

Fumonisin B₁ Carcinogenicity in a Two-Year Feeding Study Using F344 Rats and B6C3F₁ Mice

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Fumonisin B₁ (FB₁) is a mycotoxin isolated from *Fusarium* fungi that contaminate crops worldwide. A previous study demonstrated that FB₁ promoted preneoplastic foci in initiated rats and induced hepatocellular carcinomas in BD IX rats at 50 parts per million (ppm), but fundamental dose–response data were not available to assist in setting regulatory guidelines for this mycotoxin. To provide this information, female and male F344/N/Nctr BR rats and B6C3F₁/Nctr BR mice were fed for two years a powdered NIH-31 diet containing the following concentrations of FB₁: female rats, 0, 5, 15, 50, and 100 ppm; male rats, 0, 5, 15, 50, and 150 ppm; female mice, 0, 5, 15, 50, and 80 ppm; male mice, 0, 5, 15, 80, and 150 ppm. FB₁ was not tumorigenic in female F344 rats with doses as high as 100 ppm. Including FB₁ in the diets of male rats induced renal tubule adenomas and carcinomas in 0/48, 0/40, 9/48, and 15/48 rats at 0, 5, 15, 50, and 150 ppm, respectively. Including up to 150 ppm FB₁ in the diet of male mice did not affect tumor incidence. Hepatocellular adenomas and carcinomas were induced by FB₁ in the female mice, occurring in 5/47, 3/48, 1/48, 19/47, and 39/45 female mice that consumed diets containing 0, 5, 15, 50, and 80 ppm FB₁, respectively. This study demonstrates that FB₁ is a rodent carcinogen that induces renal tubule tumors in male F344 rats and hepatic tumors in female B6C3F₁ mice. **Key words:** fumonisin B₁, hepatocarcinogenicity, renal carcinoma, rodent bioassay. — *Environ Health Perspect* 109(suppl 2):277–282 (2001).

<http://ehpnet1.niehs.nih.gov/docs/2001/suppl-2/277-282howard/abstract.html>

Fumonisin is a group of hydrophilic mycotoxins produced by fungi of the *Fusarium* species. Fumonisin B₁ (FB₁) was the first fumonisin identified, and this led to the discovery of several different homologues (1–4). The *Fusarium* fungi have been shown to contaminate crops (primarily corn or maize) worldwide (5), and the severity of the infection depends on environmental conditions such as drought and heat. *Fusarium moniliforme* Sheldon (= *F. verticillioides*) is considered the dominant species of *Fusarium* on crops that produces FB₁, although other species have been shown to produce FB₁ and the other fumonisins in culture (6–13). The predominant fumonisin homologues (B₁, B₂, B₃) differ on the basis of hydroxyls at positions C5 and C10 (1–3).

The discovery of FB₁ was the result of long-term investigations into the reasons for regiospecificity in the occurrence of esophageal cancer in South Africa. Areas in the Transkei that are only 150 km apart have esophageal cancer incidences that differ 2- to 3-fold (14,15). Cytologic analysis of esophageal scrapings from patients in these areas showed that cytologically abnormal esophageal mucosal cells were present in patients from the high-risk area (16). Furthermore, investigators demonstrated a higher incidence of *F. moniliforme* in maize from households in the high-risk area than in the low-risk area of the Transkei (16,17).

These field investigations led to the isolation from household corn of several isolates of *F. moniliforme* (18–20). Including isolate *F. moniliforme* MRC 826 in the diet of BD IX rats for 22–27 months led to formation of esophageal hyperplasia, forestomach papillomas and carcinomas, hepatocellular carcinomas, and cholangiocarcinomas (21).

FB₁ was isolated as the compound present in cultures of *F. moniliforme* MRC 826 that promoted the formation of preneoplastic altered enzyme foci in the livers of dimethylnitrosamine-initiated rats (22). In a subsequent study, including FB₁ at 50 mg/kg in the diet of BD IX rats led to development of liver tumors (23). Hepatic regenerative nodules, cholangiofibrosis, and cirrhosis developed in all 10 rats maintained on the FB₁ for 20–26 months, whereas only 7 of 10 rats developed hepatocellular carcinomas (23). Although this study was limited to a single dose of FB₁ in the diet and included only a small number of rats, when combined with the tumor promotion studies (22) it strongly suggested that FB₁ was a rodent carcinogen.

FB₁ was nominated for tumorigenesis testing under the auspices of the National Toxicology Program because of the carcinogenesis of FB₁ and *F. verticillioides* MRC 826 in rats, the presence of FB₁ in maize in areas of the world with high incidences of esophageal cancer (5) attributed to the presence of FB₁ in food destined for human consumption

(maize), and because of the lack of fundamental dose–response data available to assist with setting regulatory levels.

Materials and Methods

Study Material and Feed

FB₁ was produced by aqueous cultures of *F. proliferatum* on corn. The FB₁ was extracted from the autoclaved material using methanol and purified as the ammonium salt using high-performance liquid chromatography (HPLC). The purity of the FB₁ was established as > 96% using ¹H and ¹³C nuclear magnetic resonance spectroscopy, mass spectrometry, and HPLC with evaporative light scattering detection (24).

Autoclaved-powdered NIH-31 rodent feed (Purina Corp., St. Louis, MO) was the test diet in the study, and FB₁ was added as a water-based component using a Patterson-Kelley V-blender (Patterson-Kelley Co., East Stroudsburg, PA). The FB₁ content of the control diet was below 0.06 parts per million (ppm).

Animals and Housing

Female and male Fischer 344/N/Nctr BR rats and B6C3F₁/Nctr BR mice were obtained from the National Center for Toxicological Research (NCTR) breeding colony at 4 weeks postpartum. The rats and mice were allocated to the study dose groups in a random manner that controlled for weight bias and minimized occurrence of littermates in the same dose

This article is based on a presentation at the International Conference on the Toxicology of Fumonisin held 28–30 June 1999 in Arlington, Virginia, USA.

