

UNITED STATES DEPARTMENT OF AGRICULTURE
FOOD SAFETY AND INSPECTION SERVICE
WASHINGTON, DC

FSIS NOTICE

31-07

5/22/07

NATIONWIDE YOUNG CHICKEN MICROBIOLOGICAL BASELINE DATA COLLECTION PROGRAM – UPDATE

NOTE: This notice only applies to inspection program personnel at the establishments that are included as part of the baseline data collection program.

I. PURPOSE

This notice announces that the Food Safety and Inspection Service's (FSIS) "Nationwide Young Chicken Microbiological Baseline Data Collection Program (YCBS)" will begin on June 25, 2007, and that the 90-day training ("shake down") period referred to in FSIS Notice 60-06, "Nationwide Young Chicken Microbiological Baseline Data Collection Program" has ended. Additionally, this notice:

1. cancels FSIS Notice 60-06, dated 9/14/06. The sampling instructions from that notice are included in this notice and are unchanged;
2. provides the new sampling code numbers to be used for the baseline study;
3. provides an Attachment 1 with revised questions that will appear in Block 28 of FSIS Form 10,210-3, "Requested Sample Programs" (the sampling request form); and
4. provides an Attachment 2 which answers some of the questions FSIS received during the "shake down" period.

II. BACKGROUND

During the baseline study, which will continue for a minimum of twelve months, testing will include the collection of carcass rinses at **Re-Hang** and **Post-Chill** from broiler chickens slaughtered in Federal establishments. **Re-Hang** refers to the location in the process after the picker and prior to evisceration of the bird. **Post-Chill** refers to the point in the process at the end of the drip line after all interventions have taken place, but before the bird enters the cooler or proceeds to further processing. This baseline study will provide FSIS and the regulated industry with data concerning the

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prevalence and quantitative levels of selected foodborne pathogens and microorganisms that serve as indicators of process control (e.g., *Campylobacter*, generic *Escherichia coli*, *Salmonella*, *Enterobacteriaceae*, coliforms, and plate counts of aerobic microorganisms). This data will enable the Agency and industry to target interventions and effectively work toward reducing the risk posed by foodborne pathogens in FSIS-regulated products.

III. REVIEW OF FSIS DIRECTIVES AND VIDEO MATERIALS BY INSPECTION PROGRAM PERSONNEL

Upon receipt of this notice, inspection program personnel who will be collecting the samples are to review the following materials (links provided):

1. [FSIS Directive 10,230.5](#) and the [Self-Instruction Guide for Collecting Raw Meat and Poultry Product Samples for *Salmonella* Analysis](#)

2. The DVD titled, "Sampling Raw Meat and Poultry for *Salmonella*".

The DVD is the same as the VHS video that was originally issued in 1998 with FSIS Directive 10,230.5 for the pathogen reduction sampling initiative. You should have received a DVD copy of this instruction during the "shake down" period.

3. [FSIS Directive 10,210.1, Unified Sampling Form](#). This baseline uses FSIS Form 10,210-3 (Sample Request Form).

4. [FSIS Directive 7355.1, Use of Sample Seals for Laboratory Samples and Other Applications](#).

If FSIS personnel have questions concerning the directives, video material, or the sampling procedures, they may send questions to the "Young Chicken Baseline Mailbox" in Outlook, or contact the Technical Service Center (TSC) at 1-800-233-3935.

IV. AWARENESS MEETING WITH PLANT MANAGEMENT

A. Upon receipt of this notice, the Inspector-in-Charge (IIC) is to have an awareness meeting with plant management to inform them that the establishment will be part of the national baseline study and explain the program. The actual baseline study will commence with receipt of this notice.

B. The IIC will emphasize to plant management that the results of the microbiological sampling from this baseline testing will NOT be the basis for regulatory actions. The IIC will provide plant management with a copy of this notice and review the following program points during the awareness meeting:

1. The FSIS Nationwide Young Chicken Microbiological Baseline Data Collection Program will collect carcass rinses at **Re-Hang** and **Post-Chill** from broiler chickens.

