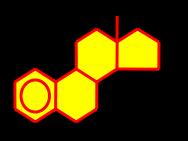
# Development and Pre-Validation of a H295R Cell Line Screening Test to Evaluate Toxicant-Induced Effects on Steroidogenesis



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#### Thanks To

MICHIGAN STATE

Ralph Cooper and John Laskey (US-EPA) Anne-Marie Vinggaard and Marie Louise Hagen (Danish Institute for Food and Veterinary Research, DK)

Yumi Akahori and Makoto Nakai (Chemicals Assessment Center, Japan)

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## Outline

- 1. Background
- 2. The H295R Steroidogenesis system
  - a) Performance criteria
  - b) Model chemical evaluation
- 3. Preliminary inter-laboratory comparison
- 4. Conclusions
- 5. Future directions





#### H295R Cell Line

× Human female adrenocortical carcinoma **¤** Produces many steroid hormones ✓ progestins ✓ androgens & estrogens Glucocorticoids & mineralocorticoids **¤** Expresses most of the important steroidogenic enzymes ✓ CYP11A, CYP11B, CYP17, CYP19, CYP21





#### H295R Cell Line

The cells maintain the capacity to synthesize most of the steroid hormones characteristic of three phenotypically distinct zones of the adult adrenal cortex

- ✓ Zona glomerulosa
- Zona fasciculata
- ✓ Zona reticularis

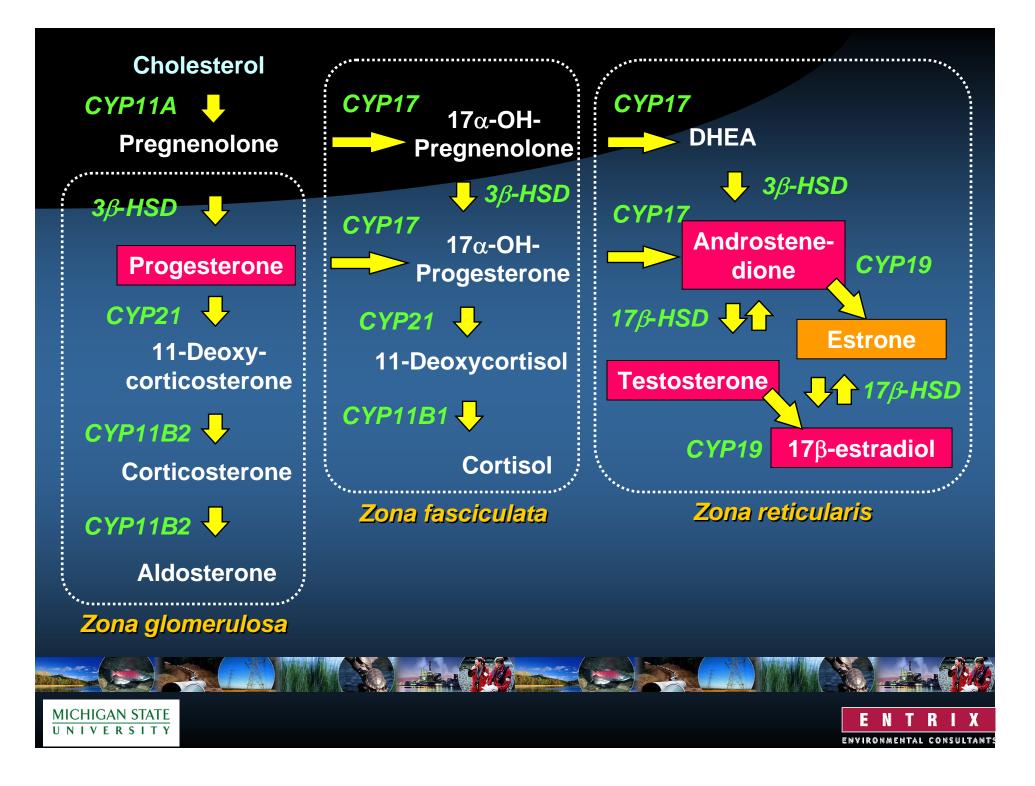




#### **Effects on Steroidogenesis**

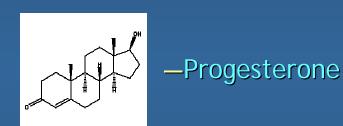
At level of expression
 measure mRNA levels: RT-PCR
 Effects on enzyme concentrations
 measure catalytic activities: selective substrates
 Effects on metabolism of steroid hormones
 measure steroid hormone concentrations

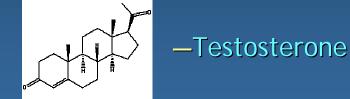


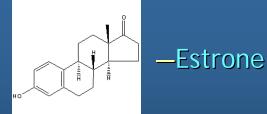


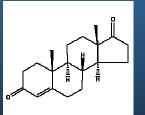
#### **Objectives**

Develop and optimize a rapid screening test to determine effects of chemicals on sex steroid synthesis:

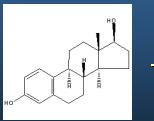








-Androstenedione



–17β-estradiol





# Objectives (cont')

- Demonstrate the performance of the assay with known inhibitors and inducers of steroidogenesis
- Assess and quantify sources of variability in the assay to:
  - Establish performance criteria for large scale screening of chemicals
  - Demonstrate flexibility and transferability of the protocol to other laboratories prior to conducting ring tests
- Develop optimized protocol for inter-laboratory validation phase.



#### Goals

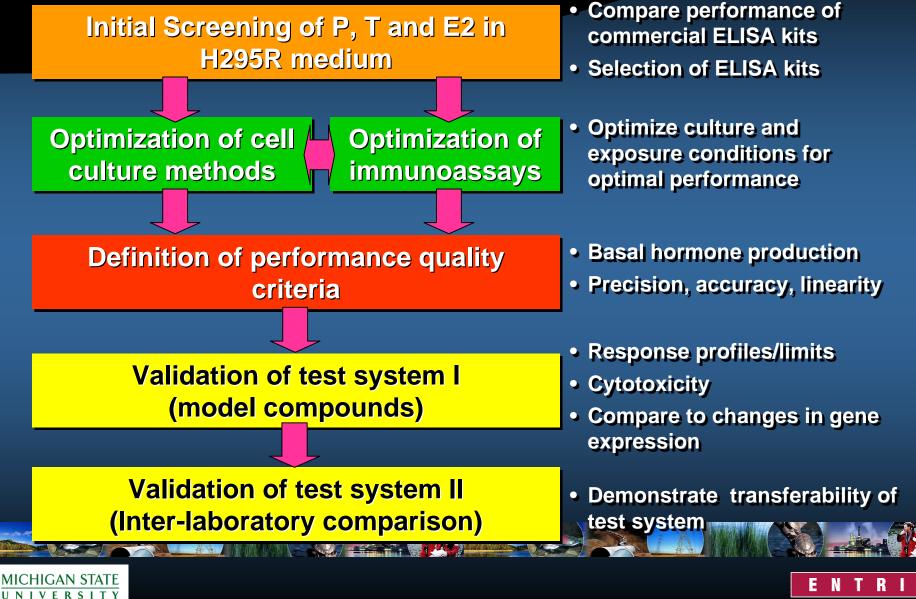
Establish an assay that will integrate possible effects on multiple parts of the steroidogenic pathway:

