

Effects of acute leg ischemia during cycling on oxygen and carbon dioxide stores

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Abstract—This study estimated changes in whole body oxygen stores (O_2s) and carbon dioxide stores (CO_2s) during steady state exercise with leg ischemia induced by leg cuff inflation. Six physically fit subjects performed 75 W steady state exercise for 15 min on a cycle ergometer. After 5 min of exercise, cuffs on the upper and lower legs were inflated to 140 mmHg. Cuffs were deflated after 5 min and exercise continued for another 5 min. O_2 uptake ($\dot{V}O_2$) and CO_2 output ($\dot{V}CO_2$) significantly increased during the first 30 s after inflation, significantly decreased between 60 and 90 s, and then rose linearly until deflation. $\dot{V}O_2$ and $\dot{V}CO_2$ significantly increased further after cuff deflation, peaking between 30 and 60 s and then returned to near baseline exercise levels. Model-estimated changes in total O_2s and CO_2s were compared with time-integrated store changes from $\dot{V}O_2$ and $\dot{V}CO_2$. During 5 min after cuff deflation, $\dot{V}O_2$ and $\dot{V}CO_2$ exceeded the model-estimated change in stores by 273 and 697 mL, respectively. These results reflect the O_2 cost repayment of the anaerobic component and lactate buffering to neutralize circulating metabolites caused by the preceding ischemia.

Key words: anaerobic exercise, bicarbonate buffering, carbon dioxide stores, ergoreflex, ischemia, lactate, oxygen deficit, oxygen stores, rehabilitation, ventilation/perfusion ratio, ventilation response.

INTRODUCTION

Progressive physical deconditioning is common in patients with chronic diseases, such as congestive heart failure and chronic obstructive pulmonary disease. One

limitation these patients face is an inability to exercise with sufficient intensity to provide adequate training stimuli. However, regional training of muscles without taxing the central circulation can improve whole-body exercise capacity in these patients [1]. An unusual potential tool to facilitate regional muscle rehabilitation is exercise training during reduced limb blood flow [2–3]. Such “ischemic

Abbreviations: ADS = anatomical dead space (mL), BE = base excess (measure of whole blood buffer base [mmol/L]), CO_2 = carbon dioxide, CO_2s = CO_2 stores (mL), f = breathing frequency (breaths/min), FIO_2 = fraction of inspired oxygen, H^+ = hydrogen ion concentration (nmol/L), Hb = hemoglobin concentration (g%), HCO_3^- = bicarbonate concentration (mmol/L), O_2 = oxygen, O_2s = O_2 stores (mL), $PACO_2$ = partial pressure of alveolar CO_2 (mmHg), PAO_2 = partial pressure of alveolar O_2 (mmHg), P_B = barometric pressure, PCO_2 = partial pressure of CO_2 (mmHg), pHa = arterial pH, PO_2 = partial pressure of O_2 (mmHg), \dot{Q} = cardiac output (L/min), RER = respiratory exchange ratio (CO_2 output/ O_2 uptake), \dot{V}_A = alveolar ventilation ([L/min] body temperature, ambient pressure, saturated), $\dot{V}CO_2$ = CO_2 output ([mL/min] standard temperature and pressure, dry), \dot{V}_E = pulmonary ventilation ([L/min] body temperature, ambient pressure, saturated), $\dot{V}O_2$ = O_2 uptake ([mL/min] standard temperature and pressure, dry), $\dot{V}O_{2max}$ = maximal $\dot{V}O_2$.

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limb training” with limb pressure cuffs has improved limb strength and exercise endurance in physically fit subjects [4–5], diminished postoperative disuse atrophy of knee extensors [6], and induced favorable biochemical and structural changes in muscles [7–8]. Ischemic limb training with low-intensity exercise in patients with congestive heart failure has also reduced exertional dyspnea [9]. We recently demonstrated that leg-extension exercise endurance was enhanced with a 6-week training program of very light leg-extension exercise with ischemia induced by thigh cuff inflation [10].

Superimposing ischemia on exercising limbs provokes the muscle metaboreflex, whereby pulmonary ventilation (\dot{V}_E) and systemic blood pressure are elevated by a chemoreflex stimulated by buildup of metabolic by-products in the ischemic limbs; the most likely candidate is hydrogen ion concentration (H^+) [11]. The oxygen (O_2) stores (O_{2s}) and carbon dioxide (CO_2) stores (CO_{2s}) in the region where blood flow is occluded, as well as in the whole body, will be affected during this ischemia and after circulation is restored as a result of ventilatory, blood flow, and biochemical perturbations. The magnitude and time course of these gas store changes will affect regional and whole-body acid-base status, will cause secondary ventilatory and gas exchange fluctuations during and after exercise, and may induce transient hypoxemia and hypercapnia, such as noted following passive changes in posture [12].

Although rapid transient changes in O_{2s} and CO_{2s} during exercise workload transitions have been studied and quantified [13], gas store changes induced by limb ischemia have received little attention. Specifically, the quantitative relationship is not well defined between O_2 repayment and CO_2 elimination after exercise requiring energy partially derived from anaerobic sources [14] and these measurements with the anaerobic component artificially superimposed have not been reported. Therefore, this study was an initial attempt to estimate the time course and magnitude of changes in O_{2s} and CO_{2s} during and after acute, temporary ischemia of the legs applied by cuff inflation during steady state exercise on a cycle ergometer.

METHODS

Subjects

Five men and one woman volunteered as subjects. Informed consent was obtained from each person, as

approved by the University of New Mexico Human Research Review Committee. All were physically fit and regularly taking part in physical recreation and fitness activities, including jogging and cycling. Their ages ranged from 24 to 62 yr, with a mean body weight and body mass index of 82.5 kg and 25.0 kg/m², respectively. Their maximal O_2 uptake ($\dot{V}O_{2max}$) averaged 48 mL·min⁻¹·kg⁻¹ (range: 42–56). The O_2 uptake ($\dot{V}O_2$) during exercise before ischemia (baseline) averaged 35.7 percent (range: 30%–42%, standard error of the mean = 1.7%) of the subjects' $\dot{V}O_{2max}$. This percentage was not related to age ($r = -0.22$).

Ergometer Exercise and Inflation Cuffs

We placed cuffs on each upper thigh (SC-17, Hokanson Co; Bellevue, Washington) and each lower leg (SC-22) using adhesive tape to keep them in position during exercise. Lower leg cuffs were used to minimize trapping of blood and to enhance ischemia of the calf muscles. Cuffs were inflated to 140 mmHg during exercise. This cuff pressure, slightly exceeding systolic pressure, was chosen after preliminary trials indicated that discomfort at this pressure could be tolerated and gas exchange transients stabilized in about 5 min at the chosen workload. Although the blood pressure response of each subject to the inflation pressure varied, we maintained the pressure at the same level for all to reduce variations in blood “pooling” and thereby reduce variability in the measured responses. Resting measurements were made for 5 min while subjects sat on the ergometer before and after exercise. Subjects cycled for 15 min at 75 W on an electrically load-controlled Bosch ergometer (model ERG 551; Munich, Germany) at 50 to 60 rpm. After 5 min, the four cuffs were simultaneously inflated over a ≈ 10 s period from a gas cylinder pressure source. Cuff pressure was maintained for 5 min and then deflated rapidly in 3 s, with exercise continuing for another 5 min.

