

NATIONAL TOXICOLOGY PROGRAM
Technical Report Series
No. 409

TOXICOLOGY AND CARCINOGENESIS

STUDIES OF

QUERCETIN

(CAS NO. 117-39-5)

IN F344/N RATS

(FEED STUDIES)

U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES
Public Health Service
National Institutes of Health

FOREWORD

The National Toxicology Program (NTP) is made up of four charter agencies of the U.S. Department of Health and Human Services (DHHS): the National Cancer Institute (NCI), National Institutes of Health; the National Institute of Environmental Health Sciences (NIEHS), National Institutes of Health; the National Center for Toxicological Research (NCTR), Food and Drug Administration; and the National Institute for Occupational Safety and Health (NIOSH), Centers for Disease Control. In July 1981, the Carcinogenesis Bioassay Testing Program, NCI, was transferred to the NIEHS. The NTP coordinates the relevant programs, staff, and resources from these Public Health Service agencies relating to basic and applied research and to biological assay development and validation.

The NTP develops, evaluates, and disseminates scientific information about potentially toxic and hazardous chemicals. This knowledge is used for protecting the health of the American people and for the primary prevention of disease.

The studies described in this Technical Report were performed under the direction of the NIEHS and were conducted in compliance with NTP laboratory health and safety requirements and must meet or exceed all applicable federal, state, and local health and safety regulations. Animal care and use were in accordance with the Public Health Service Policy on Humane Care and Use of Animals. The prechronic and chronic studies were conducted in compliance with Food and Drug Administration (FDA) Good Laboratory Practice Regulations, and all aspects of the chronic studies were subjected to retrospective quality assurance audits before being presented for public review.

These studies are designed and conducted to characterize and evaluate the toxicologic potential, including carcinogenic activity, of selected chemicals in laboratory animals (usually two species, rats and mice). Chemicals selected for NTP toxicology and carcinogenesis studies are chosen primarily on the bases of human exposure, level of production, and chemical structure. Selection *per se* is not an indicator of a chemical's carcinogenic potential.

These NTP Technical Reports are available for sale from the National Technical Information Service, U.S. Department of Commerce, 5285 Port Royal Road, Springfield, VA 22161 (703-487-4650). Single copies of this Technical Report are available without charge while supplies last from the NTP Central Data Management, NIEHS, P.O. Box 12233, MD A0-01, Research Triangle Park, NC 27709 (919-541-1371).

NTP TECHNICAL REPORT
ON THE
TOXICOLOGY AND CARCINOGENESIS
STUDIES OF QUERCETIN
(CAS NO. 117-39-5)
IN F344/N RATS
(FEED STUDIES)

NATIONAL TOXICOLOGY PROGRAM
P.O. Box 12233
Research Triangle Park, NC 27709

September 1992

NTP TR 409

NIH Publication No. 92-3140

U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES
Public Health Service
National Institutes of Health

CONTRIBUTORS

National Toxicology Program

Evaluated and interpreted results and reported findings

C.J. Alden, Ph.D.
 G.A. Boorman, D.V.M., Ph.D.
 D.A. Bridge, B.S.
 J.K. Dunnick, Ph.D.
 S.L. Eustis, D.V.M., Ph.D.
 T.J. Goehl, Ph.D.
 R.A. Griesemer, D.V.M., Ph.D.
 J.R. Hailey, D.V.M.
 J.K. Haseman, Ph.D.
 C.W. Jameson, Ph.D.
 G.N. Rao, D.V.M., Ph.D.
 D.B. Walters, Ph.D.
 K.L. Witt, M.S., Oak Ridge Associated Universities

EG&G Mason Research Institute

Conducted studies, evaluated pathology findings

H.S. Lilja, Ph.D., Principal Investigator
 A.J. Block, Ph.D.
 H.J. Esber, Ph.D.
 R.W. Fleischman, D.V.M.
 M. Hagopian, Ph.D.

Experimental Pathology Laboratories, Inc.

Provided pathology quality assessment

B. F. Hamilton, D.V.M., Ph.D, Principal Investigator

Integrated Laboratory Systems

Prepared quality assurance audits

J.C. Bhandari, D.V.M., Ph.D., Principal Investigator

NTP Pathology Working Group

Evaluated slides, prepared pathology report (27 April 1990)

R.M. Kovatch, D.V.M., Chair
 Pathology Associates, Inc.
 J. Cullen, V.M.D., Ph.D.
 North Carolina State University
 B. F. Hamilton, D.V.M., Ph.D.
 Experimental Pathology Laboratories
 S. Imoto, D.V.M., Ph.D.,
 Shin Nippon Biomedical Laboratories
 M.P. Jokinen, D.V.M.
 National Toxicology Program
 J. Mahler, D.V.M.
 National Toxicology Program
 E.E. McConnell, D.V.M.
 Consultant
 M.M. McDonald, D.V.M., Ph.D.
 National Toxicology Program

Biotechnical Services, Inc.

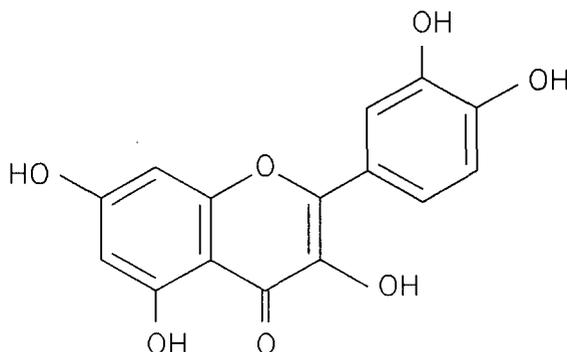
Prepared Technical Report

L.G. Cockerham, Ph.D., Principal Investigator
 P. Chaffin, M.S.
 G.F. Corley, D.V.M.
 D.D. Lambright, Ph.D.
 W.D. Sharp, B.A., B.S.

CONTENTS

| | |
|---|------------|
| ABSTRACT | 5 |
| EXPLANATION OF LEVELS OF EVIDENCE OF CARCINOGENIC ACTIVITY | 8 |
| PEER REVIEW PANEL | 9 |
| SUMMARY OF PEER REVIEW COMMENTS | 10 |
| INTRODUCTION | 11 |
| MATERIALS AND METHODS | 17 |
| RESULTS | 23 |
| DISCUSSION AND CONCLUSIONS | 35 |
| REFERENCES | 43 |
| APPENDIX A Summary of Lesions in Male Rats in the 2-Year Feed Study | 49 |
| APPENDIX B Summary of Lesions in Female Rats in the 2-Year Feed Study | 93 |
| APPENDIX C Genetic Toxicology | 137 |
| APPENDIX D Organ Weights and Organ-Weight-to-Body-Weight Ratios | 143 |
| APPENDIX E Hematology, Clinical Chemistry, and Urinalysis Results | 147 |
| APPENDIX F Chemical Characterization and Dose Formulation Studies | 153 |
| APPENDIX G Feed and Compound Consumption in the 2-Year Feed Studies | 161 |
| APPENDIX H Ingredients, Nutrient Composition, and Contaminant Levels in NIH-07 Rat and Mouse Ration | 165 |
| APPENDIX I Sentinel Animal Program | 171 |

ABSTRACT



QUERCETIN

CAS No. 117-39-5

Chemical Formula: $C_{15}H_{10}O_7$ Molecular Weight: 302.23

Synonyms: C.I. Natural Yellow 10; C.I. 75670; Cyanidelonon 1522; Flavin Meletin; Quercetine; Quercetol; Quertin; Quertine; Sophoretin; Xanthaurine; 3,3',4',5,7-Pentahydroxyflavone; 3,5,7,3',4'-Pentahydroxyflavone; 2-(3,4-Dihydroxyphenyl)-3,5,7-trihydroxy-4H-1-benzopyran-4-one

Quercetin is a member of a group of naturally occurring compounds, the flavonoids, which have a common flavone nucleus composed of two benzene rings linked through a heterocyclic pyrone ring. Quercetin is found in various plants, food products, and dyes of natural origin. The estimated average daily intake of quercetin by an individual in the United States is 25 mg. The Food and Drug Administration nominated quercetin for toxicity and carcinogenicity studies in the rat because it is a chemical that is widely distributed in foods. Quercetin was administered to rats by dosed feed since human exposure is by dietary consumption.

Information in the literature showed that quercetin administered in the diet to rats at levels up to approximately 4% caused a minor body weight effect, whereas higher dose levels produced greater than 10% reduction in body weight gains relative to controls. Based on this information, the NTP 2-year

studies were conducted by administering 0, 1,000, 10,000, or 40,000 ppm quercetin (>95% pure) in feed to groups of 50 male and female rats for 104 weeks. Ten additional animals per dose group were evaluated at 6 and 15 months.

Body Weight, Survival, and Clinical Findings in the 2-Year Studies

Body weights of exposed male and female rats given 1,000 and 10,000 ppm were within 5% of controls throughout the studies. Reduced body weight gain in male and female rats receiving 40,000 ppm was observed by week 15 and the final mean body weights were 87% of controls at week 104. Survival and feed consumption were similar among exposed and control groups throughout the studies. The average amounts of quercetin consumed per day by the 1,000, 10,000 and 40,000 ppm dose groups after week 52 were 40, 400, and 1,900 mg/kg of body weight.

Nonneoplastic and Neoplastic Effects in the 2-Year Studies

In male rats, the principal toxic effects associated with the dietary administration of quercetin for 2 years were observed in the kidney. There were dose-related increases in the severity of chronic nephropathy (control, 2.7; low-dose, 2.7; mid-dose, 3.0; high-dose, 3.2) and a slight increased incidence in focal hyperplasia of the renal tubule epithelium (1/50; 2/50; 3/50; 4/50). Parathyroid hyperplasia, indicative of renal secondary hyperparathyroidism, also increased incidence in dosed male rats (1/43, 6/45, 6/43, 17/43).

The evaluation of single sections from the left and right kidneys revealed renal tubule adenomas in three male rats and adenocarcinomas in another male rat receiving 40,000 ppm quercetin; none were seen in the controls. Examination of additional step sections of the male rat kidney identified additional hyperplasia and adenomas in all dose groups (hyperplasia: 2/50, 2/50, 6/50, 8/50; adenoma: 1/50, 2/50, 7/50, 6/50). The overall incidence of renal tubule adenoma or adenocarcinoma combined in male rats was 1/50 in controls and 9/50 in the high-dose group.

There was no apparent effect of quercetin on the kidney of female rats. A single renal tubule adenoma was seen in a female receiving 10,000 ppm; this neoplasm was not considered biologically significant.

There was a statistically significant, dose-related decrease in the incidence of mammary gland fibroadenomas in exposed female rats (29/50, 27/50, 16/50, 9/50), which may in part be attributed to lower body weight gains.

There was a treatment-related accumulation of yellow-brown granular pigment adsorbed to or absorbed by the epithelial cells of the glandular stomach, ileum, jejunum, and, to a lesser extent, the duodenum and colon. The severity of the pigmentation in these tissues increased with increased length of exposure. There were no other lesions considered to be related to chemical administration.

Genetic Toxicology

Quercetin induced gene mutations in *Salmonella typhimurium* strains TA100 and TA98 with and without exogenous metabolic activation (S9). Positive results were also obtained in tests with and without S9 for induction of sister chromatid exchanges and chromosomal aberrations in Chinese hamster ovary cells.

Conclusions

Under the conditions of these 2-year feed studies there was *some evidence of carcinogenic activity** of quercetin in male F344/N rats based on an increased incidence of renal tubule cell adenomas. There was *no evidence of carcinogenic activity* of quercetin in female F344/N rats receiving 1,000, 10,000 or 40,000 ppm. The incidence of renal tubule hyperplasia and the severity of nephropathy were increased in exposed male rats.

* Explanation of Levels of Evidence of Carcinogenic Activity is on page 8. A summary of peer review comments and the public discussion on this Technical Report appear on page 10.

Summary of the 2-Year Carcinogenesis and Genetic Toxicology Studies of Quercetin

| | Male F344/N Rats | Female F344/N Rats |
|---|--|---|
| Doses | 0, 1,000, 10,000, or 40,000 ppm in feed | 0, 1,000, 10,000, or 40,000 ppm in feed |
| Final body weights (% of controls) | 97%, 95%, 87% | 101%, 98%, 87% |
| 2-Year survival rates | 26/50, 29/50, 25/50, 25/50 | 30/50, 28/50, 35/50, 28/50 |
| Nonneoplastic effects | Kidney: renal tubule hyperplasia (single sections): 1/50, 2/50, 3/50, 4/50; (step sections): 2/50, 2/50, 6/50, 8/50; chronic nephropathy (severity grades: 2.7, 2.7, 3.0, 3.2) | None |
| Neoplastic effects | Kidney (single sections): adenoma - 0/50, 0/50, 0/50, 3/50; adenocarcinoma - 0/50, 0/50, 0/50, 1/50; (step sections): adenoma - 1/50, 2/50, 7/50, 6/50 | None |
| Level of evidence of carcinogenic activity | Some evidence | No evidence |
| Genetic toxicology | | |
| <i>Salmonella typhimurium</i> (gene mutation): Sister chromatid exchanges | Positive with and without S9 in strains TA100 and TA98 | |
| Chinese hamster ovary cells <i>in vitro</i> : Chromosomal aberrations | Positive with and without S9 | |
| Chinese hamster ovary cells <i>in vitro</i> : | Positive with and without S9 | |

EXPLANATION OF LEVELS OF EVIDENCE OF CARCINOGENIC ACTIVITY

The National Toxicology Program describes the results of individual experiments on a chemical agent and notes the strength of the evidence for conclusions regarding each study. Negative results, in which the study animals do not have a greater incidence of neoplasia than control animals, do not necessarily mean that a chemical is not a carcinogen, inasmuch as the experiments are conducted under a limited set of conditions. Positive results demonstrate that a chemical is carcinogenic for laboratory animals under the conditions of the study and indicate that exposure to the chemical has the potential for hazard to humans. Other organizations, such as the International Agency for Research on Cancer, assign a strength of evidence for conclusions based on an examination of all available evidence, including animal studies such as those conducted by the NTP, epidemiologic studies, and estimates of exposure. Thus, the actual determination of risk to humans from chemicals found to be carcinogenic in laboratory animals requires a wider analysis that extends beyond the purview of these studies.

Five categories of evidence of carcinogenic activity are used in the Technical Report series to summarize the strength of the evidence observed in each experiment: two categories for positive results (**clear evidence** and **some evidence**); one category for uncertain findings (**equivocal evidence**); one category for no observable effects (**no evidence**); and one category for experiments that cannot be evaluated because of major flaws (**inadequate study**). These categories of interpretative conclusions were first adopted in June 1983 and then revised in March 1986 for use in the Technical Report series to incorporate more specifically the concept of actual weight of evidence of carcinogenic activity. For each separate experiment (male rats, female rats, male mice, female mice), one of the following five categories is selected to describe the findings. These categories refer to the strength of the experimental evidence and not to potency or mechanism.

- **Clear evidence** of carcinogenic activity is demonstrated by studies that are interpreted as showing a dose-related (i) increase of malignant neoplasms, (ii) increase of a combination of malignant and benign neoplasms, or (iii) marked increase of benign neoplasms if there is an indication from this or other studies of the ability of such tumors to progress to malignancy.
- **Some evidence** of carcinogenic activity is demonstrated by studies that are interpreted as showing a chemical-related increased incidence of neoplasms (malignant, benign, or combined) in which the strength of the response is less than that required for clear evidence.
- **Equivocal evidence** of carcinogenic activity is demonstrated by studies that are interpreted as showing a marginal increase of neoplasms that may be chemical related.
- **No evidence** of carcinogenic activity is demonstrated by studies that are interpreted as showing no chemical-related increases in malignant or benign neoplasms.
- **Inadequate study** of carcinogenic activity is demonstrated by studies that, because of major qualitative or quantitative limitations, cannot be interpreted as valid for showing either the presence or absence of carcinogenic activity.

When a conclusion statement for a particular experiment is selected, consideration must be given to key factors that would extend the actual boundary of an individual category of evidence. Such consideration should allow for incorporation of scientific experience and current understanding of long-term carcinogenesis studies in laboratory animals, especially for those evaluations that may be on the borderline between two adjacent levels. These considerations should include:

- adequacy of the experimental design and conduct;
- occurrence of common versus uncommon neoplasia;
- progression (or lack thereof) from benign to malignant neoplasia as well as from preneoplastic to neoplastic lesions;
- some benign neoplasms have the capacity to regress but others (of the same morphologic type) progress. At present, it is impossible to identify the difference. Therefore, where progression is known to be a possibility, the most prudent course is to assume that benign neoplasms of those types have the potential to become malignant;
- combining benign and malignant tumor incidence known or thought to represent stages of progression in the same organ or tissue;
- latency in tumor induction;
- multiplicity in site-specific neoplasia;
- metastases;
- supporting information from proliferative lesions (hyperplasia) in the same site of neoplasia or in other experiments (same lesion in another sex or species);
- presence or absence of dose relationships;
- statistical significance of the observed tumor increase;
- concurrent control tumor incidence as well as the historical control rate and variability for a specific neoplasm;
- survival-adjusted analyses and false positive or false negative concerns;
- structure-activity correlations; and
- in some cases, genetic toxicology.

PEER REVIEW PANEL

The members of the Technical Reports Review Subcommittee who evaluated the NTP draft Technical Report on quercetin on March 11, 1991 are listed below. Panel members serve as independent scientists, not as representatives of any institution, company, or governmental agency. In this capacity, panel members have five major responsibilities in reviewing NTP studies:

- to ascertain that all relevant literature data have been adequately cited and interpreted,
- to determine if the design and conditions of the NTP studies were appropriate,
- to ensure that the Technical Report presents the experimental results and conclusions fully and clearly,
- to judge the significance of the experimental results by scientific criteria, and
- to assess the evaluation of the evidence of carcinogenic activity and other observed toxic responses.

Daniel S. Longnecker, M.D., Chair
Department of Pathology
Dartmouth Medical School
Hanover, NH

Paul T. Bailey, Ph.D.
Toxicology Division
Mobil Oil Corporation
Princeton, NJ

Louis S. Beliczky, M.S., M.P.H., Principal Reviewer
Department of Industrial Hygiene
United Rubber Workers International Union
Akron, OH

Gary P. Carlson, Ph.D.
Department of Pharmacology and Toxicology
Purdue University
West Lafayette, IN

Harold Davis, D.V.M., Ph.D.
School of Aerospace Medicine
Brooks Air Force Base, TX

Robert H. Garman, D.V.M., Principal Reviewer
Consultants in Veterinary Pathology
Murrysville, PA

Jay I. Goodman, Ph.D., Principal Reviewer
Department of Pharmacology and Toxicology
Michigan State University
East Lansing, MI

David W. Hayden, D.V.M., Ph.D.
Department of Veterinary Pathobiology
College of Veterinary Medicine
University of Minnesota
St. Paul, MN

Curtis D. Klaassen, Ph.D.
Department of Pharmacology and Toxicology
University of Kansas Medical Center
Kansas City, KS

Barbara McKnight, Ph.D.
Department of Biostatistics
University of Washington
Seattle, WA

Ellen K. Silbergeld, Ph.D.*
University of Maryland Medical School
Baltimore, MD

Lauren Zeise, Ph.D.
California Department of Health Services/RCHAS
Berkeley, CA

*Did not attend

SUMMARY OF PEER REVIEW COMMENTS

On March 11, 1991, the draft Technical Report on the toxicology and carcinogenesis studies of quercetin received public review by the National Toxicology Program Board of Scientific Counselors' Technical Reports Review Subcommittee and associated Panel of Experts. The review meeting was held at the National Institute of Environmental Health Sciences, Research Triangle Park, NC.

Dr. J.K. Dunnick, NIEHS, introduced the toxicology and carcinogenesis studies of quercetin by discussing the uses of the chemical and rationale for study, describing the experimental design, reporting on survival and body weight effects, and commenting on neoplasms and nonneoplastic lesions of the kidneys in male and female rats. The proposed conclusions were *some evidence of carcinogenic activity* in male rats and *no evidence of carcinogenic activity* in female rats. Dr. Dunnick added that because of the low but slightly increased number of renal neoplasms in male rats, additional step sections of residual kidneys from all control and high-dose rats were cut and evaluated.

Dr. Garman, a principal reviewer, agreed with the proposed conclusions. He thought the conclusions in male rats were quite reasonable based both on the frequencies of hyperplasia and of benign and malignant renal tubule epithelial neoplasms and on the morphology of these neoplasms. Dr. Garman asked whether the induction of neoplasms was related to hyaline droplet nephropathy, and if so, he thought this might imply a decreased level of concern with regard to human exposure to quercetin. Dr. J.R. Hailey, NIEHS, said there was no evidence for the hyaline droplet nephropathy in this study and also no evidence of kidney lesions from interim evaluations at 6 and 15 months. Dr. Garman asked for clarification of the identity and tissue location of the pigment found in the gastrointestinal tract. Dr. Hailey said the identity was not determined.

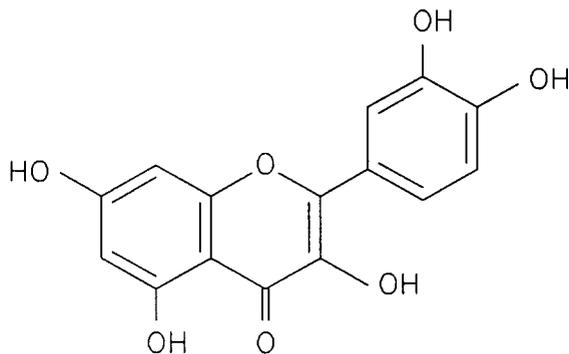
Dr. Goodman, the second principal reviewer, agreed with the proposed conclusions. He inquired what effect the procedure of step sectioning of the kidneys has on the incidence of kidney neoplasms in control and treated animals. Dr. J. Haseman, NIEHS, said that for the eight NTP studies for which step sections have been evaluated, the control rate of renal tubule neoplasms in male rats is 3.7%, or slightly more than double the rate in the current historical control database of 1.6%. Dr. Dunnick reported that the findings from the step sections in other studies have been supportive of the original diagnoses. Dr. Goodman suggested that specific references of studies on chemically induced α_2 -globulin nephropathy in male rats should be considered for inclusion in the discussion. Dr. Dunnick said they would be added.

Mr. Beliczky, the third principal reviewer, agreed with the proposed conclusions. He commented that the increased sensitivity of detection for renal neoplasms and preneoplastic lesions resulting from step sectioning was impressive. He asked whether studies on quercetin had been done in mice. Dr. Dunnick responded that several previous studies by others in mice had shown no evidence of carcinogenic effects.

Dr. Carlson said he was not convinced that two squamous cell carcinomas of the tongue in high-dose female rats were unrelated to chemical administration. Dr. Dunnick said the number was within the historical control range and microscopic analysis indicated no supporting preneoplastic lesions.

Dr. Garman moved that the Technical Report on quercetin be accepted with the revisions discussed and the conclusions as written for male rats, *some evidence of carcinogenic activity*, and for female rats, *no evidence of carcinogenic activity*. Dr. Goodman seconded the motion, which was accepted unanimously with 10 votes.

INTRODUCTION



QUERCETIN

CAS No. 117-39-5

Chemical Formula: $C_{15}H_{10}O_7$ Molecular Weight: 302.23

Synonyms: C.I. Natural Yellow 10; C.I. 75670; Cyanidelonon 1522; Flavin Meletin; Quercetine; Quercetol; Quertin; Quertine; Sophoretin; Xanthaurine; 3,3',4',5,7-Pentahydroxyflavone; 3,5,7,3',4'-Pentahydroxyflavone; 2-(3,4-Dihydroxyphenyl)-3,5,7-trihydroxy-4H-1-benzopyran-4-one

PHYSICAL AND CHEMICAL PROPERTIES, PRODUCTION, OCCURRENCE, AND USE

Quercetin is a yellow, crystalline solid with a bitter taste, which is insoluble in water, slightly soluble in alcohol, and soluble in glacial acetic acid and aqueous alkaline solutions (Weast, 1979; *Merck Index*, 1983). Quercetin is a member of a group of naturally occurring compounds, the flavonoids, which have a common flavone nucleus composed of two benzene rings linked through a heterocyclicpyrone ring. Animals are unable to synthesize the flavone nucleus; thus, flavonoids are found exclusively in the plant kingdom. Quercetin and more than 2,000 other flavonoids occur as condensation products of β -glycosides (Herrmann, 1976; Kuhnau, 1976; Brown, 1980; IARC, 1983). Quercetin is found in various food products and plants, including fruits, seeds, vegetables, tea, coffee, bracken fern, and natural

dyes. Quercetin is usually obtained from the hydrolysis of rutin (quercetin-3-rutinoside), a naturally occurring flavonoid glycoside (Griffith *et al.*, 1955), although it can also be synthesized (Shakhova *et al.*, 1962).

Flavonoids, including quercetin, were once thought to have therapeutic applications, including induction of smooth muscle relaxation, reduction of capillary fragility, and as anti-inflammatory agents. However, in 1970, the Food and Drug Administration withdrew its approval of drugs containing rutin or quercetin because there was insufficient evidence to support the reported pharmacologic effects (Brown, 1980; IARC, 1983). The total flavonoid intake in the U.S. is estimated at 1 g per person per day, with an average daily intake of the individual flavonoid, quercetin, of approximately 25 mg per person (Kuhnau, 1976).

METABOLISM AND DISTRIBUTION

Quercetin glycosides are relatively poorly absorbed by the small intestine. Microflora of the lower bowel hydrolyze the flavonide-glycoside to quercetin and the sugar, and quercetin is then absorbed into the enterohepatic system (Brown, 1980; Tamura *et al.*, 1980; Bokkenheuser *et al.*, 1987). After oral administration of quercetin to rabbits (Booth *et al.*, 1956) or rats (Petrakis *et al.*, 1959), three metabolites of quercetin were identified in the urine: 3,4-dihydroxyphenylacetic acid, 3-methoxy-4-hydroxyphenylacetic acid (homovanillic acid), and *m*-hydroxyphenylacetic acid. These metabolites are thought to be formed in the liver after fusion of the heterocyclicpyrone ring. When Brown and Griffiths (1983) administered quercetin to rats by intraperitoneal injection, they identified the 3'-*o*-methyl-ether of quercetin (isorhamnetin) as a metabolite in bile.

The distribution, metabolism and excretion of 4-[¹⁴C] quercetin in male ACI rats were studied by autoradiography and quantitation of radioactivity (Ueno *et al.*, 1983). After oral administration, 20% of the dose was absorbed from the digestive tract and then excreted into the bile and urine within 48 hours as glucuronide or sulfate conjugates. Autoradiographic analysis of a rat 3 hours after receiving a single 2.3 mg/kg oral dose of quercetin showed that most of the radioactivity remained in the digestive tract with low levels seen in the blood, liver, kidney, lung, and rib.

In five human volunteers, no quercetin was detected in the plasma or urine after oral administration of 4 g of quercetin (Gugler *et al.*, 1975).

TOXICITY

The oral LD₅₀ of quercetin was reported as 160 mg/kg in the mouse and 161 mg/kg in the rat; the LD₅₀ in the mouse by the subcutaneous route was reported as 97 mg/kg (Sullivan *et al.*, 1951). The purity of the compound used in this study was not specified. Subsequent studies have shown that rodents tolerate much higher doses of quercetin.

Rats fed diets containing up to 1% quercetin for 410 days showed no decrease in body weight gain and no compound-related histopathologic lesions (Ambrose *et al.*, 1952).

REPRODUCTIVE TOXICOLOGY

The reproductive toxicity of quercetin was studied in male and female F344 rats fed diets containing 0.1% or 0.2% quercetin from birth to breeding during week 12 or 13. During gestation and lactation, animals were fed diets without quercetin. Quercetin had no effect on mean viable litter size, live birth index, 3-day survival of pups, lactation index, or weight of pups at birth or at 21 days (Stoewsand *et al.*, 1984). When 0, 20, 200, or 2,000 mg/kg quercetin was administered to Sprague-Dawley rats from days 6 through 15 of gestation, no overt signs of toxicity were seen in the dams even at the highest dose, but average fetal weight of the 2,000 mg/kg group was reduced relative to control fetal weight. No fetal abnormalities attributable to chemical administration were observed (Willhite, 1982).

CARCINOGENICITY

Quercetin has been studied in a variety of test systems for carcinogenicity and in the majority of these studies there was no evidence of neoplasms related to chemical administration (Table 1). In a 2-year study of F344 rats, 0%, 1.25% or 5% quercetin was administered in the diet for 104 weeks, followed by an additional 8-week recovery period (Ito *et al.*, 1989). Major tissues and organ systems were examined histopathologically. Hyperplastic polyps of the cecum were found in males and females fed diets containing 5% quercetin. An adenoma and two adenocarcinomas of the cecum were observed in high-dose males, while two adenomas of the colon were observed in the high-dose females. The incidences of these neoplasms were not considered statistically significant and the authors concluded that there was no evidence for any clear carcinogenic effect.

TABLE 1
Quercetin Rodent Carcinogenicity Studies

| Strain of Rodent | Route of Administration and Dose ^a | Length of Dosing | Histopathologic Findings ^b | Reference |
|---|---|---|--|----------------------------------|
| Male and female F344/DuCrj rats | Diet 0, 1.25, 5.0% | 104 weeks | Negative | Ito <i>et al.</i> , 1989 |
| Male and female albino rats | Diet 0, 0.1% quercetin | 58 weeks | Intestinal and urinary bladder neoplasms in treated groups | Pamukcu <i>et al.</i> , 1980 |
| Male and female ACI rats | Diet 1, 5, 10% | 850 days | Negative | Hirono <i>et al.</i> , 1981 |
| Male and female F344 rats | Diet 0, 0.1% | 540 days | Negative | Takanashi <i>et al.</i> , 1983 |
| Male and female ddY mice | Diet 0, 2% | 842 days | Negative | Saito <i>et al.</i> , 1980 |
| Male and female golden hamsters | Diet 0, 1, 4% | 351 to 709 days | Negative | Morino <i>et al.</i> , 1982 |
| Female ICR/Ha Swiss mice | DMBA as initiator on skin, 25 mg quercetin applied to the skin 3 times per week for 25 weeks | 368 days | Negative (no skin neoplasm induction) | Van Duuren and Goldschmidt, 1976 |
| Male F344 rats (effects on initiation/promotion in urinary bladder) | Diet 0, 5% | a) Quercetin given for 25 weeks after initiation with 0.01% BHBN, b) 5% quercetin given as an initiator for 4 weeks followed by 0.001% BHBN for 29 weeks | No effects on initiation/promotion in urinary bladder | Hirose <i>et al.</i> , 1983 |
| Female ICR mice | Skin initiated with DMBA, promoted with telocidin twice per week, quercetin (30 μ mol) treatment applied topically with telocidin | 20 weeks | Suppressed skin neoplasm formation | Nishino <i>et al.</i> , 1984a |

(continued)

TABLE 1
Quercetin Rodent Carcinogenicity Studies (continued)

| Strain of Rodent | Route of Administration and Dose | Length of Dosing | Histopathologic Findings ^a | Reference |
|---|--|------------------|--|---------------------------|
| Male and female A(A/JJms) mice ^c | Diet 0, 5% | 23 weeks | Negative (no increase or decrease in lung neoplasms) | Hosaka and Hirono, 1981 |
| Female CD-1 mice | Skin-initiated with DMBA, promoted with TPA, quercetin (30 μ mol) applied topically after each TPA treatment | 18 weeks | Suppressed skin neoplasm formation | Kato <i>et al.</i> , 1983 |

^a DMBA = 7,12-dimethyl[a]anthracene; TPA = 12-*o*-tetradecanoylphorbol-13-acetate;

BHBN = N-butyl-N-(4-hydroxybutyl)nitrosamine

^b Negative = no evidence for neoplasms related to administration of quercetin

^c This strain develops lung neoplasms at a low incidence by week 23.