- **1.** Steroidogenic signal transduction
- 2. Regulation of cholesterol transport by the STAR-Protein
- **3.** Conversion of cholesterol to testosterone by:
  - **P450SCC**
  - **3**β-HSD & 17β-HSD
  - P450C17
- 4. Androgen conversion to estrogen by CYP19 aromatase



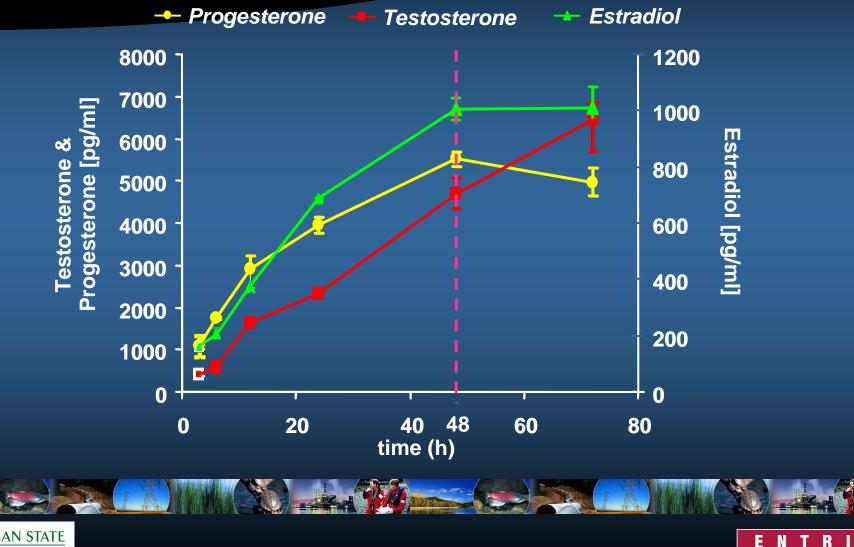


## **Overall Approach**



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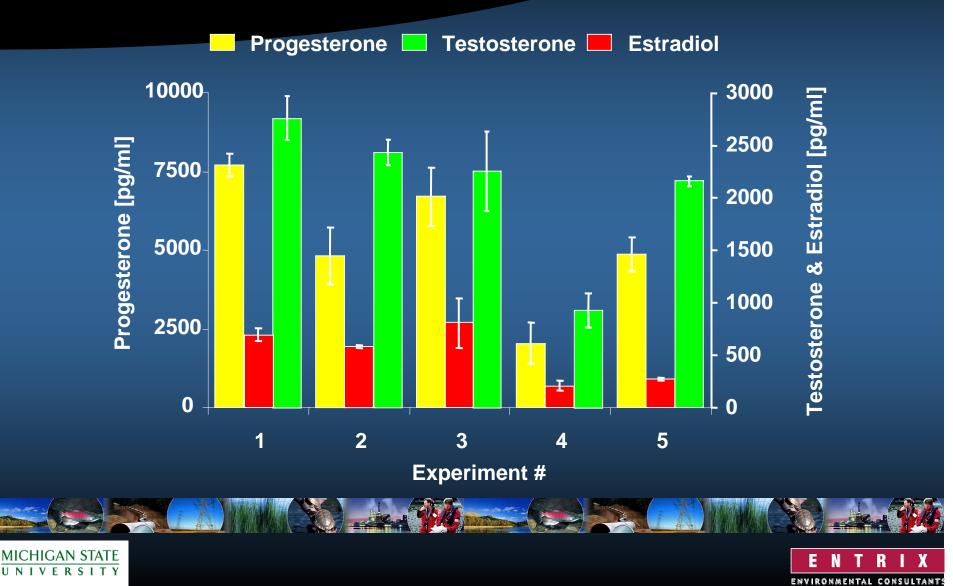
#### H295R Cell Test Development *Time Series*



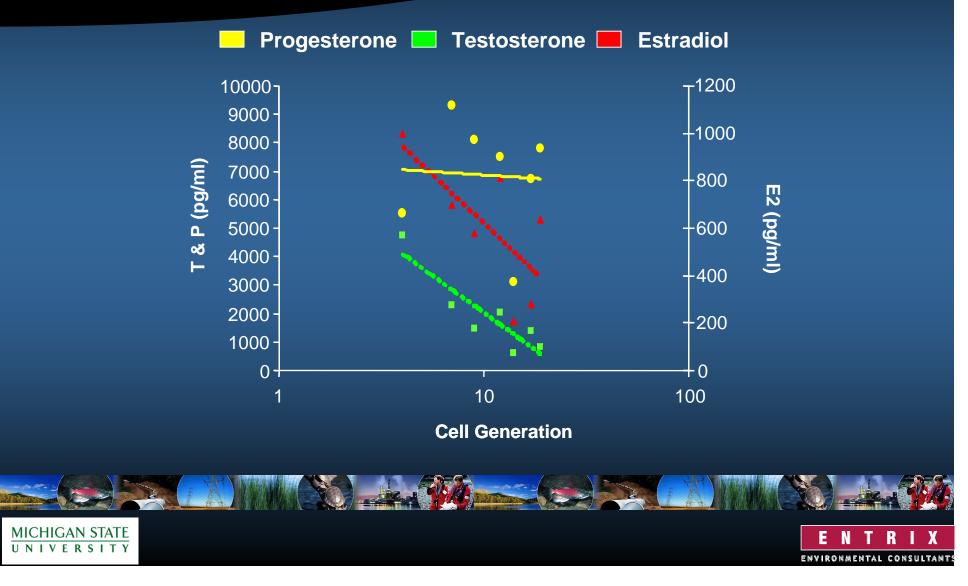
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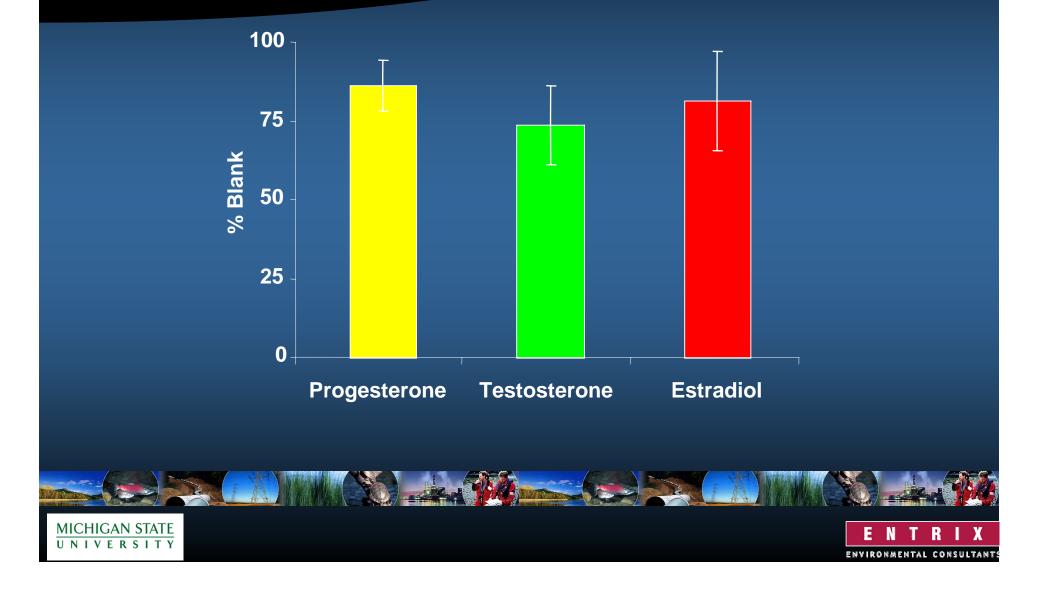
#### H295R Cell Test Development Basal Hormone Production