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This work was supported by Interagency Agreement IAG 224-93-0001 between the U.S. Food and Drug Administration and the National Institute for Environmental Health Sciences. The contents of this manuscript do not necessarily reflect the views and policies of the U.S. Food and Drug Administration or the National Institute for Environmental Health Sciences. Also, the mention of trade names or commercial products does not constitute endorsement or recommendation for their use.

Received 10 May 2000; accepted 16 August 2000.

group. The rats were housed two per cage and the mice four per cage in polycarbonate cages with autoclaved hardwood chip bedding. Cagemates were identified using an ear-clip identification system. Water was available *ad libitum*. The powdered feed was available *ad libitum* in custom feeders designed for powdered feed. The cages and water were changed twice weekly for the rats and weekly for the mice.

Study Design

The doses of FB₁ for the rats were based on toxicities detected in 28-day and 90-day subchronic studies (24–26). The doses for the bioassay were 0, 5, 15, 50, and 100 ppm FB₁ for female F344 rats and 0, 5, 15, 50, and 150 ppm FB₁ for the male F344 rats.

The doses of FB₁ for the B6C3F₁ mice were based on the response of the mice in 28- and 90-day subchronic studies (24,25). The FB₁ doses used with the female mice were 0, 5, 15, 50, and 80 ppm FB₁, whereas the doses used in the diets for the male mice were 0, 5, 15, 80, and 150 ppm FB₁.

The study was conducted in accordance with the guidelines of the National Toxicology Program (27) and the U.S. Food and Drug Administration (28). Animals were allocated 48 for each sex per dose group except for the groups receiving diets containing 5 ppm FB₁, in which there were 40 rats of each sex. The mice and rats were acclimated to the cage and control powdered feed until 6 weeks postpartum, when the dosed feed was added.

Animals were necropsied after 104 weeks of consumption of the dosed feed or upon removal as moribund or dead. The livers and kidneys from all the animals were examined microscopically, whereas other tissues were examined only for animals receiving control or high-dose diets (24). Pathologic examinations were conducted as described for the National Toxicology Program (27).

Statistical Analysis

Tests of pairwise comparisons of the neoplastic and non-neoplastic lesions for each exposed group with the controls was conducted using the Poly-k test (24,29,30). A k-value of 3 was used in these analyses (30). The analysis includes a risk–weight adjustment on animals that died before completion of the study and reports an adjusted rate of lesion incidence.

Results

The female F344 rats that consumed diets containing 100 ppm FB₁ had decreased weights compared to the body weights of the female F344 rats on the control diets (Figure 1A). No body weight differences were detected in the female F344 rats that

consumed diets containing 5, 15, and 50 ppm FB₁ compared to the female F344 rats on the control diet (Figure 1A). The daily mean consumption of diet by the male and female rats during the course of the study was indistinguishable from the dietary consumption of other F344/N/Nctr BR rats at this facility (data not shown). The mean consumption rates of compound in the female rats between weeks 51 and 104 on the dosed feed were 0, 0.27, 0.78, 2.57, and 5.24 mg FB₁ per kg body weight per day (mg/kg bw/day) for 0, 5, 15, 50, and 100 ppm diets, respectively. There were no FB₁-dependent changes in the body weights of the male F344 rats that consumed up to 150 ppm FB₁ (Figure 1B). The mean consumption rates of FB₁ between weeks 51 and 104 of the study were 0, 0.22, 0.67, 2.24, and 6.60 mg/kg bw/day for male rats consuming diets containing 0, 5, 15, 50, and 150 ppm FB₁.

There were no dose-related differences in the survival of the female F344 rats at 104 weeks (Figure 2A). The survival for female F344 rats on the control diet was 52%, whereas the survival of female F344 rats consuming the 100 ppm FB₁ diet was 60%. Consumption of the FB₁ dose used in the study did not induce toxicity that could be detected by changes in body weight throughout the 2-year study.

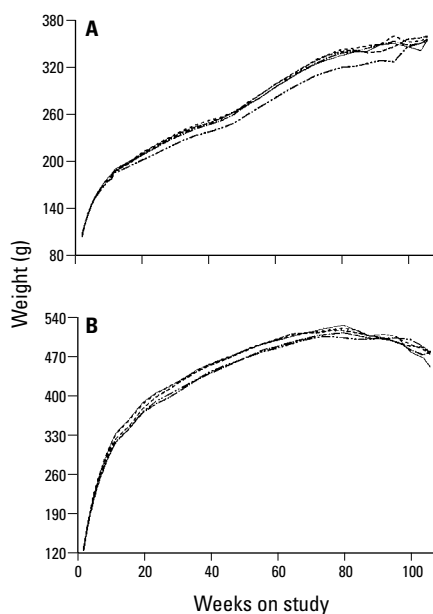


Figure 1. Body weights of female (A) and male (B) F344/N/Nctr BR rats in the 2-year tumor study. The body weights were measured weekly, and are shown at 4-week intervals starting at 20 weeks. The following FB₁ doses were given to the female F344 rats: control diets (—), 5 ppm (---), 15 ppm (- · -), 50 ppm (- · · -), and 100 ppm (- · · · -). The following FB₁ doses were given to the male F344 rats: control diets (—), 5 ppm (---), 15 ppm (- · -), 50 ppm (- · · -), and 150 ppm (- · · · -).

There were no dose-related differences in the survival rates of the male F344 rats, with 35% of the male F344 rats on the control diet and 52% of the male F344 rats consuming diets with 150 ppm FB₁ surviving for 2 years (Figure 2B).

