Forest Health Technology Enterprise Team

TECHNOLOGY TRANSFER

Invasive Species

Catalogue of Introductions of Pathogens and Nematodes for Classical Biological Control of Insects and Mites



ANN E. HAJEK, MICHAEL L. MCMANUS, ITALO DELALIBERA JÚNIOR









FHTET-2005-05 September 2005 The Forest Health Technology Enterprise Team (FHTET) was created in 1995 by the Deputy Chief for State and Private Forestry, USDA Forest Service, to develop and deliver technologies to protect and improve the health of American forests. This book was published by FHTET as part of the technology transfer series.

http://www.fs.fed.us/foresthealth/technology/



Cover photo by Vince D'Amico University of Delaware USDA-Forest Service Dept. Entomology Townsend Hall Newark, DE 19716

610-368-4289 vdamico@fs.fed.us

The U.S. Department of Agriculture (USDA) prohibits discrimination in all its programs and activities on the basis of race, color, national origin, sex, religion, age, disability, political beliefs, sexual orientation, or marital or family status. (Not all prohibited bases apply to all programs.) Persons with disabilities who require alternative means for communication of program information (Braille, large print, audiotape, etc.) should contact USDA's TARGET Center at 202-720-2600 (voice and TDD).

To file a complaint of discrimination, write USDA, Director, Office of Civil Rights, Room 326-W, Whitten Building, 1400 Independence Avenue, SW, Washington, D.C. 20250-9410 or call 202-720-5964 (voice and TDD). USDA is an equal opportunity provider and employer.

The use of trade, firm, or corporation names in this publication is for information only and does not constitute an endorsement by the U.S. Department of Agriculture.



Federal Recycling Program Printed on recycled paper.

Catalogue of Introductions of Pathogens and Nematodes for Classical Biological Control of Insects and Mites

Ann E. Hajek Department of Entomology, Cornell University Ithaca, New York USA

Michael L. McManus USDA Forest Service, Northeastern Research Station Hamden, Connecticut, USA

ITALO DELALIBERA JÚNIOR Department of Entomology, Plant Pathology, and Agricultural Zoology ESALQ-University of São Paulo Piracicaba, Brazil

For additional copies of this publication contact:

Richard Reardon FHTET, USDA Forest Service 180 Canfield St. Morgantown, WV 26505 U.S.A. (304) 285-1566 rreardon@fs.fed.us

Acknolwedgements

This publication would not have been possible without the help of many people including P. Allsopp, T. Andreadis, J. Becnel, A. Bellotti, G. Blissard, D. Boucias, G. Carner, A. Diss, J. Eilenberg, H. Evans, B. Federici, J. Fuxa, P. Gullan, I. Hall, L. Harrington, D. Haugen, E. R. Hoebeke, K. Hodge, J. Huber, R. Humber, C. Ignoffo, P. Klasmer, M. Klein, C. Lange, J. Liebherr, J. Martin, S. Merkel, M. Montgomery, D. Moore, C. Nielsen, D. Pilarska, J. Podgwaite, G. Poinar, B. Ratcliffe, M. Schlabach, A. Sharov, D. Smith, L. Smith, L. Solter, D. Sosa-Gomez, P. Stock, D. Streett, K. Teramoto, F. Vega, L. Volkman, J. Weiser, S. Woods, O. Zethner, S. Zinder, Yang Zhong-qi and many more. We also sincerely thank the excellent Entomology and Mann Libraries at Cornell University and the helpful and very knowledgeable librarians who work there.

Finally, we wish to thank Dick Reardon, USDA Forest Service, Forest Health Technology Enterprise Team (FHTET), for supporting this project, and Chuck Benedict, USDA Forest Service, FHTET/ITX, for editing, designing, and laying out this publication, and for sheparding it through production.

Table of Contents

Introduction1
Table A: Exotic viruses released and target hosts
Table B: Exotic bacteria released and target hosts 11
Table C: Exotic fungi and oomycetes released and target hosts
Table D: Exotic microsporidia released and target hosts
Table E: Exotic nematodes released and target hosts
Table F: Accidental introductions of pathogens and nematodes
Chart 1: Classification of pathogens and nematodes purposefully introduced for classical biological control of insects and mites and accidentally introduced pathogens and nematodes infecting insects and mites
Chart 2: Classification of insects and mites targeted by classical biological control programs using pathogens and nematodes, and insect and mite hosts of accidentally introduced pathogens and nematodes
References
Indexes
Scientific names of insect and mite hosts56
Families of insect and mite hosts57
Scientific names of pathogens and nematodes57
Higher classification of pathogens and nematodes58
Release countries and regions58
Source countries and regions59

Classical biological control is a strategy that has been defined as "The intentional introduction of an exotic biological control agent for permanent establishment and long-term pest control" (41). Numerous summaries of the many classical biological control programs have been published (e.g., 28, 55, 74). This strategy has been used extensively to control weeds and arthropods pests. For control of weeds phytophagous arthropods have principally been used and for control of arthropod pests parasitoids and predators have principally been used (58).

Most programs using pathogens and nematodes for control of insects and mites have focused on mass production and inundative release. As long-term solutions for insect and mite pests (i.e., use in classical biological control programs), pathogens and nematodes have been used much less frequently when compared with parasitoids and predators (46, 59). Interestingly, while some classical biological control programs using pathogens and nematodes have been very successful in controlling insect and mite pests, some accidental introductions of entomopathogenic agents have also yielded substantial and long-term control.

We believe this publication is the most complete catalogue to date of classical biological control programs that have used pathogens and nematodes to control arthropod pests. It was difficult to find many of the classical biological control programs listed in the tables that follow; probably, we have not listed them all. Likewise, it was often difficult determining whether a release program should be included in this catalogue, particularly when a program was implemented many years ago and/ or not thoroughly documented. Thus, we used the following criteria for including programs in this catalogue:

- 1. The target pest was an insect or mite.
- 2. The microbial pathogen or nematode was an exotic (non-native) in the area of release. We include programs where the species of microbe or nematode was exotic (introduced) as well as programs where only the strain or biotype released was exotic.

3. Whether the releases were successful or not, the establishment of the microbe appeared to be a goal (i.e., long-term establishment and control) and was either investigated or discussed or, for older programs, we can infer that establishment of the pathogen or nematode was a goal of the program.

(Note: Intentionally, we did not include examples of early widespread introductions of entomopathogens that were later shown to be questionably pathogenic, or widespread introductions where contaminants were actually released instead of the intended organisms [e.g., see 25, 147].)

Organization of tables and charts

Tables

Intentional and accidental releases of entomopathogens are grouped according to specific pathogen and nematode groups, and are presented in Tables A through F. Column headings and descriptions of contents are as follows:

Pest Group and Species

Only pestiferous insect and mite hosts are included. Taxonomic grouping, scientific names and synonyms for species names used in the publications cited or in the literature, are provided. If known, common names for pests are included.

Biological Control Agent

All natural enemies listed are exotic to their respective areas of release, i.e., either the species or the strain released was exotic, and include viruses, bacteria, fungi, a chromist, microsporidia and nematodes. Scientific names and synonyms are provided.

Release date (year)

The year of release is listed, providing the intent of the release was to establish the pathogen or nematode in the release area. In some cases, pathogen or nematode levels declined over time, so agents were re-introduced. In other cases, pathogens have been re-introduced throughout a region over a period of years because the agents spread slowly on their own. In both cases, we list only the year or years of the initial release; the dates of second or third introductions, or release in later years in the same general region, are included only if the initial releases were from a different source or sources. In the case of accidental introductions (Table F), the year the agent was first found is listed.

Release country, source and results from introduction

Releases are presented separately for geographically isolated areas and are listed by the country, or in a few instances by region (e.g. "Europe"), where the release was made (given in capital letters, e.g., BRAZIL). In some cases, a pathogen or nematode was released in more than one area within the same country. If release areas are isolated from one another, these introductions are considered separate introductions. For example, releases of Romanomermis culicivorax (originating from Louisiana) in both Maryland and California would be considered separate introductions although both are states within the same country because these release areas are geographically separated from one another and the sites differ in climate and topographies. The exception to this would be the release of a pathogen or nematode on proximate islands of the same country, e.g., in the many island groups in the south Pacific. If it appears that the introductions of pathogens or nematodes on proximate islands within a group were part of the same program, only the initial introduction is listed.

The geographical location where the pathogen or nematode was acquired for the release follows the release country (after Ex., e.g. Ex. China). Whenever appropriate, microbes from different source locations are listed separately.

Results of introductions are provided as brief summaries of establishment, control, and persistence. We found that it is not always easy to classify control programs by strategy (i.e., classical biological control vs. inundative augmentation) and there are multitudes of programs where pathogens and nematodes have been released inundatively. For studies to be included in this catalogue, there must be some documented evidence that, whether the pathogen persisted or not after release, the intent of the program was to establish the pathogen in the release area for long-term, not temporary, control. Some older, poorly documented programs are exceptions and are included when we inferred the goal was establishment.

Clear summaries of results from introductions cannot always be found. In some cases, this is because not enough time has transpired since the release to see an effect. Unfortunately, in other cases, especially in earlier programs, we could find no documentation of what happened after releases.

Pest origin

I = Introduced (exotic) N = Native (endemic) ? = Origin unknown

Charts

Chart 1 provides the classification for pathogens and nematodes included in the catalogue either as intentional or accidental releases.

Chart 2 provides the classification for insect and mite hosts targeted by pathogens or nematodes that were either intentionally or accidentally introduced.

References

The reference list does not include every mention of a classical biological control introduction of a pathogen or nematode. Rather, it includes selected sources providing the information presented in this catalogue. If the information included in the catalogue has not been published, the individual providing the information is cited.

TABLE A: EXOTIC VIRUSES RELEASED AND TARGET HOSTS

ORD	ER	: COLEOPTERA				
FA	MIL	Y				
	_	Scarabaeidae				
	SP	ECIES				
		Oryctes rhinoceros (L.)				
		(rhinoceros beetle)				
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
		Oryctes rhinoceros virus (OrV) (= Rhabdionvirus oryctes (Huger); = Baculovirus oryctes Huger) (Currently unassigned; previously in Baculoviridae)	1967	SAMOA (= WESTERN SAMOA) Ex. Malaysia. Established in 1 year and spread. Between 1973-75, adult infection decreased from 63 to 35% and although total population density also declined, damage was noticed again. Virus was re-released 1975-1978 with a resulting decline in damage.	Ι	14, 89, 90, 145, 163
I			1967	TOKELAU ISLANDS Ex. Malaysia. Released on Nukunonu Atoll. Established and by 1973 39% of beetles infected and only 1.5-6.5% of palm fronds damaged.	Ι	14, 145, 164
I			1970-74	FIJI Ex. Samoa (= Western Samoa). Established and by 1974 57-68% of beetles infected. Damage decreased significantly 12-18 months after virus establishment.	Ι	12, 13, 14, 145
			1970	PALAU Ex. Samoa (= Western Samoa). Established on Babeldaob Isl., controlling beetles.	I	129, 145
			1983	PALAU Ex. Samoa (= Western Samoa). Released on Peleliu Isl. and "other places where beetle problems were evident," resulting in beetle control.	I	129
				resulting in beetle control.		

DLEOPTERA	arabaeidae	yctes rhinoceros (L.)	ctes rhinoceros virus (OrV)	1970-71	WALLIS ISLAND Ex. Samoa (= Western Samoa). Established, < 2 months after release spread over entire island. In 1 year beetle populations decreased by 60- 80% and damage decreased by 82%. Average number infested palms reduced from 60% in 1967 to 20% in 1981.	I	14, 56, 62, 145
00	Sc	OL	õ	1970-71	TONGA Ex. Samoa (= Western Samoa). Released in Tongatapu. Established, epizootics developed in 5 months and virus spread at 2-3 km/month, beetles and damage reduced. After 7 years, 84% of adult beetles infected throughout population and damage remained low (< 5% of palm crowns surveyed).	Ι	145, 160, 161
				1970-72	MAURITIUS Ex. Samoa (= Western Samoa). Established, beetle populations declined sharply from 1970. At least through 1976-77, damage reduced by 60-95%.	I	14, 105
				1972	AMERICAN SAMOA Ex. Samoa (= Western Samoa). Established, virus spread 0.8- 1.6 km/month and damage declined.	I	14, 145
				1978-79	PAPUA NEW GUINEA Ex. Samoa (= Western Samoa). Released on 3 islands; established at nearly all sites, spread at 1 km/month.	I	53
				1983-84	INDIA Ex. India (Kerala). Released on Minicoy Island. Established within 9 months, pest suppressed to low levels and damage reduced. Pest remained at low levels 3.5 years after release.	Ν	101

COLEOPTERA	Scarabaeidae	Oryctes rhinoceros (L.)		1987	INDIA Ex. India (Kerala). Released at 4 locations on Andaman Islands. Palm damage reduced by 90% within 43 months of release, large reduction in numbers of adults and numbers of breeding sites. Virus spread at 1 km/year. By 1996, beetle populations remained at low levels.	N	71
				1984-85	MALDIVES Ex. Philippines, Tanzania & Malaysia. Established and caused highly significant reduction in palm damage on most islands where released. Different strains released and one strain (X2B) consistently yielded better infection and pest reduction.	Ν	31, 165
		SD					
		JF	Oryctes monoceros (Olivier)				
			(rhinoceros beetle)				
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			Oryctes rhinoceros virus (OrV) (= Rhabdionvirus oryctes (Huger); = Baculovirus oryctes Huger) (Currently unassigned; previously in Baculoviridae)	1973	SEYCHELLES Ex. Samoa. Released on Mahé, Praslin Island group & La Digue. Establishment confirmed in 1986 on Praslin Island group only, with infection 70-90%.	Ν	86, 87
				1981-83	SEYCHELLES Ex. Praslin Island group. Established on Mahé & Ste. Anne with 20-50% infection and 30% reduction in beetle population.	Ν	86, 87
				1983-87	TANZANIA Ex. Philippines & Samoa (= Western Samoa). Established at 2 sites, with 40- 60% infection after 1-1.5 years but reduction in f1020	Ν	123, 131

TERA	dae	SP	ECIES <i>Scapanes australis</i> (Boisduval) (rhinoceros beetle)				
DLEOF	arabaei		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
00	Sci		Oryctes rhinoceros virus (OrV) (= Rhabdionvirus oryctes (Huger); = Baculovirus oryctes Huger) (Currently unassigned; previously in Baculoviridae)	before 1979	SOLOMON ISLANDS Ex. Fiji. Virus released on Guadalcanal but fate unknown.	Ν	139

ORDER: LEPIDOPTERA

FAMILY

Zygaenidae

SPECIES

Harrisina brillians (Barnes & McDunnough)

(western grapeleaf skeletonizer)

	·			
BIOLOGICAL CONTROL AGENT	Release date	Country, source, and results from introduction	Pest origin	References
<i>Harrisina brillians</i> Granulovirus (HbGV) (Baculoviridae)	1981-1982	UNITED STATES OF AMERICA Ex. Mexico & USA (Arizona). Released in central California (Tulare Co.). Established, epizootics develop in high density host populations. Overall, lowers general equilibrium density of host populations.	Ι	144

MIL	_Y				
	Lymantriidae				
SP	PECIES				
	Lymantria dispar (L.)				
	(gypsy moth)				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Lymantria dispar</i> Multicapsid Nucleopolyhedrovirus (LdMNPV) (Baculoviridae)	1972	SARDINIA Ex. Serbia. Established, high levels of larval mortality year of release, > 40% infection the next year and spread over 300 hectares.	Ν	88
	SP	MILY Lymantriidae SPECIES Lymantria dispar (L.) (gypsy moth) BIOLOGICAL CONTROL AGENT Lymantria dispar Multicapsid Nucleopolyhedrovirus (LdMNPV) (Baculoviridae)	MILY Lymantriidae SPECIES Lymantria dispar (L.) (gypsy moth) BIOLOGICAL CONTROL AGENT Lymantria dispar Multicapsid Nucleopolyhedrovirus (LdMNPV) (Baculoviridae)	MILY Lymantriidae SPECIES Lymantria dispar (L.) (gypsy moth) BIOLOGICAL CONTROL AGENT Lymantria dispar Multicapsid Nucleopolyhedrovirus (LdMNPV) (Baculoviridae) RELEASE DATE SARDINIA Ex. Serbia. Established, high levels of larval mortality year of release, > 40% infection the next year and spread over 300 hectares.	MILY Lymantriidae SPECIES Lymantria dispar (L.) (gypsy moth) BIOLOGICAL CONTROL AGENT Lymantria dispar Multicapsid Nucleopolyhedrovirus (LdMNPV) (Baculoviridae) 1972 SARDINIA Ex. Serbia. Established, high levels of larval mortality year of release, > 40% infection the next year and spread over 300 hectares.

