tion, Stage I zoeae of C. septemspinosa can be distinguished from Stage I zoeae of C. franciscorum angustimana by the exopodites of the maxillipeds. The exopodites of the maxillipeds are jointed in Stage I zoeae of C. septemspinosa and are not jointed in Stage I zoeae of C. franciscorum angustimana. Also, the fifth pair of telson spines are distinctly shorter than the fourth or sixth pair in C. septemspinosa; whereas, in my Stage I zoeae of C. franciscorum angustimana, the fifth pair of telson spines are about equal in length to the fourth and sixth pairs.

The occurrence in later zoeal stages of functional exopodites on the first pair of pereopods but not on pereopodal pairs 2-5 has been used as a criterion for distinguishing larvae of the genus *Crangon* from larvae of other genera of the family Crangonidae (Williamson 1960).

I found buds of exopodites on both the first and second pair of pereopods in Stage I zoeae of C. franciscorum angustimana. Assuming zoeae of C. franciscorum angustimana undergo typical development for crangonid larvae, these buds will become functional exopodites at Stage III or IV (Needler 1941; Kurata 1964; Makarov 1967). The criterion of the absence of exopodites on the second pair of pereopods for distinguishing larvae of Crangon from other genera of the Crangonidae. therefore, is invalid for the North Pacific Ocean. Unfortunately, larvae are described for only a few species of crangonids from the North Pacific Ocean, including the genus Crangon, and confirmation of the generic characteristics of the larvae is needed.

Acknowledgment

I thank Terry Butler, Pacific Biological Station, Nanaimo, British Columbia, Canada, for identifying the ovigerous females.

Literature Cited

HAYNES, E.

1976. Description of zoeae of coonstripe shrimp, *Pandalus hypsinotus*, reared in the laboratory. Fish. Bull., U.S. 74:323-342.

IVANOV, B. G.

1971. The larvae of some eastern shrimps in relation to their taxonomic status. [In Russ., Engl. summ.] Zool. Zh. 50:657-665.

KURATA, H.

1964. Larvae of decapod Crustacea of Hokkaido. 4. Crangonidae and Glyphocrangonidae. [In Jpn., Engl. summ.] Bull. Hokkaido Reg. Fish. Res. Lab. 28:35-50. (Translated by Division of Foreign Fisheries, Natl. Mar. Fish. Serv., NOAA.)

LOVELAND, H. A., JR.

1968. Larval development under laboratory conditions of Crangon alaskensis Rathbun, (Crustacea:Decapoda). M.A. Thesis, Walla Walla Coll., Walla Walla, Wash., 22 p.

MAKAROV, R. R.

1967. Larvae of the shrimps and crabs of the west Kamtschatkan shelf and their distribution. (Translated from Russ. by B. Haigh.) Natl. Lending Libr. Sci. Technol., Boston Spa, Yorkshire, 199 p.

NEEDLER, A. B.

1938. The larval development of Pandalus stenolepis. J. Fish. Res. Board Can. 4:88-95.

1941. Larval stages of Crago septemspinosus Say. Trans. R. Can. Inst. 23:193-199.

PIKE, R. B., AND D. I. WILLIAMSON.

1961. The larvae of Spirontocaris and related genera (Decapoda, Hippolytidae). Crustaceana 2:187-208.

1964. The larvae of some species of Pandalidae (Decapoda). Crustaceana 6:265-284.

TESMER, C. A., AND A. C. BROAD.

1964. The larval development of Crangon septemspinosa (Say) (Crustacea:Decapoda). Ohio J. Sci. 64:239-250.

WILLIAMSON, D. I.

1960. Crustacea, Decapoda: Larvae VII. Caridea, Family Crangonidae, Stenopodidea. Cons. Int. Explor. Mer, Fiches Identification Zooplankton 90, 5 p.