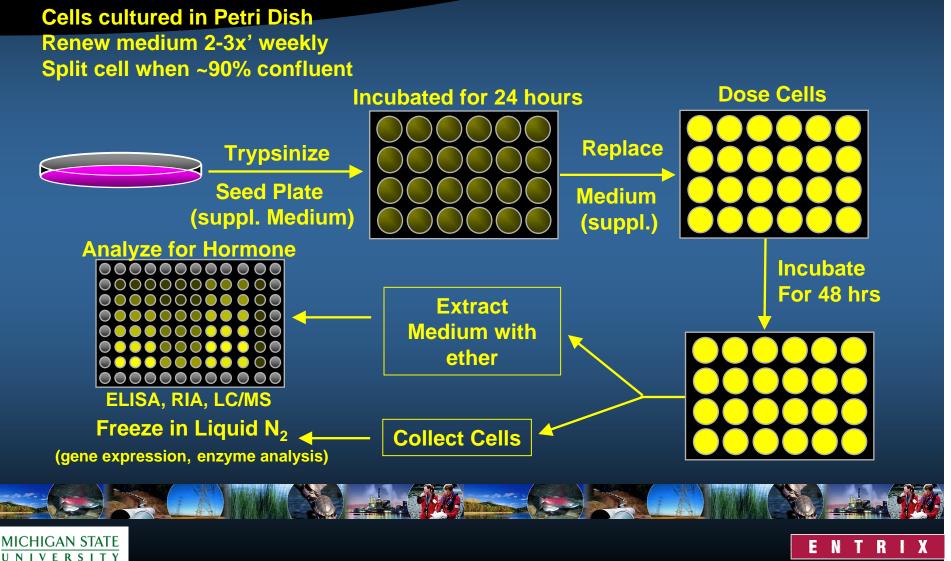
### H295R Cell Test Development Effects of Cell Passage



## H295R Cell Test Development Effect of Solvent (0.1% DMSO)



#### H295R Methods to Measure Effects on Hormone Production



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#### **Model Chemicals**

#### ¤ Prochloraz

 Imidiazol fungicide: Potent inhibitors of aromatase; Capable of affecting other P450 dependent enzymes

#### **X** Aminoglutethimide

 Generation I aromatase inhibitor; Can also downregulate synthesis of cortisol and aldosterone

#### 🛛 Forskolin

✓ General inducer: Stimulating adenylcyclase and increasing cAMP levels in adrenal cells

#### **¤** Ketoconazole

 Imidiazol fungicide: Inhibits p450 enzymes (24-hydroxylase, Cholesterol SCC and C-17,20 lyase)





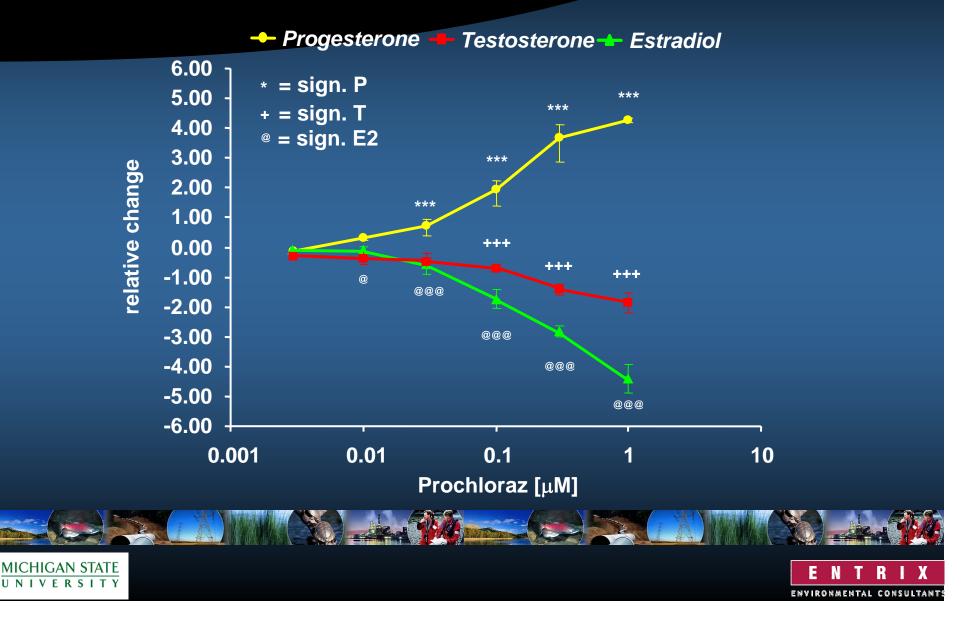
#### Cytotoxicity/Cell Viability

Prior to initiation of exposure experiments, cytotoxicity of each chemical was assessed using the MTT assay

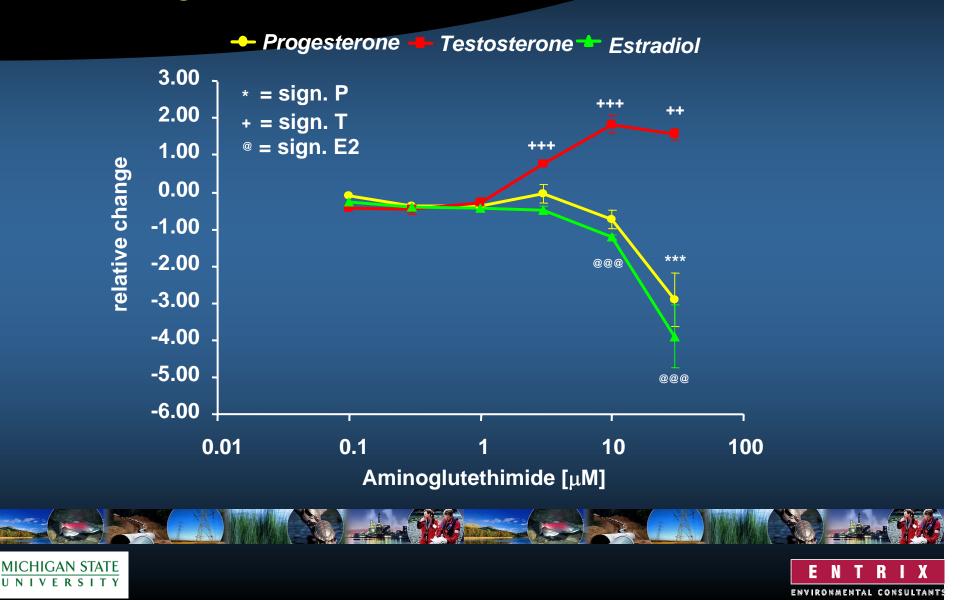
Dose ranges for all subsequent exposures represent non-cytotoxic concentrations



