Can we do more substantively? We have identified the issue. I don't think that any of us have got any more of an answer yet, though. I think Dr. Cornetta put it as well as anyone could.

DR. WILSON: I think you are right that this is a difficult issue which we probably won't be able to come to a definitive answer on today, and a number of important points have been raised for our own internal consideration and discussion. Probably, for the sake of time, we should go ahead and move on for now.

DR. SALOMON: Dr. Cornetta, I don't get any physical input because I can't see you. Are you okay with that? Again, I want to respect the fact that you brought it up.

DR. CORNETTA: No; I don't think it is a thing that you are going to be able to answer today and I think the overall point, and I think that was probably Carolyn, was that I think it is something the FDA is going to have to think hard about and, just as they went through with the retroviruses and coming up with guidelines for testing, this is going to be something that they are going to have to struggle with.

I think all the appropriate comments have

been made.

DR. SALOMON: Then we will move on. I think it would be fair to say, since I have been sort of giving you guys a hard time about this particular one, that I do think that you have--as all the other sponsors appropriately acknowledged, that this is an area of difficulty and you have used what are available guidances to try and help you to figure it out.

So I do commend you for what I think was a good-faith effort, very much so, in this and it is not your fault that we haven't solved it yet.

Are there additional in vivo studies that need to be performed is c), regarding now the safety testing of VRX. I think we have gone over that, too.

DR. WILSON: I think we have really covered c) and d) already.

DR. SALOMON: Good. I was going to try and agree with that.

Question 3. "Please discuss whether vector mobilization is considered an advantage or a safety concern for the proposed clinical trial and consider. Please consider the following, specifically." Now we clearly had a discussion of

mobilization yesterday. I know Dr. Mulligan had to leave to the airport, so we will try and remember and be faithful to some of the comments that he made yesterday.

"Are the data available from the assays to assess vector mobilization by wild-type HIV sufficient? Are there additional preclinical studies to assess vector mobilization that should be performed and, if some, discuss the optimal study design."

Dr. Allan, we have discussed the monkey study; right? So that is on the record. I make a joke of it just not to take myself too seriously but not to trivialize it. That was a good discussion.

So, are there additional studies? Do we think that the data here was sufficient? So I will jump in there and say, yes; it was sufficient to demonstrate to me that there was a lot of mobilization. Again, now, we are doing adjectives; low mobilization, high mobilization.

To me, what I calculated out in the in vitro culture system, which is reasonably what has been studied, was about 1000 copies per ml. I wrote that down. Yes; 1000 copies of

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packaged/mobilized, depending on which terminology you want to use, per ml is occurring.

To me, that is a lot when one copy of HIV theoretically can infect a cell and set off an infection of a patient.

The studies in the mouse show that there is mobilization to CD4 cells. The studies of failure to mobilize to the B-cells, I would suggest, are uninterpretable because there were not enough CD19-positive cells, when you are looking at 1 or 2 percent, to find it. So I think those, you would have to go back, if you want to sell those studies, and do much higher injury levels of CD19 cells.

DR. ALLAN: Do other target cells need to be considered there? Is there tropism, necessarily, uniformly to all non-CD4-positive cells so should macrophages or some other cell type be considered?

DR. TORBETT: And that issue is relevant because, as was mentioned before, you are changing or stressing the envelope nature of this. If you would think about it in antibody terms where an antibody has to be qualified against as panel of cell types, it would seem reasonable, since we are

trying to detect the nonprobable but bad thing to happen, that that issue be explored a bit further.

DR. SALOMON: Yes; it suggests
experiments. Again, I don't want to get down into
the detail, but there are a lot of other things
that could be relatively simply done; for example,
putting EGFP into H9 and putting H9 into the--or
Jerkit or Mold4, different cell lines into the SCID
mice to see whether it was mobilization to
noninfected T-cells, for example.

Again, the details of that; all I am saying is that, from my point of view, just to start this, I think that you have demonstrated that there is mobilization. I think the question now is, unless someone says no, I disagree with you and that is what I am waiting for, but if you agree that there is mobilization, whether it is a little mobilization or a lot of mobilization, I don't know what that means in terms of the biological significance of it, and therefore is that an issue now?

Is that a problem? Are we going to accept mobilization in this system.

DR. ALLAN: What struck me as the fact that it wasn't clear to me what was being mobilized

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and whether they are actually recombinants or not, because the data was presented, which was good, which was it directly looked at either the antisense vector or--let me see; what else was looked at. My consideration is whether you have got some sort of recombinant in there, not that it is replication-competent, but which could be, at some point later, which is like a gag/pol intermediate or something else, that has been mobilized.

So not just that the vector has been mobilized but anything else.

