

## OSHA work-practice guidelines for personnel dealing with cytotoxic (antineoplastic) drugs

**Abstract:** Work-practice guidelines for personnel dealing with cytotoxic drugs (CDs) are presented.

Current practices in the preparation, storage, administration, and disposal of CDs may expose pharmacists, nurses, physicians, and other health-care workers to high environmental levels of these drugs. OSHA has developed these guidelines to protect health-care workers from unnecessary exposure to CDs.

A brief summary of the short-term and long-term hazards known to be associated with these drugs is presented. The risks to workers handling CDs are a com-

bined result of the drugs' inherent toxicity and the extent to which workers are directly exposed to CDs via inhalation, absorption, and ingestion.

Work-practice guidelines that can limit the exposure of workers to CDs and the equipment necessary to carry out these practices properly are described.

**Index terms:** Antineoplastic agents; Equipment; Guidelines; Health care; Occupational Safety and Health Administration; Personnel; Safety; Toxicity, environmental

*Am J Hosp Pharm.* 1986; 43:1193-1204

### Preface

It is not uncommon for health-care professionals to regard themselves as immune from any harm arising from their work. Thus, during the course of treating their patients, they may inadvertently expose themselves and their staff to hazardous substances while taking every precaution to ensure that the administered drugs are protected from contamination. The guidelines that follow have been developed in response to requests for assistance from a number of disparate sources, including health-care personnel concerned with the potential harm that may result from exposure to cytotoxic drugs (CDs).

The guidelines (not to be confused with mandatory standards) are designed to assist all health-care personnel, including physicians, nurses, pharmacists, aides, and the numerous and diverse health-care support staff who may be exposed to cytotoxic drugs through inhalation, skin absorption, or trauma. We are aware that the drugs are prepared and administered in a wide variety of

places, ranging from physicians' private consulting rooms to large pharmaceutical preparation rooms. We cannot cover every situation, but we offer what we believe is the most reasonable course of action, whether dealing with the drugs as a routine health-care procedure or during an emergency situation such as a large spill. Because of the widespread use of these drugs and the general lack of written policies or standard operating procedures in most facilities, we hope this instruction will fulfill the need to provide a description of the potential hazards of CDs, as well as the proper safety procedures, personal protective equipment, and engineering controls for CDs that will reduce contamination of workers.

The reader must be reminded that persons involved in health care, whether professionals or nonprofessionals, are workers and that the hospital, clinic, pharmacy, or even consulting room is a workplace. Some of the methods recommended may seem to be more suited to a nonclinical workplace, in particular, the suggested use of goggles

---

Issued January 29, 1986, by the Office of Occupational Medicine, Directorate of Technical Support, Occupational Safety and Health Administration, U.S. Department of Labor.

Developed by Ralph E. Yodaiken, M.D., M.P.H., Director, and Dianne Bennett, M.D., M.P.H., Medical Officer, Office of Occupational Medicine, Occupational Safety and Health Adminis-

---

tration, Room N 3651, 200 Constitution Avenue, N.W., Washington, DC 20210. For further information about these guidelines, contact Dr. Yodaiken.

Editor's note: See related news report in this issue on page 1116.

where appropriate and a respirator where a biological safety cabinet is not available. Such recommendations must be considered in the light in which they are proffered; health professionals must walk the narrow line between alarming their patients unnecessarily and protecting their own health. We are aware of incidents in which nurses, ridding syringes of air, inadvertently sprayed their eyes with a drug aerosol. If health professionals are ROUTINELY careful during such procedures, goggles will not be essential. Similarly, if a biological safety cabinet is not in place, we know of no way of avoiding inhalation of a carcinogen other than by using an appropriate respirator.

Two elements are essential to ensure proper workplace practices:

- Education and training of all staff involved in handling any aspect of CDs.
- A biological safety cabinet (BSC).

The costs of implementing the former and installing the latter are relatively minor. The potential benefits are major.

Finally, we are grateful to our reviewers, including those who reviewed the document informally. We have incorporated as many of their recommendations and corrections as possible, and we are aware of the need to revise and update this document from time to time.

## I. Introduction

A. Current practices in the preparation, storage, administration, and disposal of the widely used group of antineoplastic (anti-new-growth; anticancer) drugs—also called cytotoxic drugs (CDs) because they are toxic to cells—may expose pharmacists, nurses, physicians, and other health-care workers to high environmental levels of these drugs.

1. Although little research has been done on the long-term risks at the levels of exposure encountered by unprotected health-care workers, these drugs have been associated with human cancers at high (therapeutic) levels of exposure<sup>1-9</sup> and are carcinogens and teratogens in many animal species.<sup>4,9</sup>
2. Under current work practices, CDs have demonstrated the ability to cause elevations in sister chromatid exchanges and chromosome breakage in circulating lymphocytes and mutagenic activity in urine.
3. In addition, many of these drugs have been shown to cause a variety of acute effects in humans, such as localized skin necrosis (death of tissue) after surface contact with abraded skin<sup>10</sup> or damage to normal skin.<sup>11</sup>

B. Several sets of work-practice guidelines have been issued by various professional bodies in the United States and by foreign institutions.<sup>12-25</sup> The volume of requests to OSHA indicates a broader interest among administrators and health-care professionals who are not aware of, or who have not had access to, these guidelines. Moreover, recent surveys reveal that there is little standardization of work practices and that proper practices and adequate protective equipment are not currently being used. Therefore, OSHA considers implementation of its work-practice guidelines important for protecting workers against these serious occupational hazards. The following information contained in sections II, III, and IV

1. Provides a brief summary of the short-term and long-term hazards now known to be associated with the drugs;
  2. Lists work-practice guidelines that can limit the exposure of workers to CDs and the equipment necessary to carry out these practices properly; and
  3. Lists the CDs currently in use (Table 1).
- C. These guidelines are addressed to persons who have a broad spectrum of qualifications and experience, and some readers may find them repetitious and too detailed. However, even the most qualified professionals do not always observe elementary and essential workplace safety practices; therefore, we have taken the precaution of covering as much detail as is considered essential to good work practice. Any repetition is an attempt to make each section complete.