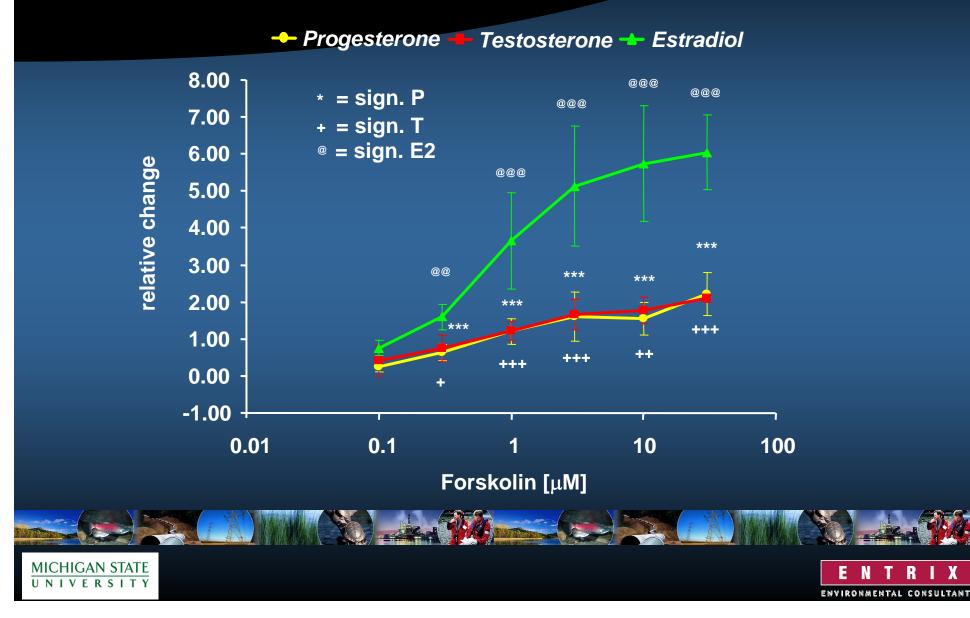
#### Model Chemical Exposure Prochloraz



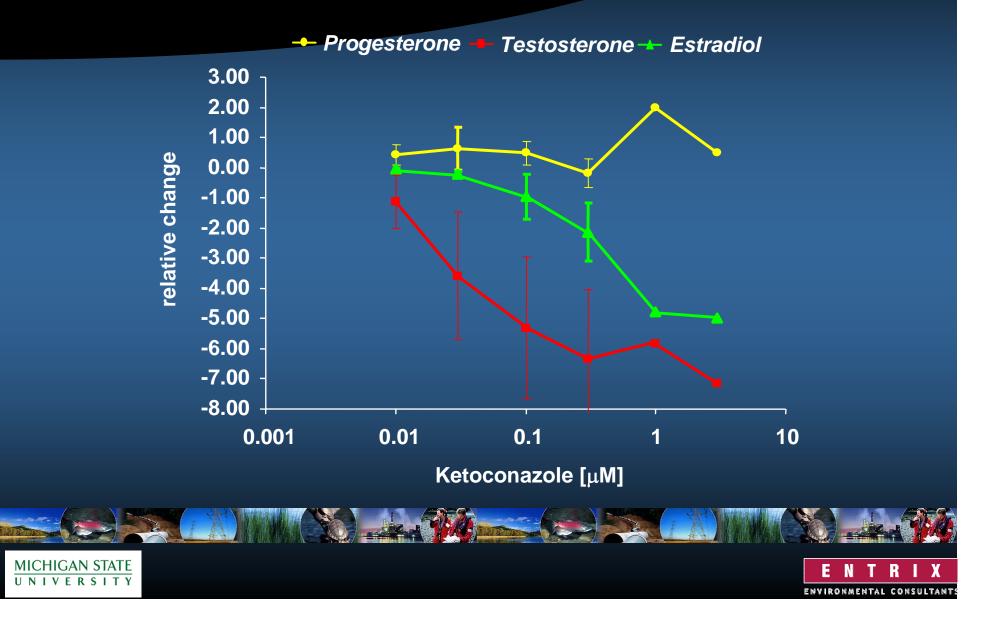
#### Model Chemical Exposure Aminoglutethimide



### Model Chemical Exposure Forskolin



### Model Chemical Exposure Ketoconazole (Preliminary Results)



#### **Inter-Laboratory Comparison**

#### **¤** Participating Laboratories:

- ✓ US Environmental Protection Agency Endocrinology Laboratory, U.S.A.
- Chemicals Assessment Center Chemical Evaluation and Research Institute, Japan
- Danish Institute for Food and Veterinary Research Department of Toxicology and Risk Assessment, Denmark

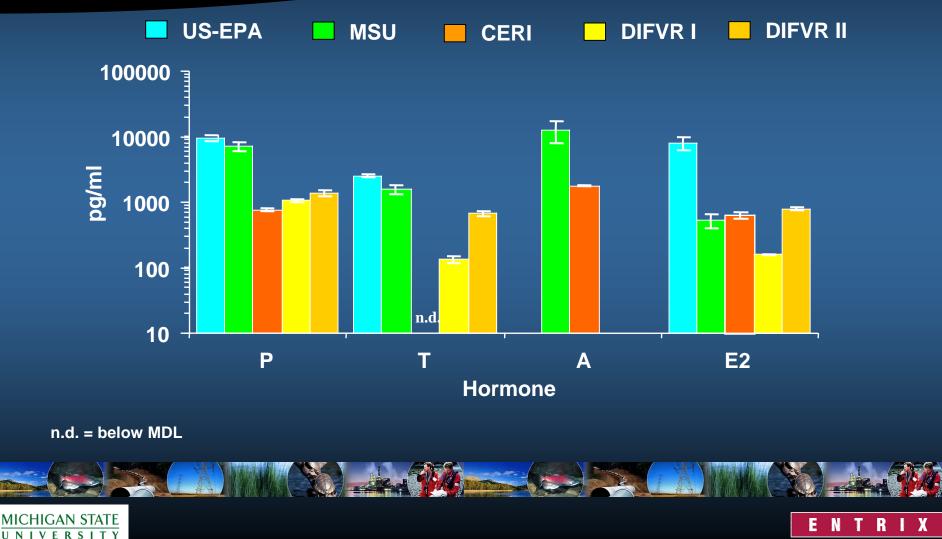


# Inter-Laboratory Comparison (cont')

Performance based comparison. Use of:
same cells/same passages
different cell culture protocols/conditions
same seeding density
same acclimation and exposure protocols/conditions
different hormone detection methods