Quercetin has been shown to inhibit the promotion of skin neoplasms in the mouse (Kato *et al.*, 1983; Nishino *et al.*, 1984a) and to suppress the formation of urinary bladder neoplasms (Hirose *et al.*, 1983). Quercetin had no effect on the formation of lung neoplasms in strain A mice (Hosaka and Hirono, 1981) and did not induce preneoplastic glutathione S-transferase placental form-positive foci in F344 rats (Ito *et al.*, 1988). When quercetin was given intraperitoneally for 6 days at a dose of 500 mg/kg per day to hepatectomized rats followed by phenobarbital treatment, there was no increase in liver neoplasms compared to rats treated in the same manner without quercetin (Kato *et al.*, 1985).

Pamukcu *et al.* (1980) reported that albino rats (Norwegian strain) fed a diet containing 0.1% quercetin for 58 weeks showed an increased incidence of intestinal and urinary bladder neoplasms in dosed animals. Other long-term rat studies have not confirmed this carcinogenic effect (Hirono *et al.*, 1981; Takanashi *et al.*, 1983; Ito *et al.*, 1989). Long-term studies in mice also showed no carcinogenic effects from quercetin (Saito *et al.*, 1980). Some of these animal studies showed that

quercetin can inhibit the promotion of neoplasms (Kato *et al.*, 1983). Follow-up studies *in vitro* suggest that quercetin can inhibit cell proliferation. Quercetin has been shown to have pleiotropic effects on the transformation of BALB 3T3 cells. At a concentration of 0.5 μ g/mL, quercetin suppresses the promoting action of 12-*o*-tetradecanoylphorbol-13-acetate on cells initiated with 20-methylcholanthrene (MCA), but at higher concentrations (5.0 μ g/mL), quercetin enhanced cell transformation by MCA (Tanaka *et al.*, 1987).

Quercetin has been shown to inhibit cell proliferation in Ehrlich ascites neoplasms cells and to inhibit thymidine incorporation (Graziani and Chayoth, 1979). Quercetin also inhibits the growth of squamous cell carcinoma lines *in vitro* (Castillo *et al.*, 1989).

Bracken fern, which contains quercetin and many other chemicals, causes mononuclear cell leukemia, intestinal neoplasms, urinary bladder carcinomas, and mammary adenocarcinomas in rats; urinary bladder neoplasms in guinea pigs; alimentary tract and urinary bladder cancers in cattle; and intestinal

carcinomas and hepatomas in toads (IARC, 1986, 1987). The neoplasms appear to be caused by known carcinogens, such as shikimic acid and tannin, also found in bracken fern (Evans, 1984; Hirono, 1986).

GENETIC TOXICITY

The mutagenicity of quercetin and other flavonoids has been reviewed by Sugimura *et al.* (1977), Brown (1980), and Nagao *et al.* (1981). Structural requirements for mutagenic activity of flavonoids in *Salmonella* are discussed in detail by MacGregor and Jurd (1978) and Nagao *et al.* (1981). Mutagenic flavonoids (primarily the 3-hydroxyflavones) generally contain a free hydroxyl group at the 3 position, a double bond between positions 2 and 3, and a keto group at the 4 position. The presence of exogenous metabolic activating systems may render some of these structural requirements nonessential. Of the flavonoids, quercetin exhibits the strongest mutagenic activity. Quercetin induces gene mutations in base substitution as well as frameshift strains of *Salmonella typhimurium*, with and without exogenous metabolic activation, although activation increases the magnitude of the mutagenic response (Bjeldanes and Chang, 1977; Hardigree and Epler, 1978; Brown and Dietrich, 1979; Bartholomew and Ryan, 1980; McCoy *et al.*, 1983; Stoewsand *et al.*, 1984; Kuroda, K. *et al.*, 1985; Kuroda, M. *et al.*, 1985; Busch *et al.*, 1986; Löfroth *et al.*, 1986; Rueff *et al.*, 1986; Brams *et al.*, 1987; Marzin *et al.*, 1987; MacGregor and Wilson, 1988). It has also been reported to induce sex-linked recessive lethal mutations in germ cells of male *Drosophila melanogaster* (Watson, 1982). Tests for chromosomal effects in mammalian cell cultures have also been positive with quercetin: it induced chromosomal aberrations and sister chromatid exchanges, in the absence of S9 metabolic activation, in Chinese hamster Don-6 and B-131 fibroblasts (Yoshida *et al.*, 1980), in Chinese hamster ovary cells (Stich *et al.*, 1981; Carver *et al.*, 1983), and in human HE2144 fibroblasts and leukocytes (Yoshida *et al.*, 1980). In addition, sister chromatid exchanges (Rueff *et al.*, 1986) and chromosomal aberrations (Marzin *et al.*, 1987) were induced by quercetin in the presence, as well as the absence, of S9 in human peripheral lymphocyte cultures. Despite the consistently positive results for quercetin *in vitro* assays for genotoxic activity, most of the *in vivo* test data were negative.

Quercetin (maximum dose of 1,000 mg/kg) did not induce micronuclei in bone marrow erythrocytes of mice exposed either by intraperitoneal injection or gavage (Aeschbacher *et al.*, 1982; MacGregor *et al.*, 1983); feed studies (5% and 10% in chow for 8 days) also yielded negative results for micronucleus induction (MacGregor *et al.*, 1983). No increase in the frequency of sister chromatid exchanges in rabbit lymphocytes was observed 1 or 7 days after intraperitoneal (i.p.) injection of 250 to 1,000 mg/kg quercetin (MacGregor *et al.*, 1983). Results of dominant lethal assays with quercetin in male Swiss mice (200 to 400 mg/kg i.p.) and Wistar rats (200 and 300 mg/kg i.p.) were also negative (Aravindakshan *et al.*, 1985).

Feces or fecal extracts from laboratory rats fed quercetin showed mutagenic activity in *Salmonella typhimurium* and urine from treated rats showed a small amount of mutagenic activity (in proportion to administered dose) (Stoewsand *et al.*, 1984; Crebelli *et al.*, 1987). Bacterial-mediated degradation of quercetin in the gut and lack of absorption may be contributing factors to the observed lack of *in vivo* genetic effects. Additionally, bacterial gene mutation studies with known metabolites of quercetin have shown little effect: 3-hydroxyphenylacetic acid, 3,4-dihydroxyphenylacetic acid, homovanillic acid, and phloroglucinol carboxylic acid were all negative for gene reversion induction in *S. typhimurium* strains TA98 and TA100 (Bjeldanes and Chang, 1977; MacGregor and Jurd, 1978; Hatcher *et al.*, 1981). However, positive results were obtained in *S. typhimurium* with dihydroquercetin (TA98) and isorhamnetin (TA98, TA100) (MacGregor and Jurd, 1978; Nagao *et al.*, 1981). 3,4-Dihydroxyphenylacetic acid (100 µg/mL) caused chromosomal aberrations in Chinese hamster ovary cells treated for 3 hours with or without S9 activation (Stich *et al.*, 1981).

STUDY RATIONALE

Quercetin was nominated by The Food and Drug Administration for toxicity and carcinogenicity studies in the rat because it is widely distributed in natural foods, and although long-term studies had been conducted previously in both rats and mice, there was conflicting information on the carcinogenicity of quercetin in the rat. Quercetin was administered to rats by dosed feed because human exposure is by dietary consumption.

MATERIALS AND METHODS

PROCUREMENT AND CHARACTERIZATION

Quercetin was obtained from Freeman Industries (Tuckahoe, NY) in two lots. It was prepared by hydrolyzing rutin (quercetin-3-rutinoside), a naturally occurring flavonoid glycoside. Both lots (lot no. 969-3790-05, anhydrous form, and lot no. 969-0483-18BL, dihydrate form) were used throughout the studies. Identity, purity, and stability analyses were conducted by the analytical chemistry laboratory, Midwest Research Institute (Kansas City, Missouri) (Appendix F). The study chemical, a yellow crystalline powder, was identified as quercetin by infrared, ultraviolet/visible, and nuclear magnetic resonance spectroscopy.

Both lots were greater than 95% pure, as determined by titration, Karl Fischer water analysis, weight loss on drying, chromatographic analysis, nuclear magnetic resonance spectroscopy, and elemental analyses. The largest impurity was identified by spectroscopy and mass spectrometry as ellagic acid (2.6% in lot 969-3790-05 and 1.1% in lot 969-0483-18BL). Stability studies performed by high-performance liquid chromatography indicated that quercetin was stable as a bulk chemical for at least 2 weeks at temperatures to 60° C when protected from light in a nitrogen atmosphere.

Based on the results of a stability study, the bulk chemical was stored at 0° ± 5° C throughout the study period. The bulk chemical was monitored periodically by the study laboratory using high-performance liquid chromatography and infrared spectroscopy. No change in the study material was detected.

PREPARATION AND ANALYSIS OF DOSE FORMULATIONS

The dose formulations were prepared by mixing appropriate amounts of quercetin and feed (Table F1). Studies were conducted by the analytical chemistry laboratory to determine the

homogeneity and stability of 10,000 ppm quercetin in feed. Homogeneity was confirmed by ultraviolet spectroscopy. Stability of dose formulations stored at temperatures up to 25° C for at least 14 days was confirmed by high-performance liquid chromatography. During the studies, the dose formulations were stored in opaque plastic bags (because of reported light sensitivity) at approximately 4° C for no longer than 2 weeks.

The study laboratory conducted periodic analyses of the quercetin dose formulations using ultraviolet spectrophotometry (Appendix F). During the 2-year studies, the dose formulations were analyzed at approximately 8-week intervals and all formulations were within 10% of the target concentrations (Table F2). Results of periodic referee analyses of the dose formulations performed by the analytical chemistry laboratory were in agreement with the results from the study laboratory (Table F3).

2-YEAR STUDIES

Study Design

Groups of 70 rats of each sex were administered 0, 1,000, 10,000, or 40,000 ppm quercetin. These doses were selected based on the literature reports which showed that quercetin administered in the diet at levels up to approximately 4% (40,000 ppm) caused a minor body weight decrement, and that this effect was more severe at doses higher than 4%. Since 1,000 ppm was the dose level used in the one study reporting carcinogenic results in rats, this concentration was selected as the low dose for these studies. Ten male and ten female rats per dose group were randomly selected and necropsied for interim evaluation after 6 months and 15 months of chemical administration.

Source and Specification of Animals

Male and female F344/N rats were obtained at 4 to 5 weeks of age from Charles River Breeding Laboratories (Portage, MI) for use in the 2-year

studies. Males were quarantined for 15 days and females were quarantined for 21 days. Five animals of each sex were randomly selected and killed for parasite evaluation and gross observation of disease. The rats were placed on study at about 7 weeks of age. The health of the animals was monitored during the course of the studies according to the protocols of the NTP Sentinel Animal Program (Appendix I).

Animal Maintenance

Rats were housed five per cage. Feed and water were available *ad libitum*. Racks were rotated in the room every 2 weeks, and cages were rotated from top to bottom within each group every 2 weeks. Further details of animal maintenance are given in Table 2.

Clinical Examinations and Pathology

All animals were observed twice daily and clinical findings were recorded weekly for 13 weeks and monthly thereafter. Rats were weighed at study initiation, once per week for 14 weeks, and once every 4 weeks thereafter. Feed consumption was measured weekly.

After 6 months, 10 male and 10 female rats from each dose group were killed for interim evaluations. An additional 10 rats from each dose group were randomly selected and killed for 15-month interim evaluations. Blood was drawn from the tails of rats to measure the following hematology parameters: erythrocytes, total leukocyte count, leukocyte differential counts, and nucleated erythrocytes. Blood collected from the jugular vein was analyzed for concentrations of blood urea nitrogen, creatinine, sodium, potassium, chloride, alanine aminotransferase, aspartate aminotransferase, and sorbitol dehydrogenase. One week prior to the 6- or 15-month interim evaluations, urine was collected over a 24-hour period, the volume was measured and the concentrations of chloride, potassium, and sodium were determined. The brain, liver, and right kidney of each animal were weighed at necropsy. Further details of the interim evaluations are presented in Table 2.

Animals found moribund, selected for the 6- or 15-month interim evaluations, or surviving to the end of the 2-year studies were killed. Necropsy was

performed on all animals. At necropsy, all organs and tissues were examined for gross lesions. All major tissues were fixed and preserved in 10% neutral buffered formalin, processed and trimmed, embedded in paraffin, sectioned, and stained with hematoxylin and eosin for microscopic examination. Histopathology examinations of the tissues were performed according to an "inverse pyramid" design (McConnell, 1983a,b). Complete histopathologic examinations were performed on all grossly visible lesions in all dose groups, on all control animals, and on animals receiving 40,000 ppm. Selected histopathology examinations were performed on 1,000 and 10,000 ppm dose group animals dying before the end of the study period. The tissues, tissue groups, and organs examined are listed in Table 2.

Upon completion of the microscopic evaluation by the laboratory pathologist, the slides, paraffin blocks, and residual wet tissues were sent to the NTP Archives for inventory, slide/block match, and wet tissue audit. The slides, individual animal data records, and pathology tables were sent to an independent pathology quality assessment laboratory. The individual animal records and pathology tables were compared for accuracy, slide and tissue counts were verified, and histotechnique was evaluated. The kidney from all male rats and the kidney, uterus, and thyroid gland from all female rats were reevaluated microscopically by a quality assessment pathologist. Additionally, the duodenum, ileum, jejunum, and glandular stomach were reviewed from all animals in all groups for pigmentation. All diagnoses of primary mammary gland tumor and squamous cell carcinoma of the tongue in females were examined. Since the urinary bladder had been affected in a previous study, 20% of the control and 40,000 ppm dose groups were randomly selected for microscopic review of the urinary bladder.

The quality assessment report and slides were submitted to the NTP Pathology Working Group (PWG) chair, who reviewed all kidneys and all segments of duodenum, ileum, jejunum and glandular stomach with a diagnosis of pigmentation. All uteri and mammary glands with a tumor diagnosis and all tongues with the diagnosis of squamous cell carcinoma from female rats were also reviewed. Representative examples of potential chemical-related nonneoplastic lesions and

neoplasms, lesions for which there was a difference in diagnosis between the study pathologist and reviewing pathologist, and lesions of general interest were selected by the chair for review by the PWG. All renal neoplasms and hyperplasias were examined by the PWG. The PWG consisted of the quality assessment pathologist and other pathologists experienced in rodent toxicologic pathology. This group examined the tissues without knowledge of dose groups or previously rendered diagnoses.

When the consensus opinion of the PWG differed from that of the laboratory pathologist, the diagnosis was changed. Thus, the final diagnoses represent a consensus of contractor pathologists and the PWG. Details of these review procedures have been described by Maronpot and Boorman (1982) and Boorman *et al.* (1985). For subsequent analysis of pathology data, the diagnosed lesions for each tissue type are evaluated separately or combined according to the guidelines of McConnell *et al.* (1986).

Statistical Methods

Survival Analyses

The probability of survival was estimated by the product-limit procedure of Kaplan and Meier (1958) and is presented in the form of graphs. Animals were censored from the survival analyses at the time they were found dead of other than natural causes; animals dying from natural causes were not censored. Statistical analyses for a possible dose-related effect on survival used the method of Cox (1972) for testing two groups for equality and Tarone's (1975) life table test to identify dose-related trends. All reported P values for the survival analyses are two sided.

Calculation of Incidence

Tables A1 and B1 summarize the incidence of neoplasms in male and female rats. Tables A5 and B5 summarize the incidence of nonneoplastic lesions in male and female rats. The incidence of neoplasms or nonneoplastic lesions is given as the ratio of the number of animals bearing such lesions at a specific anatomic site to the number of animals in which that site was examined. In most instances, the denominators include only those animals for which the site was examined histopathologically.

However, when macroscopic examination was required to detect lesions (e.g., skin or mammary gland neoplasms) prior to histologic sampling, or when lesions had multiple potential sites of occurrence (e.g., mononuclear cell leukemia), the denominators consist of the number of animals on which a necropsy was performed.

Analysis of Neoplasm Incidence

The majority of neoplasms in these studies were considered to be incidental to the cause of death or not rapidly lethal. Thus, the primary statistical method used was a logistic regression analysis, which assumed that the diagnosed neoplasms were discovered as the result of death from an unrelated cause and thus did not affect the risk of death. In this approach, neoplasm prevalence was modeled as a logistic function of chemical exposure and time. Both linear and quadratic terms in time were incorporated initially, and the quadratic term was eliminated if it did not significantly enhance the fit of the model. The dosed and control groups were compared on the basis of the likelihood score test for the regression coefficient of dose. This method of adjusting for intercurrent mortality is the prevalence analysis of Dinse and Lagakos (1983), further described and illustrated by Dinse and Haseman (1986). When neoplasms are incidental, this comparison of the time-specific neoplasm prevalence also provides a comparison of the time-specific neoplasm incidences (McKnight and Crowley, 1984).

In addition to logistic regression, alternative methods of statistical analysis were used, and the results of these tests are summarized in the appendixes. These include the life table test (Cox, 1972; Tarone, 1975), appropriate for rapidly lethal neoplasms, and the Fisher exact test and the Cochran-Armitage trend test (Armitage, 1971; Gart *et al.*, 1979), procedures based on the overall proportion of tumor-bearing animals.

Tests of significance included pairwise comparisons of each dosed group with controls and a test for an overall dose-response trend. Continuity-corrected tests were used in the analysis of tumor incidence, and reported P values are one sided. The procedures described above were also used to evaluate selected nonneoplastic lesions. (For further

discussion of these statistical methods, see Haseman, 1984).

Historical Control Data

Although the concurrent control group is always the first and most appropriate control group used for evaluation, there are certain instances in which historical control data can be helpful in the overall assessment of neoplasm incidence. Consequently, control tumor incidences from the NTP historical control database (Haseman *et al.*, 1984, 1985) are included in the NTP reports for neoplasms appearing to show compound-related effects.

Analysis of Continuous Variables

Two approaches were employed to assess the significance of pairwise comparisons between dosed and control groups in the analysis of continuous variables. Organ and body weight data, which have approximately normal distributions, were analyzed using the parametric multiple comparison procedures of Williams (1971, 1972) and Dunnett (1955). Clinical chemistry, urinalysis, and hematology data, which typically have skewed distributions, were analyzed using the nonparametric multiple comparison methods of Shirley (1977) and Dunn (1964). Jonckheere's test (Jonckheere, 1954) was used to assess the significance of dose-response trends and to determine whether a trend-sensitive test (Williams' or Shirley's test) was more appropriate for pairwise comparisons than a test

that does not assume a monotonic dose-response trend (Dunnett's or Dunn's test). Average nephropathy severity values for the 2-year studies were analyzed for significance using the Mann-Whitney U test (Hollander and Wolfe, 1973).

QUALITY ASSURANCE METHODS

The 2-year studies were conducted in compliance with FDA Good Laboratory Practice Regulations (21 CFR Part 58). In addition, as study records were submitted to the NTP Archives, they were audited retrospectively by an independent quality assurance contractor. Separate audits covering completeness and accuracy of the pathology data, pathology specimens, final pathology tables, and preliminary review draft of this NTP Technical Report were conducted. Audit procedures and findings are presented in the reports, which are on file at the NIEHS. The audit findings were reviewed and assessed by NTP staff so that all discrepancies had been resolved or were otherwise addressed during the preparation of this Technical Report.

GENETIC TOXICITY

The genetic toxicity of quercetin was assessed by testing the ability of the chemical to induce mutations in various strains of *Salmonella typhimurium* and to induce sister chromatid exchanges and chromosomal aberrations in Chinese hamster ovary cells. The protocols for these studies and tabular presentations of their findings are given in Appendix C.

TABLE 2
Experimental Design and Materials and Methods in the 2-Year Feed Studies of Quercetin

Study Laboratory

EG&G Mason Research Institute, Worcester, MA

Strain and Species

F344/N rats

Animal Source

Charles River Breeding Laboratories, Portage, MI

Date of Birth

Males: 3 - 10 May 1982

Females: 10 - 17 May 1982

Time Held Before Study

Males: 15 days

Females: 21 days

Average Age When Placed on Study

7 weeks

Date of First Dose

Males: 23 June 1982

Females: 6 July 1982

Duration of Dosing

104 weeks (7 days/week)

Date of Last Dose

Males: 15 - 21 June 1984

Females: 27 June - 6 July 1984

Necropsy Dates

6-month interim evaluation: Males: 28 - 30 December 1982; Females: 12 - 14 January 1983

15-month interim evaluation: Males: 28 - 30 September 1983; Females: 12 - 14 October 1983

2-year studies: Males: 15 - 21 June 1984; Females: 28 June - 5 July 1984

Average Age When Killed

111 weeks

Size of Study Groups

70 males and 70 females

Method of Animal Distribution

Animals of each sex randomized into cage groups, and then cages randomized to treatment groups using appropriate table of random numbers.

Animals per Cage

5

Method of Animal Identification

Ear punch

Diet

NIH-07 Rat and Mouse Ration, Open formula, pellets (Zeigler Bros., Inc., Gardners, PA), available *ad libitum*

TABLE 2
Experimental Design and Materials and Methods in the 2-Year Feed Studies of Quercetin (continued)

Feeders

Stainless steel, gang style (Scientific Cages, Inc., Bryan, TX), changed once weekly

Water

Tap water (City of Worcester Water Supply) via outside-the-cage automatic watering system (Edstrom Industries, Inc., Waterford, WI), available *ad libitum*

Cages

Solid-bottom polycarbonate (Lab Products, Inc., Rochelle Park, NJ)

Bedding

Aspen bed, heat-treated hardwood chips (American Excelsior Co., Baltimore, MD), changed twice weekly

Cage Filters

Non-woven fiber filters (Snow Filtration, Cincinnati, OH)

Animal Room Environment

Temperature: $22.5^{\circ} \pm 1.5^{\circ}$ C

Relative humidity: $47.6\% \pm 5.8\%$

Fluorescent light: 12 hours/day

Room air changes: 12/hour

Doses

0, 1,000, 10,000, or 40,000 ppm quercetin in feed

Type and Frequency of Observation

Observed twice/day; body weight initially, once/week for 14 weeks, once/month thereafter; clinical observations once/week for 13 weeks, once/month thereafter; feed consumption measured once/week.

Necropsy and Histopathology

Organ weights: Recorded for brain, right kidney, and liver of all animals sacrificed at 6 and 15 months

Necropsy: Performed on all animals

Histopathology: Complete histopathologic examinations performed on all grossly visible lesions in all dose groups and on all control and 40,000 ppm dose animals; histopathologic examinations performed on the following tissues:
 At 6 months: 10,000 ppm dose groups (large intestine, small intestine, and uterus)
 At 15 months: 1,000 ppm dose groups (large intestine); 10,000 ppm dose group (large intestine, small intestine, and stomach)
 Animals dying early and at study termination (for 1,000 and 10,000 ppm groups): (kidney, liver, pancreas, parathyroid gland, pituitary gland, small intestine, tongue, urinary bladder, and uterus)

Clinical Pathology

Blood and urine samples were collected from males and females at the 6- and 15-month interim evaluations.

Hematology: Erythrocytes, leukocytes, leukocyte differential count, and nucleated erythrocytes

Clinical chemistry: Blood urea nitrogen, creatinine, sodium, potassium, chloride, alanine aminotransferase, aspartate aminotransferase, and sorbitol dehydrogenase

Urinalysis: Urinary sodium, urinary potassium, and urinary chloride

RESULTS

2-YEAR STUDIES

6- and 15-Month Interim Evaluations

The relative kidney and liver weights of male and female rats that received 40,000 ppm were significantly greater than those of the controls at both 6 and 15 months (Tables D1 and D2). For females these differences primarily reflected the reduced body weights observed in high-dose animals. No biologically significant changes in hematology or clinical chemistry parameters were observed (Tables E1 and E2). The only abnormality noted in the urinalyses was the presence of calcium oxalate crystals in 7 of 10 high-dose males at 15 months.

Yellow-brown pigmentation occurred in several tissues and was most prevalent in the glandular stomach and the distal segments of the small intestine. The incidence and severity of pigmentation increased with dose concentration and duration. At 15 months all high-dose males had pigmentation in the glandular stomach, as did 5 of 10 high-dose females. Epithelial staining of the small intestine was present in all high-dose males and in nine high-dose females at 15 months. One high-dose male also had pigmentation in the lamina propria of the jejunum and ileum and two mid-dose females had pigmentation in the jejunal and ileal submucosa. Furthermore, at 15 months eight high-dose males and four high-dose females had pigmentation of the skulls or teeth. There were no

neoplasms or nonneoplastic lesions related to quercetin administration in male or female rats at 6 or 15 months.

Survival

Estimates of the probabilities of survival for male and female rats are shown in Table 3 and in the Kaplan-Meier survival curves in Figure 1. Exposure to quercetin had no significant effect on survival.

Body Weights, Feed Consumption, and Clinical Findings in the 2-Year Studies

Male and female rats given 40,000 ppm quercetin had lower body weight gains than those of the controls. In males, the difference was 5% at week 25, and in females, 10% at week 25 (Tables 4 and 5 and Figure 2). From 65 weeks to the end of the study, the difference among males ranged from 6% to 13%; in females, the difference ranged from 13% to 15%. Feed consumption by exposed males and females was similar to that of the controls (Tables G1 and G2). The decreased body weight gains relative to controls were attributed to quercetin toxicity. A yellowish discoloration of the hair coat, especially in the perineal area, was present in all mid- and high-dose animals, presumably due to the urinary and/or fecal excretion of quercetin and/or its metabolites.

TABLE 3
Survival of Rats in the 2-Year Feed Studies of Quercetin

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|---------|-----------|------------|------------|
| Male | | | | |
| Animals initially in study | 70 | 70 | 70 | 70 |
| 6-Month interim evaluation ^a | 10 | 10 | 10 | 10 |
| 15-Month interim evaluation ^a | 10 | 10 | 10 | 10 |
| Natural deaths | 3 | 7 | 3 | 6 |
| Moribund kills | 21 | 15 | 22 | 21 |
| Animals surviving until end of the study | 26 | 28 | 25 | 23 |
| Percent survival at end of study ^b | 52 | 56 | 50 | 47 |
| Mean survival (days) ^c | 576 | 581 | 577 | 576 |
| Survival analyses ^d | P=0.603 | P=0.838N | P=0.940 | P=0.824 |
| Female | | | | |
| Animals initially in study | 70 | 70 | 70 | 70 |
| 6-Month interim evaluation ^a | 10 | 10 | 10 | 10 |
| 15-Month interim evaluation ^a | 10 | 10 | 10 | 10 |
| Natural deaths | 1 | 4 | 2 | 3 |
| Moribund kills | 19 | 18 | 13 | 19 |
| Animals surviving until end of the study | 30 | 28 | 35 | 28 |
| Percent survival at end of study ^b | 60 | 56 | 71 | 56 |
| Mean survival (days) ^c | 590 | 574 | 586 | 576 |
| Survival analyses ^d | P=0.709 | P=0.612 | P=0.445N | P=0.656 |

^a Censored from survival analyses

^b Kaplan-Meier determinations. Survival rates adjusted for interim evaluations.

^c Mean of all deaths (uncensored, censored, terminal sacrifice).

^d The result of the life table trend test (Tarone, 1975) is in the control column, and the results of the life table pairwise comparisons (Cox, 1972) with the controls are in the dosed columns. A lower mortality in a dose group is indicated by N.