DR. TORBETT: I think the question is is the vector getting mobilized and you are asking is the vector now becoming infectious; is that right?

DR. ALLAN: Not necessarily infectious but that, beyond the vector being mobilized, which they have demonstrated, are there other things being mobilized. It is just a question of terms. I think it is terminology.

DR. TORBETT: I guess it is two different questions, really. I mean, the question of vector mobilization was very hard to assess in any of their animal models. All I can say is it occurred. To what degree, how much, is unclear to me and what

is coming out is unclear.

DR. SALOMON: So I think the suggestion would be to better characterize--I think, as an overall safety concern, is the idea that what is happening is mixing, matching and evolution of the species in the system as a model for what might happen in humans.

I think that, to understand better the mixing and matching, we go back to a discussion we have already had, that we are going to need to see more data in longer-term in vitro cultures with both characterization--I don't want to restate what we have already agreed on and restated.

I don't know whether I want to go into saying--get into the details now because I think we are then going into the study-section mode again, whether you should do it in the NOD, or the NOD-SCID, or how you should do it. Again, I would love to have those conversations, but I don't think that is appropriate for today.

DR. NOGUCHI: Some of this is sounding, to get back into that, "We would like to see something else like a nonreplicative recombinant, and an assay for that." Yet there has been a discussion as well, maybe that is too much to ask at this

time.

DR. SALOMON: I don't think that is too much to ask. I don't think anybody told you that was too much to ask. I think everybody here is saying that we want evidence that there is not a real evolution, if it is non-replicative, of viral sequences in this kind of a system and, if there is, I think we need to know about it and try and regulate on that and consider its safety intelligently.

DR. TORBETT: I think this is an important point because we are using not a self-enacting vector but something that has an active LTR which can make a full-length transcript which increases the chance for recombination.

So I think these are serious considerations. I don't know which vector is better, whether it is a SIN or an active, but I think if you are using it as a full-length transcript, then some of this information needs to come out for safety concerns.

DR. EMERMAN: This is Mike Emerman, again. It was alluded to that SIN vector wouldn't work. It would be interesting to note that was actually tested. I guess my sense of this is that the

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antiviral effect depends on copackaging. does depend on copackaging, it is hard for us to ban vector mobilization except to have it characterized. I think, in terms of an DR. SALOMON: assay design for assessing mobilization, I think what we all agree on is that it should be done in long-term, more complex, multicellular in vitro cultures. Now, whether or not you should also include animal studies, sort of building on--you do have some expertise and have been pretty successful with your SCID studies. I am, again, trying to articulate a decision here, or a recommendation. think that would be a positive. I am not certain, though, as I said, that I would insist that you do that because I think that -- I am not sure how that -- that is science. That is not necessarily simple testing. DR. DROPULIC: It is an extreme amount of work to do that. DR. SALOMON: As I said, as my laboratory does it, I absolutely agree. It is a lot of work. Are we done with Question 3?

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Part b) of Question 3 is more

DR. WILSON:

specific to the study subjects.

DR. SALOMON: Yes; I missed that right now. But you are right. So what should we do--we have gotten so focussed on this first part that we haven't talked enough about what is happening in the study subjects which is what we are being reminded of now.

So what should be done on the study subjects in terms of looking for vector mobilization?

DR. TORBETT: Many of the things that we discussed, the technology is very similar, whether it is before you put the cells in, do it in culture or coming out of the patients. That is very similar.

So I think that the same kinds of studies we have discussed in the past need to be applied to the patients as well. I am opening that up for discussion, I guess.

DR. SAUSVILLE: I would echo that. I think that is the logical follow-through, the issue about the evolution of this virus or viruses into something that--we don't know exactly what we are going to see in the clinical situation.

I would just note that, in the long list

of things that are going to be looked for in the clinical trial, I didn't go back and look, but this issue of variants that emerge, I think, is very important and should be captured. That would include whether you mobilize some aspect of the vector or whether you change the population of the HIV that is running around.

DR. SALOMON: Dr. Zaia?

DR. ZAIA: I just want to say that within the clinical-trial groups, AIDS clinical-trial groups, that are looking at fitness of virus relative to new drug additions, that technology is well worked out so you are going to very rapidly detect changes in fitness in this population of patients who clearly will have virus in their blood.

DR. REITZ: I think, also, you want to follow the virus populations with some sort of genotype analysis over a period of time. You will learn stuff from it.

DR. SALOMON: I think that there is nothing but logical follow-through here in terms of saying that the same concerns we have of mixing and matching viral parts leading to evolution of the species is as big a concern after the treatment as

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before. I think everyone has said that.