## II. Cytotoxic Drugs and Sites of Potential Risks

### A. Cancer Chemotherapy.

1. The attempt to stop or reverse the growth of malignant growing cells with drugs began in the late 1940s, when mechlorethamine hydrochloride and its derivatives were first used therapeutically. Currently, approximately 30 cancer CDs are available commercially in the United States; these are administered to an estimated 200,000–400,000 patients annually (Table 1). Consequently, several thousand health-care employees may be exposed yearly to a variety of risks.
2. From the time of their initial use, these drugs were known to be potentially harmful to workers dealing with them. The mechlorethamine drugs are extremely irritating to mucous mem-

**Table 1.**  
**Antineoplastic Agents Currently in Use<sup>a</sup>**

Type of Agent	Trade Name
<i>Alkylating Agents</i>	
Cisplatin	Platinol
Cyclophosphamide	Cytoxan
Mechlorethamine hydrochloride <sup>b</sup>	Mustargen
Thiotepa	
Carmustine	BiCNU
Streptozocin	Zanosar
Busulfan	Myleran
Chlorambucil	Leukeran
Lomustine	CeeNu
Melphalan	Alkeran
Teosulfan	
Uracil mustard	
Chlornaphazine	
Dacarbazine	DTIC-Dome
<i>Antimetabolites</i>	
Cytarabine	Cytosar-U
Fluorouracil	Adrucil
Methotrexate	Mexate
Mercaptopurine	Purinethol
Azathioprine	Imuran
Procarbazine	Matulane
<i>Antibiotics</i>	
Doxorubicin hydrochloride	Adriamycin
Bleomycin sulfate	Blenoxane
Dactinomycin	Cosmegen
Daunorubicin hydrochloride	Cerubidine
Mithramycin	Mithracin
Mitomycin	Mutamycin
<i>Mitotic Inhibitors (Vinca alkaloids)</i>	
Vincristine sulfate	Oncovin
Vinblastine sulfate	Velban
Etoposide	
<i>Miscellaneous</i>	
Asparaginase	Elspar
<i>Investigational Drugs</i>	
Azacitidine	
Amsacrine	
Teniposide	
Ifosfamide	
Mitoxantrone hydrochloride	
Vindesine	

<sup>a</sup> This is not a complete list; it requires periodic updating.

<sup>b</sup> Referred to in these guidelines as nitrogen mustard.

branes, eyes, and skin.<sup>26</sup> Other agents developed later on, such as fluorouracil, also have well-known topical effects.<sup>11</sup> Spills of agents such as doxorubicin onto abraded skin can lead to severe soft-tissue injury, such as necrosis and sloughing of exposed areas,<sup>10</sup> as well as possible effects on the fetus.<sup>27</sup> Symptoms such as lightheadedness, dizziness, nausea, headache, and possible allergic reaction also have been described in nurses after the preparation of antineoplastic drugs and their subsequent administration in unventilated areas.<sup>28,29</sup>

3. The potential for harmful effects developing over a longer term is also well known. Most CDs either bind directly to

genetic material, in the cell nucleus, or affect cellular protein synthesis and may therefore damage growth and reproduction of normal cells as well.

## B. Various Studies.

1. One study, on chlorambucil, shows that chromosome damage to cells among people to whom the drug is administered is related to both dose and duration of therapy and is cumulative.<sup>30</sup> Evidence that these drugs induce malignant tumors in animals and neoplasms and leukemias in humans to whom they are administered was available as early as the 1940s and 1950s, and continued to mount in the succeeding decades.<sup>6-9</sup>
2. In vivo, in vitro, and human studies have implicated antineoplastic drugs in chromosomal damage and teratogenesis (malformation), as well as carcinogenesis (cancer induction).<sup>8,27,30-32</sup> Testicular and ovarian dysfunction including permanent sterility have been demonstrated in male and female patients, respectively, who have received CDs either singly or in combination. Congenital malformations have been attributed to fluorouracil.<sup>27</sup> A study from Finland also indicates an association between CDs and fetal malformations in pregnant nurses who work with CDs.<sup>31</sup> While this needs to be confirmed, fetal loss associated with cytotoxic drugs has been reported among nurses in Finnish hospitals.<sup>33</sup>
3. Finally, organ damage also has been associated with CDs, not only in animals<sup>34,35</sup> and human patients receiving long-term therapy<sup>34</sup> but also among employees. Liver damage has been reported in nurses working on an oncology ward; the damage appeared to be related to the intensity and duration of work exposure.<sup>36</sup>
4. Various studies have shown that current practices expose workers to substantial amounts of these drugs and that these drugs may increase the risk of long-term harm. Detectable amounts of various drugs have been found in the urine of health-care workers handling them.<sup>37</sup> Attempts also have been made to show that mutagenic activity in urine and chromosome damage are increased in workers handling CDs without proper protection and safe workplace practices.
  - a. Pharmacists who reconstituted anticancer drugs showed increasingly mutagenic urine over the period of exposure; when

they stopped handling the drugs, mutagenic activity fell within two days to the level of unexposed controls.<sup>38</sup> Also, the installation of a vertical-flow containment hood, or biological safety cabinet (BSC), has been shown to reduce the levels of mutagenic substances in the urine of pharmacy workers preparing CDs.<sup>39</sup>

- b. A variety of chemical or physical agents have been associated with mutagenesis. Hair dyes and cigarette smoke are two that may concern health-care personnel. Smokers exposed to CDs exhibit greater urine mutagenicity than smokers who did not handle CDs.<sup>40</sup> Smokers who do not take simple protective measures such as gloving and hand washing may take in additional amounts of the drug orally through contaminated cigarettes. Other studies have shown a definite and significant reduction of urine mutagenicity in both smokers and nonsmokers who work with the agents when their work practices have improved.<sup>41</sup>
- c. Though the levels of absorption that may have taken place during work are hard to assess, it is essential to minimize exposure to these potent carcinogens and teratogens.

### III. Current Practices in Preparation, Use, and Disposal: Points of Exposure for Personnel

*Note:* The risks to workers handling CDs are a combined result of the drugs' inherent toxicity and the extent to which workers are directly exposed to CDs on the job. The main routes of exposure are through the *inhalation* of drug dusts or droplets, *absorption* through the skin, and *ingestion* through contact with contaminated food or cigarettes. Opportunity for exposure may occur at many points in the handling of these drugs.

