DEPARTMENT OF HEALTH AND HUMAN SERVICES
PUBLIC HEALTH SERVICE
FOOD AND DRUG ADMINISTRATION

MILK LABORATORY EVALUATION FORM

LABORATORY		
LOCATION		LAB #
DATE	X = DEVIATION O = NOT USED	U = UNDETERMINED NA = NOT APPLICABLE

DISINTEGRATION METHOD FOR PAPER, PAPERBOARD OR MOLDED PULP MATERIALS

[Unless otherwise stated all tolerances are ±5%]

1. Laboratory Requirements .....
a. Record time and date samples received .....
b. Record time and date samples examined .....

APPARATUS AND MATERIALS

2. See Cultural Procedures, items 1-3, 10-21, 23-32 as appropriate .....
3. Pipets, sterile .....
a. Capacity, 10 mL with 3 mm tip opening .....
b. Or, 20 mL with large-bore tip opening .....
4. Pipet containers .....
a. Used for sterilization, storage, non-toxic .....
5. Scalpel or scissors, and forceps sterile .....
6. Disintegrator blender .....
a. Sterile, high speed, electrically operated, corrosion-resistant cup .....
b. Capacity, 500 mL; optionally, 1000 mL .....
7. 70% ethyl alcohol .....
a. In a covered container to hold scalpels, scissors and forceps
8. Dilution buffer (Cultural Procedures, item 25a,c) .....
a. In containers filled to contain 300±6 mL (or 500±10 mL) .....
9. Sterile kraft paper or envelopes .....

PROCEDURE

10. Not applicable when wax, plastic or metal is food contact surface .....
11. Use sterile cutting device, cut 100g from butt roll and transfer to sterile wrapper or envelope .....
12. With sterile cutting device, trim off 5 cm of the outer edge of the sample sheet .....
13. Handling with sterile forceps, cut into 0.5 cm pieces 3g of the center portion into a sterile petri dish .....
14. Transfer this 3g into a sterile disintegrator cup containing 300 mL dilution water (5g in 500 mL) .....
15. Place cup on blender motor, run at high-speed 30 sec, check to insure no particles are on side of cup or trapped beneath blade .....
16. Continue high speed blending for a total of 2 min, depending on paper type, checking at intervals for particles on side of cup .....
17. Take great precautions to avoid dust, moisture, and other contaminants at all steps .....

PLATING

18. With sterile pipet, divide 10 mL of disintegrated sample equally among 3 plates (optionally use 5 plates) .....
19. Pour agar (see SPC, item 13), thoroughly and evenly mix with test portion in plate .....

CONTROLS

20. See SPC, item 14 .....

INCUBATION

21. See SPC, item 15 .....
a. Incubate at 32±1C for 48±3 hr .....

COUNTING COLONIES

22. See SPC, items 16, 17

REPORTS

23. Computing and Reporting Counts .....
a. Multiply the sum of the colonies on 3 (5) plates by 10 .....
b. Report as the number of colonies/g of stock .....