity of an organ or organ system to absorb additional toxic stress but will not necessarily lead to the inability of the organ or organ system to function normally.”

Sulfur dioxide was administered through a mouthpiece in many of the clinical studies. A mouth-only exposure eliminates scrubbing by the nasal mucosa and likely results in the delivery of larger doses to the lower airways. Mouth breathing is common during exercise and in individuals with blocked nasal passages,

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**Table 2-2. Summary of the Health Effects of Sulfur Dioxide<sup>a</sup>**

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Concentration (ppm)	Effect
≥0.1	Bronchoconstriction in sensitive exercising asthmatics
0.3–1	Possibly detected by taste or smell
1–2	Lung function changes in healthy nonasthmatic individuals
2	ACGIH recommended TLV-TWA
3	Easily detected odor
5	NIOSH recommended U.S. government STEL
6–12	May cause nasal and throat irritation
10	Upper respiratory irritation, some nosebleeds
20	Definitely irritating to eyes; chronic respiratory symptoms develop at this level; respiratory protection is necessary
50–100	Maximum tolerable exposures for 30–60 minutes
≥100	NIOSH recommended immediate danger to life

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<sup>a</sup>Modified from Ellenhorn and Barceloux 1988 and WHO 1979

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such as asthmatics. However, one group of investigators demonstrated similar results following oral and oronasal exposure of asthmatics to sulfur dioxide (Bethel et al. 1985; Sheppard et al. 1981). A need has been expressed for additional research in the distribution of inhaled air under various conditions (e.g., rest and exercise) and in healthy versus asthmatic subjects (Sheppard et al. 1981).

Acute studies in animals have supported the human data on pulmonary effects of sulfur dioxide (Amdur 1959; Balchum et al. 1959, 1960b). Guinea pigs, in particular, are considered a suitable animal model (Amdur 1966). The dose-response relationship in guinea pigs correlates well with that observed for exercising asthmatics. Exposure to high concentrations of sulfur dioxide has been shown to result in degenerative changes in the olfactory epithelium in mice (Min et al. 1994), cellular necrosis in the trachea and bronchus of rats (Stratmann et al. 1991), nasal epithelial metaplasia and basal cell hyperplasia in rats (Fowlie et al. 1990), bronchitic lesions in hamsters (Goldring et al. 1970), increased goblet cell numbers in rats (Lamb and Reid 1968) and decreased respiratory rate and bronchopneumonia in rabbits (Miyata et al. 1990).

The association between mortality and/or lung function changes and acute exposures to ambient levels of sulfur dioxide that have been examined in epidemiology studies is not clear. In the atmosphere, sulfur dioxide occurs with other pollutants that may also be associated with similar toxicological effects. It has been difficult to separate the potential effects of sulfur dioxide from those of other pollutants in epidemiological studies.

The 1990 Amendments to the Clean Air Act require that emissions of sulfur dioxide be decreased. In accordance with these Amendments, EPA has reviewed the human health effects data on this chemical and has established National Ambient Air Quality Standards (NAAQS). The annual arithmetic mean NAAQS is 0.03 ppm, while the 24-hour limit is 0.14 ppm.

### **Minimal Risk Levels for Sulfur Dioxide**

#### ***Inhalation MRLs***

- An MRL of 0.01 ppm has been derived for acute-duration exposure (14 days or less) to sulfur dioxide. This MRL is derived from the study by Sheppard et al. (1981) in which exercising mild asthmatics were exposed to  $\geq 0.1$  ppm sulfur dioxide for 10 minutes. The two most sensitive subjects developed slight bronchoconstriction after inhaling 0.1 ppm sulfur dioxide. These two subjects showed dose-related increases in airway resistance after exposure to 0.1-0.5 ppm sulfur dioxide. Significant increases in airway resistance were observed in other asthmatics exposed to  $\geq 0.25$  ppm sulfur dioxide



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during moderate exercise. Other studies in asthmatics have shown lung function changes at concentrations as low as 0.25 ppm (Bethel et al. 1985; Horstman et al. 1986; Myers et al. 1986a, 1986b). A few studies have not reported respiratory effects in asthmatics at 0.25 ppm (Linn et al. 1983b, 1987,1990).

The dose level of 0.1 ppm sulfur dioxide can be considered a minimal LOAEL. This concentration was divided by an uncertainty factor of 9 (3 for the use of a minimal LOAEL and 3 for human variability) to yield a calculated MRL of 0.01 ppm. The uncertainty factor for human variability addresses varying sensitivity among asthmatics and possible increased sensitivity in children. There is concern of increased sensitivity in young children but there is not sufficient data to confirm it (See Section 2.6).

No intermediate-duration or chronic-duration MRLs were derived because the data are insufficient.

***Oral MRLS***

No oral MRLs were derived since data were not sufficient, and this is not a clinically relevant route of exposure in humans.

**Death.** Several case studies of human deaths following acute inhalation exposures to high concentrations of sulfur dioxide have been reported (Atkinson et al. 1993; Charan et al. 1979; Harkonen et al. 1983; Huber and Loving 1991; Rabinovitch et al. 1989). It is not possible, however, to draw accurate or definitive conclusions from these reports as to the levels of exposure. Effects that were described in these reports included asphyxia, secondary to passive pulmonary and visceral congestion, lungs filled with proteinaceous edema fluid, and irreversible airway obstruction. A concentration of 100 ppm is considered immediately dangerous to life and health (HSDB 1998). Excess mortality among the elderly or the chronically sick has been observed in humans exposed to sulfur dioxide, smoke, and particulate matter during acute air pollution episodes (WHO 1979). The deaths were due primarily to respiratory failure. Associations between daily acute mortality and exposure to ambient levels of sulfur dioxide are difficult to assess because of confounders such as the influence of other air pollutants.

In one limited acute study in the mouse, an acute LC<sub>50</sub> of 3,000 ppm was reported (Hilado and Machado 1977). For rats (n=8/group) continuously exposed to 590, 925, 2350, 50,000, or 500,000 ppm sulfur dioxide, average times of death were 1866, 750, 176, <10 and <2 minutes respectively (Cohen et al. 1973). The numbers of rats (n=8/group) which died within 2 weeks following exposure to 224, 593, 965, 1,168, or 1,319 ppm sulfur dioxide for 4 hours were 0, 0, 3, 5, and 8 respectively (Cohen et al. 1973).

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There are no reports of fatalities in humans or animals exposed by the oral route or solely by the dermal route.

**Systemic Effects**

***Respiratory Effects.*** Available data indicate that the respiratory system is the primary target system for sulfur dioxide toxicity following inhalation exposure at low levels (EPA 1994a, 1994b; Sheppard et al. 1981). Numerous controlled studies in humans have documented that acute-duration inhalation exposure to sulfur dioxide causes constriction of the airways, particularly in asthmatics and other sensitive individuals (Sheppard et al. 1981). Bronchoconstriction has been observed in asthmatics exposed to sulfur dioxide at concentrations as low as 0.1 ppm (Sheppard et al. 1981). Studies have also shown that exercise enhances the responsiveness to sulfur dioxide. Other factors that can exacerbate the respiratory effects of sulfur dioxide include exposure to cold or dry air and other pollutants such as sulfuric acid, nitrogen dioxide, and ozone. Healthy nonasthmatics respond to higher concentrations of sulfur dioxide (i.e.,  $\geq 1.0$  ppm) (EPA 1994a, 1994b).

Acute exposures to high concentrations of sulfur dioxide can result in severe and irreversible airway obstruction and in edematous lung tissue (Charan et al. 1979; Harkonen et al. 1983). However, actual concentrations of sulfur dioxide causing these effects have not been reported. Bronchitis has been reported following brief, accidental exposures to 100 ppm sulfur dioxide (Skalpe 1964).

Epidemiological studies on the relationship between sulfur dioxide and respiratory effects are limited because of the difficulties in separating potential effects of sulfur dioxide from those of particulates and other pollutants. Some epidemiological studies of children reveal reversible lung function changes such as decreases in FVC and FEV<sub>1</sub> but no effects on respiratory performance were noted in other studies (Agocs et al. 1997; Buchdahl et al. 1996; Dockery et al. 1982; Dodge et al. 1985; Shy et al. 1973). The effects of particulates cannot be separated from those of sulfur dioxide. Likewise, although lung function changes were reported in several occupational studies in which workers were also exposed to other chemicals such as arsenic (Archer and Gillan 1978; Smith et al. 1977). No significant relationship between pulmonary function and exposure to  $\geq 5$  ppm sulfur dioxide was noted in studies of potential lung function changes in copper smelter workers (Lebowitz et al. 1979) and steel plant workers (Lowe et al. 1970). Breathing difficulties and cough were reported by workers in a broom manufacturing facility who were exposed to 0.29-57 ppm sulfur dioxide and also to high concentrations of dust (Savic et al. 1987). Irritation and shortness of breath upon

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exertion were reported by workers in a refrigeration unit plant who were exposed to sulfur dioxide levels of 5-100 ppm (Kehoe et al. 1932). Cough, sputum production, and difficulty breathing were reported by pulp mill workers exposed to 2-36 ppm sulfur dioxide (Skalpe 1964).

Studies in experimental animals have indicated pulmonary effects of sulfur dioxide following inhalation exposure. Increased airway resistance and decreased compliance were noted in groups of guinea pigs exposed to 2.6 ppm sulfur dioxide for 1 hour (Amdur 1959). Increased pulmonary flow resistance was noted in guinea pigs exposed to 0.16-835 ppm sulfur dioxide (Amdur 1966). Similar effects were observed in dogs exposed to 1.1-141 ppm sulfur dioxide (Balchum et al. 1959). Acute-duration exposure to high concentrations of sulfur dioxide can result in biochemical, clinical, and histological changes in the respiratory systems of the mouse and rabbit. Degenerative changes in the olfactory epithelium were seen in mice exposed to 20 ppm sulfur dioxide for 120 minutes (Min et al. 1994). Rats exposed to 25 ppm sulfur dioxide for up to 5 days displayed nasal epithelial metaplasia and basal cell hyperplasia (Fowlie et al. 1990).

In an intermediate-duration study, mild bronchitic lesions were seen in hamsters exposed to 650 ppm sulfur dioxide for 19-74 days (Goldring et al. 1970). Inflammation of the bronchial mucosa was observed in rats exposed to 3040 ppm sulfur dioxide for 1 hour/day, 5 days/week, for 12 weeks (Krasnowska et al. 1998). Nasopharyngitis and lipid peroxidation of lung tissue were observed in guinea pigs exposed to 10 ppm sulfur dioxide for 1 hour/day for 30 days (Haider 1985). Increased numbers of goblet cells were observed in the airways of rats exposed to 400 ppm sulfur dioxide for 3 hours/day, 5 days/week (Basbaum et al. 1990; Lamb and Reid 1968).

The respiratory toxicity of sulfur dioxide following chronic-duration inhalation exposure has been studied in animals. No effects on lung function were observed in guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972). Likewise, no lung function changes or histopathological alterations in the lung were observed in monkeys exposed to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975).

***Cardiovascular Effects.*** Human, nonasthmatic subjects exposed by inhalation to 1-8 ppm sulfur dioxide for acute durations showed increased pulse rate (Amdur et al. 1953). No studies were located regarding cardiovascular toxicity after oral or dermal exposure.

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No evidence of histological lesions in the heart was found in monkeys following exposure to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). Likewise, no microscopic lesions were detected in the hearts of guinea pigs exposed by inhalation to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972). Lipid peroxidation was observed in the hearts of guinea pigs exposed to 10 ppm sulfur dioxide for 1 hour/day for 30 days (Haider 1985).

***Gastrointestinal Effects.*** Nausea and vomiting were observed in humans exposed to >40 ppm sulfur dioxide during an accident at a copper mine (Rabinovitch et al. 1989). No studies were located regarding gastrointestinal effects after oral or dermal exposure. No studies were located regarding gastrointestinal effects in animals.