2. The purpose of the baseline study is to collect data concerning the prevalence and quantitative levels of selected foodborne pathogens and microorganisms that serve as indicators of process control.
3. The laboratories will analyze the sample results, and FSIS will use the results for program development.
4. Inspection program personnel will only collect broilers for this baseline study. All broiler chickens that will have the head and feet removed and that will be eviscerated during the slaughter process are eligible for sampling in this program.
5. Inspection program personnel will collect samples from both production shifts.
6. The laboratory will test the samples for *Salmonella*, *Campylobacter*, generic *Escherichia coli*, *Enterobacteriaceae*, coliforms, and plate counts of aerobic microorganisms. Inspection program personnel will send all samples to the FSIS contract laboratory, Food Safety Net Services LTD, 221 West Rhapsody, San Antonio, Texas 78216.
7. The study will continue for a minimum of one year.
8. Once FSIS has completed the baseline study, the Agency will publish the study as the official baseline report. The report will present a summary of baseline findings on a national basis. FSIS will not publish individual plant results.
9. FSIS will not post the microbiological results in the Laboratory Electronic Application for Results Notification (LEARN) system.
10. The Agency expects each establishment to carry out any sampling described in its food safety system or required by regulation (9 CFR 381.94) for raw poultry regardless of whether FSIS collects a baseline sample.
11. There will be no regulatory action taken based on the results. Consequently, establishments need not hold product.

V. GENERAL SAMPLING PROCEDURES FOR THE BASELINE STUDY

NOTE: The following sampling procedure instructions are for this baseline study only. Inspection program personnel that are assigned to establishments that are participating in this study are to use the sample collection methods in this notice in lieu of the sample collection instructions in FSIS Directive 10,230.5.

- A. Inspection program personnel will follow the methodologies for collecting samples as directed in this notice.
- B. Sample request forms with the same collection week represent companion samples that form a sample pair. A sample pair consists of one carcass rinse collected

at **Re-Hang** and a second carcass rinse from the same grow-out flock/house slaughtered during the same specified production shift collected at **Post-Chill**.

C. Block 14 of FSIS Form 10,210-3 indicates the project ID code for the sample to be collected. There will be a sample ID code listed for the **Re-Hang** sample and a sample ID code listed for the **Post-Chill** sample, and the code will indicate the production shift from which the sample pair is to be collected.

The project ID codes for the sample pairs by production shift are as follows:

First Shift Pair	Second Shift Pair
B46REHG1 - Re-Hang Shift 1	B46REHG2 - Re-Hang Shift 2
B46POST1 - Post-Chill Shift 1	B46POST2 - Post-Chill Shift 2

D. Each sample request form identifies the week of sample collection (Block 4). To the extent possible, collect all samples during the designated week. Collect both samples from the same grow-out flock/house. Collected samples will go to the FSIS contract laboratory. Block 9 of FSIS Form 10,210-3 will have the name and address of the designated laboratory preprinted in it. The laboratory will accept samples collected outside of the designated week if resources (supplies, time, or personnel) or production timing do not permit submission of samples during the designated week.

E. Block 18 of FSIS Form 10,210-3 will denote the type of sample and from which production shift inspection program personnel are to collect the samples. The shift specified for sample collection will alternate during the study. For this study, the production shift corresponds with the shift for which inspection program personnel enter the slaughter totals into the Electronic Animal Disposition Reporting System (eADRS).

F. During this baseline study, the questions found in Attachment 1 of this notice will appear in Block 28 of FSIS Form 10,210-3 for **Re-Hang** sample requests (**B46REHG1** and **B46REHG2**).