#### A. Survey of Current Work Practices.

A 1982 survey of 21 cancer centers in the United States revealed little standardization of work practices, equipment, or training for personnel administering injectable CDs.<sup>42</sup> In only 10 centers were the necessary BSCs used for preparation. A 1983 survey of 10 hospital oncology clinics<sup>43</sup> showed that 9 did not use BSCs for preparation and that use of gloves was routine in only 3 clinics. Eating and drinking occurred in seven of the preparation rooms, greatly increasing the probability of oral intake of the drugs. Wastes were disposed of in covered receptacles in only seven of the preparation rooms. Air monitoring showed substantial air levels of the two drugs in most common use in these clinics. Based on these studies, the need for the

adoption of guidelines, the provision of proper protection and equipment, and the use of appropriate training is crucial.

#### B. Pharmacy or Other Preparation Areas.

1. In large oncology centers, CDs are usually prepared in the pharmacy by pharmacy personnel, but in many hospitals and smaller centers they may be prepared by physicians or nurses in patient-care or staff areas that are often inadequately ventilated.<sup>28</sup> Many CDs must be dissolved, transferred from one container to another, or manipulated before they can be administered to patients. Even if care is taken, opportunity for absorption through inhalation or direct skin contact may occur.<sup>15,43</sup> Examples of manipulations that can cause splattering, spraying, and aerosol generation include:
  - a. Withdrawing needles from drug vials,
  - b. Transferring drug with syringes and needles or filter straws,
  - c. Breaking open ampuls, and
  - d. Expelling air from a drug-filled syringe.
2. Aerosols can be generated by these activities, exposing not only the employee immediately involved but also staff and patients in the surrounding areas.<sup>25</sup> A properly enclosed and ventilated work area, respiratory and skin protection, and training in the proper handling of these drugs is essential.<sup>20,25,28,43,44</sup> Smoking, drinking, applying cosmetics, and eating where these drugs are prepared, stored, or used should *never* take place because these practices greatly increase the chance of exposure.<sup>25</sup>
3. Even in the pharmacy, where protective clothing and gloves are worn and careful aseptic techniques are used as a matter of course, opportunities for exposure can occur. A horizontal-airflow, clean workbench is often used to provide an aseptic environment for the preparation of injectable drugs. Because this unit provides a flow of filtered air originating at the back of the work space and exiting toward the employee using the unit, it provides protection for the drugs but increases the likelihood of exposure to the preparer of the drugs and the other personnel who may be in the room. The preparer and others are exposed to the aerosols generated during preparation procedures.
  - a. Class II vertical-flow containment hoods, also called biological safety cabinets (BSCs), provide appropriate protection.<sup>44,45</sup>

- b. Type A BSCs are the minimal requirement. Type A hoods that are vented (some of these are now classified as type B3) are preferred.<sup>18</sup>

### C. Administration of Drugs to Patients.

1. The administration of drugs to patients is generally carried out by nurses or physicians. Injecting the drug into the i.v. line, clearing air from the syringe or infusion line, and leakage at the tubing, syringe, or stopcock connection present opportunities for both skin contact and aerosol generation leading to respiratory exposure. Clipping used needles and crushing used syringes, standard practice in some work situations, may produce a considerable aerosol.<sup>43</sup>
2. Excreta from patients who have received certain antineoplastic drugs may contain high concentrations of the drug or hazardous metabolites. For example, patients receiving cyclophosphamide excrete large amounts of the drug and mutagenic metabolites.<sup>46,47</sup> Patients treated with cisplatin excrete potentially hazardous amounts of the drug.<sup>48</sup> Handling urine or urine-soaked sheets may lead to dangerous exposure. Nursing and housekeeping personnel may be exposed to CDs if they are not made aware of the potential hazard and not trained to take precautions.

### D. Disposal of Drugs and Contaminated Materials.

1. Materials that have been used in the preparation and administration of CDs, such as gloves, gowns, syringes, or vials, present a possible source of exposure or injury to support and housekeeping staff, as well as other health-care workers not involved with their preparation and administration. The use of properly labeled, sealed, and covered containers, handled only by trained and protected personnel, should be routine. Spills also represent a hazard, and all employees should be familiar with appropriate spill procedures for their own protection.

## IV. Guidelines for the Handling of Cytotoxic Drugs

### A. Drug Preparation.

#### 1. Personal Protective Equipment.

- a. New research indicates that surgical latex gloves are less permeable to many CDs than the polyvinyl chloride gloves recommended in older guidelines.<sup>18,49,50</sup> Surgical latex gloves therefore should be used for the preparation of CDs unless the

manufacturer specifically stipulates that some other glove provides better protection. (Powdered gloves should *never* be used.) A double layer of gloves is substantially less permeable and should be used if double-gloving does not interfere with technique. Because all gloves are permeable to some extent and their permeability increases with time,<sup>49</sup> they should be changed regularly (hourly is preferable) or immediately if they are torn or punctured.

- b. A protective disposable gown made of lint-free, low-permeability fabric with a closed front, long sleeves, and elastic or knit-closed cuffs must be worn, with the cuffs tucked under the gloves. Gowns and gloves in use should not be worn outside the preparation area.
  - c. A BSC is essential for the preparation of CDs, but where one is not currently available, a respirator with a high-efficiency filter, preferably a powered air-purifying respirator used by personnel who have been trained to use respirators, provides the best protection until a BSC is installed. We realize that this is a departure from the usual hospital/clinic/pharmacy procedures, but in this case we are dealing with a variety of known carcinogens and therefore appropriate preventive measures are necessary.
  - d. Surgical masks do *not* protect against the breathing of aerosols. A plastic face shield or splash goggles complying with American National Standards Institute 28.7.1-1968 criteria also should be worn if a BSC is not in use and an eyewash fountain made available. (See the Preface.)
2. *Preparation Area.* It is suggested that all CDs be prepared in one central area. If this is not practical, the number of areas used for preparation should be minimized. If possible, an isolated BSC where only CDs are prepared should be designated. Warning signs designating the area as a CD-preparation area, which should not be entered by unauthorized staff, should be clearly posted. Procedures for handling spills should also be posted. Eating, drinking, smoking, chewing gum, applying cosmetics, and storing food in or near the preparation area should be forbidden.
    - a. A Class II BSC that meets current National Sanitation Foundation Standard 49<sup>51</sup> must be used.<sup>17-19,44,52</sup> The blower on the vertical-airflow hood should be on at all times, 24 hours a day, seven days a week. Venting to the outside is preferable where feasible and is required with a Type B BSC. If the hood has an outside exhaust system, filtered exhaust to the outside should be at an appropriate level and away from air-

intake units. BSCs should be certified by a qualified technician every six months or any time the cabinet is moved.