***Hematological Effects.*** Exposure to sulfur dioxide levels of 5-100 ppm did not appear to affect blood cell numbers of workers in a refrigeration unit plant (Kehoe et al. 1932). Methemoglobin levels were increased in the blood of workers exposed to 0.29-57 ppm sulfur dioxide in a broom manufacturing facility (Savic et al. 1987).

An increase in erythrocyte deformability was observed in rats exposed to 1 ppm sulfur dioxide for 24 hours (Baskurt et al. 1990). Lipid peroxidation has been observed in the erythrocytes of rats exposed to 10 ppm sulfur dioxide for 1 hour/day for 45 days (Etlik et al. 1997) or for 1 hour/day, 7 days/week, for 8 weeks (Gumuslu et al. 1998). Osmotic hemolysis and sulfhemoglobin levels were increased in erythrocytes of rats exposed to 0.9 ppm sulfur dioxide for 24 hours (Baskurt et al. 1988). An increase in the erythrocyte deformability index and lipid peroxidation was noted in the erythrocytes of guinea pigs exposed to 10 ppm sulfur dioxide for 30 days (Dikmenoglu et al. 1991). No hematological effects were observed in a chronic toxicity study of monkeys exposed to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). Likewise, no hematological effects were noted in guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972). However, statistically significant increases in white and red blood cell counts, as well as hematocrit and hemoglobin levels were observed in rats exposed to 10 ppm sulfur dioxide for 1 hour/day for 45 days (Etlik et al. 1997). Increased hematocrit levels were also observed in rats exposed to 0.9 ppm sulfur dioxide for 24 hours (Baskurt et al. 1988).

***Musculoskeletal Effects.*** No studies were located regarding musculoskeletal effects in humans or animals after exposure to sulfur dioxide.

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**Hepatic Effects.** No adverse hepatic effects have been reported in humans exposed by inhalation to sulfur dioxide. No studies were located regarding hepatic effects in humans or animals after oral or dermal exposure.

Information on hepatic effects in animals following inhalation of sulfur dioxide is very limited. In a chronic toxicity study in monkeys, no histological lesions were detected in monkeys exposed to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). In contrast, microscopic examination of guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks showed an increase in the size of hepatocytes accompanied by cytoplasmic vacuolation (Alarie et al. 1972). Liver triglyceride and esterified cholesterol levels were increased in rats fed either a standard diet or a high cholesterol diet and exposed to 10 ppm sulfur dioxide for 24 hours/day, 7 days/week, for 15 days (Lovati et al. 1996). However, exposure of diabetic rats to 5 or 10 ppm sulfur dioxide under the same conditions resulted in a significant reduction of liver triglyceride levels. Liver cholesterol levels decreased in guinea pigs exposed to 10 ppm sulfur dioxide for 1 hour/day for 30 days (Haider 1985).

**Renal Effects.** No adverse renal effects have been reported in humans exposed by inhalation to sulfur dioxide. No studies were located regarding renal effects in humans or animals after oral or dermal exposure.

Information on renal effects in animals following inhalation of sulfur dioxide is very limited. No renal effects were observed in a chronic toxicity study of monkeys exposed to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). Likewise, no renal effects were noted in guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972).

**Dermal Effects.** Liquid sulfur dioxide from pressurized containers is a severe skin irritant in humans (Department of Labor 1975). Skin irritation has been observed in rats and guinea pigs exposed to 10 ppm sulfur dioxide for 1 hour/day for 30 days (Haider 1982, 1985).

**Ocular Effects.** In a case report of an industrial accident in which five persons were exposed by inhalation to high concentrations (not specified) of sulfur dioxide for less than 5 minutes, reversible conjunctivitis and superficial corneal burns were noted (Charan et al. 1979). In another case report dealing with a pyrite dust explosion that resulted in nine persons being exposed to high levels of sulfur dioxide, conjunctival irritation and corneal erosion were observed (Harkonen et al. 1983). The concentration of sulfur dioxide was not measured, but, in experimental explosions, the concentration was estimated to be 30-1,600 ppm. Burning of

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the eyes was reported in a case study of an accident at a copper mine in which miners were exposed to >40 ppm sulfur dioxide (Rabinovitch et al. 1989).

Sulfur dioxide gas is an eye irritant, causing burning and lacrimation (Department of Labor 1975). A concentration of 20 ppm is considered definitely irritating to the eyes (Ellenhorn and Barceloux 1988). Direct application of sulfur dioxide gas to the eyes of rabbits has resulted in severe damage to the cornea and conjunctiva, similar to that observed with chemical burns (Grant 1974).

***Body Weight Effects***- No body weight effects have been reported in humans following exposure to sulfur dioxide.

Following 5 weeks of inhalation exposure to 70-300 ppm sulfur dioxide, body weight gains of treated animals were 25% lower than those of controls (Miyata et al. 1990). No body weight effects were observed in a chronic toxicity study of monkeys exposed to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). Likewise, no body weight effects were noted in guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972) or in rats exposed to 10 ppm sulfur dioxide for 1 hour/day, 7 days/week for 8 weeks (Gumuslu et al. 1998).

**Endocrine Effects.** No studies were located regarding endocrine effects in humans after inhalation exposure to sulfur dioxide.

Significant dose-related decreases in plasma insulin levels were observed in rats exposed to 5 or 10 ppm sulfur dioxide for 24 hours/day, 7 days/week, for 15 days, but significant effects on insulin levels were not noted in diabetic rats exposed to sulfur dioxide under the same conditions (Lovati et al 1996).

**Immunological and Lymphoreticular Effects.** One study was located that examined the effect of sulfur dioxide on the airway response of human subjects with mild asthma to inhaled house dust mite allergen (Devalia et al. 1994). Prior inhalation of 0.2 ppm sulfur dioxide for 6 hours did not significantly affect the airway response to inhaled antigen, as measured by the dose of allergen required to produce a 20% decrease in forced expiratory volume in 1 second (PD<sub>20</sub> FEV<sub>1</sub>). However, exposure to both 0.2 ppm sulfur dioxide and 0.4 ppm nitrogen dioxide significantly reduced the PD<sub>20</sub> FEV<sub>1</sub>.

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Epidemiological studies have indicated that sulfur dioxide air pollution may increase the prevalence of allergies (Soyseth et al. 1996) and incidence of respiratory infections (French et al. 1973) in children. However, both studies were limited by inadequate control of confounding factors and a lack of personal exposure data.

Sulfites are potent sensitizers, and anaphylaxis can occur from exposure to residues in food (IARC 1992; Sheppard 1988; WHO 1979). Since sulfite-containing foods can off-gas sulfur dioxide, it is possible that some of the effects of sulfite ingestion may be due to inhalation of sulfur dioxide (Sheppard 1988).

Increased sensitization to antigen was reported in a study of guinea pigs exposed by inhalation to sulfur dioxide (Riedel et al. 1992). In this experiment, groups of 6 guinea pigs were exposed to air or 5 ppm sulfur dioxide for 8 hours/day for 5 days, with intermittent inhalation of ovalbumin (8 mg aerosolized) in order to sensitize the animals. The guinea pigs were also given two sensitizing injections of *Canida albicans* prior to sulfur dioxide exposure. Animals were exposed to the ovalbumin for 45 minutes following sham or sulfur dioxide exposure on days 3,4, and 5. Seven days after the last exposure, the guinea pigs were tested for bronchial sensitization to ovalbumin. Antibodies against ovalbumin were measured before and after sham and sulfur dioxide exposures. Sulfur dioxide-treated animals were sensitized to ovalbumin. Pretreatment with anti-inflammatory drugs (methylprednisolone, indomethacin, or nedocromilsodium) for 6 days beginning 12 hours before the first sulfur dioxide exposure prevented the sensitization to ovalbumin. Exposure of guinea pigs to 5 ppm sulfur dioxide for 4 hours/day, 5 days/week, for 6 weeks resulted in increased delayedtype dyspneic symptoms (Kitabatake et al. 1995).

In an acute-duration inhalation study with hamsters, there was a significant reduction in endocytosis by pulmonary macrophage following exposure to 50 ppm sulfur dioxide for 4 hours while exercising (Skornik and Brain 1990). However, pulmonary defense mechanisms were not affected in rats and mice exposed to 0.32-0.43 ppm sulfur dioxide together with 87-113  $\mu\text{g}/\text{m}^3$  sulfate for 4 hours prior to or 17 hours following infection with *Staphylococcus aureus* or Group C *Streptococci* (Goldstein et al. 1979).

**Neurological Effects.** Variations in reflex response time were observed in workers of a refrigeration unit plant who were exposed to sulfur dioxide concentrations ranging from 5-100 ppm (Kehoe et al. 1932). The effect was not consistent with neurological injury but may have been a psychological response to general sulfur dioxide toxicity. Seizures and prostration were observed in rats following exposure to 2350, 50,000 and 500,000 ppm sulfur dioxide for an average of 176 minutes, <10 minutes, and <2 minutes respectively

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(Cohen 1973). Lipid peroxidation has been observed in the brains of guinea pigs exposed to sulfur dioxide for 1 hour/day for 30 days (Haider et al. 1982). Transient changes in the frequency of certain behavioral patterns, such as grooming and digging, were noted in male and female mice continuously exposed to 30 ppm sulfur dioxide and observed during the first 9 days of a reproductive study. There was no evidence of neurological effects in the offspring of the mice (Petruzzi et al. 1996).

**Reproductive Effects.** In a cross-sectional study of spontaneous abortions in an industrial community in Finland, no evidence was found that exposure to sulfur dioxide was associated with a risk of spontaneous abortions (Hemminki and Niemi 1982). A study conducted in the Czech Republic found that abnormal sperm morphology was associated with sulfur dioxide, which was used as a surrogate for all air pollutants (Selevan et al. 1995). Reproductive effects, as measured by completed pregnancies, litter size, sex ratio, and neonatal mortality, were not observed in male and female mice continuously exposed to 5, 12, or 30 ppm sulfur dioxide for 9 days prior to mating until days 12-14 of pregnancy (Petruzzi et al. 1996). The offspring were not assessed for reproductive function.

**Developmental Effects.** An association between exposure to increased levels of air pollution during pregnancy and a reduction in birth weight has been reported in humans (Wang et al. 1997). Decreased birth weights were noted for exposure to both sulfur dioxide and total suspended particulates (TSPs). However evaluation of these results is complicated by limitations such as confounding effects of various air pollutants and a lack of personal exposure data. Therefore, differentiation of potential developmental effects associated with individual pollutants of a mixture is not possible.

No developmental effects were observed in a study in which pregnant mice were exposed to 32-250 ppm sulfur dioxide on gestation days 7-17 (Singh 1982). Neurological effects, measured by reflexes and learning ability, were not observed in the offspring of mice continuously exposed to 5-30 ppm sulfur dioxide 9 days prior to mating through pregnancy days 12-14 (Petruzzi et al. 1996). Although there were some transient behavioral effects in dams, there were no signs of systemic maternal toxicity. In another developmental study in mice in which pregnant females were exposed to 32 ppm or 65 ppm sulfur dioxide on gestation days 7-18, increased time for righting reflex on postnatal day 1 and increased negative geotaxis on postnatal day 10 were reported (Singh 1989). The duration of exposure for each day was not stated. No visible signs of maternal toxicity and no effect on the number of live births were observed. Sulfur dioxide at a concentration of 65 ppm significantly decreased the birth weight (about 89% of controls) of the pups. Minor skeletal variations were reported in offspring of mice exposed to 25 ppm sulfur dioxide for 7 hours/day on gestation



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days 6-15 and rabbits exposed to 70 ppm sulfur dioxide for 7 hours/day on gestation days 6-18 (Murray et al. 1979). Fetal body weights were also lower in exposed mice. Reduced food intake was the only sign of maternal toxicity.