G. Inspection program personnel are to sample from the specified production shift (based on eADRS data entry).

H. Inspection program personnel are to review Section 2, *Supplies*, and Section 5, *Preparation for Sample Collection*, of FSIS Directive 10,230.5, Attachment 1, for all samples collected. The supplies for this baseline study are different. The following is a list of the supplies for each sample request:

M20 box (shipper) with baseline sorting labels (project labels) plus two sets of supplies:

Supply List for **Re-Hang**:

- 1-15" x 20" sterile plastic bag
- 1- pair of sterile gloves

- 1- sterile wide-mouth container containing 400 ml of sterile Buffered Peptone Water (BPW), also to be reused to ship the sample rinsate to the contract lab
- 1- quart resealable ziplock-type bag
- 1- 6" x 12" plastic sleeve for FSIS Form 10,210-3
- 1- FSIS Form 7355-2A/2B (Laboratory sample container seals)

Supply List for **Post-Chill**:

- 1-15" x 20" sterile plastic bag
- 1- pair of sterile gloves
- 1- sterile wide-mouth container containing 400 ml of sterile BPW, also used to ship the sample rinsate to the contract lab
- 1- quart resealable ziplock-type bag
- 1- 6" x 12" plastic sleeve for FSIS Form 10,210-3
- 1- FSIS Form 7355-2A/2B (Laboratory sample container seals)

- Absorbent pad
- Coolboard separator
- Gel coolant pack(s)
- FedEx (preprinted) billable air bill

I. Inspection program personnel are to refrigerate the BPW upon receipt. Ensuring that the BPW is prechilled is CRITICAL to this baseline study. Only use prechilled BPW.

VI. PROCEDURES FOR RE-HANG SAMPLING

A. Inspection program personnel mark an "R" on the BPW container designated for the **Re-Hang** sample with an indelible marker.

B. Inspection program personnel are to mark an "R" on the sample request form identified with the project ID codes **B46REHG1** and **B46REHG2** in the upper right-hand corner of the form. Keep the sample request forms identified for the **Re-Hang** samples and the **Re-Hang** collected rinsate samples together. Marking the BPW container prior to selecting the samples prevents confusion. Do not write over the preprinted bar code.

C. When preparing a collection of **Re-Hang** samples, inspection program personnel are to prepare an ICE BATH to place collected samples in immediately after collection of the rinsate and are to take the ice bath to the sample collection area and randomly select a **Re-Hang** sample.

NOTE: To make the ice bath, use a small container (pail or small plastic tub). Place about 4 inches of ice in the bottom of the container. The purpose of the ice bath is to reduce the temperature of samples collected at **Re-Hang** as quickly as possible. The establishment has not eviscerated the carcasses selected at this point in the process, and they are still hot. Placing the rinsate sample into the ice bath will reduce the temperature and minimize the outgrowth of microorganisms. Alternatively, if there is no way of creating an ice bath, place the samples under refrigeration, or similar method,

within five (5) minutes of selecting the sample to ensure that the temperature of the sample is reduced to minimize the outgrowth of microorganisms. Remember, an ice bath is the preferred method for this baseline study.

VII. PROCEDURES FOR POST-CHILL SAMPLING

A. For **Post-Chill**, inspection program personnel are to determine the times at which carcasses from the same grow-out flock/house will reach the end of the drip line or the equivalent point in air-chill systems. Inspection program personnel are to follow the same procedure for selecting a poultry carcass from the **Post-Chill** area (after all interventions have taken place) that originated from the same grow-out flock/house.

B. Inspection program personnel are to change disposable gloves after they collect each sample, and whenever necessary, to prevent cross-contamination of birds and samples. Avoid contamination of carcasses and rinse bottles.

VIII. COLLECTING RINSATE

A. Inspection program personnel are to take all necessary precautions not to contaminate the BPW container once opened. After pouring in the contents of the BPW container, inspection program personnel are to replace the lid on the container, being careful not to contaminate the inside of the lid or container lip.