Necropsy and microscopic evaluation of the tissues of the male F344 rat revealed an increase in renal tubule adenomas and carcinomas (Table 1). No tumors were present in the kidneys of the male F344 rats that consumed diets containing 0, 5, or 15 ppm FB₁. Renal tubule adenomas were present in 2 of the 48 male F344 rats consuming 50 ppm FB₁, and renal tubule carcinomas were present in 7 of the 48 rats on this dose. This produced an adjusted incidence of 25.7% for renal adenomas and carcinomas in the male rats at the 50 ppm FB₁ dose. The development of renal tubule adenomas and carcinomas was more pronounced in male F344 rats that consumed 150 ppm FB₁ (Table 1), with 5 of the 48 rats developing adenomas and 10 of the 48 rats developing carcinomas, for an adjusted incidence of 38.1% of the rats at the 150 ppm FB₁ dose with adenomas or carcinomas. These increases in adenomas at 150 ppm, carcinoma at 50 and 150 ppm, and adenoma or carcinoma at 50 and 150 ppm were significant for the control group (Table 1).

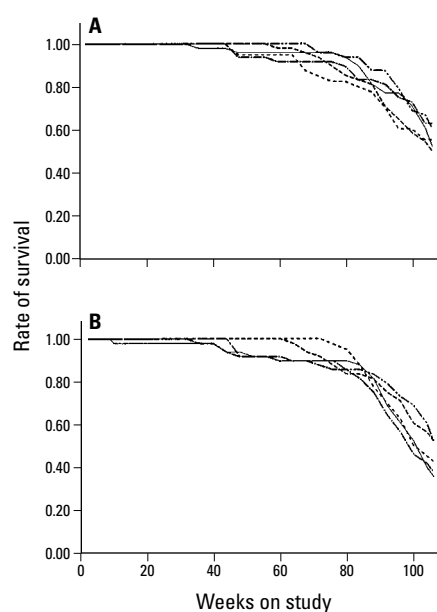


Figure 2. Survival of female (A) and male (B) F344/N/Nctr BR rats in the 2-year tumor study. Survival rates are expressed as the fraction of rats initially loaded into the study that survived to the end of the indicated week. The following FB₁ doses were given to the female F344 rats: control diets (—), 5 ppm (---), 15 ppm (- · -), 50 ppm (- · · -), and 100 ppm (- · · · -). The following FB₁ doses were given to the male F344 rats: control diets (—), 5 ppm (---), 15 ppm (- · -), 50 ppm (- · · -), and 150 ppm (- · · · -).

Among the female F344 rats, there were no FB₁-dependent changes in the incidence of tumors. One renal adenoma was detected in a female F344 rat consuming 50 ppm FB₁, and one renal tubule carcinoma was detected in a female F344 rat that consumed 100 ppm FB₁; however, from the Poly-k analysis, the low frequency of these tumors did not indicate a dose-related trend.

The renal tubule adenomas were characterized by a defined focus of expansive tubule cells. The nuclear and cell volumes were increased in adenoma cells compared to normal adjacent cells. The cytoplasm of the adenoma cells stained clear to basophilic. A representative adenoma is shown in Figure 3.

The renal tubule carcinomas were characterized as growths of abnormal and atypical cells that compressed and invaded neighboring normal tissue (Figure 4). The cells within the growing boundary of the carcinoma contained basophilic cytoplasm with typically increased volume and hyperchromatic nuclei. Necrosis was evident within the interior of the larger carcinomas. In many of the carcinomas, small renal tubule-like structures were evident.

These renal tubule carcinomas metastasized to the lung and lymphatic tissues.

We detected no differences in the body weights of the female B6C3F₁ mice (Figure 5A) or in the consumption of the diets containing FB₁ (data not shown) when compared to those of mice receiving control diets. The mean daily consumption of FB₁ between weeks 51 and 104 of the study were 0, 0.65, 1.91, 6.62, and 12.76 mg/kg bw/day for the groups receiving 0, 5, 15, 50 and 80 ppm FB₁, respectively. Similarly, the body weights of the male B6C3F₁ mice consuming diets containing FB₁ did not differ from the body weights of the male B6C3F₁ mice on control diets (Figure 5B). The mean daily consumption of FB₁ in the male mice receiving 0, 5, 15, 50, and 150 ppm FB₁ was 0, 0.53, 1.55, 9.04, and 15.41 mg/kg bw/day, respectively, between weeks 51 and 104 of the study. The body weights of the male mice were approximately 15% less than the body weights of B6C3F₁/Nctr BR mice in other studies conducted at NCTR, whereas the body weights of the female mice were 30% less than expected. Analysis of the feed consumption

rates indicated that the mice were consuming approximately 30% less feed than mice in other studies at NCTR. The lower consumption of feed apparently was not caused by palatability, because feed consumption was reduced in the control groups. Availability of the feed through the screen feeders was reduced, although particle size analysis of the feed did not indicate that the powdered feed was altered by the addition of FB₁.

Survival of the female B6C3F₁ mice consuming 80 ppm FB₁ decreased compared to survival of mice consuming control diets or diets containing 0–50 ppm FB₁ (Figure 6A). This decrease in survival started at approximately 1 year of age and continued until the end of the study. Female B6C3F₁ mice consuming the other FB₁ diets had survival rates indistinguishable from those of female mice consuming control diets (Figure 6A). Exposure to FB₁ had no effect on survival of male B6C3F₁ mice at any of the doses (Figure 6B).

Hepatocellular adenomas were present in 11.7% of the female B6C3F₁ mice given control diet for 2 years (Table 2). Hepatocellular adenomas were present at adjusted rates of 6.5 and 2.1% at 5 and 15 ppm FB₁; these values were not statistically significantly different from the incidence in the control group. In females consuming 50 and 80 ppm FB₁, the adjusted rates for the incidence of adenoma increased to 36.3 and 73.7%, respectively. Hepatocellular carcinomas were not present in female B6C3F₁ mice given 0, 5, or 15 ppm FB₁. Hepatocellular carcinomas were present at adjusted rates of 22.5 and 23% among female B6C3F₁ mice that consumed 50 and 80 ppm FB₁, respectively (Table 2). Consumption of FB₁ increased the adjusted rate of incidence of hepatocellular adenomas and carcinomas from 11.7% of the

Table 1. Incidences of neoplastic and non-neoplastic lesions in male rat kidneys.