**Genotoxic Effects.** There are some limited data showing clastogenic effects in humans following occupational exposure to sulfur dioxide. Increases in chromosome aberrations and sister chromatid exchanges were detected in lymphocytes from 42 workers of an Indian fertilizer factory who were exposed to an average concentration of  $41.7 \text{ mg/m}^3$  (15.92 ppm) of sulfur dioxide (Yadav and Kaushik 1996). Confounding factors were not discussed. Similar findings were observed in another study of 40 workers occupationally exposed to sulfur dioxide at a Chinese sulfuric acid plant (Meng and Zhang 1990a). Exposure concentrations ranged from  $0.34 \text{ mgm}^3$  to  $11.97 \text{ mg/m}^3$  (0.13-4.57 ppm). In addition, increases in the frequencies of lymphocytes with micronuclei were noted (Meng and Zhang 1990b). A significant increase in the frequency of chromosomal aberrations was found among 19 workers at a sulfite pulp factory (Nordenson et al. 1980). Exposure concentrations were not reported and workers were also exposed to chlorine and dust. One study of potential chromosomal abnormalities in workers exposed to sulfur dioxide in the aluminum industry revealed that sulfur dioxide was without effect (Sorsa et al. 1982).

Mutations were not observed in germ cells following *in vivo* exposure of animals to sulfites (metabolites of sulfur dioxide). In a dominant lethal test, mutations were not observed in offspring of male mice treated by peritoneal injection with 300 mg/Kg sodium bisulfite on 38 out of 54 days or 400 mg/kg sodium bisulfite 5 days/week for a total of 20 treatments (Russell and Kelly 1975; Shapiro et al. 1977). A specific-locus test was conducted in the same wild type mice from the 400 mg/kg treatment group, which were mated to females with homozygous marker genes for seven loci (Russell and Kelly 1975). Mutations associated with abnormalities in sperm were not observed in 13,568 offspring which were scored for mutations at seven loci. Table 2-3 summarizes data on genotoxicity of sulfur dioxide *in vivo*.

Table 2-4 summarizes data on the genotoxicity of sulfur dioxide *in vitro*. Chromosomal effects were observed following *in vitro* testing with sulfur dioxide. Lymphocytes from healthy donors were cultured and treated with 100 cc of 5.7 ppm sulfur dioxide, which was bubbled through the culture media (Schneider and Calkins 1970). Twenty percent of the hypotonic smears obtained from treated cells contained abnormalities which were characterized primarily by clumping of chromosomes. Abnormalities were not observed in any of the air-treated controls.

TABLE 2-3 Genotoxicity of Sulfur Dioxide *In Vivo*

Species (test system)	End point	Results	Reference
<i>Mammalian cells:</i>			
Human peripheral lymphocytes	Sister chromatid exchanges and chromosomal aberrations	+	Meng and Zhang 1990a
Human peripheral lymphocytes	Micronuclei	+	Meng and Zhang 1990b
Human peripheral lymphocytes	Chromosomal aberrations	+	Nordenson et al. 1980
Human peripheral lymphocytes	Sister chromatid exchanges and chromosomal aberrations	-	Sorsa et al. 1982
Human peripheral lymphocytes	Sister chromatid exchanges and chromosomal aberrations	+	Yadar and Kaushik 1996
Mice (Dominant-Lethal test)	Gene mutations	-	Russell and Kelly 1975; Shapiro et al. 1977
Mice (Specific-locus test)	Gene mutations	-	Russell and Kelly 1975

- = negative result; + = positive result

TABLE 2-4 Genotoxicity of Sulfur Dioxide *In Vitro*

Species (test system)	End point	Results		Reference
		With activation	Without activation	
<i>Eukaryotic organisms:</i>				
Animal:				
Cow Oocytes	Chromosomal Aberration	+		Jagiello et al. 1975
Ewe Oocytes	Chromosomal Aberration	+		Jagiello et al. 1975
Plant:				
<i>Tradescantia paludosa</i> clone O <sub>3</sub> (liquid absorption treatment)	Micronuclei	No data	(+)	Ma et al. 1984
<i>Tradescantia paludosa</i> clone 4430 (gaseous exposure)	Micronuclei	No data	(+)	Ma et al. 1973
Fungi:				
<i>Saccharomyces cerevisiae</i>	Gene mutation	+	No data	Guerra et al. 1981

+ = positive result; (+) = weakly positive result

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Chromosomes from mouse, cow, and ewe oocytes appeared fuzzy following *in vitro* exposure to 250 µg/m<sup>3</sup> sodium sulfite and fragmentation and clumping were observed in the ewe and cow oocytes (Jagiello et al. 1975). Meiosis was inhibited in mouse and cow oocytes treated with 150 and 500 µg/m<sup>3</sup> sodium sulfite respectively. However, oocyte meiosis was not inhibited in mice administered 1-5 mg sodium sulfite intravenously, thus supporting the conclusion that *in vivo* exposure of sulfur dioxide will not result in genotoxic effects of germ cells (Jagiello et al. 1975).

Sulfur dioxide induced micronuclei in eukaryotic plants (Ma et al. 1973, 1984). Sulfur dioxide caused gene mutation in *Saccharomyces cerevisiae* (Guerra et al. 1981).

The available *in vivo* and *in vitro* data indicate a genotoxic potential for sulfur dioxide. The mechanism of the potential genotoxicity of sulfur dioxide is unknown; however, a metabolite of sulfur dioxide, i.e., sulfite, can cause an inhibition of DNA synthesis and induce chromosomal aberrations in human lymphocytes (Yadav and Kaushik 1996).

**Cancer.** There are no definitive data in humans or animals that indicate a carcinogenic potential for sulfur dioxide. Several epidemiological studies have been conducted on workers in the copper smelting and pulp and paper industries in which exposure to sulfur dioxide can occur (Enterline et al. 1987; IARC 1992; Lubin et al. 1981; Welch et al. 1982). However, many of the copper smelter studies focused primarily on the association between arsenic exposure and respiratory cancer (Enterline et al. 1987; Lubin et al. 1981; Welch et al. 1982). It has been difficult to separate the potential effects of sulfur dioxide and arsenic exposures in the copper smelter studies. IARC has classified sulfur dioxide as a Group 3 carcinogen, i.e., not classifiable as to its carcinogenicity. EPA has not assigned sulfur dioxide a cancer classification.

One chronic-duration animal study investigated the potential carcinogenicity of inhaled sulfur dioxide in mice (Peacock and Spence 1967). Male and female mice were exposed to 500 ppm sulfur dioxide for 5 minutes/day, 5 days/week, for 2 years. A significant increase in the incidence of lung tumors (13/30 adenomas and carcinomas versus 5/30 in controls; 4/30 primary carcinomas versus none in controls) was observed in the female mice. The incidence of lung adenomas and carcinomas was also higher in the treated males (15/28 versus 11/35 in controls), but the increase was not significant. The incidence of primary carcinomas in treated males was similar to that of the controls. Although limited by small sample size and testing of just one dose, the study suggests that sulfur dioxide may be a carcinogen in mice. Studies in additional species are required before a conclusion can be made about carcinogenic potential.

## 2.6 CHILDREN'S SUSCEPTIBILITY

This section discusses potential health effects from exposures during the period from conception to maturity at 18 years of age in humans, when all biological systems will have fully developed. Potential effects on offspring resulting from exposures of parental germ cells are considered, as well as any indirect effects on the fetus and neonate due to maternal exposure during gestation and lactation. Relevant animal and in vitro models are also discussed.

Children are not small adults. They differ from adults in their exposures and may differ in their susceptibility to hazardous chemicals. Children's unique physiology and behavior can influence the extent of their exposure. Exposures of children are discussed in section 5.6 Exposures of Children.

Children sometimes differ from adults in their susceptibility to hazardous chemicals, but whether there is a difference depends on the chemical (Guzelian et al. 1992; NRC 4993). Children may be more or less susceptible than adults to health effects and the relationship may change with developmental age (Guzelian et al. 1992; NRC 1993). Vulnerability often depends on developmental stage. There are critical periods of structural and functional development during both pre-natal and post-natal life and a particular structure or function will be most sensitive to disruption during its critical period(s). Damage may not be evident until a later stage of development. There are often differences in pharmacokinetics and metabolism between children and adults. For example, absorption may be different in neonates because of the immaturity of their gastrointestinal tract and their larger skin surface area in proportion to body weight (Morselli et al. 1980; NRC 1993); the gastrointestinal absorption of lead is greatest in infants and young children (Ziegler et al. 1978). Distribution of xenobiotics may be different; for example, infants have a larger proportion of their bodies as extracellular water and their brains and livers are proportionately larger (Widdowson and Dickerson 1964; Foman et al. 1982; Owen and Brozek 1966; Altman and Dittmer 1974; Foman 1966). The infant also has an immature blood-brain barrier (Adinolfi 1985; Johanson 1980) and probably an immature blood-testis barrier (Setchell and Waites 1975). Many xenobiotic metabolizing enzymes have distinctive developmental patterns and at various stages of growth and development, levels of particular enzymes may be higher or lower than those of adults and sometimes unique enzymes may exist at particular developmental stages (Leeder and Kearns 1997; Komori 1990; Vieira et al. 1996; NRC 1993). Whether differences in xenobiotic metabolism make the child more or less susceptible also depend on whether the relevant enzymes are involved in activation of the parent compound to its toxic form or in detoxification. There may also be differences in excretion, particularly in the newborn who has a low glomerular filtration rate and has not

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developed efficient tubular secretion and resorption capacities (Altman and Dittmer 1974; West et al. 1948; NRC 1993). Children and adults may differ in their capacity to repair damage from chemical insults. Children also have a longer lifetime in which to express damage from chemicals; this potential is particularly relevant to cancer.

Certain characteristics of the developing human may increase exposure or susceptibility while others may decrease susceptibility to the same chemical. For example, the fact that infants breathe more air per kilogram of body weight than adults may be somewhat counterbalanced by their alveoli being less developed, so there is a disproportionately smaller surface area for absorption (NRC 1993).

Clinical studies were conducted to examine respiratory function in healthy, atopic, or asthmatic adolescents (n=8-9, aged 12-17 years) and 10 healthy male seniors (aged 55-73 years) following a mouth only exposure to a mixture of 1 ppm sulfur dioxide and 1 mg/m<sup>3</sup> sodium chloride aerosol (Koenig et al. 1982a, 1982b, 1983; Rondinelli et al. 1987). Adolescents exercised for 10 minutes of a 40 minute exposure period and the seniors exercised for 10 minutes of a 30 minute exposure period. Statistically significant decreases in forced expiratory volume in 1 second (FEV<sub>1</sub>) were observed in asthmatic adolescents (-23%), atopic adolescents (-18%), seniors (-8%), and healthy adolescents (-6%). This study concluded that respiratory health status and not age is the primary factor in determining susceptibility to sulfur dioxide in adolescents and adults because asthmatic adolescents were most sensitive to sulfur dioxide and normal adolescents were least sensitive (Rondinelli et al. 1987). However, the study did not examine susceptibility to sulfur dioxide in younger children and adults, so conclusions about sensitivity in infants and preadolescent children cannot be made. Subjects were exposed to sulfur dioxide in combination with sodium chloride aerosols because sulfur dioxide is found together with one or more droplet aerosols in the atmosphere. In two studies, it was verified that exposure to a mixture of sulfur dioxide and sodium chloride aerosol resulted in effects which did not differ significantly from those obtained from exposure to sulfur dioxide alone (Koenig et al. 1982b, 1982a).