B. Inspection program personnel are to remove the chicken aseptically from the sample bag before collecting the rinsate. To do this:

1. carefully open the bag containing the bird;
2. work the plastic bag down around the carcass so that you can firmly grip one leg, without touching the inside of the plastic bag;
3. while holding the bag with the one hand, carefully remove the bird from the bag with the other hand; and
4. place the bird back on the conveyor or table.

C. Inspection program personnel are to collect rinsate samples from carcasses immediately after they select the carcass by:

1. removing the lid from the empty BPW container and place the lid once again on the table;
2. being careful not to contaminate the inside of the bottle or the lid, and by not allowing the bag to contact the interior surfaces of the bottle;
3. using the "V" formed by the bag at the lower corner as a pouring spout, carefully pour the rinsate into the open BPW container;

4. collecting as much of the BPW rinsate as possible, but at least 200 ml, and by making sure to use the BPW container marked with an “R” for the **Re-Hang** rinsate sample;
5. placing the cap back on the container and checking to be sure that the lid is securely in place;
6. placing the rinse bottles for **Re-Hang** samples in an ice bath immediately after sample collection, or refrigerating the sample within five (5) minutes of collection;
7. placing collected and labeled sample containers in a ziplock-type bag; and
8. holding the samples under refrigeration and FSIS control until shipment to the laboratory.

IX. SAMPLE STORAGE PRIOR TO SHIPMENT

A. Inspection program personnel are to review and follow Section 8, *Sample Storage Prior to Shipment*, of FSIS Directive 10, 230.5, Attachment 1, for all samples collected. All samples are to be refrigerated or placed on ice immediately after sampling and maintained under refrigeration at 40°F, or lower, until shipped. All samples are to be kept secure. Inspection program personnel are not to freeze samples.

B. Inspection program personnel are to avoid storing sample boxes near heaters or areas exposed to excessive heat. The laboratory will discard rinse samples that arrive at or above 10°C (50°F). **It is critical to the success of this baseline study that the sample temperature is maintained during collection and shipment.**

X. SHIPPING OF SAMPLES

A. Inspection program personnel are to review and follow the instructions in Section 9, *Sample Shipment*, of FSIS Directive 10,230.5, Attachment 1. Samples are to be collected and shipped to the laboratory the same day when possible.

1. First shift samples should be shipped the same day collected or else they will be discarded by the laboratory. First shift samples may be collected Monday through Friday.

2. Second shift samples should only be collected Monday through Thursday, because of shipping-related issues. Samples collected on the second shift and shipped to the contract laboratory the next day will not be discarded.

NOTE: Both halves of a sample pair must be received in order to ensure that the results from the sample pair are accepted into the Young Chicken Baseline project database.

B. The sample pair is to be shipped in the same shipping container and is to be shipped to the laboratory contracted by FSIS to conduct this testing [Food Safety Net Services LTD, 221 West Rhapsody, San Antonio, TX 78216] listed in Block 9 of FSIS Form 10,210-3.

XI. OBTAINING RESULTS OF FSIS MICROBIOLOGICAL TESTING OF YOUNG CHICKEN (BROILER) BASELINE SAMPLES

Inspection program personnel will not receive laboratory testing results for the non-regulatory samples analyzed at the contract laboratory. These non-regulatory testing results are not posted in LEARN.

XII. CLARIFICATION OF COMMON QUESTIONS

During the 90-day training phase for this baseline study, many questions were fielded at the Technical Service Center and through the “Young Chicken Baseline Mailbox” in Outlook. Many of these questions are addressed in Attachment 2 of this notice, *Questions and Answers Concerning Young Chicken Nationwide Microbiological Baseline Data Collection Program*.

Inspectors are to direct questions to the “Young Chicken Baseline Mailbox” in Outlook, or the Technical Service Center at 1-800-233-3935.