LEPIDOPTERA Lymantriidae

SPECIES				
Lymantria monacha (L.)				
(nun moth)				
BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
<i>Lymantria monacha</i> Nucleopolyhedrovirus (Baculoviridae)	1973-4	DENMARK Ex. Sweden and West Germany. Released in Silkeborg (1973), 90% infection year of release and, in 1974, no serious defoliation within and directly around virus-release stands while insecticides had to be applied to other areas. In 1975, no virus was found in the few larvae collected. In Grindsted (1974), the population collapsed the year of virus release but it is suggested that other factors, including the native virus, played important parts.	Ν	167

FAMILY

	Noctuidae				
SF	PECIES				
	Anticarsia gemmatalis (Hübner)				
	(velvetbean caterpillar)				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Anticarsia gemmatalis</i> Multicapsid Nucleopolyhedrovirus (AgMNPV) (Baculoviridae)	1979-80	UNITED STATES OF AMERICA Ex. Brazil (Santa Catarina). Released in South Carolina. 59-86% infection the season of release but no infections found 1 year after release.	Ν	8, 23
		1990-91	UNITED STATES OF AMERICA Ex. Brazil. Released in soybean fields in Louisiana. Established, causing 25-100% infection the year of release and 4-49% infection for years 2-4 after release, even in rotated fields.	Ν	47

		SF	ECIES				
R			Trichoplusia ni (Hübner)				
핕	tuidae		(cabbage looper)				
PDOP			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
LER	Noc		<i>Trichoplusia ni</i> Nucleopolyhedrovirus (TnNPV) (Baculoviridae)	1970	COLOMBIA Ex. USA (California). Persisted after release, controlling subsequent pest generations.	Ι	15, 33
		58	'EUIES Regudantusia includons (Walkov	-)			
			(sovbean looper))			
				RELEASE	COUNTRY, SOURCE, AND	PEST	
			BIOLOGICAL CONTROL AGENT	DATE	RESULTS FROM INTRO.	ORIGIN	REFERENCES
			<i>Pseudoplusia includens</i> Singlecapsid Nucleopolyhedrovirus (PiSNPV) (Baculoviridae)	1975-77	UNITED STATES OF AMERICA Ex. Guatemala. Released in soybean fields in Louisiana, established, 38-63% infection 12-15 years after introduction.	Ν	48
		SP	PECIES	(a www.iiill.a.w)			
			Agrotis segetum (Denis & Schift (turnin moth)	ermuller)			
				RELEASE	COUNTRY SOURCE AND	PEST	
			BIOLOGICAL CONTROL AGENT	DATE	RESULTS FROM INTRO.	ORIGIN	REFERENCES
			<i>Agrotis segetum</i> Granulovirus (AsGV) (Baculoviridae)	1975-80	DENMARK Ex. Austria. Released in Lammefjord. Caused 65-70% reduction in damage soon after release and thought to have spread 10 m from release. One year after release, ca. 99% of infectivity of virus applied to soils had been lost.	Ν	166, 168, 171

OF	٦D	ER	: HYMENOPTERA				
	FA	MIL	_Y				
			Diprionidae				
		SF	ECIES				
			Neodiprion sertifer (Geoffrey)				
			(European pine sawfly)				
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			<i>Neodiprion sertifer</i> Nucleopolyhedrovirus (NeseNPV) (Baculoviridae)	1950	CANADA Ex. Sweden. Released in southern Ontario, near Strathroy. > 90% mortality 14 days after release and virus persisted. Widely distributed for release in pine plantations, e.g., one introduction in 1951 controlled an infestation over 100 acres within 3 years. After introduction, this virus replaced insecticides for controlling hosts and provided long-term control. Today, host is a minor pest of plantations and ornamentals but occasionally can increase locally as natural spread and effectiveness of the virus is much reduced at low host densities.	Ι	19, 34, 39, 92
				1951-52	UNITED STATES OF AMERICA Ex. Canada. Released in New Jersey, established and spread (ca. 300 m from individual trees after release). Provided complete control.	Ι	39

HYMENOPTERA	Diprionidae	Neodiprion sertifer (Geoffrey)	Neodiprion sertifer	1952	UNITED STATES OF AMERICA Ex. USA (New Jersey). Released in Illinois. By 19 days after treatment, 82-100% control. In 1953, spread was up to 80 m from treated area. Excellent control achieved. Virus from Canada released in Indiana from 1953 and reported as maintaining adequate control over several years through recurring epizootics after establishment.	Ι	39, 130
				1961	UNITED KINGDOM (SCOTLAND) Ex. Canada. 85% of colonies had infected individuals 24 days after release, resulting in very good control. In 1962, found to persist in treated areas but minimal spread.	Ν	35, 126
		SP	PECIES				
			Gilpinia hercyniae (Hartig) (= Di	prion hercy	niae (Hartig))		
			(European spruce sawfly)				
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			<i>Gilpinia hercyniae</i> Nucleopolyhedrovirus (GhNPV) (Baculoviridae)	1943-45	CANADA Ex. Canada (Mainland). Released in Newfoundland. Established and by 1946 reported as prevalent over considerable areas surrounding release areas.	I	4, 92
				1950	CANADA Ex. Canada (New Brunswick). Released in an isolated host population in Sault Ste. Marie, Ontario, 160 km (100 miles) beyond western distribution of insect. Established and spread rapidly through infested area.	Ι	20

TABLE B: EXOTIC BACTERIA RELEASED AND TARGET HOSTS

DEF	R: COLEOPTERA				
MIL	_Y				
_	Scarabaeidae				
SP	PECIES				
	Papuana huebneri (Halmahera)				
	(taro beetle)				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Paenibacillus popilliae</i> (Dutky) (Bacillaceae)	1976	KIRIBATI Ex. Papua New Guinea & Solomon Islands. Released on So. Tarawa. Isolate from Papua New Guinea (type A1) caused infections 1 year after release.	I	149
SP	PECIES				
	Oryctes rhinoceros (L.)				
	(rhinoceros beetle)				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Paenibacillus popilliae</i> (Dutky) (Bacillaceae)	1951	PALAU Ex. USA (strain from Japanese beetle, <i>Popillia japonica</i> Newman). Not recovered after release.	Ι	145
		1957	AMERICAN SAMOA Ex. USA (strain from Japanese beetle, <i>Popillia japonica</i> Newman). Not recovered after release.	Ι	145
SP	ECIES				
	Schizonycha sp.				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Paenibacillus popilliae</i> (Dutky) (Bacillaceae)	1956	KENYA Ex. USA (A & B strains from Japanese beetle, <i>Popillia</i>	Ν	54

RA		SP	PECIES Cochliotis melolonthoides (Gers	staecker)			
OPTE	aeidae		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
COLE	Scarab		<i>Paenibacillus popilliae</i> (Dutky) (Bacillaceae)	1968	TANZANIA Ex. Probably USA. Japanese beetle, <i>Popillia</i> <i>japonica</i> Newman. Seemed to become established but this is not certain due to presence of an indigenous milky disease.	Ν	54

TABLE C: EXOTIC FUNGI RELEASED AND TARGET HOSTS

ORDEF	R: ORTHOPTERA				
FAMI	LY				
	Acrididae				
S	PECIES				
	Phaulacridium vittatum (Sjösted	dt)			
	(wingless grasshopper)				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Entomophaga grylli</i> (Fresenius) Batko, pathotype I (Zygomycetes: Entomophthoraceae)	1984	AUSTRALIA Ex. USA (Arizona). Released near Canberra. Epizootics did not occur and permanent establishment questioned, efficacy unlikely.	Ν	96
	Various species, including <i>Mela</i> <i>pellucida</i> Scudder (twostriped grasshopper, migra	tory grassh RELEASE	opper, clearwinged grasshopp COUNTRY, SOURCE, AND	per) PEST	.), Camnula
	<i>Entomophaga grylli</i> (Fresenius) Batko, pathotype III (Zygomycetes: Entomophthoraceae)	1989-91	UNITED STATES OF AMERICA Ex. Australia. Isolate chosen based on biology, similarity of climates and ability to infect species in both Oedipodinae and Melanoplinae, but not <i>Hesperotettix viridis</i> (Scudder). Released in North Dakota. Populations of some species declined in 1991-92 with 23% infection in 1992 at < 1 km from release, 1.7% in 1993 and no infection in 1994 when host populations were low. Long-term establishment questionable	N	18, 24, 25

ERA		SP	ECIES Melanoplus sanguinipes (F.) (migratory grasshopper)				
THOPT	Ididae		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
OR	ACLI		<i>Entomophaga grylli</i> (Fresenius) Batko, pathotype III (Zygomycetes: Entomoph- thoraceae)	1990	UNITED STATES OF AMERICA Ex. Australia. Released in Alaska. No establishment reported.	Ν	24, 124

RD	ER	: HEMIPTERA				
FA	MIL	_Y				
		Cercopidae				
	SP	ECIES				
		Aeneolamia flavilatera (Ulrich)				
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
		<i>Metarhizium anisopliae</i> (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales)	1944	GUYANA Ex. Trinidad. Introduced by releasing infected adult froghoppers. Established, considered unsuccessful for control but < 1 year later abundant infections ca. 32 km away. Unknown whether this was due to introduced or indigenous fungus.	Ν	29

LY				
Cicadellidae				
PECIES				
Empoasca fabae (Harris)				
(potato leafhopper)				
BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
<i>Zoophthora radicans</i> (Brefeld) Batko (Zygomycetes: Entomophthoraceae)	1984	UNITED STATES OF AMERICA Ex. Brazil. Introduced to Illinois. No establishment.	I	65, 93
	LY Cicadellidae ECIES Empoasca fabae (Harris) (potato leafhopper) BIOLOGICAL CONTROL AGENT Zoophthora radicans (Brefeld) Batko (Zygomycetes: Entomophthoraceae)	Y Cicadellidae PECIES Empoasca fabae (Harris) (potato leafhopper) BIOLOGICAL CONTROL AGENT Zoophthora radicans (Brefeld) Batko (Zygomycetes: Entomophthoraceae)	Y Cicadellidae Cicadellidae ECIES Empoasca fabae (Harris) (potato leafhopper) BIOLOGICAL CONTROL AGENT RELEASE COUNTRY, SOURCE, AND DATE Zoophthora radicans (Brefeld) 1984 UNITED STATES OF AMERICA Ex. Brazil. Batko 1984 UNITED STATES OF AMERICA Ex. Brazil. Introduced to Illinois. No establishment. Introduced to Illinois. No	Y Cicadellidae ECIES Empoasca fabae (Harris) (potato leafhopper) RELEASE COUNTRY, SOURCE, AND PEST BIOLOGICAL CONTROL AGENT RELEASE COUNTRY, SOURCE, AND PEST Zoophthora radicans (Brefeld) 1984 UNITED STATES OF I Batko AMERICA Ex. Brazil. Introduced to Illinois. No I Entomophthoraceae) establishment. I

		SP	PECIES				
2	е		Unspecified species				
Ë	lida		(leafhopper)				
HEMIP	Cicadel		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			Unknown fungus	1906	UNITED STATES OF AMERICA Ex. Australia and Fiji, where they infected leafhopper eggs. Released in Hawaii. Establishment not confirmed.	Ι?	114
	FA	MIL	_Y				
			Aphididae				
		SF	PECIES				
			Therioaphis maculata (Buckton)				
			(spotted alfalfa aphid)				
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			<i>Zoophthora radicans</i> (Brefeld) Batko (Zygomycetes: Entomophthoraceae)	1979	AUSTRALIA Ex. Israel. Isolate chosen in part due to similar climate. Released in New South Wales. Became widely distributed in New South Wales and southern Queensland, causing epizootics in late summer/autumn: only the first	Ι	97, 98, 99

S	PECIES				
L	Aphis gossypii Glover				
2	(cotton aphid)				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
S	Neozygites fresenii (Nowakowski) Batko (Zygomycetes: Neozygitaceae)	1994-95	UNITED STATES OF AMERICA Ex. USA (Arkansas). Released in San Joaquin Valley, California. Cycling during release seasons with infection levels that would have initiated epizootics in Arkansas but epizootics did not occur. Persisted until end of release seasons but not recovered 1997-2001, so long-term establishment questionable.		52, 142
ľ	Metopolophium dirhodum (Walk	er), plus ot	her cereal aphids		
L	(rose-grain aphid)		·		
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES

		SP	ECIES				
&			Macrosiphum solanifolii (Ashme	ead)			
ЩЩ	lae		(potato aphid)				
HEMIF	Aphidic		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			Probably in <i>Lecanicillium</i> (= <i>Verticillium</i>) <i>lecanii</i> species complex (Reported as <i>Acrostalagmus</i> sp.) (Ascomycetes: Anamorph of Hypocreales)	1955	UNITED STATES OF AMERICA Ex. USA (Hawaii). Released in Maine. Diseased aphids found 3 weeks after release and one infected specimen found in 1958. Unknown if permanently established.	Ν	132, 133
	FA	MIL	.Y				
			Aleyrodidae				
		SP	ECIES				
			Singhiella citrifolii (Morgan) (= L	Dialeurodes	citrifolii Morgan)		
			(cloudywinged whitefly)				
			BIOLOGICAL CONTROL AGENT		COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			BIOLOGICAL CONTROL AGENT	DAIL		•••••	