Therefore, if you agree, the advice would be that it should be characterized biologically, on multiple cell lines with different tropisms but, also, some of it has to be done molecularly either through genotyping or through specific--I think what Dr. Zaia is saying is interesting because it also implies that some of the things could be just looking at changes in drug resistance which could be done more rapidly than full sequencing and trying to imply changes on that basis.

Is that right, Dr. Zaia?

DR. ZAIA: Yes, but I was just thinking about fitness of the virus, per se. If we are most concerned about that, we could easily do that.

DR. SALOMON: So fitness would be like a quantitative assay that demonstrated just an increase in the rate of spread through a given indicator cell line. Okay; I stand corrected.

Dr. Long?

DR. LONG: I would just like the ask the panel to consider what kind of assay would be appropriate given the fact that they only detect one variant when the screen 240-some clones sequencing.

DR. SALOMON: No; you didn't get that right. There were hundreds of variants. About 80 percent of them were changed. That is a rough calculation. It was 264 clones. 240 had various mutations.

DR. DROPULIC: No. They had all deletions. Most of them were deletions. There were relatively few that had the base substitutions that we are seeing. 91 percent were deletions.

DR. COHEN: I am Ruben Cohen. I manage the clean room for VIRXSYS. I would like to just offer kind of a global perspective in terms of patient safety. I have graduated from GTI so I know a little about the patient trials there having been involved in training the medical centers that were involved in collaboration.

I also would like to say that one of the reasons I am happy with the way VIRxSYS is handling this is because I have also come through the agricultural world. The fact that I consider this vector is under a sentinel control, its expression is limited to the cells that basically have some activity going on with relationship to what the problem is.

One of the lessons I would like the FDA to

be thinking about is that, having come out of the agricultural world and working in molecular biology there, the antisense tomato is not something we buy in the market. The reason, I think, for its failure is because it was under a constitutive promotor, it was always expressed. It was everywhere and everybody was afraid of that.

The fact that this has a kind of a sentinel function, it only works where it is needed, I think is both expressing something towards both the safety and the appeal of the way this product is being handled.

DR. NOGUCHI: I think the point is well taken. However, I think that the reason that we are actually having this extensive discussion is that the absence of evidence is not evidence of absence. The fact that this is to be a sentinel-only function is, in fact, the question.

We agree that the scientific basis for this is to have it work only in that which is affected, but what biology teaches us, and, from GTI, you know this as well, what we believe today is not what we know to be true tomorrow and it is very likely not to be what will be approved.

So we need to be able to go through this

in an open a fashion as we possibly can. I think the fact that we have spent such a grueling set of time here really going over all the pluses and minuses and the pros and cons really does illustrate the intense interest in being able to develop something that does have this type of specificity.

As is often said, now we are getting down to the details and both god and the devil are always in that. So your point it absolutely well taken, but we need to be assured that, in fact, the sentinel remains specific for only one particular cell.

DR. DELPH: I have a question. If mobilization occurs into other tissues, is it necessary to look at viral reservoirs and characterize HIV species there. I am asking that because the species that you find in viral reservoirs may be very different from what you find in blood.

DR. SALOMON: I think that is fair. One of the difficulties, of course, is sampling and characterizing viral reservoirs. So, again, I would defer to my colleagues who do HIV as their primary business.

Dr. Zaia?

DR. ZAIA: I would suggest that that is a question for the phase II study.

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DR. ALLAN: You could actually look at macrophages and see if you get mobilization of the macrophages and, if you do, what effect that might have on function. So I think there are some things you can do even within the blood compartment.

DR. DELPH: You can look at semen.

DR. SALOMON: Yes. You can. That is a point. I think we should let it stand and I think Dr. Zaia's comment about whether that should come in in a phase I or a phase II is also well taken.

I think we are done in the sense that, what I was going to say is Question 4, we have really covered. I think we all agree on the basic principles here. I think we have also articulated for you up to the edge how far we can go without it getting gray.

Yes? Dr. Zaia?

DR. ZAIA: There is one aspect, though, of Question 4 that I think needs to be discussed and that is the safety assessment is linked to the rules of escalation. The rules say that we will look at 28 days and look for toxicity. If the

toxicity is not there, we will escalate.

I just have a problem with that. I don't think 28 days convinces me that you can see some of the things that we have been talking about. But, more importantly, I don't even think this study needs to be a dose escalation, unless you want to dose escalate the transducing agent. But that is not the research agent, or the investigational agent.

So I would really encourage the sponsors to rethink the design of this study because they are not asking how to get T-cells expanded and infused safely. I mean, there are several other studies I am sure the FDA knows more about than I do where they are infusing cells around the same level, 3 times 10<sup>10</sup>, and we all know how safe that is.