Measurements and Calculations

We measured gas exchange at the mouth while subjects sat on the ergometer at rest, during exercise, and at rest after exercise, using a TrueMax 2400 breath-by-breath automated system (Parvomedics, Inc; Sandy, Utah) with incorporated software and model 2700 Rudolph breathing valve and mouthpiece (Hans Rudolph, Inc; Shawnee, Kansas). The measurements included $\dot{V}O_2$, CO_2 output ($\dot{V}CO_2$), \dot{V}_E , calculated respiratory exchange ratio (RER), and $\dot{V}_E/\dot{V}CO_2$ as an index of ventilatory drive. Alveolar ventilation (\dot{V}_A) was calculated from anatomical

dead space (ADS) taken as apparatus dead space + milliliter = body weight in pounds [15] and breathing frequency (f) as $\dot{V}_A = \dot{V}_E - f \times \text{ADS}$. Experiments were conducted at an average barometric pressure (P_B) of 631 mmHg (range: 630–635 mmHg) and ambient fraction of inspired O₂ (FIO₂) of 0.2094. Partial pressure of CO₂ (PCO₂) in alveoli (PACO₂) and partial pressure of O₂ (PO₂) in alveoli (PAO₂) were calculated from alveolar gas equations [16]:

$$\text{PACO}_2 = (\dot{V}\text{CO}_2 \times 0.863) / \dot{V}_A \quad (1)$$

and

$$\text{PAO}_2 = (P_B - 47.1) \text{FIO}_2 - \text{PACO}_2 [\text{FIO}_2 + (1 - \text{FIO}_2) / \text{RER}]. \quad (2)$$

We averaged breath-by-breath measurements continuously over 30 s intervals for each subject throughout exercise and the pre- and postexercise rest periods. We then averaged these values for the six subjects to obtain representative temporal patterns for analysis.

Average changes in O₂s and CO₂s were calculated from differences between measured and predicted gas exchange time courses integrated over time. We based predicted values on baseline gas exchange measurements during the 5th min, assuming these represented steady state values required by the workload, and an increase during ischemia based on assumptions given in the subsequent section for predicted gas exchange. An increase in O₂s was indicated when measured $\dot{V}\text{O}_2$ is greater than predicted $\dot{V}\text{O}_2$ over time, and a decrease in CO₂s was indicated when measured $\dot{V}\text{CO}_2$ is greater than predicted $\dot{V}\text{CO}_2$ and vice versa. Differences in these gas store changes during and after blood flow restriction were attributed to the ischemia. In addition, we obtained total body gas stores present during baseline, 5th min during cuff inflation, and 5th min after cuff deflation from a model using gas exchange, blood flow, and blood volume values. We also used differences between these modeled total store values and the time-integrated measured values of changes in O₂s and CO₂s to extract effects of leg ischemia.

Predicted Gas Exchange

During cuff inflation, we assumed the predicted time course for $\dot{V}\text{O}_2$ would increase linearly during the 6th through 10th min from the steady state exercise value at 5 min because of—

1. A gradual loss of mechanical efficiency by increasing recruitment of ancillary muscles of the hip, torso, and arms to maintain leg work as fatigue increased.

2. Increased O₂ cost of ventilation stimulated by the metaboreflex, which may account for as much as one-third of the observed $\dot{V}\text{O}_2$ rise [17–18].
3. The partial restoration of curtailed leg circulation by the reflex rise in blood pressure that would enhance O₂ delivery to the legs despite restricted blood flow during cuff inflation.
4. The subjects' subjective reports that the last minute of exercise seemed less stressful than the previous minutes, indicating that the anaerobic component of the energy supply had stabilized.

During the 5 min following cuff deflation, $\dot{V}\text{O}_2$ was assumed to decline exponentially to the baseline exercise value by 15 min because the factors just listed were removed by cuff deflation and the elevated $\dot{V}\text{O}_2$ was expected to return similarly to that following the removal of an additional acute exercise workload. The predicted $\dot{V}\text{CO}_2$ was similarly assumed to increase linearly from baseline to 10 min, but to a value calculated as measured $\dot{V}\text{O}_2 \times$ measured baseline RER before cuff inflation (for correcting the elevated $\dot{V}\text{CO}_2$ from the increase in \dot{V}_E resulting from the metaboreflex), and then decline exponentially to the baseline value by 15 min.

Total Gas Stores Model with Blood Flow and Volume Redistribution

Computations and assumptions are shown in the following list for compartmental and total whole body O₂s and CO₂s during exercise at three exercise conditions A, B, and C: A = baseline, 5th min before cuff inflation; B = 5th min of cuff inflation; and C = 5th min after cuff deflation. Arterial and mixed venous blood O₂ and CO₂ contents and mixed venous PO₂ and PCO₂ were calculated from a computer model integrating gas exchange and blood flow values [19–20].

- Blood volume.
 - Total = 71.5 mL/kg body weight = 5,900 mL.
 - Venous compartment for exercise conditions A and C = total \times 0.8 = 4,720 mL.
 - Arterial compartment for exercise conditions A and C = total \times 0.2 = 1,180 mL.
 - During condition B, a 300 mL blood volume shift from the venous to arterial compartment was predicted based on transient increases in measured $\dot{V}\text{O}_2$ and a $\dot{V}\text{CO}_2$ from 30 to 60 s after cuff deflation.
- Lung: O₂ and CO₂ were calculated from PAO₂ and PACO₂ and an assumed functional residual capacity of 4.0 L.

- Arterial O₂: Content based on Hb (hemoglobin concentration) = 15 g%, arterial PO₂ = PAO₂, saturation = standard dissociation curve [21] at pH_a (arterial pH, the negative log of H⁺ in arterial blood) calculated to maintain whole blood base excess (BE) equal to baseline [22], where a pH_a value of 7.420 was assumed.
- Venous O₂: Content from Fick equation with arterial content and measured $\dot{V}O_2$ at exercise conditions A, B, and C and cardiac output (\dot{Q}) = 15 L/min at conditions A and C, with 1 L/min reduction during condition B, based on observations during cuff-induced ischemia by Asmussen and Nielsen [23].
- Tissue O₂.
 - PO₂ from venous content and saturation from standard curve.
 - PO₂ × body weight (82.5 kg) × 0.64 × 0.024 [24].
- Arterial CO₂.
 - Content based on arterial PCO₂ = PACO₂.
 - Content from CO₂ dissociation curve at Hb and pH_a [25].
- Venous CO₂: Content from Fick equation with arterial CO₂ content and measured $\dot{V}CO_2$ and predicted \dot{Q} at exercise conditions A, B, and C.
- Tissue CO₂.
 - PCO₂ for venous content from CO₂ dissociation curve.
 - PCO₂ × body weight × 1.02.