SI	PECIES <i>Dialeurodes citri</i> (Ashmead) (citrus whitefly)				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	Aschersonia spp. (Ascomycetes: Anamorphs of Hypocreales)	1960-1964	GEORGIA Ex. 11 species and forms from China, Trinidad, Vietnam, Cuba, Japan & India. No information on whether all strains became established. The most aggressive was <i>A.</i> <i>placenta</i> Berkeley & Broome from Vietnam and China, giving up to 90% infection in Adzharia in favorable weather but was inhibited by drought. In 1980-84, in areas where large complexes of natural enemies occurred in citrus plantations (including fungi in this genus), the pest was kept below the economic threshold.	I	70, 120
		1961-1964	AZERBAIJAN Ex. 11 species and forms from China, Trinidad, Vietnam, Cuba, Japan, USA & India. No information on whether all strains became established. About 80% larval mortality in citrus plantations under favorable conditions and fungus spread to new plantations. In 1980-84, in areas where large complexes of natural enemies occurred in citrus plantations (including fungi in this genus), the pest was kept below the economic threshold.	Ι	70, 122
6					
5	Dialeurodes sp.				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	Aschersonia aleyrodis Webber (Ascomycetes: Anamorph of Hypocreales)	1926	BERMUDA Ex. USA (Florida). Establishment and persistence not reported.	?	111

eurothrixus ly) RELEASE DATE before 1920 RELEASE DATE 1909	floccosus (Maskell) COUNTRY, SOURCE, AND RESULTS FROM INTRO. VIRGIN ISLANDS Ex. Unknown. No establishment due to high winds and drought. COUNTRY, SOURCE, AND RESULTS FROM INTRO. UNITED STATES OF AMERICA Ex. USA (Elorida)	PEST ORIGIN N? PEST ORIGIN ?	REFERENCES 154 REFERENCES 81
ly) RELEASE DATE before 1920 RELEASE DATE 1909	COUNTRY, SOURCE, AND RESULTS FROM INTRO. OVIRGIN ISLANDS Ex. Unknown. No establishment due to high winds and drought. COUNTRY, SOURCE, AND RESULTS FROM INTRO. UNITED STATES OF AMERICA Ex. USA (Elorida)	PEST ORIGIN N? PEST ORIGIN ?	REFERENCES 154 REFERENCES 81
RELEASE DATE before 1920 RELEASE DATE 1909	COUNTRY, SOURCE, AND RESULTS FROM INTRO. VIRGIN ISLANDS Ex. Unknown. No establishment due to high winds and drought. COUNTRY, SOURCE, AND RESULTS FROM INTRO. UNITED STATES OF AMERICA Ex. USA (Elorida)	PEST ORIGIN N? PEST ORIGIN ?	REFERENCES 154 REFERENCES 81
RELEASE DATE 1909	VIRGIN ISLANDS Ex. Unknown. No establishment due to high winds and drought. COUNTRY, SOURCE, AND RESULTS FROM INTRO. UNITED STATES OF AMERICA Ex. USA (Elorida)	N? PEST ORIGIN ?	154 REFERENCES
RELEASE DATE 1909	COUNTRY, SOURCE, AND RESULTS FROM INTRO. UNITED STATES OF AMERICA EX. USA (Elorida)	PEST ORIGIN ?	REFERENCES
RELEASE DATE 1909	COUNTRY, SOURCE, AND RESULTS FROM INTRO. UNITED STATES OF AMERICA EX, USA (Elorida)	PEST ORIGIN ?	REFERENCES
RELEASE DATE 1909	COUNTRY, SOURCE, AND RESULTS FROM INTRO. UNITED STATES OF AMERICA EX, USA (Elorida)	PEST ORIGIN ?	REFERENCES
RELEASE DATE 1909	COUNTRY, SOURCE, AND RESULTS FROM INTRO. UNITED STATES OF AMERICA EX, USA (Elorida)	PEST ORIGIN ?	REFERENCES
1909	UNITED STATES OF AMERICA Ex. USA (Florida)	?	81
	One species released in Hawaii against whitefly. Results not reported.		01
_		-	
RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
1928 or before	UNITED STATES OF AMERICA Ex. USA (Florida). Released in Hawaii. Established and provided effective control.	Ι	69, 146
	RELEASE DATE 1928 or before	RELEASE DATECOUNTRY, SOURCE, AND RESULTS FROM INTRO.1928 or beforeUNITED STATES OF AMERICA Ex. USA (Florida). Released in Hawaii. Established and provided effective control.	RELEASE DATECOUNTRY, SOURCE, AND RESULTS FROM INTRO.PEST ORIGIN1928 or beforeUNITED STATES OF AMERICA Ex. USA (Florida). Released in Hawaii. Established and provided effective control.I

HEMIPTERA	Coccidae	SP	ECIES Coccus viridis (Green), Eucalyn (green scale, tessellated scale, BIOLOGICAL CONTROL AGENT Lecanicillium lecanii (Zimmerman) Gams & Zare (= Verticillium lecanii (Zimmerman); = Cephalosporium lecanii Zimmerman) (Ascomycetes: Anamorph of	nnatus tess red wax sca RELEASE DATE 1911	ellatus (Signoret), Ceroplastes Ile) COUNTRY, SOURCE, AND RESULTS FROM INTRO. SEYCHELLES Ex. Sri Lanka. Established and largely controlled scale populations.	rubens (I PEST ORIGIN I	Maskell) REFERENCES 1
		SP	Hypocreales) ECIES Various species of lecaniine sca BIOLOGICAL CONTROL AGENT Lecanicillium lecanii (Zimmerman) Gams & Zare (= Verticillium lecanii (Zimmerman);	ales RELEASE DATE before 1933	COUNTRY, SOURCE, AND RESULTS FROM INTRO. SEYCHELLES Ex. India. Well established on lecaniine scales, especially Coccus	PEST ORIGIN ?	REFERENCES 139
		SP	 Cephalosporium lecanii Zimmerman) (Ascomycetes: Anamorph of Hypocreales) 		viridis (Green) on coffee, and spread widely.		
			Unspecified species of coccids				
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			Unknown fungi	1897	UNITED STATES OF AMERICA Ex. Unrecorded (Unknown). 2 species released in Hawaii against Coccidae. Establishment and spread over most parts of the islands.	?	79

۶A	FA	MIL	.Y Diaspididae				
Π		SP	ECIES				
MIP			Aonidiella aurantii (Maskell)				
H			(California red scale)				
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			Fusarium coccophilum (Desmazieres) Wollenweber & Reinking (= Fusarium episphaerea f. coccophila Tul.); teleomorph = Nectria flammea (Tulasne & Tulasne) Dingley (Ascomycetes: Anamorph of Hypocreales)	1900	ARGENTINA Ex. USA. Established and occasionally caused up to 90% mortality in northeastern and northwestern regions.	1	32
		SP	ECIES <i>Cornuaspis beckii</i> (Newman) (= (purple scale, mussel scale)	Lepidosapi	hes beckii (Newman))		
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			Fusarium coccophilum (Desmazieres) Wollenweber & Reinking (= Sphaerostilbe coccophila Tul.); teleomorph = Nectria flammea (Tulasne & Tulasne) Dingley (Ascomycetes: Anamorph of Hypocreales)	1905 or before	UNITED STATES OF AMERICA Ex. USA (Florida). Introduced to Hawaii. Established and locally abundant but control only partial.	I	80
				1926	BERMUDA Ex. USA (Florida). Establishment and persistence not recorded.	Ι	111
			<i>Podonectria coccicola</i> Petch (Ascomycetes: Tubeufiaceae)	1926	BERMUDA Ex. USA (Florida). Establishment and persistence not recorded.	I	111
			<i>Triblidium caespitosum</i> Cooke & Masee (= <i>Myrangium duriaei</i> Montagne & Berkeley) (Ascomycetes: Triblidiaceae)	1926	BERMUDA Ex. USA (Florida). Establishment and persistence not recorded.	I	111

PECIES				
Aspidiotus destructor Signoret				
(coconut scale)				
BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCE
Fusarium juruanum P. Hennings (= Pseudomicrocera henningsii (Koord.) Petch) (Ascomycetes: Anamorph of Hypocreales)	1929	SEYCHELLES Ex. Sierra Leone. Did not establish.	I	40
PECIES				
Quadraspidiotus perniciosus (C	omstock) (<i>= Aspidiotus perniciosus</i> (Com	stock))	
(San Jose scale)				
BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
Fusarium coccophilum (Desmazieres) Wollenweber & Reinking (= Sphaerostilbe coccophila Tul.); teleomorph = Nectria flammea (Tulasne & Tulasne) Dingley (Ascomycetes: Anamorph of	1897	UNITED STATES OF AMERICA Ex. USA (Florida). Released in California. As a result of this introduction, or a native fungus, scale nearly exterminated in southern California.	I	157
Hypocreales)	1897	UNITED STATES OF AMERICA Ex. USA (Florida). Released in New Jersey. Established, overwintered, with abundant infection the following Sept. but this pathogen alone failed to provide adequate control.	Ι	134, 135
	1898	UNITED STATES OF AMERICA Ex. USA (Florida). Released in Illinois by tying twigs with infected scales to trees. Overwintered and many scales infected but healthy scales still abundant. Hypothesized this fungus could add to effects of other natural enemies to provide a permanent check of scale	Ι	45

ORI	ORDER: THYSANOPTERA							
E.	FAMILY							
		٦	Thripidae					
		SP	ECIES					
	L		Thrips tabaci Lindeman					
	L	_	(onion thrips)					
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES	
			<i>Neozygites parvispora</i> (MacLeod & Carl) Remaudiére & Keller (Zygomycetes: Neozygitaceae)	1973-76	BARBADOS Ex. Switzerland. Released in onion field but no establishment.	Ι	29	

ER	: COLEOPTERA				
ЛГ	Y				
ç	Scarabaeidae				
SP	ECIES				
	Oryctes rhinoceros (L.)				
	(rhinoceros beetle)				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Metarhizium anisopliae</i> (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales)	1939	SAMOA (= WESTERN SAMOA) Ex. Java. This fungal species recovered after release but whether it was the introduced strain or a native strain is uncertain.	Ι	145
		1952	WALLIS ISLAND Ex. Argentina. Results from release unknown.	Ι	145
		1967	TOKELAU ISLANDS Ex. Samoa (= Western Samoa). Results from release unknown.	Ι	145
		1969	TONGA Ex. Samoa (= Western Samoa). High levels of infection directly after release, infections still present 3 years later but prevalence extremely low.	I	145, 160

SF	PECIES				
	(taro beetle)				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Metarhizium anisopliae</i> (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales)	1976	KIRIBATI Ex. Unknown. Released on southern Tarawa by Latch. Establishment not confirmed.	Ι	95
		1995	KIRIBATI Ex. Papua New Guinea. Released on southern Tarawa. Persisted in soil through 2003, spread and exerted some control.	I	95, 149
SP	PECIES Phyllophaga smithi (Arrow) (= L smithi (Arrow)	achnostern	a smithi (Arrow); = Clemora sı	nithi (Arro	ow); = Phytalus
	(sugar cane white grub)				
		RELEASE	COUNTRY, SOURCE, AND	PEST	
	BIOLOGICAL CONTROL AGENT	DATE	RESULTS FROM INTRO.	ORIGIN	REFERENCES
	BIOLOGICAL CONTROL AGENT Beauveria bassiana (Balsamo) Vuillemin (= Beauveria densa (Link) Vuillemin) (Ascomycetes: Anamorph of Hypocreales)	DATE 1932	RESULTS FROM INTRO. MAURITIUS Ex. Unknown isolate from Imperial Bureau of Mycology, United Kingdom. Host population gradually declined and diseases may have played a part.	ORIGIN I	REFERENCES 54, 102, 103, 104
	BIOLOGICAL CONTROL AGENT Beauveria bassiana (Balsamo) Vuillemin (= Beauveria densa (Link) Vuillemin) (Ascomycetes: Anamorph of Hypocreales)	DATE 1932	RESULTS FROM INTRO. MAURITIUS Ex. Unknown isolate from Imperial Bureau of Mycology, United Kingdom. Host population gradually declined and diseases may have played a part.	ORIGIN I	REFERENCES 54, 102, 103, 104
	BIOLOGICAL CONTROL AGENT Beauveria bassiana (Balsamo) Vuillemin (= Beauveria densa (Link) Vuillemin) (Ascomycetes: Anamorph of Hypocreales) Metarhizium anisopliae (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales)	DATE 1932 1932	RESULTS FROM INTRO. MAURITIUS Ex. Unknown isolate from Imperial Bureau of Mycology, United Kingdom. Host population gradually declined and diseases may have played a part. MAURITIUS Ex. Unknown isolate from Imperial Bureau of Mycology, UK. Host population gradually declined and diseases may have played a part.	I	REFERENCES 54, 102, 103, 104 54, 102, 103, 104
	BIOLOGICAL CONTROL AGENT Beauveria bassiana (Balsamo) Vuillemin (= Beauveria densa (Link) Vuillemin) (Ascomycetes: Anamorph of Hypocreales) Metarhizium anisopliae (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales)	DATE 1932 1932	RESULTS FROM INTRO. MAURITIUS Ex. Unknown isolate from Imperial Bureau of Mycology, United Kingdom. Host population gradually declined and diseases may have played a part. MAURITIUS Ex. Unknown isolate from Imperial Bureau of Mycology, UK. Host population gradually declined and diseases may have played a part.	I	REFERENCES 54, 102, 103, 104 54, 102, 103, 104
SP	BIOLOGICAL CONTROL AGENT Beauveria bassiana (Balsamo) Vuillemin (= Beauveria densa (Link) Vuillemin) (Ascomycetes: Anamorph of Hypocreales) Metarhizium anisopliae (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales) PECIES Alissonotum impressicolle Arro	DATE 1932 1932 w	RESULTS FROM INTRO. MAURITIUS Ex. Unknown isolate from Imperial Bureau of Mycology, United Kingdom. Host population gradually declined and diseases may have played a part. MAURITIUS Ex. Unknown isolate from Imperial Bureau of Mycology, UK. Host population gradually declined and diseases may have played a part.	I	REFERENCES 54, 102, 103, 104 54, 102, 103, 104
SF	BIOLOGICAL CONTROL AGENT Beauveria bassiana (Balsamo) Vuillemin (= Beauveria densa (Link) Vuillemin) (Ascomycetes: Anamorph of Hypocreales) Metarhizium anisopliae (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales) PECIES Alissonotum impressicolle Arro BIOLOGICAL CONTROL AGENT	DATE 1932 1932 1932 w RELEASE DATE	RESULTS FROM INTRO. MAURITIUS Ex. Unknown isolate from Imperial Bureau of Mycology, United Kingdom. Host population gradually declined and diseases may have played a part. MAURITIUS Ex. Unknown isolate from Imperial Bureau of Mycology, UK. Host population gradually declined and diseases may have played a part. COUNTRY, SOURCE, AND RESULTS FROM INTRO.	ORIGIN 	REFERENCES 54, 102, 103, 104 54, 102, 103, 104 REFERENCES

	SP	ECIES				
		Dermolepida albohirtum (Waterh	nouse)			
dae		(greyback cane beetle)				
arabaeid		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
Sci		<i>Metarhizium anisopliae</i> (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales)	about 1914	AUSTRALIA Ex. Samoa. Released in Queensland but before release, had already been found infecting this host in Queensland. Post release, at times considerable numbers of grubs of intended host and <i>Rhabdoscelus obscurus</i> (Boisduval) killed by this fungus.	Ν	155
	~ ~	ECIES				
	SP					
	SP	Lepidiota pruinosa Wied., Leuco	opholis irro	rata Chevrolat		
	SP	Lepidiota pruinosa Wied., Leuco (white grubs in sugar cane)	opholis irro	rata Chevrolat		
	SP	Lepidiota pruinosa Wied., Leuco (white grubs in sugar cane) BIOLOGICAL CONTROL AGENT	RELEASE DATE	rata Chevrolat COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	SP	Lepidiota pruinosa Wied., Leuco (white grubs in sugar cane) BIOLOGICAL CONTROL AGENT Metarhizium anisopliae (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales)	RELEASE DATE 1928	rata Chevrolat COUNTRY, SOURCE, AND RESULTS FROM INTRO. PHILIPPINES Ex. Australia (Queensland). Not effective control and "undoubtedly already present."	PEST ORIGIN N	REFERENCES 125
	SP	Lepidiota pruinosa Wied., Leuco (white grubs in sugar cane) BIOLOGICAL CONTROL AGENT Metarhizium anisopliae (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales)	RELEASE DATE 1928	rata Chevrolat COUNTRY, SOURCE, AND RESULTS FROM INTRO. PHILIPPINES Ex. Australia (Queensland). Not effective control and "undoubtedly already present."	PEST ORIGIN N	REFERENCES 125
	SP	Lepidiota pruinosa Wied., Leuco (white grubs in sugar cane) BIOLOGICAL CONTROL AGENT Metarhizium anisopliae (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales) ECIES	RELEASE DATE 1928	rata Chevrolat COUNTRY, SOURCE, AND RESULTS FROM INTRO. PHILIPPINES Ex. Australia (Queensland). Not effective control and "undoubtedly already present."	PEST ORIGIN N	REFERENCES 125
	SP	Lepidiota pruinosa Wied., Leuco (white grubs in sugar cane) BIOLOGICAL CONTROL AGENT Metarhizium anisopliae (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales) ECIES Various scarabs (including Anop	RELEASE DATE 1928	rata Chevrolat COUNTRY, SOURCE, AND RESULTS FROM INTRO. PHILIPPINES Ex. Australia (Queensland). Not effective control and "undoubtedly already present." sp., Lepidiota sp., etc.)	PEST ORIGIN N	REFERENCES 125
	SP	Lepidiota pruinosa Wied., Leuco (white grubs in sugar cane) BIOLOGICAL CONTROL AGENT Metarhizium anisopliae (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales) ECIES Various scarabs (including Anop (white grubs in sugar cane)	RELEASE DATE 1928	rata Chevrolat COUNTRY, SOURCE, AND RESULTS FROM INTRO. PHILIPPINES Ex. Australia (Queensland). Not effective control and "undoubtedly already present." sp., Lepidiota sp., etc.)	PEST ORIGIN N	REFERENCES 125
	SP	Lepidiota pruinosa Wied., Leuco (white grubs in sugar cane) BIOLOGICAL CONTROL AGENT Metarhizium anisopliae (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales) ECIES Various scarabs (including Anop (white grubs in sugar cane) BIOLOGICAL CONTROL AGENT	RELEASE DATE 1928 DIognathus RELEASE DATE	rata Chevrolat COUNTRY, SOURCE, AND RESULTS FROM INTRO. PHILIPPINES Ex. Australia (Queensland). Not effective control and "undoubtedly already present." sp., Lepidiota sp., etc.) COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN N PEST ORIGIN	REFERENCES 125 REFERENCES