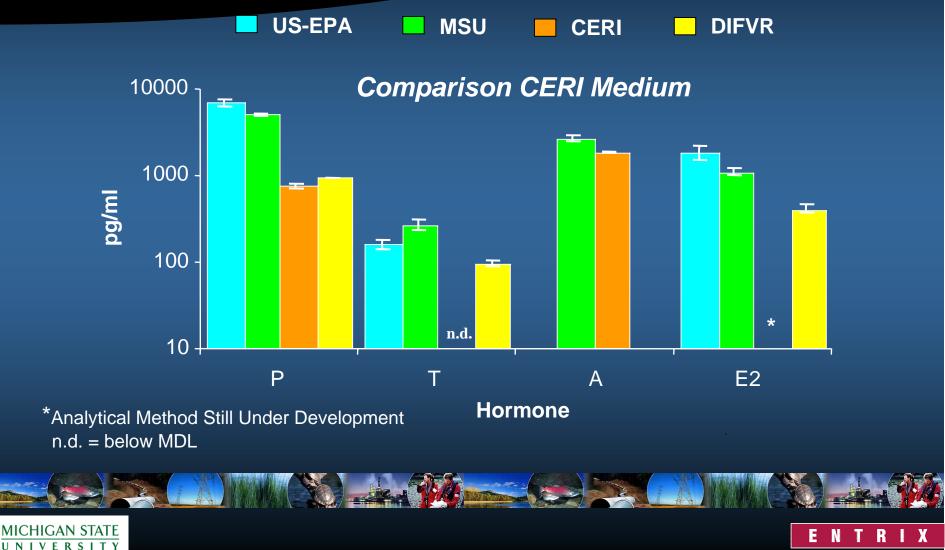


#### Phase I - General Test Performance Basal Hormone Production



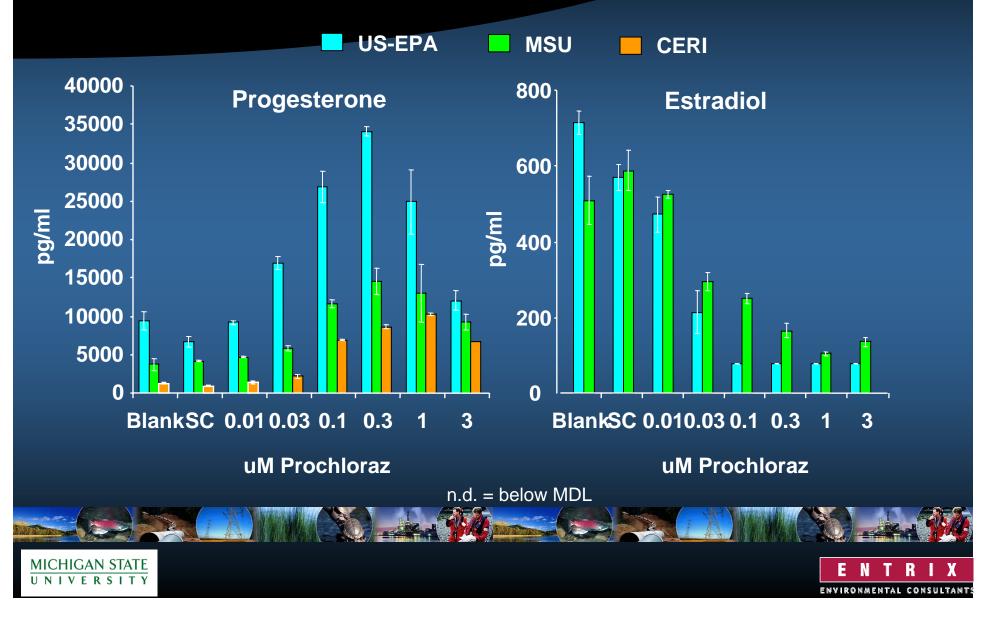
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#### Phase I - General Test Performance Hormone Detection Systems

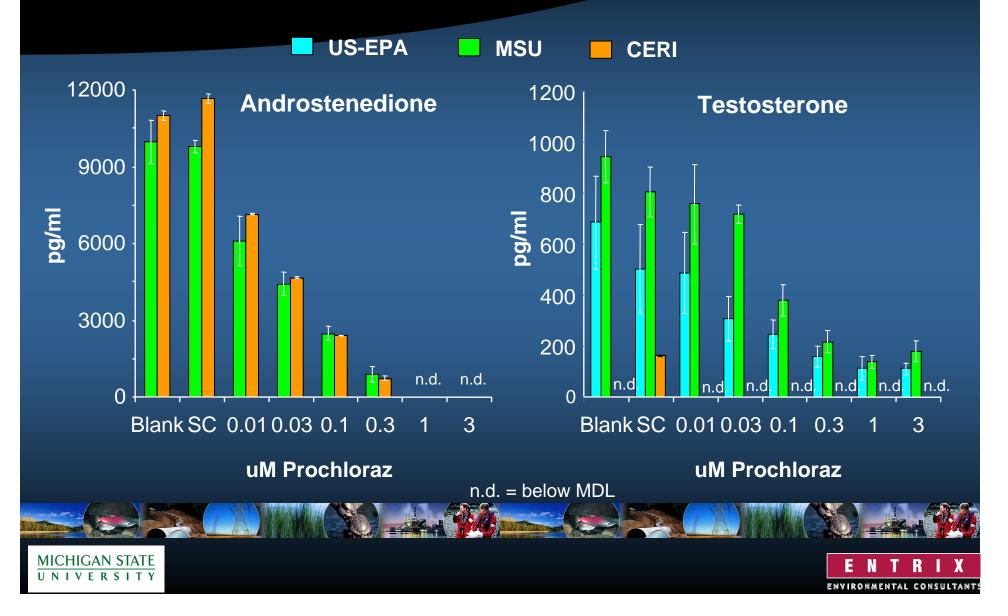


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#### Phase I – General Test Performance Comparison MSU Prochloraz Exposure



#### Phase I – General Test Performance Comparison MSU Prochloraz Exposure



#### Phase I - General Test Performance Summary & Conclusions

Some variation due to different hormone detection systems:

- Different antibody cross-reactivities (MSU P values explainable by cross-reaction with pregnenolone)
- ➡ Differences in clean-up/extraction procedures?