EVAN HAYNES

Northwest and Alaska Fisheries Center Auke Bay Laboratory National Marine Fisheries Service, NOAA P.O. Box 155 Auke Bay, AK 99821

LENGTH-WEIGHT RELATIONSHIPS OF WESTERN ATLANTIC BLUEFIN TUNA, THUNNUS THYNNUS¹

The Atlantic bluefin tuna, Thunnus thynnus, is seasonally distributed over most of the North Atlantic Ocean from Newfoundland to Brazil and from Norway to the Canary Islands (Gibbs and Collette 1967). There has been a great reduction in the Atlantic-wide catch (including Mediterranean) from 38,500 metric tons (t) in 1964 to 12,500 t in 1973 (Miyake et al. 1974). Because of this, a number of studies have been made and are being continued in order to understand the reason for this decline (Parks 1977; Shingu and Hisada 1977).

¹Contribution Number SEFC 80-01M, Southeast Fisheries Center Miami Laboratory, National Marine Fisheries Service, NOAA, Miami, Fla.

Length-weight, length-to-length, and weight-to-weight relationships are necessary in population analyses for converting one measurement to another. In this paper I present the relationships of the following: round weight-straight fork length, round weight-dressed weight, and straight fork length-curved fork length.

During my review of bluefin tuna literature, I found a lack of information on size relationships. Mather and Schuck (1960) used a length-weight curve based on 778 bluefin tuna from Cape Cod to estimate length. They did not indicate, however, when these fish were collected. They did not give a regression formula for the length-weight relationship, but they did present a straight length-curved length relationship based on 185 measurements fitted by inspection. Rodriguez-Roda (1964, 1971) collected 793 bluefin tuna and then determined the length-weight relationship. Of these, 467 bluefin tuna (prespawning) were entering the Mediterranean during May and June and 326 bluefin tuna (postspawning) were leaving the Mediterranean during July and August 1956, 1958, 1959, and 1961. Butler (1971) determined the length-weight relationship by the standard least squares regression method for 237 giant bluefin tuna caught during July through September 1966 from Conception Bay, Newfoundland. Mather et al. (1974) presented regression equations for converting from weight to length for bluefin tuna from Newfoundland, Libya, and the Bahamas from data supplied by the Fisheries Research Board of Canada, the International Council for the Exploration of the Sea, and the Woods Hole Oceanographic Institution. They also presented an equation for converting dressed weight to round weight. The method of determining the equations, the sample sizes, and time period sampled were not presented. Coan (1976) gave a length, weight, and age conversion table for bluefin tuna of both sexes. He converted length to weight based on a length-weight regression given in Sakagawa and Coan (1974), who had in turn, obtained this regression from Frank J. Mather. Woods Hole Oceanographic Institution. Unfortunately, there was no mention of sample size, location, or date.

Methods

Bluefin tuna length and weight measurements were collected during 1974 through 1977 from various landing points and processing plants along the east coast of the United States from Florida to Maine and from the Bahamas. These fish had been caught by purse seine, rod and reel, handline, and harpoon. Straight fork length (centimeters) was measured by caliper, and curved fork length (centimeters) was measured along the body contour by tape. Round weight (total weight of fish when caught) and dressed weight (head, viscera, and tail removed) were recorded in pounds and later converted to kilograms.

Ricker (1973) showed that the geometric mean (GM) regression can be used for a majority of biological situations as a reasonable and consistent estimate of the functional slope because most of the variability is natural.

The functional (GM) regression was calculated for the logarithmic transformation of the length-weight relationship for 3,578 bluefin tuna taken from May through October. The GM regression was also calculated for the relationship between round weight and dressed weight for 685 bluefin tuna taken from July through September, and for the straight fork length to curved fork length relationship for 606 bluefin tuna taken from July through October.

The general equation for the GM regression as given by Ricker is: Y = u + vX, with variables X and Y, and u is the y-axis intercept, where $u = \overline{Y} - v\overline{X}$, v is the slope, and $v = [\Sigma y^2/\Sigma x^2]^{\frac{1}{2}}$, where $y_i = Y_i - \overline{Y}$ and $x_i = X_i - \overline{X}$. The limits on all Σ are $i = 1, \ldots, n$.