We obtained half-times for rest-to-exercise (“on”) responses and (“off”) transitions from exponential fits to the 10 measured breath-by-breath intervals. We used paired *t*-tests to determine significance (*p* < 0.05) of selected individual transient changes over time and used least squares linear regressions to estimate the significance of relationships between selected variables.

RESULTS

The average $\dot{V}O_2$ and $\dot{V}CO_2$ measurements during rest, exercise, and postexercise rest are shown in **Figure 1**. A plateau for both measurements was reached after ≈3 min of exercise, because the 5th min values were not significantly above the 3 min values (*p* > 0.13). Transient changes induced by ischemia and cuff deflation appeared to have stabilized by the end of exercise. The baseline mechanical efficiency at 75 W for a $\dot{V}O_2$ of 1,410 mL/min (minus the resting $\dot{V}O_2$ of 335 mL/min) was 20.0 percent,

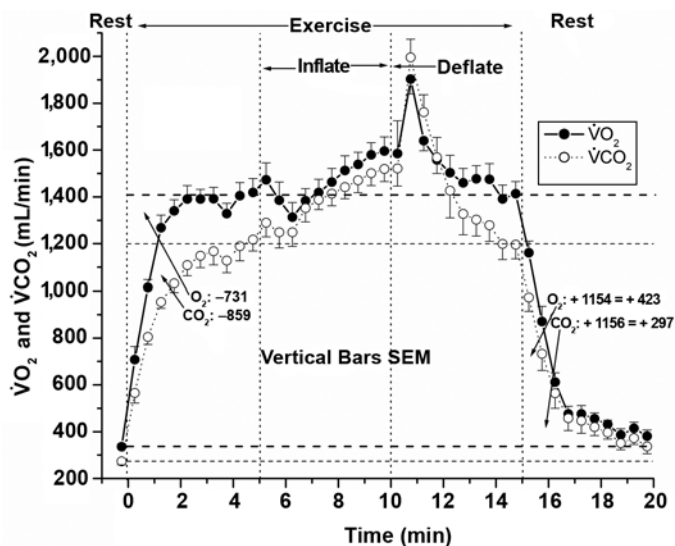


Figure 1. $\dot{V}O_2$ and $\dot{V}CO_2$ during rest, 15 min of cycle ergometer exercise at 75 W, and 5 min postexercise rest. Each point is an average of 30 s values for six subjects. Values (milliliters) are shown for total 5 min exercise onset deficit and 5 min postexercise rest and sum for $\dot{V}O_2$ and $\dot{V}CO_2$. Postexercise excess is significantly greater than preexercise deficit for O₂ (*p* = 0.003) and CO₂ (*p* = 0.031). CO₂ = carbon dioxide, O₂ = oxygen, SEM = standard error of the mean, $\dot{V}CO_2$ = CO₂ output, $\dot{V}O_2$ = O₂ uptake.

decreasing to 17.1 percent at 1,595 mL/min by the end of inflation. During the 5 min postexercise rest period, the total excess $\dot{V}O_2$ and $\dot{V}CO_2$ were both significantly larger than the 5 min $\dot{V}O_2$ deficits following exercise onset. The averages of the corresponding changes in gas stores calculated from time-integrated values for measured and predicted $\dot{V}O_2$ and $\dot{V}CO_2$ are detailed in **Figure 2**.

Oxygen

Measured $\dot{V}O_2$ increased significantly during the first 30 s after cuffs were inflated (*p* = 0.042) and then declined transiently, but significantly, at 6.5 min by 72 mL/min (*p* = 0.049). $\dot{V}O_2$ then rose steadily until cuffs were deflated. The O₂s cumulative loss over 5 min of cuff inflation was 227 mL (**Figure 2**). $\dot{V}O_2$ peaked 45 s after cuff deflation, being 150 mL above adjacent measurements (*p* = 0.001). The 5 min postdeflation exercise $\dot{V}O_2$ excess indicated that O₂s increased by 518 mL.

Carbon Dioxide

Measured $\dot{V}CO_2$ during ischemia is related to similar circulatory and biochemical events affecting $\dot{V}O_2$ but is partially overridden by the large increase in \dot{V}_E (**Figure 3**),

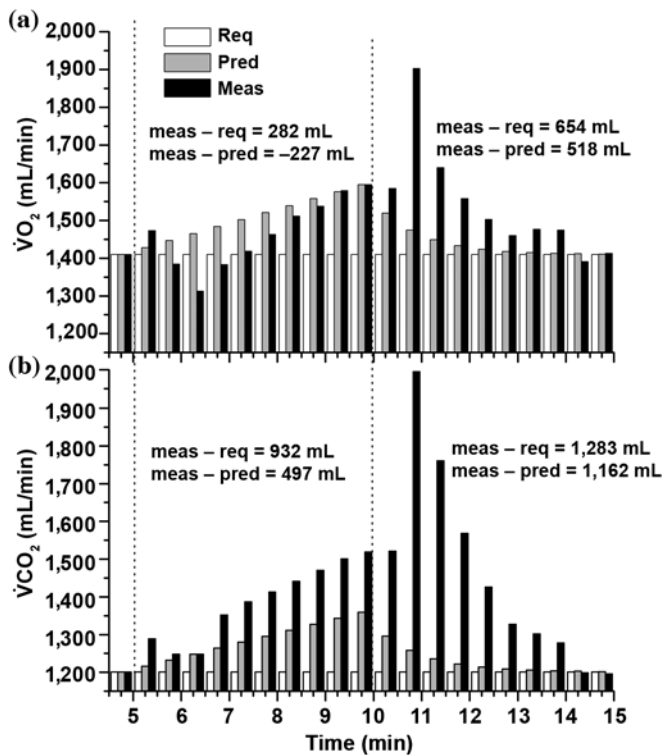


Figure 2.

Changes in gas stores represented by differences between measured (meas) (Figure 1) and predicted (pred) time course for (a) oxygen uptake ($\dot{V}O_2$) and (b) carbon dioxide output ($\dot{V}CO_2$) during 5 min of cuff inflation and 5 min after cuff deflation at 10 min. Values averaged for 30 s. Predicted time course for $\dot{V}O_2$ and $\dot{V}CO_2$ is described in main text. Values for changes in stores (milliliters) are indicated for time-integrated totals over 5 min of inflation and 5 min after cuff deflation. req = required.

because of the metaboreflex stimulation by leg ischemia. CO₂s decreased by 497 mL by the end of the 5 min inflation, as indicated in Figure 2. Similar to $\dot{V}O_2$, the $\dot{V}CO_2$ peaked 45 s after cuff deflation, indicating an additional 180 mL loss in CO₂s above the adjacent measurements ($p = 0.002$), corresponding to the 150 mL of O₂ taken up. The loss in CO₂s over 5 min after cuff deflation was 1,162 mL, about double that of the O₂s gain (518 mL). Over the 10 min of exercise during cuff inflation and deflation, the total O₂s gain was $-227 + 518 = 291$ mL and the total CO₂s loss was $497 + 1,162 = 1,659$ mL.