Type of lesion	Number of rats examined				
	48	40	48	48	48
Renal tubule epithelial hyperplasia	2	1	4	14 ^a	8 ^a
Renal tubule adenoma	0	0	0	2	5
Adjusted rate (percent) ^b	0	0	0	5.7	12.9
Significance ^b	–	–	–	<i>p</i> = 0.2293	<i>p</i> = 0.0314
Renal tubule carcinoma	0	0	0	7	10
Adjusted rate (percent)	0	0	0	20.0	25.4
Significance	–	–	–	<i>p</i> = 0.0059	<i>p</i> = 0.0008
Renal tubule adenoma or carcinoma	0	0	0	9	15
Adjusted rate (percent)	0	0	0	25.7	38.1
Significance	–	–	–	<i>p</i> = 0.0011	<i>p</i> = 0.0001

^aSignificantly different from control group (*p* < 0.05) using the Poly-k test described in "Methods." ^bThe adjusted incidence rate and significance were determined using the Poly-k test.

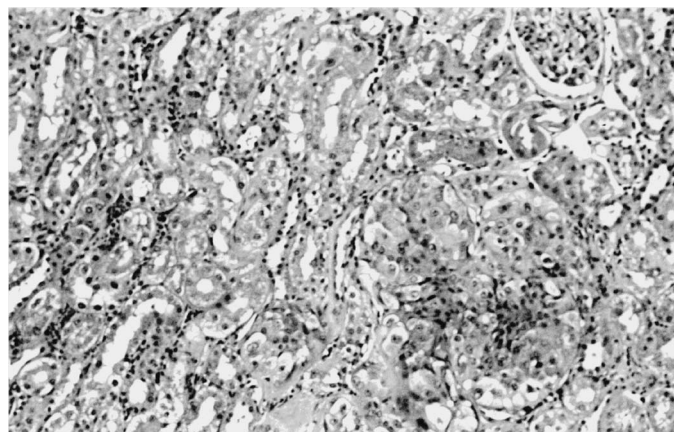


Figure 3. Photomicrograph of renal tubule adenoma from a male F344/N/Nctr BR rat that consumed a diet containing 150 ppm FB₁. The adenoma is present in the lower left side of the photomicrograph as a focus of cells that disrupted normal tubule organization.

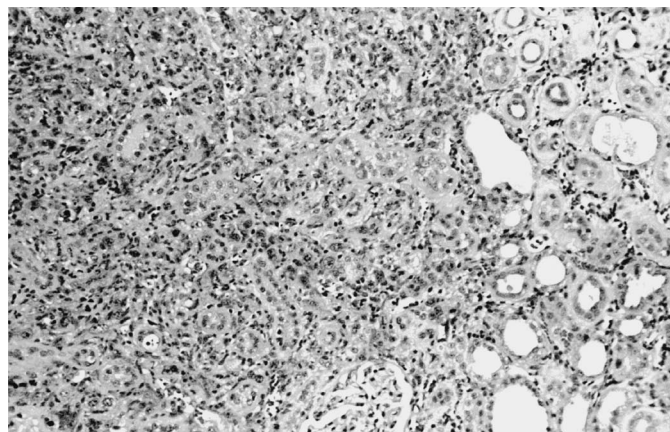


Figure 4. Photomicrograph of a renal tubule carcinoma from a male F344/N/Nctr BR rat that consumed a diet containing 150 ppm FB₁. The normal kidney is present on the right side of the photomicrograph where renal tubules are present. The carcinoma (left side of photomicrograph) contains cells of atypical morphology, although some tubule-like organization is found within the tumor.

female B6C3F₁ mice on control diets to 6.5, 2.1, 42.7, and 88.3% of mice consuming diets that contained 5, 15, 50, and 80 ppm FB₁, respectively (Table 2). The increased rates of adenomas or carcinomas at 50 and 80 ppm FB₁ were statistically significantly different from those of the control group.

The hepatocellular adenomas were characterized by distinct foci of cells that were either eosinophilic or basophilic and that routinely compressed the adjacent normal parenchymal cells (Figure 7). The carcinomas were characterized by poorly differentiated and anaplastic cells within the liver (Figure 8).

Among male B6C3F₁ mice, exposure to FB₁ did not affect the incidence of neoplasia of any type, including in the liver, where

approximately 25% of the mice had hepatocellular adenomas or carcinomas (Table 2).

The increase in numbers of hepatocellular adenomas or carcinomas in female mice was accompanied by increases in hepatocellular hypertrophy at 50 and 80 ppm FB₁ (Table 2). Although hypertrophy correlated with tumor incidence in female mice, it was present in the livers of male mice at 80 and 150 ppm (Table 2), but there was no increase in tumor incidence.

Discussion

These studies show that FB₁ is a renal carcinogen when included in the diets of male F344 rats. Renal tubule adenomas or carcinomas were absent in the male F344 rats consuming

control diets and diets containing 5 or 15 ppm FB₁ for 2 years (Table 1). Induction of renal tubule carcinomas was evident at doses of 50 and 150 ppm FB₁ (Table 1), with adjusted rates of 22.5 and 23.0%, respectively. This suggests that a no-observed-effect-level (NOEL) for the induction of renal tumors in male rats lies between 15 and 50 ppm FB₁. The highest dose of FB₁ included in the diets of the female F344 rats was 100 ppm; this did not increase tumor incidence.