I don't think there is anything about these cells that make them more dangerous in regards to infusion-related toxicity. What is different about these cells is these other toxicities that we are talking about which are more, I guess, virological or less easy to elucidate within the normal observation periods that we look at in the standard trial.

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So I think that is a critical question that the sponsor has to contend with, I think, is how can they use this dose-escalation rule for the kinds of toxicities that we are most concerned with.

DR. SALOMON: Actually, that is perfect. What I was going to say is I think we are kind of done with the questions. What I would like to do now, in coming to a close, is for people to weigh in with other things here because I have got a list, too, and I wonder if some of you don't.

We have addressed the questions. I would say that there were a couple of things that bothered me. That was one. I don't think it is very clear to me whether the dose escalation--and the way I get it is the dose escalation is in different patients. Part of me is going, well, why not--you could do a dose escalation in the same patient.

Certainly, I would echo Dr. Zaia's comments that a 28-day--the things that are really safety concerns to me; 28 days? No; I don't think so. I would say more like three to six months.

DR. CHAMPLIN: Although these are not dose-related issues so much.

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DR. SALOMON: No.

DR. CHAMPLIN: So the issue of infusion toxicity is sort of all you are looking for realistically in the short term toxicity analysis and the emergence of resistant virus, of course, over a longer period of time and is probably going to be dose independent. So you could sort of minimize the importance of the dose escalation because that is just to establish the tolerance to the infusions. But, really, you are looking at the sort of long-term phase-I aspects of the biology of the whole approach.

DR. SALOMON: That is good point. But, then, I might say, that you don't need 28 days to find out. I guess I am a little confused on the premises here. There is toxicity from infusion. You know that in two or three days.

DR. HIGH: But if what we are talking about is doing better characterization of what is happening to HIV variants, you would like to at least wait 28 days, probably, before you even drew that blood. Then, once you have got it, it is going to take a little while to characterize it.

So you might not want to enroll the second patient before you have analyzed at least, at 28

days, what is going on with the first one.

DR. SALOMON: Yes. I guess what I am saying is I agree with you, Dr. High, and that is the conundrum. I think the way Dr. Zaia and I were coming at it was 28 days is not long enough to see the safety issues that we are concerned. But Dr. Champlin said, but the safety issues concerned with the dose escalation is simply a dose effect of the infusion.

My response to that is, okay, good point, but that is three to five days. We have got to be consistent here, logically. If the concern is an evolution of the viral species present, then I think a couple of months is probably appropriate. But that should be discussed.

DR. SAUSVILLE: Recognizing that that gets into the issue of why the dose escalation. It does lead to the protocol-design issue as to what are you going to consider your endpoint here.

Certainly, matters related to the infusion, while we don't think they are going to be a big issue, they are formally something that does have to be captured and scoped out.

Certainly, if we are looking at the incidence of variants mobilization et cetera, a

dose response relationship to input would be certainly of interest to also capture. So I guess I sort of come down in the middle, but I agree 28 days is probably a little soon. I do believe there is a role for dose escalation here and I do believe that there is a role for longer periods of observation before doing the dose escalation.

Let's put it that way.

DR. SALOMON: Dr. High, did that capture kind of where you were going?

DR. HIGH: Yes.

DR. SALOMON: Dr. Allan and then Dr. Torbett.

DR. ALLAN: Safety is also--you are hoping to get some efficacy with this or you wouldn't be doing it. But you know there is always the possibility that it is not going to have a good outcome. That outcome might not happen for six months or longer.

So my question, then, is do you need to rush doing dose escalation anyway. What is the need to have to do it after 28 days. If this is the first patient or the first three patients who get this, do we want to sit back and go, okay, what is going to happen, let's see what happens before

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we do anymore and let's see if we get a--not to look at efficacy but to look at is there an outside possibility that it may not be a good outcome.

So I would say you want to wait.

DR. CHAMPLIN: You need well defined early stopping criteria, what events would make you not enter another patient. You need to define exactly what those things are and then work out your accrual of patients accordingly. So, obviously, the faster you do the initial process that allows additional accrual, the faster they can complete the trial.

If you ask for a year follow up for every patient before you enter another one, it is going to take forever. So there has to be some sort of middle ground where you have an accrual of a reasonable number of patients that you can observe and then make it a go/no-go kind of determination should you continue the trial.

DR. SALOMON: Dr. Torbett and then Dr. Sausville.