The standard error of the slope was computed for each regression equation using the following equation from Ricker (1973): $S_v = [S_{yx}^2/\Sigma x^2]^{\frac{1}{2}}$, where S_v is the standard error of the slope and S_{yx}^2 is the mean square or variance of the observations from the regression line in the vertical direction.

Results and Discussion

Based on the classification system of Rivas and Mather (in press), the fish sampled mainly consisted of two size categories, giant bluefin tuna (>180 cm straight fork length and 130 kg round weight) and small bluefin tuna (<130 cm straight fork length or <45 kg round weight). Based on previous growth studies by Mather and Schuck (1960), the giant fish are probably age 9 and older and the small bluefin tuna are most likely age 4 or younger. Very few medium bluefin tuna (130-180 cm straight fork length and 45-130 kg round weight) probably ages 5 through 8 were sampled.

The functional (GM) regressions for straight fork length-round weight (log transformation), round weight-dressed weight, and straight fork length-curved fork length are presented in Table 1. All of these relationships were characterized by high correlation coefficients. The data points are plotted with regression lines in Figures 1-3. The data points show that the GM regression model fits the data reasonably well for the size ranges studied. Extrapolation beyond the size range of observations may yield erroneous predictions. Regression statistics for each relationship are presented in Table 2.

The use of logarithmic transformations may lead to bias in data estimates (Pienaar and Thomson 1969; Beauchamp and Olson 1973; Lenarz 1974). However, since the mean square error for the round weight-straight fork length logarithmic transformation is low (Table 2), the bias in the data estimate was found to be minimal (1%).

Previous publications have not included standard errors or confidence limits or statistics necessary for their estimation. Therefore, comparisons with my data could not be made. To compare results from my study with studies by other authors, I compared estimates of Y using both their regression equations and mine. Whenever possible, I

TABLE 1.—Functional (GM) regression equation and correlation coefficient for the relationships between round weight (Y) and straight fork length (X), round weight (Y) and dressed weight (X), and straight fork length (Y) and curved fork length (X) for western Atlantic bluefin tuna. Weights in kilograms and lengths in centimeters.

Geometric mean regression equation			
Log ₁₀ round weight - log ₁₀ straight fork length:			
$\log_{10} Y = -4.52307 + 2.91920 \log_{10} X$	0.997		
Round weight - dressed weight:			
Y = -7.92240 + 1.29607 X	0.935		
Straight fork length - curved fork length:			
Y = -2.06971 + 0.963300 X	0.892		

selected X values at each end of their range of values that corresponded with my range of values. I also compared estimates of Y for an X value taken at the middle of their size range.

My estimates of round weight from straight fork length using the functional (GM) regression agreed most closely with my estimates obtained using the regression equation of Sakagawa and Coan (1974), with the greatest difference in estimates of only 2% occurring for a 270 cm fork length (FL) bluefin tuna. My calculated functional regression estimates next most closely agreed with estimates obtained using the length-weight relationship of Butler (1971), with the largest difference of 6% occurring at 250 cm FL. My esti-

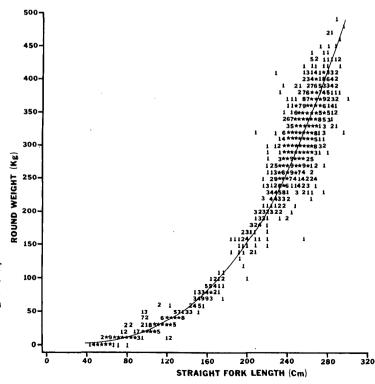


FIGURE 1.—Functional (GM) regression of round weight on straight fork length for 3,578 western Atlantic bluefin tuna 1974-77. (Number of fish indicated, star signifies number >9.)