In the present study, including FB₁ in doses as high as 150 ppm in the diets of male F344 rats and as high as 100 ppm in the diet of female F344 rats over the 2-year feeding period did not increase mortality compared to that of rats consuming control diets (Figure 2). Similarly, the body weights of the male and female rats consuming diets containing FB₁ did not decrease compared to the body weights of rats on the control diets (Figure 1). Therefore, we can conclude that the dietary levels of FB₁ that induced tumors in male F344 rats (15 ppm < NOEL ≤ 50 ppm) were not close to the maximum tolerated dose (MTD), as evidenced by an absence of effect on the growth and survival of the dosed rats.

In a feeding study using male BD IX rats, Gelderblom et al. (23) included 50 ppm FB₁ in the diet for up to 26 months. The diet contained 75% sifted white corn, and the purity of the FB₁ was reported as > 90% (23). The FB₁-fed rats developed hepatic regenerative nodules and cholangiofibrosis (synonymous with adenofibrosis), whereas rats on the control diets did not. The liver dysplasia progressed to hepatic cirrhosis and hepatocellular carcinomas in 10 of the 15 rats sacrificed between 18 and 26 months. In an additional study, FB₁ was fed to male BD IX rats at 0, 1, 10, and 25 ppm for 24 months (31). These levels of FB₁ failed to induce the hepatocellular carcinomas that were induced in the previous study with 50 ppm FB₁. As a result, the studies with male BD IX rats suggest that a NOEL exists between 25 and 50 ppm FB₁ for the formation of hepatocellular tumors in male BD IX rats. Renal tumors in male rats

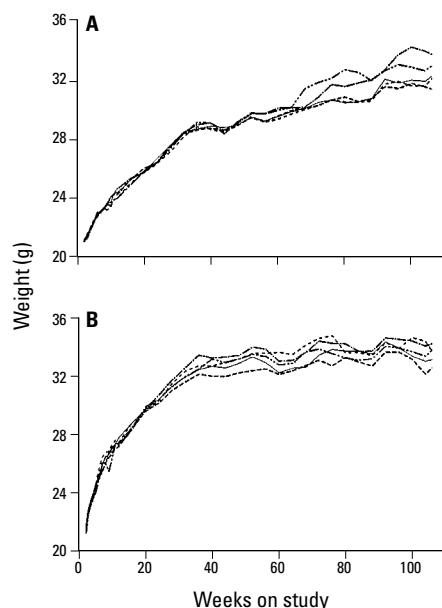


Figure 5. Body weights of female (A) and male (B) B6C3F₁/Nctr BR mice in the 2-year tumor study. The body weights were measured weekly and are shown at 4-week intervals starting at 20 weeks. The following FB₁ doses were given to the female mice: control diets (—), 5 ppm (---), 15 ppm (- · - ·), 50 ppm (- · · -), and 80 ppm (- · · · -). The following FB₁ doses were given to the male mice: control diets (—), 5 ppm (---), 15 ppm (- · - ·), 80 ppm (- · · -), and 150 ppm (- · · · -).

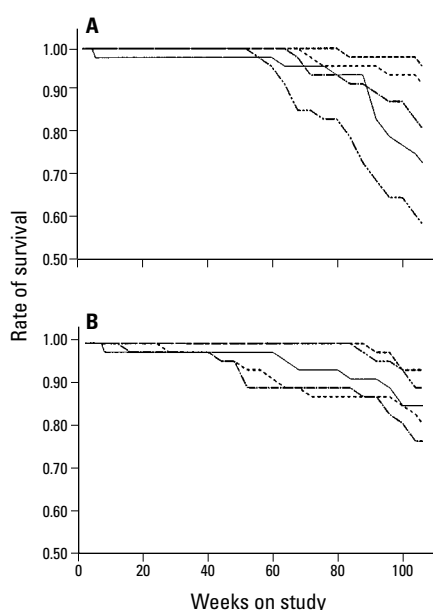


Figure 6. Survival of the female (A) and male (B) B6C3F₁/Nctr BR mice in the 2-year tumor study. Survival rates are expressed as the fraction of mice initially loaded into the study that survived to the end of the indicated week. The following FB₁ doses were given to the female mice: control diets (—), 5 ppm (---), 15 ppm (- · - ·), 50 ppm (- · · -), and 80 ppm (- · · · -). The following FB₁ doses were given to the male mice: control diets (—), 5 ppm (---), 15 ppm (- · - ·), 80 ppm (- · · -), and 150 ppm (- · · · -).

Table 2. Incidences of neoplastic and non-neoplastic lesions in mouse liver.

Type of lesion	Number of female mice examined					Number of male mice examined				
	47	48	48	47	45	47	47	48	48	48
Diffuse hepatocellular hypertrophy	0	0	0	27 ^a	31 ^a	10	9	24 ^a	25 ^a	30 ^a
Hepatocellular adenoma	5	3	1	16	31	9	7	7	6	8
Adjusted rate (percent) ^b	11.7	6.5	2.1	36.3	73.7					
Significance ^b	—	<i>p</i> = 0.3314	<i>p</i> = 0.0862	<i>p</i> = 0.0047	<i>p</i> = 0.0001					
Hepatocellular carcinoma	0	0	0	10	9	4	3	4	3	2
Adjusted rate (percent)	0	0	0	22.5	23.0					
Significance	—	—	—	<i>p</i> = 0.0007	<i>p</i> = 0.0007					
Hepatocellular adenoma or carcinoma	5	3	1	19	39	12	9	9	9	10
Adjusted rate (percent)	11.7	6.5	2.1	42.7	88.3					
Significance ^b	—	<i>p</i> = 0.3314	<i>p</i> = 0.0862	<i>p</i> = 0.0005	<i>p</i> = 0.0001					

^aSignificantly different from control group (*p* < 0.01) using the Poly-k test described in "Methods." ^bThe adjusted incidence rate and significance were determined using the Poly-k test.

can be induced by compounds that bind to $\alpha_{2\mu}$ -globulin (32,33). The morphology of the tumors was not consistent with this type of mechanism. The reasons for the different organospecificities in tumor formation (i.e., livers in the male BD IX rats and kidneys in the male F344 rats) remain to be elucidated and warrant additional studies.