➡ Differences in sensitivity





#### Phase I - General Test Performance Summary & Conclusions (cont')

Variation of basal hormone concentrations measured at different laboratories

Different medium composition:

-Supplemented vs. non-supplemented medium

-Use of antibiotics





#### Phase I - General Test Performance Summary & Conclusions (cont')

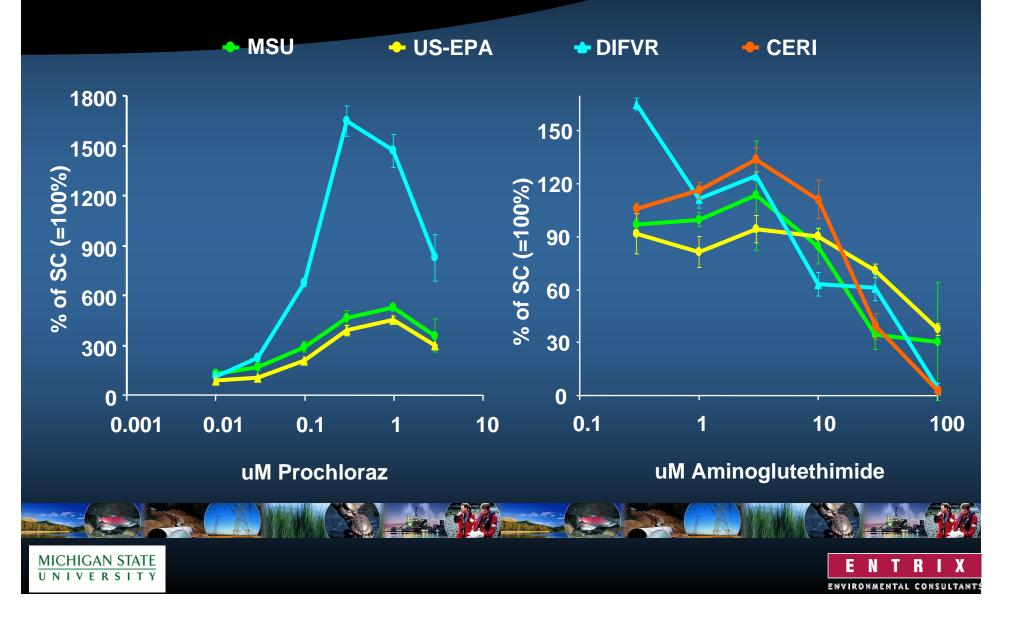
✗ Good reproducibility of results at each laboratory:

- Low intra-assay variation
- ✓ Low inter-assay variation
- ✓ Good linearity
- Good recovery of hormone spikes

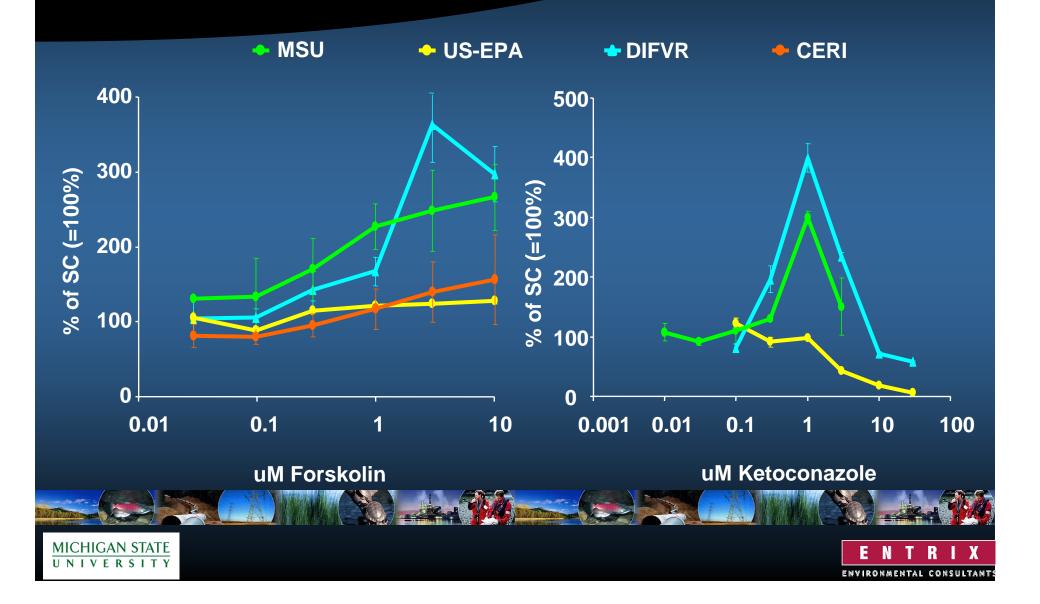




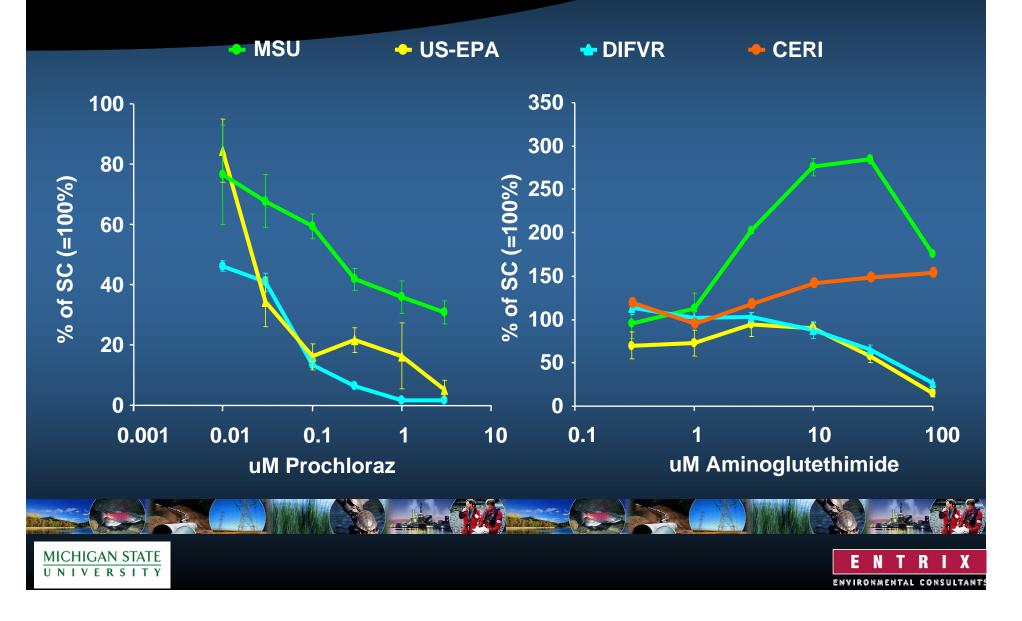
### Phase II - Model Chemicals *Progesterone* (Preliminary Data)