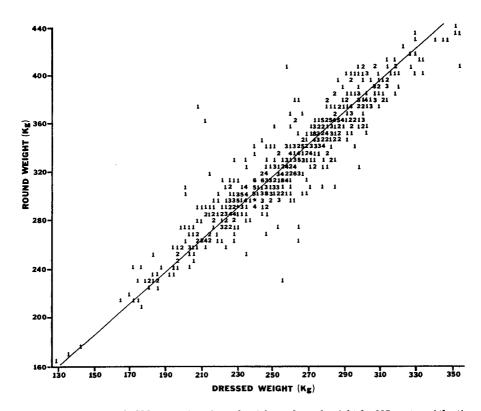


FIGURE 2.—Functional (GM) regression of round weight on dressed weight for 685 western Atlantic bluefin tuna 1974-77. (Number of fish indicated, star signifies number >9.)

TABLE 2.—Regression statistics for \log_{10} round weight $(Y) - \log_{10}$ straight fork length (X), round weight (Y) - dressed weight(X), and straight fork length (Y) - curved fork length(X) of western Atlantic bluefin tuna. Weights in kilograms and lengths in centimeters.

$\overline{\overline{X}}$	$\overline{\overline{Y}}$	$\sum x^2$	$\sum y^2$	Σχγ	S _{VX} ²	Sv
	Log					· ·-
2.19254	1.87739	222.745	1,898.17	648.054	0.00356051	0.00399809
		Round weight	- dressed weig	ht		
256.993	325.158	832,635	1,398,650	1,009,090	257.261	0.0175776
		Straight fork length	- curved fork	length		
271.477	259.444	120,979	112,262	103,959	37.9615	0.0177140
	2.19254 256.993	2.19254 1.87739 256.993 325.158	Log ₁₀ round weight 2.19254 1.87739 222.745 Round weight 256.993 325.158 832,635 Straight fork length	Log ₁₀ round weight - log ₁₀ straight fc 2.19254 1.87739 222.745 1,898.17 Round weight - dressed weig 256.993 325.158 832,635 1,398,650 Straight fork length - curved fork	Log ₁₀ round weight — log ₁₀ straight fork length 2.19254 1.87739 222.745 1,898.17 648.054 Round weight — dressed weight 256.993 325.158 832,635 1,398,650 1,009,090 Straight fork length — curved fork length	Log ₁₀ round weight - log ₁₀ straight fork length 2.19254 1.87739 222.745 1,898.17 648.054 0.00356051 Round weight - dressed weight 256.993 325.158 832,635 1,398,650 1,009,090 257.261 Straight fork length - curved fork length

mates of weight from length differed most from estimates which I calculated using equations of Rodriguez-Roda (1964, 1971). The largest variation (12%) was found for a prespawning fish measuring 48 cm.

No size range was reported by Mather et al. (1974) for estimating length from weight. However, estimated length corresponding to the extremes and middle of the size range in weight I studied agree closely to values I calculated using their regression equation for Newfoundland, with the greatest difference being only 3% for a 5 kg fish. A greater difference (13%) was noted when comparing estimates from my functional (GM) re-

gression with estimates obtained using their regression equation for the Bahamas for a 5 kg fish. This large difference may have resulted from their not including fish in this size range when calculating their equation because differences at the middle and upper end of my size range were small, 4% or less. There appears to be a typographical error in the equation these authors gave for bluefin tuna from Libya, so no comparison was made.