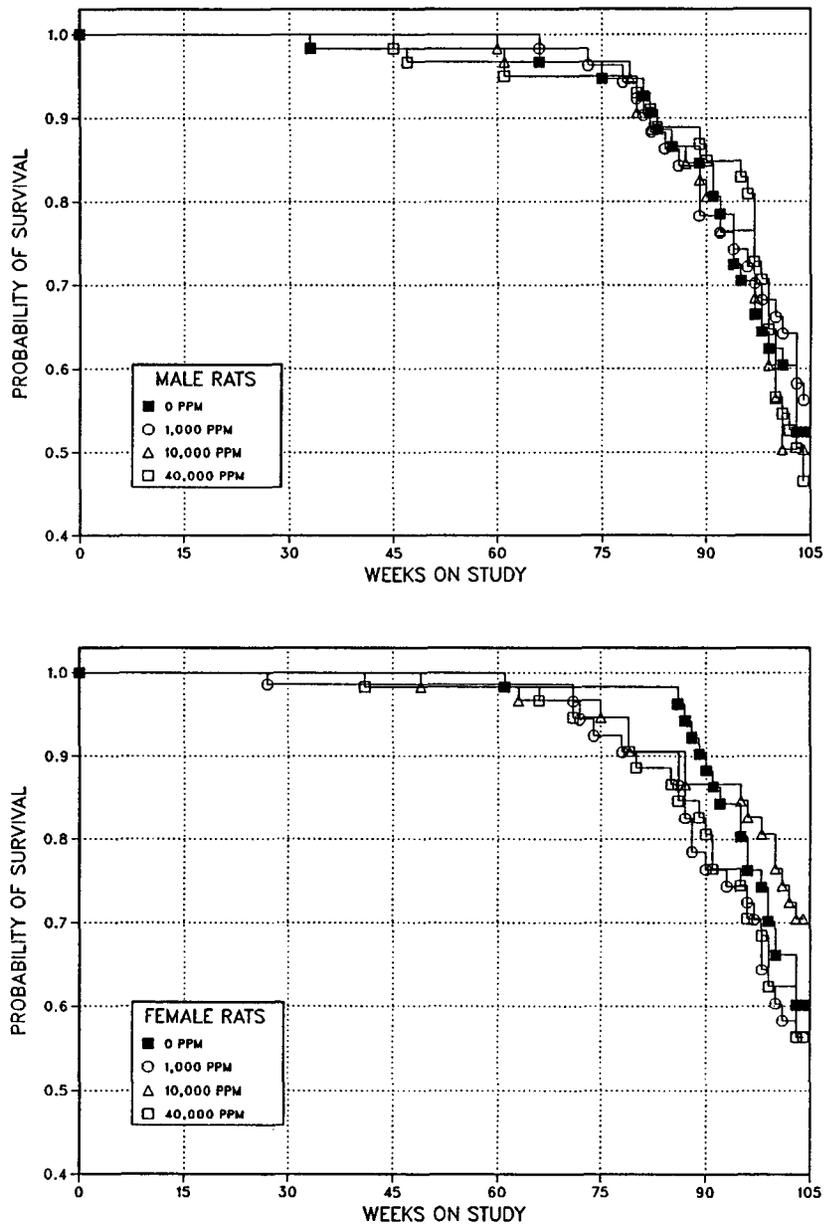


FIGURE 1
Kaplan-Meier Survival Curves for Rats Administered Quercetin in Feed for 2 Years

TABLE 4
Mean Body Weights and Survival of Male Rats in the 2-Year Feed Study of Quercetin

| Weeks on Study | 0 ppm | | 1,000 ppm | | | 10,000 ppm | | | 40,000 ppm | | |
|---------------------------|----------------|---------------------|----------------|------------------------|---------------------|----------------|------------------------|---------------------|----------------|------------------------|---------------------|
| | Av. Wt. (g) | No. of Survivors | Av. Wt. (g) | Wt. (% of controls) | No. of Survivors | Av. Wt. (g) | Wt. (% of controls) | No. of Survivors | Av. Wt. (g) | Wt. (% of controls) | No. of Survivors |
| 1 | 162 | 70 | 160 | 99 | 70 | 165 | 102 | 70 | 167 | 103 | 70 |
| 2 | 195 | 70 | 196 | 101 | 70 | 203 | 104 | 70 | 198 | 102 | 70 |
| 3 | 225 | 70 | 230 | 102 | 70 | 233 | 103 | 70 | 228 | 101 | 70 |
| 4 | 253 | 70 | 255 | 101 | 70 | 257 | 102 | 70 | 252 | 99 | 70 |
| 5 | 271 | 70 | 272 | 100 | 70 | 272 | 100 | 70 | 259 | 95 | 70 |
| 6 | 284 | 70 | 278 | 98 | 70 | 287 | 101 | 70 | 282 | 99 | 70 |
| 7 | 300 | 70 | 298 | 100 | 70 | 304 | 102 | 70 | 300 | 100 | 70 |
| 8 | 310 | 70 | 304 | 98 | 70 | 312 | 101 | 70 | 309 | 100 | 70 |
| 9 | 322 | 70 | 321 | 100 | 70 | 325 | 101 | 70 | 320 | 99 | 70 |
| 10 | 319 | 70 | 323 | 101 | 70 | 336 | 106 | 70 | 332 | 104 | 70 |
| 11 | 342 | 70 | 343 | 100 | 70 | 346 | 101 | 70 | 339 | 99 | 70 |
| 12 | 340 | 70 | 337 | 99 | 70 | 334 | 98 | 70 | 328 | 97 | 70 |
| 13 | 354 | 70 | 350 | 99 | 70 | 343 | 97 | 70 | 343 | 97 | 70 |
| 14 | 364 | 70 | 363 | 100 | 70 | 359 | 99 | 70 | 355 | 97 | 70 |
| 17 | 376 | 70 | 375 | 100 | 70 | 373 | 99 | 70 | 364 | 97 | 70 |
| 21 | 399 | 70 | 399 | 100 | 70 | 395 | 99 | 70 | 382 | 96 | 70 |
| 25 | 416 | 70 | 412 | 99 | 70 | 409 | 98 | 70 | 393 | 95 | 70 |
| 29 ^a | 434 | 60 | 438 | 101 | 60 | 430 | 99 | 60 | 413 | 95 | 60 |
| 30 | 445 | 60 | 438 | 98 | 60 | 435 | 98 | 60 | 409 | 92 | 60 |
| 33 | 456 | 60 | 456 | 100 | 60 | 448 | 98 | 60 | 426 | 93 | 60 |
| 37 | 457 | 59 | 460 | 101 | 60 | 458 | 100 | 60 | 432 | 95 | 60 |
| 41 | 464 | 59 | 466 | 101 | 60 | 464 | 100 | 60 | 439 | 95 | 60 |
| 45 | 469 | 59 | 470 | 100 | 60 | 464 | 99 | 60 | 442 | 94 | 60 |
| 49 | 481 | 59 | 486 | 101 | 60 | 482 | 100 | 60 | 453 | 94 | 58 |
| 53 | 484 | 59 | 487 | 101 | 60 | 481 | 100 | 60 | 453 | 94 | 58 |
| 57 | 487 | 59 | 491 | 101 | 60 | 488 | 100 | 60 | 460 | 95 | 58 |
| 61 | 478 | 59 | 487 | 102 | 60 | 484 | 101 | 59 | 457 | 96 | 58 |
| 65 | 485 | 59 | 491 | 101 | 60 | 483 | 100 | 58 | 453 | 94 | 57 |
| 68 ^a | 486 | 58 | 493 | 102 | 49 | 490 | 101 | 48 | 458 | 94 | 47 |
| 73 | 492 | 48 | 497 | 101 | 49 | 491 | 100 | 48 | 458 | 93 | 47 |
| 81 | 492 | 47 | 492 | 100 | 46 | 483 | 98 | 45 | 451 | 92 | 46 |
| 85 | 485 | 44 | 488 | 101 | 43 | 480 | 99 | 44 | 444 | 92 | 44 |
| 89 | 476 | 43 | 477 | 100 | 42 | 473 | 99 | 41 | 436 | 92 | 43 |
| 93 | 473 | 39 | 482 | 102 | 38 | 465 | 98 | 38 | 426 | 90 | 42 |
| 97 | 479 | 35 | 485 | 101 | 36 | 450 | 94 | 38 | 418 | 87 | 40 |
| 101 | 447 | 31 | 451 | 101 | 33 | 427 | 96 | 28 | 402 | 90 | 28 |
| 104 | 464 | 26 | 451 | 97 | 29 | 440 | 95 | 25 | 403 | 87 | 25 |
| Terminal sacrifice | | 26 | | | 29 | | | 25 | | | 25 |
| Mean for weeks | | | | | | | | | | | |
| 1-13 | 283 | | 282 | 100 | | 286 | 101 | | 281 | 100 | |
| 14-52 | 433 | | 433 | 100 | | 429 | 99 | | 410 | 95 | |
| 53-104 | 479 | | 482 | 101 | | 472 | 99 | | 440 | 92 | |

^a Interim evaluation occurred.

TABLE 5
Mean Body Weights and Survival of Female Rats in the 2-Year Feed Study of Quercetin

| Weeks on Study | 0 ppm | | 1,000 ppm | | | 10,000 ppm | | | 40,000 ppm | | |
|---------------------------|----------------|---------------------|----------------|------------------------|---------------------|----------------|------------------------|---------------------|----------------|------------------------|---------------------|
| | Av. Wt. (g) | No. of Survivors | Av. Wt. (g) | Wt. (% of controls) | No. of Survivors | Av. Wt. (g) | Wt. (% of controls) | No. of Survivors | Av. Wt. (g) | Wt. (% of controls) | No. of Survivors |
| 1 | 138 | 70 | 141 | 102 | 70 | 139 | 101 | 70 | 141 | 102 | 70 |
| 2 | 153 | 70 | 155 | 102 | 70 | 152 | 99 | 70 | 152 | 100 | 70 |
| 3 | 163 | 70 | 165 | 101 | 70 | 162 | 100 | 70 | 162 | 99 | 70 |
| 4 | 165 | 70 | 167 | 101 | 70 | 167 | 101 | 70 | 165 | 100 | 70 |
| 5 | 177 | 70 | 178 | 101 | 70 | 177 | 100 | 70 | 176 | 100 | 70 |
| 6 | 187 | 70 | 186 | 100 | 70 | 183 | 98 | 70 | 181 | 97 | 70 |
| 7 | 191 | 70 | 193 | 101 | 70 | 189 | 99 | 70 | 186 | 97 | 70 |
| 8 | 199 | 70 | 199 | 100 | 70 | 194 | 98 | 70 | 190 | 96 | 70 |
| 9 | 200 | 70 | 204 | 102 | 70 | 198 | 99 | 70 | 194 | 97 | 70 |
| 10 | 208 | 70 | 210 | 101 | 70 | 203 | 98 | 70 | 197 | 95 | 70 |
| 11 | 210 | 70 | 211 | 101 | 70 | 205 | 98 | 70 | 198 | 94 | 70 |
| 12 | 215 | 70 | 214 | 100 | 70 | 202 | 94 | 70 | 195 | 91 | 70 |
| 13 | 215 | 70 | 219 | 102 | 70 | 207 | 96 | 70 | 192 | 89 | 70 |
| 14 | 216 | 70 | 218 | 101 | 70 | 212 | 98 | 70 | 200 | 93 | 70 |
| 17 | 225 | 70 | 226 | 101 | 70 | 217 | 97 | 70 | 203 | 91 | 70 |
| 21 | 233 | 70 | 233 | 100 | 70 | 222 | 95 | 70 | 209 | 90 | 70 |
| 25 | 244 | 70 | 246 | 101 | 70 | 232 | 95 | 70 | 220 | 90 | 70 |
| 29 ^a | 255 | 60 | 257 | 101 | 60 | 237 | 93 | 60 | 220 | 86 | 60 |
| 33 | 257 | 60 | 263 | 102 | 60 | 242 | 94 | 60 | 225 | 88 | 60 |
| 37 | 268 | 60 | 276 | 103 | 60 | 252 | 94 | 60 | 231 | 86 | 60 |
| 41 | 279 | 60 | 288 | 103 | 60 | 261 | 94 | 60 | 239 | 86 | 60 |
| 45 | 292 | 60 | 299 | 102 | 60 | 273 | 94 | 60 | 246 | 84 | 59 |
| 49 | 301 | 60 | 305 | 102 | 60 | 279 | 93 | 60 | 248 | 82 | 59 |
| 53 | 311 | 60 | 317 | 102 | 60 | 290 | 93 | 59 | 256 | 83 | 59 |
| 57 | 319 | 60 | 329 | 103 | 60 | 299 | 94 | 59 | 265 | 83 | 59 |
| 61 | 327 | 60 | 337 | 103 | 60 | 310 | 95 | 59 | 277 | 85 | 59 |
| 65 | 336 | 59 | 344 | 103 | 60 | 320 | 95 | 58 | 285 | 85 | 59 |
| 69 ^a | 343 | 49 | 349 | 102 | 50 | 331 | 96 | 48 | 291 | 85 | 48 |
| 73 | 350 | 49 | 355 | 102 | 48 | 335 | 96 | 48 | 296 | 85 | 47 |
| 77 | 355 | 49 | 364 | 102 | 47 | 340 | 96 | 47 | 303 | 85 | 47 |
| 81 | 362 | 49 | 368 | 102 | 45 | 345 | 95 | 45 | 308 | 85 | 44 |
| 85 | 365 | 49 | 367 | 101 | 45 | 348 | 95 | 45 | 311 | 85 | 44 |
| 89 | 369 | 46 | 371 | 101 | 39 | 352 | 95 | 43 | 314 | 85 | 42 |
| 93 | 369 | 42 | 376 | 102 | 38 | 355 | 96 | 43 | 318 | 86 | 38 |
| 97 | 360 | 38 | 367 | 102 | 36 | 340 | 94 | 41 | 312 | 87 | 35 |
| 101 | 365 | 33 | 368 | 101 | 30 | 351 | 96 | 38 | 317 | 87 | 31 |
| 104 | 357 | 30 | 360 | 101 | 28 | 349 | 98 | 35 | 311 | 87 | 28 |
| Terminal sacrifice | | 30 | | | 28 | | | 35 | | | 28 |
| Mean for weeks | | | | | | | | | | | |
| 1-13 | 186 | | 188 | 101 | | 183 | 99 | | 179 | 97 | |
| 14-52 | 257 | | 261 | 103 | | 243 | 95 | | 224 | 88 | |
| 53-104 | 349 | | 355 | 102 | | 333 | 95 | | 297 | 85 | |

^a Interim evaluation occurred.

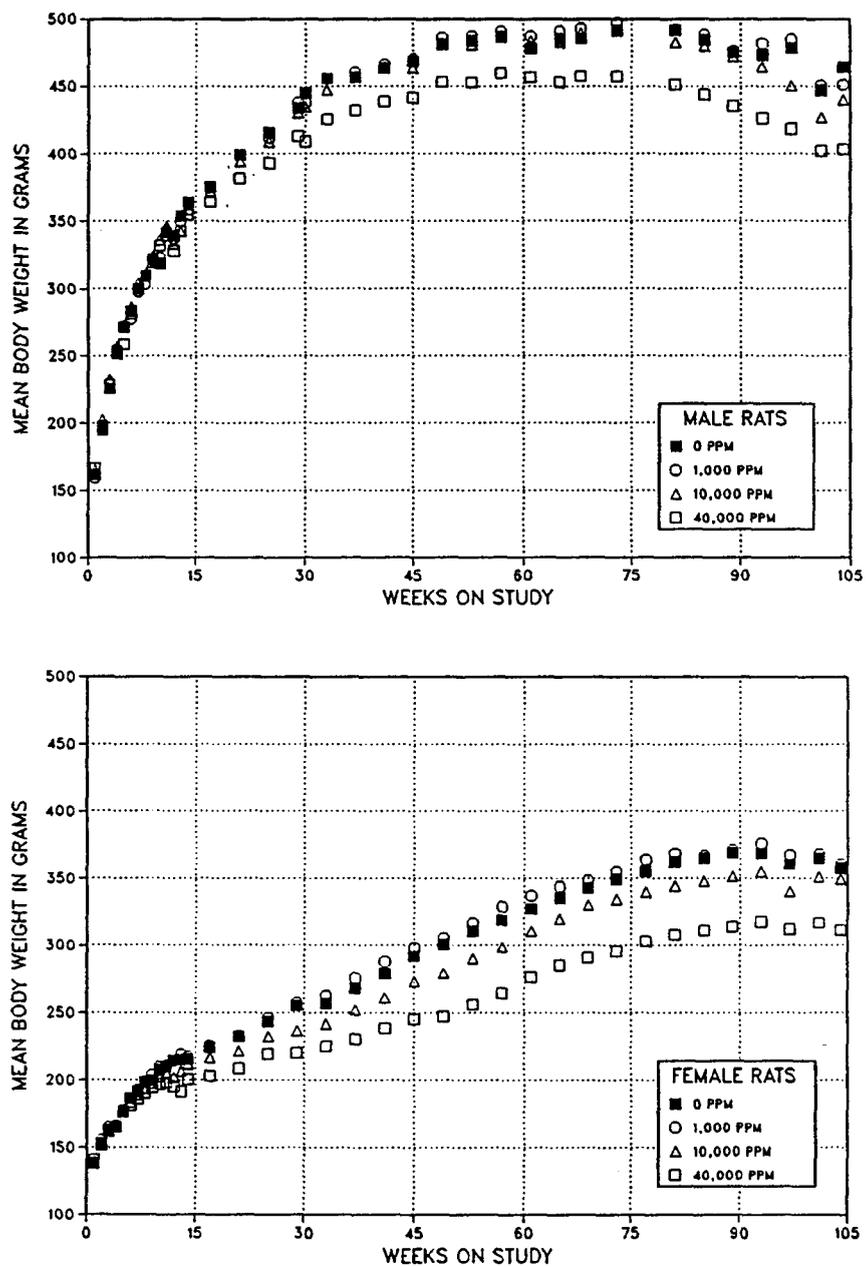


FIGURE 2
Growth Curves for Rats Administered Quercetin in Feed for 2 Years

Pathology and Statistical Analyses of Results of the 2-Year Studies

Summaries of the incidences of neoplasms and nonneoplastic lesions, individual animal tumor diagnoses, statistical analyses of primary neoplasms that occurred with an incidence of at least 5% in at least one animal group, and historical control incidences for the biologically significant neoplasms mentioned in this section are presented in Appendixes A for male rats and B for female rats.

Kidney: Initially, single sections of the left and right kidneys from each rat were examined microscopically. Renal tubule neoplasms were seen in four high-dose male rats, whereas none were observed in controls (Table 6). Three of these neoplasms were adenomas, and one was an adenocarcinoma. Renal tubule neoplasms are relatively uncommon in aged rats. The combined incidence of these neoplasms in the study laboratory historical control males for feed studies is 3/99 (3%, range 0%-6%). The combined incidence in control male rats is 8/499 (1.6%, range 0%-6%) in all NTP feed studies (Table A4). Additionally, there was a slight dose-related increase in the incidence of renal tubule hyperplasia in males.

Because of the low number of neoplasms in the high-dose males, the residual halves of the formalin-fixed kidneys from all males and females were step sectioned to provide approximately eight additional sections per rat for microscopic examination.

During this reevaluation, renal tubule focal hyperplasia was observed in eight high-dose males (one of these animals had been identified in the initial evaluation), and renal tubule adenomas were observed in six high-dose males (one of these animals had been identified in the initial evaluation) (Table 6). Focal hyperplasia was seen in two additional control males and a renal tubule adenoma was observed in one control male. The increased incidences of renal tubule hyperplasia and renal tubule neoplasms in high-dose males is supportive of some evidence of carcinogenicity.

In the initial evaluation, a renal tubule adenoma was seen in one mid-dose female rat, and an adenoma was found in one control female during the evaluation of the step sections. Thus, there was no evidence of a chemical-related increased incidence in kidney neoplasms in females (Table 7).

Renal tubule cell hyperplasias in male rats were focal lesions characterized by increased numbers of tubule epithelial cells forming multiple layers which partially or totally filled the lumen and usually caused slight tubule dilation (Plate 1). The appearance of the hyperplastic cells ranged from those of normal tubule epithelial cells to enlarged polygonal cells resembling cells of the adenomas (Plate 2).

In general, the adenomas were small (400-800 μm) and were distinguished from tubule hyperplasia by larger size and lack of a definite tubular structure. Many adenomas had a prominent microtubular pattern (Plates 3 and 4). Adenomas were expansile and frequently compressed surrounding parenchyma (Plate 5). The neoplasms consisted of large polygonal cells with abundant eosinophilic cytoplasm and large, pale-staining nuclei. The adenocarcinoma was 0.7 cm in diameter, expansile, and was composed of variably sized tubule-like structures which were filled with cells and often contained necrotic centers (Plate 6). Adenocarcinoma cells were clearly more anaplastic and were often characterized by marked pleomorphism, large nuclei, large nucleoli, and atypical mitotic figures (Plate 7).

The nephropathy was significantly more severe in male rats receiving 40,000 ppm than in the controls (Table 8). There was no significant increase in the severity of nephropathy in dosed female rats. Nephropathy was typical of the spontaneously occurring kidney lesion in aging F344/N rats. Severity grades were based upon the extent of nephropathy and the amount of renal parenchyma affected. Nephropathy consisted of a spectrum of lesions, including varying degrees of tubule dilation, distortion with occasional cyst formation, proteinaceous casts, atrophy, regeneration and hypertrophy of tubule epithelium, thickening of tubular and glomerular basement membranes, interstitial fibrosis, scattered foci of suppurative inflammation (primarily within degenerating tubules), and a scattering of varying numbers and aggregates of mononuclear inflammatory cells within the interstitium. Regenerating tubule epithelial cells had basophilic nuclei, scant cytoplasm, and usually formed a single cell layer. There was also a dose-related increase in the incidence of renal pelvic transitional epithelial hyperplasia in males. This change is associated with severe nephropathy.

TABLE 6
Selected Kidney Lesions in Male Rats in the 2-Year Feed Study of Quercetin

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|----------------|-----------|------------|-------------|
| Initial Evaluation (Single Sections) | | | | |
| Renal Tubule: Hyperplasia | | | | |
| Overall rates ^a | 1/50 (2%) | 2/50 (4%) | 3/50 (6%) | 4/50 (8%) |
| Logistic regression ^b | P=0.079 | P=0.752N | P=0.492 | P=0.182 |
| Renal Tubule: Adenoma | | | | |
| Overall rates | 0/50 (0%) | 0/50 (0%) | 0/50 (0%) | 3/50 (6%) |
| Adjusted rates ^c | 0.0% | 0.0% | 0.0% | 11.1% |
| Terminal rates ^d | 0/26 (0%) | 0/28 (0%) | 0/25 (0%) | 2/23 (9%) |
| First incidence (days) | - ^e | - | - | 676 |
| Logistic regression | P=0.042 | - | - | P=0.122 |
| Renal Tubule: Adenocarcinoma | | | | |
| Overall rates | 0/50 (0%) | 0/50 (0%) | 0/50 (0%) | 1/50 (2%) |
| Renal Tubule: Adenoma or Adenocarcinoma ^f | | | | |
| Overall rates | 0/50 (0%) | 0/50 (0%) | 0/50 (0%) | 4/50 (8%) |
| Adjusted rates | 0.0% | 0.0% | 0.0% | 15.3% |
| Terminal rates | 0/26 (0%) | 0/28 (0%) | 0/25 (0%) | 3/23 (13%) |
| First incidence (days) | - | - | - | 676 |
| Logistic regression | P=0.002 | - | - | P=0.064 |
| Evaluation of Step Sections | | | | |
| Renal Tubule: Hyperplasia | | | | |
| Overall rates | 2/50 (4%) | 2/50 (4%) | 6/50 (12%) | 8/50 (16%) |
| Renal Tubule: Adenoma | | | | |
| Overall rates | 1/50 (2%) | 2/50 (4%) | 7/50 (14%) | 6/50 (12%) |
| Single and Step Sections Combined | | | | |
| Renal Tubule: Hyperplasia | | | | |
| Overall rates | 3/50 (6%) | 3/50 (6%) | 8/50 (16%) | 11/50 (22%) |
| Logistic regression | P=0.006 | P=0.655N | P=0.099 | P=0.022 |
| Renal Tubule: Adenoma | | | | |
| Overall rates | 1/50 (2%) | 2/50 (4%) | 7/50 (14%) | 8/50 (16%) |
| Logistic regression | P=0.012 | P=0.526 | P=0.032 | P=0.018 |
| Renal Tubule: Adenoma or Adenocarcinoma | | | | |
| Overall rates | 1/50 (2%) | 2/50 (4%) | 7/50 (14%) | 9/50 (18%) |
| Logistic regression | P=0.005 | P=0.526 | P=0.032 | P=0.010 |

^a Number of lesion-bearing animals/number of animals examined at site

^b Beneath the control incidence are the P values associated with the trend test. Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the controls and that dosed group. The logistic regression tests regard these lesions as nonfatal. A lower incidence in a dose group is indicated by N.

^c Kaplan-Meier estimated neoplasm incidence at the end of the study after adjustment for intercurrent mortality

^d Observed incidence at terminal kill

^e Not applicable; no neoplasms in animal group

^f Historical incidence for 2-year NTP feed studies with untreated control groups (mean \pm standard deviation): 4/499 (0.8% \pm 1.1%, range 0%-4%)

TABLE 7
Selected Kidney Lesions in Female Rats in the 2-Year Feed Study of Quercetin^a

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|-----------|-----------|------------|------------|
| Initial Evaluation (Single Sections) | | | | |
| Renal Tubule: Hyperplasia Overall rates | 1/49 (2%) | 1/49 (2%) | 3/50 (6%) | 1/50 (2%) |
| Renal Tubule: Adenoma ^b Overall rates | 0/49 (0%) | 0/49 (0%) | 1/50 (2%) | 0/50 (0%) |
| Evaluation of Step Sections | | | | |
| Renal Tubule: Hyperplasia Overall rates | 1/49 (2%) | - | - | 3/50 (6%) |
| Renal Tubule: Adenoma Overall rates | 1/49 (2%) | - | - | 0/50 (0%) |
| Single and Step Sections Combined | | | | |
| Renal Tubule: Hyperplasia Overall rates | 2/49 (4%) | - | - | 4/50 (8%) |
| Renal Tubule: Adenoma Overall rates | 1/49 (2%) | - | - | 0/50 (0%) |

^a Step sections were not evaluated in the 1,000 ppm and 10,000 ppm dose groups.

^b Historical incidence for 2-year NTP feed studies of untreated control groups (mean \pm standard deviation): 1/499 (0.2% \pm 0.6%), range 0%-2%

TABLE 8
Incidences and Severity of Nephropathy in Male and Female Rats in the 2-Year Feed Studies of Quercetin^a

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|------------------------|----------------|----------------|----------------|------------------|
| Male | | | | |
| Minimal (grade 1) | 1/48 | 2/50 | 1/50 | 2/49 |
| Mild (grade 2) | 18/48 | 19/50 | 13/50 | 7/49 |
| Moderate (grade 3) | 19/48 | 20/50 | 23/50 | 16/49 |
| Marked (grade 4) | 10/48 | 9/50 | 13/50 | 24/49 |
| Average severity grade | 2.7 \pm 0.14 | 2.7 \pm 0.11 | 3.0 \pm 0.11 | 3.2 \pm 0.14** |
| Female | | | | |
| Minimal (grade 1) | 12/48 | 7/48 | 9/50 | 8/48 |
| Mild (grade 2) | 20/48 | 25/48 | 30/50 | 21/48 |
| Moderate (grade 3) | 13/48 | 12/48 | 10/50 | 14/48 |
| Marked (grade 4) | 3/48 | 4/48 | 1/50 | 5/48 |
| Average severity grade | 2.1 \pm 0.13 | 2.2 \pm 0.12 | 2.1 \pm 0.10 | 2.2 \pm 0.14 |

** Statistically significant ($P \leq 0.01$) from the control group by the Mann-Whitney U test

^a Number of animals with severity grade/number of animals with nephropathy. Severity grade was based on the percentage of parenchyma involved: Minimal - usually less than 25% of cortex; mild - 25% to 50% of cortex; moderate - 50% to 75% of cortex; marked - greater than 75% of cortex. Average severity grade given as the mean \pm standard error.

Parathyroid gland: Hyperplasia of the parathyroid glands, characterized by bilateral diffuse enlargement of the glands, occurred with a dose-related increased incidence in male rats (0 ppm, 1/43; 1,000 ppm, 6/45; 10,000 ppm, 6/43; 40,000 ppm, 17/43) (Table A5). Typically, the hyperplasias occurred with greater frequency and severity in animals with marked nephropathy. This is characteristic of renal secondary hyperparathyroidism.

Tongue: A single squamous cell papilloma of the tongue was present in a high-dose male (Table A1). Two squamous cell carcinomas were present in high-dose females (Table B1). The historical control incidence of oral cavity neoplasms for female rats is 4/500 (0.8%, range 0%-2%) for all NTP studies (Table B4b).

Mammary gland: Fibroadenomas of the mammary gland occurred with a highly significant, dose-related, negative trend in female rats (Table 9). The incidences of fibroadenoma in the mid- and high-dose female groups were significantly lower than that in the controls. Fibroadenoma is the most common neoplasm of the mammary gland in female rats, occurring in 178/500 (35.6%, range 8%-56%) of NTP untreated historical controls (Table B4c). Although the incidence of fibroadenoma in the controls of this study (58%) slightly exceeds the range for historical controls, the incidence in the high-dose group is about one-half of the mean rate of historical controls. The lower number of female rats with fibroadenomas in the high-dose group is considered chemically related, and may be associated with the lower body weights in this group.

Uterus: Uterine stromal polyps occurred more frequently in mid-dose female rats than in controls (7/50, 9/50, 16/50, 11/50) (Table B1). The incidence of stromal polyps in the high-dose group, however, was similar to controls. A uterine stromal sarcoma occurred in one female rat in the mid-dose group as well. Due to the marginal increased incidence in stromal polyps, the lack of a dose response, and because only the incidence in the mid-dose group exceeded the range in untreated NTP historical controls (Table B4d), these neoplasms were not considered related to quercetin administration.

Gastrointestinal tract: There was a significant dose-related accumulation of a fine granular yellow to light brown pigment in the epithelial cells lining the glandular stomach, jejunum, ileum, and, to a lesser extent, the duodenum and colon (Table 10). Special stains to further characterize the pigment were not used, but the pigment was believed to be quercetin or one of its metabolites.

Other organs/tissues: Other nonneoplastic lesions occurred with statistically significant increased incidence, but the biological significance of their occurrence is uncertain. High-dose female rats had a marginally increased incidence of chronic inflammation involving the liver (Table B5). However, all male groups had higher incidence than the high-dose females. High-dose male rats had a reduced incidence of bile duct hyperplasia, an increased incidence of lymphatic ectasia within mesenteric lymph nodes, and a marginal dose-related increased incidence in testicular interstitial cell hyperplasia (Table A5).

TABLE 9
Mammary Gland Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|-------------|-------------|-------------|------------|
| Fibroadenoma^a | | | | |
| Overall rates ^b | 29/50 (58%) | 27/50 (54%) | 16/50 (32%) | 9/50 (18%) |
| Adjusted rates ^c | 66.4% | 72.3% | 38.4% | 30.1% |
| Terminal rates ^d | 16/30 (53%) | 18/28 (64%) | 10/35 (29%) | 8/28 (29%) |
| First incidence (days) | 597 | 597 | 605 | 549 |
| Logistic regression tests ^e | P<0.001N | P=0.553N | P=0.008N | P<0.001N |

^a Historical incidence for 2-year NTP feed studies of untreated control groups (mean \pm standard deviation): 178/500 (35.6% \pm 15.0%), range 8%-56%

^b Number of neoplasm-bearing animals/number of animals necropsied

^c Kaplan-Meier estimated neoplasm incidence at the end of the study after adjustment for intercurrent mortality

^d Observed incidence at terminal kill

^e Beneath the control incidence are the P values associated with the trend test. Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the controls and that dosed group. The logistic regression tests regard these lesions as nonfatal. For all tests, a negative trend or a lower incidence in a dose group is indicated by N.