Assistant Administrator
Office of Policy, Program, and Employee Development

**Changes to Block 28 Questions on
FSIS Form 10,210-3**

The information in this Attachment supersedes the information in paragraphs VI. F. and G. from FSIS Notice 60-06. FSIS has revised the questions in Block 28 of the forms for this study. FSIS removed some of the questions asked during the training period (i.e. “shake down”) and added others.

VI. F. The following questions will appear in Block 28 for **Re-Hang** sample requests (B46REHG1 and B46REHG2). The information requested on both forms is essential for the correct analysis of baseline data. The italicized text under the questions provides guidance that will not appear on the form.

1. During which production shift was the bird slaughtered?

First ___ Second ___

Identify the production shift based on how you enter slaughter totals into eADRS. This will confirm you collected samples from the specified shift.

2. Did the shift during which this bird was slaughtered occur immediately after a total clean-up/pre-op?

Yes ___ No ___

Check the appropriate response to identify if this production shift began immediately following a clean-up or pre-operational inspection.

Recall that the production shift number should correspond with the shift for which slaughter totals will be entered in eADRS.

3. Approximately what percent of young chickens slaughtered during this shift were broilers? _____%

Response is a number ranging from 1 to 100, rounded to the nearest percent.

Broilers are young chickens (usually under 13 weeks of age), of either sex, that are tender-meated with soft, pliable, smooth-textured skin and flexible breastbone cartilage. (9 CFR 381.170(a) (1) (iii))

If there is doubt as to the class of the chicken (broilers versus roasters versus Cornish game hens), use the establishment’s label designation as final identification of class. For this study, this label designation is “broiler.”

4. How many broilers did the plant slaughter during the same production shift as this carcass (in thousands)? _____ (If less than or equal to 1000, enter 1)

Response is a number ranging from 1 to 150, rounded to the nearest whole number, where 1 represents ≤1000 broilers slaughtered per shift and 150 represents 150,000 broilers slaughtered per shift.

5. **What is the estimated live weight of this bird (in pounds)?** _____

Response is a number ranging from 0.5 to 10.0, rounded to the nearest half-pound.

6. **FORM NUMBER OF COMPANION POST-CHILL SAMPLE** _____

Provide the form number from Block 1 of the **Post-Chill** Sample Request Form (FSIS Form 10,210-3) to be collected on the same shift from the same grow-out flock/house. (B46REHG1/B46POST1 or B46REHG2/B46POST2)

VI. G. The following questions will appear in Block 28 for **Post-Chill** sample requests (**B46POST1** and **B46POST2**). The italicized clarifications under the questions provide guidance that will not appear on the form.

1. **What is the estimated post-evisceration weight of this bird (in pounds)?**

Expected response is a number ranging from 0.5 to 10.0, rounded to the nearest half-pound.

2. **Which antimicrobial treatment is used in the on-line reprocessing system? CIRCLE THE TREATMENT USED.**

No on-line reprocessing Acidified Sodium Chlorate TSP FreshFx

Chlorine Dioxide Inspexx Hypochlorous Acid

Other _____

An on-line reprocessing system refers to the on-line removal of feces, digestive tract contents, or extraneous materials that are contaminating the abdominal cavity of dressed carcasses and requires a waiver from the New Technology Staff. (This is in contrast to off-line reprocessing that occurs at a designated off-line work station.) An antimicrobial treatment refers to the use of any safe and suitable processing aid which reduces carcass bacterial loads.

Circle the appropriate response:

Acidified sodium chlorite refers to the product known as Sanova®.

TSP refers to the product, trisodium phosphate.

FreshFx® refers to an aqueous solution of citric, phosphoric, and hydrochloric acids produced by SteriFX®.

Inspexx® is a peroxyacetic acid-based antimicrobial surface treatment produced by Ecolab®.

Hypochlorous acid refers to the chlorine-based antimicrobial treatment produced by Tomco2 ®.

If an antimicrobial treatment other than those listed is in use, please circle "Other" and fill in the product name. Limit this response to 16 characters or less.