Ventilation

After exercise termination, the off-responses for $\dot{V}CO_2$ and \dot{V}_E (Figure 3) were similar to each other and their on-responses (36–39 s) but slower than the on-response for $\dot{V}O_2$. $\dot{V}O_2$ and $\dot{V}CO_2$ were slightly above baseline at the

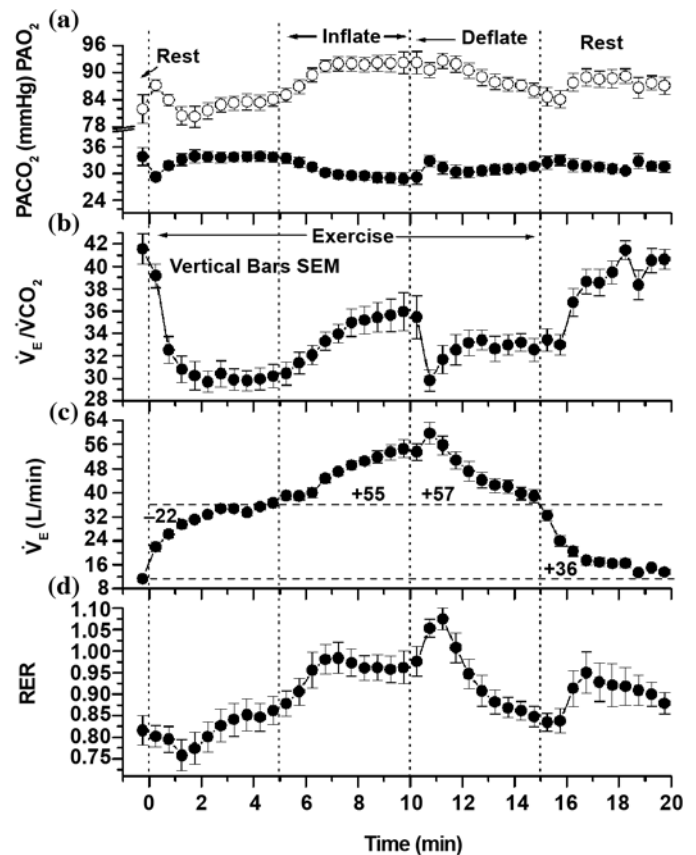


Figure 3.

Average values for six subjects for (a) alveolar gases; (b) $\dot{V}_E/\dot{V}CO_2$; (c) \dot{V}_E ; and (d) RER during rest, 15 min of exercise at 75 W, and 5 min postexercise rest. Values (liters) are shown for cumulative time-integrated \dot{V}_E deficit following exercise onset, cumulative excess during 5 min of cuff inflation, 5 min after cuff deflation, and 5 min of postexercise rest. Latter value is significantly greater than deficit at beginning of exercise. PACO₂ = partial pressure of alveolar carbon dioxide, PAO₂ = partial pressure of alveolar oxygen, RER = respiratory exchange ratio, SEM = standard error of the mean, $\dot{V}CO_2$ = carbon dioxide output, \dot{V}_E = pulmonary ventilation.

end of the 5 min postexercise rest period (Figure 1). The RER was significantly higher during the 5th min postexercise rest compared with the preexercise rest because $\dot{V}CO_2$ was significantly higher (30%) than $\dot{V}O_2$ (18%), indicating a residual enhanced ventilatory drive.

Whole Body CO₂s

By superimposing controlled hyperventilation, one can obtain estimates of whole-body CO₂s during exercise. From measurements in these “hyperventilation” experiments during ischemic exercise, the whole-body CO₂ capacitance (dissociation curve) was $1.2 \text{ L} \cdot \text{mmHg}^{-1} \cdot \text{kg}^{-1}$, as calculated

from the excess of measured vs predicted $\dot{V}CO_2$ (497 mL) (Figure 2) per change in $PACO_2$ (5 mmHg) (Figure 3) per body weight (82.5 kg).

Model of Total and Changing Gas Stores

Table 1 shows the compartmental and total gas stores calculated for the three exercise conditions from the flow and volume redistribution model. Because lactate, bicarbonate concentration (HCO_3^-), and BE changes are linearly related [22], we incorporated a decrease in whole blood BE of 4 mmol/L estimated from other studies (see "Discussion") during the 5th min after cuff deflation to account for circulating lactate. The values from the total stores model from Table 1 are indicated in Figure 4 in relation to the 5 min-integrated stores changes obtained from measured gas exchange (Figure 2). According to the model, during cuff inflation, total O_2s did not change and CO_2s decreased 164 mL, whereas the 5 min totals (Figure 2) decreased 227 and 497 mL, respectively. The

difference indicates that the redistribution of blood volume and flow, the anaerobic work component, and hyperventilation resulted in losses of 227 mL and 333 mL in O_2s and CO_2s , respectively. During the 5th min after cuff deflation, O_2s increased by 18 mL and CO_2s decreased another 465 mL, whereas the 5 min totals showed that O_2s increased by 518 mL and CO_2s decreased by 1,162 mL. For O_2s , reducing the 518 mL gain after cuff deflation by the 18 mL increase in total stores, as well as the 227 mL deficit during prior inflation (which is being repaid), leaves a net gain of 273 mL used to repay the anaerobic cost during ischemia. The 1,162 mL 5 min loss in CO_2s after cuff deflation exceeds the 465 mL loss in absolute stores by 697 mL (Figure 4). Over the total 10 min, 5 min before and 5 min after inflation, the ratio of the total loss in CO_2s versus gain in O_2s is 3.7 (1,030/273), which includes the hyperventilation "artifact" during ischemia.

Table 1.

Estimated oxygen stores (O_2s) and carbon dioxide stores (CO_2s) (milliliters) during three conditions (A, B, and, C) of 15 min exercise: 5th min baseline, 5th min of inflation, and 5th min after cuff deflation, respectively.

| Condition | Location | PO_2 | O_2 Stores | PCO_2 | CO_2 Stores |
|--|----------|--------|--------------|---------|---------------|
| A. 5th Min Baseline (BE = -1.8 mmol/L, pHa = 7.420, \dot{Q} = 15 L/min) | Arterial | 84.1 | 232 | 33.8 | 479 |
| | Venous | 28.9 | 486 | 45.4 | 2,293 |
| | Tissue | 28.9 | 37 | 45.4 | 3,824 |
| | Lung | 84.1 | 576 | 33.8 | 231 |
| Total | — | — | 1,331 | — | 6,827 |
| B. 5th Min Cuff Inflation (BE = -1.8 mmol/L, pHa = 7.462, \dot{Q} = 14 L/min) | Arterial | 92.1 | 293 | 28.9 | 559 |
| | Venous | 25.3 | 375 | 44.7 | 2,146 |
| | Tissue | 25.3 | 32 | 44.7 | 3,760 |
| | Lung | 92.1 | 631 | 28.9 | 198 |
| Total | — | — | 1,331 | — | 6,663 |
| B - A | — | — | 0 | — | -164 |
| C. 5th Min Cuff Deflation* (BE = -5.8 mmol/L, pHa = 7.370, \dot{Q} = 15 L/min) | Arterial | 86.6 | 232 | 31.4 | 402 |
| | Venous | 30.3 | 486 | 42.7 | 1,986 |
| | Tissue | 30.3 | 38 | 42.7 | 3,595 |
| | Lung | 86.6 | 593 | 31.4 | 215 |
| Total | — | — | 1,349 | — | 6,198 |
| C - B | — | — | 18 | — | -465 |
| C - A | — | — | 18 | — | -629 |