TABLE 10
Gastrointestinal Pigmentation in Rats in the 2-Year Feed Studies of Quercetin^a

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--------------------------|-------|-----------|------------|------------|
| Male | | | | |
| Glandular Stomach | | | | |
| Epithelium | 0/50 | 0/50 | 3/49 | 34/48** |
| Large Intestine | | | | |
| Colon, epithelium | 0/49 | 0/12 | 0/11 | 1/47 |
| Small Intestine | | | | |
| Duodenum, epithelium | 0/48 | 0/47 | 0/48 | 3/46 |
| Ilium, epithelium | 0/47 | 1/47 | 15/48** | 28/45** |
| Jejunum, epithelium | 0/48 | 0/44 | 2/42 | 19/44** |
| Female | | | | |
| Glandular Stomach | | | | |
| Epithelium | 0/50 | 0/49 | 8/50** | 38/50** |
| Small Intestine | | | | |
| Duodenum, epithelium | 0/50 | 0/48 | 0/50 | 1/49 |
| Ileum, epithelium | 0/49 | 0/48 | 19/49** | 32/49** |
| Jejunum, epithelium | 0/50 | 0/47 | 3/49 | 20/49** |

** Significantly different ($P \leq 0.01$) from the control group by the logistic regression test

^a Number of lesion-bearing animals/number of tissues examined

GENETIC TOXICOLOGY

Exposure to quercetin (0.3-1,000 $\mu\text{g}/\text{plate}$) produced a strong, dose-related increase in gene mutations in *Salmonella typhimurium* strains TA100 and TA98 both in the presence and in the absence of Aroclor 1254-induced male Sprague-Dawley rat or Syrian hamster liver S9 (Table C1). In cytogenetic tests with Chinese hamster ovary cells, quercetin induced marked increases in both sister chromatid exchanges and chromosomal aberrations, with and without metabolic activation (Tables C2 and C3). In the sister chromatid exchange test without S9, positive

responses were observed over a dose range of 0.67 to 20 $\mu\text{g}/\text{mL}$ quercetin; with S9, effective doses ranged from 2 to 45 $\mu\text{g}/\text{mL}$. In the chromosomal aberration test, the trials conducted in the absence of S9 activation employed a delayed harvest protocol to offset quercetin toxicity; positive responses occurred with 10 to 50 $\mu\text{g}/\text{mL}$ quercetin. With S9, standard harvest times were employed and strong increases in aberrations were observed with 25 to 75 $\mu\text{g}/\text{mL}$ quercetin. At the highest dose (75 $\mu\text{g}/\text{mL}$), all cells scored contained aberrations.

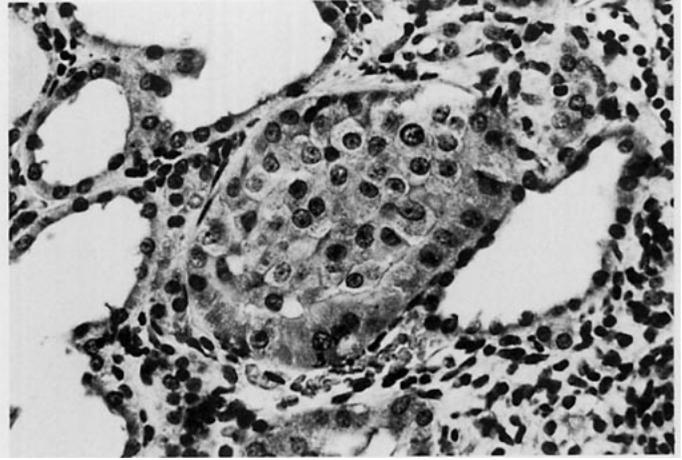
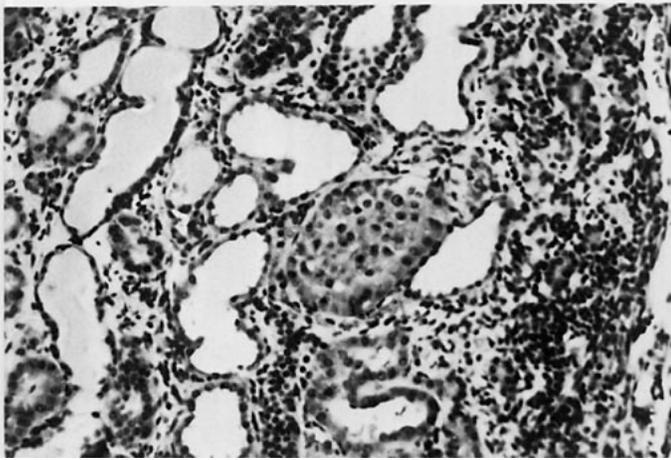


PLATE 1

Renal tubule hyperplasia with hyperplastic cells filling the lumen in the kidney of a male F344/N rat administered 40,000 ppm quercetin in feed for 2 years. H&E, 50X.

PLATE 2

Higher magnification of Plate 1 demonstrating cellular morphology typical of hyperplasia. H&E, 100X.

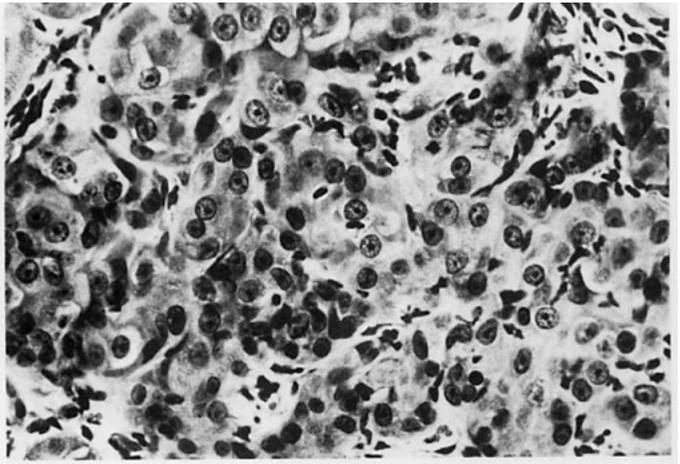


PLATE 3

Renal tubule adenoma with microtubular pattern in the kidney of a male F344/N rat administered 40,000 ppm quercetin in feed for 2 years. H&E, 50X.

PLATE 4

Higher magnification of Plate 3 demonstrating cellular morphology typical of the adenomas. H&E, 100X.

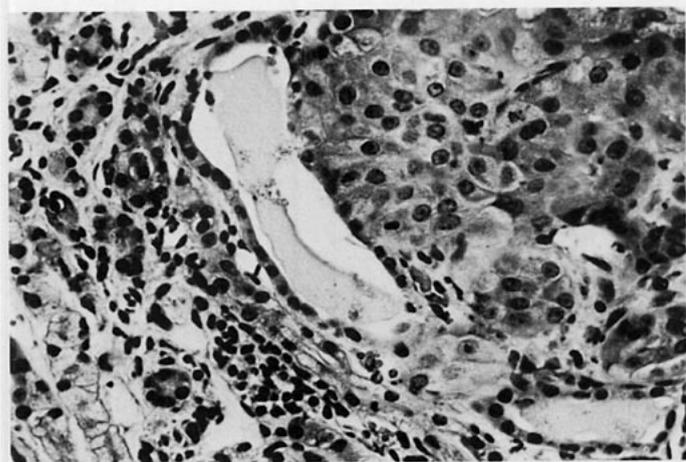


PLATE 5

Expansile renal tubule adenoma with compression of an adjacent tubule in the kidney of a male F344/N rat administered 40,000 ppm quercetin in feed for 2 years. H&E, 100X.

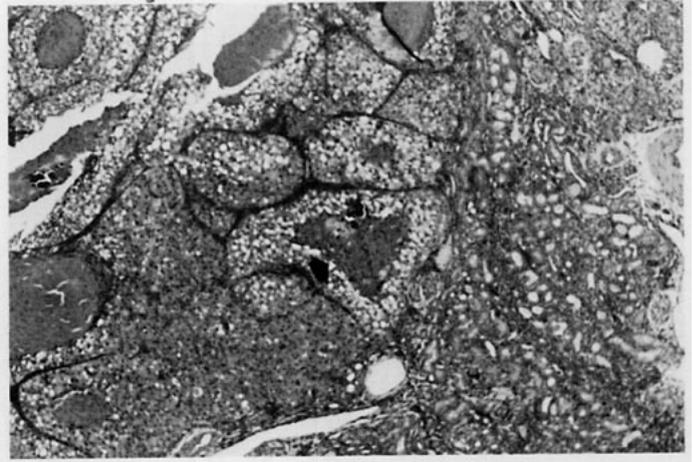


PLATE 6

Renal tubule adenocarcinoma with variably sized tubular structures in the kidney of a male F344/N rat administered 40,000 ppm quercetin in feed for 2 years. Tubular structures often had necrotic centers. H&E, 10X.

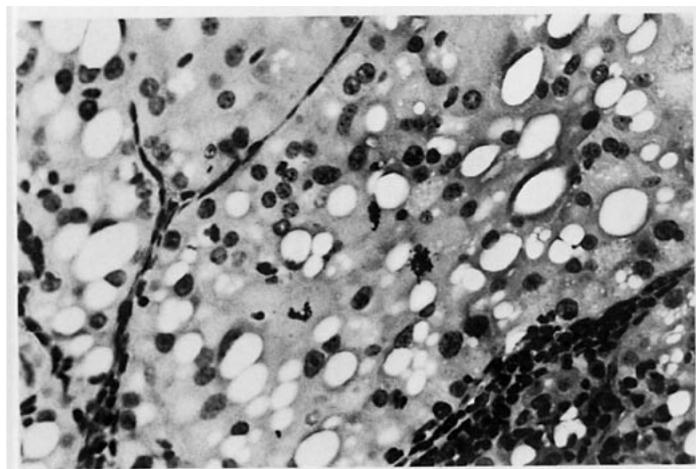


PLATE 7

Higher magnification of Plate 6 demonstrating cellular anaplasia and atypical mitotic figures. H&E, 100X.

DISCUSSION AND CONCLUSIONS

Quercetin, a flavonoid, is found in many food plants including citrus fruits, berries, leafy vegetables, roots, herbs and spices, legumes, cereal grains, tea, and cocoa (Brown, 1980). The flavonoids as a group are reported to have a wide range of possible uses in medicine. To date, though, there have been no reported controlled clinical trials or toxicity testing of these compounds to demonstrate efficacy as antiviral or anticancer agents. Thus, flavonoids are not approved for drug use in the United States (Havsteen, 1983; Cody, 1988; Gilman *et al.*, 1990). Interest in the toxicity and carcinogenicity of the flavonoids began in the mid-1970's when it was shown that some of these naturally occurring chemicals were mutagenic in the *Salmonella typhimurium* assay system. Quercetin is one of the most common flavonoids in plants and is also a component of bracken fern, a plant shown to cause toxicity and death in cattle. Quercetin was nominated by the Food and Drug Administration for toxicity and carcinogenicity studies in the rat because it is widely distributed in natural foods and because of conflicting information in the literature regarding the carcinogenicity of quercetin in previous animal studies.

Previous studies have shown that quercetin administered in feed to rodents at levels up to 5% (the maximum dose usually used in feed studies without adverse effects on nutrition) caused little organ toxicity and had no effect on mortality. Rutin, a glycoside conjugate of quercetin, has also been tested and found to be negative for carcinogenicity when fed in the diet to rats for up to 850 days (Hirono *et al.*, 1981). However, long-term administration of quercetin in feed has resulted in gradually decreased body weights of dosed animals. Hirono *et al.* (1981) observed that male rats fed 5% quercetin in the diet for more than 100 days had an approximate 15% decrease in body weight from that of the control animals. Ito *et al.* (1989) reported that F344/DuCrj rats fed 1.25% or 5.0% quercetin in CRF-1 diet for 104 weeks had final body weights that were 91% and 93% of those of the controls for males and females.

In the present NTP studies, quercetin administered at 4% (40,000 ppm) in the diet also caused a gradual reduction in body weight gain; final mean body weights of male and female rats were 87% of those of the controls. The depressed weight gain observed in the high-dose rats in these studies demonstrates that the doses were sufficient to elicit general toxicity. The high dose (4%) also approached the maximum level of quercetin that could be given in the diet (5%) without adverse nutritional effects. Experience with 2-year rodent studies has shown that control and treated animals must have body weight differences within approximately 10% to 15% to maintain similar rates of background tumors. Thus, these 2-year studies of quercetin were considered adequate for assessing toxicity and carcinogenic activity because the differences in body weights between the high-dose and control groups were within this limit.

The principal lesions associated with the administration of quercetin occurred in the kidney of dosed male rats and included severe chronic nephropathy, renal tubule hyperplasia, and renal tubule adenomas. Chronic nephropathy, a common condition in aging rats, showed a treatment-related increased severity. Hyperplasia of the renal pelvic epithelium (transitional epithelium overlying the renal papilla) is a component of severe nephropathy and occurred in males with a similar treatment-related positive trend. This transitional hyperplasia is characteristic of chemical-related toxicity and is not considered to be a preneoplastic lesion. The dose-related increased incidence of parathyroid hyperplasia in male rats is an indication that nephropathy was severe enough to compromise renal function. Hyperparathyroidism frequently accompanies severe nephropathy in rats because the progressive loss of renal function disrupts calcium and phosphorus homeostasis, which leads to prolonged parathyroid gland stimulation. This results in hyperplasia and elevated levels of parathyroid hormone. An associated mineralization was present in vascular walls and several organs. Severe nephropathy induced by chemical exposure may be life-threatening and has been the cause of

reduced survival among chemical-exposed rats in several NTP 2-year studies. However, the survival rates of exposed rats in the quercetin studies were similar to that of the controls.

The treatment-related increase in the severity of nephropathy was seen only in males. This greater sensitivity to quercetin toxicity is apparently due to a greater susceptibility of male rats to spontaneous nephropathy during aging and the exacerbation of this disease by chemical administration. Changes in glomerular permeability, resulting in proteinuria, progressive glomerular sclerosis, tubule damage, inflammation, and interstitial fibrosis, are associated with the process of aging in rats.

One factor that may contribute in part to the greater severity of tubule damage in male rats than in female rats is their production of more α_{2u} -globulin, a low molecular weight protein. Vandoren *et al.* (1983) showed that female rats excrete less than 1% of the amount of α_{2u} -globulin excreted by male rats. Further, in males, approximately 60% of the α_{2u} -globulin is reabsorbed by epithelial cells of the proximal convoluted tubule, primarily in the P₂ segment, where it is slowly or poorly hydrolyzed and accumulates in lysosomes (Charbonneau *et al.*, 1988). Short *et al.* (1987) showed that cells containing the α_{2u} -globulin undergo degeneration, necrosis, and a higher rate of cell turnover in the P₂ segment compared with other segments of the proximal convoluted tubule.

Histopathologically, the cell turnover is normally cellular regeneration, but under conditions not fully understood, hyperplasia results. Several authors suggest that this hyperplastic response of renal tubules in spontaneous nephropathy may be similar to those of tubules responding to chemical toxins. Konishi and Ward (1989) observed increased ³H-thymidine labeling indices in the tubule epithelium with corresponding increased severity of nephropathy. Short *et al.* (1987) also showed increased cell necrosis and regeneration associated with an accumulation of α_{2u} -globulin induced by chemical administration.

In the initial evaluation of single sections from each left and right kidney, three renal tubule adenomas and one adenocarcinoma were seen in the high-dose male rats with a concomitant slight dose-

related increase in the incidence of renal tubule hyperplasia. Although the incidence of renal neoplasms in the high-dose group was not significantly greater than in the controls by pairwise comparisons, the trend test was significant. Even though renal neoplasms are relatively uncommon in NTP untreated historical control male rats (8/499, mean 1.6%, range 0%-6%; Table A4a), the low number of neoplasms was difficult to interpret.

The NTP and Kurokawa *et al.* (1983) have found that multiple sectioning of the kidney may enable a more precise evaluation of the potential chemical-related induction of renal tubule neoplasms compared with observations from single-section sampling. The majority of renal neoplasms in these studies are microscopic (i.e., not observed by macroscopic examination at necropsy), thus, multiple sections might be expected to increase the number of neoplasms observed and allow for a more rigorous statistical evaluation. The residual halves of the formalin-fixed kidneys from all the rats were step sectioned to provide approximately eight additional tissue sections for microscopic examination. Renal tubule focal hyperplasia was observed in eight high-dose males (one of these animals had been identified in the initial evaluation), and renal tubule adenomas were observed in six high-dose males (one of these animals had been identified in the initial evaluation). Focal renal tubule hyperplasia was seen in two additional control males and a renal tubule adenoma was observed in one control male.

The renal tubule hyperplasia observed in these studies was distinguished from background regenerative hyperplasia, which commonly accompanies the degenerative tubule changes of age-related or chemical-induced nephropathy, on the basis of cellular atypia and prominent stratification of the epithelium. These cytological features suggest a loss of cell growth regulation and failure of cellular differentiation. This lesion is similar to those induced by potent renal carcinogens and appears to represent the early stages of renal tubule adenoma and adenocarcinoma development (Hard, 1986; Tsuda *et al.*, 1986). Although focal hyperplasia, adenoma, and adenocarcinoma constitute a morphological continuum, the rates of possible progression or regression of hyperplasia or adenoma are not known and likely vary with the inducing agent and the

mechanism of induction. It has been postulated that increases in cellular proliferation secondary to chemical-related cytotoxicity may create the appropriate environment for the development of neoplasia, perhaps by increasing the frequency of spontaneous mutations through clonal expansion of initiated cells or by other means (Farber, 1980; Pitot and Sirica, 1980; Stott *et al.*, 1981; Butterworth, 1989; Cohen and Ellwein, 1990). An increase in chemical-related accumulation of α_2 -globulin in the P₂ segment of the nephron has been associated with the development of renal neoplasms (Goldsworthy and Popp, 1987; Goldsworthy *et al.*, 1988). This syndrome, also known as hyaline droplet nephropathy or α_2 -globulin nephropathy, is best identified in 13-week studies; such studies were not conducted by the NTP with quercetin. However, the linear tubule mineralization within the papilla, which is commonly seen in the 2-year studies of chemicals producing this syndrome, was not seen in the quercetin studies.

Previous rodent toxicity and carcinogenicity studies of quercetin did not identify the kidney as a target organ (Saito *et al.*, 1980; Hirono *et al.*, 1981; Takanashi *et al.*, 1983; Ito *et al.*, 1989). However, in the NTP 2-year studies of quercetin, the renal tubule cell neoplasms observed in male rats were judged to show some evidence for carcinogenicity due to supportive evidence for this neoplastic response by an increase in renal tubule hyperplasia. Step-sectioned kidneys showed an increase in the incidence of kidney neoplasms, which supported the original findings of only a few neoplasms at this site. Since most of the neoplasms were adenomas, this effect was judged to be some evidence rather than clear evidence for carcinogenic activity.

In a series of NCI/NTP long-term rodent carcinogenicity studies of chemicals, treatment-related kidney neoplasms were found more frequently in male rats (23) than in female rats (8), male mice (3), or female mice (1) (Table 11). Based on this information, the kidneys of male rats appear to be more sensitive to chemical-induced formation than are the kidneys of female rats or mice of either sex. The reasons for the susceptibility of the male rat kidney to chemical toxicity or carcinogenicity may vary from chemical to chemical. There is no one particular chemical structure that is associated with the induction of kidney neoplasms in male rats and

some of the chemicals causing kidney tumors demonstrate genetic toxicity in *in vitro* tests while others do not. An increase in chemical concentration in the kidney of male F344/N rats, an animal with a significant age-related background of kidney disease, may make this animal particularly susceptible to the induction of renal tubule cell neoplasms.

The two squamous cell carcinomas of the tongue observed in the high-dose female rats were not considered to be related to treatment. Squamous cell papillomas or carcinomas have been observed sporadically in NTP study animals in both treated and control groups, occurring with a historical mean incidence of 0.8% in untreated controls (4/500, range 0%-2%; Table B4b). Due to the occurrence of these oral cavity neoplasms, a complete histopathologic examination was performed on the tongue. There was no supportive evidence for hyperplasia or other neoplasms at this site. Chemicals which have been shown to cause oral cavity neoplasms, such as benzidine congeners or dyes, are generally characterized as potent genotoxic agents and cause neoplasms at a variety of other sites. In summary, the oral cavity neoplasms observed in the high-dose female rats were not considered to be related to chemical administration because of the low incidence, lack of supportive nonneoplastic lesions at this site, and lack of supportive evidence of neoplasms in related tissues of epidermal origin.

There was a dose-related decrease in mammary gland fibroadenomas in female rats. Previous studies showed that decreases in some naturally occurring benign neoplasms, especially neoplasms of the mammary gland and reproductive organs, are associated with decreased body weight relative to controls (Rao *et al.*, 1987). Some investigators have reported that quercetin may have some undefined antitumor activity (Hirose *et al.*, 1983; Kato *et al.*, 1983; Nishino *et al.*, 1984b).

While most rat studies of quercetin have shown no evidence of carcinogenicity, Pamukcu *et al.* (1980) reported that 0.1% quercetin administered in the diet to Norwegian rats for 58 weeks caused an 80% increase in the incidence of intestinal neoplasms and a 20% increase in the incidence of urinary bladder neoplasms. The mechanism for this increase could not be explained based on the information given but

may be due to different experimental conditions, study animals, or chemical preparations. The NTP quercetin studies showed no evidence of chemical-induced neoplasms of the urinary bladder or intestine.

Quercetin has a marked ability to cause mutations in various genetic toxicity tests including the *Salmonella typhimurium* assay systems. Due to these results, the chemical might be expected to cause neoplasms at a variety of sites in male rats besides the kidney. The mutagenic flavonoids generally contain a free hydroxy group at the 3 position. Brown and Griffiths (1983) have shown that rats are capable of metabolizing quercetin and other 3-hydroxyl flavonoids to the 3'-*o*-methyl ethers. The 3'-*o*-methyl ether of quercetin is considerably

less mutagenic in the *Salmonella* assay than the parent compound (MacGregor and Jurd, 1978). This ability of rats to form 3'-*o*-methyl ethers may be important in protecting against the carcinogenic action of quercetin at various sites in the body.

CONCLUSIONS

Under the conditions of these 2-year feed studies there was *some evidence of carcinogenic activity** of quercetin in male F344/N rats based on an increased incidence of renal tubule cell adenomas. There was *no evidence of carcinogenic activity* of quercetin in female F344/N rats receiving 1,000, 10,000 or 40,000 ppm. Incidence of renal tubule hyperplasia and the severity of nephropathy were increased in exposed male rats.

* Explanation of Levels of Evidence of Carcinogenic Activity is on page 8. A summary of peer review comments and the public discussion on this Technical Report appear on page 10.

TABLE 11
Evidence of Kidney Neoplasms and *Salmonella* Mutagenicity in Rats and Mice
for Selected Chemicals Tested by the National Toxicology Program

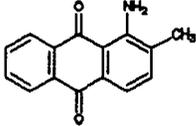
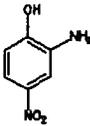
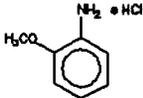
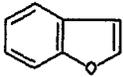
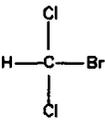
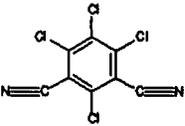
| Chemical Name and Structure | Technical Report Number | Kidney Neoplasms ^a | | | | NTP <i>Salmonella</i> Results |
|--|----------------------------|-------------------------------|--------|--------|--------|----------------------------------|
| | | ♂ Rats | ♀ Rats | ♂ Mice | ♀ Mice | |
| 1-Amino-2-Methylantraquinone  | 111 | + | | | | + |
| 2-Amino-4-Nitrophenol  | 339 | + | | | | + |
| <i>o</i> -Anisidine  | 89 | + ^b | | | | + |
| Benzofuran  | 370 | | + | | | - |
| Bromodichloromethane  | 321 | + | + | + | | - |
| Chlorinated Paraffins CH ₃ (CH ₂ CHClCH ₂ CHClCH ₂) ₂ CH ₂ Cl (approximation) | 308 | + | | | | - |
| Chlorothalonil  | 41 | + | + | | | - |

TABLE 11
Evidence of Kidney Neoplasms and *Salmonella* Mutagenicity in Rats and Mice
for Selected Chemicals Tested by the National Toxicology Program (continued)

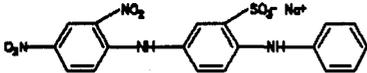
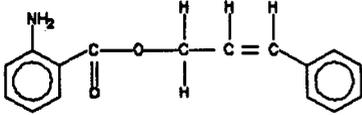
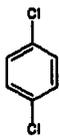
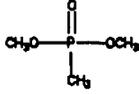
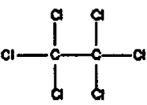
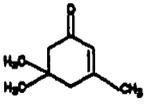
| Chemical Name and Structure | Technical Report Number | Kidney Neoplasms | | | | NTP <i>Salmonella</i> Results |
|---|----------------------------|------------------|--------|--------|--------|----------------------------------|
| | | ♂ Rats | ♀ Rats | ♂ Mice | ♀ Mice | |
| C.I. Acid Orange 3  | 335 | | + | | + | |
| Cinnamyl Anthranilate  | 196 | + | | | | - |
| 1,4-Dichlorobenzene  | 319 | + | | | | - |
| Dimethyl Methylphosphonate  | 323 | + ^c | | | | - |
| Hexachloroethane  | 361 | + | | NT | NT | - |
| Hydroquinone  | 366 | + | | | | - |
| Isophorone  | 291 | + | | | | - |

TABLE 11
 Evidence of Kidney Neoplasms and *Salmonella* Mutagenicity in Rats and Mice
 for Selected Chemicals Tested by the National Toxicology Program (continued)

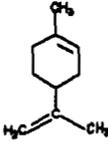
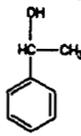
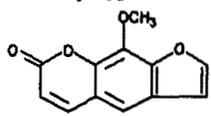
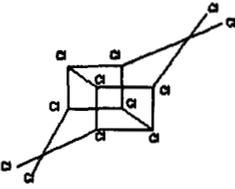
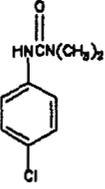
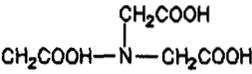
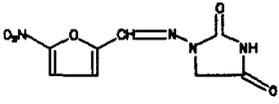
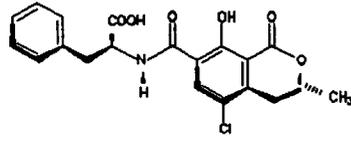
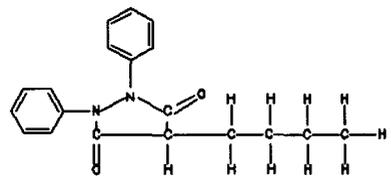
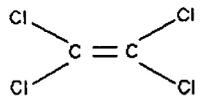
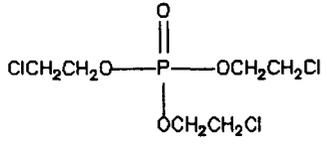
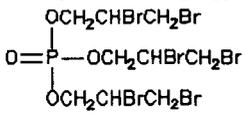
| Chemical Name and Structure | Technical Report Number | Kidney Neoplasms | | | | NTP <i>Salmonella</i> Results |
|---|----------------------------|------------------|--------|--------|--------|----------------------------------|
| | | ♂ Rats | ♀ Rats | ♂ Mice | ♀ Mice | |
| <p><i>d</i>-Limonene</p>  | 347 | + | | | | - |
| <p>α-Methylbenzyl Alcohol</p>  | 369 | + | | | | - |
| <p>8-Methoxyopsoralen</p>  | 359 | + | | NT | NT | + |
| <p>Mirex</p>  | 313 | + ^b | | | | - |
| <p>Monuron</p>  | 266 | + | | | | - |
| <p>Nitriiotriacetic Acid</p>  | 6 | + | | + | + | - |

TABLE 11
Evidence of Kidney Neoplasms and *Salmonella* Mutagenicity in Rats and Mice
for Selected Chemicals Tested by the National Toxicology Program (continued)

| Chemical Name and Structure | Technical Report Number | Kidney Neoplasms | | | | NTP <i>Salmonella</i> Results |
|--|----------------------------|------------------|--------|--------|--------|----------------------------------|
| | | ♂ Rats | ♀ Rats | ♂ Mice | ♀ Mice | |
| Nitrofurantoin  | 341 | + | | | | + |
| Ochratoxin  | 358 | + | + | NT | NT | - |
| Phenylbutazone  | 367 | | + | | | - |
| Tetrachloroethylene  | 311 | + | | | | - |
| Tris(2-Chloroethyl) Phosphate  | 391 | + ^b | + | | | - |
| Tris(2,3-Dibromopropyl) Phosphate  | 76 | + | + | + | | + |

^a Primarily renal tubule cell neoplasms observed unless otherwise noted; + = present, NT = species not tested.