- b. Technicians servicing these cabinets or changing high-efficiency particulate air (HEPA) filters should be warned of the nature of CDs and should use the same personal protective equipment as an employee dealing with a large spill (see A.I.). Special containment procedures that should be used to avoid contamination, both of the service personnel and the room, have been described in detail previously.<sup>44</sup>
  - c. All used gowns and gloves and disposable materials used in preparation should be disposed of according to the hospital's toxic-waste procedures and as described in the section on waste disposal (see IV, D.).
3. *Preparation Equipment.* Work with CDs must be carried out in a BSC on a disposable, plastic-backed paper liner, which should be changed after preparation is completed for the day, or after a shift, whichever comes first. Syringes and i.v. sets with Luer-Lok fittings should always be used, and syringes should always be large enough so that they need never be more than three-fourths full. A nonsplash disposal-collection vessel, such as a plastic or metal tray lined with sterile gauze pads, should be on hand to collect excess solution. All necessary items should be placed within the BSC before work is begun, and all extraneous items should be kept out of the work area in order to avoid contamination.
- a. The work areas should be provided with a closable, puncture-resistant, shatter-proof container for disposal of contaminated sharp and breakable materials. Labeled sealable plastic or wire-tie bags, as described in the section on waste disposal, should be on hand so that all boxes and other contaminated materials, including gloves, gowns, and paper liners, can be immediately placed in them and disposed of according to the hospital's toxic-waste procedures.
  - b. The cabinet should be cleaned daily with 70% alcohol and decontaminated weekly, whenever spills occur, or when the cabinet requires service or certification. Ordinary decontamination procedures, which include fumigation with a germicidal agent, are inappropriate in a BSC used for CDs because such procedures do not deactivate the drugs and in addition may cause chemical reactions.<sup>52</sup> Decontamination should consist of surface cleaning with high pH agents followed by thorough rinsing. Removable work trays, if present, should be removed, and the back of the

work tray and the sump below should be included in the cleaning.

4. *Work Practices in Preparation.* Proper aseptic techniques are essential for worker protection, but because it is generally accepted that these techniques are essential for patient safety, it is assumed they will already be standard practice in drug preparation. Therefore, general principles of aseptic technique will not be described here. It should be noted, however, that BSC benches differ from horizontal-flow units in several ways, thus requiring special precautions: Manipulations should not be performed close to the work surface, and unsterilized items, including liners and hands, must be kept downstream from the working area. Operators should be trained in these techniques.<sup>52</sup>
- a. *Syringes and I.V. Bottles.* These should be labeled with the patient's name and room number, drug name, and quantity per total volume, route of administration, date and time prepared, dose, expiration date, and storage requirements if the drug is not to be transported immediately. All syringes, i.v. bags, and bottles containing CDs should be labeled with a distinctive warning label such as "Chemotherapy—handle with gloves—dispose of properly."
  - b. *Needles.* The use of large-bore needles, #18 or #20, will ensure that high-pressure syringing of the solutions is avoided. However, some experienced personnel believe that large-bore needles are more likely to drip. The needle should be chosen with these advantages or disadvantages in mind.
    1. Drug administration sets should be attached and primed within the hood, before the drug is added to the fluid, to obviate the need to prime the set in a less well controlled environment and to ensure that any fluid that escapes during priming contains no drug.
    2. All syringes and needles used in the course of preparation should be placed in the puncture-proof container for disposal without being crushed, clipped, or capped. (Some professionals believe that capping the needle before disposal reduces the generation of aerosols; others warn that it increases the chances of needle sticks.)
  - c. *Handling Vials.* Medication vials should not be vented unless a BSC is used as the work area or unless a hydrophobic filter-needle unit is available to eliminate pressure.<sup>53</sup> Syringe and needle fittings should

be of the Luer-Lok variety.

1. Diluent should be added slowly to the vial by alternately injecting small amounts and allowing displaced air to escape into the syringe. (All the diluent should not be injected at once because a large volume of displaced air can cause the syringe's plunger to back up and possibly spray the drug or cause leakage around the needles.) When all diluent has been added, a small amount of additional air may be withdrawn to create a negative pressure in the vial, but this should *not* be expelled into room air because it may contain drug residue. It should either be injected into a vacuum vial or remain in the syringe to be discarded.
  2. A sterile gauze pad should be wrapped around the needle and vial top when solution is withdrawn (employees should take care to avoid needle sticks during this procedure). The drug should be withdrawn from the vial while negative pressure is maintained. (The technique has been described previously.<sup>34</sup>) If this use of negative pressure is considered impossible, a syringe should be filled with air equal to the volume of drug required, and the solution should be withdrawn by alternately injecting small amounts of air into the vial and withdrawing equal amounts of liquid until the required volume is withdrawn. The drug should be cleared from the needle and hub (neck) of the syringe before separation to avoid spraying on separation.
- d. **Handling Ampuls.** Any material remaining in the top of an ampul should be tapped down before the ampul is opened. A sterile gauze pad should be wrapped around the ampul neck before the top is broken to protect against cuts and to catch aerosolized material.
1. The ampul top should not be removed close to the employee's face. If diluent is to be added, it should be injected slowly down the inside wall of the ampul. The ampul should be tilted gently to ensure that all the powder is wet before it is agitated to dissolve the contents.
  2. The needle should be held vertically with the needle upward; the syringe should be tapped to remove air bubbles, and the air bubbles should be expelled into sterile gauze, not into the air.