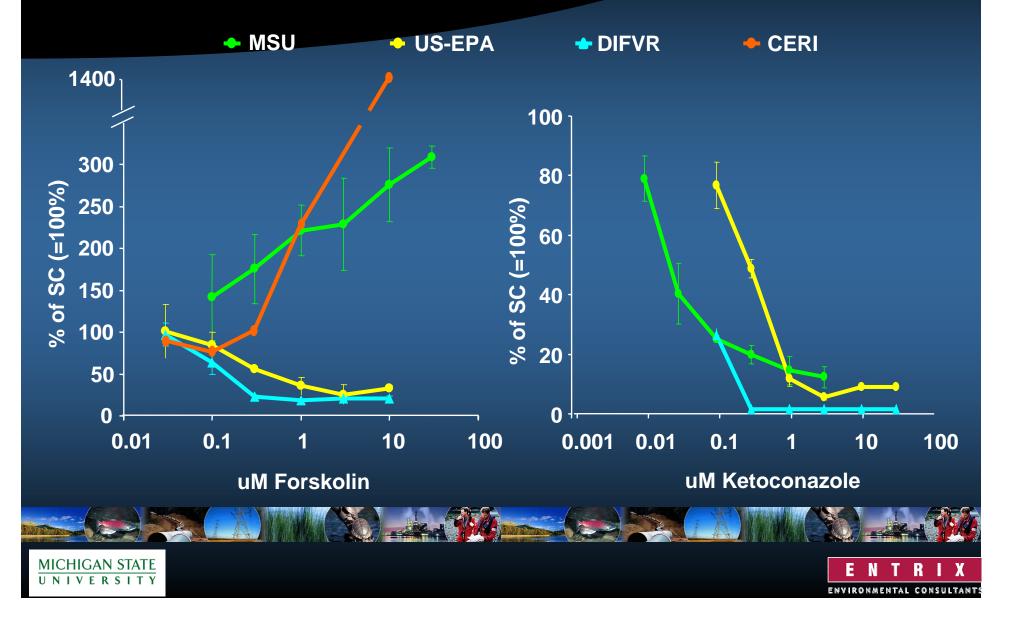
### Phase II - Model Chemicals *Progesterone* (Preliminary Data)



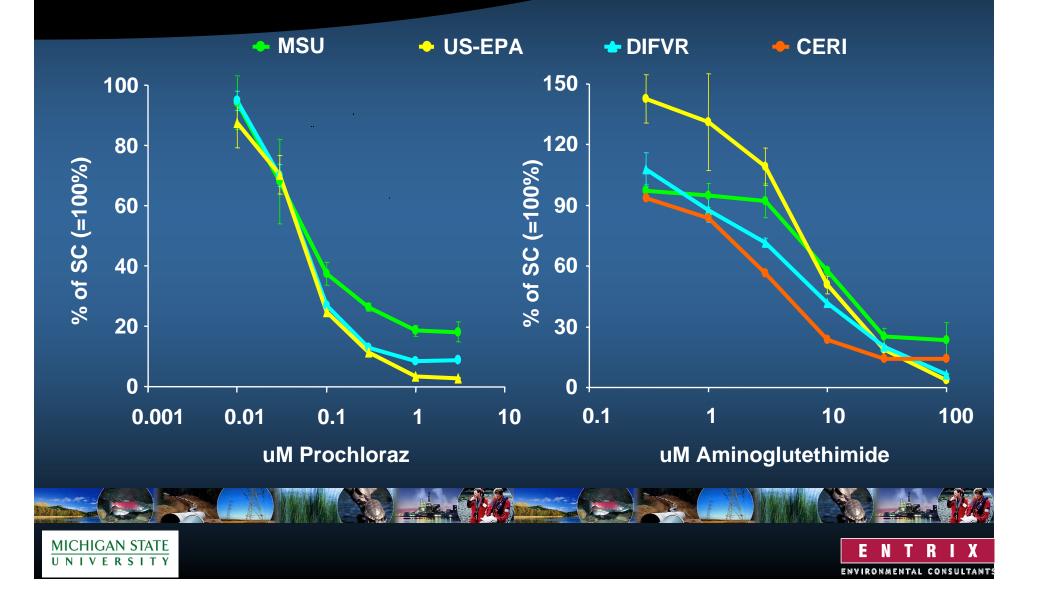
#### Phase II - Model Chemicals Testosterone (Preliminary Data)



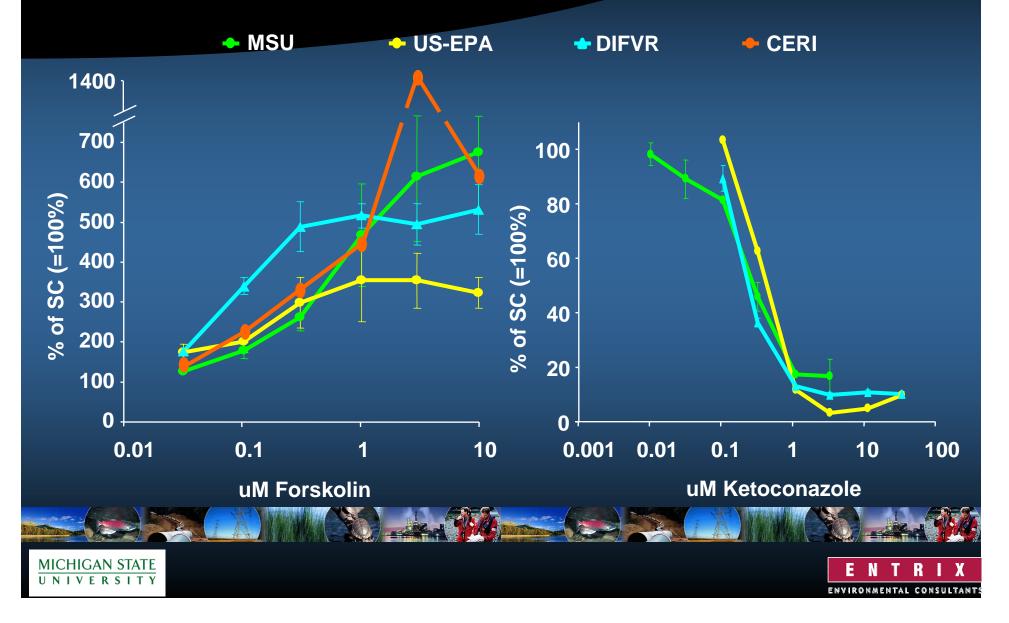
#### Phase II - Model Chemicals Testosterone (Preliminary Data)



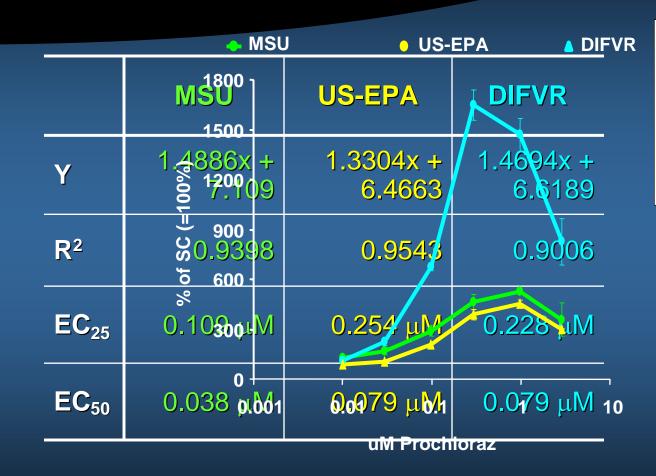
### Preliminary Inter-Lab Comparison Estradiol (Preliminary Data)

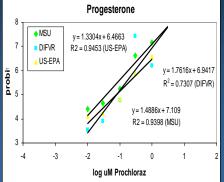


## Phase II - Model Chemicals Estradiol (Preliminary Data)



## Preliminary Inter-Lab Comparison Prochloraz (Progesterone)



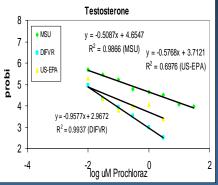






## Preliminary Inter-Lab Comparison Prochloraz (Testosterone)



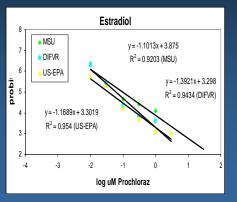






## Preliminary Inter-Lab Comparison Prochloraz (Estradiol)

MSU OUS-EPA OIFVR			
	MSU	US-EPA	DIFVR
Y	-1,101x +	-1.109x + 3.302	-1.007x + 3.936
R <sup>2</sup>	=) 20.9203 50	0.9540	0.9415
<b>EC</b> <sub>25</sub>	<b>≈</b> 0.39 <b>2</b> ⁰µ́М	0.133 μM	0.411-μM
<b>EC</b> <sub>50</sub>	o 0.0950.001	0.035 μM.	