Exposure of 9 atopic adolescents (aged 12-18 years) to 0.1 ppm sulfur dioxide by mouth did not result in statistically significant changes in respiratory function parameters (Koenig et al. 1989). However, there was some evidence that exposure to 0.1 ppm sulfur dioxide may potentiate sulfuric acid-induced respiratory effects. Exposure to 0.068 mg/m<sup>3</sup> sulfuric acid by itself resulted in a nonsignificant, 12% increase in total respiratory resistance. However, a statistically significant, 15% increase in respiratory resistance was observed following exposure to both 0.1 ppm sulfur dioxide and 0.068 mg/m<sup>3</sup> sulfuric acid.

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Epidemiological evidence indicates a possible association between sulfur dioxide levels in air and respiratory symptoms in children. In a longitudinal study of children in Arizona, increased prevalence of coughing was noted following exposure to high intermittent levels of sulfur dioxide with a peak 3 hour mean  $>2.5 \text{ mg/m}^3$  (0.95 ppm) and average levels of particulates (Dodge et al. 1985). In Steubenville, OH, small transient reductions in forced vital capacity (FVC) and forced expiratory volume in 0.75 seconds ( $\text{FEV}_{0.75}$ ) were observed in children following acute episodes of air pollution with sulfur dioxide levels  $\geq 0.17 \text{ ppm}$  and total suspended particulate levels that exceeded the 1979 standard (Dockery et al. 1982).

In New York seasonal levels of sulfur dioxide,  $63\text{-}71 \text{ }\mu\text{g/m}^3$  (0.024-0.027 ppm), and other air pollutants (total suspended particulates (TSPs), suspended sulfates, and nitrogen dioxide) were associated with reduced  $\text{FEV}_{0.75}$ , in children aged 9-13 years but not in children aged 5-8 years (Shy et al. 1973). Three years prior to the study, levels of sulfur dioxide, TSPs, and suspended sulfates were greatly reduced. Sulfur dioxide levels previously ranged between  $364\text{-}435 \text{ }\mu\text{g/m}^3$  (0.14-0.17 ppm). It was concluded that persistent respiratory effects in the older children resulted from exposure to high levels of sulfur dioxide, TSPs, and suspended sulfates during the first 5-10 years of life. Effects associated with individual air pollutants were not separated and there was no control for other confounding factors.

There was no association between peak expiratory flow rates (PEFR) and seasonal levels of sulfur dioxide,  $11\text{-}185 \text{ }\mu\text{g/m}^3$  (0.004-0.071 ppm), and particulate pollution in relation to 60 asthmatic children (aged 9-14 years) residing in Budapest (Agocs et al. 1997). Air pollutants were evaluated separately and numerous confounding factors were controlled. However, the authors discussed limitations such as medication use by subjects and a lack of personal exposure data.

In London, emergency room visits for acute wheezing episodes in children increased by 12% for each  $14 \text{ }\mu\text{g/m}^3$  (0.005 ppm) increase in sulfur dioxide level (Buchdahl et al. 1996). Effects from other air pollutants (ozone and nitrogen dioxide) were not separated in the evaluation of sulfur dioxide but were controlled for in the evaluation of ozone. The association with wheezing episodes in children was strongest for ozone.

Increased prevalence of allergies was observed in 556 children (aged 7-13 years) living near an aluminum smelter in Norway for 7 years or more (Soyseth et al. 1996). Prevalence was highest in children who were exposed to  $20\text{-}24 \text{ }\mu\text{g/m}^3$  (0.008-0.009 ppm) sulfur dioxide between the ages of 19-36 months. However, there was no evidence of a dose response relationship and fluoride levels were also increased in the area.

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There was some evidence of increased incidence of respiratory infection in children who lived within four regions of the United States with high levels of sulfur dioxide, 63-275  $\mu\text{g}/\text{m}^3$  (0.024-0.1 ppm) and other air pollutants (French et al. 1973). In the Salt Lake Basin and Rocky Mountain region, the highest incidence of respiratory infection was observed in children 4 years-old and younger. However, in Chicago, there was no association between respiratory illness and air pollution in children who were too young to attend nursery school. Polluted areas of the Salt Lake Basin, Rocky Mountain regions, and Chicago were also characterized by high levels of sulfates, metals, and TSPs, respectively. Differences in the chemical composition of air pollutants or inadequate control of confounding factors are possible reasons for the different findings in very young children.

As previously mentioned, epidemiological studies are limited due to confounding effects of particulates and other air pollutants. It has been reported that particulate sulfates, which form when sulfur oxides are released into the atmosphere, have a greater impact on health than sulfur dioxide (Shy 1977). Sulfate levels as low as 0.009-0.013  $\text{mg}/\text{m}^3$  may adversely affect lung function and increase the incidence of acute respiratory disease in children. The exposure levels are similar to those associated with adverse cardiopulmonary effects in adults (0.006-0.010  $\text{mg}/\text{m}^3$ ). Additional research is required to assess the impact of individual air pollutants on children's health.

Asthmatics in general are most susceptible to sulfur dioxide exposure, especially during periods of exercise (EPA 1994a, 1994b). Prevalence of asthma is highest in African Americans, children between the ages of 8 to 11, and urban residents (EPA 1994b). Mortality rates associated with asthma are higher in nonwhite individuals, but the reason is not known (EPA 1994b). Increased susceptibility to sulfur dioxide is therefore expected in asthmatic, minority children living in urban areas.

Following inhalation of radiolabeled sulfur dioxide by dogs, a very small percentage of radiolabel was detected in gonads, 0.01-0.18% in ovaries, and 0.006-0.1% in testicles, but the chemical form of the radiolabel was not known (Balchum et al. 1959). There are no known studies which examine whether sulfur dioxide metabolites can pass the placenta or be excreted into breast milk. Because sulfur dioxide and its metabolites are water soluble, accumulation in maternal tissues and subsequent mobilization during pregnancy or lactation is not likely.

Evidence of reproductive and developmental effects in humans is limited. No association was observed between exposure to sulfur dioxide and spontaneous abortion in 1,792 cases of a cross-sectional study of an



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industrial community in Finland (Hemminki and Niemi 1982). In the Czech Republic an association was noted between a high sulfur dioxide level (as a marker for all air pollutants) and an increased incidence of abnormality and reduced motility of sperm in 325 18-year-old males (Selevan et al. 1995). A developmental study conducted in China indicated an association between exposure to sulfur dioxide pollution during pregnancy and decreased infant birth weight in 74,671 women (Wang et al. 1997). However, interpretation of such studies is complicated by limitations such as confounding effects of various air pollutants and a lack of personal exposure data.

Reproductive effects following exposure to sulfur dioxide have not been observed in animals. Exposure of male and female mice (10/sex/group) to 5-30 ppm sulfur dioxide for 9 days prior to mating until days 12-14 of pregnancy had no effect upon completed pregnancies, litter size, sex ratio, and neonatal mortality (Petruzzi et al. 1996). Significant differences in pregnancy rate, implantations, resorptions, or live fetuses were not observed in 20 rabbits exposed to 70 ppm sulfur dioxide on gestation days 6-18 or 32 mice exposed to 25 ppm sulfur dioxide on gestation days 5-15 for 7 hours/day (Murray et al. 1977, 1979).

Serious developmental effects following exposure to sulfur dioxide have only been observed in one study. Increased time for righting reflex on postnatal day 1 and negative geotaxis on postnatal day 10 were observed in the offspring of mice exposed to 32 or 65 ppm sulfur dioxide on gestation days 7-18 (Singh 1989). However, neurological effects were not noted in the offspring of 10 mice exposed to 5-30 ppm sulfur dioxide 9 days prior to mating until days 12-14 of pregnancy (Petruzzi et al. 1996). Exposure of mice (13-17/group) to 32-250 ppm sulfur dioxide on gestation days 7-17 did not result in adverse developmental effects (Singh 1982). Serious malformations were not observed in 20 rabbits exposed to 70 ppm sulfur dioxide on gestation days 6-18 and 32 mice exposed to 25 ppm sulfur dioxide on gestation days 5-15 for 7 hours/day, but minor skeletal variations were noted (Murray et al. 1977, 1979). Effects in rabbits included nonossified sections of bone, fusion of cranial bones, and extra ribs. Delayed ossification and decreased fetal weight were observed in treated mice.

Genotoxic compounds could potentially affect children's health through the induction of mutations in the parental germ cells. *In vitro* experiments have demonstrated a genotoxic effect of sulfur dioxide metabolites on animal germ cells, but adverse effects have not been observed following *in vivo* exposure of animals. Mouse, cow, and ewe oocytes were incubated in cell culture media containing sodium sulfite (Jagiello et al. 1975). An inhibitory effect on meiosis was noted in the mouse and cow oocytes at 150 and 500  $\mu\text{m}^3$  sodium sulfite, respectively. At 250  $\mu\text{g}/\text{cm}^3$  sodium sulfite, the chromosomes of all three species appeared

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fuzzy. Fragmentation and clumping were also observed in the chromosomes of ewe and cow oocytes. Genetic disorders can occur in the offspring which develop from oocytes containing fragmented chromosomes. Therefore, the effects of sodium sulfite on oocytes were studied following intravenous administration of 1-5 mg sodium sulfite to 6 mice (Jagiello et al. 1975). In *vivo* administration of sodium sulfite had no effect on the meiotic division of the oocytes.

In a dominant-lethal test, mutations were not observed in the offspring of 10 male mice injected peritoneally with 300 mg/kg sodium bisulfite on 38 out of 54 days or 400 mg/kg sodium bisulfite 5 days/week for a total of 20 treatments (Russell and Kelly 1975; Shapiro et al. 1977). Seven and a half weeks later, the mice (wildtype) from the latter treatment group were mated to females with homozygous marker genes for seven loci (Russell and Kelly 1975). No mutations that may have resulted from an abnormality in sperm were observed in a total of 13,568 offspring which were scored for mutations at the seven loci. The authors estimated the upper limit of genetic risk to humans and concluded that exposure to sulfur dioxide would not likely result in adverse genetic effects in germ cells. Studies which examined effects of *in utero* sulfur dioxide exposure on developing germ cells and possible effects on future generations were not located.

One study has also indicated that maternal nutrition may affect the offspring's susceptibility to sulfur dioxide later in life. Increased evidence of pulmonary injury, measured by protein in bronchoalveolar lavage fluid, was observed in 10 rats exposed to a low protein diet (60 g caisin/kg) *in utero* and 0.29 mg/m<sup>3</sup> (0.11 ppm) sulfur dioxide later in life (Langley-Evans et al. 1997).

Studies which specifically examine metabolism in children were not identified. Upon contact with moisture in airways, sulfur dioxide is converted to a mixture of sulfite, bisulfite, and hydrogen ions (Gunnison et al. 1987). Sulfites are then converted to sulfates by sulfite oxidase. Age-related differences in sulfite oxidase activity have been demonstrated in rats (Cohen 1974). In 1-day-old rats, liver sulfite oxidase activity was approximately 1/10 the level of adult rats. Activity increased as the rats matured and at 32 days of age, the liver sulfite oxidase activity was approximately one half the level of adult rats. Decreased sulfite oxidase activity may result in increased susceptibility to oxidative effects due to increased formation of a sulfur trioxide radical intermediate (Constantin 1996). Animal studies have demonstrated an inverse relationship between sulfite oxidase activity and susceptibility to intraperitoneally administered bisulfite as measured by lethality (Tejnorova 1978).

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Pharmacokinetic studies or PBPK models which specifically look at absorption, distribution, and excretion in children or immature animals were not identified. The solubility of gases determine where in the respiratory tract absorption will occur and absorption in the lungs is based upon the blood-to-gas partition coefficient (Amdur et al. 1991). Because solubility and blood-to-gas partition coefficient are constants, absorption of sulfur dioxide in children is expected to be similar to that of adults. Distribution and excretion are also expected to be similar in children and adults. General information about pharmacokinetics can be obtained in Section 2.3.