## B. Drug Administration.

### 1. *Personal Protective Equipment.* Consider-

ation should be given by personnel administering CDs to wearing the following items:

- a. A gown, as described in the section on drug preparation.
- b. Disposable surgical latex gloves, double pairs if appropriate.
- c. A surgical mask also may be used, but it should be noted that this provides only minimal protection against CD aerosols and is no substitute for the proper procedures, which have been described.

To avoid alarm or misunderstanding, patients should be informed that any protective equipment in use is necessary for the protection of workers against the directly irritating effects of the drugs to eyes and skin (long-term as well as short-term effects of the drug already should have been discussed with patients in terms of potential risks to themselves).

### 2. *CD-Administration Equipment.* Protective equipment and other necessary items may be packaged together and labeled as a CD-administration kit, which should include:

- a. Personal protective equipment as described in IV, A.1;
- b. Gauze (4" X 4") for cleanup;
- c. Alcohol wipes;
- d. Disposable plastic-backed absorbent liner;
- e. Empty vials to be used as receptacles for excess drug solution;
- f. A puncture-proof container for needles and syringes;
- g. A 4-mil sealable plastic or wire-tie bag (with warning label) large enough to contain waste materials, and accessory warning labels;
- i. If additional preparation that cannot be done in a BSC is required before administration, a respirator (see IV, A.1.) used perhaps in a specially assigned side room, splash-proof goggles, and a 32-oz bottle of sterile isotonic eye and face wash should be readily available for emergencies.

### 3. *Work Practices.* Personnel should follow safe work practices including the following:

- a. Hands should be washed before gloves are put on. Gowns or gloves that become contaminated should be changed immediately.
- b. Infusion sets and pumps, which should have Luer-Lok fittings, should be watched for signs of leakage during use. A plastic-backed absorbent pad should be placed under the tubing during administration to catch any leakage. The line should be bled into gauze inside a sealable plastic bag.

- c. Priming should be carried out under a BSC. However, if i.v. sets are primed or air is expelled from syringes at the bedside, gauze in a plastic bag should be used as a receptacle. Syringes, i.v. bottles and bags, and pumps should be wiped clean of any drug contamination with an alcohol wipe. Needles and syringes should not be crushed or clipped but should be placed in a puncture-resistant container to go into the CD-disposal bag, along with all other contaminated materials. The bag should be disposed of in accordance with the hospital's toxic-waste-disposal procedures.
- d. Protective goggles should be wiped several times with an alcohol wipe and properly rinsed. Hands should be washed after removal of gloves. All gauze and alcohol wipes must be put in an appropriate container for disposal.

*Note:* Currently, a large number of investigational CDs are under clinical study in health-care facilities. Personnel not directly involved in the investigation should not administer these drugs unless they have received adequate instructions regarding safe handling procedures.

#### C. Caring for Patients Receiving CDs.

1. *Personal Protective Equipment.* Personnel dealing with blood, vomitus, or excreta from patients who have received CDs in the last 48 hours should wear surgical latex gloves and disposable gowns, to be discarded after each use as described in the section on waste disposal. (No protective equipment is necessary for ordinary patient contact for employees not dealing with drug administration or bodily secretions.) Hands should be washed after removal of gloves or after contact with the above substances.
2. *Linen.* Linen contaminated with CDs, blood, vomitus, or excreta from a patient who has received CDs up to 48 hours before should be placed in a specially marked laundry bag, and the laundry bag should be placed in a labeled impervious bag. This laundry bag and its contents should be prewashed, and then the linens should be added to other laundry for an additional wash. Laundry personnel should wear surgical latex gloves and gowns while handling this material. (No additional gain is made by autoclaving items contaminated with CDs, unless they are also contaminated with infectious waste.)

#### D. Waste Disposal.

1. *Equipment.* Cytotoxic-waste-disposal sealable plastic or wire-tie bags of 4-mil-

thick polyethylene or 2-mil-thick polypropylene, labeled with a cytotoxic hazard label and colored differently from other hospital trash bags, should be used for the routine accumulation and collection of used containers, syringes, discarded gloves, gowns, goggles, and any other disposable material. All CD-related wastes should be put into these bags and not into any other container.

- a. Needles, syringes, and breakable items should be placed in a plastic vial or puncture-proof box before they are placed into the bag. Needles should not be clipped or capped, and syringes should not be crushed. The bag should be kept inside a covered waste container clearly labeled "cytotoxic waste only."
  - b. At least one such receptacle should be located in every area where the drugs are prepared or administered so that the waste need not be moved from one area to another. The bag should be sealed when it is filled, and the carton should be taped.
2. *Handling.* Housekeeping personnel must wear gowns and surgical latex gloves when handling the waste containers, and they should be instructed on the necessity of handling this waste with care and on procedures governing spills and leaks.
  3. *Disposal.* These wastes must be handled separately from other hospital trash and must be regarded as toxic (hazardous) wastes and disposed of in accordance with applicable regulations.<sup>55</sup>
    - a. Disposal in a licensed sanitary landfill for toxic wastes is an acceptable alternative. If waste is to be picked up by a commercial disposal firm, the company must be licensed, and the waste must be held in a secure area in covered, labeled drums lined with 6.5-mil polyethylene liners.
    - b. Chemical inactivation of CDs is often ineffective and may produce by-products that are more mutagenic than the parent drug.<sup>18</sup> Therefore, with the exception of nitrogen mustard, which can be safely inactivated by sodium thiosulfate, chemical inactivation should be avoided until safe chemical procedures are developed.