Note: O_2s and CO_2s are based on model given in "Methods" of main text with assumptions:

- Total blood volume = 5,900 mL.
- Venous volume = total \times 0.8 = 4,720 mL; arterial = total \times 0.2 = 1,180 mL in conditions A and C.
- 300 mL was shifted from venous to arterial compartment in condition B; i.e., venous = 4,420 mL and arterial = 1,480 mL.

*Adjusted for $\Delta BE = -4.0$ mmol/L.

BE = base excess (measure of whole blood buffer base), pHa = arterial pH (negative log of H^+ in arterial blood), PCO_2 = partial pressure of carbon dioxide, PO_2 = partial pressure of oxygen, \dot{Q} = cardiac output.

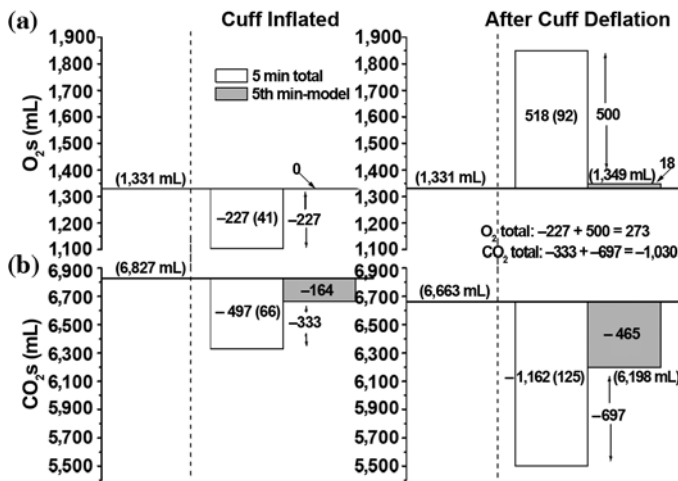


Figure 4.

Values for total gas stores from model in **Table 1** in relation to changes in gas stores from time-integrated $\dot{V}O_2$ and $\dot{V}CO_2$ shown in **Figure 2**. Standard error of the mean is shown for latter in parentheses. Total increase in (a) O₂s for 5 min of inflation and 5 min postinflation phase is 273 mL, and decrease in (b) CO₂s is 1,030 mL. CO₂ = carbon dioxide, CO₂s = CO₂ stores, O₂ = oxygen, O₂s = O₂ stores, $\dot{V}CO_2$ = CO₂ output, $\dot{V}O_2$ = O₂ uptake.

DISCUSSION

The initial increase in $\dot{V}O_2$ during the 1st min of ischemia can be accounted for by the bolus of venous blood from the legs moving into the central circulation during cuff inflation and its oxygenation to arterial blood as it traverses the pulmonary capillaries. This ≈ 30 mL of O₂ (**Figure 2**) would reoxygenate 300 mL of venous blood having an O₂ content of 10 vol%. This rise in $\dot{V}O_2$ and the ≈ 30 mL significant simultaneous loss of CO₂ ($p = 0.004$) indicated a 300 mL shift of blood from the venous to arterial compartment. A redistribution of blood flow accounted for the transient reduction in $\dot{V}O_2$ during the 2nd min of ischemia, whereby cuffs restricted O₂ delivery to the legs by arterial blood, reducing $\dot{V}O_2$ temporarily and increasing O₂ content of mixed venous blood. Similar cardiovascular readjustments with breath holds during exercise have been noted to reduce $\dot{V}O_2$ [26]. The linear rise during the last 3 min with ischemia reflects the decreasing mechanical efficiency and the progressive partial restoration of leg circulation. The peak 45 s after cuff deflation signifies lung reoxygenation of venous blood returning from the legs, extracting more O₂ to repay the aerobic and anaerobic deficit incurred during the prior ischemia. Most of the anaerobic deficit was repaid over the last 3 min of

uncuffed exercise as $\dot{V}O_2$ returned to near baseline exercise levels. However, some residual debt repayment probably occurred during the postexercise rest because the repayment exceeded the deficit at the start of exercise by 423 mL (**Figure 1**) and the half-time of the off-response (37 s) was significantly ($p = 0.001$) slower than the on-response (27 s); the latter value agreed with previous reports [27–28].

The estimated CO₂ capacitance value of 1.2 L·mmHg⁻¹·kg⁻¹ is lower than that (1.6) interpolated for the same exercise workload from a report [29] during 15 min of hyperventilation, although values twice as high have also been reported [30]. Capacitance values are directly related to the length of experiments, because more CO₂ is then washed out of slower compartments [31]. Because leg perfusion was impaired during our experiments, one would have expected a relatively low capacitance value because CO₂ in blood and tissue of the legs are then washed out at a slower rate, being somewhat isolated from the lung. Another consideration is that CO₂s change significantly slower than O₂s, having a half-time of 4.0 min versus 0.5 min for O₂s, based on studies on dogs by Farhi and Rahn [24]. This finding suggests that part of the loss in CO₂s following cuff deflation may be attributed to the hyperventilation during the prior ischemia.

After cuff deflation, the larger CO₂s loss relative to O₂s gain resulted from the HCO₃⁻ buffering of lactate entering the circulation. Correlation of lactate levels with excess $\dot{V}CO_2$ in relation to $\dot{V}O_2$ during and after heavy exercise resulted in the “anaerobic threshold” concept [32–33]. Excess $\dot{V}CO_2$ during exercise has also been used to estimate lactate accumulation in physically fit subjects [34] and cardiac patients [35]. The elevated $\dot{V}CO_2$ and CO₂s depletion is caused by carbonic acid, arising from the combination of H⁺ with HCO₃⁻; dissolved CO₂ from the muscle tissue being transported to the lungs once circulation is restored; and elevated $\dot{V}E$. As shown by $\dot{V}E/\dot{V}CO_2$ in **Figure 3**, the metaboreflex ventilatory drive was quickly diminished after cuff deflation, but the drive was then taken over by the chemoreflex stimulated by elevated H⁺ and PCO₂ in blood arriving at central chemoreceptors and continuing during subsequent rest.