		SP	FCIES				
RA	aeidae	0.	Adoretus tenuimaculatus Water	house			
OPTE			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
COLE	Scarab		<i>Metarhizium anisopliae</i> (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales)	before 1918	FIJI Ex. Unknown. Some signs that this fungus acted as a check on the beetles.	Ι	78
. :	FA	MIL	Y Curculionidae				
		0.0					
		SP	ECIES Sitona discoidous Gyllonhal				
			(Sitona weevii)				
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			<i>Beauveria bassiana</i> (Balsamo) Vuillemin (Ascomycetes: Anamorph of Hypocreales)	1984	AUSTRALIA Ex. France (Montpellier). Released in southern Australia. No infections ever found.	I	3
		SP	ECIES				
			Otiorhynchus nodosus (Müller)	, Otiorhyncl	hus arcticus (Fabricius)		
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			<i>Metarhizium anisopliae</i> (Metschnikoff) Sorokin (Ascomycetes: Anamorph of Hypocreales)	2003	ICELAND Ex. Faroe Islands (Havnardalur). Released in eroded areas in Haukadalur. Too early to determine effect.	Ν	108

AMII	Y					
_	Culicidae					
SPECIES						
	Aedes polynesiensis Marks					
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES	
	<i>Coelomomyces stegomyiae</i> Keilin (Chytridiomycetes: Coelomomycetaceae)	1958	TOKELAU ISLANDS Ex. Singapore. Released on Nukunonu Atoll. Established, by 1963 infected larvae found in 13 of 35 habitats.	Ν	63, 82	
SF	PECIES Culex tarsalis Coquillett					
	(western encephalitis mosquito)					
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES	

* Member of the Kingdom Chromista, Phylum Oomycota. All other entries in this list belong to in the Kingdom Fungi.

OR	RDER	R: LEPIDOPTERA				
	FAMII	LY				
	_	Lymantriidae				
	SF	PECIES				
		Lymantria dispar (L.)				
		(gypsy moth)				
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
		<i>Entomophaga maimaiga</i> Humber, Shimazu & Soper (Zygomycetes: Entomophthoraceae)	1910-1911	UNITED STATES OF AMERICA Ex. Japan (Nishigahara). Released in Massachusetts (Boston area). Not established.	I	138
			1985	UNITED STATES OF AMERICA Ex. Japan (Ishikawa Prefecture). Released in southwestern New York State. Not established.	Ι	60
			1986	UNITED STATES OF AMERICA Ex. Japan (Ishikawa Prefecture). Released in northern Virginia. Not established.	I	60
	l		1996	BULGARIA Ex. USA (Connecticut). Released in Levishte (northeastern Bulgaria). No infection in 1997.	Ν	117
	l		1999	BULGARIA Ex. USA (Massachusetts). Released in Karlovo (central Bulgaria). Established but negligible control.	Ν	118
			2000	BULGARIA Ex. USA (Connecticut). Released in Levishte. Infections found in 2002, 2003 & 2004.	Ν	117
			2002	RUSSIA Ex. USA (Virginia). Released in Novosibirsk region. Establishment not confirmed.	Ν	2

SUB	UBCLASS: ACARI							
FA	MI	LY						
		Eriophyidae						
	SF	PECIES						
		Eriophyes sheldoni (Ewing)						
		(citrus bud mite)						
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES		
I		<i>Hirsutella thompsonii</i> Fisher var. <i>vinacea</i> Samson, McCoy & O'Donnell (Ascomycetes: Anamorph of Hypocreales)	1985	ARGENTINA Ex. USA (North Carolina). Released on lemon trees in Tucuman. Initially 92% decrease in mites but persistence unknown.	Ι	136, 137		
	L							
	SF	PECIES						
	Eriophyes sheldoni (Ewing), Phyllocoptruta oleivora (Ashmead)							
		(citrus bud mite, citrus rust mite)					
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES		
		<i>Hirsutella thompsonii</i> Fisher var. <i>synnematosa</i> Samson, McCoy & O'Donnell (Ascomycetes: Anamorph of Hypocreales)	1985	ARGENTINA Ex. Zimbabwe. Released in Tucuman. About 50% infection for both mite species after release but persistence unknown.	Ι	136, 137		
FA	AMIL	LY Tetrancvchidae						
	SP	PECIES						
		Mononychellus tanajoa (Bondar)					
		(cassava green mite)	•					
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES		
		Neozygites tanajoae Delalibera, Hajek & Humber (prev. referred to as Neozygites floridana (Weiser & Muma) Remaudiére & Keller) (Zygomycetes: Neozygitaceae)	1998-1999	BENIN Ex. northeastern Brazil. Established, epizootics occurring in 2002 & 2003 at release sites. Molecular probes developed to confirm that epizootics were caused by exotic strains of the introduced pathogen rather than a closely related native strain.	I	36, 66		

TABLE D: EXOTIC MICROSPORIDIA RELEASED AND TARGET HOSTS

ORD	E	R: ORTHOPTERA						
F/	١M	LY						
	Acrididae							
	SPECIES							
	Various species. Principal targets are Melanoplinae: <i>Dichroplus maculipennis</i> (Blanchard), <i>Dichroplus elongatus</i> (Giglio-Tos), <i>Dichroplus pratensis</i> Bruner, <i>Scotussa lemniscata</i> Stål							
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES		
		Paranosema locustae (Canning) (= Nosema locustae Canning; = Antonospora locustae (Canning)) (Microsporidia: Nosematidae)	1978-82	ARGENTINA Ex. USA (Idaho). Released in central Argentina. Established and, in 1994-5, found 75 km from release sites. Epizootics occur with accompanying host declines but levels of infection in susceptible species usually average < 10%.	Ν	84		
OR	DE	R: DIPTERA						
F/	١MA	LY						
		Culicidae						
	S	PECIES						
		Culex pipiens quinquefasciatus	Say (= <i>C. pi</i>	ipiens fatigans Wiedemann; = (C. fatigans	s Wiedemann)		
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES		

Pleistophora culicis (= Plistophora 1967 *culicis)* Weiser (Microsporidia: Pleistophoridae)

- NAURU Ex. Nigeria (Lagos). Establishment not confirmed.
- Ν 82

ORD	ER	R: LEPIDOPTERA				
FA	MII	LY				
	_	Pyralidae				
	SF	PECIES				
		Ostrinia nubilalis (Hübner)				
		(European corn borer)				
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
		Nosema pyrausta (Paillot) (= Perezia pyraustae Paillot; = Glugea pyraustae (Paillot)) (Microsporidia: Nosematidae)	Unk betw. 1952 & 1960	UNITED STATES OF AMERICA Ex. USA (Iowa). Infected larvae distributed at scattered localities throughout Illinois. Disease became prevalent and kept host populations at low levels.	Ι	37
	_					
FA	MII	LY				
		Lymantriidae				
	SF	PECIES				
		Lymantria dispar (L.)				
					DEOT	
		BIOLOGICAL CONTROL AGENT	DATE	RESULTS FROM INTRO.	ORIGIN	REFERENCES
I		Nosema portugal Maddox & Vávra (= <i>Microsporidium</i> sp.) (Microsporidia: Nosematidae)	1986	UNITED STATES OF AMERICA Ex. Portugal. Released in Maryland. Established, low levels of infection in 1987 which persisted for 3 years.	Ι	72, 73, 94
			1992, 1993	UNITED STATES OF AMERICA Ex. Portugal. Released in Michigan. Low levels of infection during the season of release, persistence not confirmed.	I	6, 7
		<i>Endoreticulatus</i> sp. (= <i>Vavraia</i> sp.) (Microsporidia: Pleistophoridae)	1986	UNITED STATES OF AMERICA Ex. Portugal. Released in Maryland. Not established.	I	72, 73, 94

TABLE E: EXOTIC NEMATODES RELEASED AND TARGET HOSTS

DER	: ORTHOPTERA				
FAMII	LY				
	Gryllotalpidae				
SF	'ECIES				
	Scapteriscus abbreviatus Scudo	ler, Scapteri	iscus borelli Gigli-Tos, Scapter	riscus vic	inus Scudder
	(mole crickets)				
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Steinernema scapterisci</i> Nguyen & Smart (Rhabditida: Steinernematidae)	1985	UNITED STATES OF AMERICA Ex. Uruguay. Released in Florida. Established, host populations declined by 85-98%, by 1988 infected hosts collected 23 km from release site. Establishment on golf courses not as successful but > 27% reduction in hosts when	Ι	113

ORI	DER	R: COLEOPTERA				
F	AMII	LY				
		Scarabaeidae				
	SF	PECIES				
		Oryctes rhinoceros (L.)				
		(rhinoceros beetle)				
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
1		<i>Rhabditis</i> sp. (Rhabditida: Rhabditidae)	1954	FIJI Ex. Sri Lanka. Results of release not reported.	Ι	28
I			1957	FIJI Ex. Madagascar. Recovered after release, persistence not confirmed.	I	145
I			1957	AMERICAN SAMOA Ex. Madagascar. Results from release unknown.	Ι	145
		Rhabditis sp. nr. maupasi Seurat in Maupas (Rhabdita: Rhabditidae)	1957	SAMOA (= WESTERN SAMOA) Ex. Sri Lanka. Results from release unknown.	I	145

COLEOPTERA	Scarabaeidae	Oryctes rhino. (L.)	<i>Rhabditis</i> sp.	1957 1957	AMERICAN SAMOA Ex. Sri Lanka. Results from release unknown. WALLIS ISLAND Ex. Sri Lanka. Results from release unknown.	I	145 145
	FA	MIL	_Y				
			Curculionidae				
		SP	PECIES				
			Sitona discoideus Gyllenhal				
			(sitona weevil)				
			BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			<i>Heterorhabditis heliothidis (</i> Khan, Brooks & Hirschmann) (Rhabditida: Heterorhabditidae)	1982	AUSTRALIA Ex. New Zealand. Released in So. Australia. No infections ever found.	I	3

ORDER: DIPTERA

			ī.	1.2	
E7	ΔU	M	I	L	Y

	Culicidae				
SP	PECIES				
	Culex pipiens quinquefasciatus	Say			
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Romanomermis culicivorax</i> Ross & Smith (= <i>Reesimermis nielseni</i> Tsai & Grundmann) (Mermithidae)	1971-72	TAIWAN Ex. USA (Louisiana). Released in Taipei. Infection rates low after release and no indication of establishment.	Ν	100
		1974	TAIWAN Ex. USA (Louisiana). Released in Taipei. Recycling occurred through 196 days after release but continued persistence not confirmed.	Ν	27
		1972	THAILAND Ex. USA (Louisiana). Large releases in ditches and drains in Bangkok, infection 0-27%, no recycling.	Ν	26 (in 115)

	Aedes spp., Ocnierotatus spp. (10 species t	otal)		
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Romanomermis culicivorax</i> Ross & Smith (= <i>Reesimermis nielseni</i> Tsai & Grundmann) (Mermithidae)	1974	CANADA Ex. USA (Louisiana). Released in snow melt pools in Winnipeg, Manitoba. No infection.	Ν	49 (in 115)
		1975-76	CANADA Ex. USA (Louisiana). Released in snow melt pools in Winnipeg, Manitoba. Meager parasitism after one winter and continued persistence questionable.	Ν	50
Γ					
SI	PECIES				
	Aedes polynesiensis Marks, Aed	des aegypti	(L.)		
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Romanomermis culicivorax</i> Ross & Smith (= <i>Reesimermis nielseni</i> Tsai & Grundmann) (Mermithidae)	1978	TOKELAU ISLANDS Ex. USA (Louisiana). Released on Fakatao Atoll in tree holes and man-made containers. Established in 35 of 41 sites with 14-22% infection. Persisted at least 3 years.	N/I	83
SF	PECIES				
	Anopheles nyssorhynchus albir	nanus Wied	emann		
	BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
	<i>Romanomermis culicivorax</i> Ross & Smith (= <i>Reesimermis nielseni</i> Tsai & Grundmann)	1983	COLOMBIA Ex. USA (Louisiana). Released in El Valle. Established and cycled	Ν	127

among schoolchildren.

SPECIES

DIPTERA Culicidae Anopheline species: Anopheles dthali Patton, Anopheles superpictus Grassi, Anopheles sergentii (Theobald), Anopheles turkhudi Liston, Anopheles culicifacies Giles

BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
<i>Romanomermis culicivorax</i> Ross & Smith (<i>= Reesimermis nielseni</i> Tsai & Grundmann) (Mermithidae)	1984-85	IRAN Ex. USA (Louisiana). Established, 56-61% parasitism immediately post-release but only minor reductions in host populations. 8% parasitism at 1 of 13 sites 1 year after release. Effective long-term control unlikely.	Ν	162

SPECIES

Anopheles punctipennis (Say), Anopheles crucians Weidemann, Aedes vexans (Meig.), Culex restuans Theobald, Culex pipiens L.

BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
<i>Romanomermis culicivorax</i> Ross & Smith (= <i>Reesimermis nielseni</i> Tsai & Grundmann) (Mermithidae)	1975	UNITED STATES OF AMERICA Ex. USA (Louisiana). Released in Maryland. Established, 50-100% host mortality even 2 years after release.	Ν	106

SPECIES

Anopheles freeborni Aitken, Culex tarsalis Coquillett

BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
<i>Romanomermis culicivorax</i> Ross & Smith (= <i>Reesimermis nielseni</i> Tsai & Grundmann) (Mermithidae)	1975-76	UNITED STATES OF AMERICA Ex. USA (Louisiana). Released in California rice fields. Continuous partial control through rice growing season with mean weekly infection for both species > 60%. Survived chemicals, drying, harvest, winter and cultivation and parasitized hosts next summer.	N	115

		SP	ECIES				
			Anopheles nyssorhynchus albin	nanus Weide	emann, Anopheles punctipenni	s (Say)	
TERA	cidae		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
DIP	Culi		<i>Romanomermis culicivorax</i> Ross & Smith (<i>= Reesimermis nielseni</i> Tsai & Grundmann) (Mermithidae)	1977	EI SALVADOR Ex. USA (Louisiana). Released in Lake Apasteque. Releases through year yielded 46-96% parasitism; up to 17x reduction in host populations. Recycling not reported, questionable.	N/I?	116, 127
		SP	PECIES				
			Culex pipiens quinquefasciatus	Sav. Aedes	aegypti (L.)		
			BIOLOGICAL CONTROL AGENT	RELEASE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			<i>Octomyomermis muspratti</i> Obiamiwe & Macdonald (Mermithidae)	1967	NAURU Ex. Zambia. After release, parasitism found in several tree holes but long-term establishment not reported.	N/I	82, 119

ORDER: LEPIDOPTERA

E	Λ	N	Л	I	L	v
		Π	Т	Ц	_	

Lymantriidae

SPECIES

Lymantria dispar (L.)