Mention of companies or commercial products does not imply recommendation or endorsement by the USDA over others not mentioned. USDA neither guarantees nor warrants the standards of any product mentioned. Product names are mentioned solely as examples.

3. **Was chlorine used in the inside/outside birdwasher (IOBW) on the shift this bird was slaughtered?**

Yes ___ **No** ___ **No IOBW** ___

An IOBW uses high pressure water spray to flush the interior and exterior of the carcass prior to chilling.

If a high pressure IOBW is not used during this shift, then check "No IOBW" for this question. Otherwise, check the appropriate response.

4. **Are Post-Chill interventions (e.g., Post-Chill dips) routinely used?**

Yes _____ **No** _____

Were any used today?

Yes _____ **No** _____

Two responses are requested here. The first response refers to routine practice in the plant and the second response refers to any specific action taken on the day and shift of sample collection.

5. **Name of Grower** _____

Specify the name of the grower of the flock (the specific house of origin) represented by the sample pair. Use abbreviations to limit this response to sixteen (16) characters.

6. **FORM NUMBER OF COMPANION RE-HANG SAMPLE** _____
(B46POST1/B46REHG1 or B46POST2/B46REHG2)

*Provide the form number from Block 1 of the **Re-Hang** sample request form (FSIS Form 10,210-3) to be collected on the same shift from the same grow-out flock/house.*

**Questions and Answers Concerning the Nationwide Young Chicken
Microbiological Baseline Data Collection Program**

1. **Question: How would I determine if a plant had been selected to be included in the baseline data collection program?**

Answer: The list of plants selected for the baseline study is maintained by the Office of Public Health Science (OPHS), Washington, D.C. If an establishment is included in the baseline study, the IIC will receive a copy of the FSIS Notice and special sampling supplies, and individual sample requests will be identified in the PREP Schedule report for the program. You may also send an e-mail to the “Young Chicken Baseline Mailbox” in Outlook to inquire about the status of establishments.

2. **Question: Can a plant receive its individual results from samples collected during the Young Chicken Baseline Study (YCBS)?**

Answer: FSIS does not intend to report sample results back to plant management or inspection program personnel for these non-regulatory samples.

3. **Question: If the feet are not removed until after post-mortem inspection, are these birds eligible to be included in the YCBS?**

Answer: Yes, these birds are still eligible to be included in the baseline study. If the feet cannot be removed because of interference with the normal mechanical evisceration process, then the best alternative is to rinse the attached feet prior to sampling/rinsing the birds for the baseline study. A gentle, potable water rinse to remove any attached debris from the feet would be best. Rinse the feet only to approximately the level of the hock joint and avoid rinsing the main portion of the carcass. Then take the rinse sample from the carcass with the feet still attached.

4. **Question: Are birds that are labeled as “roasters,” but actually meet the definition of a young chicken, eligible for sampling?**

Answer: Yes, birds being processed that meet the regulatory definition of a broiler (even if the plant elects to label it as a roaster) qualify for this baseline study. If the poultry being processed meets the definition of a “roaster,” then we do not want to sample these birds. Inspection program personnel should monitor the process to be sure that only birds that are eligible for this baseline study are sampled. The definitions for poultry are found in 9 CFR 381.170.

(a)(1)(iii) Broiler or fryer. A broiler or fryer is a young chicken (usually under 13 weeks of age), of either sex, that is tender-meated with soft, pliable, smooth-textured skin and flexible breastbone cartilage.

(a)(1)(iv) Roaster or roasting chicken. A bird of this class is a young chicken (usually 3-5 months of age), of either sex, that is tender-meated with soft, pliable, smooth-textured skin and breastbone cartilage that may be somewhat less flexible than that of a broiler or fryer.

5. **Question: Which seal should go on the shipping container, the Re-Hang sample seal or the Post-Chill sample seal?**

Answer: Either of the supplied sample seals may be used to seal the shipping container as long as it matches one of the samples inside the shipping container. It is also acceptable to apply both sample seals to the shipping container if the container contains both types of samples.