In studies somewhat similar to this one, a rise of arterial blood lactate of ≈ 4 mmol/L was reported 4 to 5 min after cuff deflation [36] and also a 4 mmol/L loss of plasma HCO₃⁻ [37]. This amount of lactate release was incorporated into the model shown in **Table 1** and **Figure 4**. If 4 mmol/L of lactate release from the legs to central

circulation was entirely buffered by HCO_3^- during the 5 min postinflation period, it would amount to a CO_2s loss of $4 \text{ mmol/L} \times 5.9 \text{ L} \times 22.3 \text{ mL/mmol} = 526 \text{ mL}$ [33]. This amount accounts for 75 percent of the 697 mL estimate. However, the ratio of CO_2 loss to O_2 gain of 2.6 (697/273) suggests that a part of the lactate may have been converted by oxidation, in addition to being buffered [38]. These and other biochemical processes must have continued beyond the postexercise resting measurement period to fully restore O_2s and CO_2s to baseline levels of 1,331 and 6,827 mL, respectively. However, most of the excess CO_2 was eliminated by the time exercise stopped because $\dot{V}\text{CO}_2$ had returned to baseline (Figure 1). Without prolonged lactate turnover measurements, we can only generalize that the majority of the lactate was buffered in preference to other chemical pathways to account for the CO_2s loss exceeding the O_2s gain. Qualitatively, \dot{V}_E increases during exercise with cuffs inflated, depleting CO_2s , while the partially anaerobic exercise continues. When cuffs are deflated and after exercise stops, metabolic by-products from the legs returning to the central circulation keep ventilation elevated to repay O_2s , while CO_2s remains below baseline for a longer time.

Clearly, the assumptions in the total gas stores model demonstrated in Table 1 and Figure 4 will affect the absolute values and changes in gas store values. Some quantities, such as tissue water and arterial and venous blood volumes, are not easily measured and were taken from estimates in the literature. To quantify the effect of variations in these assumed values, in Table 2, we show changes in total O_2s and CO_2s resulting from variations

in values from those used in Table 1 during the three exercise conditions. We varied indicated values for relevant physiological components individually, assuming the other variables remained constant. Table 2 indicates that calculations of total O_2s and CO_2s and phase differences in stores are most sensitive to values for Hb and reductions in \dot{Q} during the ischemic phase. Any alveolar-arterial differences in PO_2 and PCO_2 greatly influence total stores, especially CO_2s , but the effect on store differences is smaller, somewhat similar to changing values for the other components. Therefore, performing invasive measurements, including arterial and mixed venous blood gases and lactate, in more definitive future studies is important.

Most studies using cuffs to induce acute exercise ischemia have focused on the \dot{V}_E response following cuff deflation to study CO_2 chemoreceptor response mechanisms. Data from some of these reports [23,36–37,39–40] allowed a gas store pattern estimation to compare with this study and are shown in Table 3. Generalizations from these limited data include (1) an inverse relationship between cuff pressure and O_2s reduction during inflation, (2) a direct relationship between workload and the increase in O_2s and reduction in CO_2s after cuff deflation, and (3) the CO_2s loss after cuff deflation exceeds the change during inflation and also exceeds the O_2s gain in recovery. From the time trends in the present study and those prior studies where time resolution was presented [37,39], apparently during inflation, the decrease in O_2s is attenuated as exercise duration increases. This finding is probably associated with the increasing $\dot{V}\text{O}_2$ required by the

Table 2.

Effect on total gas stores of variations in assumed values for gas stores model during three conditions A, B, and C of 15 min exercise: 5th min baseline, 5th min of cuff inflation, and 5th min after cuff deflation, respectively.

| Variable | Value | Exercise Condition | Value Change | O_2s Difference | | CO_2s Difference | |
|----------------------------------|------------------------------------|--------------------|--------------|---------------------------------|-------|----------------------------------|-------|
| | | | | Total* (%)† | Diff‡ | Total* (%)† | Diff‡ |
| Functional Residual Capacity (L) | 4.0 | A, B, C | ±10% | 60 (4.5) | 5 | 21 (0.3) | 3 |
| Blood Volume (L) | 5.9 | A, B, C | ±10% | 69 (5.2) | 8 | 263 (4.0) | 19 |
| Hb (g%) | 15.0 | A, B, C | ±10% | 118 (8.8) | 2 | 110 (1.7) | 34 |
| Alv-Art Diff (mmHg) | PCO_2 & $\text{PO}_2 = 0$ | A, B, C | 3, 13 | 112 (8.4) | 10 | 536 (8.2) | 32 |
| H^+ (nmol/L) | pHa = 7.42 at base | A, B, C | ±10% | 4 (0.3) | 2 | 193 (2.9) | 18 |
| \dot{Q} Decrease (L/min) | 1.0 | B | 0 & 2 | 37 (2.8) | 37 | 141 (2.1) | 141 |
| BE Decrease (mmol/L) | 4.0 | C | -3 & -5 | 2 (0.1) | 2 | 73 (1.2) | 73 |

Note: CO_2s decreases 73 mL per 1.0 mmol/L decrease in BE.

*Mean absolute differences in total gas stores (milliliters) from values in Table 1 (see main text).

†These mean differences as % of values in Table 1.

‡Mean of differences in gas store changes between conditions from those in Table 1.

Alv-Art = alveolar-arterial, BE = base excess (measure of whole blood buffer base), CO_2s = carbon dioxide stores, diff = differences, H^+ = hydrogen ion concentration, Hb = hemoglobin concentration, O_2s = oxygen stores, PCO_2 = partial pressure of carbon dioxide, pHa = arterial pH, PO_2 = partial pressure of oxygen, \dot{Q} = cardiac output.

Table 3.

Cumulative time-integrated oxygen uptake ($\dot{V}O_2$), carbon dioxide output ($\dot{V}CO_2$), and pulmonary ventilation (\dot{V}_E) differences from baseline during leg cuff inflation and after cuff deflation.

| Studies | n | Work (W) | Cuff Pressure (mmHg) | Work Time Before Inflation (min) | Inflation | | | Deflation | | | | |
|---|------------|----------|----------------------|----------------------------------|----------------------|-------------------|--------------------|-----------------|---------------------|-------------------|--------------------|-----------------|
| | | | | | Inflation Time (min) | $\dot{V}O_2$ (mL) | $\dot{V}CO_2$ (mL) | \dot{V}_E (L) | Recovery Time (min) | $\dot{V}O_2$ (mL) | $\dot{V}CO_2$ (mL) | \dot{V}_E (L) |
| Stegemann, 1963 [1] | None given | 0 | 250–300 | 0 | 10 | –495 | –387 | –13 | 20 | 448 | 599 | 14 |
| | None given | “mild” | 250–300 | 20 | 6 | –233 | –121 | 0 | 10 | 228 | 399 | 17 |
| Asmussen & Nielsen, 1964 (CO ₂ added to maintain PACO ₂) [2] | 1 | 31 | 300–350 | 10–15 | 3 | –358 | — | — | — | — | — | — |
| | 4 | 62 | 300–350 | 10–15 | 3 | –984 | — | — | — | — | — | — |
| | 2 | 124 | 300–350 | 10–15 | 2 | –1,183 | — | — | — | — | — | — |
| Sargeant et al., 1981 [3] | 5 | 100 | 250 | 0 | 2 | –130 | 350 | 32 | 4 | 770 | 1,591 | 45 |
| Stanley et al., 1985 [4] | 8 | 49 | 200 | 6 | 2 | –274 | 129 | 8 | 4 | 547 | 799 | 16 |
| | 8 | 98 | 200 | 6 | 2 | –428 | 106 | 10 | 4 | 643 | 1,151 | 23 |
| Roth et al., 1988 [5] | 9 | ≈17 | 200 | 6 | 2 | –740 | — | — | 4 | 1,440 | — | — |
| This Study | 6 | 75 | 140 | 5 | 5 | 282 | 932 | 55 | 5 | 654 | 1,283 | 57 |
| | | | | | (2) | –42 | 167 | 10 | | | | |