^b Transitional cell neoplasms

^c Transitional cell neoplasms and renal tubule cell neoplasms

REFERENCES

- Aeschbacher, H.U. (1982). The significance of mutagens in food. In *Mutagens in Our Environment*, pp. 349-362. A.R. Liss, New York.
- Ambrose, A.M., Robbins, D.J., and DeEds, F. (1952). Comparative toxicities of quercetin and quercitrin. *J. Am. Pharm. Assoc.* **41**, 119-122.
- Aravindakshan, M., Chauhan, P.S., and Sundaram, K. (1985). Studies on germinal effects of quercetin, a naturally occurring flavonoid. *Mutat. Res.* **144**, 99-106.
- Armitage, P. (1971). *Statistical Methods in Medical Research*, pp. 362-365. John Wiley and Sons, New York.
- Bartholomew, R.M., and Ryan, D.S. (1980). Lack of mutagenicity of some phytoestrogens in the *Salmonella*/mammalian microsome assay. *Mutat. Res.* **78**, 317-321.
- Bjeldanes, L.F., and Chang, G.W. (1977). Mutagenic activity of quercetin and related compounds. *Science* **197**, 577-578.
- Bokkenheuser, V.D., Shackleton, C.H.L., and Winter, J. (1987). Hydrolysis of dietary flavonoid glycosides by strains of intestinal *Bacteroides* from humans. *Biochem. J.* **248**, 953-956.
- Boorman, G.A., Montgomery, C.A., Jr., Eustis, S.L., Wolfe, M.J., McConnell, E.E., and Hardisty, J.F. (1985). Quality assurance in pathology for rodent carcinogenicity studies. In *Handbook of Carcinogen Testing* (H.A. Milman and E.K. Weisburger, Eds.), pp. 345-357. Noyes Publications, Park Ridge, NJ.
- Booth, A.N., Murray, C.W., Jones, F.T., and DeEds, F. (1956). The metabolic rate of rutin and quercetin in the animal body. *J. Biol. Chem.* **223**, 251-257.
- Borghoff, S.J., Short, B.G., and Swenberg, J.A. (1990). Biochemical mechanisms and pathobiology of α_2 -globulin nephropathy. *Annu. Rev. Pharmacol. Toxicol.* **30**, 349-367.
- Brams, A., Buchet, J.P., Crutzen-Fayt, M.C., De Meester, C., Lauwerys, R., and Leonard, A. (1987). A comparative study, with 40 chemicals, of the efficiency of the *Salmonella* assay and the SOS chromotest (kit procedure). *Toxicol. Lett.* **38**, 123-133.
- Brown, J.P. (1980). A review of the genetic effects of naturally occurring flavonoids, anthraquinones and related compounds. *Mutat. Res.* **75**, 243-277.
- Brown, J.P., and Dietrich, P.S. (1979). Mutagenicity of plant flavonols in the *Salmonella*/mammalian microsome test. Activation of flavonol glycosides by mixed glycosidases from rat cecal bacteria and other sources. *Mutat. Res.* **66**, 223-240.
- Brown, S., and Griffiths, L.A. (1983). New metabolites of the naturally-occurring mutagen, quercetin, the pro-mutagen, rutin and of taxifolin. *Experientia* **39**, 198-200.
- Busch, D.B., Hatcher, J.F., and Bryan, G.T. (1986). Urine recovery experiments with quercetin and other mutagens using the Ames test. *Environ. Mutagen.* **8**, 393-399.
- Butterworth, B.E. (1989). Nongenotoxic carcinogens in the regulatory environment. *Regul. Toxicol. Pharmacol.* **9**, 244-256.
- Carver, J.H., Carrano, A.V., and MacGregor, J.T. (1983). Genetic effects of the flavonols quercetin, kaempferol and galangin on Chinese hamster ovary cells *in vitro*. *Mutat. Res.* **113**, 45-60.

- Castillo, M.H., Perkins, E., Campbell, J.H., Doerr, R., Hassett, J.M., Kandaswami, C., and Middleton, E., Jr. (1989). The effects of the bioflavonoid quercetin on squamous cell carcinoma of head and neck origin. *Am. J. Surg.* **158**, 351-355.
- Charbonneau, M., Stasser, J., Borghoff, S.J., and Swenberg, J.A. (1988). *In vitro* hydrolysis of [¹⁴C]- α_2 -globulin isolated from male rat kidney. *Toxicologist* **8**, 135. (Abstr).
- Code of Federal Regulations* (CFR) **21**, part 58.
- Cody, V. (1988). Crystal and molecular structures of flavonoids. In *Plant Flavonoids in Biology and Medicine II: Biochemical, Cellular and Medicinal Properties*. (V. Cody, E. Middleton, Jr., J.B. Harborne, and A. Beretz, Eds.). pp. 29-44. A.R. Liss, NY.
- Cohen, S.M., and Ellwein, L.B. (1990). Cell proliferation in carcinogenesis. *Science* **249**, 1007-1011.
- Cox, D.R. (1972). Regression models and life tables. *J. R. Stat. Soc.* **B34**, 187-220.
- Crebelli, R., Aquilina, G., Falcone, E., and Carere, A. (1987). Urinary and faecal mutagenicity in Sprague-Dawley rats dosed with the food mutagens quercetin and rutin. *Food Chem. Toxicol.* **25**, 9-15.
- Dinse, G.E., and Haseman, J.K. (1986). Logistic regression analysis of incidental-tumor data from animal carcinogenicity experiments. *Fundam. Appl. Toxicol.* **6**, 44-52.
- Dinse, G.E., and Lagakos, S.W. (1983). Regression analysis of tumor prevalence data. *Appl. Statist.* **32**, 236-248.
- Dunn, O.J. (1964). Multiple comparisons using rank sums. *Technometrics* **6**, 241-252.
- Dunnett, W. (1955). A multiple comparison procedure for comparing several treatments with a control. *J. Am. Stat. Assoc.* **50**, 1095-1121.
- Evans, I.A. (1984). Bracken carcinogenicity. *Am. Chem. Soc.* **18**, 1171-1204.
- Farber, E. (1980). The sequential analysis of liver cancer induction. *Biochem. Biophys. Acta* **605**, 149-166.
- Galloway, S.M., Bloom, A.D., Resnick, M., Margolin, B.H., Nakamura, F., Archer, P., and Zeiger, E. (1985). Development of a standard protocol for *in vitro* cytogenetic testing with Chinese hamster ovary cells: Comparison of results for 22 compounds in two laboratories. *Environ. Mutagen.* **7**, 1-51.
- Galloway, S.M., Armstrong, M.J., Reuben, C., Colman, S., Brown, B., Cannon, C., Bloom, A.D., Nakamura, F., Ahmed, M., Duk, S., Rimpo, J., Margolin, B.H., Resnick, M.A., Anderson, B., and Zeiger, E. (1987). Chromosome aberrations and sister chromatid exchanges in Chinese hamster ovary cells: Evaluations of 108 chemicals. *Environ. Mol. Mutagen.* **10** (Suppl. 10), 1-175.
- Gart, J.J., Chu, K.C., and Tarone, R.E. (1979). Statistical issues in interpretation of chronic bioassay tests for carcinogenicity. *J. Natl. Cancer Inst.* **62**, 957-974.
- Gilman, A.G., Rall, T.W., Nies, A.S., and Taylor, P. (Eds.) (1990). *The Pharmacological Basis of Therapeutics*, 8th ed. Pergamon, Elmsford, NY.
- Goldsworthy, T.L., and Popp, J.A. (1987). Chlorinated hydrocarbon-induced peroxisomal enzyme activity in relation to species and organ carcinogenicity. *Toxicol. Appl. Pharmacol.* **88**, 225-233.
- Goldsworthy, T.L., Lyght, O., Burnett, V.L., and Popp, J.A. (1988). Potential role of α_2 -globulin, protein droplet accumulation, and cell replication in the renal carcinogenicity of rats exposed to trichloroethylene, perchloroethylene, and pentachloroethane. *Toxicol. Appl. Pharmacol.* **96**, 367-379.
- Graziani, Y., and Chayoth, R. (1979). Regulation of cyclic AMP level and synthesis of DNA, RNA and protein by quercetin in Ehrlich ascites tumor cells. *Biochem. Pharmacol.* **28**, 397-403.
- Griffith, J.Q., Krewson, C.F., and Naghski, J. (1955). *Rutin and Related Flavonoids*, pp. 234-242. Mack, Easton, PA.

- Gugler, R., Leschik, M., and Dengler, H.J. (1975). Disposition of quercetin in man after single oral and intravenous doses. *Eur. J. Clin. Pharmacol.* **9**, 229-234.
- Hard, G.C. (1986). Experimental models for the sequential analysis of chemically-induced renal carcinogenesis. *Toxicol. Pathol.* **14**, 112-122.
- Hardigree, A.A., and Epler, J.L. (1978). Comparative mutagenesis of plant flavonoids in microbial systems. *Mutat. Res.* **58**, 231-239.
- Haseman, J.K. (1984). Statistical issues in the design, analysis and interpretation of animal carcinogenicity studies. *Environ. Health Perspect.* **58**, 385-392.
- Haseman, J.K., Huff, J., and Boorman, G.A. (1984). Use of historical control data in carcinogenicity studies in rodents. *Toxicol. Pathol.* **12**, 126-135.
- Haseman, J.K., Huff, J.E., Rao, G.N., Arnold, J.E., Boorman, G.A., and McConnell, E.E. (1985). Neoplasms observed in untreated and corn oil gavage control groups of F344/N rats and (C57BL/6N x C3H/HeN)_{F1} (B6C3F₁) mice. *JNCI* **75**, 975-984.
- Hatcher, J.F., Pamukcu, A.M., and Bryan, G.T. (1981). Quercetin (Q) and kaempferol (K) content of bracken fern (BF) and mutagenic activity in urine of rats ingesting Q, rutin (R), or BF. *Proc. Am. Assoc. Cancer Res. Am. Soc. Clin. Oncol.* **22**, 114. (Abstr.)
- Havsteen, B. (1983). Flavonoids, a class of natural products of high pharmacological potency. *Biochem. Pharmacol.* **32**, 1141-1148.
- Haworth, S., Lawlor, T., Mortelmans, K., Speck, W., and Zeiger, E. (1983). *Salmonella* mutagenicity test results for 250 chemicals. *Environ. Mutagen.* **5** (Suppl. 1), 3-142.
- Herrmann, K. (1976). Flavonols and flavones in food plants: A review. *J. Food Technol.* **11**, 433-448.
- Hirono, I. (1986). Carcinogenic principles isolated from bracken fern. *CRC Crit. Rev. Toxicol.* **17**, 1-22.
- Hirono, I., Ueno, I., Hosaka, S., Takanashi, H., Matsushima, T., Sugimura, T., and Natori, S. (1981). Carcinogenicity examination of quercetin and rutin in ACI rats. *Cancer Lett.* **13**, 15-21.
- Hirose, M., Fukushima, S., Sakata, T., Inui, M., and Ito, N. (1983). Effect of quercetin on two-stage carcinogenesis of the rat urinary bladder. *Cancer Lett.* **21**, 23-27.
- Hollander, M., and Wolfe, D.A. (1973). *Nonparametric Statistical Methods*, pp. 120-123. John Wiley and Sons, New York.
- Hosaka, S., and Hirono, I. (1981). Carcinogenicity test of quercetin by pulmonary-adenoma bioassay in strain A mice. *Jpn. J. Cancer Res.* **72**, 327-328.
- International Agency for Research on Cancer (IARC) (1983). Quercetin. IARC Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Humans, Vol. 31. IARC, Lyon, France.
- International Agency for Research on Cancer (IARC) (1986). Bracken fern and some of its constituents. IARC Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Humans, Vol. 40. IARC, Lyon, France.
- International Agency for Research on Cancer (IARC) (1987). Overall evaluations of carcinogenicity. IARC Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Humans, Vol. 7. IARC, Lyon, France.
- Ito, N., Tsuda, H., Tatematsu, M., Inoue, T., Tagawa, Y., Aoki, T., Uwagawa, S., Kagawa, M., Ogiso, T., Masui, T., Imaida, K., Fukushima, S., and Asamoto, M. (1988). Enhancing effect of various hepatocarcinogens on induction of preneoplastic glutathione S-transferase placental form positive foci in rats - an approach for a new medium-term bioassay system. *Carcinogenesis* **9**, 387-394.
- Ito, N., Hagiwara, A., Tamano, S., Kagawa, M., Shibata, M.-A., Kurata, Y., and Fukushima, S. (1989). Lack of carcinogenicity of quercetin in F344/DuCrj rats. *Jpn. J. Cancer Res.* **80**, 317-325.

- Jonckheere, A. (1954). A distribution-free k-sample test against ordered alternatives. *Biometrika* **41**, 133-145.
- Kaplan, E.L., and Meier, P. (1958). Nonparametric estimation of incomplete observations. *J. Am. Stat. Assoc.* **53**, 457-481.
- Kato, R., Nakadate, T., Yamamoto, S., and Sugimura, T. (1983). Inhibition of 12-*o*-tetradecanoylphorbol-13-acetate-induced tumor promotion and ornithine decarboxylase activity by quercetin: Possible involvement of lipoxygenase inhibition. *Carcinogenesis* **4**, 1301-1305.
- Kato, K., Mori, H., Tanaka, T., Fujii, M., Kawai, T., Nishikawa, A., Takahashi, M., and Hirono, I. (1985). Absence of initiating activity by quercetin in the rat liver. *Ecotoxicol. Environ. Safety* **10**, 63-69.
- Konishi, N., and Ward, J.M. (1989). Increased levels of DNA synthesis in hyperplastic renal tubules of aging nephropathy in female F344/NCr rats. *Vet. Pathol.* **26**, 6-10.
- Kühnau, J. (1976). The flavonoids. A class of semi-essential food components: Their role in human nutrition. *World Rev. Nutr. Diet.* **24**, 117-191.
- Kuroda, K., Yoo, Y.S., Yoshikura, T., and Ishibashi, T. (1985). Mutagenicity of natural food additives. *AOISD* **47**, 24-30.
- Kuroda, M., Yoshida, D., and Mizusaki, S. (1985). Mutagenicity of pyrolyzates of natural substances toward *Salmonella typhimurium* TA97. *Agric. Biol. Chem.* **49**, 1893-1895.
- Kurokawa, Y., Hayashi, Y., Maekawa, A., Takahashi, M., Kokubo, T., and Odashima, S. (1983). Carcinogenicity of potassium bromate administered orally to F344 rats. *JNCI* **71**, 965-971.
- Löfroth, G., Lazaridis, G., and Rudling, L. (1986). Mutagenicity assay of emission extracts from wood stoves: Comparison with other emission parameters. *Sci. Total Environ.* **58**, 199-208.
- MacGregor, J.T., and Jurd, L. (1978). Mutagenicity of plant flavonoids: Structural requirements for mutagenic activity in *Salmonella typhimurium*. *Mutat. Res.* **54**, 297-309.
- MacGregor, J.T., Wehr, C.M., Manners, G.D., Jurd, L., Minkler, J.L., and Carrano, A.V. (1983). *In vivo* exposure to plant flavonols: Influence on frequencies of micronuclei in mouse erythrocytes and sister chromatid exchange in rabbit lymphocytes. *Mutat. Res.* **124**, 255-270.
- MacGregor, J.T., and Wilson, R.E. (1988). Flavone mutagenicity in *Salmonella typhimurium*: Dependence on the pKM101 plasmid and excision-repair deficiency. *Environ. Mol. Mutagen.* **11**, 315-322.
- Maronpot, R.R., and Boorman, G.A. (1982). Interpretation of rodent hepatocellular proliferative alterations and hepatocellular tumors in chemical safety assessment. *Toxicol. Pathol.* **10**, 71-80.
- Marzin, D., Phi, H.V., Olivier, P., and Sauzieres, J. (1987). Study of mutagenic activity of troxerutin, a flavonoid derivative. *Toxicol. Lett.* **35**, 297-305.
- McConnell, E.E. (1983a). Pathology requirements for rodent two-year studies. I. A review of current procedures. *Toxicol. Pathol.* **11**, 60-64.
- McConnell, E.E. (1983b). Pathology requirements for rodent two-year studies. II. Alternative approaches. *Toxicol. Pathol.* **11**, 65-76.
- McConnell, E.E., Solleveld, H.A., Swenberg, J.A., and Boorman, G.A. (1986). Guidelines for combining neoplasms for evaluation of rodent carcinogenesis studies. *JNCI* **76**, 283-289.
- McCoy, E.C., Anders, M., and Rosenkranz, H.S. (1983). The basis of the insensitivity of *Salmonella typhimurium* strain TA98/1,8-DNP₆ to the mutagenic action of nitroarenes. *Mutat. Res.* **121**, 17-23.
- McKnight, B., and Crowley, J. (1984). Tests for differences in tumor incidence based on animal carcinogenesis experiments. *J. Am. Stat. Assoc.* **79**, 639-648.
- The Merck Index.* (1983). 10th ed. (M. Windholz, Ed.), p. 1160. Merck and Company, Rahway, NJ.
- Morino, K., Matsukura, N., Kawachi, T., Ohgaki, H., Sugimura, T., and Hirono, I. (1982). Carcinogenicity test of quercetin and rutin in golden hamsters by oral administration. *Carcinogenesis* **3**, 93-97.

- Nagao, M., Morita, N., Yahagi, T., Shimizu, M., Kuroyanagi, M., Fukuoka, M., Yoshihira, K., Natori, S., Fujino, T., and Sugimura, T. (1981). Mutagenicities of 61 flavonoids and 11 related compounds. *Environ. Mutagen* 3, 401-419.
- National Cancer Institute (NCI) (1976). Guidelines for Carcinogen Bioassay in Small Rodents. Technical Report Series No. 1. NIH Publication No. 76-801. U.S. Department of Health, Education, and Welfare; Public Health Service; National Institutes of Health, Bethesda, MD.
- National Institutes of Health (NIH) (1978). Open Formula Rat and Mouse Ration (NIH-07). NIH Publication No. 11-1335. National Institutes of Health, Bethesda, MD.
- Neuhaus, O.W., Flory, W., Biswas, N., and Hollerman, C.E. (1981). Urinary excretion of α_2 -globulin and albumin by adult male rats following treatment with nephrotoxic agents. *Nephron* 28, 133-140.
- Nishino, H., Iwashima, A., Fujiki, H., and Sugimura, T. (1984a). Inhibition by quercetin of the promoting effect of teleocidin on skin papilloma formation in mice initiated with 7,12-dimethylbenz[a]anthracene. *Jpn. J. Cancer Res.* 75, 113-116.
- Nishino, H., Naito, E., Iwashima, A., Tanaka, K., Matsuura, T., Fujiki, H., and Sugimura, T. (1984b). Interaction between quercetin and Ca^{2+} -calmodulin complex: Possible mechanism for anti-tumor-promoting action of the flavonoid. *Jpn. J. Cancer Res.* 75, 311-316.
- Pamukcu, A.M., Yalciner, S., Hatcher, J.F., and Bryan, G.T. (1980). Quercetin, a rat intestinal and bladder carcinogen present in bracken fern (*Pteridium aquilinum*). *Cancer Res.* 40, 3468-3472.
- Petrakis, P.L., Kallianos, A.G., Wender, S.H., and Shetlar, M.R. (1959). Metabolic studies of quercetin labeled with C^{14} . *Arch. Biochem. Biophys.* 85, 264-271.
- Pitot, H.C., and Sirica, A.E. (1980). The stages of initiation and promotion in hepatocarcinogenesis. *Biochem. Biophys. Acta* 605, 191-215.
- Rao, G.N., Piegorsch, W.W., and Haseman, J.K. (1987). Influence of body weight on the incidence of spontaneous tumors in rats and mice of long-term studies. *Am. J. Clin. Nutr.* 45, 252-260.
- Rueff, J., Laires, A., Borba, H., Chaveca, T., Gomes, M.I., and Halpern, M. (1986). Genetic toxicology of flavonoids: The role of metabolic conditions in the induction of reverse mutation, SOS functions and sister chromatid exchanges. *Mutagenesis* 1, 179-183.
- Sadtler Standard Spectra. IR Nos. 594 and 1009, UV No. 18272. Sadtler Research Laboratories, Philadelphia.
- Saito, D., Shirai, A., Matsushima, T., Sugimura, T., and Hirono, I. (1980). Test of carcinogenicity of quercetin, a widely distributed mutagen in food. *Teratogenesis Carcinog. Mutagen.* 1, 213-221.
- Shakhova, M.K., Samokhvalov, G.I., and Preobrazhenskii, N.A. (1962). Synthetic investigations in the field of flavonoids. III. total synthesis of quercetin-3- β -rutinoside, rutin. *Zh. Obshch. Khim.* (USSR) 32, 390-396.
- Shirley, E. (1977). A non-parametric equivalent of Williams' test for contrasting increasing dose levels of a treatment. *Biometrics* 33, 386-389.
- Short, B.G., Burnett, V.L., Cox, M.G., Bus, J.S., and Swenberg, J.A. (1987). Site-specific renal cytotoxicity and cell proliferation in male rats exposed to petroleum hydrocarbons. *Lab. Invest.* 57, 564-577.
- Stich, H.F., Rosin, M.P., Wu, C.H., and Powrie, W.D. (1981). The action of transition metals on the genotoxicity of simple phenols, phenolic acids and cinnamic acids. *Cancer Lett.* 14, 251-260.
- Stoewsand, G.S., Anderson, J.L., Boyd, J.N., Hrazdina, G., Babish, J.G., Walsh, K.M., and Losco, P. (1984). Quercetin: A mutagen, not a carcinogen, in Fischer rats. *J. Toxicol. Environ. Health* 14, 105-114.
- Stott, W.T., Reitz, R.H., Schumann, A.M., and Watanabe, P.G. (1981). Genetic and nongenetic events in neoplasia. *Food Cosmet. Toxicol.* 19, 567-576.

- Sugimura, T., Nagao, M., Matsushima, T., Yahagi, T., Seino, Y., Shirai, A., Sawamura, M., Natori, S., Yoshihira, K., Fukuoka, M., and Kuroyanagi, M. (1977). Mutagenicity of flavone derivatives. *Proc. Jpn. Acad.* B53, 194-197.
- Sullivan, M., Follis, R.H., Jr., and Hilgartner, M. (1951). Toxicology of podophyllin. *Proc. Soc. Exp. Biol. Med.* 77, 269-272.
- Swenberg, J.A., Short, B., Borghoff, S., Strasser, J., and Charbonneau, M. (1989). The Comparative Pathobiology of α_2 -globulin nephropathy. *Toxicol. Appl. Pharmacol.* 97, 35-46.
- Takanashi, H., Aiso, S., Hirono, I., Matsushima, T., and Sugimura, T. (1983). Carcinogenicity test of quercetin and kaempferol in rats by oral administration. *J. Food Safety* 5, 55-60.
- Tamura, G., Gold, C., Ferro-Luzzi, A., and Ames, B.N. (1980). Fecalase: A model for activation of dietary glycosides to mutagens by intestinal flora. *Proc. Natl. Acad. Sci. USA* 77, 4961-4965.
- Tanaka, K., Ono, T., and Umeda, M. (1987). Pleiotropic effects of quercetin on the transformation of BALB 3T3 cells. *Jpn. J. Cancer Res.* 78, 819-825.
- Tarone, R.E. (1975). Tests for trend in life table analysis. *Biometrika* 62, 679-682.
- Tsuda, H., Hacker, H.J., Katayama, H., Masui, T., Ito, N., and Bannasch, P. (1986). Correlative histochemical studies on preneoplastic and neoplastic lesions in the kidney of rats treated with nitrosamines. *Virchows Arch. [Cell Pathol.]* 51, 385-404.
- Ueno, I., Nakano, N., and Hirono, I. (1983). Metabolic fate of [14 C]quercetin in the ACI rat. *Jpn. J. Exp. Med.* 53, 41-50.
- Vandoren, G., Mertens, B., Heyns, W., Van Baelen, H., Rombauts, W., and Verhoeven, G. (1983). Different forms of α_2 -globulin in male and female rat urine. *Eur. J. Biochem.* 134, 175-181.
- Van Duuren, B.L., and Goldschmidt, B.M. (1976). Cocarcinogenic and tumor-promoting agents in tobacco carcinogenesis. *J. Natl. Cancer Inst.* 56, 1237-1242.
- Watson, W.A.F. (1982). The mutagenic activity of quercetin and kaempferol in *Drosophila melanogaster*. *Mutat. Res.* 103, 145-147.
- Weast, R.C. (Ed.). (1979). *Handbook of Chemistry and Physics*, 60th ed. CRC, Boca Raton, FL.
- Willhite, C.C. (1982). Teratogenic potential of quercetin in the rat. *Food Chem. Toxicol.* 20, 75-79.
- Williams, D.A. (1971). A test for differences between treatment means when several dose levels are compared with a zero dose control. *Biometrics* 27, 103-117.
- Williams, D.A. (1972). The comparison of several dose levels with a zero dose control. *Biometrics* 28, 519-531.
- Yoshida, M.A., Sasaki, M., Sugimura, K., and Kawachi, T. (1980). Cytogenetic effects of quercetin on cultured mammalian cells. *Proc. Jpn. Acad. Ser. B* 56, 443-447.
- Zeiger, E., Anderson, B., Haworth, S., Lawlor, T., and Mortelmans, K. (1988). *Salmonella* mutagenicity tests. IV. Results from the testing of 300 chemicals. *Environ. Mol. Mutagen* 11 (Suppl. 12), 1-158.

APPENDIX A
SUMMARY OF LESIONS IN MALE RATS
IN THE 2-YEAR FEED STUDY
OF QUERCETIN

| | | |
|-----------------|---|-----------|
| TABLE A1 | Summary of the Incidence of Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin | 50 |
| TABLE A2 | Individual Animal Tumor Pathology of Male Rats in the 2-Year Feed Study of Quercetin | 54 |
| TABLE A3 | Statistical Analysis of Primary Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin | 78 |
| TABLE A4 | Historical Incidence of Renal Tubule Neoplasms in Untreated Male F344/N Rats . . . | 83 |
| TABLE A5 | Summary of the Incidence of Nonneoplastic Lesions in Male Rats in the 2-Year Feed Study of Quercetin | 84 |

TABLE A1
Summary of the Incidence of Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|--------|-----------|------------|------------|
| Disposition Summary | | | | |
| Animals initially in study | 70 | 70 | 70 | 70 |
| 6-Month interim evaluation | 10 | 10 | 10 | 10 |
| 15-Month interim evaluation | 10 | 10 | 10 | 10 |
| Early deaths | | | | |
| Natural deaths | 3 | 7 | 3 | 6 |
| Moribund | 21 | 15 | 22 | 21 |
| Survivors | | | | |
| Moribund | | | 1 | 1 |
| Terminal sacrifice | 25 | 27 | 22 | 19 |
| Died last week of study | 1 | 1 | 2 | 3 |
| Animals examined microscopically | 50 | 50 | 50 | 50 |
| Alimentary System | | | | |
| Intestine large, cecum | (49) | (11) | (13) | (46) |
| Intestine large, colon | (49) | (12) | (11) | (47) |
| Intestine small, duodenum | (48) | (47) | (48) | (46) |
| Intestine small, ileum | (47) | (47) | (48) | (45) |
| Adenoma | | | | 1 (2%) |
| Intestine small, jejunum | (48) | (44) | (42) | (44) |
| Liver | (50) | (50) | (50) | (50) |
| Cholangiocarcinoma | | | 1 (2%) | |
| Hemangiosarcoma | | | 1 (2%) | |
| Hepatocellular carcinoma | | 1 (2%) | | 1 (2%) |
| Hepatocellular adenoma | 3 (6%) | 1 (2%) | | |
| Hepatocellular adenoma, multiple | | | 1 (2%) | |
| Neoplastic nodule | | 1 (2%) | 2 (4%) | |
| Neoplastic nodule, multiple | | 1 (2%) | 1 (2%) | |
| Sarcoma, metastatic, uncertain primary site | 1 (2%) | | | |
| Mesentery | (7) | (6) | (5) | (5) |
| Sarcoma, poorly differentiated | | | | 1 (20%) |
| Pancreas | (50) | (50) | (50) | (47) |
| Adenoma | | 2 (4%) | 2 (4%) | |
| Salivary glands | (16) | (14) | (12) | (10) |
| Stomach | (50) | (50) | (50) | (49) |
| Stomach, forestomach | (49) | (49) | (49) | (46) |
| Stomach, glandular | (50) | (50) | (49) | (48) |
| Tongue | (45) | (48) | (47) | (44) |
| Papilloma squamous | | | | 1 (2%) |
| Cardiovascular System | | | | |
| Heart | (50) | (18) | (18) | (50) |
| Schwannoma benign | 1 (2%) | | | |
| Schwannoma malignant | | | 1 (6%) | |

TABLE A1
Summary of the Incidence of Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|----------|-----------|------------|------------|
| Endocrine System | | | | |
| Adrenal gland, cortex | (50) | (18) | (21) | (49) |
| Adrenal gland, medulla | (50) | (18) | (21) | (49) |
| Pheochromocytoma malignant | 1 (2%) | | 1 (5%) | 2 (4%) |
| Pheochromocytoma benign | 8 (16%) | 4 (22%) | 9 (43%) | 9 (18%) |
| Pheochromocytoma benign, multiple | 4 (8%) | | 1 (5%) | 2 (4%) |
| Islets, pancreatic | (47) | (15) | (13) | (41) |
| Adenoma | | 2 (13%) | | 3 (7%) |
| Carcinoma | | | 1 (8%) | |
| Parathyroid gland | (43) | (45) | (43) | (43) |
| Adenoma | 1 (2%) | | | |
| Pituitary gland | (46) | (49) | (50) | (48) |
| Pars distalis, adenoma | 14 (30%) | 14 (29%) | 18 (36%) | 11 (23%) |
| Pars distalis, adenoma, multiple | | 3 (6%) | 1 (2%) | 1 (2%) |
| Pars distalis, pars intermedia, pars nervosa, leukemia mononuclear | | | 1 (2%) | |
| Thyroid gland | (50) | (17) | (22) | (49) |
| C-cell, adenoma | 4 (8%) | 1 (6%) | 4 (18%) | 1 (2%) |
| C-cell, carcinoma | 1 (2%) | 2 (12%) | 1 (5%) | |
| Follicular cell, adenocarcinoma | | | | 1 (2%) |
| Follicular cell, adenoma | 1 (2%) | | | 1 (2%) |
| General Body System | | | | |
| Tissue NOS | | (1) | | |
| Basosquamous tumor benign | | 1 (100%) | | |
| Genital System | | | | |
| Epididymis | (50) | (13) | (13) | (49) |
| Preputial gland | (13) | (22) | (19) | (15) |
| Adenoma | 2 (15%) | 5 (23%) | 3 (16%) | 1 (7%) |
| Carcinoma | 1 (8%) | | 1 (5%) | 3 (20%) |
| Squamous cell carcinoma | | 1 (5%) | | |
| Prostate | (49) | (14) | (12) | (48) |
| Seminal vesicle | (50) | (22) | (23) | (49) |
| Testes | (50) | (46) | (48) | (50) |
| Interstitial cell, adenoma | 44 (88%) | 43 (93%) | 45 (94%) | 45 (90%) |
| Hematopoietic System | | | | |
| Bone marrow | (11) | (12) | (12) | (9) |
| Lymph node | (49) | (29) | (26) | (50) |
| Carcinoma, metastatic, thyroid gland | | | 1 (4%) | |
| Mediastinal, sarcoma, metastatic, uncertain primary site | 1 (2%) | | | |
| Lymph node, mandibular | (46) | (18) | (15) | (47) |
| Carcinoma, metastatic, thyroid gland | | 1 (6%) | | |
| Lymph node, mesenteric | (22) | (14) | (17) | (19) |
| Spleen | (50) | (25) | (38) | (50) |
| Hemangioma | | | 1 (3%) | |
| Hemangiosarcoma | | | 1 (3%) | |
| Sarcoma | | | 1 (3%) | |
| Thymus | (14) | (11) | (13) | (5) |
| Mediastinum, basosquamous tumor NOS | 1 (7%) | | | |