#### E. Spills.

1. *General Procedure.* Spills and breakages should be cleaned up immediately by a properly protected person trained in the appropriate procedures. Broken glass should be carefully removed. A spill should be identified with a warning sign so that other persons in the area will not be contaminated.
2. *Personnel Contamination.* Overt contami-



nation of gloves or gowns, or direct skin or eye contact should be treated as follows:

- a. Immediately remove the gloves or gown;
  - b. Wash the affected skin area immediately with soap (not germicidal cleaner) and water. For eye exposure, immediately flood the affected eye with water or isotonic eyewash designated for that purpose for at least five minutes;
  - c. Obtain medical attention immediately.
3. **Cleanup of Small Spills.** Spills of less than 5 mL or 5 g outside a hood should be cleaned immediately by personnel wearing gowns and double surgical latex gloves and eye protection.
- a. Liquids should be wiped with absorbent gauze pads; solids should be wiped with wet absorbent gauze. The spill areas then should be cleaned (three times) using a detergent solution followed by clean water.
  - b. Any glass fragments should be placed in a small cardboard or plastic container and then into a CD-disposal bag, along with the used absorbent pads and any non-cleanable contaminated items.
  - c. Glassware or other contaminated reusable items should be placed in a plastic bag and washed in a sink with detergent by a trained employee wearing double surgical latex gloves.
4. **Cleanup of Large Spills.** For spills of amounts larger than 5 mL or 5 g, spread should be limited by gently covering with absorbent sheets or spill-control pads or pillows or, if a powder is involved, with damp cloths or towels. Be sure not to generate aerosols. Access to the spill areas should be restricted.
- a. Protective apparel should be used (see E.3.), with the addition of a respirator when there is any danger of airborne powder or an aerosol being generated. The dispersal of CD particles into surrounding air and the possibility of inhalation is a serious matter and should be treated as such.
  - b. Chemical inactivators, with the exception of sodium thiosulfate, which can be used safely to inactivate nitrogen mustard, may produce hazardous by-products<sup>18</sup> and should not be applied to the absorbed drug.
  - c. All contaminated surfaces should be thoroughly cleaned with detergent solution and then wiped with clean water. All contaminated absorbents and other materials should be disposed of in the CD-disposal bag.
5. **Spills in Hoods.** Decontamination of all interior hood surfaces may be required

after the *above procedures have been followed*. If the HEPA filter of a hood is contaminated, the unit must be labeled "Do not use—contaminated," and the filter must be changed and disposed of properly as soon as possible by trained personnel wearing protective equipment. Protective goggles should be cleaned with an alcohol wipe after the cleanup.

6. **Spill Kits.** Spill kits, clearly labeled, should be kept in or near preparation and administrative areas. It is suggested that kits include a respirator, chemical splash goggles, two pairs of gloves, two sheets (12" X 12") of absorbent material, 250-mL and 1-L spill-control pillows, and a small scoop to collect glass fragments. Absorbents should be incinerable. Finally, the kit should contain two large CD-waste-disposal bags.

#### F. Medical Surveillance.

1. **Routine Procedures.**
  - a. All employees with potential exposure to CDs through preparation, administration, housekeeping, waste disposal, transport, or storage of CDs, in addition to being fully informed of all potential dangers and the need to take adequate precautions, should have a preplacement physical examination. Care should be taken to note any risk factors in the history, and a complete blood count including differential should be taken to provide a baseline.
  - b. Currently, no techniques exist for screening individual employees that would indicate the level of exposure reliably, though group screening for urine mutagenesis or for the presence of certain CDs in urine may be recommended by medical staff.
  - c. A registry of all staff who routinely prepare or administer CDs should be permanently maintained, with the number recorded of each drug the employee has prepared or administered, if this is feasible.
2. **Acute Exposures.** After an acute exposure, the treatment procedure described in section E on spills should be followed, and the employee should receive a physical examination with particular attention to the eyes, buccal and nasal mucous membranes, and skin. Acute exposures include needle sticks from needles attached to syringes containing the drugs. However, needle sticks received by laboratory personnel dealing with the blood of patients being treated with CDs do not constitute a special hazard and require only ordinary needle-stick pro-

cedures. Needle sticks, as with all other acute exposures, should be recorded both on incident forms and in the employee's medical record.

3. **Pregnancy.** On the basis of the available evidence, it seems reasonable to assume that if appropriate procedures are followed and proper equipment and protection are provided, reproductive hazards will be reduced.
  - a. Employees should be fully informed of the potential reproductive hazards and, if they so request, staff members who are pregnant or breast-feeding should be transferred to comparable duties that do not involve handling CDs.
  - b. A similar policy covering male or female personnel who are actively trying to conceive a child should be established.

#### G. Storage and Transport.

1. **Storage Areas.** Access to areas where CDs are stored should be limited to authorized personnel. Such areas should be posted with a large warning sign, a list of all drugs covered by CD policies, and a sign detailing spill procedures. Facilities used for storing CDs, if possible, should not be used for other drugs and should be designed to prevent containers from falling to the floor. Warning labels should be applied to all CD containers, as well as the shelves and bins where these containers are permanently stored.
2. **Receiving Damaged CD Packages.** Damaged cartons should be opened in an isolated area by an employee wearing the same protective equipment as is used in preparation (including a powered air-purifying respirator) without a hood.
  - a. Broken containers and contaminated packaging mats should be placed in a puncture-resistant receptacle and then in CD disposal bags, which should be closed and placed into appropriate receptacles, both of which are described in section D on waste disposal.
  - b. The appropriate protective equipment and waste-disposal materials should be kept in the area where shipments are received, and employees should be trained in their use and the risks of exposure to CDs.
3. **Transport.** Within the medical facility, drugs should be securely capped or sealed and packaged in impervious packing material for transport.
  - a. Personnel involved in transporting CDs should be cautioned and trained in the necessary procedures should a spill occur, including sealing off the contaminated

area and calling for appropriate assistance.

- b. All drugs should be labeled with a warning label and clearly identified as cytotoxics. Transport methods that produce stress on contents, such as pneumatic tubes, should not be used to transport CDs.