DR. TORBETT: I guess I have a little bit more fundamental question. In some of the earlier data, it was shown that, depending on the number of integrins per cell--that is, the number of hits per

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cell--you can or can't select quickly the variants Am I correct? coming out. DR. DROPULIC: Can you repeat that? The question is, I think you TORBETT: showed us originally that, if you had one integrin per cell, or an MOI of 10 or 5, 10 and 20, the breakthrough came slower or faster. Right. So, as you dose DR. DROPULIC: down the vector -- I am not so sure faster. I am sorry about that word. DR. TORBETT: DR. DROPULIC: We saw breakthrough at a transduction MOI of 5 and a challenge MOI of 0.1 in that Sup-T1-cell experiment. DR. TORBETT: The reason I am bringing this up, it comes to the whole question -- and it wasn't clear from the document that I had what the MOI is per number of cells. But depending on the 17 number MOI per cells, even if you can get 100 18 percent and you increase the number of integrins, 19 then the time period to look for variants coming 20 out would vary. Is that logical? 21 At the optimally transduced DR. DROPULIC: 2.2 cells, we haven't see any. 23

Maybe it is the way I am -- it DR. TORBETT: is late in the day and I haven't had any more

sugar, so maybe that is part of the problem here. 1 2 But if you had one integrin per cell and, at a 3 certain time, you had a turnover virus, say you had more integrins, less, and you had 10 integrins per 4 5 cell--I am just making these numbers up--and you 6 had pretty good control but it took a longer time, 7 the length of time that you would want to look 8 could very well be longer and the time that you would want to sample each time would vary. 9 10 DR. SLEPUSHKIN: I just would like to 11 answer it. In any case, we have specification for copy number per cell. That won't change during 12 escalation. So there will be about the same amount 13 14 of integrins on all steps of escalation. 15 are just changing the amount of cell injury. 16 DR. TORBETT: So the MOI is going to be 17 constant. 18 DR. SLEPUSHKIN: Yes. 19 DR. TORBETT: Just, out of curiosity, what 20 is it? 2.1 DR. SLEPUSHKIN: I probably will be about 22 200 as it was in the animal or the clinical 23 experiment. 24 DR. TORBETT: So it is going to be an MOI

of 200.

1 DR. DROPULIC: That is not copy number. DR. SLEPUSHKIN: 2 Not copy number. 3 MOI; yes. 4 DR. TORBETT: I understand, per X number 5 of cells. Okay. 6 DR. SLEPUSHKIN: Copy number 7 specifications; copy number per cell should be between 1 and 10. And, in the clinical animal 8 9 experiments, it was 6. 10 DR. TORBETT: So is there any data suggesting -- well, I guess you can't control with 11 the variability. Maybe that is something that 12 shouldn't be brought up here. 13 14 DR. SALOMON: Unless you guys want me to, 15 I am not going to try and come to consensus on each of these points. I think we have done our job. 16 But I would like to, in this kind of concluding 17 process, have everyone share with you, and with the 18 FDA staff, additional things that were not in your 19 20 question list. So I believe my job as chair has suddenly 21 now to just making sure that everybody gets a 22 comment and everyone gets heard. So unless someone 23 is insisting that I come to consensus, I am not 24 25 going to.

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Dr. Torbett?

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DR. TORBETT: This is a first trial and I would like to commend the company for going It was a pretty brave step and I think, forward. without too many boundaries, they done a fairly admirable job of starting up the stairway.

DR. DELPH: I would like to echo that. would also like to express some concern about the patient population that has been selected for this I really think a CD4 count of 600 as an trial. upper limit is way to high. right now, even the DHHS guidelines for treatment are really not coming out in favor of recommending treatment until CD4 counts drop below 350.

In the European guidelines, they are even dropping them as low as 200. Generally speaking and, of course, there are exceptions, patients with CD4 counts of over 200 are not that ill. would certainly -- I understand the difficulties that the company says that they may have in getting Tcells from patients who have CD4 counts of under 200, but I think we need to also need to balance that with the need for the patient population and what is safe for them.

The other thing, and I don't know what

your actual inclusion criteria are for "failing or discontinued HAART therapy." But I think that needs to be far more clearly defined and I would recommend that you define it in terms of the number of drug classes that someone is resistant to. I would recommend that someone be resistant to at least two of the drug classes currently on the market and possibly even three, but at least two protease inhibitors.

You are going to look at people who really have few, if any, drug options left. So those are my major comments.

DR. DROPULIC: I appreciate those comments and I just want to assure you that we want to work with the FDA to finalize these patient criteria. It is not set in stone now and your comments are well taken.

DR. DELPH: I also suggest that you work with the HIV community on that.

DR. DROPULIC: Okay. Yes. Thank you. Yes.