6. **Question: Are inspection program personnel required to split or share the sample with the establishment?**

Answer: No, inspection program personnel are not required to split or share the sample with the establishment. Splitting/sharing the sample with the plant is not part of the baseline study design. However, if the plant is interested in doing their own study, they can certainly use their own supplies to collect a rinse from a different bird at approximately the same time the FSIS sample is collected.

7. **Question: What do I do if the establishment has been identified as being part of the YCBS but does not slaughter broilers?**

Answer: Check box 60 in block 33 of all sample request forms and return to the contract lab indicated in block 9.

8. **Question: In plants that use an automated process, with manual back-up, to transfer carcasses from the picking line to the evisceration line, where should FSIS collect their samples?**

Answer: It is recommended that you collect carcasses that are automatically transferred since the majority of birds are transferred in this manner. The carcasses collected for this baseline study should reflect the “typical” process in an establishment. The preferred collection point would be where you can safely obtain the carcass after the auto-transfer and after the feet have been removed.

9. **Question: In order to prevent accidental squeezing of feces from the carcass, would it be best to insert a clean foam plug into the vent prior to collecting the Re-Hang rinse sample?**

Answer: No, not unless the vent is normally plugged prior to **Re-Hang**. The carcasses collected for this baseline should reflect routine processes in an establishment.

10. Question: Should birds with head or feet still attached, slaughtered under religious exemption (for example, Halal), be sampled?

Answer: Birds that are slaughtered under 9 CFR 381.11 religious exemptions are NOT eligible for sampling in this baseline study because they do not bear the mark of inspection depicted in 9 CFR 381.96. Birds that receive the mark of inspection ARE eligible for sampling even if there is a special labeling claim concerning a religious authority.

11. Question: Can a plant be scheduled to sample for both PR/HACCP *Salmonella* performance standard verification sampling and the YCBS at the same time?

Answer: Yes, it is possible to be conducting both sampling projects concurrently.

12. Question: Should establishments that do split-carcass type processing prior to chilling be included in the baseline sampling?

Answer: If front and rear portions of the carcass are chilled separately, then the **Post-Chill** collection procedure will involve retrieving two carcass components. The inspector should collect a front half at **Post-Chill** at the approximate time a portion from the original grow-out flock/house would reach the end of the drip line. At the same time, collect a saddle in the same bag for simultaneous rinsing of both carcass portions. If necessary, due to loss of specific matching identity for both carcass portions, more than one *Name of Grower* can be entered in block 28.

13. Question: Should night shift personnel leave the birds in a bag in the refrigerator for day shift personnel to complete the sampling?

Answer: No, the bird itself should be rinsed and the RINSATE held overnight under refrigeration until shipped. This is a sampling procedure change for this baseline study only.

14. Question: What PBIS procedure code should be used when conducting the sampling?

Answer: The 05B02 Directed Sampling ISP code should be used.

15. Question: Why does the sample request form ask for the form number of the companion sample?

Answer: Two forms (B46REHG1 and B46POST1 or B46REHG2 and B46POST2) make up a sample pair. Since the form numbers for the two sample request forms are different, this information allows for matching the samples to confirm that both samples were collected as a pair.

16. Question: What are some common reasons for samples to be discarded?

Answer: All samples will be analyzed with the following exceptions:

- there is NOT enough rinsate (at least 200 ml are required);

- the temperature requirements are NOT met upon receipt at the contract laboratory;
- a copy and NOT an original form accompanies the samples;
- the original form has been altered (i.e., do not cross out anything on the sample form);
- sample containers other than those provided for this baseline study were used to submit samples;
- sample containers (bottles or bags) are leaking; and
- first shift samples not received the day following collection.