Note: See legend to **Figure 2** for “predicted” values for $\dot{V}O_2$ and $\dot{V}CO_2$ for this study; here all “predicted” values were assumed equal to baseline.

1. Stegemann J. [On the mechanism of pulse frequency regulation by metabolism. I. The influence of metabolism in a muscle group isolated from the circulation on the behavior of the pulse frequency]. *Pflügers Arch Gesamte Physiol Menschen Tiere*. 1963;276:481–92. German. [PMID: 13983630]
2. Asmussen E, Nielsen M. Experiments on nervous factors controlling respiration and circulation during exercise employing blocking of the blood flow. *Acta Physiol Scand*. 1964;60:103–11. [PMID: 14131818]
3. Sargeant AJ, Rouleau MY, Sutton JR, Jones NL. Ventilation in exercise studied with circulatory occlusion. *J Appl Physiol*. 1981;50(4):718–23. [PMID: 6790486]
4. Stanley WC, Lee WR, Brooks GA. Ventilation studied with circulatory occlusion during two intensities of exercise. *Eur J Appl Physiol Occup Physiol*. 1985;54(3):269–77. [PMID: 3933976]
5. Roth DA, Stanley WC, Brooks GA. Induced lactacidemia does not affect postexercise O₂ consumption. *J Appl Physiol*. 1988;65(3):1045–49. [PMID: 3182473]

elevated \dot{V}_E and extra muscular effort and partial restoration of leg blood flow that diminish the O₂s deficit and increase the CO₂s deficit. Apparently, leg cuff pressures must be >90 mmHg during exercise to affect measured $\dot{V}CO_2$ and $\dot{V}O_2$ during exercise [41–42].

SUMMARY AND CONCLUSIONS

The events in these experiments can be described as a respiratory alkalosis during ischemia, followed by a metabolic acidosis after cuff deflation when metabolites from the anaerobic portion of leg work return to the central circulation. Changes in O₂s depend mainly on perfusion through lung and tissue, while CO₂s changes are primarily determined by \dot{V}_E , venous blood redistribution, and HCO₃[–] buffering of lactate. This study estimated that the ischemia required a repayment of 273 mL of O₂ and produced 697 mL of CO₂. These values depend on workload,

work duration with ischemia, the cuff pressure determining the perfusion impairment, and the intensity of the metaboreflex. The amount of anaerobic debt incurred and tolerated and the recovery from a given ischemic exercise scenario will depend on the aerobic fitness of the subject and related blood pressure reflex response. These factors must be considered if this form of exercise is further evaluated and implemented for rehabilitation.

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REFERENCES

1. Tyni-Lenné R, Dencker K, Gordon A, Jansson E, Sylvén C. Comprehensive local muscle training increases aerobic working capacity and quality of life and decreases neurohormonal activation in patients with chronic heart failure. *Eur J Heart Fail.* 2001;3(1):47–52. [\[PMID: 11163735\]](#)
2. Eiken O. Responses to dynamic leg exercise in man as influenced by changes in muscle perfusion pressure. *Acta Physiol Scand Suppl.* 1987;566:1–37. [\[PMID: 3480686\]](#)
3. Sundberg CJ, Kaijser L. Effects of graded restriction of perfusion on circulation and metabolism in the working leg; quantification of a human ischaemia-model. *Acta Physiol Scand.* 1992;146(1):1–9. [\[PMID: 1442118\]](#)
4. Takarada Y, Sato Y, Ishii N. Effects of resistance exercise combined with vascular occlusion on muscle function in athletes. *Eur J Appl Physiol.* 2002;86(4):308–14. [\[PMID: 11990743\]](#)
5. Teramoto M, Golding LA. Low-intensity exercise, vascular occlusion, and muscular adaptations. *Res Sports Med.* 2006;14(4):259–71. [\[PMID: 17214403\]](#)
6. Takarada Y, Takazawa H, Ishii N. Applications of vascular occlusion diminish disuse atrophy of knee extensor muscles. *Med Sci Sports Exerc.* 2000;32(12):2035–39. [\[PMID: 11128848\]](#)
7. Burgomaster KA, Moore DR, Schofield LM, Phillips SM, Sale DG, Gibala MJ. Resistance training with vascular occlusion: metabolic adaptations in human muscle. *Med Sci Sports Exerc.* 2003;35(7):1203–8. [\[PMID: 12840643\]](#)
8. Nygren AT, Sundberg CJ, Göransson H, Esbjörnsson-Liljedahl M, Jansson E, Kaijser L. Effects of dynamic ischaemic training on human skeletal muscle dimensions. *Eur J Appl Physiol.* 2000;82(1–2):137–41. [\[PMID: 10879455\]](#)
9. Piepoli M, Clark AL, Volterrani M, Adamopoulos S, Sleight P, Coats AJ. Contribution of muscle afferents to the hemodynamic, autonomic, and ventilatory responses to exercise in patients with chronic heart failure: effects of physical training. *Circulation.* 1996;93(5):940–52. [\[PMID: 8598085\]](#)
10. Loeppky JA, Gurney B, Kobayashi Y, Icenogle MV. Effects of ischemic training on leg exercise endurance. *J Rehabil Res Dev.* 2005;42(4):511–22. [\[PMID: 16320146\]](#)
11. Scott AC, Wensel R, Davos CH, Georgiadou P, Kemp M, Hooper J, Coats AJ, Piepoli MF. Skeletal muscle reflex in heart failure patients: role of hydrogen. *Circulation.* 2003;107(2):300–306. [\[PMID: 12538432\]](#)
12. Loeppky JA, Luft UC. Fluctuations in O₂ stores and gas exchange with passive changes in posture. *J Appl Physiol.* 1975;39(1):47–53. [\[PMID: 1150591\]](#)
13. Chuang ML, Ting H, Otsuka T, Sun XG, Chiu FY, Beaver WL, Hansen JE, Lewis DA, Wasserman K. Aerobically generated CO₂ stored during early exercise. *J Appl Physiol.* 1999;87(3):1048–58. [\[PMID: 10484576\]](#)
14. Baldwin KM. Comments on classical papers. *J Appl Physiol.* 2005;99(4):1241–42. [\[PMID: 16160016\]](#)
15. Luft UC, Loeppky JA, Mostyn EM. Mean alveolar gases and alveolar-arterial gradients in pulmonary patients. *J Appl Physiol.* 1979;46(3):534–40. [\[PMID: 438024\]](#)
16. Rahn H, Fenn WO. A graphical analysis of the respiratory gas exchange: The O₂-CO₂ diagram. Washington (DC): American Physiological Society; 1955. p. 40.
17. McGregor M, Becklake MR. The relationship of oxygen cost of breathing to respiratory mechanical work and respiratory force. *J Clin Invest.* 1961;40:971–80. [\[PMID: 13773979\]](#)
18. Vella CA, Marks D, Robergs RA. Oxygen cost of ventilation during incremental exercise to VO₂ max. *Respirology.* 2006;11(2):175–81. [\[PMID: 16548903\]](#)
19. Vidal Melo MF, Loeppky JA, Caprihan A, Luft UC. Alveolar ventilation to perfusion heterogeneity and diffusion impairment in a mathematical model of gas exchange. *Comput Biomed Res.* 1993;26(2):103–20. [\[PMID: 8477584\]](#)
20. Loeppky JA, Caprihan A, Altobelli SA, Icenogle MV, Scotto P, Vidal Melo MF. Validation of a two-compartment model of ventilation/perfusion distribution. *Respir Physiol Neurobiol.* 2006;151(1):74–92. [\[PMID: 16024300\]](#)
21. Severinghaus JW. Simple, accurate equations for human blood O₂ dissociation computations. *J Appl Physiol.* 1979;46(3):599–602. [\[PMID: 35496\]](#)
22. Loeppky JA, Fletcher ER, Roach RC, Luft UC. Relationship between whole blood base excess and CO₂ content in vivo. *Respir Physiol.* 1993;94(1):109–20. [\[PMID: 8272578\]](#)
23. Asmussen E, Nielsen M. Experiments on nervous factors controlling respiration and circulation during exercise employing blocking of the blood flow. *Acta Physiol Scand.* 1964;60:103–11. [\[PMID: 14131818\]](#)
24. Farhi LE, Rahn H. Gas stores of the body and the unsteady state. *J Appl Physiol.* 1955;7(5):472–84. [\[PMID: 14367232\]](#)
25. Loeppky JA, Luft UC, Fletcher ER. Quantitative description of whole blood CO₂ dissociation curve and Haldane effect. *Respir Physiol.* 1983;51(2):167–81. [\[PMID: 6405469\]](#)
26. Lindholm P, Linnarsson D. Pulmonary gas exchange during apnoea in exercising men. *Eur J Appl Physiol.* 2002;86(6):487–91. [\[PMID: 11944095\]](#)
27. Linnarsson D. Dynamics of pulmonary gas exchange and heart rate changes at start and end of exercise. *Acta Physiol Scand Suppl.* 1974;415:1–68. [\[PMID: 4621315\]](#)