TABLE A1
Summary of the Incidence of Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|----------|-----------|------------|------------|
| Integumentary System | | | | |
| Mammary gland | (13) | (10) | (14) | (9) |
| Fibroadenoma | 5 (38%) | 1 (10%) | 5 (36%) | 3 (33%) |
| Skin | (20) | (18) | (19) | (18) |
| Basal cell carcinoma | 1 (5%) | | | |
| Basosquamous tumor benign | | | 1 (5%) | |
| Keratoacanthoma | | | 1 (5%) | |
| Papilloma squamous | 2 (10%) | | 3 (16%) | 1 (6%) |
| Subcutaneous tissue, fibroma | 2 (10%) | 1 (6%) | 1 (5%) | 3 (17%) |
| Subcutaneous tissue, fibrosarcoma | 1 (5%) | | | |
| Subcutaneous tissue, lipoma | 1 (5%) | | | |
| Subcutaneous tissue, myxoma | | | 1 (5%) | |
| Subcutaneous tissue, sarcoma | | 1 (6%) | 1 (5%) | 1 (6%) |
| Musculoskeletal System | | | | |
| Bone | (10) | (12) | (15) | (23) |
| Osteoma | 1 (10%) | | | |
| Skeletal muscle | (1) | (2) | | |
| Sarcoma | 1 (100%) | | | |
| Nervous System | | | | |
| Brain | (50) | (15) | (12) | (50) |
| Meningioma benign | | 1 (7%) | | |
| Choroid plexus, sarcoma, metastatic, uncertain primary site | 1 (2%) | | | |
| Respiratory System | | | | |
| Lung | (50) | (28) | (31) | (50) |
| Alveolar/bronchiolar adenoma | 2 (4%) | | 1 (3%) | 1 (2%) |
| Alveolar/bronchiolar carcinoma, multiple | | | 1 (3%) | |
| Carcinoma, metastatic, thyroid gland | 1 (2%) | 1 (4%) | 1 (3%) | |
| Cholangiocarcinoma, metastatic, liver | | | 1 (3%) | |
| Osteosarcoma, metastatic, bone | 1 (2%) | | | |
| Pheochromocytoma malignant, metastatic, adrenal gland | 1 (2%) | | | 1 (2%) |
| Sarcoma, metastatic, uncertain primary site | 1 (2%) | | | |
| Squamous cell carcinoma | | | | 1 (2%) |
| Mediastinum, schwannoma malignant, metastatic, heart | | | 1 (3%) | |
| Nose | (44) | (48) | (49) | (49) |
| Papilloma squamous | | | | 1 (2%) |
| Glands, carcinoma | | 1 (2%) | | |
| Trachea | (50) | (12) | (12) | (50) |
| Special Senses System | | | | |
| Eye | (2) | (9) | (6) | (3) |

TABLE A1
Summary of the Incidence of Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|----------|-----------|------------|------------|
| Urinary System | | | | |
| Kidney | (50) | (50) | (50) | (50) |
| Sarcoma, metastatic, uncertain primary site | 1 (2%) | | | |
| Proximal convoluted renal tubule, adenoma | | | | 1 (2%) |
| Renal tubule, adenocarcinoma | | | | 1 (2%) |
| Renal tubule, adenoma | | | | 2 (4%) |
| Urinary bladder | (50) | (49) | (49) | (48) |
| Systemic Lesions | | | | |
| Multiple organs ^a | (50) | (50) | (50) | (50) |
| Leukemia mononuclear | 16 (32%) | 18 (36%) | 22 (44%) | 13 (26%) |
| Lymphoma malignant histiocytic | | | 1 (2%) | |
| Lymphoma malignant lymphocytic | 3 (6%) | | | |
| Lymphoma malignant mixed | | 1 (2%) | 1 (2%) | |
| Lymphoma malignant undifferentiated cell | | 1 (2%) | | |
| Mesothelioma benign | | 1 (2%) | 1 (2%) | |
| Mesothelioma malignant | 4 (8%) | 3 (6%) | 2 (4%) | 1 (2%) |
| Tumor Summary | | | | |
| Total animals with primary neoplasms ^b | 50 | 50 | 50 | 48 |
| Total primary neoplasms | 125 | 111 | 139 | 113 |
| Total animals with benign neoplasms | 49 | 48 | 50 | 48 |
| Total benign neoplasms | 95 | 82 | 102 | 88 |
| Total animals with malignant neoplasms | 27 | 27 | 32 | 21 |
| Total malignant neoplasms | 29 | 29 | 37 | 25 |
| Total animals with metastatic neoplasms | 4 | 1 | 3 | 1 |
| Total metastatic neoplasms | 8 | 2 | 4 | 1 |
| Total animals with malignant neoplasms of uncertain primary site | 1 | | | |
| Total animals with neoplasms uncertain-benign or malignant | 1 | | | |
| Total uncertain neoplasms | 1 | | | |

^a Number of animals with any tissue examined microscopically

^b Primary neoplasms: all neoplasms except metastatic neoplasms

TABLE A2
Individual Animal Tumor Pathology of Male Rats in the 2-Year Feed Study of Quercetin: 0 ppm

| | | | | | | | | | | | | | | | | | | | | | | | | | | | |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|--|
| Number of Days on Study | 2 | 4 | 5 | 5 | 5 | 5 | 5 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 7 | 7 | 7 | 7 | 7 | 7 | | |
| | 2 | 5 | 1 | 6 | 7 | 7 | 9 | 2 | 3 | 3 | 4 | 5 | 5 | 5 | 6 | 7 | 7 | 8 | 8 | 0 | 1 | 1 | 2 | 2 | 2 | | |
| | 6 | 8 | 9 | 2 | 3 | 5 | 0 | 0 | 6 | 6 | 0 | 4 | 4 | 6 | 4 | 4 | 9 | 2 | 9 | 4 | 7 | 7 | 0 | 1 | 4 | | |
| Carcass ID Number | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | | |
| | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 0 | 1 | 1 | 0 | 1 | 0 | 1 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 1 | 0 | 0 | | |
| | 1 | 3 | 3 | 2 | 1 | 2 | 6 | 4 | 1 | 2 | 1 | 9 | 1 | 8 | 0 | 0 | 3 | 5 | 1 | 3 | 6 | 1 | 3 | 5 | 4 | | |
| | 4 | 5 | 5 | 3 | 5 | 4 | 4 | 4 | 2 | 3 | 4 | 5 | 3 | 2 | 4 | 3 | 4 | 3 | 2 | 3 | 3 | 1 | 4 | 2 | 3 | | |
| Alimentary System | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Esophagus | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Intestine large | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | A | + | + | + | + | + | + | + | + | |
| Intestine large, cecum | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | A | + | + | + | + | + | + | + | + | |
| Intestine large, colon | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | A | + | + | + | + | + | + | + | + | |
| Intestine large, rectum | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | A | + | + | + | + | + | + | + | + | |
| Intestine small | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | A | + | + | + | + | + | + | + | + | |
| Intestine small, duodenum | + | M | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | A | + | + | + | + | + | + | + | + | |
| Intestine small, ileum | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | M | + | A | + | + | + | + | + | + | + | M | |
| Intestine small, jejunum | + | + | + | + | A | + | + | + | + | + | + | + | + | + | + | + | + | A | + | + | + | + | + | + | + | + | |
| Liver | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Hepatocellular adenoma | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Sarcoma, metastatic, uncertain primary site | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Mesentery | | | | | | + | | | | | + | + | | | | | | + | | | | | | | | | |
| Pancreas | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Salivary glands | + | + | + | + | + | + | | + | + | + | + | | | | | | | + | | | | | | | | M | |
| Stomach | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Stomach, forestomach | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Stomach, glandular | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Tongue | | + | + | + | M | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Cardiovascular System | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Blood vessel | | | | | | | | | | | | | | | | | | | | | | | | | | + | |
| Heart | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Schwannoma benign | | | | | | | | | | | | | | | | | | | | | | | | | | X | |
| Endocrine System | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Adrenal gland | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Adrenal gland, cortex | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Adrenal gland, medulla | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Pheochromocytoma malignant | | | | | | | | | | | | | | | | | | | | | | | | | | X | |
| Pheochromocytoma benign | | | | | | | | | | | | | | | | | | | | | | | | | | X | |
| Pheochromocytoma benign, multiple | | | | | | | | | | | | | | | | | | | | | | | | | | X | |
| Islets, pancreatic | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | |
| Parathyroid gland | + | M | M | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | M | M | + | + | + | + | + | + | |
| Adenoma | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Pituitary gland | A | + | + | + | M | + | + | + | + | + | + | + | + | + | + | + | + | M | + | + | + | + | + | + | + | + | |
| Pars distalis, adenoma | | | | | | | | | | | | | | | | | | | | | | | | | | X | |

+: Tissue examined microscopically
A: Autolysis precludes examination

M: Missing tissue
I: Insufficient tissue

X: Lesion present
Blank: Not examined

TABLE A2
Individual Animal Tumor Pathology of Male Rats in the 2-Year Feed Study of Quercetin: 10,000 ppm

| | |
|-----------------------------------|---|
| Number of Days on Study | 4 4 5 5 5 5 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 7 7 7 |
| | 2 2 4 5 5 7 9 0 1 2 4 4 7 7 7 7 8 8 8 9 9 9 0 0 0 |
| | 0 7 9 5 5 4 2 8 7 4 3 3 3 3 4 4 0 4 7 0 5 9 3 4 4 |
| Carcass ID Number | 0 |
| | 4 3 4 2 3 3 4 3 4 3 3 4 3 4 3 3 3 3 2 3 4 3 3 3 3 |
| | 0 9 2 9 0 7 2 6 1 8 6 1 7 2 2 3 1 9 9 6 1 8 7 6 6 |
| | 5 5 3 3 1 5 2 5 4 4 4 5 2 1 4 4 3 3 2 3 2 3 4 1 2 |
| Alimentary System | |
| Esophagus | + + + + M + + + + + + + |
| Intestine large | + + + + + + + A + + + + + |
| Intestine large, cecum | + + + + + + + A + + + + + |
| Intestine large, colon | + + + + + + + A + + + + + |
| Intestine large, rectum | + + M + + + + A + + + + + |
| Intestine small | + + + + + + + A + + + + + + + + + + + + + + + + + |
| Intestine small, duodenum | + + + + + + + A + + + + + + M + + + + + + + + + + + |
| Intestine small, ileum | + + + + + + + A + + + + + + + + + + + + + + + + + |
| Intestine small, jejunum | + + + + + + + A + + + + + + + + + + M + + + + + M + |
| Liver | + |
| Cholangiocarcinoma | |
| Hemangiosarcoma | |
| Hepatocellular adenoma, multiple | |
| Neoplastic nodule | |
| Neoplastic nodule, multiple | |
| Mesentery | + |
| Pancreas | + |
| Adenoma | |
| Salivary glands | + |
| Stomach | + |
| Stomach, forestomach | + |
| Stomach, glandular | + |
| Tongue | + + + + + + + + M + + + + + + + + + + + + + + + + + + |
| Cardiovascular System | |
| Blood vessel | |
| Heart | + + + + + + + + + + + + + |
| Schwannoma malignant | |
| Endocrine System | |
| Adrenal gland | + |
| Adrenal gland, cortex | + |
| Adrenal gland, medulla | + |
| Pheochromocytoma malignant | |
| Pheochromocytoma benign | |
| Pheochromocytoma benign, multiple | |
| Islets, pancreatic | + |
| Carcinoma | |
| Parathyroid gland | M I + M M + + + + I + + + + + + + + + + + + + + + + + + |
| Pituitary gland | + |
| Pars distalis, adenoma | |
| Pars distalis, adenoma, multiple | |

TABLE A3
Statistical Analysis of Primary Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|-------------|-------------------------|--------------------------|-------------|
| Adrenal Medulla: Pheochromocytoma Benign | | | | |
| Overall rates ^a | 12/50 (24%) | 4/18 (22%) ^e | 10/21 (48%) ^e | 11/49 (22%) |
| Adjusted rates ^b | 37.6% | | | 36.0% |
| Terminal rates ^c | 7/26 (27%) | | | 6/23 (26%) |
| First incidence (days) | 575 | | | 662 |
| Life table tests ^d | | | | P=0.568N |
| Logistic regression tests ^d | | | | P=0.503N |
| Fisher exact test ^d | | | | P=0.522N |
| Adrenal Medulla: Pheochromocytoma (Benign or Malignant) | | | | |
| Overall rates | 13/50 (26%) | 4/18 (22%) ^e | 11/21 (52%) ^e | 12/49 (24%) |
| Adjusted rates | 39.7% | | | 38.5% |
| Terminal rates | 7/26 (27%) | | | 6/23 (26%) |
| First incidence (days) | 575 | | | 662 |
| Life table tests | | | | P=0.574N |
| Logistic regression tests | | | | P=0.501N |
| Fisher exact test | | | | P=0.523N |
| Kidney (Renal Tubule): Adenoma (Single Sections) | | | | |
| Overall rates | 0/50 (0%) | 0/50 (0%) | 0/50 (0%) | 3/50 (6%) |
| Adjusted rates | 0.0% | 0.0% | 0.0% | 11.1% |
| Terminal rates | 0/26 (0%) | 0/28 (0%) | 0/25 (0%) | 2/23 (9%) |
| First incidence (days) | - | | | 676 |
| Life table tests | P=0.007 | - | - | P=0.114 |
| Logistic regression tests | P=0.009 | - | - | P=0.122 |
| Cochran-Armitage test ^d | P=0.008 | | | |
| Fisher exact test | | | | P=0.121 |
| Kidney (Renal Tubule): Adenoma (Single and Step Sections) | | | | |
| Overall rates | 1/50 (2%) | 2/50 (4%) | 7/50 (14%) | 8/50 (16%) |
| Adjusted rates | 3.8% | 7.1% | 22.5% | 27.5% |
| Terminal rates | 1/26 (4%) | 2/28 (7%) | 4/25 (16%) | 5/23 (22%) |
| First incidence (days) | 724 (T) | 724 (T) | 617 | 667 |
| Life table tests | P=0.008 | P=0.526 | P=0.032 | P=0.016 |
| Logistic regression tests | P=0.012 | P=0.526 | P=0.032 | P=0.018 |
| Cochran-Armitage test | P=0.012 | | | |
| Fisher exact test | | P=0.500 | P=0.030 | P=0.015 |
| Kidney (Renal Tubule): Adenoma or Adenocarcinoma (Single Sections) | | | | |
| Overall rates | 0/50 (0%) | 0/50 (0%) | 0/50 (0%) | 4/50 (8%) |
| Adjusted rates | 0.0% | 0.0% | 0.0% | 15.3% |
| Terminal rates | 0/26 (0%) | 0/28 (0%) | 0/25 (0%) | 3/23 (13%) |
| First incidence (days) | | | | 676 |
| Life table tests | P=0.001 | - | - | P=0.056 |
| Logistic regression tests | P=0.002 | - | - | P=0.064 |
| Cochran-Armitage test | P=0.002 | | | |
| Fisher exact test | | - | - | P=0.059 |

TABLE A3
Statistical Analysis of Primary Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin
 (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|------------|-------------------------|------------------------|------------|
| Kidney (Renal Tubule): Adenoma or Adenocarcinoma (Single and Step Sections) | | | | |
| Overall rates | 1/50 (2%) | 2/50 (4%) | 7/50 (14%) | 9/50 (18%) |
| Adjusted rates | 3.8% | 7.1% | 22.5% | 31.5% |
| Terminal rates | 1/26 (4%) | 2/28 (7%) | 4/25 (16%) | 6/23 (26%) |
| First incidence (days) | 724 (T) | 724 (T) | 617 | 667 |
| Life table tests | P=0.003 | P=0.526 | P=0.032 | P=0.008 |
| Logistic regression tests | P=0.005 | P=0.526 | P=0.032 | P=0.010 |
| Cochran-Armitage test | P=0.005 | | | |
| Fisher exact test | | P=0.500 | P=0.030 | P=0.008 |
| Liver: Neoplastic Nodule or Hepatocellular Adenoma | | | | |
| Overall rates | 3/50 (6%) | 3/50 (6%) | 4/50 (8%) | 0/50 (0%) |
| Adjusted rates | 11.5% | 10.7% | 16.0% | 0.0% |
| Terminal rates | 3/26 (12%) | 3/28 (11%) | 4/25 (16%) | 0/23 (0%) |
| First incidence (days) | 724 (T) | 724 (T) | 724 (T) | - |
| Life table tests | P=0.105N | P=0.631N | P=0.478 | P=0.142N |
| Logistic regression tests | P=0.105N | P=0.631N | P=0.478 | P=0.142N |
| Cochran-Armitage test | P=0.082N | | | |
| Fisher exact test | | P=0.661N | P=0.500 | P=0.121N |
| Liver: Neoplastic Nodule, Hepatocellular Adenoma, or Hepatocellular Carcinoma | | | | |
| Overall rates | 3/50 (6%) | 4/50 (8%) | 4/50 (8%) | 1/50 (2%) |
| Adjusted rates | 11.5% | 14.3% | 16.0% | 4.3% |
| Terminal rates | 3/26 (12%) | 4/28 (14%) | 4/25 (16%) | 1/23 (4%) |
| First incidence (days) | 724 (T) | 724 (T) | 724 (T) | 724 (T) |
| Life table tests | P=0.208N | P=0.541 | P=0.478 | P=0.348N |
| Logistic regression tests | P=0.208N | P=0.541 | P=0.478 | P=0.348N |
| Cochran-Armitage test | P=0.161N | | | |
| Fisher exact test | | P=0.500 | P=0.500 | P=0.309N |
| Mammary Gland: Fibroadenoma | | | | |
| Overall rates | 5/50 (10%) | 1/50 (2%) | 5/50 (10%) | 3/50 (6%) |
| Adjusted rates | 16.3% | 3.6% | 17.3% | 13.0% |
| Terminal rates | 3/26 (12%) | 1/28 (4%) | 3/25 (12%) | 3/23 (13%) |
| First incidence (days) | 590 | 724 (T) | 673 | 724 (T) |
| Life table tests | P=0.590 | P=0.094N | P=0.612 | P=0.401N |
| Logistic regression tests | P=0.546N | P=0.100N | P=0.624 | P=0.350N |
| Cochran-Armitage test | P=0.551N | | | |
| Fisher exact test | | P=0.102N | P=0.630N | P=0.357N |
| Pancreatic Islets: Adenoma or Carcinoma | | | | |
| Overall rates | 0/47 (0%) | 2/15 (13%) ^e | 1/13 (8%) ^e | 3/41 (7%) |
| Adjusted rates | 0.0% | | | 11.9% |
| Terminal rates | 0/25 (0%) | | | 1/18 (6%) |
| First incidence (days) | - | | | 694 |
| Life table tests | | | | P=0.101 |
| Logistic regression tests | | | | P=0.100 |
| Fisher exact test | | | | P=0.097 |

TABLE A3
Statistical Analysis of Primary Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin
 (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|-------------|-------------|-------------|-------------|
| Pituitary Gland (Pars Distalis): Adenoma | | | | |
| Overall rates | 14/46 (30%) | 17/49 (35%) | 19/50 (38%) | 12/48 (25%) |
| Adjusted rates | 42.9% | 45.4% | 54.0% | 34.5% |
| Terminal rates | 8/25 (32%) | 9/27 (33%) | 10/25 (40%) | 3/23 (13%) |
| First incidence (days) | 519 | 546 | 592 | 422 |
| Life table tests | P=0.278N | P=0.401 | P=0.219 | P=0.444N |
| Logistic regression tests | P=0.191N | P=0.413 | P=0.245 | P=0.360N |
| Cochran-Armitage test | P=0.192N | | | |
| Fisher exact test | | P=0.412 | P=0.287 | P=0.360N |
| Skin: Squamous Papilloma | | | | |
| Overall rates | 2/50 (4%) | 0/50 (0%) | 3/50 (6%) | 1/50 (2%) |
| Adjusted rates | 7.7% | 0.0% | 12.0% | 2.6% |
| Terminal rates | 2/26 (8%) | 0/28 (0%) | 3/25 (12%) | 0/23 (0%) |
| First incidence (days) | 724 (T) | - | 724 (T) | 676 |
| Life table tests | P=0.632N | P=0.221N | P=0.482 | P=0.516N |
| Logistic regression tests | P=0.589N | P=0.221N | P=0.482 | P=0.495N |
| Cochran-Armitage test | P=0.586N | | | |
| Fisher exact test | | P=0.247N | P=0.500 | P=0.500N |
| Skin (Subcutaneous Tissue): Fibroma | | | | |
| Overall rates | 2/50 (4%) | 1/50 (2%) | 1/50 (2%) | 3/50 (6%) |
| Adjusted rates | 7.1% | 3.0% | 2.1% | 8.8% |
| Terminal rates | 1/26 (4%) | 0/28 (0%) | 0/25 (0%) | 1/23 (4%) |
| First incidence (days) | 717 | 704 | 555 | 574 |
| Life table tests | P=0.241 | P=0.480N | P=0.525N | P=0.476 |
| Logistic regression tests | P=0.265 | P=0.489N | P=0.502N | P=0.500 |
| Cochran-Armitage test | P=0.263 | | | |
| Fisher exact test | | P=0.500N | P=0.500N | P=0.500 |
| Skin (Subcutaneous Tissue): Fibroma or Fibrosarcoma | | | | |
| Overall rates | 3/50 (6%) | 1/50 (2%) | 1/50 (2%) | 3/50 (6%) |
| Adjusted rates | 8.9% | 3.0% | 2.1% | 8.8% |
| Terminal rates | 1/26 (4%) | 0/28 (0%) | 0/25 (0%) | 1/23 (4%) |
| First incidence (days) | 458 | 704 | 555 | 574 |
| Life table tests | P=0.351 | P=0.292N | P=0.330N | P=0.635 |
| Logistic regression tests | P=0.392 | P=0.320N | P=0.328N | P=0.664N |
| Cochran-Armitage test | P=0.378 | | | |
| Fisher exact test | | P=0.309N | P=0.309N | P=0.661N |
| Skin (Subcutaneous Tissue): Fibroma, Fibrosarcoma, or Sarcoma | | | | |
| Overall rates | 3/50 (6%) | 2/50 (4%) | 2/50 (4%) | 4/50 (8%) |
| Adjusted rates | 8.9% | 6.2% | 5.0% | 10.7% |
| Terminal rates | 1/26 (4%) | 0/28 (0%) | 0/25 (0%) | 1/23 (4%) |
| First incidence (days) | 458 | 704 | 555 | 555 |
| Life table tests | P=0.282 | P=0.474N | P=0.519N | P=0.476 |
| Logistic regression tests | P=0.318 | P=0.509N | P=0.523N | P=0.501 |
| Cochran-Armitage test | P=0.304 | | | |
| Fisher exact test | | P=0.500N | P=0.500N | P=0.500 |

TABLE A3

Statistical Analysis of Primary Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin
(continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|--------------|-------------------------|-------------------------|--------------|
| Testes: Adenoma | | | | |
| Overall rates | 44/50 (88%) | 43/46 (93%) | 45/48 (94%) | 45/50 (90%) |
| Adjusted rates | 100.0% | 100.0% | 100.0% | 100.0% |
| Terminal rates | 26/26 (100%) | 26/26 (100%) | 24/24 (100%) | 23/23 (100%) |
| First incidence (days) | 573 | 509 | 420 | 555 |
| Life table tests | P=0.252 | P=0.477N | P=0.341 | P=0.335 |
| Logistic regression tests | P=0.617 | P=0.226 | P=0.218 | P=0.521 |
| Cochran-Armitage test | P=0.521N | | | |
| Fisher exact test | | P=0.287 | P=0.264 | P=0.500 |
| Thyroid Gland (C-cell): Adenoma or Carcinoma | | | | |
| Overall rates | 5/50 (10%) | 3/17 (18%) ^e | 4/22 (18%) ^e | 1/49 (2%) |
| Adjusted rates | 15.6% | | | 2.6% |
| Terminal rates | 2/26 (8%) | | | 0/23 (0%) |
| First incidence (days) | 654 | | | 676 |
| Life table tests | | | | P=0.116N |
| Logistic regression tests | | | | P=0.103N |
| Fisher exact test | | | | P=0.107N |
| All Organs: Mononuclear Leukemia | | | | |
| Overall rates | 16/50 (32%) | 18/50 (36%) | 22/50 (44%) | 13/50 (26%) |
| Adjusted rates | 42.4% | 39.2% | 58.8% | 42.5% |
| Terminal rates | 6/26 (23%) | 2/28 (7%) | 11/25 (44%) | 7/23 (30%) |
| First incidence (days) | 590 | 546 | 549 | 662 |
| Life table tests | P=0.257N | P=0.470 | P=0.168 | P=0.393N |
| Logistic regression tests | P=0.164N | P=0.394 | P=0.151 | P=0.322N |
| Cochran-Armitage test | P=0.164N | | | |
| Fisher exact test | | P=0.417 | P=0.151 | P=0.330N |
| All Organs: Malignant Lymphoma (Histiocytic, Lymphocytic, Mixed, or Undifferentiated Cell Type) | | | | |
| Overall rates | 3/50 (6%) | 2/50 (4%) | 2/50 (4%) | 0/50 (0%) |
| Adjusted rates | 8.3% | 5.5% | 6.2% | 0.0% |
| Terminal rates | 0/26 (0%) | 1/28 (4%) | 1/25 (4%) | 0/23 (0%) |
| First incidence (days) | 654 | 458 | 592 | - |
| Life table tests | P=0.117N | P=0.488N | P=0.514N | P=0.118N |
| Logistic regression tests | P=0.102N | P=0.533N | P=0.502N | P=0.121N |
| Cochran-Armitage test | P=0.107N | | | |
| Fisher exact test | | P=0.500N | P=0.500N | P=0.121N |
| All Organs: Mesothelioma (Benign or Malignant) | | | | |
| Overall rates | 4/50 (8%) | 4/50 (8%) | 3/50 (6%) | 1/50 (2%) |
| Adjusted rates | 10.8% | 11.8% | 6.9% | 3.6% |
| Terminal rates | 1/26 (4%) | 2/28 (7%) | 0/25 (0%) | 0/23 (0%) |
| First incidence (days) | 573 | 599 | 420 | 704 |
| Life table tests | P=0.133N | P=0.626N | P=0.501N | P=0.192N |
| Logistic regression tests | P=0.111N | P=0.635 | P=0.534N | P=0.179N |
| Cochran-Armitage test | P=0.120N | | | |
| Fisher exact test | | P=0.643N | P=0.500N | P=0.181N |

TABLE A3
Statistical Analysis of Primary Neoplasms in Male Rats in the 2-Year Feed Study of Quercetin
 (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|--------------|----------------|----------------|--------------|
| All Organs: Benign Tumors | | | | |
| Overall rates | 49/50 (98%) | 48/50 (96%) | 50/50 (100%) | 48/50 (96%) |
| Adjusted rates | 100.0% | 100.0% | 100.0% | 100.0% |
| Terminal rates | 26/26 (100%) | 28/28 (100%) | 25/25 (100%) | 23/23 (100%) |
| First incidence (days) | 226 | 509 | 420 | 422 |
| Life table tests | P=0.340 | P=0.359N | P=0.413 | P=0.467 |
| Logistic regression tests | P=0.643 | P=0.307N | P=0.627 | P=0.612N |
| Cochran-Armitage test | P=0.464N | | | |
| Fisher exact test | | P=0.500N | P=0.500 | P=0.500N |
| All Organs: Malignant Tumors | | | | |
| Overall rates | 29/50 (58%) | 27/50 (54%) | 32/50 (64%) | 21/50 (42%) |
| Adjusted rates | 64.9% | 54.7% | 73.5% | 57.0% |
| Terminal rates | 11/26 (42%) | 6/28 (21%) | 14/25 (56%) | 9/23 (39%) |
| First incidence (days) | 458 | 458 | 420 | 422 |
| Life table tests | P=0.170N | P=0.392N | P=0.339 | P=0.177N |
| Logistic regression tests | P=0.047N | P=0.438N | P=0.340 | P=0.081N |
| Cochran-Armitage test | P=0.050N | | | |
| Fisher exact test | | P=0.420N | P=0.341 | P=0.081N |
| All Organs: Benign or Malignant Tumors | | | | |
| Overall rates | 50/50 (100%) | 50/50 (100%) | 50/50 (100%) | 48/50 (96%) |
| Adjusted rates | 100.0% | 100.0% | 100.0% | 100.0% |
| Terminal rates | 26/26 (100%) | 28/28 (100%) | 25/25 (100%) | 23/23 (100%) |
| First incidence (days) | 226 | 458 | 420 | 422 |
| Life table tests | P=0.427 | P=0.419N | P=0.470 | P=0.523 |
| Logistic regression tests | P=0.058N | - ^g | - ^g | P=0.162N |
| Cochran-Armitage test | P=0.042N | | | |
| Fisher exact test | | P=1.000N | P=1.000N | P=0.247N |

(T)Terminal sacrifice

^a Number of tumor-bearing animals/number of animals examined. Denominator is number of animals examined microscopically for adrenal gland, bone marrow, brain, clitoral gland, epididymis, gallbladder (mouse), heart, kidney, larynx, liver, lung, nose, ovary, pancreas, parathyroid gland, pituitary gland, preputial gland, prostate gland, salivary gland, spleen, testes, thyroid gland, and urinary bladder; for other tissues, denominator is number of animals necropsied.

^b Kaplan-Meier estimated tumor incidence at the end of the study after adjustment for intercurrent mortality

^c Observed incidence at terminal kill

^d Beneath the control incidence are the P values associated with the trend test. Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the controls and that dosed group. The life table analysis regards tumors in animals dying prior to terminal kill as being (directly or indirectly) the cause of death. The logistic regression tests regard these lesions as nonfatal. The Cochran-Armitage and Fisher exact tests compare directly the overall incidence rates. For all tests, a negative trend or a lower incidence in a dose group is indicated by N.

^e Tissue was examined microscopically only when it was observed to be abnormal at necropsy; thus statistical comparisons with the controls are not appropriate.

^f Not applicable; no tumors in animal group.

^g Value of statistic cannot be computed.