Studies which determine if mechanisms of toxicity differ in children were not identified. Evidence indicates that sulfur dioxide-induced bronchoconstriction occurs through reflexes which are mediated by cholinergic and noncholinergic factors. The percent contribution from cholinergic versus noncholinergic mechanisms is not known and it could potentially differ between children and adults. However, this has not been verified through research. General mechanisms of toxicity are discussed in Section 2.4.

There are no studies which look at interactions with other chemicals that are unique to children. Studies in animals have indicated that exposure to other air pollutants may potentiate reproductive or developmental effects of sulfur dioxide. Exposure of 20 rabbits to a mixture of 70 ppm sulfur dioxide and 250 ppm carbon monoxide 7 hours/day on gestation days 6-18 resulted in a significant increase in resorptions/litter, which was not observed with exposure to sulfur dioxide or ozone alone (Murray et al. 1977, 1979). Exposure of 32 mice to a combination of 25 ppm sulfur dioxide and 250 ppm carbon monoxide 7 hours/day on gestation days 5-15 potentiated the decrease in fetal weight gain which was observed with exposure to sulfur dioxide alone (Murray et al. 1977, 1979). Exposure of pregnant mice to ozone alone had the opposite effect and resulted in a significant increase in fetal weight gain. A significant decrease in fetal crown-rump length was also observed in the same mice following exposure of dams to the mixture of pollutants but not sulfur dioxide alone. Effects on fetal crown-rump length following exposure to ozone alone were not discussed.

It is not known whether biomarkers or methods for reducing toxicity to exposure differ in children.

In conclusion, studies of susceptibility in children are limited. Clinical studies have indicated that compared to senior citizens, adolescents are not at increased risk of respiratory effects following inhalation of sulfur dioxide. Adverse respiratory effects may be associated with exposure of children to air pollution, but it is difficult to determine if sulfur dioxide is directly responsible for those effects. The genotoxic potential of sulfur dioxide metabolites has been demonstrated through *in vitro* studies, but *in vivo* exposure of animals

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has indicated that mutations in germ cells are unlikely following inhalation exposure of sulfur dioxide. Difficulties in separating the effects of individual air pollutants also complicate the evaluation of sulfur dioxide pollution on human reproduction and gestational development. Results from animal developmental studies are equivocal. Most animal studies have indicated a lack of serious developmental effects but other studies have reported developmental delays which might have resulted from decreased food intake by dams. Lastly, decreased sulfite oxidase activity in young animals has been demonstrated. If the same is true for children, they may be at increased risk for oxidative damage.

**2.7 BIOMARKERS OF EXPOSURE AND EFFECT**

Biomarkers are broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility (NAUNRC 1989).

Due to a nascent understanding of the use and interpretation of biomarkers, implementation of biomarkers as tools of exposure in the general population is very limited. A biomarker of exposure is a xenobiotic substance or its metabolite(s), or the product of an interaction between a xenobiotic agent and some target molecule(s) or cell(s) that is measured within a compartment of an organism (NRC 1989). The preferred biomarkers of exposure are generally the substance itself or substance-specific metabolites in readily obtainable body fluid(s) or excreta. However, several factors can confound the use and interpretation of biomarkers of exposure. The body burden of a substance may be the result of exposures from more than one source. The substance being measured may be a metabolite of another xenobiotic substance (e.g., high urinary levels of phenol can result from exposure to several different aromatic compounds). Depending on the properties of the substance (e.g., biologic half-life) and environmental conditions (e.g., duration and route of exposure), the substance and all of its metabolites may have left the body by the time samples can be taken. It may be difficult to identify individuals exposed to hazardous substances that are commonly found in body tissues and fluids (e.g., essential mineral nutrients such as copper, zinc, and selenium). Biomarkers of exposure to sulfur dioxide are discussed in Section 2.7.1.

Biomarkers of effect are defined as any measurable biochemical, physiologic, or other alteration within an organism that, depending on magnitude, can be recognized as an established or potential health impairment or disease (NAS/NRC 1989). This definition encompasses biochemical or cellular signals of tissue dysfunction (e.g., increased liver enzyme activity or pathologic changes in female genital epithelial cells), as well as physiologic signs of dysfunction such as increased blood pressure or decreased lung capacity. Note that these

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markers are not often substance specific. They also may not be directly adverse, but can indicate potential health impairment (e.g., DNA adducts). Biomarkers of effects caused by sulfur dioxide are discussed in Section 27.2.

A biomarker of susceptibility is an indicator of an inherent or acquired limitation of an organism's ability to respond to the challenge of exposure to a specific xenobiotic substance. It can be an intrinsic genetic or other characteristic or a preexisting disease that results in an increase in absorbed dose, a decrease in the biologically effective dose, or a target tissue response. If biomarkers of susceptibility exist, they are discussed in Section 2.9, Populations That Are Unusually Susceptible.

### 2.7.1 Biomarkers Used to Identify or Quantify Exposure to Sulfur Dioxide

No studies pertaining to specific biomarkers used to identify or quantify exposure to sulfur dioxide were located.

Sulfur dioxide metabolites are rapidly absorbed from the upper respiratory passages and are readily distributed throughout the body (Balchum et al. 1960a; Frank et al. 1967; Kleinman 1984; Speizer and Frank 1966; Yokoyama et al. 1971). Sulfur dioxide reacts with plasma proteins to form *S*-sulfonates. *S*-Sulfonate levels in human plasma showed a positive correlation with atmospheric levels (> 0.3 ppm) of sulfur dioxide (Gunnison and Palmes 1974). Therefore, plasma *S*-sulfonate levels may serve as a potential indicator of exposure to sulfur dioxide.

Inhaled sulfur dioxide is excreted in the urine as sulfate. However, sulfur dioxide is not the only source of sulfate in the urine. Sulfates are also metabolites of sulfur-containing amino acids and are therefore normal constituents of intracellular fluids and urine (Vander et al. 1975; Lentner 1981). Concentrations of sulfates in urine range from 53  $\mu\text{mol/dL/Kg}$  in 1-day-old infants to 500  $\mu\text{mol/dL/Kg}$  in young men (Lentner 1981). The level of urinary sulfate is a measure of the quantity and quality of proteins in the diet.

### 2.7.2 Biomarkers Used to Characterize Effects Caused by Sulfur Dioxide

No specific biomarkers used to characterize effects caused by sulfur dioxide were located.

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The upper respiratory system is the primary target of sulfur dioxide. Alterations in pulmonary function parameters, such as increased airway resistance, decreased airway conductance, and decreased expiratory volume, might be considered potential biomarkers to characterize effects caused by sulfur dioxide, but alterations in pulmonary function parameters are not specific for sulfur dioxide and may indicate exposure to other air pollutants such as nitrogen dioxide or ozone.

An increase in the number of leukocytes in bronchoalveolar lavage fluid is a biomarker for inflammatory effects of sulfur dioxide exposure (Sandstrom 1989a, 1989b). However, the effect is not specific to sulfur dioxide because inflammation can occur following exposure to numerous irritating substances.

DNA adduct formation in the nasal mucosa is a potential biomarker which could be used in the future. The formation of adducts between the DNA base cytosine and bisulfite, a metabolite of sulfur dioxide, has been demonstrated *in vitro* (Shapiro 1977). Adduct formation in placentas of women from industrial and agricultural areas in the Czech Republic has been compared in order to assess toxicity from exposure to pollution, and no differences were found between the two groups (Topinka et al. 1995). However, the nasal mucosa is the optimal tissue for the development of biomarker techniques because it is the first to contact inhaled toxicants (Flato et al. 1996). A technique for measuring adduct formation in cells obtained from nasal lavage has been demonstrated (Flato et al. 1996). However, studies are needed to verify the formation of bisulfite adducts in nasal mucosa. Adduct formation would not be specific to sulfur dioxide because adducts can form after exposure to numerous carcinogenic and mutagenic compounds.

Clastogenic effects in humans following occupational exposure to sulfur dioxide via the inhalation route have been reported. Increases in chromosome aberrations and sister chromatid exchanges were detected in lymphocytes from 42 workers who were occupationally exposed to an average concentration of 41.7 mg/m<sup>3</sup> (15.92 ppm) of sulfur dioxide (Yadav and Kaushik 1996). Similar findings were observed in another study of 40 workers occupationally exposed to sulfur dioxide (Meng and Zhang 1990a). Exposure concentrations ranged from 0.34 to 11.97 mg/m<sup>3</sup> (0.13 to 4.57 ppm). In addition, increases in the frequencies of lymphocytes with micronuclei were noted (Meng and Zhang 1990b). Such clastogenic effects may serve as potential biomarkers for genotoxic effects of sulfur dioxide. However, these clastogenic effects are not specific for sulfur dioxide and could indicate exposure to other chemicals.

For more information on biomarkers for renal and hepatic effects of chemicals see ATSDR/CDC Subcommittee Report on Biological Indicators of Organ Damage (1990).

## 2.8 INTERACTIONS WITH OTHER CHEMICALS

Epidemiological studies of the associations between effects of sulfur dioxide and exposure to other chemicals are limited because of the presence of other air pollutants as confounders. A number of other air pollutants such as nitrogen dioxide, sulfuric acid, particulate matter, and ozone can result in respiratory effects similar to those of sulfur dioxide. There is some evidence that particulate matter, which is capable of oxidizing sulfur dioxide to sulfuric acid, can cause a three- to four-fold potentiation of the irritant response in guinea pigs (Amdur 1969). It is postulated that binding of acids to carbon particles may impede neutralization by ammonia in the airways or buffering system in epithelium (Jakab 1996). Under conditions of high relative humidity, sulfate is produced from mixtures of sulfur dioxide and carbon particles. Acute exposure of mice to a mixture of carbon particles and sulfur dioxide at 85% relative humidity resulted in an inhibition of phagocytosis by alveolar macrophages, which was not observed from exposure to sulfur dioxide alone (Jakab 1996).

Concurrent exposures to sulfur dioxide, smoke, and particulates have been associated with symptoms including: increased respiratory effects, increased frequencies of respiratory illness, excess mortality, and worsening of existing respiratory disease (WHO 1979). Table 2-5 summarizes the effects of concurrent exposure to sulfur dioxide, smoke, and particulates, and it designates which portions of the population are affected.

In a controlled study of asthmatic subjects, an enhanced responsiveness to 0.75 ppm sulfur dioxide after exposure to 0.25 ppm nitrogen dioxide for 30 minutes was reported (Jorres and Magnussen 1990). The authors suggested that acute exposure of asthmatics to nitrogen dioxide at rest enhances airway responsiveness to hyperventilation of sulfur dioxide without altering airway tone. In another study of asthmatics, the influence of prior exposure to a low concentration of ozone (0.12 ppm) on the pulmonary response to a subthreshold concentration of sulfur dioxide (0.1 ppm) was examined (Koenig et al. 1990). Prior exposure to ozone potentiated subsequent responses to sulfur dioxide in asthmatic subjects.

The effects of exposure to a combination of sulfur dioxide (0.2 ppm) and nitrogen dioxide (0.4 ppm) on the airway response of mild asthmatic patients to allergen inhalation have been investigated (Devalia et al. 1994). Subjects were exposed for 6 hours to sulfur dioxide and nitrogen dioxide alone, or in combination, in exposure chambers. The subjects were then challenged with predetermined concentrations of *Dermatophagoides pteronyssinus* allergen 10 minutes after each exposure. The cumulative breath units of allergen

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**Table 2-5. Symptoms Associated with Concurrent Exposure to Sulfur Dioxide, Smoke, and Particulates<sup>a</sup>**

Sulfur Dioxide Concentration (ppm)	Effects
0.01	Increased respiratory symptoms among the general population and increased frequencies of respiratory illness among children
0.04	Excess mortality among the elderly or the chronically sick
0.04	Worsening of the condition of patients with existing respiratory disease

<sup>a</sup>WHO 1979



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required to produce a 20% fall in forced expiratory volume in 11 second ( $PD_{20} FEV_1$ ) were measured after each exposure. The results showed that only the combination of sulfur dioxide and nitrogen dioxide significantly ( $p=0.005$ ) decreased  $PD_{20} FEV_1$ .