DR. LEVINE: Let me address the issue of the cell number, if I could. Once you get below 200 cells, there are increasing difficulties with the transduction and the culture. I think if we

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could make an analogy about immunologic treatment in a different setting, with Cliff Lane's studies, where he has given IL2 to the patients, there was a real dichotomy with the effectiveness of raising the CD4 count with patients who came in with a count above 200 versus those that came in with a count below 200; that is, there was not an effective of the IL2 once you had patients coming in below 200.

so I think, at that point, the immune system has suffered what may be irreparable injury and would make any trial safety and feasibility more difficult.

DR. NOGUCHI: The question is not simply one of difficulty. I will point out that interleukin 2 has been actually approved for the treatment of renal-cell carcinoma so that the parameters of experience are far different than what we are talking about here. But we do understand the technical aspects of this particular approach which is trying to take cells and expand them. There very well may be areas that we will need to really fine tune.

DR. CHAMPLIN: They are few and far between, but the availability of identical twins

sometimes can give you sort of an opportunity to treat a seriously ill patient who now has a normal twin that you can get unlimited numbers of leukocytes from. So, for proof of principle kinds of things, that offers, sometimes, unique opportunities.

DR. SALOMON: Dr. Golding.

DR. GOLDING: I just to bring a little perspective from the Office of Vaccines. Our group is responsible for a lot of HIV vaccine including therapeutic vaccines. Most of the therapeutic vaccines, as you know, are done in the context of antiviral therapies. So what are sort of guidelines that we are using in terms of the safety monitoring of the patient.

Of course, just like in this case, when an outcome can be either no effect or worse disease progression, is the same thing we have to deal with when we deal with when you deal with therapeutic vaccine, that even though you gave something that is supposed to help to control viremia, you actually have a negative effect.

I think that can be seen relatively quickly by measuring viral-load changes in the patient. So I think it is important, once the

cells have been transduced into the individuals is to really take multiple measurements of viral loads over this first period, whether it is one month or six weeks, and to actually have a sense of the slope of viral load measurement as well, of course, of CD4 counts because you are talking about all kinds of toxicities.

There is the infusion toxicity. There is potential emergence of more fit viruses that, down the road, can dominate the patients. But I think if you have a really adverse reaction, you have something really bad in your product, what you are going to see is much more rapid increase in viral load.

For that, you have to have patients that are not in the millions of viral loads to start with. You have to have a window that would really allow you to see some really enhanced increased.

DR. SALOMON: Right. The logic to what you have said, Dr. Golding, though, is that you are probably going to want to choose a patient population that is not all that far gone and wildly rising HIV viral loads on the day that treatment is initiated.

DR. GOLDING: I wouldn't say total failure

of anti-HAART with a million copies because I really don't think you will be able to see this type of adverse reaction that you want to use to allow you to go to the next patient.

DR. SALOMON: So we have to put that in the context with what Dr. Delph shared with us in terms of the patient selection.

I had some other concerns I wanted to put on the table. One would be, in terms of the CD3, CD28, beads, you have less than 100 beads per 3 million cells which means you could literally be putting in thousands of beads into someone. I do not buy that one at all. I think you would cause a pulmonary embolus. But maybe you have experience to answer that.

DR. LEVINE: That number was developed in consultation with the FDA. There have been toxicity studies done by what used to be Baxter Immunotherapy infusing a large number of beads into rats looking for that sort of thing. There were no toxic effects at levels of beads very much higher than what we are infusing.

We are also very much below that number of 100 beads per 3 million cells. What we are able to achieve currently is a depletion of greater than

six logs to what we start with. So if we were to stimulate 50 million cells, 100 million cells, a billion cells, and have, say, a 3-bead-to-cell ratio of, let's say, up to 2 to 3 billion, we would anticipate easily being able to deplete six logs of those beads.

DR. SALOMON: Another issue; I think I understand why you want to activate these cells is because you think you are going to have very low numbers and you want to get up to these higher numbers of 10<sup>10</sup> and higher in your infusion.

Pheresis, even--I am not getting into the state of the T-cells when you are really getting down to 150, 200 CD4 cells. Your comments stand on that. But I don't understand that. I don't understand why everyone wants to ignore the biology of T-cells and activate them and culture them in nonphysiological concentrations of interleukin 2, inject them back in the patients.

I mean, the whole purpose of a lentiviral vector is it is incorporated into non-replicating cells. So my murine Moloney leukemia virus backbone, I have to activate my T-cells and I am not happy about it. But you don't. For studies that you are trying to maintain a normal immune

repertoire yet you are doing these things that I don't think there is any data here demonstrating what the immune repertoire is that is left in these cells after you do this. I just don't understand why you want you to go there.

DR. LEVINE: I can tell you that the maintenance of the repertoire after 60 days in culture is published in 1996 in the Journal of Immunology showing that we do maintain the entire repertoire.