The current study is the first to examine the carcinogenicity of FB₁ in mice. Our results show that FB₁ was hepatocarcinogenic in the female B6C3F₁ mice at doses of 50 and 80 ppm, with carcinoma formation in 22.5 and 23% of the mice, respectively (Table 2). There was an increased incidence of hepatocellular adenomas in the mice fed 50 and 80 ppm FB₁ (Table 2). These results suggest that the NOEL for adenoma or carcinoma in the female B6C3F₁ mouse is between 15 and 50 ppm. Failure to increase the incidence of hepatocellular adenomas and carcinomas in male B6C3F₁ mice and the lack of an increase in tumors in any other sites in response to the consumption of FB₁-containing diets suggests a NOEL for tumor formation in male B6C3F₁ mice > 150 ppm.

Little information is available on the MTD of FB₁ in mice to allow comparison of the FB₁ doses required for tumor induction versus MTD. In a 90-day feeding study with B6C3F₁ mice, Voss et al. (25) demonstrated that 81 ppm FB₁ in the diet was not toxic to male or female B6C3F₁ mice. In male B6C3F₁ mice fed FB₁ for 28 days, decreased body weights were detected in the group that consumed 484 ppm FB₁ but not in the group fed 234 ppm FB₁ (24). The body weights of female B6C3F₁ mice that consumed up to 484 ppm FB₁ for 28 days were not affected (24). Additionally, there were no deaths among the male or female B6C3F₁ mice consuming FB₁ for 28 days. The body weights and survival rates of the male B6C3F₁ mice

in our 2-year study were not affected by the FB₁ (Figures 5 and 6), suggesting that the MTD for FB₁ in a chronic study is > 150 ppm. Although the body weights of the female mice were unaffected by doses of FB₁ as high as 80 ppm, their survival rates decreased beginning after approximately 1 year of consuming diets containing FB₁ (Figure 6). The cause of death in the mice consuming 80 ppm FB₁ was primary liver cancer (data not shown). Therefore, the MTD for FB₁ in female B6C3F₁ mice seems to be between 50 and 80 ppm FB₁ because of the appearance of hepatocarcinogenicity at 80 ppm.

The body weights of the male and female B6C3F₁ mice in the 2-year study were low when compared to the historical body weights of B6C3F₁ mice at NCTR. This apparently was caused by an inadvertent restriction of feed in the feeders used in this study. With a commercial blender, the FB₁ was added as an aqueous solution to predried powdered NIH-31 rodent feed. Particle analysis of the feed did not demonstrate any difference in the particle size before or after application of the FB₁ (data not shown), but the free flow of the powdered feed in the mouse feeders apparently was restricted, which meant that the mice consumed only about 70% of the amount of feed expected for an *ad libitum* study. This explains a reduction in body weight of approximately 15% for the male B6C3F₁ mice and approximately 20% for the female mice at 52 weeks in this study, compared to *ad libitum* studies with other B6C3F₁/Nctr BR mice at NCTR. The effects of reduced body weight through feed restriction were increased longevity and reduced incidence of spontaneous tumor formation (34). Given the body weight of the male mice at 52 weeks, a liver tumor incidence of 20% would have been expected (35). The liver tumor rate in the control

group of male B6C3F₁ mice in this study was 26%; therefore, it appears that the inadvertent feed restriction resulted in the predicted liver tumor rate among the male mice. In another study (36), dietary restriction of female B6C3F₁ mice to 60% of *ad libitum* reduced liver tumors from 55 to 12% of the mice. The adjusted liver tumor rate in our study was 11.7% for the female B6C3F₁ mice consuming control diets. Therefore, it appears that the feed restriction in our study resulted in a decrease in liver tumor rates in the female B6C3F₁ mice similar to those previously reported (36).

F. moniliforme MRC 826 culture material was isolated from an area of South Africa that has a high incidence of esophageal cancer (18–20). Including this fungal isolate at 0.5% (wt/wt) in the diet of BD IX rats led to the development of esophageal hyperplasia and the development of forestomach and liver tumors (21). Further studies with the MRC 826 fungal isolate led to the discovery of FB₁ as the compound responsible for the induction of preneoplastic foci in initiated rats (22) and eventually to hepatic tumor formation in BD IX rats (23). The purification of FB₁ from *F. moniliforme* MRC 826 has led researchers to conclude that FB₁ is a potential human esophageal carcinogen. In this study we were unable to detect any hyperplasia or tumors in the esophageal tissue of rats or mice treated for 2 years with FB₁. Although transient increases in esophageal epithelium labeling index have been reported following gavage administration of FB₁ to rats (37), other reports have indicated a lack of effect of FB₁ consumption on rat esophageal epithelial tissue (22–26).

Esophageal tumors have been induced by many compounds in rat feeding studies. Most of these compounds are *N*-nitrosamines (38), which are structurally dissimilar from the

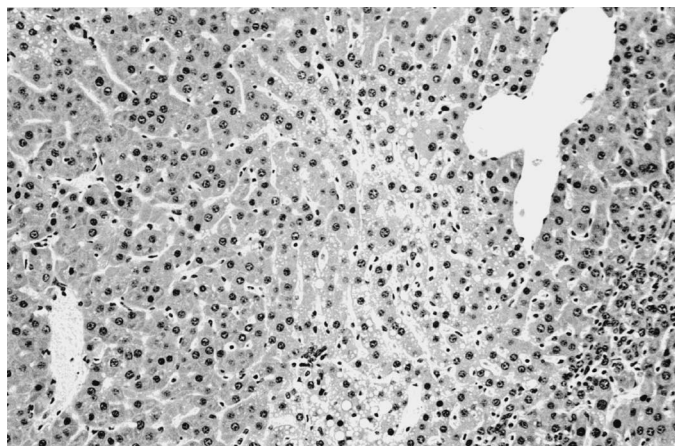


Figure 7. Photomicrograph of a hepatocellular adenoma in a B6C3F₁/Nctr BR mouse that consumed a diet containing 80 ppm FB₁. The adenoma is present in the upper center of the photomicrograph as a focus of basophilic cells of altered morphology.