Exposure to sulfur dioxide may enhance ozone absorption in the lung. Absorption of ozone boluses were compared in healthy nonsmokers before and after exposure to 0.36 ppm sulfur dioxide for 2 hours (Rigas et al. 1997). Sulfur dioxide exposure resulted in an increased absorption of ozone in males, but the results were not statistically significant. It was postulated that anatomical variations, such as differences in dead space volume, may have contributed to the sex-specific differences in absorption.

Sulfur dioxide (0.1 ppm) by itself had no statistically significant effect on forced expiratory volume in one second ( $FEV_1$ ) or total respiratory resistance in 9 atopic adolescents (aged 12-18 years) who were exposed by mouth, while exercising for 10 minutes of a 40 minute exposure period (Koenig et al. 1989). However, sulfur dioxide may potentiate sulfuric acid-induced respiratory effects. Exposure to  $0.068 \text{ mg/m}^3$  sulfuric acid (0.6  $\mu\text{m}$  aerosol) by itself resulted in a nonsignificant 12% increase in total respiratory resistance. However, a statistically significant 15% increase in respiratory resistance was observed following exposure to both 0.1 ppm sulfur dioxide and  $0.068 \text{ mg/m}^3$  sulfuric acid.

A synergistic toxic action between sulfur dioxide and sulfuric acid has been observed in studies of bronchoconstriction in guinea pigs (Amdur 1959, 1974). Particle size of the aerosol was found to be an important factor in the potentiation of the response to sulfur dioxide. For instance, a potentiation of the response to sulfur dioxide was noted when the sulfuric acid particle size was 0.8 microns ( $\mu$ ) but not when the particle size was 2.5  $\mu$ .

Sulfite may potentiate effects induced by peroxyxynitrite, a compound commonly found in lungs of individuals with inflammatory diseases such as asthma (Reist et al. 1998). Peroxyxynitrite inactivates  $\alpha_1$ -antitrypsinase, which inhibits lung damaging enzymes such as elastase. In vitro experiments have demonstrated potentiation by sulfite only if its concentration does not exceed that of peroxyxynitrite.

### 2.9 POPULATIONS THAT ARE UNUSUALLY SUSCEPTIBLE

A susceptible population will exhibit a different or enhanced response to sulfur dioxide than will most persons exposed to the same level of sulfur dioxide in the environment. Reasons may include genetic

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makeup, age, health and nutritional status, and exposure to other toxic substances (e.g., cigarette smoke). These parameters may result in reduced detoxification or excretion of sulfur dioxide, or compromised function of target organs affected by sulfur dioxide. Populations who are at greater risk due to their unusually high exposure to sulfur dioxide are discussed in Section 5.7, Populations With Potentially High Exposure.

Exercising asthmatics are recognized as the most susceptible group to sulfur dioxide inhalation (EPA 1994a, 1994b). Some sensitive asthmatics have been shown to respond to sulfur dioxide at a concentration as low as 0.1 ppm (Sheppard et al. 1981). In particular, physically active asthmatics would be at special risk (EPA 1994a, 1994b). In addition, based on data pertaining to the prevalence of asthma and mortality rates from asthma, certain minority group individuals (e.g., African American, Hispanic) may also represent population segments at increased potential risk for sulfur dioxide respiratory effects (EPA 1994a, 1994b) given the higher rates of asthma mortality in nonwhite populations. A sulfite-sensitive subpopulation of asthmatics, who have a relative deficiency of sulfite oxidase, has been postulated to exist (IARC 1992). In contrast, healthy nonasthmatic individuals do not experience respiratory effects at concentrations of up to 1.0 ppm. Nonasthmatic individuals who are unusually sensitive to cold air may also be more susceptible to the respiratory effects of sulfur dioxide.

Elderly adults with preexisting respiratory or cardiovascular disease may be susceptible to the increased risk of mortality associated with acute-duration exposure to sulfur dioxide (WHO 1979). Children may be particularly susceptible to increased frequencies of respiratory illness following chronic-duration exposure to sulfur dioxide (WHO 1979).

Individuals who are deficient in sulfite oxidase may be susceptible to oxidative effects following exposure to sulfur dioxide. *In vitro* studies have demonstrated increased formation of a sulfur trioxide radical intermediate in polymorphonuclear leukocytes of individuals with decreased sulfite oxidase activity (Constantin 1996). Elderly adults and individuals with Down's syndrome may also be susceptible to oxidative stress because limited evidence indicates increased formation of the sulfur trioxide radical regardless of sulfite oxidase activity (Constantin, 1996). Animal studies have demonstrated an inverse relationship between sulfite oxidase activity and susceptibility to intraperitoneally administered bisulfite (Tejnorova 1978).

## **2.10 METHODS FOR REDUCING TOXIC EFFECTS**

This section will describe clinical practice and research concerning methods for reducing toxic effects of exposure to sulfur dioxide. However, because some of the treatments discussed may be experimental and unproven, this section should not be used as a guide for treatment of exposures to sulfur dioxide. When specific exposures have occurred, poison control centers and medical toxicologists should be consulted for medical advice. For specific information about treatment following exposures to sulfur dioxide, the reader should consult the text of Bronstein AC and Currence PL (1994): *Emergency Care for Hazardous Materials Exposure*.

### **2.10.1 Reducing Peak Absorption Following Exposure**

There are no specific methods available to reduce the absorption of sulfur dioxide following exposure. Supportive treatment includes administration of 100% humidified supplemental oxygen with assisted ventilation as required, endotracheal intubation or tracheostomy if upper airway obstruction is present, and the use of inhaled sympathomimetic bronchodilators, such as albuterol, metaproterenol, or cromolyn sodium, for bronchoconstriction (HSDB 1996). For eye contamination, copious irrigation is recommended (HSDB 1998).

### **2.10.2 Reducing Body Burden**

There are no known methods for reducing the body burden of sulfur dioxide. Following absorption, inhaled sulfur dioxide dissolves on the walls of the moist airways to form sulfite and bisulfite, which in turn, can be oxidized to sulfates, a reaction catalyzed by sulfite oxidase (Gunnison et al. 1987). An inverse correlation between sulfite oxidase activity and sensitivity to bisulfite toxicity has been noted (IARC 1992; Tejnorova 1978).

### **2.10.3 Interfering with the Mechanism of Action for Toxic Effects**

Sulfur dioxide-induced increase in airway resistance is due to reflex bronchoconstriction (Frank et al. 1962; Nadel et al. 1965). Injection of atropine prevented the increase in airway resistance in healthy subjects exposed by inhalation to 4-6 ppm sulfur dioxide for 10 minutes. Indomethacin, a prostaglandin synthetase inhibitor, and zafirlukast, a leukotriene receptor antagonist, were demonstrated to reduce airway

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responsiveness in asthmatics challenged with sulfur dioxide (Field et al. 1996; Lazarus et al. 1997). Inhaled sympathomimetic bronchodilators can also be used to reverse bronchospasm. Inhaled corticosteroids may be used to alleviate bronchoconstriction associated with reactive airways dysfunction syndrome (RADS) (Kennedy et al. 1992).

Several techniques have been demonstrated to reduce sulfur dioxide-induced toxicity in animals. Administration of low molecular weight heparin to rats during or after exposure to 30-40 ppm sulfur dioxide for 1 hour/day, 5 days/week, for 12 weeks reduced inflammation of the bronchial mucosa (Krasnowska et al. 1998). Treatment with nebulized S-carboxymethylcysteine, a medication prescribed for sinusitis in the United Kingdom and Japan, restored ciliary function and accelerated mucosal repair in the sinuses of rabbits exposed to 20 ppm sulfur dioxide 4 hours/day for 4 weeks (Sugiura et al. 1997).

Administration of vitamins E and C to 7 rats exposed to 10 ppm sulfur dioxide for 1 hour/day for 45 days reduced oxidative effects such as lipid peroxidation and membrane damage in erythrocytes (Etlík et al. 1997).

Once absorbed by the respiratory tract, sulfur dioxide can form sulfite and bisulfite. The enzyme sulfite oxidase can oxidize sulfite and bisulfite to sulfates. Sulfite and/or bisulfite may be the chemicals that play a role in the sulfur dioxide-induced bronchoconstriction (Sheppard 1988). Interference with the formation of sulfite and/or bisulfite may be a potential strategy, albeit untested, to reduce bronchoconstriction from sulfur dioxide toxicity.

### 2.11 ADEQUACY OF THE DATABASE

Section 104(I)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of sulfur dioxide is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of sulfur dioxide.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that

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all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

### 2.11.1 Existing Information on Health Effects of Sulfur Dioxide

The existing data on health effects of inhalation, oral, and dermal exposure of humans and animals to sulfur dioxide are summarized in Figure 2-3. The purpose of this figure is to illustrate the existing information concerning the health effects of sulfur dioxide. Each dot in the figure indicates that one or more studies provide information associated with that particular effect. The dot does not necessarily imply anything about the quality of the study or studies, nor should missing information in this figure be interpreted as a “data need.” A data need, as defined in ATSDR’s *Decision Guide for Identifying Substance-Specific Data Needs Related to Toxicological Profiles* (ATSDR 1989), is substance-specific information necessary to conduct comprehensive public health assessments. Generally, ATSDR defines a data gap more broadly as any substance-specific information missing from the scientific literature.

There are human data on inhalation exposure to sulfur dioxide that provide information on acute and chronic systemic effects. There are also limited data on genotoxic and carcinogenic effects. Data on potential chronic systemic, genotoxic, and carcinogenic effects are limited because of confounding factors such as multiple exposure. There are no oral or dermal exposure data.

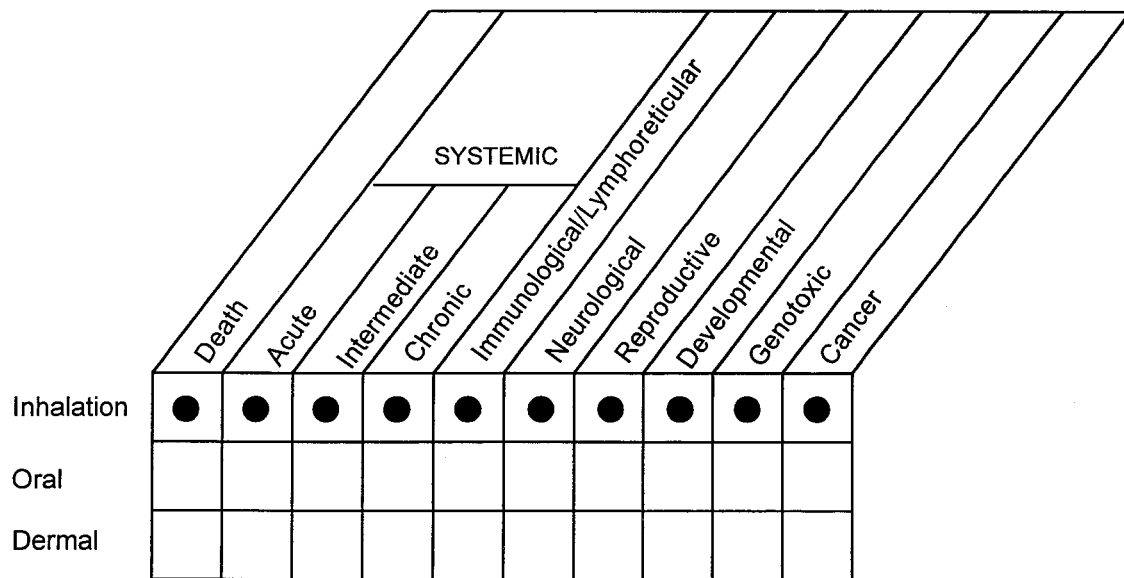
Animal data on inhalation exposure cover primarily acute respiratory effects. Data on other acute-duration effects in animals are limited. There are limited data concerning intermediate- and chronic-duration systemic effects in animals. One chronic study is available that examines potential carcinogenicity of sulfur dioxide in mice. Studies which examine ocular and dermal effects from exposure to the liquid and gaseous form of sulfur dioxide are available. There are no oral exposure data.