#### H. Training and Information Dissemination.

1. **Training and Personnel.** All personnel involved in any aspect of the handling of CDs (shipment-receiving personnel, physicians, nurses, pharmacists, housekeepers, employees involved in the transport or storage of drugs) must receive an orientation to CDs, including their known risks, relevant techniques and procedures for their handling, the proper use of protective equipment and materials, spill procedures, and medical policies (including those dealing with pregnancy and with staff actively trying to conceive children). Prospective temporary and permanent employees who will be required to work with CDs should receive notice of this requirement. Medical staff who are not hospital employees should be informed of hospital policies and of the expectation that they will comply with these policies.
2. **Evaluation of Staff Performance.** Knowledge and competence of personnel should be evaluated after the first orientation or training session, and then yearly, or more often if a need is perceived. Evaluation may involve direct observation of an individual's performance on the job. In addition, non-CD solutions may be used to evaluate preparation technique; quinine, which will fluoresce under ultraviolet light, provides an easy mechanism for the detection of clumsy technique.
3. **Information.** The pharmacy should maintain a loose-leaf, index card, or computerized file containing information on the toxicity, acute-exposure treatment, chemical inactivators, solubility, and stability of the CDs used in the institution. This file should be available to employees. (Any special instructions for the handling of specific drugs should be included in the orientation and training sessions.) If drugs are administered in a centralized area, such as an oncology floor, a copy of this file should be available there. Summaries of relevant procedures should be posted in the appropriate work areas. A complete policy and procedures manual should be made available to all employees.

## References

- Berk PD, Goldberg JD, Silverstein MN et al. Increased incidence of acute leukemia in polycythemia vera associated with chlorambucil therapy. *N Engl J Med.* 1981; 304:441-7.
- Harris CC. A delayed complication of cancer therapy—cancer. *J Natl Cancer Inst.* 1979; 63:275-7.
- Hunstein W. Tumor inductions by cytostatics in man. *Recent Results Cancer Res.* 1975; 52:50-6.
- IARC Working Group on the Evaluation of the Carcinogenic Risk of Chemicals to Humans. Some antineoplastic and immunosuppressive agents. IARC monographs on the evaluation of the carcinogenic risk of chemicals to humans. Vol 26. May 1981.
- Reimer RR, Hoover R, Fraumeni JB et al. Acute leukemia after alkylating-agent therapy of ovarian cancer. *N Engl J Med.* 1977; 22:177-81.
- Rosner F. Acute leukemia as a delayed consequence of cancer chemotherapy. *Cancer.* 1976; 37:1033-6.
- Sieber SM. Cancer chemotherapeutic agents and carcinogenesis. *Cancer Chemother Rep.* 1975; 59:915-8.
- Sieber SM, Adamson RH. Toxicity of antineoplastic agents in man: chromosomal aberrations, antifertility effects, congenital malformations, and carcinogenic potential. *Adv Cancer Res.* 1975; 22:57-155.
- Weisburger JH, Griswold DP, Prejean JD et al. I. Tumor induction by cytostatics. The carcinogenic properties of some of the principal drugs used in clinical cancer chemotherapy. *Recent Results Cancer Res.* 1975; 52:1-17.
- Rudolph R, Suzuki M, Luce JK. Experimental skin necrosis produced by adriamycin. *Cancer Treat Rep.* 1979; 63:529-37.
- Slickson AS, Mottaz J, Weiss LW. Effects of topical fluorouracil on normal skin. *Arch Dermatol.* 1975; 111:1301-6.
- Directorate of Labour Inspection. Guidelines concerning the handling of cytostatic agents. Oslo, Norway: Aug 1980.
- Hakanasson L, Landersjo L. Instructions for handling and administering of cytostatics. Stockholm, Sweden: National Social Welfare Board; Oct 1978.
- Health and Safety Executive (U.K.). Precautions for the safe handling of cytotoxic drugs. Guidance Note MS 21; 1984.
- Henderson IWD, Sproul J. Guidelines for safe handling of cytotoxic agents. Canada: Bureau of Human Prescription Drugs.
- National Cancer Institute. Safety standards for research involving chemical carcinogens. Rockville, MD: National Cancer Institute, 1975; DHEW publication no. (NIH) 76-900.
- National Institute of Health. NIH guidelines for the laboratory use of chemical carcinogens. Bethesda, MD: National Institute of Health, 1981; NIH publication no. 81-2385.
- National Study Commission on Cytotoxic Exposure. Consensus to unresolved questions concerning cytotoxic agents. Jeffrey LP, chairman. Providence, RI: Rhode Island Hospital; Mar 1984.
- National Study Commission of Cytotoxic Exposure. Recommendations for handling cytotoxic agents. Louis P. Jeffrey LP, chairman. Providence, RI: Rhode Island Hospital; 1984.
- Ontario Hospital Association. A guide for the safe preparation and disposal of antineoplastic drugs. Toronto: Ontario Hospital Association. Oct 1982.
- Society of Hospital Pharmacists of Australia. Guidelines for safe handling of cytotoxic drugs in pharmacy departments and hospital wards. *Hosp Pharm.* 1981; 16:17-20.
- Stolar MH, Power LA. Recommendations for handling cytotoxic drugs in hospitals. *Am J Hosp Pharm.* 1983; 40:1163-71.
- American Society of Hospital Pharmacists. ASHP technical assistance bulletin on handling cytotoxic drugs in hospitals. *Am J Hosp Pharm.* 1985; 42:131-7.
- U.S. Department of Health and Human Services. Recommendations for the safe handling of parenteral antineoplastic drugs. Washington, DC: U.S. Government Printing Office; 1983.
- Zimmerman PF, Larsen RK, Barkley EW et al. Recommendations for the safe handling of injectable antineoplastic drug products. *Am J Hosp Pharm.* 1981; 38:1693-5.
- Levantine A, Almeyda J. Cutaneous reactions to cytostatic agents. *Br J Dermatol.* 1974; 90:239-42.
- Stephens JD, Golbus MS, Miller TR et al. Multiple congenital abnormalities in a fetus exposed to 5-fluorouracil during the first trimester. *Am J Obstet Gynecol.* 1980; 137:747-9.
- Crudi CB. A compounding dilemma: I've kept the drug sterile but have I contaminated myself? *Natl I.V. Ther J.* 1980; 3:77-80.
- Reynolds RD, Ignoffo R, Lawrence J et al. Adverse reactions to AMSA in medical personnel. *Cancer Treat Rep.* 1982; 66:1885.
- Palmer RG, Dore CJ, Denman AM. Chlorambucil-induced chromosome damage to human lymphocytes is dose-dependent and cumulative. *Lancet.* 1984; 1:246-9.
- Hemminki K, Kyyronen P, Lindholm ML. Spontaneous abortions and malformations in the offspring of nurses exposed to anaesthetic gases, cytostatic drugs, and other potential hazards in hospitals, based on registered information of outcome. *J Epidemiol Community Health.* 1985; 39: (In press.)
- Schafer AI. Teratogenic effects of antileukemic therapy. *Arch Intern Med.* 1981; 141:514-5.
- Selevan SG, Lindholm ML, Hornung RW et al. A study of occupational exposure to antineoplastic drugs and fetal loss in nurses. *N Engl J Med.* 1985; 313:1173-221.
- Menard DB, Gisselbrecht C, Marty M et al. Antineoplastic agents and the liver. *Gastroenterology.* 1980; 78:142-64.
- Sternberg SS, Philips FS, Cronin AP. Renal tumors and other lesions in rats following a single intravenous injection of Daunomycin. *Cancer Res.* 1972; 32:1029-36.
- Sotaniemi EA, Sutinen S, Arranto AJ et al. Liver damage in nurses handling cytostatic agents. *Acta Med Scand.* 1983; 214:181-9.
- Hirst M, Tse S, Mills DG et al. Occupational exposure to cyclophosphamide. *Lancet.* 1984; 1:186-8.
- Macek C. Hospital personnel who handle anticancer drugs may face risks. *JAMA.* 1982; 247:11-2.
- Anderson RW, Puckett WH, Dana WJ et al. Risk of handling injectable antineoplastic agents. *Am J Hosp Pharm.* 1982; 39:1881-7.
- Bos RP, Leenars AO, Theuvs JL et al. Mutagenicity of urine from nurses handling cytostatic drugs, influence of smoking. *Int Arch Occup Environ Health.* 1982; 50:359-69.
- Kolmodin-Hedman B, Hartvig P, Sorsa M et al. Occupational handling of cytostatic drugs. *Arch Toxicol.* 1983; 54:25-33.
- Leroy ML, Roberts MJ, Theisen JA. Procedures for handling antineoplastic injections in comprehensive cancer centers. *Am J Hosp Pharm.* 1983; 40:601-3.
- Neal AD, Wadden RA, Chiou WL. Exposure of hospital workers to airborne antineoplastic agents. *Am J Hosp Pharm.* 1983; 40:597-601.
- Avis KE, Levchuck JW. Special considerations in the use of vertical laminar flow workbenches. *Am J Hosp Pharm.* 1984; 41:81-7.
- Donner AL. Possible risks of working with antineoplastic drugs in horizontal laminar flow hoods. *Am J Hosp Pharm.* 1978; 35:900. Letter.
- Juma FD, Rogers HJ, Trounce JR et al. Pharmacokinetics of intravenous cyclophosphamide in man, estimated by gas-liquid chromatography. *Cancer Chemother Pharmacol.* 1978; 1:229-31.
- Siebert D, Simon U. Cyclophosphamide: pilot study of genetically active metabolites in the urine of a treated human patient. *Mutat Res.* 1973; 19:65-72.
- Venitt S, Crofton-Sleigh C, Hunt J et al. Monitoring exposure of nursing and pharmacy personnel to cytotoxic drugs: urinary mutation assays and urinary platinum as markers of absorption. *Lancet.* 1984; 1:74-6.
- Connor TH, Laidlaw JL, Theiss JC et al. Permeability of latex and polyvinyl chloride gloves to carmustine. *Am J Hosp Pharm.* 1984; 41:676-9.
- Laidlaw JL, Connor TH, Theiss JC et al. Permeability of gloves to a spectrum of cytotoxic drugs. Paper presented to