DR. SALOMON: Defining repertoire as the CD4/CD8 ratio?

DR. LEVINE: As 24V beta families as analyzed by the CDR3.

DR. SALOMON: Okay. I guess, again, this is not a comment coming from the chair. We are not going to try and get consensus, so just a comment to you. I just don't believe that these assays maintain the normal T-cell phenotype. To go into these initial studies at the early low-dose effects where you don't have to activate and you don't have to treat with interleukin 2 just seems to me you are adding another variable to prepare yourself for a later thing based on an assumption that you have maintained your repertoire.

DR. LEVINE: I would say that we have experience with these T-cell infusions, with CD4-cell infusions and with bulk T-cell infusions in HIV patients and in cancer patients. We, ourselves, have done 51 infusions in HIV patients. CellGenesis has done with CD3-28-stimulated cells I am guessing 60 or 80 infusions and, in cancer patients, even more.

So I think we just have to agree to disagree.

DR. SALOMON: Right. That's fine. That is perfectly fine.

MS. KNOWLES: I would like to take Dr.

Delph's comments one step further and caution--in

terms of her comments about the other

pharmacological agents in the research pipeline

because she is right. There are more things coming

down the pipeline. As such, I would like to

caution the sponsor to not put the message forward

that your proposed clinical trial is going to be a

last-ditch treatment effort for people with HIV

because it is one potential of the armamentarium.

DR. DELPH: I have another question because it wasn't clear from what you have given us about the protocol. Are these subjects going to be

on antiretrovirals or not?

DR. DROPULIC: They will be failing HAART and, if they are not on a therapy, then they are not on. But we are not going to require them to come off therapy. We think that that is unethical. So, if they are on one or two drugs and they are failing therapy, then they can enroll in the study. That is how we have defined it so far but, again, we can negotiate this with the FDA to see how we approach this. That is how we have characterized it presently.

MS. KNOWLES: If they go on study drugs, are you going to pay for them? Who is going to pay for the drugs?

DR. DROPULIC: I hadn't thought of that. We will think about that one.

DR. TORBETT: You propose in your set of criteria that you follow these individuals for life. Who would pay for those, assuming that the company had problems?

DR. DROPULIC: We plan to be around a long time.

DR. TORBETT: In any event that you don't, is there going to be--I am just curious. Would you take out insurance to make sure that that is done?

This is a serious consideration. It has been discussed before. I am just curious.

DR. DROPULIC: If that is a requirement, we can do that.

DR. TORBETT: I just wanted your thoughts on that.

DR. DROPULIC: Haven't thought about it, quite frankly, because we expect to be around a long time.

DR. SALOMON: I think one of the comments
I have, and this is not specific to VIRxSYS, but
that I think the focus of these discussions in the
last two days have, and perhaps very appropriately,
focused on the biggest risk, the low-hanging fruit,
if you will, of the replication-competent
lentivirus and shuffling of the DNA species, et
cetera, which is fine.

I guess I still feel like, as part of this sort of last number of comments here--it continues to bother me what is happening also to the trans gene that is being delivered, the payload, if you will. That, to me, is as much a part of the product as the issues of safety.

Here, you get close to this gray area of "okay." But remember this is phase I and we want

safety not efficacy. But, as Dr. Noguchi said, when there is significant risk and unclear benefit, it is very hard to construct risk/benefit ratios and I think the rules change.

We have been through that with xenotransplantation. So, going back to that, I just think that--one of my personal comments here is that, at some point, we need to also consider how we are characterizing the quality and the integrity of the payload through all these changes because everything we have talked about, up to now, has not really dealt with that.

DR. NOGUCHI: I would actually disagree. I think there has been a lot of very good discussion on that, and you note that Dr. Wilson and Takefman are diligently noting these things. It is actually central to some of the evaluation because it does appear as though the payload may actually push the virus to recombine and do different mutations, deletions and so forth which clearly is an activity we need to be monitoring even from just the safety aspect.

So I think we have actually gotten very good advice on that.

DR. CORNETTA: This is Ken Cornetta. Can

I make a comment?

DR. SALOMON: Yes; go ahead.

DR. CORNETTA: I guess just maybe to pick up a little bit of what you were saying about the T-cell function after transduction and the stimulation process. That bothered me, too, as I was reading through. While a lot of cancer patients have gotten T-cells that have been manipulated and given back, our experience, although limited, has been that those cells don't function very well, at least after allogeneic transplantation.