## Phase II - Exposure to Model Chemicals Summary & Conclusions

≍ Good reproducibility of dose-response profiles across laboratories:

E2 production of cells exposed to all model chemicals

P production of cells exposed to all model chemicals with the exception of one lab in the ketoconazole experiment



## Phase II - Exposure to Model Chemicals Summary & Conclusions

Different dose-response profiles for T production of cells exposed to Aminoglutethimide and Forskolin:

Need to evaluate different cell exposure and hormone detection methods

Need to assess effects on other androgens (Androstenedione)



## Conclusions

H295R test system:

Rapid and easy to use

Constitutive basal production of estradiol, testosterone and progesterone

Can measure both increase and decrease of hormone production over several orders of magnitude

Can determine changes in hormone production with high precision and accuracy

**¤**Reproducible





## Conclusions (cont')

#### **¤H295**R test system:

- Flexible can be tailored to identify effects at multiple biological levels in the same system:
  - Gene expression
  - Catalytic enzyme activities
  - Hormone production
- Has the potential to identify multiple mechanisms of action
- Significant reduction of whole animal tests



## Conclusions (cont')

#### ¤H295R test system:

- ✓ Cost effective:
  - ELISA: approx. 200 US\$/sample/hormone<sup>a</sup> + approx. 2 person hrs/sample/hormone<sup>a</sup>
  - Cell culture and exposure: between 0.05 (48 well plate) and 0.15 (6 well plate) person hrs/sample
- Rapid and economic screen of chemicals for their potential to alter Steroidogenesis (priority setting, Tier 1 screening)
- <sup>a</sup> Calculation based on of triplicate measures of 6 different doses + solvent control per sample (chemical)





#### **Conclusion II**

#### **¤** Results can be related to other endpoints

- Pre-screening with certain model compounds resulted in responses that correlated with earlier studies on changes in expression patterns of steroidogenic genes
- Preliminary tests show great promise regarding the transferability of this test system for P and E2
- Need to address variation in responses of T concentrations in media
  - Measurement of alternative androgen endpoints such as androstenedione (currently under-way)





#### **Conclusion II**

- Ankley et al. 2005: Prochloraz suppresses plasma estrogen and androgen concentrations in female and male fathead minnows, respectively
- Vinggaard et al. 2005: Prochloraz suppresses testicular testosterone production and increases testicular progesterone production in rat offspring
- Monteiro et al. 2000: Aminoglutethimide increases androstenedione and decreases estradiol in the flounder; Ketoconazole decreases androgen and estradiol concentrations



Extend hormone analyses to other steroids:
 ✓ Estrone (under-way)
 ✓ Androstenedione (under-way)
 ✓ Cholesterol
 Confirm hypothesized mode of action by measuring actual enzyme activities (e.g., aromatase)



Identify sources for inter-laboratory variability of basal hormone concentrations

Identify causalities for different T patterns observed at different laboratories



 Establish optimized H295R test protocol
 Validation of H295R steroidogenesis test system in extended inter-laboratory trials
 Use of optimized and standardized protocol
 Reduced number of endpoints (2 hormones)
 Larger number of particiating laboratories (10 - 20)



Establish exposure profiles (dose response/ time response) for model compounds with other modes of action

Apply test system to selected priority substances





Validate transferability of test system (currently underway)

Compare to ex vivo and in vivo data

–Xenopus laevis - MSU (*ex vivo*)

-Fathead minnow - US-EPA lab Duluth (*in vivo* and *ex vivo*)

-Minced testis assay

-Uterotrophic assay





#### **Publications**

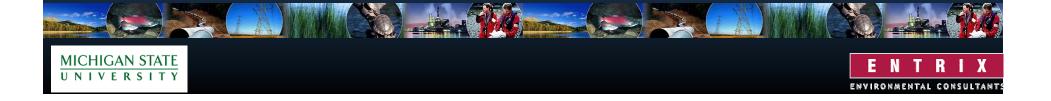
- Hilscherova, K.; Jones, P. D.; Gracia, T.; Newsted, J. L.; Zhang, X.; Sanderson, J. T.; Yu, R. M. K.; Wu, R. S. S.; Giesy, J. P. Assessment of the Effects of Chemicals on the Expression of Ten Steroidogenic Genes in the H295R Cell Line Using Real-Time PCR. *Toxicol. Sci.* 2004, 81, 78-89.
- Zhang, X, Yu, R, Jones, P.D., Newsted, J.L., Gracia, T., Hecker, M., Hilscherova, K., Sanderson, J.T., Wu, R., and Giesy, J.P. 2005. Quantitative RT-PCR Methods for Evaluating Toxicant-Induced Effects on Steroidogenesis Using the H295R Cell Line. *Environ. Sci. Technol.* 39: 2777-2785.
- Hecker, M., Newsted, J.L., Murphy, M.B., Higley, E.B., Tompsett, A.R., Jones, P.D., and Giesy, J.P. 2005. Effects of Prochloraz, Aminoglutethimide, Forskolin and Ketoconazole on steroid hormone production human adrenocarcinoma cells (H295R) cells. *Environ. Sci. Technol.*, submitted for publication



#### **Publications**

Gracia, T., Hilscherova, K., Jones, P.D., Newsted, J.L., Zhang, X., Hecker, M., Higley, E.B., Sanderson, J. T., Yu, R.M.K., Wu, R.S.S. and Giesy, J.P. 2005. Effects of Chemical Mixtures on the Expression of Ten Steroidogenic Genes in the H295R Cell Line. Comp. Physiol. Biochem. B, submitted for publication.

Blaha, L., Hilscherova, K., Mazurova, E., Hecker, M., Jones, P.D., Bradley, P., Gracia, T.R., Duris, Z., Holoubek, I. and Giesy J.P. Alteration of steroidogenesis in H295R cells by organic sediment contaminants and relationships to other endocrine disrupting effects. In prep.



#### Thank You!

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#### H295R cell line

Derived from the NCI-H295 pluripotent adrenocortical carcinoma cell line (Gazdar, et al. 1990) from a carcinoma of the adrenal cortex that arose in a 48 y.o. black female.

Modified cells retain the ability to produce aldosterone, cortisol and C19 steroids (adrenal androgens).



# ×Validate transferability of test system (currently underway)

- ✓ Within the same laboratory (completed)
- Between different laboratories (underway)



## Model Chemical Exposure Vinclozolin