28. Özyener F, Rossiter HB, Ward SA, Whipp BJ. Influence of exercise intensity on the on- and off-transient kinetics of pulmonary oxygen uptake in humans. *J Physiol.* 2001; 533(Pt 3):891–902. [\[PMID: 11410644\]](#)
29. Jones NL, Jurkowski JE. Body carbon dioxide storage capacity in exercise. *J Appl Physiol.* 1979;46(4):811–15. [\[PMID: 457560\]](#)
30. Ozcelik O, Ward SA, Whipp BJ. Effect of altered body CO₂ stores on pulmonary gas exchange dynamics during incremental exercise in humans. *Exp Physiol.* 1999;84(5): 999–1011. [\[PMID: 10502667\]](#)
31. Farhi LE. Gas stores of the body. In: Fenn WO, Rahn H, editors. *Handbook of physiology. Section 3: Respiration. Vol I.* Washington (DC): American Physiological Society; 1964. p. 873–85.
32. Stringer W, Wasserman K, Casaburi R. The VCO₂/VO₂ relationship during heavy, constant work rate exercise reflects the rate of lactic acid accumulation. *Eur J Physiol Occup Physiol.* 1995;72(1–2):25–31. [\[PMID: 8789566\]](#)
33. Whipp BJ, Mahler M. Dynamics of pulmonary gas exchange. In: West JB, editor. *Pulmonary gas exchange.* New York (NY): Academic Press; 1980. p. 33–96.
34. Hirakoba K, Maruyama A, Misaka K. Prediction of blood lactate accumulation from excess CO₂ output during constant exercise. *Appl Human Sci.* 1996;15(5):205–10. [\[PMID: 8979401\]](#)
35. Wilson JR, Ferraro N, Weber KT. Respiratory gas analysis during exercise as a noninvasive measure of lactate concentration in chronic congestive heart failure. *Am J Cardiol.* 1983;51(10):1639–43. [\[PMID: 6407294\]](#)
36. Roth DA, Stanley WC, Brooks GA. Induced lactacidemia does not affect postexercise O₂ consumption. *J Appl Physiol.* 1988;65(3):1045–49. [\[PMID: 3182473\]](#)
37. Sargeant AJ, Rouleau MY, Sutton JR, Jones NL. Ventilation in exercise studied with circulatory occlusion. *J Appl Physiol.* 1981;50(4):718–23. [\[PMID: 6790486\]](#)
38. Kelley KM, Hamann JJ, Navarre C, Gladden LB. Lactate metabolism in resting and contracting canine skeletal muscle with elevated lactate concentration. *J Appl Physiol.* 2002; 93(3):865–72. [\[PMID: 12183479\]](#)
39. Stanley WC, Lee WR, Brooks GA. Ventilation studied with circulatory occlusion during two intensities of exercise. *Eur J Appl Physiol Occup Physiol.* 1985;54(3):269–77. [\[PMID: 3933976\]](#)
40. Stegemann J. [On the mechanism of pulse frequency regulation by metabolism. I. The influence of metabolism in a muscle group isolated from the circulation on the behavior of the pulse frequency]. *Pflügers Arch Gesamte Physiol Menschen Tiere.* 1963;276:481–92. German. [\[PMID: 13983630\]](#)
41. Greiner A, Esterhammer R, Pilav S, Arnold W, Santner W, Neuhauser B, Fraedrich G, Jaschke WR, Schocke MF. High-energy phosphate metabolism in the calf muscle during moderate isotonic exercise under different degrees of cuff pressure: a phosphorus 31 magnetic resonance spectroscopy study. *J Vasc Surg.* 2005;42(2):259–67. [\[PMID: 16102624\]](#)
42. Smith SA, Gallagher KM, Norton KH, Querry RG, Welch-O'Connor RM, Raven PB. Ventilatory responses to dynamic exercise elicited by intramuscular sensors. *Med Sci Sports Exerc.* 1999;31(2):277–86. [\[PMID: 10063818\]](#)

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