So, in the process, their ability to do what the T-cell initially was designed to do seems to be lacking. So one of the real advantages I saw to lentiviruses was that you might be able to avoid this in vitro stimulation. It bothered me a little bit that there seemed to be a fair amount of stimulation that would occur in these cells and that, again, the concern that their ability to function, once they got back into the patient, would be not as we hoped.

DR. SALOMON: Do you want to comment on that?

DR. LEVINE: Yes. We have several lines

of evidence, and I could spend an hour talking about them, that, by stimulating by CD3 and 28, we reverse defects of the T-cells as they are removed from both cancer patients and HIV patients.

We have recently completed at phase I study in lymphoma where we have looked at intracellular cytokine response following TMA and antimycin stimulation at Day 0 and at Day 12 of in vitro culture and showed that we can reverse what is a substantially diminished response at Day 0 that is increased at Day 12.

In the HIV setting, we have looked at response to allogeneic mixed lymphocyte reaction and show that we can increase in the study population that we did several years ago--we increased that allo-MLR response.

With respect to a CCR5 population that is different from the population with this study, we looked at CCR5 in vivo expressed on CD4 patients, both before and after infusions, dose-escalating infusions of 3, 10 and 30 times 109 and could show, specifically on the CD4 cells, that we have reduced the CCR5 levels.

Also, in vitro, we have looked at cytokine production in the HIV patients both before and

21.

after stimulation and it is very much higher. So I think there is a wide spectrum of T-cell functions that are improved following CD3-28 stimulation.

I think the point also is that it may be nonphysiologic but that might be better. So, by not stimulating CHLA4, by stimulating CD28 specifically, you are upregulating BCL2 protecting against apoptosis. So there really is a wide spectrum of things that are improved following CD28 stimulation.

DR. SAUSVILLE: I guess I would offer that, certainly if this is successful to the extent that we work through some of these issues, or the sponsor works through these issues and gets into the clinic, one can imagine many different flavors of mix and match. That would be the subject for future clinical investigations.

I certainly would agree that, being vested in this particular way of doing it, this is one way to do. And I would leave it at that.

DR. SALOMON: Fine. Again, as I said, we had agreed to disagree a bit ago and nothing has changed.

Any other comments or can we come to a close?

DR. GAYLOR: I haven't said anything for the last two days and feel compelled to earn my way here somewhat. But, as a statistician, I need data to work with. There is obviously a paucity of data here for understandable reasons. It is a brand-new area. So my role, I think, has changed from being one that could have any scientific input to really how does the man on the street feel about this, somebody that not really been terribly involved in this.

I feel very comfortable with the discussions I have heard. There has been a lot of thought. A lot of questions have been raised. There has been a lot of good discussion and I guess, again, it is a theoretical comfort because I don't have a lot of data to look at.

But I think the committee and the people involved, the research that has been done, makes me feel like everything is being done that could be done at this point.

DR. SALOMON: Again, anyone else have any final comments that they would like to make?

DR. NOGUCHI: On behalf of the FDA, I hope I could take this liberty to really thank the committee, VIRxSYS especially, for being so bold as

2.4

to come here and face the stings and arrows, I guess, as best we can put it.

The committee, especially, for this round, has been exceptional both in its civility as well as rigor in pursuing obvious and not-so-obvious questions. I would especially like to thank our chair for keeping us on keel and getting us through this very difficult set of questions.

I think that, on our side, we can say that, with this advice, we are confident we will be able to make the appropriate decisions to move the entire field forward and we thank you for that.

DR. SALOMON: Thank you, Phil. Then, as chair, let me speak for everyone. I think that VIRXSYS, you guys did a really good job. I have always said, going back a couple of years now, that this committee functions the best when a sponsor can step up and provide us a real protocol to look at. That is when we can really deal with the kinds of specifics that allow the fields to move forward.

You guys have done that and I respect that. I also thank the sponsors who presented yesterday for doing the exact same thing in a situation that they even have more to lose, if you will, because we were taking them on on some of

their things that they hadn't even brought quite as far as you guys have. Again, I thank them.

I think everyone from the committee for two to three days, depending on which group you are in, hanging in there with us. To Rosanna Harvey and Gail Depolito and the rest of the FDA staff who worked so hard to put all this together, to get us here, to take us to dinner, to move us around in hotel rooms, and to the audiovisual staff and everyone else involved.

Thank you very much. Everyone travel safe.

[Whereupon, at 1:50 p.m., the meeting was adjourned.]

## CERTIFICATE

I, ALICE TOIGO, the Official Court Reporter for Miller Reporting Company, Inc., hereby certify that I recorded the foregoing proceedings; that the proceedings have been reduced to typewriting by me, or under my direction and that the foregoing transcript is a correct and accurate record of the proceedings to the best of my knowledge, ability and belief.

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