My functional (GM) regression estimates of round weight from dressed weight agree well with the estimates I obtained using the regression equation of Mather et al. (1974). The largest dif-

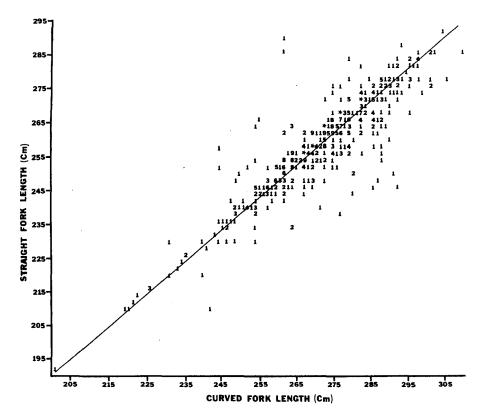


FIGURE 3.—Functional (GM) regression of straight fork length on curved fork length for 606 western Atlantic bluefin tuna 1974-77. (Number of fish indicated, star signifies number >9.)

ference I found was 3% for a 130 cm bluefin tuna. Again I used my range of values for dressed weight since the range was not given by these authors.

My functional (GM) regression estimates of straight fork length from curved fork length agree very closely over my entire size range with estimates I obtained using the regression equation given by Mather and Schuck (1960). The largest difference I found, only 1%, occurred at the lower end of my range of curved fork length values of 200 cm.

Acknowledgments

I thank Gary Sakagawa of the Southwest Fisheries Center, National Marine Fisheries Service (NMFS), NOAA; Emma Henderson of the Northeast Fisheries Center, NMFS, NOAA; Frederick Berry, Raymon Conser, Mark Farber, Michael Parrack, William Richards, Luis Rivas, James Tyler, and James Zuboy of the Southeast Fisheries Center, NMFS, NOAA; and Arvind Khilnani of Stanford University for their helpful comments on the manuscript. I also thank Tom Chewning of the Southeast Fisheries Center for computer programming assistance.

Literature Cited

BEAUCHAMP, J. J., AND J. S. OLSON.

 Corrections for bias in regression estimates after logarithmic transformation. Ecology 54:1403-1407.

BUTLER, M.

1971. Biological investigation on aspects of the life history of the bluefin tuna 1970-1971. Newfoundland and Labrador Tourist Development Office, St. Johns, 169 p.

COAN, A. L.

1976. Length, weight and age conversion tables for Atlantic tunas. Int. Comm. Conserv. Atl. Tunas, Collect. Vol. Sci. Pap. 5 (SCRS-1975):64-66.

GIBBS, R. H., JR., AND B. B. COLLETTE.

1967. Comparative anatomy and systematics of the tunas, genus *Thunnus*. U.S. Fish Wildl. Serv., Fish. Bull. 66:65-130.

LENARZ, W. H.

 Length-weight relations for five eastern tropical Atlantic scombrids. Fish. Bull., U.S. 72:848-851.

MATHER, F. J., III, J. M. MASON, AND A. C. JONES. 1974. Distribution, fisheries and life history data relevant to identification of Atlantic bluefin tuna stocks. Int. Comm. Conserv. Atl. Tunas, Collect. Vol. Sci. Pap. 2 (SCRS-1973):234-258.

MATHER, F. J., III, AND H. A. SCHUCK.

 Growth of bluefin tuna of the western North Atlantic. U.S. Fish Wildl. Serv., Fish. Bull. 61:39-52.

MIYAKE, M. P., A. A. DEBOISSET, AND S. MANNING (compilers). 1974. International Commission for the Conservation of Atlantic Tunas. Stat. Bull. 5, Unnumbered pages. PARKS. W. W.

1977. Cohort and equilibrium yield-per-recruit analyses for the Atlantic bluefin tuna fisheries system accounting two system configurations and two natural mortality models. Int. Comm. Conserv. Atl. Tunas, Collect. Vol. Sci. Pap. 6 (SCRS-1976):385-401.

PIENAAR, L. V., AND J. A. THOMSON.

1969. Allometric weight-length regression model. J. Fish. Res. Board Can. 26:123-131.

RICKER, W. E.

 Linear regressions in fishery research. J. Fish. Res. Board. Can. 30:409-434.

RIVAS, L. R., AND F. J. MATHER III.