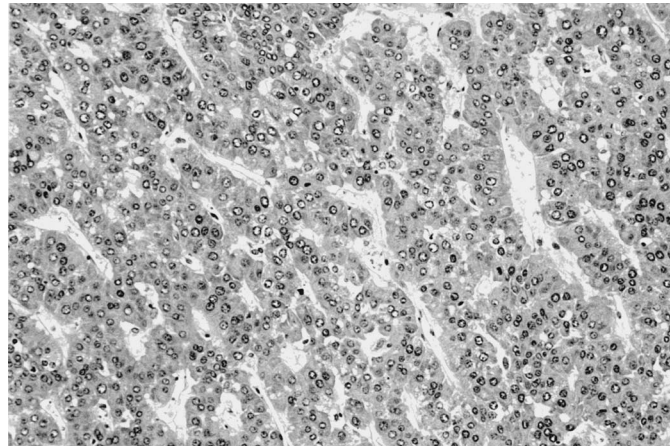


Figure 8. Photomicrograph of a hepatocellular carcinoma in a female B6C3F₁/Nctr BR mouse that consumed a diet containing 80 ppm FB₁. Some normal hepatic tissue is shown at the right side of the photomicrograph; the carcinoma is present in the left center of the photomicrograph.

fumonisin. Whereas *N*-nitrosamines are DNA alkylators, FB₁ is a nongenotoxic compound. We have reported that the incubation of methanolic extracts of *Fusarium* cultures with DNA in the presence of rat liver S9 proteins results in the formation of DNA adducts (39). The chromatographic characteristics of these unidentified DNA adducts suggest they are hydrophobic (39). Therefore, the possibility exists that compounds present in *Fusarium* fungi might alkylate DNA and participate in the induction of *Fusarium*-induced rodent esophageal dysplasia and forestomach tumors. Further research is required to establish whether FB₁ has a role in the development of esophageal cancer in humans.