### 2.11.2 Identification of Data Needs

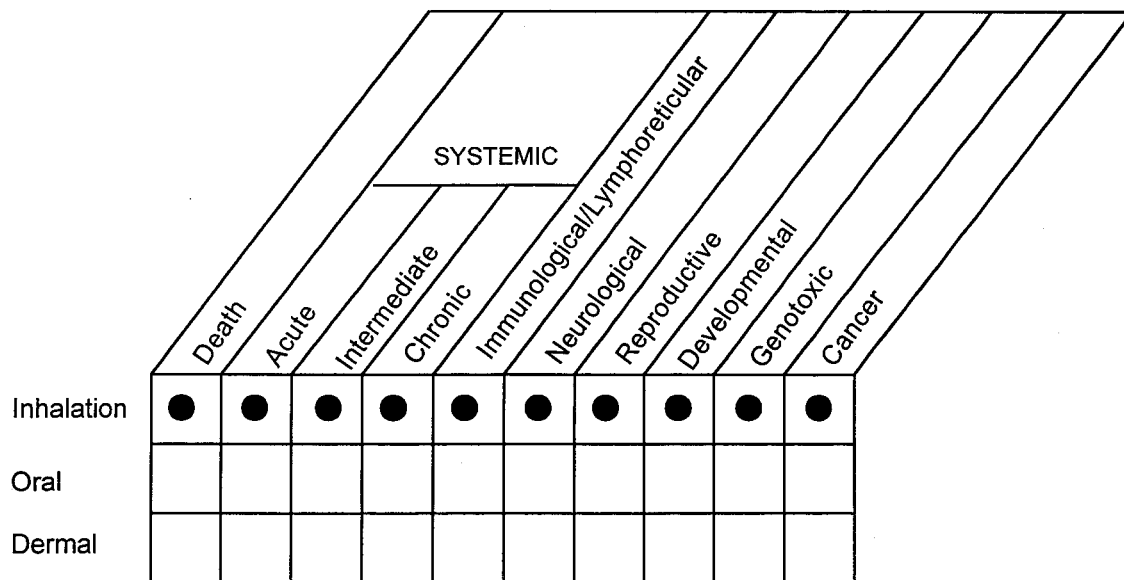
**Acute-Duration Exposure.** There are several case reports of deaths in humans (Atkinson et al. 1993; Charan et al. 1979; Harkonen et al. 1983; Huber and Loving 1991; Rabinovitch et al. 1989). However, estimates of exposure concentrations were not often reported in these studies.

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**FIGURE 2-3. Existing Information on Health Effects of Sulfur Dioxide**



**Human**



**Animal**

● Existing Studies

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There are numerous acute-duration controlled inhalation studies in humans concerning respiratory effects. These studies have established that acute-duration exposure to sulfur dioxide causes constriction of the airways, especially in exercising asthmatics (Bethel et al. 1985; EPA 1994a, 1994b; Horstman et al. 1986; Linn et al. 1983a, 1983b, 1987, 1990; Roger et al. 1985; Sheppard et al. 1980, 1981). Pulmonary function tests in exercising, mild asthmatics indicate that 0.1 ppm sulfur dioxide may be close to the threshold for bronchoconstriction (Sheppard et al. 1981). Based on a minimal LOAEL of 0.1 ppm for increased airway resistance observed in the Sheppard et al. (1981) study, an acute-duration MRL of 0.01 ppm was derived. Studies in experimental animals have supported the pulmonary effects of sulfur dioxide following inhalation exposure. Increased airway resistance and decreased compliance were noted in guinea pigs exposed to 2.6 ppm sulfur dioxide for 1 hour (Amdur 1959).

In recent years, there has been concern about the potential health significance of 5-10 minute exposures to peak levels of sulfur dioxide currently occurring near heavy industrial areas (EPA 1994a, 1994b). Additional studies on the frequency of occurrence of 5-10 minute peak sulfur dioxide levels in the ambient air and the number of asthmatics that may be potentially exposed to such peak levels of sulfur dioxide would be useful in assessing the health effects of exposure to peak levels of sulfur dioxide. In addition, information on the frequency at which asthmatics would be potentially exposed to peak levels of sulfur dioxide would be helpful. Although epidemiological studies on the association between short-term peaks of sulfur dioxide and potential health effects in asthmatics, including exercising asthmatics, would be useful, these kinds of studies are most likely difficult to design and conduct.

The controlled human exposure studies of sulfur dioxide are typically restricted to mild asthmatics. Thus, it is not certain if such studies, although consistent in demonstrating the sensitivity of asthmatics to sulfur dioxide, reflect the characteristics of the asthmatic population as a whole. Individuals with severe asthma may be more susceptible to the effects of sulfur dioxide because of their lower reserve of lung function. However, severe asthmatics may actually be protected against sulfur dioxide effects because they are less prone to strenuous outdoor exercise and more likely to take medication before participating in outdoor activities (EPA 1994b). Additional information on this issue would be useful.

Although oral and dermal data regarding the effects of sulfur dioxide were not identified, human exposure would be expected to be principally by inhalation. Oral and dermal exposure studies may be clinically relevant but are not a high priority.

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**Intermediate-Duration Exposure.** There are no human data on intermediate-duration inhalation exposures to sulfur dioxide. Studies on intermediate-duration exposures to sulfur dioxide would be useful for assessing potential health risks to humans. No intermediate-duration oral or dermal studies of sulfur dioxide were identified. Because inhalation is the primary route of concern, oral and dermal exposure studies should not be a high priority.

Intermediate-duration inhalation exposure studies in animals are very limited. In an intermediate-duration study, mild bronchitic lesions were seen in hamsters exposed to 650 ppm sulfur dioxide for 19-74 days (Goldring et al. 1970). Decreased respiratory rate, rhinitis, tracheitis, and bronchopneumonia were observed in rabbits exposed to 70-300 ppm sulfur dioxide for 6 weeks (Miyata et al. 1990). Inflammation of the bronchial mucosa was observed in rats exposed to 30-40 ppm sulfur dioxide for 1 hour/day, 5 days/weeks, for 12 weeks (Krasnowska et al. 1998). Increased numbers of goblet cells were observed in the airways of rats exposed to 400 ppm sulfur dioxide for 3 hours/day, 5 days/week, for 3 weeks (Basbaum et al. 1990; Lamb and Reid 1968). Nasopharyngitis and lipid peroxidation of lung tissue were observed in guinea pigs exposed to 10 ppm sulfur dioxide for 1 hour/day for 30 days (Haider 1985). Systemic effects have also been noted following inhalation of sulfur dioxide. Lipid peroxidation was observed in erythrocytes of rats exposed to 10 ppm sulfur dioxide for 1 hour/day for 45 days (Etlik et al. 1997) or for 1 hour/day, 7 days/week, for 8 weeks (Gumuslu et al. 1998). A reduction in insulin plasma levels and increase in liver triglyceride levels were observed in rats exposed to 10 ppm sulfur dioxide for 24 hours/day, 7 days/week, for 15 days (Lovati et al. 1996). Studies were inadequate for the development of an MRL. Doses administered in animal studies were well above levels which produce toxic effects in humans. In addition, multiple doses were not administered and a dose-response relationship could not be assessed. Additional well designed animal studies on the respiratory and systemic effects following intermediate-duration exposure, and the relevance of the effects to exposed humans, would be useful.

**Chronic-Duration Exposure and Cancer.** There are limited data on chronic occupational exposures to sulfur dioxide. These studies indicate a potential association between sulfur dioxide exposure and respiratory effects (lung cancer and lung function changes). However, these occupational studies have limitations concerning accurate exposure assessments and concomitant exposures. Epidemiological studies on the relationship between ambient air pollution and lung cancer (Ponka et al. 1993; Tango 1994), although limited, have suggested that the risk of lung cancer from sulfur dioxide exposure in the environment is nonsignificant. Epidemiological studies of children and acute air episodes of sulfur dioxide are confounded by the presence of other air pollutants that are also associated with lung function changes. Additional



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supporting data from well-designed epidemiological studies would be useful. There are no known data on chronic human oral or dermal exposures to sulfur dioxide. Oral and dermal routes may be clinically relevant to humans, but inhalation is the primary route of concern. Therefore, oral and dermal data should not be high priority.

There are only a few limited chronic inhalation studies in two species of animals. No effects on lung function were observed in guinea pigs exposed to 5.72 ppm sulfur dioxide for 22 hours/day, 7 days/week, for 52 weeks (Alarie et al. 1972). Likewise, no lung function changes or histopathological alterations in the lung were observed in monkeys exposed to 5.12 ppm sulfur dioxide for 23.3 hours/day, 7 days/week, for 78 weeks (Alarie et al. 1975). Additional chronic-duration animal studies following inhalation exposure would be useful for providing supporting data regarding potential chronic-duration effects of sulfur dioxide. The chronic-duration studies are too limited to be used for developing a chronic-duration inhalation MRL; because only one dose was administered, a dose response relationship could not be assessed.

Several epidemiological studies were conducted to address cancer associated with occupational or environmental exposure to sulfur dioxide pollution, but definitive conclusions could not be drawn because of confounding factors (Bond et al. 1986; Enterline et al. 1987; Lubin et al. 1981; Ponka et al. 1993; Tango et al. 1994; Welch et al. 1982). A chronic cancer study in mice was also limited due to small sample size and the administration of only one dose. Well-designed studies which evaluate carcinogenicity potential following inhalation exposure in additional species are needed. Oral and dermal exposure studies are not of high priority because inhalation is the primary route of concern

**Genotoxicity.** *In vivo* clastogenic effects of sulfur dioxide in humans have been reported. Increases in chromosome aberrations and sister chromatid exchanges were detected in lymphocytes from 42 workers who were occupationally exposed to an average concentration of 41.7 mg/m<sup>3</sup> (15.9 ppm) of sulfur dioxide, a level which is eight times greater than the TLV (Yadav and Kaushik 1996). Similar findings have been observed in other studies of workers (Meng and Zhang 1990a; Nordenson et al. 1980). In addition, increases in the frequencies of lymphocytes with micronuclei were noted (Meng and Zhang 1990b). One study of potential chromosomal abnormalities in workers exposed to 8-hour workshift levels of about 1 ppm (2.62 mg/m<sup>3</sup>) sulfur dioxide in the aluminum industry revealed that sulfur dioxide was without effect (Sorsa et al. 1982). Studies which examine genotoxic effects in animals following inhalation exposure were not located but would be useful. Studies with animal germ cells have indicated that genotoxicity is possible following *in vitro* exposure but unlikely from intraperitoneal administration of sulfur dioxide metabolites (Jagiello et al. 1975;

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Russell and Kelly 1975). Further human and animal studies to explore possible dose-response relationships and to provide mechanistic data would be useful.

**Reproductive Toxicity.** In a cross-sectional study of spontaneous abortions in an industrial community in Finland, no evidence was found that would associate sulfur dioxide exposure with a risk of spontaneous abortions (Hemminki and Niemi 1982). However, abnormalities in sperm have been associated with sulfur dioxide, as a surrogate of all air pollutants, in a limited study conducted in the Czech Republic (Selevan et al. 1995).