In press. Proposed terminology for size groups of the north Atlantic bluefin tuna *Thunnus thynnus*. Int. Comm. Conserv. Atl. Tunas, Collect. Vol. Sci. Pap. (SCRS-1978). RODRIGUEZ-RODA, J.

1964. Biología del Atún, Thunnus thynnus (L.), de la costa sudatlántica de España. Invest. Pesq. 25:33-146.

1971. Investigations of tuna (Thunnus thynnus) in Spain. In Report for biennial period 1970-71, Part II, p. 110-113. Int. Comm. Conserv. Atl. Tunas.

SAKAGAWA, G. T., AND A. L. COAN.

1974. A review of some aspects of the bluefin tuna Thunnus thynnus thynnus fisheries of the Atlantic Ocean. Int. Comm. Conserv. Atl. Tunas, Collect. Vol. Sci. Pap. 2 (SCRS-1973):259-313.

SHINGU, C., AND K. HISADA.

1977. A review of the Japanese Atlantic longline fishery for bluefin tuna and the consideration of the present status of the stocks. Int. Comm. Conserv. Atl. Tunas, Collect. Vol. Sci. Pap 6 (SCRS-1976):366-384.

RAYMOND E. BAGLIN, JR.

Southeast Fisheries Center Miami Laboratory National Marine Fisheries Service, NOAA 75 Virginia Beach Drive, Miami, Fl 33149 Present address: National Marine Fisheries Service, NOAA c/o Alaska Department of Fish and Game P.O. Box 686, Kodiak, AK 99615

DEVELOPMENTAL ANATOMY AND INFLATION OF THE GAS BLADDER IN STRIPED BASS, MORONE SAXATILIS

In 1974, a percentage of striped bass, *Morone saxatilis*, fingerlings reared at the Cooperative Fishery Research Laboratory, Southern Illinois University, lacked an inflated gas bladder. The purpose of this study was to describe the de-

velopmental anatomy of the gas bladder and its associated structures in striped bass so that a better understanding of the inflation mechanism could be obtained.

With regard to gas bladder morphology, bony fishes are classified as physostomes or physoclists. Generally, the more ancient, soft-rayed fishes (Malacopterygii) are physostomous, while the more modern, spiny-rayed fishes (Acanthoptervgii) are physoclistic (Lagler et al. 1962). A physotome possesses a hollow connection, the pneumatic duct, between the gut and the gas bladder throughout its entire life. Some physotomes gulp surface air through the pneumatic duct to initiate inflation of the gas bladder (Tait 1960). Fish that are physoclistic do not possess this open connection as adults. Some physoclists, however, do possess a pneumatic duct as larvae, but the duct atrophies prior to adulthood. Günther's (1880) examinations have shown that adult striped bass are physoclistic. Doroshev and Cornacchia (1979) give a partial description of the development of the gas bladder in striped bass.

Several theories have been advanced to explain how the gas bladder is initially inflated in fishes that do not gulp surface air or are physoclistic prior to initial inflation. Some of these theories include: gases produced by the disintegration of organic materials (Powers 1932); production of gasses as a result of digestion (Johnston 1953); vacuolation of the gas bladder epithelia (McEwen 1940); and functioning of a rete mirabile, or gas gland (Schwarz 1971).

Methods

Histomorphological Studies

Striped bass larvae were obtained from the Hudson River, N.Y., and Lake Charles, La. Upon arrival, the 1- to 4-day-old larvae were transferred into 200 l aquaria and maintained at 16°-18° C. Brine shrimp, Artemia salina, were fed regularly to the larvae. Eighty-three striped bass larvae 4.3-24 days old (from the time of hatching) were removed from the aquaria and prepared for histological study. The larvae were fixed in either 10% Formalin¹ or Bouin's fluid, dehydrated in a series of graded alcohols, cleared in benzene, and embedded in Carbowax. From a representative

¹Reference to trade names does not imply endorsement by the National Marine Fisheries Service, NOAA.