REFERENCES AND NOTES

1. Bezuidenhout SC, Gelderblom WCA, Gorst-Allman CP, Horak RM, Marasas WFO, Spiteller G, Vleggaar R. Structure elucidation of the fumonisins, mycotoxins from *Fusarium moniliforme*. *J Chem Soc Chem Commun* 1988:743–745 (1988).
2. Plattner RD, Weisleder D, Shackelford DD, Peterson R, Powell RG. A new fumonisin from solid cultures of *Fusarium moniliforme*. *Mycopathologia* 117:23–28 (1992).
3. Scott PM. Fumonisins. *Int J Food Microbiol* 18:257–270 (1993).
4. Branham BE, Plattner RD. Isolation and characterization of a new fumonisin from liquid cultures of *Fusarium moniliforme*. *J Nat Prod* 56:1630–1633 (1993).
5. Dutton MF. Fumonisins, mycotoxins of increasing importance: their nature and their effects. *Pharmacol Ther* 70:137–161 (1996).
6. Nelson PE, Plattner RD, Shackelford DD, Desjardins AE. Fumonisin B₁ production by *Fusarium* species other than *F. moniliforme* in section *Liseola* and by some related species. *Appl Environ Microbiol* 58:984–989 (1992).
7. Thiel PG, Marasas WFO, Sydenham EW, Shephard GS, Gelderblom WCA, Nieuwenhuis JJ. Survey of fumonisin production by *Fusarium* species. *Appl Environ Microbiol* 57:1089–1093 (1991).
8. Nelson PE. Taxonomy and biology of *Fusarium moniliforme*. *Mycopathologia* 117:29–36 (1992).
9. Norred WP. Fumonisins—Mycotoxins produced by *Fusarium moniliforme*. *J Toxicol Environ Health* 38:309–328 (1993).
10. Bullerman LB, Tsai WYJ. Incidence and level of *Fusarium moniliforme*, *Fusarium proliferatum* and fumonisins in corn. *J Food Prot* 57:541–547 (1994).
11. Meireless MCA, Correa B, Fischman O, Gambale W, Paula CR, Chacon-Reche NO, Pozzi CR. Mycoflora of the toxic feeds associated with equine leukoencephalomalacia (ELEM) outbreaks in Brazil. *Mycopathologia* 127:183–188 (1994).
12. Abbas HK, Ocamb CM, Xie WP, Mirocha CJ, Shier WT. First report of fumonisin B₁, B₂, and B₃ produced by *Fusarium oxysporum* var *redolens*. *Plant Dis* 79:968 (1995).
13. Abbas HK, Ocamb CM. First report of production of fumonisin B₁ by *Fusarium polyphialidicum* collected from seeds of *Pinus strobus*. *Plant Dis* 79:642 (1995).
14. Rose EF, McGlashan ND. The spatial distribution of oesophageal carcinoma in the Transkei, South Africa. *Br J Cancer* 31:197–206 (1975).
15. Rose EF. Esophageal cancer in Transkei—the pattern and associated risk factors. In: *Cancer of the Esophagus* (Piffier CJ, ed). Boca Raton, FL: CRC Press, 1982:19–28.
16. Marasas WFO, Jaskiewicz K, Venter FS, van Sckalkwyk DJ. *Fusarium moniliforme* contamination of maize in oesophageal cancer areas in Transkei. *S. African Med J* 74:110–114 (1988).
17. Rheeder JP, Marasas WFO, Thiel PG, Sydenham EW, Shepherd GS, van Sckalkwyk DJ. *Fusarium moniliforme* and fumonisins in corn in relation to esophageal cancer in Transkei. *Phytopathology* 82:353–357 (1992).
18. Kriek NPJ, Kellerman TS, Marasas WFO. A comparative study of the toxicity of *Fusarium verticillioides* (= *F. moniliforme*) to horses, primates, pigs, sheep and rats. *Understepoort J Vet Res* 48:129–131 (1981).
19. Kriek NPJ, Marasas WFO, Thiel PG. Hepato- and cardiotoxicity of *Fusarium verticillioides* (*F. moniliforme*) isolates from southern African maize. *Food Cosmet Toxicol* 19:447–456 (1981).
20. Jaskiewicz K, van Rensburg SJ, Marasas WF, Gelderblom WC. Carcinogenicity of *Fusarium moniliforme* culture material in rats. *J Natl Cancer Inst* 78:321–325 (1987).
21. Marasas WFO, Kriek NPJ, Fincham JE, van Rensburg SJ. Primary liver cancer and oesophageal basal cell hyperplasia in rats caused by *Fusarium moniliforme*. *Int J Cancer* 34:383–387 (1984).
22. Gelderblom WCA, Jaskiewicz K, Marasas WFO, Thiel PG, Horak RM, Vleggaar R, Kriek NPJ. Fumonisins—Novel mycotoxins with cancer-promoting activity produced by *Fusarium moniliforme*. *Appl Environ Microbiol* 54:1806–1811 (1988).
23. Gelderblom WCA, Kriek NP, Marasas WF, Thiel PG. Toxicity and carcinogenicity of the *Fusarium moniliforme* metabolite, fumonisin B₁, in rats. *Carcinogenesis* 12:1247–1251 (1991).
24. NTP. Toxicology and Carcinogenesis Studies on Fumonisin B₁ in F344/N Rats and B6C3F₁ Mice (Feed Studies). Technical Report Series, No. 496. NIH Publication No. 99-3955. Research Triangle Park, NC: National Toxicology Program, (1999).
25. Voss KA, Chamberlain WJ, Bacon CW, Herbert RA, Walters DB, Norred WP. Subchronic feeding study of the mycotoxin fumonisin B₁ in B6C3F₁ mice and Fischer 344 rats. *Fund Appl Toxicol* 24:102–110 (1995).
26. Voss KA, Chamberlain WJ, Bacon CW, Norred WP. A preliminary investigation on renal and hepatic toxicity in rats fed purified fumonisin B₁. *Natural Toxins* 1:222–228 (1993).
27. NTP. Specification for the Conduct of Studies to Evaluate the Toxic and Carcinogenic Potential of Chemical, Biological, and Physical Agents in Laboratory Animals for the National Toxicology Program. Research Triangle Park, NC: National Toxicology Program, 1994.
28. U.S. FDA. Toxicological Principles for the Safety Assessment of Direct Food Additives and Color Additives Used in Food. Redbook II. Washington, DC: U.S. Food and Drug Administration, 1993.
29. Portier CJ, Hedges JC, Hoel DG. Age-specific models of mortality and tumor onset for historical control animals in the National Toxicology Program's carcinogenicity experiments. *Cancer Res* 46:4372–4378 (1986).
30. Bailer AJ, Portier CJ. Effects of treatment-induced mortality and tumor-induced mortality on tests for carcinogenicity in small samples. *Biometrics* 44:417–431 (1988).
31. Gelderblom WC, Smuts CM, Abel S, Snyman SD, van der Westhuizen L, Huber WW, Swanevelter S. Effect of fumonisin B₁ on the levels and fatty acid decomposition of selected lipids in rat liver *in vivo*. *Food Chem Toxicol* 35:647–656 (1997).
32. Short BG, Burnett VL, Swenberg JA. Elevated proliferation of proximal tubule cells and localization of accumulated α₂_u-globulin in F344 rats during chronic exposure to unleaded gasoline or 2,2,4-trimethylpentane. *Toxicol Appl Pharmacol* 101:414–431 (1989).
33. Dietrich DR, Swenberg JA. The presence of α₂_u-globulin is necessary for d-limonene promotion of male rat kidney tumors. *Cancer Res* 51:3512–3521 (1991).
34. Sheldon WG, Bucc TJ, Hart RW, Turturro A. Age-related neoplasia in a lifetime study of *ad libitum*-fed and food-restricted B6C3F₁ mice. *Toxicol Pathol* 23:458–476 (1995).
35. Leakey JEA, Seng JE, Barnas CR, Baker VM, Hart RW. A mechanistic basis for the beneficial effects of caloric restriction on longevity and disease: consequences for the interpretation of rodent toxicity studies. *Int J Toxicol* 17:5–56 (1998).
36. Haseman JK. National Toxicology Program experience with dietary restriction: does the manner in which reduced body weight is achieved affect tumor incidence? *Int J Toxicol* 17:119–134 (1998).
37. Lim CW, Parker HM, Vesonder RF, Haschek WM. Intravenous fumonisin B₁ induces cell proliferation and apoptosis in the rat. *Nat Toxins* 4:34–41 (1996).
38. Summary of the Carcinogenic Potency Database by Target Organ. Available: <http://potency.berkeley.edu/pathology.table.html> [cited 1 August 2000].
39. Bever RJ Jr, Couch LH, Sutherland JB, Williams AJ, Beger RD, Churchwell MI, Doerge DR, Howard PC. DNA adduct formation by *Fusarium* culture extracts: lack of role of fusarin C. *Chem-Biol Interact* 128:141–157 (2000).