17. Question: How can I best ensure that temperature requirements are met?

Answer: Inspection program personnel should ensure the following:

- collected samples should be refrigerated as soon as possible after collection;
- make sure the rinsate samples have been cooled down prior to shipping;
- make sure the cold packs are completely frozen;
- use sufficient coolant to maintain sample temperatures during shipment;
- pre-chill the shipping container; and
- pack the shipper as close to the expected FedEx pick-up time as possible.

18. Question: Should Block 21 (product temperature) on the sample request form (FSIS Form 10210-3) be filled in?

Answer: Leave Block 21 blank. FSIS Directive 10,210.1 states that, *If REQUESTED in the specific program instructions, enter the product temperature at the time the sample was collected.* Block 18 of the sample request form (10,210-3) for the YCBS indicates the blocks on the form that must be completed for this study (Blocks 19, 20, 28-32). Block 21 is NOT required for this study. Leaving this block blank will not affect the analysis of submitted samples.

19. Question: What should I do if the FedEx air bill is not included when I receive the sampling supplies?

Answer: Send an e-mail to the “Young Chicken Baseline Mailbox” to request a replacement FedEx air bill.

20. Question: Can I use sample boxes or rinsate other than the supplies sent to me for the YCBS?

Answer: No, inspection program personnel are to use only the supplies sent for the YCBS. Conversely, do not use the YCBS sampling supplies for any other directed sampling not related to the YCBS.

21. Question: What do I do if I have more than 400 ml of rinsate?

Answer: Only 400 ml of rinsate is required for the baseline study, so discard any

excess.

22. Question: How were poultry slaughter facilities selected to be included in the YCBS?

Answer: The OPHS Young Chicken Baseline working group determined that those Federal establishments (including Talmadge-Aiken plants) that slaughter a minimum of 100,000 young chickens per year would be eligible for selection to participate in this baseline study based on the established study criteria. Eligible establishments will receive multiple sample requests during the baseline study, with the frequency of sampling in an establishment based on production volume.

23. Question: How often might YCBS sample requests be received?

Answer: Samples might be requested up to three (3) times per month in the larger establishments, or only once every other month in smaller establishments.

24. Question: I have received the Sample Request Forms for selecting YCBS samples, but we have not received any supplies. How do I obtain the supplies needed?

Answer: The Sample Request Forms and the sampling supplies for this baseline study are sent separately. The Sample Request Forms are printed and distributed at the beginning of each month and may arrive 1 to 3 weeks before you receive the sampling supplies. The laboratory ships sampling supplies to the designated establishments the week prior to the scheduled sampling. If sampling forms have been received, and no supplies have arrived by the beginning of the designated sampling week, send an e-mail to the “Young Chicken Baseline Mailbox.”

25. Question: What are the Project ID codes for the Full Implementation Young Chicken Baseline Study?

Answer: The Project ID Codes for the Full Implementation Study are:

First Shift Pair	Second Shift Pair
B46REHG1 – Re-Hang Shift 1	B46REHG2 – Re-Hang Shift 2
B46POST1 – Post-Chill Shift 1	B46POST2 – Post-Chill Shift 2

26. Question: When should samples be collected and shipped?

Answer: Inspection program personnel are to follow the instructions in Section 9, Sample Shipment, of FSIS Directive 10,230.5, Attachment 1. **Samples are to be collected and shipped to the laboratory the same day when possible.**

First shift samples should be shipped the same day collected or else they will be discarded by the laboratory. First shift samples may be collected Monday through Friday.

Second shift samples should only be collected Monday through Thursday. Samples collected on the second shift and shipped to the contract laboratory the next day will not be discarded.

27. Question: Are the questions in Block 28 of FSIS Form 10,210-3 for the Full Implementation Young Chicken Baseline Study the same as the questions asked during the “shake down” period?

Answer: Some of the questions and instructions have changed. The questions have been modified to collect information on the antimicrobial treatments being used in the on-line reprocessing systems and any **Post-Chill** interventions being used.