Reproductive effects were not observed in rats exposed to 5-30 ppm sulfur dioxide for 9 days prior to mating until 12-14 days of pregnancy (Petruzzi et al. 1996) in mice exposed to 25 ppm sulfur dioxide 7 hours/day on gestation days 6-15 or in rabbits exposed to 70 ppm sulfur dioxide 7 hours/day on gestation days 6-18 (Murray et al. 1979). The studies are limited because the reproductive function of the offspring were not assessed. A well-designed multigeneration reproductive study would be useful to assess the potential reproductive toxicity of sulfur dioxide. Because inhalation is the primary route of concern, oral and dermal exposure studies should not be a high priority.

**Developmental Toxicity.** There is evidence of an association between environmental exposure to sulfur dioxide during pregnancy and reduced birth weight. Like other epidemiological studies of air pollution mixtures, interpretation of the results is complicated by confounding factors such as other air pollutants and a lack of personal exposure information. Though additional, well-designed epidemiological studies would be useful, separation of effects associated with individual components of an air pollution mixture is unlikely considering the currently available technology.

Numerous developmental studies have been conducted in animals. One study reported no developmental effects in the offspring of pregnant mice that were exposed to 32-250 ppm sulfur dioxide on gestation days 7-17 (Singh 1982). In another developmental study in mice in which pregnant females were exposed to 32 ppm or 65 ppm on gestation days 7-18, increased time for righting reflex on postnatal day 1 and increased negative geotaxis on postnatal day 10 were reported (Singh 1989). The duration of exposure for each day was not stated. No visible signs of maternal toxicity and no effect on the number of live births were observed. Sulfur dioxide at a concentration of 65 ppm significantly decreased the birth weight (about 89% of controls) of the pups. Minor skeletal variations were reported in offspring of mice exposed to 25 ppm sulfur dioxide 7 hours/day on gestation days 6-15 and rabbits exposed to 70 ppm sulfur dioxide 7 hours/day on

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gestation days 6-18 (Murray et al. 1979). Fetal body weights were also lower in exposed mice. Reduced food intake was the only sign of maternal toxicity. Neurological effects were not observed in the offspring of mice exposed to 5-30 ppm sulfur dioxide for 9 days prior to mating until 12-14 days of pregnancy (Petruzzi et al. 1996). Studies which examine developmental effects following oral or dermal exposure should not be high priority since inhalation is the primary route of concern.

**Immunotoxicity.** Epidemiological studies have indicated that sulfur dioxide air pollution may increase the prevalence of allergies (Soyseth et al. 1996) and incidence of respiratory infections (French et al. 1973) in children. However, both studies were limited by inadequate control of confounding factors and a lack of personal exposure data. Increased sensitization to antigen was reported in a study of guinea pigs exposed by inhalation to sulfur dioxide (Riedel et al. 1992). In an acute-duration inhalation study with hamsters, there was a significant reduction in endocytosis by pulmonary macrophage following exposure to 50 ppm sulfur dioxide for 4 hours while exercising (Skornik and Brain 1990). However, pulmonary defense mechanisms were not affected in rats and mice exposed to 0.32-0.43 ppm sulfur dioxide together with 87-113  $\mu\text{g}/\text{m}^3$  sulfate for 4 hours prior to or 17 hours following infection with *Staphylococcus aureus* or Group C *Streptococci* (Goldstein et al. 1979). Additional studies to determine if inhalation of sulfur dioxide increases susceptibility to infection or allergen sensitization in humans or animals would be useful. Because inhalation is the primary route of concern, studies of oral or dermal exposure should not be high priority.

**Neurotoxicity.** One study has indicated that reflex response times may have been affected in workers exposed to sulfur dioxide, and was most likely due to a psychological response to systemic toxicity (Kehoe et al. 1932). Seizures and prostration were observed in rats following exposure to 2,350,50,000 and 500,000 ppm sulfur dioxide for an average of 176 minutes, <10 minutes, and <2 minutes respectively (Cohen 1973). Lipid peroxidation has been observed in the brains of guinea pigs exposed to sulfur dioxide for 1 hour/day for 30 days (Haider et al. 1982). Additional neurotoxicity studies would be useful to characterize the effects of sulfur dioxide on the nervous system.

**Epidemiological and Human Dosimetry Studies.** There are several epidemiological studies that have examined the potential association between respiratory effects (i.e., lung cancer and lung function changes) and inhalation exposure to sulfur dioxide (Agocs et al. 1997; Archer and Gillan 1978; Buchdahl et al. 1996; Dockery et al. 1982; Dodge et al. 1985; Lebowitz et al. 1979; Lowe et al. 1970; Shy et al. 1973; Skalpe 1964; Smith et al. 1977). There are also epidemiological studies that have examined the association between daily acute mortality and sulfur dioxide exposure (Anderson et al. 1996; Bacharova, 1996; Ballester

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et al. 1996; Katsouyanni et al. 1997; Loomis et al. 1996; Mazumdar et al. 1982; Moolgavkar et al. 1995a, 1995b; Spix, 1996; Sunyer et al. 1996; Thurston et al. 1989; Touloumi et al. 1996; Verhoeff et al. 1996). Reproductive effects have also been examined in epidemiological studies (Selevan et al. 1995; Wang et al. 1997). However, these epidemiological studies are limited by confounders. Further studies that employ more precise measurements of exposure, control of exposure to other chemicals, and follow-up of occupational cohorts would be useful. Monitoring of populations around industrial areas where there are exposures to peak levels of sulfur dioxide would also be useful.

**Biomarkers of Exposure and Effect**

**Exposure.** No specific biomarkers of exposure for sulfur dioxide have been identified. Potential biomarkers include plasma *S*-sulfonate levels and urinary levels of sulfate (Balchum et al. 1960a; Frank et al. 1967; Kleinman 1984; Speizer and Frank 1966; Yokoyama et al. 1971). Further studies examining the suitability of these potential biomarkers would be useful.

**Effect.** No specific biomarkers of effect for sulfur dioxide have been identified. Potential nonspecific biomarkers include lung function changes (EPA 1994a, 1994b; Sheppard et al. 1981) and clastogenic effects (Meng and Xhang 1990a, 1990b; Yadav and Kaushik 1996), leukocytes in bronchioalveolar lavage fluid (Sandstrom 1989a, 1989b), or adduct formation in nasal mucosa (Topinka et al. 1995). Further studies examining the suitability of these potential biomarkers, and identification of potential specific biomarkers, would be useful.

**Absorption, Distribution, Metabolism, and Excretion.** Sulfur dioxide, a highly water-soluble gas, is rapidly absorbed by the mucosa of the nose and upper respiratory tract (Kleinman 1984; Speizer and Frank 1966). Absorption in the lower respiratory tract is increased with enhanced ventilation associated with a transition from nasal to oronasal breathing (EPA 1986d). Absorbed sulfur dioxide metabolites are taken up by the blood and are readily distributed throughout the body (Balchum et al. 1960a; Frank et al. 1967; Yokoyama et al. 1971). Once absorbed, sulfur dioxide is rapidly metabolized to sulfates in the liver by the enzyme, sulfite oxidase (Gunnison et al. 1987) or through the generation of sulfur trioxide radical intermediate (Constantin et al. 1996). Sulfur dioxide may also react with proteins to form *S*-sulfonate (Gunnison and Palmes 1974). Sulfur dioxide is excreted primarily in the urine as sulfate. There is also evidence that glutathione is involved in the detoxification of sulfur dioxide (Kagadel et al. 1986; Langley-Evans et al. 1996). Studies which compare the quantitative distribution of sulfur dioxide within the

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respiratory tract during nasal, oronasal, and oral breathing would be useful. Studies which examine the relationship between sulfite oxidase activity and levels of sulfur dioxide metabolites in blood would also be helpful. Identification of additional factors involved in the metabolism of sulfites would also be useful information.

**Comparative Toxicokinetics.** There are no studies that directly compare the toxicokinetics across species. PBPK models have not been developed. Sulfite oxidase activity varies among species. Therefore, studies providing quantitative data necessary to develop PBPK models would be useful.

**Methods for Reducing Toxic Effects.** Other than removing the subject from exposure, there is no specific method to reduce the absorption of sulfur dioxide. There are no known methods for specifically reducing the body burden of sulfur dioxide. Supportive treatment includes administration of 100% humidified supplemental oxygen with assisted ventilation as required, endotracheal intubation or tracheostomy if upper airway obstruction is present, and the use of inhaled sympathomimetic bronchodilators for bronchoconstriction (HSDB 1998). Studies examining methods to enhance the oxidation of sulfur dioxide to increase elimination might be useful. Studies to determine effects of antioxidant therapy in humans may also be useful.

**Children's Susceptibility.** Clinical studies have indicated that compared to healthy senior citizens, healthy adolescents are not at increased risk of respiratory effects following inhalation of sulfur dioxide (Koenig et al. 1982b; Rondinelli et al. 1987). Epidemiological studies indicate associations between sulfur dioxide pollution and respiratory symptoms (Buchdahl et al. 1996; Dodge et al. 1985; WHO 1979) and transient effects on respiratory function in children (Dockery et al. 1982; Shy et al. 1973). However, such studies are limited due to confounding effects of other air pollutants. Asthmatics in general are most susceptible to sulfur dioxide exposure, but it is not known if asthmatic children are more sensitive than asthmatic adults (EPA 1994a, 1994b). It is unlikely that additional epidemiological studies would provide conclusive information. However, additional controlled studies in asthmatic and healthy children would be useful in determining doses which produce effects, and whether these children are more susceptible to sulfur dioxide-induced respiratory effects than asthmatic and healthy adults. Controlled studies in young versus mature animals would also be useful.

Studies in humans and animals indicate that serious developmental effects are not likely from maternal inhalation of sulfur dioxide. However, developmental delays, such as reduced birth weight in humans and

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decreased fetal weight and delayed ossification in animals, have been observed (Murray et al. 1977, 1979; Wang 1997). Data requirements for developmental effects are discussed under the Developmental Toxicity heading of this section (2.11.2 Identification of Data Needs).

Metabolic studies have demonstrated reduced liver sulfite oxidase activity in young rats (Cohen 1974). Comparisons of sulfite oxidase activity in infants, children, and adults and also in young and mature animals would be useful. Measurement of liver sulfite oxidase activity in young versus mature animals of other species would also be useful. Studies in which liver sulfite oxidase activity in young and mature animals is compared in relation to blood levels of sulfates, sulfites, and *S*-sulfonates following sulfur dioxide inhalation would also be useful. Pharmacokinetic studies in dogs have demonstrated that following inhalation of radio-labeled sulfur dioxide, a small amount of radio-label reaches the ovaries and testes (Balchum 1959). Experiments to identify the sulfur dioxide metabolite present in gonads would be useful. Studies which examine pharmacokinetics in immature or pregnant animals were not identified. Studies which examine the distribution of sulfur dioxide metabolites in pregnant animals to determine if they cross the placenta or are transferred to breast milk might be useful. Accumulation in maternal tissues is not expected for sulfur dioxide or its metabolites due to its high water solubility. Studies which examine absorption, metabolism, distribution, and excretion in immature animals would also be helpful in evaluating the impact of sulfur dioxide exposure in children.

Studies of genotoxicity in germ cells have demonstrated that mutations and clastogenic effects occur with *in vitro* exposure but not *in vivo* exposure (Iagiello et. al. 1975; Russell and Kelly 1975; Shapiro et al. 1977). Although genotoxicity has been observed in lymphocytes of workers exposed to sulfur dioxide (Meng and Zhang 1990a, 1990b; Yadav and Kaushik 1996), available data in animals indicates that genotoxicity in germ cells is unlikely from exposure to sulfur dioxide metabolites (Russell and Kelly 1975). Inhalation studies in animals could be done to verify that genotoxicity in germ cells does not occur following inhalation exposure to sulfur dioxide.

Child health data needs relating to exposure are discussed in 5.8.1 Data Needs: Exposures of Children.

### 2.11.3 Ongoing Studies

Ongoing animal, occupational, or epidemiological studies of the health effects of sulfur dioxide were not identified.