(gypsy moth)

BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
<i>Hexamermis</i> sp. (Mermithidae)	1974	UNITED STATES OF AMERICA Ex. Austria. Released in New Jersey. Not established.	Ι	30
	1976	UNITED STATES OF AMERICA Ex. Japan (Hokkaido). Released in Pennsylvania. Not established.	Ι	30

OR	DRDER: HYMENOPTERA								
F	AMI	LY							
	_	Siricidae							
	SF	PECIES							
		Sirex noctilio F.							
		(European woodwasp, Sirex was	sp)						
		BIOLOGICAL CONTROL AGENT	RELEASE DATE	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES			
		<i>Deladenus siricidicola</i> Bedding (= <i>Beddingia siricidicola</i> (Bedding)) (Rhabditida: Neotylenchidae)	1967	NEW ZEALAND Ex. New Zealand (North Island). Released on South Island. 29-76% infection reported within first year of release and, by 1970, natural spread of ca. 50 km. Releases continued at least through 1974. Lack of establishment at some sites linked with low density host populations.	Ι	170			
			1970	AUSTRALIA Ex. Hungary. Released in Tasmania. Established, reached high levels of parasitism rapidly. In one forest, six years after release of 50 parasitized females, trees were no longer being killed by woodwasps. Spread to nearby forests and also released in other areas. Considered the key biological agent controlling <i>Sirex</i> .	Ι	9, 64			
			1971	AUSTRALIA Ex. Tasmania (plus other locations). Released in Victoria. Established, dispersed by woodwasps in local forests and by humans between forests. Use of this nematode became a cornerstone in the National Sirex Control strategy. Released over many years in many areas; 147,000 radiata pines inoculated in the Green Triangle in 1987 alone. With over 20 years of in vitro production, strain lost virulence resulting in replacement of strain used for releases.	Ι	11, 64			

		Bedding	1987	URUGUAY Ex. New Zealand. Established, yielding 18% parasitism.	l	17, 121, 128
HYMENOPTERA	Sirex noctilio F.	Deladenus siricidicola	1989, 1990, 1994	BRAZIL Ex. Australia. Principally released in 3 southern provinces. After loss of infectivity, new strain (Kamona from Tasmania) introduced in 1994, yielding 50-80% parasitism. Overall established, parasitism levels variable but provided substantial control in some areas.	Ι	11, 67, 68
			1995	SOUTH AFRICA Ex. Australia. Released Kamona strain in southwestern Cape Province. Established, with 23% parasitism reported in 1996. In 1998, along with cultural control, credited with containing the spread of the pest.	Ι	150, 151
			1999	ARGENTINA Ex. Brazil. Released in Patagonia. Established, 50-60% parasitism reported at release site in 2000.	I	76, 77

TABLE F: ACCIDENTAL INTRODUCTIONS OF PATHOGENS AND NEMATODES

ORD	DRDER: LEPIDOPTERA								
FA	FAMILY								
	Zygaenidae								
	SP	PECIES							
		Harrisina brillians Barnes & McI	Dunnough						
		(western grapeleaf skeletonizer)							
		BIOLOGICAL CONTROL AGENT	YEAR FOUND	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES			
		<i>Harrisina brillians</i> Granulovirus (HbGV) (Baculoviridae)	early 1950s	UNITED STATES OF AMERICA Found in San Diego Co., California, probably inadvertently introduced with parasitoids from Mexico and USA (Arizona). Infections observed in field and virus continually wiped out colonies for rearing parasitoids	Ι	143, 144			
				for rearing paraonolido.					
E									
FA	MIL	_Y							
FA	MIL	_Y Pyralidae							
FA	AMIL SP	-Y Pyralidae PECIES							
FA	AMIL SP	-Y Pyralidae PECIES Ostrinia nubilalis (Hübner)							
FA	AMIL SP	-Y Pyralidae PECIES <i>Ostrinia nubilalis</i> (Hübner) (European corn borer)							
FA	SP	-Y Pyralidae PECIES Ostrinia nubilalis (Hübner) (European corn borer) BIOLOGICAL CONTROL AGENT	YEAR FOUND	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES			

	FAMILY									
RA			Lymantriidae							
H		SP	ECIES							
ğ			<i>Lymantria dispar</i> (L.)							
Π			(gypsy moth)							
۳			BIOLOGICAL CONTROL AGENT	YEAR FOUND	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES			
			Entomophaga maimaiga Humber, Shimazu & Soper (Zygomycetes: Entomophthoraceae)	1989	UNITED STATES OF AMERICA Ex. Japan. First found in 1989 in 7 northeastern states but spread naturally and through releases. Established in Connecticut, Delaware, Maine, Massachusetts, Maryland, Michigan, North Carolina, New Hampshire, New Jersey, New York, Ohio, Pennsylvania, Rhode Island, Virginia, Vermont, Wisconsin, West Virginia and in Ontario, Canada. Host populations remain low the majority of years and sites, although localized increases can occur infrequently.	I	42, 57, 60, 107			
			<i>Lymantria dispar</i> Multicapsid Nucleopolyhedrovirus (LdMNPV) (Baculoviridae)	1907	UNITED STATES OF AMERICA (probably Massachusetts). Thought to have been introduced after 1900 from Europe with parasitoids released for classical biological control or with plant material and spread through the host population. Until <i>E. maimaiga</i> became established, caused epizootics in high density, defoliating host populations, resulting in spectacular population crashes. Spreads naturally after the host population spreads into new areas.	I	51, 57			

ORD	RDER: HYMENOPTERA							
FA	FAMILY							
		Siricidae						
	SP	PECIES						
		Sirex noctilio F.						
		(European woodwasp, Sirex was	p)					
		BIOLOGICAL CONTROL AGENT	YEAR FOUND	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES		
		<i>Deladenus siricidicola</i> Bedding (= <i>Beddingia siricidicola</i> (Bedding)) (Rhabditida: Neotylenchidae)	1962	NEW ZEALAND (North Island). Thought to have arrived with host. Attributed with being the most important agent controlling host on the North Island.	Ι	10, 169		
FA	MIL	_Y						
		Diprionidae	_		_			
	SP	'ECIES		unio e Haustin)				
		(Europeon opruoe cowfly)	rion nercy	iniae Hartig)				
			VEAD		DEST			
		BIOLOGICAL CONTROL AGENT	FOUND	RESULTS FROM INTRO.	ORIGIN	REFERENCES		
		<i>Gilpinia hercyniae</i> Nucleopolyhedrovirus (GhNPV) (Baculoviridae)	1936	CANADA and UNITED STATES OF AMERICA Probably ex. Europe. Believed introduced with parasitoids. Virus first found in New Brunswick and then Maine, Vermont and New Hampshire, after which it spread from south to north and first found in Quebec in 1940. Also transfered to sites in Quebec and Ontario but some transfers were unsuccessful and virus spread on its own. By 1942, virus was distributed throughout most of the infested areas and was credited as cause of rapid decline in pest outbreak after 1942. Virus plus parasitoids appear to have permanently solved problems due to this pest in eastern North America.	Ι	5, 21, 34, 35, 92		

HYMENOPTERA	Diprionidae	Gilpinia hercyniae (Hartig)	GhNPV	1970 or 1971	UNITED KINGDOM (WALES) Probably ex. Europe. Spread from small epicenter and controlled pest outbreak by 1974.	Ι	35, 43
	EA	мп	v				
			-I				
	Г	_	Formicidae	_		_	
		SP	PECIES				
			Solenopsis invicta Buren				
			(red imported fire ant)				
			BIOLOGICAL CONTROL AGENT	YEAR FOUND	COUNTRY, SOURCE, AND RESULTS FROM INTRO.	PEST ORIGIN	REFERENCES
			<i>Thelohania solenopsae</i> Knell, Allen & Hazard (Microsporidia: Thelohaniidae)	1996	UNITED STATES OF AMERICA Ex. South America. Found in Florida. Infects mostly polygynous colonies. Chronic debilitation of infected queens yields smaller colony sizes and possibly prolonged death of colonies.	I	112, 153

CHART 1: CLASSIFICATION OF PATHOGENS AND NEMATODES PURPOSEFULLY INTRODUCED FOR CLASSICAL BIOLOGICAL CONTROL OF INSECTS AND MITES OR ESTABLISHED AFTER ACCIDENTAL INTRODUCTION¹

Virus

Family Baculoviridae
Agrotis segetum Granulovirus (AsGV)
Anticarsia gemmatalis Multicapsid Nucleopolyhedrovirus (AgMNPV)
Gilpinia hercyniae Nucleopolyhedrovirus (GhNPV)
Harrisina brillians Granulovirus (HbGV)
Lymantria dispar Multicapsid Nucleopolyhedrovirus (LdMNPV)
Lymantria monacha Nucleopolyhedrovirus
Neodiprion sertifer Nucleopolyhedrovirus (NeseNPV)
Pseudoplusia includens Singlecapsid Nucleopolyhedrovirus (PiSNPV)
Trichoplusia ni Nucleopolyhedrovirus (TnNPV)
Family Unassigned
Oryctes rhinoceros Virus (Orv)

Domain Bacteria

Phylum Firmicutes Class Bacilli Order Bacillales Family Bacillaceae Paenibacillus popilliae

Domain Eukarya

Kingdom Fungi Phylum Chytridiomycota Class Chytridiomycetes Order Blastocladiales Family Coelomomycetaceae Coelomomyces stegomyiae Phylum Zygomycota Class Zygomycetes Order Entomophthorales Family Entomophthoraceae Entomophaga grylli Entomophaga maimaiga Pandora neoaphidis Zoophthora radicans Family Neozygitaceae Neozygites fresenii Neozygites parvispora Neozygites tanajoae

Phylum Ascomycota Class Loculoascomycetes Order Pleosporales Family Tubeufiaceae Podonectria coccophila Class Discomycetes Order Rhytismatales Family Triblidiaceae Triblidium caespitosum Class Pyrenomycetes Order Hypocreales (anamorphic/asexual forms) Aschersonia aleyrodis Aschersonia goldiana Aschersonia spp. Beauveria bassiana Beauveria brongniartii Fusarium coccophilum Fusarium juruanum Hirsutella thompsonii var. synnematosa Hirsutella thompsonii var. vinacea Lecanicillium lecanii Metarhizium anisopliae Phylum Microsporidia Class Microsporea Order Nosematidida Family Nosematidae Nosema portugal Nosema pyrausta Paranosema locustae Order Microsporida Family Pleistophoridae Endoreticulatus sp. Pleistophora culicis Family Thelohaniidae Thelohania solenopsae Kingdom Chromista (= Kingdom Straminipila)

Phylum Oomycota Class Oomycetes Order Pythiales Family Pythiaceae *Lagenidium giganteum* Kingdom Animalia Phylum Nematoda Class Chromadorea Order Rhabditida Family Steinernematidae Steinernema scapterisci Family Neotylenchidae Deladenus siricidicola Family Rhabditidae Rhabditis sp. Rhabditis sp. nr. maupasi Family Heterorhabditidae Heterorhabditis heliothidis Class Enoplea Order Mermithida Family Mermithidae Hexamermis sp. Octomyomermis muspratti Romanomermis culicivorax

¹ Organization of domains based on Woese et al. (156). Placement of the Phylum Microsporidia in the Kingdom Fungi is a recent change (75) and, while we have adopted this change here, the higher order affiliation of this group is still somewhat in a state of flux. Nematode classification based on De Ley & Blaxter (38).

CHART 2: CLASSIFICATION OF INSECTSAND MITES TARGETED BY CLASSICAL BIOLOGICAL CONTROL PROGRAMS USING PATHOGENS OR NEMATODES, OR HOSTS OF ACCIDENTAL INTRODUCTIONS OF PATHOGENS OR NEMATODES

Kingdom Animalia Phylum Arthropoda Class Insecta Order Orthoptera Family Gryllotalpidae Scapteriscus abbreviatus Scapteriscus borelli Scapteriscus vicinus Family Acrididae Camnula pellucida Dichroplus elongatus Dichroplus maculipennis Dichroplus pratensis Melanoplus bivittatus Melanoplus sanguinipes Phaulacridium vittatum Scotussa lemniscata Order Hemiptera Family Cercopidae Aeneolamia flavilatera Family Cicadellidae Empoasca fabae Family Aphididae Aphis gossypii Macrosiphum solanifolii Metopolophium dirhodum Therioaphis maculata Family Aleyrodidae Aleurodicus cocois Aleurothrixus floccosus Dialeurodes sp. Dialeurodes citri Singhiella citrifolii Family Coccidae Ceroplastes rubens Coccus viridis Eucalymnatus tessellatus Family Diaspididae Aonidiella aurantii Aspidiotus destructor Cornuaspis beckii Quadrispidiotus perniciosus Order Thysanoptera Family Thripidae Thrips tabaci

Order Coleoptera Family Scarabaeidae Adoretus tenuimaculatus Alissonotum impressicolle Anoplognathus sp. Cochliotis melolonthoides Dermolepida albohirtum Lepidiota sp. Lepidiota pruinosa Leucopholis irrorata Oryctes monoceros Oryctes rhinoceros Papuana huebneri Phyllophaga smithi Scapanes australis Schizonycha sp. Family Curculionidae Otiorhyncus arcticus Otiorhyncus nodosus Sitona discoideus Order Diptera Family Culicidae Aedes spp. Aedes aegypti Aedes polynesiensis Aedes vexans Anopheles crucians Anopheles culicifacies Anopheles dthali Anopheles freeborni Anopheles nyssorhynchus albimanus Anopheles punctipennis Anopheles sergentii Anopheles superpictus Anopheles turkhudi Culex pipiens Culex pipiens quinquefasciatus Culex restuans Culex tarsalis Ochlerotatus spp. Order Lepidoptera Family Zygaenidae Harrisina brillians Family Pyralidae Ostrinia nubilalis Family Lymantriidae Lymantria dispar Lymantria monacha

Family Noctuidae Agrotis segetum Anticarsia gemmatalis Pseudoplusia includens Trichoplusia ni Order Hymenoptera Family Siricidae Sirex noctilio Family Diprionidae Gilpinia hercyniae Neodiprion sertifer Family Formicidae Solenopsis invicta

Class Arachnida Subclass Acari Order Prostigmata Family Eriophyidae *Eriophyes sheldoni Phyllocoptruta oleivora* Family Tetranychidae *Mononychellus tanajoa*

References

- 1. Anonymous. 1919. Notes on insects in Seychelles. Ms. received fr. Colonial Off. Sept. 6, 1919. In: Rev. Appl. Entomol., Ser. A 7: 483-484.
- 2. Bahvalov, S. 2004. Institute of Systematics and Ecology of Animals, Siberian branch of the Russian Academy of Sciences, Novosibirsk, Russia. Personal communication to A. Sharov.
- 3. Bailey, P. & R. Milner. 1985. *Sitona discoideus*: A suitable case for control with pathogens? Proc. 4th Austr. Appl. Ent. Res. Conf., p. 210-217.
- 4. Balch, R. E. 1946. The disease of the European spruce sawfly. Can. Dept. Agric. Forest Insect Invest. Bimon. Progr. Rpt. 2(5): 1.
- 5. Balch, R. E. & F. T. Bird. 1944. A disease of the European spruce sawfly, *Gilpinia hercyniae* (Htg.) and its place in natural control. Sci. Agr. 25: 65-80.
- Bauer, L. S., F. J. Sapio, M. L. McManus, J. V. Maddox, M. R. Jeffords & D. W. Onstad. 1993. Interactions of *Microsporidium* and gypsy moth in Michigan field plots. USDA, Forest Service, Gen. Tech. Rpt. NE-179: 22.
- Bauer, L. S., F. J. Sapio, M. L. McManus, J. V. Maddox, M. R. Jeffords & D. W. Onstad. 1994. Interactions of *Microsporidium* and gypsy moth in Michigan field plots. USDA, Forest Service, Gen. Tech. Rpt. NE-188: 5.
- Beach, R. M., G. R. Carner & S. G. Turnipseed. 1984. Field efficacy and persistence of a nuclear polyhedrosis virus of the velvetbean caterpillar in soybeans. J. Agric. Entomol. 1: 296-304.