- 41st ASHP Annual Meeting. Boston, MA: 1984 Jun 7.
51. National Sanitation Foundation. National Sanitation Foundation Standard No. 49 for class II (laminar flow) biohazard cabinetry. Ann Arbor, MI: National Sanitation Foundation; May 1983.
  52. Power L. Handling cytotoxics: minimizing risks. *Frankly Speaking*. 1984; 3:1-6.
  53. Hoy RH, Stump LM. Effect of an air-venting filter device on aerosol production from vials. *Am J Hosp Pharm*. 1984; 41:324-6.
  54. Wilson JP, Solimando DA. Aseptic technique as a safety precaution in the preparation of antineoplastic agents. *Hosp Pharm*. 1981; 16:575-6, 589-81.
  55. Vaccari PL, Tonat K, DeChristoforo R et al. Disposal of antineoplastic wastes at the NIH. *Am J Hosp Pharm*. 1984; 41:87-92.
- Additional Background References**
1. Faick K, Grohn P, Sorsa M et al. Mutagenicity in urine of nurses handling cytostatic drugs. *Lancet*. 1979; 1:1250-1.
  2. Knowles RS, Virden JF. Handling of injectable antineoplastic agents. *Br Med J*. 1980; 281:589-91.
  3. Marquardt H, Philips FS, Sternberg SS. Tumorigenicity in vivo and induction of malignant transformation and mutagenesis in cell cultures by adriamycin and daunomycin. *Cancer Res*. 1976; 36:2065-9.
  4. Norppa H, Sorsa M, Vainio H et al. Increased sister chromatid exchange frequencies in lymphocytes of nurses handling cytostatic drugs. *Scand J Work Environ Health*. 1980; 6:299-301.
  5. Perlman M, Walker R. Acute leukemia following cytotoxic chemotherapy. *JAMA*. 1973; 224:250. Letter.
  6. Waksvik H, Klepp O, Brogger A. Chromosome analyses of nurses handling cytostatic drugs. *Cancer Treat Rep*. 1981; 65:607-10.
  7. Wall RL, Clausen KP. Carcinoma of the urinary bladder in patients receiving cyclophosphamide. *N Engl J Med*. 1975; 293:271-3.
  8. Thielde T, Christensen BC. Bladder tumors induced by chlor-naphazin. *Acta Med Scand*. 1969; 185:133-7.
  9. Tortorici MP. Precautions followed by personnel involved with the preparation of parenteral antineoplastic medications. *Hosp Pharm*. 1980; 15:295-301.