TABLE A4
Historical Incidence of Renal Tubule Neoplasms in Untreated Male F344/N Rats^a

| Study | Incidence in Controls | | |
|--|-----------------------|--------------------------------|--|
| | Adenoma | Adenocarcinoma or Carcinoma | Adenoma, Adenocarcinoma or Carcinoma |
| Historical Incidence at EG&G Mason Research Institute | | | |
| 4-Hydroxyacetanilide | 3/50 | 0/50 | 3/50 |
| Pentaerythritol tetranitrate | 0/49 | 0/49 | 0/49 |
| Total | 3/99 (3.0%) | | 3/99 (3.0%) |
| Overall Historical Incidence | | | |
| Total | 4/499 (0.8%) | 4/499 (0.8%) | 8/499 (1.6%) |
| Standard deviation | 1.9% | 1.1% | 2.3% |
| Range | 0%-6% | 0%-4% | 0%-6% |

^a Data as of 17 September 1990

TABLE A5
Summary of the Incidence of Nonneoplastic Lesions in Male Rats in the 2-Year Feed Study of Quercetin

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--------------------------------------|----------|-----------|------------|------------|
| Disposition Summary | | | | |
| Animals initially in study | 70 | 70 | 70 | 70 |
| 6-Month interim evaluation | 10 | 10 | 10 | 10 |
| 15-Month interim evaluation | 10 | 10 | 10 | 10 |
| Early deaths | | | | |
| Natural deaths | 3 | 7 | 3 | 6 |
| Moribund | 21 | 15 | 22 | 21 |
| Survivors | | | | |
| Moribund | | | 1 | 1 |
| Terminal sacrifice | 25 | 27 | 22 | 19 |
| Died last week of study | 1 | 1 | 2 | 3 |
| Animals examined microscopically | 50 | 50 | 50 | 50 |
| Alimentary System | | | | |
| Intestine large, cecum | (49) | (11) | (13) | (46) |
| Parasite metazoan | 4 (8%) | 1 (9%) | | 4 (9%) |
| Intestine large, colon | (49) | (12) | (11) | (47) |
| Parasite metazoan | 9 (18%) | 2 (17%) | 1 (9%) | 3 (6%) |
| Epithelium, pigmentation | | | | 1 (2%) |
| Intestine large, rectum | (49) | (11) | (10) | (46) |
| Parasite metazoan | 3 (6%) | | | 4 (9%) |
| Intestine small | (49) | (50) | (49) | (46) |
| Autolysis | | | 1 (2%) | |
| Intestine small, duodenum | (48) | (47) | (48) | (46) |
| Epithelium, pigmentation | | | | 3 (7%) |
| Intestine small, ileum | (47) | (47) | (48) | (45) |
| Necrosis, coagulative | | 1 (2%) | | |
| Epithelium, pigmentation | | 1 (2%) | 15 (31%) | 28 (62%) |
| Peyer's patch, hyperplasia | | | 1 (2%) | |
| Intestine small, jejunum | (48) | (44) | (42) | (44) |
| Epithelium, pigmentation | | | 2 (5%) | 19 (43%) |
| Liver | (50) | (50) | (50) | (50) |
| Angiectasis | | 2 (4%) | 2 (4%) | 2 (4%) |
| Basophilic focus | 16 (32%) | 17 (34%) | 18 (36%) | 20 (40%) |
| Clear cell focus | 7 (14%) | 10 (20%) | 5 (10%) | 8 (16%) |
| Congestion | 1 (2%) | | | |
| Cyst | | | | 1 (2%) |
| Cyst multilocular | | | | 1 (2%) |
| Cytoplasmic alteration | 2 (4%) | 3 (6%) | 1 (2%) | |
| Degeneration | 4 (8%) | 2 (4%) | 1 (2%) | |
| Degeneration, cystic | 6 (12%) | 5 (10%) | 9 (18%) | 2 (4%) |
| Eosinophilic focus | 9 (18%) | 5 (10%) | 7 (14%) | 3 (6%) |
| Fatty change | 5 (10%) | 12 (24%) | 5 (10%) | 8 (16%) |
| Fibrosis | 2 (4%) | | | |
| Hemorrhage | 1 (2%) | 4 (8%) | 3 (6%) | 3 (6%) |
| Hepatodiaphragmatic nodule | 2 (4%) | 1 (2%) | | 1 (2%) |
| Hyperplasia, focal | 1 (2%) | 2 (4%) | | |
| Inflammation, chronic | 24 (48%) | 30 (60%) | 26 (52%) | 23 (46%) |
| Mixed cell focus | 3 (6%) | 4 (8%) | 4 (8%) | 3 (6%) |
| Mixed cell focus, multiple | | | 1 (2%) | |
| Necrosis, coagulative | 7 (14%) | 8 (16%) | 9 (18%) | 7 (14%) |
| Thrombus | | 1 (2%) | 1 (2%) | 1 (2%) |
| Bile duct, hyperplasia | 46 (92%) | 48 (96%) | 47 (94%) | 30 (60%) |
| Centrilobular, necrosis, coagulative | | 1 (2%) | 1 (2%) | 2 (4%) |

TABLE A5
Summary of the Incidence of Nonneoplastic Lesions in Male Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|----------|-----------|------------|------------|
| Alimentary System (continued) | | | | |
| Mesentery | (7) | (6) | (5) | (5) |
| Fibrosis | | 1 (17%) | 1 (20%) | |
| Inflammation, chronic | 2 (29%) | | | 1 (20%) |
| Inflammation, granulomatous, chronic | | | | 2 (40%) |
| Mineralization | | | 1 (20%) | 1 (20%) |
| Necrosis, coagulative | 1 (14%) | 2 (33%) | 1 (20%) | 1 (20%) |
| Pigmentation | 1 (14%) | | 1 (20%) | |
| Pancreas | (50) | (50) | (50) | (47) |
| Atrophy | 23 (46%) | 26 (52%) | 23 (46%) | 24 (51%) |
| Cyst | | | 1 (2%) | 1 (2%) |
| Cytoplasmic alteration | | 2 (4%) | | 3 (6%) |
| Ectopic liver | | 1 (2%) | | 1 (2%) |
| Fibrosis | | | 2 (4%) | |
| Hyperplasia | | 3 (6%) | 3 (6%) | 1 (2%) |
| Inflammation, chronic | 32 (64%) | 28 (56%) | 30 (60%) | 34 (72%) |
| Necrosis, coagulative | | | 1 (2%) | |
| Pigmentation | 1 (2%) | | 2 (4%) | |
| Thrombus | | | 1 (2%) | |
| Artery, fibrosis | | 3 (6%) | | |
| Artery, inflammation, necrotizing, chronic active | | 1 (2%) | 2 (4%) | |
| Artery, mineralization | | | | 1 (2%) |
| Duct, dilatation | 1 (2%) | 1 (2%) | | |
| Perivascular, inflammation, chronic | | 3 (6%) | | |
| Serosa, hyperplasia | | | 1 (2%) | |
| Salivary glands | (16) | (14) | (12) | (10) |
| Parotid gland, vacuolization cytoplasmic | | | 1 (8%) | |
| Stomach, forestomach | (49) | (49) | (49) | (46) |
| Acanthosis | | 6 (12%) | 2 (4%) | 5 (11%) |
| Edema | | | | 1 (2%) |
| Fibrosis | | | | 1 (2%) |
| Hyperkeratosis | | 3 (6%) | 2 (4%) | 5 (11%) |
| Hyperplasia, basal cell | 3 (6%) | 9 (18%) | 8 (16%) | 2 (4%) |
| Hyperplasia, pseudoepitheliomatous | | | | 1 (2%) |
| Inflammation, acute | | 1 (2%) | | |
| Inflammation, chronic active | 1 (2%) | 1 (2%) | | 3 (7%) |
| Mineralization | | | 3 (6%) | 2 (4%) |
| Ulcer | | 1 (2%) | 1 (2%) | |
| Muscularis, pigmentation | | | 1 (2%) | |
| Stomach, glandular | (50) | (50) | (49) | (48) |
| Edema | | | | 1 (2%) |
| Hemorrhage | | | 1 (2%) | |
| Inflammation, chronic active | 1 (2%) | 1 (2%) | | 1 (2%) |
| Necrosis, coagulative | | 1 (2%) | 1 (2%) | |
| Epithelium, pigmentation | | | 3 (6%) | 34 (71%) |
| Mucosa, mineralization | | | 3 (6%) | 7 (15%) |
| Muscularis, mineralization | 1 (2%) | | 1 (2%) | |
| Submucosa, fibrosis | 1 (2%) | | | |
| Tongue | (45) | (48) | (47) | (44) |
| Hemorrhage | | 1 (2%) | | |
| Inflammation, necrotizing, acute | | | | 1 (2%) |
| Metaplasia, osseous | 1 (2%) | | | |
| Artery, mineralization | | | 1 (2%) | 4 (9%) |
| Artery, endothelium, hyperplasia | | | | 1 (2%) |

TABLE A5
Summary of the Incidence of Nonneoplastic Lesions in Male Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|----------|-----------|------------|------------|
| Cardiovascular System | | | | |
| Blood vessel | (1) | | (1) | (2) |
| Aorta, mineralization | 1 (100%) | | 1 (100%) | 2 (100%) |
| Heart | (50) | (18) | (18) | (50) |
| Cardiomyopathy | 48 (96%) | 14 (78%) | 15 (83%) | 49 (98%) |
| Cytomegaly | | 1 (6%) | | |
| Edema | | | | 1 (2%) |
| Inflammation, chronic | | | | 1 (2%) |
| Metaplasia, osseous | | | 2 (11%) | 1 (2%) |
| Mineralization | | | | 2 (4%) |
| Thrombus | 6 (12%) | 1 (6%) | 5 (28%) | 1 (2%) |
| Artery, mineralization | | | | 3 (6%) |
| Coronary artery, inflammation, chronic active | 1 (2%) | | | |
| Endocrine System | | | | |
| Adrenal gland, cortex | (50) | (18) | (21) | (49) |
| Angiectasis | 6 (12%) | 1 (6%) | 2 (10%) | 8 (16%) |
| Atrophy | 1 (2%) | | | |
| Congestion | 1 (2%) | 3 (17%) | | 2 (4%) |
| Hematopoietic cell proliferation | 1 (2%) | | | |
| Hemorrhage | | | 1 (5%) | |
| Hyperplasia | 10 (20%) | 2 (11%) | 1 (5%) | 14 (29%) |
| Necrosis, coagulative | | 2 (11%) | 1 (5%) | 1 (2%) |
| Vacuolization cytoplasmic | 26 (52%) | 12 (67%) | 8 (38%) | 27 (55%) |
| Adrenal gland, medulla | (50) | (18) | (21) | (49) |
| Angiectasis | 2 (4%) | | | 1 (2%) |
| Atrophy | 1 (2%) | | | |
| Congestion | | | | 2 (4%) |
| Hyperplasia | 22 (44%) | 2 (11%) | 3 (14%) | 21 (43%) |
| Necrosis, coagulative | 1 (2%) | | | 1 (2%) |
| Islets, pancreatic | (47) | (15) | (13) | (41) |
| Hyperplasia | 1 (2%) | 1 (7%) | | 1 (2%) |
| Parathyroid gland | (43) | (45) | (43) | (43) |
| Hyperplasia | 1 (2%) | 6 (13%) | 6 (14%) | 17 (40%) |
| Pituitary gland | (46) | (49) | (50) | (48) |
| Autolysis | | 1 (2%) | | 3 (6%) |
| Hemorrhage | | | | 1 (2%) |
| Pars distalis, angiectasis | 2 (4%) | 1 (2%) | 1 (2%) | 1 (2%) |
| Pars distalis, autolysis | | 1 (2%) | 2 (4%) | |
| Pars distalis, congestion | | | 1 (2%) | |
| Pars distalis, cyst | 7 (15%) | 6 (12%) | 4 (8%) | 7 (15%) |
| Pars distalis, hyperplasia | 18 (39%) | 24 (49%) | 17 (34%) | 20 (42%) |
| Pars distalis, necrosis | | 1 (2%) | | |
| Pars distalis, pigmentation | | 5 (10%) | 1 (2%) | |
| Pars intermedia, angiectasis | | 2 (4%) | 3 (6%) | |
| Pars intermedia, crystals | | | 1 (2%) | |
| Pars intermedia, cyst | 6 (13%) | 10 (20%) | 14 (28%) | 7 (15%) |
| Pars intermedia, ectopic tissue | | | 1 (2%) | |
| Pars intermedia, hyperplasia | | | 1 (2%) | |
| Pars nervosa, cyst | | | | 1 (2%) |
| Pars nervosa, ectopic tissue | | | 1 (2%) | |
| Pars nervosa, hyperplasia | | | 1 (2%) | |
| Rathke's cleft, cyst | | | 1 (2%) | |

TABLE A5
Summary of the Incidence of Nonneoplastic Lesions in Male Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|----------|-----------|------------|------------|
| Endocrine System (continued) | | | | |
| Thyroid gland | (50) | (17) | (22) | (49) |
| Congestion | | 1 (6%) | | |
| Inflammation, acute | 1 (2%) | | | |
| Ultimobranchial cyst | 2 (4%) | | | |
| C-cell, hyperplasia | 17 (34%) | 3 (18%) | 6 (27%) | 16 (33%) |
| Follicle, pigmentation | 1 (2%) | | 1 (5%) | 6 (12%) |
| Follicular cell, cyst | | | | 3 (6%) |
| Follicular cell, hyperplasia | 2 (4%) | | 1 (5%) | 1 (2%) |
| General Body System | | | | |
| None | | | | |
| Genital System | | | | |
| Coagulating gland | (2) | (1) | (2) | |
| Adventitia, inflammation, chronic active | 1 (50%) | | | |
| Preputial gland | (13) | (22) | (19) | (15) |
| Abscess | | 5 (23%) | 2 (11%) | |
| Cyst | 1 (8%) | | | |
| Hyperplasia | | | | 1 (7%) |
| Inflammation, chronic | 12 (92%) | 19 (86%) | 13 (68%) | 13 (87%) |
| Duct, dilatation | | 1 (5%) | | |
| Prostate | (49) | (14) | (12) | (48) |
| Fibrosis | | | | 1 (2%) |
| Hemorrhage | | 1 (7%) | | |
| Inflammation, acute | | 1 (7%) | | |
| Inflammation, chronic | | | | 1 (2%) |
| Inflammation, chronic active | 25 (51%) | 9 (64%) | 10 (83%) | 30 (63%) |
| Epithelium, hyperplasia | 2 (4%) | | | 2 (4%) |
| Seminal vesicle | (50) | (22) | (23) | (49) |
| Atrophy | 36 (72%) | 9 (41%) | 9 (39%) | 39 (80%) |
| Testes | (50) | (46) | (48) | (50) |
| Infarct | | 1 (2%) | | |
| Necrosis, coagulative | 1 (2%) | | | 1 (2%) |
| Interstitial cell, hyperplasia | 34 (68%) | 35 (76%) | 41 (85%) | 44 (88%) |
| Seminiferous tubule, atrophy | 44 (88%) | 44 (96%) | 43 (90%) | 46 (92%) |
| Hematopoietic System | | | | |
| Bone marrow | (11) | (12) | (12) | (9) |
| Fibrosis | | | | 1 (11%) |
| Lymph node | (49) | (29) | (26) | (50) |
| Angiectasis | | 1 (3%) | | |
| Artery, pancreatic, thrombus | | | 1 (4%) | |
| Inguinal, ectasia | | 1 (3%) | | |
| Lumbar, ectasia | 1 (2%) | 3 (10%) | | |
| Lumbar, hemorrhage | 1 (2%) | 1 (3%) | 1 (4%) | |
| Lumbar, hyperplasia, plasma cell | | | 1 (4%) | |
| Lumbar, infiltration cellular, histiocyte | | 1 (3%) | | |
| Lumbar, inflammation, acute | | 1 (3%) | | |
| Lumbar, pigmentation | | 1 (3%) | | |

TABLE A5
Summary of the Incidence of Nonneoplastic Lesions in Male Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|-------------|-------------|-------------|-------------|
| Hematopoietic System (continued) | | | | |
| Lymph node (continued) | | | | |
| Mediastinal, depletion lymphoid | | | | 1 (2%) |
| Mediastinal, ectasia | | 2 (7%) | 3 (12%) | |
| Mediastinal, hemorrhage | 6 (12%) | 4 (14%) | 3 (12%) | 6 (12%) |
| Mediastinal, infiltration cellular, histiocyte | | | 1 (4%) | |
| Mediastinal, pigmentation | 1 (2%) | 5 (17%) | 2 (8%) | 1 (2%) |
| Pancreatic, ectasia | 1 (2%) | | | |
| Pancreatic, hemorrhage | 3 (6%) | 2 (7%) | 2 (8%) | 2 (4%) |
| Pancreatic, hyperplasia, plasma cell | 1 (2%) | | | |
| Pancreatic, infiltration cellular, histiocyte | 3 (6%) | 5 (17%) | 2 (8%) | |
| Pancreatic, pigmentation | 3 (6%) | 2 (7%) | 3 (12%) | |
| Renal, ectasia | 1 (2%) | 2 (7%) | 2 (8%) | 1 (2%) |
| Renal, fibrosis | | 1 (3%) | | |
| Renal, hemorrhage | 5 (10%) | 4 (14%) | 5 (19%) | 5 (10%) |
| Renal, hyperplasia, lymphoid | | | 1 (4%) | |
| Renal, hyperplasia, plasma cell | 1 (2%) | | | 1 (2%) |
| Renal, infiltration cellular, histiocyte | | 4 (14%) | 2 (8%) | 2 (4%) |
| Renal, pigmentation | 6 (12%) | 6 (21%) | 3 (12%) | 5 (10%) |
| Lymph node, mandibular | (46) | (18) | (15) | (47) |
| Angiectasis | 1 (2%) | | | |
| Congestion | | 1 (6%) | | 1 (2%) |
| Depletion lymphoid | | | | 1 (2%) |
| Ectasia | 3 (7%) | 1 (6%) | 3 (20%) | 10 (21%) |
| Hemorrhage | 12 (26%) | 4 (22%) | 2 (13%) | 12 (26%) |
| Hyperplasia, plasma cell | 3 (7%) | | | 1 (2%) |
| Infiltration cellular, histiocyte | | | | 3 (6%) |
| Pigmentation | | | | 2 (4%) |
| Lymph node, mesenteric | (22) | (14) | (17) | (19) |
| Ectasia | | | 1 (6%) | 12 (63%) |
| Hemorrhage | 3 (14%) | 1 (7%) | 4 (24%) | 3 (16%) |
| Hyperplasia, plasma cell | 1 (5%) | | | |
| Infiltration cellular, histiocyte | 11 (50%) | 10 (71%) | 14 (82%) | 9 (47%) |
| Pigmentation | 9 (41%) | 9 (64%) | 10 (59%) | 8 (42%) |
| Spleen | (50) | (25) | (38) | (50) |
| Congestion | 1 (2%) | 2 (8%) | 1 (3%) | |
| Depletion lymphoid | 9 (18%) | 14 (56%) | 13 (34%) | 9 (18%) |
| Fibrosis | 6 (12%) | 5 (20%) | 8 (21%) | 1 (2%) |
| Hematopoietic cell proliferation | 1 (2%) | 2 (8%) | 1 (3%) | 3 (6%) |
| Hyperplasia, lymphoid | | | 1 (3%) | |
| Infarct | 2 (4%) | | | |
| Necrosis, coagulative | 1 (2%) | | | |
| Pigmentation | 1 (2%) | | 1 (3%) | |
| Thrombus | | | 1 (3%) | |
| Thymus | (14) | (11) | (13) | (5) |
| Congestion | | | 1 (8%) | |
| Depletion lymphoid | 7 (50%) | 5 (45%) | 5 (38%) | 5 (100%) |
| Hemorrhage | 1 (7%) | 1 (9%) | 1 (8%) | |

TABLE A5
Summary of the Incidence of Nonneoplastic Lesions in Male Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|----------|-----------|------------|------------|
| Integumentary System | | | | |
| Mammary gland | (13) | (10) | (14) | (9) |
| Abscess | | 1 (10%) | | |
| Galactocele | 1 (8%) | 1 (10%) | | 1 (11%) |
| Hyperplasia | 7 (54%) | 9 (90%) | 9 (64%) | 6 (67%) |
| Pigmentation | | 3 (30%) | 3 (21%) | 1 (11%) |
| Skin | (20) | (18) | (19) | (18) |
| Acanthosis | 1 (5%) | | 1 (5%) | 3 (17%) |
| Cyst epithelial inclusion | 2 (10%) | 3 (17%) | | |
| Fibrosis | | | 1 (5%) | |
| Hyperkeratosis | 1 (5%) | 1 (6%) | 1 (5%) | 3 (17%) |
| Hyperplasia, basal cell | | | | 1 (6%) |
| Inflammation, necrotizing, acute | | | | 1 (6%) |
| Subcutaneous tissue, abscess | 1 (5%) | | | 1 (6%) |
| Subcutaneous tissue, edema | 1 (5%) | | | |
| Subcutaneous tissue, inflammation, chronic active | | | 1 (5%) | |
| Subcutaneous tissue, inflammation, granulomatous | 1 (5%) | 1 (6%) | | 1 (6%) |
| Subcutaneous tissue, necrosis, coagulative | | | | 1 (6%) |
| Musculoskeletal System | | | | |
| Skeletal muscle | (1) | (2) | | |
| Hindlimb, mineralization | | 1 (50%) | | |
| Nervous System | | | | |
| Brain | (50) | (15) | (12) | (50) |
| Hemorrhage | 2 (4%) | 2 (13%) | | 2 (4%) |
| Cerebellum, infarct | 1 (2%) | | | |
| Cerebrum, degeneration, focal | | | 1 (8%) | |
| Respiratory System | | | | |
| Lung | (50) | (28) | (31) | (50) |
| Congestion | | 1 (4%) | 1 (3%) | 1 (2%) |
| Edema | | | 2 (6%) | 1 (2%) |
| Hemorrhage | 4 (8%) | 6 (21%) | 5 (16%) | 1 (2%) |
| Infiltration cellular, histiocyte | 31 (62%) | 18 (64%) | 18 (58%) | 43 (86%) |
| Inflammation, chronic active | | | 1 (3%) | |
| Metaplasia, osseous | | | 1 (3%) | 2 (4%) |
| Necrosis, coagulative | 1 (2%) | | | |
| Alveolar epithelium, hyperplasia | | 4 (14%) | 2 (6%) | 3 (6%) |
| Artery, mineralization | 34 (68%) | 13 (46%) | 17 (55%) | 43 (86%) |
| Bronchiole, epithelium, hyperplasia | | | 1 (3%) | 1 (2%) |

TABLE A5
Summary of the Incidence of Nonneoplastic Lesions in Male Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|----------|-----------|------------|------------|
| Respiratory System (continued) | | | | |
| Nose | (44) | (48) | (49) | (49) |
| Foreign body | | 1 (2%) | | |
| Metaplasia, squamous | | 1 (2%) | 2 (4%) | 1 (2%) |
| Glands, inflammation, acute | 11 (25%) | 11 (23%) | 4 (8%) | 5 (10%) |
| Lumen, hemorrhage | | 1 (2%) | | |
| Lumen, inflammation, acute | 2 (5%) | 3 (6%) | 9 (18%) | 3 (6%) |
| Mucosa, congestion | | | 1 (2%) | |
| Nasopharyngeal duct, inflammation, acute | | 1 (2%) | | |
| Nasopharyngeal duct, inflammation, chronic | 1 (2%) | | | |
| Special Senses System | | | | |
| Eye | (2) | (9) | (6) | (3) |
| Atrophy | | | | 1 (33%) |
| Synechia | | | | 1 (33%) |
| Artery, mineralization | | | 1 (17%) | |
| Cornea, fibrosis | 1 (50%) | 1 (11%) | | 1 (33%) |
| Cornea, inflammation, chronic | | 1 (11%) | | |
| Posterior chamber, inflammation, chronic | | | | 1 (33%) |
| Retina, degeneration | 1 (50%) | | | 1 (33%) |
| Sclera, metaplasia, osseous | | 1 (11%) | | |
| Harderian gland | (1) | | (1) | (1) |
| Hyperplasia | 1 (100%) | | | |
| Inflammation, chronic | | | | 1 (100%) |
| Urinary System | | | | |
| Kidney | (50) | (50) | (50) | (50) |
| Autolysis | | 2 (4%) | | |
| Congestion | 2 (4%) | | | 2 (4%) |
| Cyst | 2 (4%) | 6 (12%) | 1 (2%) | 7 (14%) |
| Hemorrhage | | 1 (2%) | 1 (2%) | |
| Hydronephrosis | | 1 (2%) | 1 (2%) | |
| Nephropathy | 48 (96%) | 50 (100%) | 50 (100%) | 49 (98%) |
| Artery, inflammation, necrotizing, chronic active | | 1 (2%) | | |
| Collecting tubule, mineralization | | | 1 (2%) | |
| Interstitial tissue, inflammation, acute | | | | 1 (2%) |
| Interstitial tissue, proximal convoluted renal tubule, inflammation, acute | | 1 (2%) | | |
| Proximal convoluted renal tubule, mineralization | | 1 (2%) | 2 (4%) | 5 (10%) |
| Proximal convoluted renal tubule, necrosis | | 1 (2%) | | 1 (2%) |
| Proximal convoluted renal tubule, epithelium, pigmentation | 5 (10%) | 2 (4%) | 3 (6%) | |
| Renal tubule, hyperplasia | 1 (2%) | 1 (2%) | 2 (4%) | 4 (8%) |
| Renal tubule, hyperplasia, cystic | | 1 (2%) | 1 (2%) | |
| Transitional epithelium, hyperplasia | 14 (28%) | 9 (18%) | 16 (32%) | 27 (54%) |
| Transitional epithelium, mineralization | 1 (2%) | | | |

TABLE A5
Summary of the Incidence of Nonneoplastic Lesions in Male Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|--------|-----------|------------|------------|
| Urinary System (continued) | | | | |
| Urethra | | | | (1) |
| Calculus micro observation only | | | | 1 (100%) |
| Urinary bladder | (50) | (49) | (49) | (48) |
| Calculus micro observation only | 2 (4%) | | | 1 (2%) |
| Inflammation, chronic | | 2 (4%) | 1 (2%) | |
| Artery, mineralization | | | 1 (2%) | |
| Serosa, mineralization | | | | 1 (2%) |
| Submucosa, hemorrhage | | 1 (2%) | | |
| Subserosa, mineralization | | 1 (2%) | 1 (2%) | |
| Transitional epithelium, hyperplasia | | | 1 (2%) | |
| Wall, mucosa, muscularis, inflammation, necrotizing, acute, diffuse | | | | 1 (2%) |

APPENDIX B

SUMMARY OF LESIONS IN FEMALE RATS IN THE 2-YEAR FEED STUDY OF QUERCETIN

| | | |
|------------------|---|------------|
| TABLE B1 | Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin | 94 |
| TABLE B2 | Individual Animal Tumor Pathology of Female Rats in the 2-Year Feed Study of Quercetin | 98 |
| TABLE B3 | Statistical Analysis of Primary Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin | 122 |
| TABLE B4a | Historical Incidence of Renal Tubule Neoplasms in Untreated Female F344/N Rats . | 127 |
| TABLE B4b | Historical Incidence of Oral Cavity Neoplasms in Untreated Female F344/N Rats . . . | 127 |
| TABLE B4c | Historical Incidence of Mammary Gland Fibroadenomas in Untreated Female F344/N Rats | 128 |
| TABLE B4d | Historical Incidence of Uterine Stromal Polyps in Untreated Female F344/N Rats | 128 |
| TABLE B5 | Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Feed Study of Quercetin | 129 |

TABLE B1
Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|--------|-----------|------------|------------|
| Disposition Summary | | | | |
| Animals initially in study | 70 | 70 | 70 | 70 |
| 6-Month interim evaluation | 10 | 10 | 10 | 10 |
| 15-Month interim evaluation | 10 | 10 | 10 | 10 |
| Early deaths | | | | |
| Natural deaths | 1 | 4 | 2 | 3 |
| Moribund | 19 | 18 | 13 | 19 |
| Survivors | | | | |
| Terminal sacrifice | 29 | 28 | 35 | 27 |
| Moribund | | | | 1 |
| Died last week of study | 1 | | | |
| Animals examined microscopically | 50 | 50 | 50 | 50 |
| Alimentary System | | | | |
| Intestine large, cecum | (50) | (11) | (8) | (48) |
| Intestine large, colon | (50) | (11) | (7) | (48) |
| Intestine large, rectum | (48) | (11) | (7) | (47) |
| Polyp adenomatous | | | | 1 (2%) |
| Intestine small, duodenum | (50) | (48) | (50) | (49) |
| Leiomyoma | | 1 (2%) | | |
| Intestine small, ileum | (49) | (48) | (49) | (49) |
| Intestine small, jejunum | (50) | (47) | (49) | (49) |
| Liver | (50) | (50) | (50) | (50) |
| Neoplastic nodule | | 1 (2%) | | 1 (2%) |
| Mesentery | (2) | (6) | (2) | (1) |
| Pancreas | (50) | (49) | (50) | (50) |
| Adenoma | | | | 1 (2%) |
| Salivary glands | (7) | (12) | (7) | (11) |
| Stomach, forestomach | (49) | (50) | (50) | (47) |
| Stomach, glandular | (50) | (49) | (50) | (50) |
| Tongue | (29) | (43) | (43) | (39) |
| Squamous cell carcinoma | | | | 2 (5%) |
| Cardiovascular System | | | | |
| Heart | (50) | (13) | (7) | (50) |
| Alveolar/bronchiolar carcinoma, metastatic, lung | | 1 (8%) | | |
| Fibrosarcoma, metastatic, skin | | | 1 (14%) | |
| Endocrine System | | | | |
| Adrenal gland | (50) | (14) | (13) | (50) |
| Adrenal gland, cortex | (50) | (13) | (13) | (50) |
| Adenoma | | | 1 (8%) | 1 (2%) |
| Fibrosarcoma, metastatic, skin | | | 1 (8%) | |
| Adrenal gland, medulla | (50) | (13) | (12) | (50) |
| Pheochromocytoma malignant | 1 (2%) | 1 (8%) | | |
| Pheochromocytoma benign | 3 (6%) | | 3 (25%) | 1 (2%) |
| Islets, pancreatic | (44) | (15) | (8) | (49) |
| Adenoma | 2 (5%) | 3 (20%) | 1 (13%) | |