- 9. Bedding, R. A. 1979. Manipulating the entomophagousmycetophagous nematode, *Deladenus siricidicola*, for biological control of the woodwasp *Sirex noctilio* in Australia. In (W. E. Waters, Ed.) *Current Topics in Forest Entomology*, USDA, Forest Service, Gen. Tech. Pap. WO-8: 144-147.
- Bedding, R. A. 1984. Nematode parasites of Hymenoptera, pp. 755-795. In (W. R. Nickle, Ed.) *Plant and Insect Parasitic Nematodes*, Marcel Dekker, NY.
- Bedding, R. A. 1993. Biological control of Sirex noctilio using the nematode Deladenus siricidicola, pp. 11-20. In (R. Bedding, R. Akhurst, H. Kaya, Eds.). Nematodes and the Biological Control of Insect Pests. CSIRO, East Melbourne, Victoria, Australia.
- 12. Bedford, G. O. 1976. Use of a virus against the coconut palm rhinoceros beetle in Fiji. PANS 22: 11-25.
- 13. Bedford, G. O. 1977. Virus against coconut rhinoceros beetle in Fiji. South Pac. Bull. 27: 27-34.
- 14. Bedford, G. O. 1980. Biology, ecology, and control of palm rhinoceros beetles. Annu. Rev. Entomol. 25: 309-339.
- 15. Bellotti, A.C. & J.A. Reyes. 1980. South and Central America. Proc. Workshop on Insect Pest Management with Microbial Agents: Recent Achievements, Deficiencies, and Innovations, pp. 20-21. Boyce Thompson Institute, Ithaca, NY.
- Benjamin, D. M., J. D. Larson & A. T. Drooz. 1955. The European pine sawfly on the Henderson State Forest, Illinois, with notes on its biology and control. J. For. 53: 359-362.

- 17. Bianchi, M. 2004. Personal communication to P. Klasmer.
- Bidochka, M. J., S. R. A. Walsh, M. E. Ramos, R. J. St. Leger, J. C. Silver & D. W. Roberts. 1996. Fate of biological control introductions: Monitoring an Australian fungal pathogen of grasshoppers in North America. Proc. Natl. Acad. Sci. USA 93: 918-921.
- 19. Bird, F. T. 1955. Virus diseases of sawflies. Can. Entomol. 87: 124-127.
- 20. Bird, F. T. & J. M. Burk. 1961. Artificially disseminated virus as a factor controlling the European spruce sawfly, *Diprion hercyniae* (Htg.), in the absence of introduced parasites. Can. Entomol. 93: 228-238.
- 21. Bird, F. T. & D. E. Elgee. 1957. A virus disease and introduced parasites as factors controlling the European spruce sawfly, *Diprion hercyniae* (Htg.), in central New Brunswick. Can. Entomol. 89: 371-378.
- 22. Brooks, W. M. 1988. Entomogenous protozoa, pp. 1-150. In (C. M. Ignoffo, Ed.) Handbook of Natural Pesticides. Vol. V. Microbial Pesticides. Part A. Entomogenous Protozoa and Fungi. CRC Press, Boca Raton, FL.
- 23. Carner, G. R. 2004. Personal communication.
- 24. Carruthers, R. I. & J. A. Onsager. 1993. Perspective on the use of exotic natural enemies for biological control of pest grasshoppers (Orthoptera: Acrididae). Environ. Entomol. 22: 885-903.
- 25. Carruthers, R. I., M. E. Ramos, T. S. Larkin, D. L. Hostetter & R. S. Soper. 1996. The *Entomophaga grylli* (Fresenius) Batko species complex: Its biology, ecology, and use for biological control of pest grasshoppers. Mem. Entomol. Soc. Can. 171: 329-353.
- 26. Chapman, H. C., C. P. Pant, H. L. Mathis, M. J. Nelsen & B. Phantomachinda. 1972. Field release of the nematode *Reesimermis nielseni* for the control of *Culex p. fatigans* in Bangkok, Thailand. WHO/BC 72: 412.
- 27. Chen, P.-S. 1976. A study on *Reesimermis nielseni* for control of *Culex pipiens fatigens* in Taiwan. Bull. Inst. Zool., Acad. Sinica 15: 21-28.
- Clausen, C. P. (Ed.). 1978. Introduced Parasites and Predators of Arthropod Pests and Weeds: A World Review. USDA, Agric. Handbook No. 480, 545 pp.
- 29. Cock, M. J. W. (Ed.). 1985. A Review of Biological Control of Pests in the Commonwealth Caribbean and Bermuda up to 1982. Commonw. Inst. Biol. Contr. Tech. Comm. 9.
- Coulson, J. R. 1981. Nematoda: Mermithidae, pp. 370-372. In (C. C. Doane & M. L. McManus, Eds.) The Gypsy Moth: Research Toward Integrated Pest Management. USDA Tech. Bull. 1584.
- Crawford, A. M., B. Zelazny & A. R. Alfiler. 1986. Genotypic variation in geographical isolates of *Oryc*tes baculovirus. J. Gen. Virol. 67: 949-952.

- 32. de Crouzel, I. S. 1983. El control biológico en la Argentina. Symp. Contr. Biol. Contr. Integr. Plagas Latinoamerica. IX Claz Peru, Oct., pp. 169-174.
- 33. Cújar, A. & H. Alcaraz. 1973. La poliedrosis nuclear una enfermedad virosa del *Trichoplusia ni* (Hüber) como medida de control biologico en el algodonero. Fitotech. Latinoamer. 9: 28-35.
- 34. Cunningham, J. C. 1998. North America, pp. 313-331. In (Hunter-Fujita, P. F. Entwistle, H. F. Evans & N E. Crook, Eds.), *Insect Viruses and Pest Management*, Wiley, Chichester.
- 35. Cunningham, J. C. & P. F. Entwistle. 1981. Control of sawflies by baculovirus, pp. 379-407. In (H. D. Burges, Ed.) *Microbial Control of Pests and Plant Diseases 1970-1980*. Academic Press, London.
- 36. Delalibera Júnior, I., unpublished data.
- 37. Decker, G. 1960. Microbial insecticides---and their future. Agric. Chemicals 15(1): 30-33, 93.
- 38. De Ley, P. & M. Blaxter. 2002. A new system for Nematoda: Combining morphological characters with molecular trees, and translating clades into ranks and taxa. Nematology 4: 141-142.
- 39. Dowden, P. B. & H. B. Girth. 1953. Use of a virus disease to control European pine sawfly. J. Econ. Entomol. 46: 525-526.
- 40. Dupont, P. R. 1931. Entomological and mycological notes. Ann. Rep. Dept. Agric. Seychelles 1930, pp. 11-13. In: Rev. Appl. Entomol., Ser. A 19: 685.
- 41. Eilenberg, J., A. Hajek & C. Lomer. 2001. Suggestions for unifying the terminology in biological control. BioControl 46: 387-400.
- Elkinton, J. S., A. E. Hajek, G. H. Boettner & E. E. Simons. 1991. Distribution and apparent spread of *Entomophaga maimaiga* (Zygomycetes: Entomophthorales) in gypsy moth (Lepidoptera: Lymantriidae) populations in North America. Environ. Entomol. 20: 1601-1605.
- 43. Evans, H. F. & P. F. Entwistle. 1982. Epizootiology of the nuclear polyhedrosis virus of European spruce sawfly with emphasis on persistence of virus outside the host, pp. 449-461. In (E. Kurstak, Ed.) *Microbial and Viral Pesticides*, Dekker, New York.
- 44. Fetter-Lasko, J. L. & R. K. Washino. 1977. A three year study of the ecology of *Lagenidium giganteum*, infections of *Culex tarsalis* in California. Proc. Pap. Calif. Mosq. Control Assoc. 45: 106.
- 45. Forbes, S. A. 1899. Recent work on the San Jose scale in Illinois. Univ. Ill. Agric. Exp. Stn. Bull. 56: 270-280.
- Fuxa, J. R. 1987. Ecological considerations for the use of entomopathogens in IPM. Annu. Rev. Entomol. 32: 225-251.

- Fuxa, J. R. & A. R. Richter. 1999. Classical biological control in an ephemeral crop habitat with *Anticarsia gemmatalis* nucleopolyhedrovirus. BioControl 44: 403-419.
- 48. Fuxa, J. R., A. R. Richter & P. J. McLeod. 1992. Virus kills soybean looper years after its introduction into Louisiana. La. Agric. 35: 20-24.
- 49. Galloway, T. D. 1975. Application of a mermithid nematode (*Reesimermis nielseni* Tsai and Grundmann) from Louisiana for mosquito control in Manitoba. Proc. Alberta Mosquito Abatement Symp., Univ. Alberta, pp. 191-205.
- 50. Galloway, T. D. & R. A. Brust. 1976. Field application of the mermithid nematode, *Romanomermis culicivorax* Ross and Smith, for the control of mosquitoes, *Aedes* spp., in spring in Manitoba. Manitoba Entomol. 10: 18-25.
- 51. Glaser, R. W. 1915. Wilt of gipsy-moth caterpillar. J. Agric. Res. 4:101-128.
- 52. Godfrey, K., D. Steinkraus & M. McGuire. 2001. Fungal pathogens of the cotton and green peach aphids in the San Joaquin Valley. Southwest. Entomol. 26: 297-302.
- 53. Gorick, B. D. 1980. Release and establishment of the baculovirus disease of *Oryctes rhinoceros* (L.) (Coleoptera: Scarabaeidae) in Papua New Guinea. Bull. Entomol. Res. 70: 445-453.
- Greathead, D. J. 1971. A Review of Biological Control in the Ethiopian Region. Commonw. Inst. Biol. Contr. Tech. Commun. No. 5.
- 55. Greathead, D. J. & A. H. Greathead. 1992. Biological control of insect pests by insect parasitoids and predators: The BIOCAT database. Biocontrol News Info. 13: 61N-68N.
- 56. Gutierrez, J. 1981. Updating of data on economic entomology on Wallis and Futuna. ORSTOM (Noumea, New Caledonia): 24 pp.
- 57. Hajek, A. E., Unpublished data.
- 58. Hajek, A. E. 2004. Natural Enemies: An Introduction to Biological Control. Cambridge Univ. Press, Cambridge, UK.
- Hajek, A. E., I. Delalibera Junior & M. L. McManus. 2000. Introduction of exotic pathogens and documentation of their establishment and impact, pp. 339-369. In (L. A. Lacey & H. K. Kaya, Eds.) *Field Manual of Techniques in Invertebrate Pathology*, Kluwer Acad. Publ., Dordrecht, Netherlands.
- 60. Hajek, A. E., R. A. Humber & J. S. Elkinton. 1995. Mysterious origin of *Entomophaga maimaiga* in North America. Amer. Entomol. 41: 31-42.
- 61. Hall, I. M. 1952. Observations on *Perezia pyraustae* Paillot, a microsporidian parasite of the European corn borer. J. Parasitol. 38: 48-52.

- 62. Hammes, C. & P. Monsarratt. 1974. Recherches sur Oryctes rhinoceros L. Cah. ORSTOM Biol. 22: 44-111.
- 63. Harper, J. 1978. Introduction and colonization of entomopathogens, pp.3-13. In (G. E. Allen, C. M. Ignoffo & R. P. Jaques, Eds.), *Microbial Control of Insect Pests: Future Strategies in Pest Management Systems*, NSF-USDA-Univ. Florida Workshop.
- 64. Haugen, D. A. & M. G. Underwood. 1990. *Sirex noctilio* control program in response to the 1987 Green Triangle outbreak. Austral. For. 53: 33-40.
- 65. Hodge, K. T., A. J. Sawyer & R. A. Humber. 1995. RAPD-PCR for identification of *Zoophthora radicans* isolates in biological control of potato leafhopper. J. Invertebr. Pathol. 65: 1-9.
- 66. Hountondji, F. C. C., C. J. Lomer, R. Hanna, A. J. Cherry & S. K. Dara. 2002. Field evaluation of Brazilian isolates of *Neozygites floridana* (Entomophthorales: Neozygitaceae) for the microbial control of cassava green mite in Benin, West Africa. Biocontr. Sci. Tech. 12: 361-370.
- 67. Iede, E. T., S. R. Chiarello & M. S. Pereira. 1998. Utilização do nematóde *Deladenus siricidicola* (Nematodea: Neotylenchidae) no controle biológico de *Sirex noctilio* (Hymenoptera: Siricidae), praga de *Pinus* spp. [Abstract] Primer Congres. Latinoamer. IUFRO, Valdívia, Chile.
- 68. Iede, E. T., S. Penteado & E. Schaitza. 1998. Sirex management in Brazil. Biocontrol News & Information 19(1): 4N. http://pest.cabweb.org/Journals/BNI/ BNI19-1/genews.htm.
- 69. Illingworth, J. F. 1929. Preliminary notes on pests of agricultural crops of Kona, March 15, 1928. Proc. Haw. Ent. Soc. 7: 248-254.
- Izhevskii, S. S. & A. D. Orlinskii. 1985. Biological suppression of citrus whitefly. Zash. Rast. 1985(4): 30-31. [English abstract in CAB Abstracts]
- 71. Jacob, T. K. 1996. Introduction and establishment of baculovirus for the control of rhinoceros beetle *Oryctes rhinoceros* (Coleoptera: Scarabaeidae) in the Andaman Islands. Bull. Entomol. Res. 86: 257-262.
- 72. Jeffords, M. R., J. V. Maddox, M. L. McManus, R. E. Webb & A. Wieber. 1988. Egg contamination as a method for the inoculative release of exotic Microsporidia of the gypsy moth. J. Invertebr. Pathol. 51: 190-196.
- 73. Jeffords, M. R., J. V. Maddox, M. L. McManus, R. E. Webb & A. Wieber. 1989. Evaluation of the overwintering success of two European microsporidia inoculatively released into gypsy moth populations in Maryland. J. Invertebr. Pathol. 53: 235-240.

- 74. Julien, M.H. & M.W. Griffiths (Eds.) 1998. *Biological Control of Weeds: A World Catalogue of Agents and their Target Weeds, 4th ed.* CABI Publ., Wallingford, Oxon, UK.
- 75. Keeling, P. J. 2003. Congruent evidence from α-tubulin and β-tubulin gene phylogenies for a zygomycete origin of microsporidia. Fungal Gen. Biol. 38: 298-309.
- 76. Klasmer, P., E. N. Botto, J. C. Corley, J. M. Villacide & V. Fernandez Arhex. 2000. Advances in *Sirex noctilio* biological control in Patagonian region of Argentina. Ser.Tecnica IPEF 13:21-30.
- 77. Klasmer, P., E. N. Botto, J. C. Corley & J. M. Villacide. 2004. Evaluación del nematodo *Deladenus siricidicola* Bedding (Nematoda: Neotylenchidae) como potencial agente para el control de *Sirex noctilio* F. (Hymenoptera: Siricidae) en la Patagonia Argentina. Inv. For. Serv. Prod. II. Secr. Agric. Ganad. Pesc. Aliment., Proy. For. Desarrollo, Buenos Aires. pp. 70-73.
- Knowles, C. H. 1919. Division of Entomology, Fiji Dept. Agric. Ann. Rpt. 1918, pp. 12-15. In: Rev. Appl. Entomol., Ser. A 8: 297-298.
- 79. Koebele, A. 1897. Report of the entomologist of the Hawaiian government. Planters' Monthly 16: 67-85.
- Kotinsky, J. 1906. Report of the assistant entomologist. Haw. Board Comm. Agr. & For. Ann. Rpt. (1905), pp. 113-144.
- Kotinsky, J. 1909. Report of superintendent of entomology for June, 1909. Haw. Forester & Agr. 6: 337-339.
- Laird, M. 1971. Microbial control of arthropods of medical importance, pp. 387-406. In (H. D. Burges & N. W. Hussey, Eds.), *Microbial Control of Insects and Mites*. Academic Press, London.
- 83. Laird, M., J. Urdang & I. Tinielu. 1982. Establishment and long-term survival of *Romanomermis culicivorax* in mosquito habitats, Tokelau Islands. Mosq. News 42: 86-92.
- 84. Lange, C. E. & M. L. de Wysiecki. 1996. The fate of Nosema locustae (Microsporidia: Nosematidae) in Argentine grasshoppers (Orthoptera: Acrididae). Biol. Contr. 7: 24-29.
- 85. Latteur, G. & J. Godefroid. 1982. Trial of field treatments against cereal aphids with mycelium of *Erynia neoaphidis* (Entomophthorales) produced *in vitro*, pp. 2-10. In (R. Cavalloro & A. A. Balkema, Eds.), *Aphid Antagonists*. Rotterdam, NL.
- 86. Lomer, C. J. 1985. Ecology of *Oryctes monoceros* in the Seychelles. Antenna 9: 28-29.
- Lomer, C. J. 1986. Release of *Baculovirus oryctes* into Oryctes monoceros populations in the Seychelles. J. Invertebr. Pathol. 47: 237-246.

- 88. Magnoler, A. 1974. Field dissemination of a nucleopolyhedrosis virus against the gypsy moth, *Lymantria dispar* L. Z. Pflkrankh. 9: 497-511.
- 89. Marschall, K. J. 1970. Introduction of a new virus disease of the coconut rhinoceros beetle in Western Samoa. Nature 225: 288-289.
- Marschall, K.J. & I. Ioane. 1981. The effect of re-release of *Oryctes rhinoceros* Baculovirus in the biological control of rhinoceros beetles in Western Samoa. J. Invertebr. Pathol. 39: 267-276.
- 91. McCray Jr., E. M., D. J. Womeldorf, R. C. Husbands & D. A. Eliason. 1973. Laboratory observations and field tests with *Lagenidium* against California mosquitoes. Proc. Pap. Calif. Mosq. Control Assoc. 41: 123-128.
- 92. McGugan, B. M. & H. C. Coppel. 1962. A review of the biological control attempts against insects and weeds in Canada. II. Biological control of forest insects, 1910-1958. Commonw. Inst. Biol. Contr. Tech. Comm. 2: 35-216.
- 93. McGuire, M. R., J. V. Maddox & E. J. Armbrust. 1987. An epizootic caused by *Erynia radicans* (Zygomycetes: Entomophthoraceae) isolated from *Empoasca fabae* (Homoptera: Cicadellidae). J. Invertebr. Pathol. 50: 78-80.
- 94. McManus, M. L. 2004. Personal communication.
- 95. Milner, R. J. 2003. Personal communication.
- 96. Milner. R. J. 1985. Field tests of a strain of *Entomophaga grylli* from the USA for biocontrol of the Australian wingless grasshopper, *Phaulacridium vittatum*, pp. 255-261. Proc. 4th Australasian Conf. Grassl. Invert. Ecol., (R. B. Chapman, Ed.), Lincoln Coll., Canterbury, Caxton Press.
- 97. Milner, R. J. 1986. Pathogen importation for biological control: Risks and benefits, pp. 115-121. In (A. J. Gibbs & H. R. C. Meischke, Eds.), *Pests* and Parasites as Migrants, Cambridge Univ. Press, Cambridge, UK.
- Milner, R. J. & R. S. Soper. 1981. Bioassay of Entomophthora against the spotted alfalfa aphid Therioaphis trifolii f. maculata. J. Invertebr. Pathol. 37: 168-173.
- 99. Milner, R. J., R. S. Soper & G. G. Lutton. 1982. Field release of an Israeli strain of the fungus Zoophthora radicans (Brefeld) Batko for biological control of Therioaphis trifolii (Monell) f. maculata. J. Aust. Ent. Soc. 21: 113-118.
- 100. Mitchell, C. J., P. S. Chen & H. C. Chapman. 1974. Exploratory trials utilizing a mermithid nematode as a control agent for *Culex* mosquito in Taiwan. J. Formosan Med. Assoc. 73: 241-254.

- 101. Mohan, K. S. & G. B. Pillai. 1993. Biological control of Oryctes rhinoceros (L.) using an Indian isolate of Oryctes baculovirus. Insect Sci. Applic. 14: 551-558.
- 102. Monty, J. 1978. The coconut palm rhinoceros beetle, *Oryctes rhinoceros* (L.) (Col., Dynastidae), in Mauritius and its control. Rev. Agric. Sucr. Ile Maurice 57: 60-76.
- 103. Moutia, L. A. 1933. Campaign against *Phytalis* smithi Arrow. Rep. Dep. Agric. Mauritius, 1932, pp. 43-51.
- 104. Moutia, L. A. 1934. Campaign against *Phytalis smithi* Arrow. Rep. Dep. Agric. Mauritius, 1933, pp. 25-29.
- 105. Moutia, L. A. & R. Mamet. 1946. A review of twentyfive years of economic entomology in the island of Mauritius. Bull. Entomol. Res. 36: 439-472.
- 106. Nickle, W. R. 1979. Probable establishment and overwintering of a mermithid nematode parasite of mosquitoes in Maryland. Proc. Helminth. Soc. Wash. 46: 21-27.
- 107. Nielsen, C., M. G. Milgroom & A. E. Hajek. 2005. Genetic diversity in the gypsy moth fungal pathogen *Entomophaga maimaiga* from founder populations in North America and source populations in Asia. Mycol. Res. (in press).
- 108. Oddsdottir, E., C. Nielsen, T. Levisson, S. Harding, G. Haldorddon & J. Eilenberg. 2005. Personal communication.
- 109. Ogilvie, L. 1926. Report of the plant pathologist for the year 1925. Rept. Dept. Agric. 1925, pp. 36-63. In: Rev. Appl. Entomol., Ser. A 14: 624-626.
- 110. Ogilvie, L. 1928. Notes on the growing of citrus in Bermuda. Agric. Bull. Bermuda Dept. Agric. VI (11), pp. 3-5, (12) 4-5; VII (2), pp. 3-6, (3), pp. 4-6. In: Rev. Appl. Entomol., Ser. A 16: 445-446.
- 111. Ogilvie, L. 1928. Report of the plant pathologist for the year 1926. Rept. Dept. Agric. Bermuda 1926, pp. 35-41. In: Rev. Appl. Entomol., Ser. A 16: 17-18.
- 112. Oi, D. H. & D.F. Williams. 2002. Impact of *The-lohania solenopsae* (Microsporidia: Thelohaniidae) on polygyne colonies of red imported fire ants (Hymenoptera: Formicidae). J. Econ. Entomol. 95: 558-562.
- 113. Parkman, J. P. & Smart, Jr., G. C. 1996. Entomopathogenic nematodes, a case study: Introduction of *Steinernema scapterisci* in Florida. Biocontr. Sci. Technol. 6: 413-419.
- 114. Perkins, R. C. L. 1906. Leaf hoppers and their natural enemies (introduction). Haw. Sugar Planters' Assoc. Ent. Series Bul. No. 1, 32 pp.

- Petersen, J. J. 1985. Nematodes as biological control agents: Part I. Mermithidae. Adv. Parasitol. 24: 307-346.
- 116. Petersen, J. J., H. C. Chapman, O. R. Willis & T. Fukuda. 1978. Release of *Romanomermis culicivorax* for the control of *Anopheles albimanus* in El Salvador. II. Application of the nematode. Amer. J. Trop. Med. Hyg. 27: 1268-1273.
- 117. Pilarska, D. 2004. Personal communication.
- 118. Pilarska, D., M. McManus, A. E. Hajek, F. Hérard, F. E. Vega, P. Pilarska & G. Markova. 2000. Introduction of the entomopathogenic fungus *Entomophaga maimaiga* Hum., Shim. & Sop. (Zygomycetes: Entomophthorales) to a *Lymantria dispar* (L.) (Lepidoptera: Lymantriidae) population in Bulgaria. Anz. Schadlingsk. 73: 125-126.
- 119. Poinar, Jr., G. O. 1979. Nematodes for Biological Control of Insects. CRC Press, Boca Raton, FL.
- 120. Ponomarenko, N. G., H. A. Prilepskaya, M. Ya. Murvanidze & L. A. Stolyarova. 1975. *Aschersonia* against whiteflies. Zashchita Rastenii 1975(5): 44-45. [English abstract in CAB Abstracts]
- 121. Porcile Maderni, J. F. 1998. *Sirex noctilio* F.: Present status in Uruguay, pp. 81-82. In (E. T. Iede, E. Schaitza, S. Penteado, R. C. Reardon & S. T. Murphy, Eds.), Proceedings of a Conference: Training in the Control of *Sirex noctilio* by the Use of Natural Enemies. USDA, Forest Service FHTET-98-13.
- 122. Protsenko, E. P. 1967. The importance of the fungus Aschersonia in nature and its practical use by main in the biological control of insects. Sb. Karantinu Rast. 19: 147-215. In: McCoy, C.W., R. A. Samson & D. G. Boucias. 1988. Entomogenous fungi, pp. 151-236. In (C. M. Ignoffo, Ed.) Handbook of Natural Pesticides. Vol. V. Microbial Pesticides. Part A. Entomogenous Protozoa and Fungi. CRC Press, Boca Raton, FL.
- 123. Purrini, K. 1989. *Baculovirus oryctes* release into *Oryctes monoceros* population in Tanzania, with special reference to the interaction of virus isolates used in our laboratory infection experiments. J. Invertebr. Pathol. 53: 285-300.
- 124. Ramos, M. E. 2004. Personal communication.
- 125. Rao, V. P., M. A. Ghani, T. Sankaran & K. C. Mathur. 1971. A Review of the Biological Control of Insects and Other Pests in South-East Asia and the Pacific Region. Commonw. Inst. Biol. Contr. Tech. Comm. No. 6.
- 126. Rivers, C. F. 1962. The use of a polyhedral virus disease in the control of the pine sawfly *Neodiprion sertifer* Geoffr. in north-west Scotland. Entomophaga Mém. hor. Sér. 2: 477-480.

- 127. Rojas, W., J. Northrup, O. Gallo, A. E. Montoya, F. Montoya, M. Restrepo, G. Nimnich, M. Arango & M. Echavarria. 1987. Reduction of malaria prevalence after introduction of *Romanomermis culicivorax* (Mermithidae: Nematoda) in larval *Anopheles* habitats in Colombia. Bull. WHO 65: 331-337.
- 128. Schaitza, E. 2004. Personal communication.
- 129. Schreiner, I. 1989. Biological control introductions in the Caroline and Marshall Islands. Proc. Haw. Ent. Soc. 29: 57-69.
- Schuder, D. L. 1956. A specific virus disease for control of the European pine sawfly, *Neodiprion sertifer* (Geoff.). Proc. Ind. Acad. Sci. 66: 101-102.
- 131. Seguni, Z., B. Lohr & W. Mwaiko. 1999. Introducing Baculovirus oryctes Huger into Oryctes monoceros Oliv. populations in Tanzania. J. Appl. Ent. 123-427-431.
- 132. Shands, W. A., C. G. Thompson, G. W. Simpson & H. E. Wave. 1958. Preliminary studies of entomopathogenous fungi for the control of potato-infesting aphids in Maine. J. Econ. Entomol. 51: 184-186.
- 133. Shands, W. A., G. W. Simpson & I. M. Hall. 1963. Importance of entomogenous fungi in controlling aphids on potatoes in northeastern Maine. Maine Agric. Exp. Stn. Tech. Bull. 6: 1-42.
- 134. Smith, J. B. 1898. The San Jose or pernicious scale. New Jersey Exp. Sta. Rpt. 19: 443-446.
- 135. Smith, J. B. 1903. Report of the Entomologist. New Jersey Exp. Sta. Rpt. 24: 555-569.
- 136. Sosa Gomez, D. R. 1987. Control microbiano de *Phyllocoptruta oleivora* (Ashm). y *Eriophyes sheldoni* Ewing mediante pulverizaciones de conídios de tres variedades de *Hirsutella thompsonii* Fisher. XI Congr. Bras. Entomol. Res. 1: 167.
- 137. Sosa Gomez, D. R. & F. Moscardi. 1991. Microbial control and insect pathology in Argentina. Ciencia e Cultura 43(5): 375-379.
- 138. Speare, A. T. & R. H. Colley. 1912. The Artificial Use of the Brown-tail Fungus in Massachusetts, with Practical Suggestions for Private Experiment, and a Brief Note on a Fungous Disease of the Gypsy Caterpillar. Wright & Potter, Boston.
- 139. Squibbs, F. L. 1935. Work connected with insect pests and fungus diseases. Rep. Dep. Agric. Seychelles 1933, pg. 5, Victoria, Seychelles, 1934. In: Rev. Appl. Entomol., Ser A 23: 241.
- 140. Stapley, J.H. 1980. Annual report of the entomologist for 1979. Rept., Ministry Agric. & Lands, Solomon Islands. 30 pp.
- 141. Steinhaus, E. A. 1951. Report on diagnoses of diseased insects 1944-50. Hilgardia 20: 629-678.

- 142. Steinkraus, D. C., G. O. Boys & J. A. Rosenheim. 2002. Classical biological control of *Aphis gossypii* (Homoptera: Aphididae) with *Neozygites fresenii* (Entomophthorales: Neozygitaceae) in California cotton. Biol. Contr. 25: 297-304.
- 143. Stern, V. & B. Federici. 1990. Granulosis virus: Biological control of western grapeleaf skeletonizer. Calif. Agric. 44: 21-22.
- 144. Stern, V. & B. Federici 1990. Biological control of western grapeleaf skeletonizer, *Harrisina brillians* Barnes and McDunnough (Lepidoptera: Zygaenidae), with a granulosis virus in California, pp. 167-176. In (N. J. Bostanian, L. T. Wilson & T. J. Dennehy, Eds.) *Monitoring and Integrated Management of Arthropod Pests of Small Fruit Crops*. Intercept, Andover, Hampshire, UK.
- 145. Swan, D. I. 1974. A review of the work on predators, parasites and pathogens for the control of *Oryctes rhinoceros* (L.) (Coleoptera: Scarabaeidae) in the Pacific area. Commonw. Inst. Biol. Contrl. Misc. Publ. 7. Commonw. Agric. Bur., Farnham, Slough, UK. 64 pp.
- 146. Tanada, Y. 1957. An annotated list of infectious diseases of insects in Hawaii. Proc. 8th Pac. Sci. Congr., Vol. 3A. Oceanogr. Zool. National Res. Council, Manila, Phil.
- 147. Tanada, Y. & H. Kaya. 1993. *Insect Pathology*. Academic Press, San Diego.
- 148. Teakle, R. E. 1998. Australasia, pp. 303-312. In (F. R. Hunter-Fujita, P. F. Entwistle, H. F. Evans & N E. Crook, Eds.), *Insect Viruses and Pest Management*, Wiley, Chichester.
- 149. Theunis, W. & N. Teuriara. 1998. Biological control of *Papuana huebneri* (Coleoptera, Scarabaeidae) in Kiribati: field trials with *Metarhizium anisopliae* and *Bacillus popilliae*. J. So. Pac. Agric. 15: 46-51.
- 150. Tribe, G. 1998. Biological control of *Sirex noctilio* in South Africa, p. 91. In (E. T. Iede, E. Schaitza, S. Penteado, R. C. Reardon & S. T. Murphy, Eds.), Proceedings of a Conference: Training in the Control of *Sirex noctilio* by the Use of Natural Enemies. USDA, Forest Service FHTET-98-13.
- 151. Tribe, B. 1998. *Sirex* spreads in South Africa. Biocontrol News & Information 19(1). http://pest.cabweb. org/Journals/BNI/BNI19-1/genews.htm.
- 152. Washino, R. K., J. L. Fetter, C. K. Fukushima & K. Gonot. 1976. The establishment of *Lagenidium giganteum*, an aquatic fungal parasite of mosquitoes, three years after field introduction. Proc. Pap. Calif. Mosq. Control Assoc. 44: 52.

- 153. Williams, D. F., G. J. Knue & J. J. Becnel. 1998. Discovery of *Thelohania solenopsae* from the red imported fire ant, *Solenopsis invicta*, in the United States. J. Invertebr. Pathol. 71: 175-176.
- 154. Wilson, C. E. 1921. Report of the entomologist. Rept., Virgin Isl. Agric. Expt. Stn. 1920, pp. 20-35. In: Rev. Appl. Entomol., Ser. A 9: 429-431.
- 155. Wilson, F. 1960. A Review of the Biological Control of Insects and Weeds in Australia and Australian New Guinea. Commonw. Inst. Biol. Contr. Tech. Comm. No. 1.
- 156. Woese, C. R., O. Kandler & M. L. Wheelis. 1990. Towards a natural system of organisms: Proposal for the domains Archaea, Bacteria, and Eucarya. Proc. Natl. Acad. Sci. USA 87: 4576-4579.
- 157. Woodbridge, S. M. 1906. Diseases of scale insects. Bull. So. Calif. Acad. Sci. 5: 29-31.
- 158. Yasukawa, S. 1925. Experiments with the fungus *Metarrhizium anisopliae* Motsch., parasitic upon insects injurious to the sugar-cane. Dept. Agric., Formosa Govt. Res. Inst. 11, 81 pp. In: Rev. Appl. Entomol., Ser. A 13: 361.
- 159. Yen, D. F. 1977. Microbial control of insect pests in Taiwan. N.T.U. Phytopath. Entomol. 5: 1-14.
- 160. Young, E. C. 1974. The epizootiology of two pathogens of the coconut palm rhinoceros beetle. J. Invertebr. Pathol. 24: 82-92.
- 161. Young, E. C. & J. F. Longworth. 1981. The epizootiology of the baculovirus of the coconut palm rhinoceros beetle (*Oryctes rhinoceros*) in Tonga. J. Invertebr. Pathol. 38: 362-369.
- 162. Zaim, M., H. Ladonni, M. R. Y. Ershadi, A. V. Manouchehri, Z. Sahabi, M. Nazari & H. Shahmohammadi. 1988. Field application of *Romanomermis culicivorax* (Mermithidae: Nematoda) to control anopheline larvae in southern Iran. J. Am. Mosq. Contr. Assn. 4: 351-355.
- 163. Zelazny, B. 1973. Studies on *Rhabdionvirus oryctes*. III. Incidence in the *Oryctes rhinoceeros* population in Western Samoa. J. Invertebr. Pathol. 22: 359-363.
- 164. Zelazny, B. A. 1977. *Oryctes rhinoceros* populations and behavior influenced by a baculovirus. J. Invertebr. Pathol. 29: 210-215.
- 165. Zelazny, B., A. Lolong & A. M. Crawford. 1990. Introduction and field comparison of baculovirus strains against *Oryctes rhinoceros* (Coleoptera: Scarabaeidae) in the Maldives. Environ. Entomol. 19: 1115-1121.
- 166. Zethner, O. 2004. Personal communication.
- 167. Zethner, O. 1976. Control experiments on the nun moth (*Lymantria monacha* L.) by nuclear-polyhedrosis virus in Danish coniferous forests. Z. Angew. Entomol. 81: 192-207.

- 168. Zethner. O. 1980. Control of *Agrotis segetum* (Lep.: Noctuidae) in root crops by granulosis virus. Entomophaga 25: 27-35.
- 169. Zondag, R. 1969. A nematode infection of Sirex noctilio (F.) in New Zealand. New Zealand J. Sci. 12: 732-747.
- 170. Zondag, R. 1979. Control of *Sirex noctilio* F. with *Deladenus siricidicola* Bedding. Part II. Introductions and establishments in the South Island 1968-1975. New Zealand J. For. Sci. 9: 68-76.
- 171. Øgaard, L., C. F. Williams, C. C. Payne & O. Zethner. 1988. Activity persistence of granulosis viruses (Baculoviridae) in soils in United Kingdom and Denmark. Entomophaga 33: 73-80.

Index

Scientific names of insect and mite hosts

A

Adoretus tenuimaculatus 26 Aedes aegypti 34, 36 Aedes polynesiensis 27, 34 Aedes spp. 34 Aedes vexans 35 Aeneolamia flavilatera 14 Agrotis segetum 8 Aleurodicus cocois 19 Aleurothrixus floccosus 19 Alissonotum impressicolle 24 Anopheles crucians 35 Anopheles culicifacies 35 Anopheles dthali 35 Anopheles freeborni 35 Anopheles nyssorhynchus albimanus 34, 36 Anopheles punctipennis 35-36 Anopheles sergentii 35 Anopheles superpictus 35 Anopheles turkhudi 35 Anoplognathus sp. 25 Anticarsia gemmatalis 7 Aonidiella aurantii 21 Aphis gossypii 16 Aspidiotus destructor 22 Aspidiotus perniciosus 22

С

Camnula pellucida 13 Ceroplastes rubens 20 Clemora smithi 24 Coccus viridis 19-20 Cochliotis melolonthoides 12 Cornuaspis beckii 21 Culex fatigans 30 Culex pipiens 35 Culex pipiens fatigans 30 Culex pipiens quinquefasciatus 30, 33, 36 Culex restuans 35 Culex tarsalis 27, 35

D

Dermolepida albohirtum 25 Dialeurodes citri 18 Dialeurodes citrifolii 17 Dialeurodes sp. 18 Dichroplus elongatus 30 Dichroplus maculipennis 30 Dichroplus pratensis 30 Diprion hercyniae 10, 41-42

Е

Empoasca fabae 14 Eriophyes sheldoni 29 Eucalymnatus tessellatus 20

G

Gilpinia hercyniae 10, 41-42

Η

Harrisina brillians 6, 39

L

Lachnosterna smithi 24 Lepidiota pruinosa 25 Lepidiota sp. 25 Lepidosaphes beckii 21 Leucopholis irrorata 25 Lymantria dispar 6, 28, 31, 36, 40 Lymantria monacha 7

M

Macrosiphum solanifolii 17 Melanoplus bivittatus 13 Melanoplus sanguinipes 13-14 Metopolophium dirhodum 16 Mononychellus tanajoa 29

Ν

Neodiprion sertifer 9-10

0

Ochlerotatus spp. 34 Oryctes monoceros 5 Oryctes rhinoceros 3-5, 11, 23, 32-33 Ostrinia nubilalis 31, 39 Otiorhynchus arcticus 26 Otiorhynchus nodosus 26

P

Papuana huebneri 11, 24 Phaulacridium vittatum 13 Phyllocoptruta oleivora 29 Phyllophaga smithi 24 Phytalus smithi 24 Pseudoplusia includens 8

Q

Quadraspidiotus perniciosus 22

S

Scapanes australis 6 Scapteriscus abbreviatus 32 Scapteriscus borelli 32 Scapteriscus vicinus 32 Schizonycha sp. 11 Scotussa lemniscata 30 Singhiella citrifolii 17 Sirex noctilio F. 37-38, 41 Sitona discoideus 26, 33 Solenopsis invicta 42

T

Therioaphis maculata 15 Thrips tabaci 23 Trichoplusia ni 8

Families of insect and mite hosts

Acrididae 13-14, 30 Alevrodidae 17-19 Aphididae 15-17 Cercopidae 14 Cicadellidae 14-15 Coccidae 19-20 Culicidae 27, 30, 33-36 Curculionidae 26, 33 Diaspididae 21-22 Diprionidae 9-10, 41-42 Eriophyidae 29 Formicidae 42 Gryllotalpidae 32 Lymantriidae 6-7, 28, 31, 36, 40 Noctuidae 7-8 Pyralidae 31, 39 Scarabaeidae 3-6, 11-12, 23-26, 32-33 Siricidae 37-38, 41 Tetranychidae 29 Thripidae 23 Zygaenidae 6, 39

Scientific names of pathogens and nematodes

A

Agrotis segetum Granulovirus (AsGV) 8 Anticarsia gemmatalis Multicapsid Nucleopolyhedrovirus (AgMNPV) 7 Antonospora locustae 30 Aschersonia aleyrodis 19 Aschersonia goldiana 17 Aschersonia spp. 18

B

Baculovirus oryctes 3-6 Beauveria bassiana 24, 26 Beauveria brongniartii 25 Beauveria densa 24 Beddingia siricidicola 37-38, 41 Botrytis tenella 25

С

Cephalosporium lecanii 24 Coelomomyces stegomyiae 27

D

Deladenus siricidicola 37-38, 41

E

Endoreticulatus sp. 31 Entomophaga grylli 13-14 Entomophaga maimaiga 28, 40

F

Fusarium coccophilum 21-22 Fusarium episphaerea 21 Fusarium juruanum 22

G

Gilpinia hercyniae Nucleopolyhedrovirus (GhNPV) 10, 41-42 *Glugea pyraustae* 31, 39

Η

Harrisina brillians Granulovirus (HbGV) 6, 39 Heterorhabditis heliothidis 33 Hexamermis sp. 36 Hirsutella thompsonii 29

L

Lagenidium giganteum 27 Lecanicillium sp. 17, 19 Lecanicillium lecanii 20 Lymantria dispar Multicapsid Nucleopolyhedrovirus (LdMNPV) 6, 40 Lymantria monacha Nucleopolyhedrovirus 7

Μ

Metarhizium anisopliae 14, 23-26 Microsporidium sp. 31 Myrangium duriaei 21

N

Nectria flammea 21-22 Neodiprion sertifer Nucleopolyhedrovirus (NeseNPV) 9-10 Neozygites floridana 29 Neozygites fresenii 16 Neozygites parvispora 23 Neozygites tanajoae 29 Nosema locustae 30 Nosema portugal 31 Nosema pyrausta 31, 39

0

Octomyomermis muspratti 36 Oryctes rhinoceros virus (OrV) 3-6

P

Paenibacillus popilliae 11-12 Pandora neoaphidis 16 Paranosema locustae 30 Perezia pyraustae 31, 39 Pleistophora culicis 30 Plistophora culicis 30 Podonectria coccicola 21 Pseudomicrocera henningsii 22 Pseudoplusia includens Singlecapsid Nucleopolyhedrovirus (PiSNPV) 8

R

Reesimermis nielseni 33-36 Rhabdionvirus oryctes 3-6 Rhabditis sp. 32 Rhabditis sp. nr. maupasi 32-33 Romanomermis culicivorax 33-36

S

Sphaerostilbe coccophila 21-22 Steinernema scapterisci 32

T

Thelohania solenopsae 42 Triblidium caespitosum 21 Trichoplusia ni Nucleopolyhedrovirus (TnNPV) 8

V

Vavraia sp. 31 Verticillium sp. 17, 19 Verticillium lecanii 20

Ζ

Zoophthora radicans 14-15

Higher classification of pathogens and nematodes

Bacillaceae 11-12 Baculoviridae 6-10, 39, 40-42 Coelomomycetaceae 27 Entomophthoraceae 13-16, 28, 40 Heterorhabditidae 33 Hypocreales 14, 17-26, 29 Mermithidae 33-36 Neotylenchidae 37-38, 41 Neozygitaceae 16, 23, 29 Nosematidae 30-31, 39 Pleistophoridae 30-31 Pythiaceae 27 Rhabditidae 32-33 Steinernematidae 32 Thelohaniidae 42 Triblidiaceae 21 Tubeufiaceae 21 Unassigned virus family 3-6

Release countries and regions (capitalized as in the text)

AMERICAN SAMOA 4, 11, 32-33 ARGENTINA 21, 29-30, 38 AUSTRALIA 13, 15, 25-26, 33, 37 **AZERBAIJAN 18 BARBADOS 23 BELGIUM 16** BENIN 29 BERMUDA 17-18, 21 BRAZIL 38 **BULGARIA 28** CANADA 9-10, 34, 41 COLOMBIA 8,34 DENMARK 7-8 El SALVADOR 36 FIJI 3, 26, 32 **GEORGIA 18 GUYANA 14 ICELAND 26** INDIA 4-5 IRAN 35 **KENYA 11** KIRIBATI 11,24 MALDIVES 5 MAURITIUS 4,24 NAURU 30, 36 NEW ZEALAND 37, 41 PALAU 3, 11 PAPUA NEW GUINEA 4 PHILIPPINES 25 **RUSSIA 28** SAMOA 3, 23, 32 SARDINIA 6 SEYCHELLES 5, 20, 22 SOLOMON ISLANDS 6 SOUTH AFRICA 38 **TAIWAN 24, 33** TANZANIA 5,11 THAILAND 33 TOKELAU ISLANDS 3, 23, 27, 34 **TONGA 4,23** UNITED KINGDOM (SCOTLAND) 10 UNITED KINGDOM (WALES) 42 UNITED STATES OF AMERICA 6-10, 13-17, 19-22, 27-28, 31-32, 35-36, 39-42 URUGUAY 38 VIRGIN ISLANDS 19 WALLIS ISLAND 4, 23, 33

Source countries and regions Argentina 23 Australia 13-15, 25, 38 Austria 8,36 Brazil 7, 14, 16, 29, 38 Canada 9-10 China 18 Cuba 18 Europe 39-42 Faroe Islands 26 Fiii 6, 15 France 25-26 Germany 7 Guatemala 8 Hungary 37 India 4-5, 18, 20 Israel 15 Japan 18, 28, 36, 40 Java 23 Madagascar 32 Malaysia 3, 5 Mexico 6, 39 New Zealand 33, 37-38 Nigeria 30 Papua New Guinea 11, 24 Philippines 5 Portugal 31 Praslin Island group 5 Samoa 3-5, 23, 25 Serbia 6 Sierra Leone 22 Singapore 27 Solomon Islands 11 South America 42 Sri Lanka 20, 32-33 Sweden 7,9 Switzerland 23 Tanzania 5 Tasmania 37 Trinidad 14, 18 Unknown 19-20, 24, 26 Uruguay 32 USA 6, 8, 10-13, 16-19, 21-22, 24, 27-31, 33-36, 39 Vietnam 18 Zambia 36 Zimbabwe 29