TABLE B1
Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin
 (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|----------|-----------|------------|------------|
| Endocrine System (continued) | | | | |
| Parathyroid gland | (40) | (39) | (36) | (43) |
| Adenoma | 1 (3%) | | | |
| Pituitary gland | (50) | (49) | (50) | (49) |
| Pars distalis, adenoma | 32 (64%) | 27 (55%) | 26 (52%) | 25 (51%) |
| Pars distalis, adenoma, multiple | 5 (10%) | 4 (8%) | 9 (18%) | 2 (4%) |
| Pars distalis, carcinoma | | 1 (2%) | | 1 (2%) |
| Pars distalis, pars intermedia, pars nervosa, leukemia mononuclear | 1 (2%) | | | |
| Thyroid gland | (50) | (43) | (47) | (50) |
| C-cell, adenoma | 6 (12%) | 3 (7%) | 4 (9%) | 2 (4%) |
| C-cell, carcinoma | 2 (4%) | 3 (7%) | 2 (4%) | 1 (2%) |
| Follicular cell, adenoma | | | | 1 (2%) |
| General Body System | | | | |
| None | | | | |
| Genital System | | | | |
| Clitoral gland | (14) | (20) | (14) | (12) |
| Adenoma | 4 (29%) | 4 (20%) | 3 (21%) | 4 (33%) |
| Carcinoma | 1 (7%) | 1 (5%) | 2 (14%) | |
| Sarcoma | | | 1 (7%) | |
| Ovary | (50) | (17) | (15) | (48) |
| Granulosa cell tumor benign | 1 (2%) | | | |
| Granulosa-theca tumor malignant | | | | 1 (2%) |
| Uterus | (50) | (50) | (50) | (50) |
| Adenocarcinoma | | | 1 (2%) | |
| Leiomyoma | 1 (2%) | | | |
| Polyp stromal | 7 (14%) | 8 (16%) | 16 (32%) | 10 (20%) |
| Polyp stromal, multiple | | 1 (2%) | | 1 (2%) |
| Sarcoma stromal | | 2 (4%) | 1 (2%) | |
| Hematopoietic System | | | | |
| Bone marrow | (8) | (11) | (7) | (11) |
| Lymph node | (48) | (25) | (17) | (49) |
| Lumbar, fibrosarcoma, metastatic, skin | | | 1 (6%) | |
| Lymph node, mandibular | (46) | (19) | (10) | (46) |
| Lymph node, mesenteric | (9) | (14) | (12) | (9) |
| Spleen | (50) | (23) | (20) | (50) |
| Hemangioma | 1 (2%) | | | |
| Thymus | (8) | (10) | (7) | (9) |
| Alveolar/bronchiolar carcinoma, metastatic, lung | | 1 (10%) | | |

TABLE B1
Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin
 (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|----------|-----------|------------|------------|
| Integumentary System | | | | |
| Mammary gland | (36) | (36) | (24) | (22) |
| Adenocarcinoma | 1 (3%) | | 3 (13%) | 2 (9%) |
| Fibroadenoma | 21 (58%) | 17 (47%) | 14 (58%) | 6 (27%) |
| Fibroadenoma, multiple | 8 (22%) | 10 (28%) | 2 (8%) | 3 (14%) |
| Skin | (11) | (15) | (8) | (11) |
| Subcutaneous tissue, carcinosarcoma, poorly differentiated | 1 (9%) | | | |
| Subcutaneous tissue, fibroma | 2 (18%) | 3 (20%) | 1 (13%) | |
| Subcutaneous tissue, fibrosarcoma | | | 1 (13%) | |
| Subcutaneous tissue, sarcoma, poorly differentiated | | | | 1 (9%) |
| Musculoskeletal System | | | | |
| Skeletal muscle | | | | (3) |
| Sarcoma | | | | 1 (33%) |
| Nervous System | | | | |
| Brain | (50) | (15) | (8) | (50) |
| Carcinoma, extension, metastatic, pituitary gland | | | | 1 (2%) |
| Medulla, carcinoma, metastatic, pituitary gland | | 1 (7%) | | |
| Spinal cord | (1) | (2) | | (2) |
| Respiratory System | | | | |
| Lung | (50) | (20) | (20) | (50) |
| Alveolar/bronchiolar adenoma | 5 (10%) | | 1 (5%) | |
| Alveolar/bronchiolar carcinoma | | 1 (5%) | | |
| Carcinosarcoma, metastatic, skin | 1 (2%) | | | |
| Granulosa-theca tumor malignant, metastatic, ovary | | | | 1 (2%) |
| Hepatocellular carcinoma, metastatic, uncertain primary site | | | | 1 (2%) |
| Pheochromocytoma malignant, metastatic | | 1 (5%) | | |
| Sarcoma, metastatic, lung | | 1 (5%) | | |
| Sarcoma, poorly differentiated, metastatic, skin | | | | 1 (2%) |
| Pleura, alveolar/bronchiolar carcinoma, metastatic, lung | | 1 (5%) | | |
| Nose | (7) | (12) | (7) | (10) |
| Special Senses System | | | | |
| Ear | (1) | (2) | (1) | |
| Fibrosarcoma | 1 (100%) | | | |
| Zymbal's gland | (2) | (2) | | |
| Adenoma | 1 (50%) | | | |
| Squamous cell carcinoma | 1 (50%) | 2 (100%) | | |

TABLE B1
Summary of the Incidence of Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin
 (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|---------|-----------|------------|------------|
| Urinary System | | | | |
| Kidney | (49) | (49) | (50) | (50) |
| Renal tubule, adenoma | | | 1 (2%) | |
| Urinary bladder | (50) | (49) | (50) | (50) |
| Papilloma | 1 (2%) | | | |
| Systemic Lesions | | | | |
| Multiple organs ^a | (50) | (50) | (50) | (50) |
| Leukemia monocytic | | | | 1 (2%) |
| Leukemia mononuclear | 9 (18%) | 10 (20%) | 13 (26%) | 12 (24%) |
| Tumor Summary | | | | |
| Total animals with primary neoplasms ^b | 49 | 48 | 43 | 44 |
| Total primary neoplasms | 118 | 103 | 106 | 81 |
| Total animals with benign neoplasms | 49 | 44 | 42 | 35 |
| Total benign neoplasms | 101 | 82 | 82 | 59 |
| Total animals with malignant neoplasms | 15 | 19 | 19 | 19 |
| Total malignant neoplasms | 17 | 21 | 24 | 22 |
| Total animals with metastatic neoplasms | 1 | 4 | 1 | 4 |
| Total metastatic neoplasms | 1 | 6 | 3 | 4 |
| Total animals with malignant neoplasms of uncertain primary site | | | | 1 |

^a Number of animals with any tissue examined microscopically

^b Primary neoplasms: all neoplasms except metastatic neoplasms

TABLE B2
Individual Animal Tumor Pathology of Female Rats in the 2-Year Feed Study of Quercetin: 0 ppm (continued)

| Number of Days on Study | 7 | 2 2 2 2 2 2 2 2 2 2 2 2 2 2 3 3 3 3 3 3 3 3 3 3 3 | 4 4 4 4 4 5 5 8 9 9 9 9 9 9 1 1 1 1 1 1 1 1 2 2 2 2 2 |
|----------------------------------|---|---|---|
| Carcass ID Number | 0 | 5 6 6 6 6 5 6 6 5 6 6 7 5 5 5 6 6 6 7 5 6 6 6 6 6 | 9 4 5 6 9 8 6 2 9 1 5 6 0 7 9 9 3 5 8 0 8 1 2 3 6 |
| | 4 1 4 4 1 2 3 1 2 1 2 1 2 1 1 3 1 1 1 1 1 1 2 2 2 2 | | Total Tissues/Tumors |
| Alimentary System | | | |
| Esophagus | + | + | 50 |
| Intestine large | + | + | 50 |
| Intestine large, cecum | + | + | 50 |
| Intestine large, colon | + | + | 50 |
| Intestine large, rectum | + | + | 48 |
| Intestine small | + | + | 50 |
| Intestine small, duodenum | + | + | 50 |
| Intestine small, ileum | + | + | 49 |
| Intestine small, jejunum | + | + | 50 |
| Liver | + | + | 50 |
| Mesentery | | | 2 |
| Pancreas | + | + | 50 |
| Salivary glands | | | 7 |
| Stomach | + | + | 50 |
| Stomach, forestomach | + | + | 49 |
| Stomach, glandular | + | + | 50 |
| Tongue | | | 29 |
| Cardiovascular System | | | |
| Heart | + | + | 50 |
| Endocrine System | | | |
| Adrenal gland | + | + | 50 |
| Adrenal gland, cortex | + | + | 50 |
| Adrenal gland, medulla | + | + | 50 |
| Pheochromocytoma malignant | | X | 1 |
| Pheochromocytoma benign | | X | 3 |
| Islets, pancreatic | + | + | 44 |
| Adenoma | | | 2 |
| Parathyroid gland | + | + | 40 |
| Adenoma | | | 1 |
| Pituitary gland | + | + | 50 |
| Pars distalis, adenoma | X | X | 32 |
| Pars distalis, adenoma, multiple | | | 5 |
| Thyroid gland | + | + | 50 |
| C-cell, adenoma | | | 6 |
| C-cell, carcinoma | | | 2 |

TABLE B2
Individual Animal Tumor Pathology of Female Rats in the 2-Year Feed Study of Quercetin: 0 ppm (continued)

| | | |
|----------------------------------|---|--------------------------------------|
| Number of Days on Study | 7 | |
| | 2 2 2 2 2 2 2 2 2 2 2 2 2 3 3 3 3 3 3 3 3 3 3 | |
| | 4 4 4 4 4 5 5 8 9 9 9 9 9 1 1 1 1 1 1 1 2 2 2 2 | |
| Carcass ID Number | 0 | Total Tissues/ Tumors |
| | 5 6 6 6 6 5 6 6 5 6 6 6 7 5 5 5 6 6 6 7 5 6 6 6 | |
| | 9 4 5 6 9 8 6 2 9 1 5 6 0 7 9 9 3 5 8 0 8 1 2 3 6 | |
| | 4 1 4 4 1 2 3 1 2 1 2 1 2 1 1 3 1 1 1 1 1 2 2 2 2 | |
| Respiratory System | | |
| Lung | + | 50 |
| Alveolar/bronchiolar adenoma | X | 5 |
| Carcinosarcoma, metastatic, skin | | 1 |
| Nose | | 7 |
| Trachea | + | 49 |
| Special Senses System | | |
| Ear | | 1 |
| Fibrosarcoma | | 1 |
| Eye | + + | 6 |
| Zymbal's gland | | 2 |
| Adenoma | | 1 |
| Squamous cell carcinoma | | 1 |
| Urinary System | | |
| Kidney | + | 49 |
| Urinary bladder | + | 50 |
| Papilloma | | 1 |
| Systemic Lesions | | |
| Multiple organs | + | 50 |
| Leukemia mononuclear | | 9 |

TABLE B2
Individual Animal Tumor Pathology of Female Rats in the 2-Year Feed Study of Quercetin: 1,000 ppm
 (continued)

| | |
|--|---|
| Number of Days on Study | 1 4 5 5 5 5 5 6 6 6 6 6 6 6 6 6 6 6 6 6 7 7 7 7 7 |
| | 8 9 0 1 4 9 9 0 0 1 1 2 4 6 7 8 8 8 8 9 0 2 2 2 2 |
| | 3 7 4 5 3 6 7 3 3 2 3 6 5 8 6 0 0 3 7 6 4 1 3 3 3 |
| Carcass ID Number | 0 |
| | 8 7 8 7 7 8 7 7 7 7 7 7 7 7 8 7 7 7 7 8 7 8 7 7 7 |
| | 0 6 3 3 8 1 2 2 2 3 1 3 5 1 3 1 3 7 8 2 2 0 3 5 6 |
| | 3 5 4 5 5 4 5 2 3 4 5 3 4 2 3 4 2 4 4 5 1 2 1 3 2 |
| General Body System | |
| None | |
| Genital System | |
| Clitoral gland | + + + + + + + + + + + |
| Adenoma | |
| Carcinoma | |
| Ovary | + + + + + + + + + + + M + + |
| Uterus | + |
| Polyp stromal | X |
| Polyp stromal, multiple | |
| Sarcoma stromal | X X X |
| Hematopoietic System | |
| Blood | |
| Bone marrow | + + A + + + + + + + + + + |
| Lymph node | + + + + + + + + + + + + + + + + + |
| Lymph node, mandibular | + + + + + + + + + + + + + + + + + |
| Lymph node, mesenteric | + + + + + + + + + + + + + + + + + |
| Spleen | + + + + + + + + + + + + + + + + + |
| Thymus | + + M + + + + + + + + M M + + |
| Alveolar/bronchiolar carcinoma, metastatic, lung | X |
| Integumentary System | |
| Mammary gland | + + + + + + + + + + + + + + + + + + |
| Fibroadenoma | X X X X X X X X X X |
| Fibroadenoma, multiple | |
| Skin | + + + + + + + + + + + + + + + + + |
| Subcutaneous tissue, fibroma | X |
| Musculoskeletal System | |
| Bone | + + + + + + + + + + + + + + + |
| Nervous System | |
| Brain | + + + + + + + + + + + + + + + |
| Medulla, carcinoma, metastatic, pituitary gland | X |
| Spinal cord | |

TABLE B2
Individual Animal Tumor Pathology of Female Rats in the 2-Year Feed Study of Quercetin: 1,000 ppm
 (continued)

| | |
|--|---|
| Number of Days on Study | 1 4 5 5 5 5 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 7 7 7 7 7 |
| | 8 9 0 1 4 9 9 0 0 1 1 2 4 6 7 8 8 8 8 9 0 2 2 2 2 |
| | 3 7 4 5 3 6 7 3 3 2 3 6 5 8 6 0 0 3 7 6 4 1 3 3 3 |
| Carcass ID Number | 0 |
| | 8 7 8 7 7 8 7 7 7 7 7 7 7 8 7 7 7 7 8 7 8 7 7 7 |
| | 0 6 3 3 8 1 2 2 2 3 1 3 5 1 3 1 3 7 8 2 2 0 3 5 6 |
| | 3 5 4 5 5 4 5 2 3 4 5 3 4 2 3 4 2 4 4 5 1 2 1 3 2 |
| Respiratory System | |
| Lung | + |
| Alveolar/bronchiolar carcinoma | X |
| Pheochromocytoma malignant, metastatic | X |
| Sarcoma, metastatic, lung | |
| Pleura, alveolar/bronchiolar carcinoma, metastatic, lung | X |
| Nose | + |
| Trachea | + |
| Special Senses System | |
| Ear | |
| Eye | + I + I I I |
| Harderian gland | + + |
| Zymbal's gland | + + |
| Squamous cell carcinoma | X X |
| Urinary System | |
| Kidney | + + A + |
| Urinary bladder | + |
| Systemic Lesions | |
| Multiple organs | + |
| Leukemia mononuclear | X X X X X X X X |

TABLE B2
Individual Animal Tumor Pathology of Female Rats in the 2-Year Feed Study of Quercetin: 40,000 ppm
 (continued)

| | | |
|---|---|--------------------------------------|
| Number of Days on Study | 7 | |
| | 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 3 3 3 3 3 3 3 3 | |
| | 3 3 3 3 3 3 4 4 4 4 4 4 4 4 5 5 9 1 1 1 1 1 1 1 | |
| Carcass ID Number | 1 1 1 1 1 1 0 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 | Total Tissues/ Tumors |
| | 0 0 0 0 1 1 9 0 0 0 0 0 0 0 0 0 1 0 0 0 0 0 0 1 1 | |
| | 7 7 8 9 2 2 9 0 2 3 4 4 6 9 1 2 0 2 2 3 3 4 6 0 2 | |
| | 1 2 2 2 1 2 1 3 4 2 2 4 2 3 4 3 2 1 2 1 3 1 1 1 3 | |
| Respiratory System | | |
| Lung | + | 50 |
| Granulosa-theca tumor malignant, metastatic, ovary | | 1 |
| Hepatocellular carcinoma, metastatic, uncertain primary site | | 1 |
| Sarcoma, poorly differentiated, metastatic, skin | | 1 |
| Nose | | 10 |
| Trachea | + | 50 |
| Special Senses System | | |
| Eye | | 5 |
| Harderian gland | | 2 |
| Urinary System | | |
| Kidney | + | 50 |
| Urinary bladder | + | 50 |
| Systemic Lesions | | |
| Multiple organs | + | 50 |
| Leukemia monocytic | | 1 |
| Leukemia mononuclear | | 12 |

TABLE B3
Statistical Analysis of Primary Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|-------------|------------------------|-------------------------|----------------|
| Adrenal Medulla: Pheochromocytoma (Benign or Malignant) | | | | |
| Overall rates ^a | 3/50 (6%) | 1/13 (8%) ^e | 3/12 (25%) ^e | 1/50 (2%) |
| Adjusted rates ^b | 7.8% | | | 3.6% |
| Terminal rates ^c | 1/30 (3%) | | | 1/28 (4%) |
| First incidence (days) | 625 | | | 723 (T) |
| Life table tests ^d | | | | P=0.340N |
| Logistic regression tests ^d | | | | P=0.305N |
| Fisher exact test ^d | | | | P=0.309N |
| Lung: Alveolar/bronchiolar Adenoma or Carcinoma | | | | |
| Overall rates | 5/50 (10%) | 1/20 (5%) ^e | 1/20 (5%) ^e | 0/50 (0%) |
| Adjusted rates | 14.3% | | | 0.0% |
| Terminal rates | 3/30 (10%) | | | 0/28 (0%) |
| First incidence (days) | 641 | | | - _f |
| Life table tests | | | | P=0.044N |
| Logistic regression tests | | | | P=0.036N |
| Fisher exact test | | | | P=0.028N |
| Mammary Gland: Fibroadenoma | | | | |
| Overall rates | 29/50 (58%) | 27/50 (54%) | 16/50 (32%) | 9/50 (18%) |
| Adjusted rates | 66.4% | 72.3% | 38.4% | 30.1% |
| Terminal rates | 16/30 (53%) | 18/28 (64%) | 10/35 (29%) | 8/28 (29%) |
| First incidence (days) | 597 | 597 | 605 | 549 |
| Life table tests | P<0.001N | P=0.531 | P=0.008N | P<0.001N |
| Logistic regression tests | P<0.001N | P=0.553N | P=0.008N | P<0.001N |
| Cochran-Armitage test ^d | P<0.001N | | | |
| Fisher exact test | | P=0.420N | P=0.008N | P<0.001N |
| Mammary Gland: Adenocarcinoma | | | | |
| Overall rates | 1/50 (2%) | 0/50 (0%) | 3/50 (6%) | 2/50 (4%) |
| Adjusted rates | 2.5% | 0.0% | 7.9% | 6.0% |
| Terminal rates | 0/30 (0%) | 0/28 (0%) | 2/35 (6%) | 0/28 (0%) |
| First incidence (days) | 672 | - | 660 | 686 |
| Life table tests | P=0.299 | P=0.521N | P=0.340 | P=0.468 |
| Logistic regression tests | P=0.309 | P=0.492N | P=0.304 | P=0.492 |
| Cochran-Armitage test | P=0.318 | | | |
| Fisher exact test | | P=0.500N | P=0.309 | P=0.500 |
| Mammary Gland: Fibroadenoma or Adenocarcinoma | | | | |
| Overall rates | 30/50 (60%) | 27/50 (54%) | 17/50 (34%) | 11/50 (22%) |
| Adjusted rates | 67.2% | 72.3% | 40.8% | 34.3% |
| Terminal rates | 16/30 (53%) | 18/28 (64%) | 11/35 (31%) | 8/28 (29%) |
| First incidence (days) | 597 | 597 | 605 | 549 |
| Life table tests | P<0.001N | P=0.538N | P=0.008N | P=0.002N |
| Logistic regression tests | P<0.001N | P=0.468N | P=0.009N | P<0.001N |
| Cochran-Armitage test | P<0.001N | | | |
| Fisher exact test | | P=0.343N | P=0.008N | P<0.001N |

TABLE B3
Statistical Analysis of Primary Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin
 (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|-------------|-------------------------|------------------------|-------------|
| Pancreatic Islets: Adenoma | | | | |
| Overall rates | 2/44 (5%) | 3/15 (20%) ^e | 1/8 (13%) ^e | 0/49 (0%) |
| Adjusted rates | 6.3% | | | 0.0% |
| Terminal rates | 1/27 (4%) | | | 0/28 (0%) |
| First incidence (days) | 687 | | | - |
| Life table tests | | | | P=0.245N |
| Logistic regression tests | | | | P=0.225N |
| Fisher exact test | | | | P=0.221N |
| Pituitary Gland (Pars Distalis): Adenoma | | | | |
| Overall rates | 37/50 (74%) | 31/49 (63%) | 35/50 (70%) | 27/49 (55%) |
| Adjusted rates | 80.3% | 74.7% | 77.5% | 68.5% |
| Terminal rates | 21/30 (70%) | 18/28 (64%) | 25/35 (71%) | 16/28 (57%) |
| First incidence (days) | 597 | 183 | 441 | 549 |
| Life table tests | P=0.166N | P=0.380N | P=0.218N | P=0.157N |
| Logistic regression tests | P=0.064N | P=0.185N | P=0.441N | P=0.062N |
| Cochran-Armitage test | P=0.056N | | | |
| Fisher exact test | | P=0.175N | P=0.412N | P=0.039N |
| Pituitary Gland (Pars Distalis): Adenoma or Carcinoma | | | | |
| Overall rates | 37/50 (74%) | 32/49 (65%) | 35/50 (70%) | 28/49 (57%) |
| Adjusted rates | 80.3% | 75.3% | 77.5% | 71.2% |
| Terminal rates | 21/30 (70%) | 18/28 (64%) | 25/35 (71%) | 17/28 (61%) |
| First incidence (days) | 597 | 183 | 441 | 549 |
| Life table tests | P=0.197N | P=0.446N | P=0.218N | P=0.199N |
| Logistic regression tests | P=0.083N | P=0.239N | P=0.441N | P=0.094N |
| Cochran-Armitage test | P=0.073N | | | |
| Fisher exact test | | P=0.235N | P=0.412N | P=0.060N |
| Skin (Subcutaneous Tissue): Fibroma | | | | |
| Overall rates | 2/50 (4%) | 3/50 (6%) | 1/50 (2%) | 0/50 (0%) |
| Adjusted rates | 6.3% | 10.2% | 2.1% | 0.0% |
| Terminal rates | 1/30 (3%) | 2/28 (7%) | 0/35 (0%) | 0/28 (0%) |
| First incidence (days) | 718 | 704 | 521 | - |
| Life table tests | P=0.114N | P=0.461 | P=0.475N | P=0.266N |
| Logistic regression tests | P=0.108N | P=0.449 | P=0.470N | P=0.256N |
| Cochran-Armitage test | P=0.107N | | | |
| Fisher exact test | | P=0.500 | P=0.500N | P=0.247N |
| Skin (Subcutaneous Tissue): Fibroma or Fibrosarcoma | | | | |
| Overall rates | 2/50 (4%) | 3/50 (6%) | 2/50 (4%) | 0/50 (0%) |
| Adjusted rates | 6.3% | 10.2% | 4.1% | 0.0% |
| Terminal rates | 1/30 (3%) | 2/28 (7%) | 0/35 (0%) | 0/28 (0%) |
| First incidence (days) | 718 | 704 | 441 | - |
| Life table tests | P=0.120N | P=0.461 | P=0.672N | P=0.266N |
| Logistic regression tests | P=0.105N | P=0.449 | P=0.608N | P=0.256N |
| Cochran-Armitage test | P=0.112N | | | |
| Fisher exact test | | P=0.500 | P=0.691N | P=0.247N |

TABLE B3
Statistical Analysis of Primary Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin
 (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|------------|------------|------------|------------|
| Skin (Subcutaneous Tissue): Fibroma, Fibrosarcoma, or Sarcoma | | | | |
| Overall rates | 2/50 (4%) | 3/50 (6%) | 2/50 (4%) | 1/50 (2%) |
| Adjusted rates | 6.3% | 10.2% | 4.1% | 2.0% |
| Terminal rates | 1/30 (3%) | 2/28 (7%) | 0/35 (0%) | 0/28 (0%) |
| First incidence (days) | 718 | 704 | 441 | 284 |
| Life table tests | P=0.313N | P=0.461 | P=0.672N | P=0.527N |
| Logistic regression tests | P=0.262N | P=0.449 | P=0.608N | P=0.339N |
| Cochran-Armitage test | P=0.297N | | | |
| Fisher exact test | | P=0.500 | P=0.691N | P=0.500N |
| Thyroid Gland (C-cell): Adenoma | | | | |
| Overall rates | 6/50 (12%) | 3/43 (7%) | 4/47 (9%) | 2/50 (4%) |
| Adjusted rates | 20.0% | 12.5% | 11.8% | 6.4% |
| Terminal rates | 6/30 (20%) | 3/24 (13%) | 3/32 (9%) | 1/28 (4%) |
| First incidence (days) | 723 (T) | 723 (T) | 709 | 688 |
| Life table tests | P=0.179N | P=0.358N | P=0.325N | P=0.154N |
| Logistic regression tests | P=0.171N | P=0.358N | P=0.351N | P=0.161N |
| Cochran-Armitage test | P=0.162N | | | |
| Fisher exact test | | P=0.324N | P=0.410N | P=0.134N |
| Thyroid Gland (C-cell): Carcinoma | | | | |
| Overall rates | 2/50 (4%) | 3/43 (7%) | 2/47 (4%) | 1/50 (2%) |
| Adjusted rates | 5.6% | 12.5% | 5.7% | 3.6% |
| Terminal rates | 1/30 (3%) | 3/24 (13%) | 1/32 (3%) | 1/28 (4%) |
| First incidence (days) | 660 | 723 (T) | 707 | 723 (T) |
| Life table tests | P=0.294N | P=0.408 | P=0.666N | P=0.528N |
| Logistic regression tests | P=0.282N | P=0.371 | P=0.669 | P=0.521N |
| Cochran-Armitage test | P=0.271N | | | |
| Fisher exact test | | P=0.428 | P=0.668 | P=0.500N |
| Thyroid Gland (C-cell): Adenoma or Carcinoma | | | | |
| Overall rates | 8/50 (16%) | 6/43 (14%) | 6/47 (13%) | 3/50 (6%) |
| Adjusted rates | 25.2% | 25.0% | 17.1% | 9.9% |
| Terminal rates | 7/30 (23%) | 6/24 (25%) | 4/32 (13%) | 2/28 (7%) |
| First incidence (days) | 660 | 723 (T) | 707 | 688 |
| Life table tests | P=0.095N | P=0.561N | P=0.334N | P=0.123N |
| Logistic regression tests | P=0.087N | P=0.609 | P=0.408N | P=0.127N |
| Cochran-Armitage test | P=0.082N | | | |
| Fisher exact test | | P=0.508N | P=0.436N | P=0.100N |
| Tongue: Squamous Cell Carcinoma | | | | |
| Overall rates | 0/50 (0%) | 0/50 (0%) | 0/50 (0%) | 2/50 (4%) |
| Adjusted rates | 0.0% | 0.0% | 0.0% | 4.7% |
| Terminal rates | 0/30 (0%) | 0/28 (0%) | 0/35 (0%) | 0/28 (0%) |
| First incidence (days) | - | - | - | 492 |
| Life table tests | P=0.039 | - | - | P=0.227 |
| Logistic regression tests | P=0.047 | - | - | P=0.327 |
| Cochran-Armitage test | P=0.042 | | | |
| Fisher exact test | | - | - | P=0.247 |

TABLE B3
Statistical Analysis of Primary Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin
 (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|-------------|-------------|-------------|-------------|
| Uterus: Stromal Polyp | | | | |
| Overall rates | 7/50 (14%) | 9/50 (18%) | 16/50 (32%) | 11/50 (22%) |
| Adjusted rates | 18.8% | 27.7% | 41.4% | 33.2% |
| Terminal rates | 4/30 (13%) | 6/28 (21%) | 13/35 (37%) | 8/28 (29%) |
| First incidence (days) | 597 | 515 | 521 | 284 |
| Life table tests | P=0.262 | P=0.333 | P=0.059 | P=0.178 |
| Logistic regression tests | P=0.308 | P=0.387 | P=0.028 | P=0.265 |
| Cochran-Armitage test | P=0.314 | | | |
| Fisher exact test | | P=0.393 | P=0.028 | P=0.218 |
| Uterus: Stromal Polyp or Stromal Sarcoma | | | | |
| Overall rates | 7/50 (14%) | 11/50 (22%) | 17/50 (34%) | 11/50 (22%) |
| Adjusted rates | 18.8% | 31.2% | 44.0% | 33.2% |
| Terminal rates | 4/30 (13%) | 6/28 (21%) | 14/35 (40%) | 8/28 (29%) |
| First incidence (days) | 597 | 515 | 521 | 284 |
| Life table tests | P=0.361 | P=0.180 | P=0.040 | P=0.178 |
| Logistic regression tests | P=0.419 | P=0.233 | P=0.017 | P=0.265 |
| Cochran-Armitage test | P=0.420 | | | |
| Fisher exact test | | P=0.218 | P=0.017 | P=0.218 |
| All Organs: Leukemia (Monocytic or Mononuclear) | | | | |
| Overall rates | 9/50 (18%) | 10/50 (20%) | 13/50 (26%) | 12/50 (24%) |
| Adjusted rates | 22.8% | 26.3% | 29.6% | 32.6% |
| Terminal rates | 3/30 (10%) | 3/28 (11%) | 5/35 (14%) | 5/28 (18%) |
| First incidence (days) | 422 | 504 | 548 | 623 |
| Life table tests | P=0.286 | P=0.420 | P=0.323 | P=0.264 |
| Logistic regression tests | P=0.336 | P=0.586 | P=0.258 | P=0.331 |
| Cochran-Armitage test | P=0.322 | | | |
| Fisher exact test | | P=0.500 | P=0.235 | P=0.312 |
| All Organs: Benign Tumors | | | | |
| Overall rates | 49/50 (98%) | 44/50 (88%) | 42/50 (84%) | 35/50 (70%) |
| Adjusted rates | 98.0% | 93.5% | 87.5% | 84.9% |
| Terminal rates | 29/30 (97%) | 25/28 (89%) | 29/35 (83%) | 22/28 (79%) |
| First incidence (days) | 422 | 183 | 441 | 284 |
| Life table tests | P=0.052N | P=0.503N | P=0.041N | P=0.060N |
| Logistic regression tests | P<0.001N | P=0.069N | P=0.021N | P<0.001N |
| Cochran-Armitage test | P<0.001N | | | |
| Fisher exact test | | P=0.056N | P=0.015N | P<0.001N |
| All Organs: Malignant Tumors | | | | |
| Overall rates | 15/50 (30%) | 19/50 (38%) | 19/50 (38%) | 20/50 (40%) |
| Adjusted rates | 36.4% | 44.0% | 42.5% | 46.1% |
| Terminal rates | 6/30 (20%) | 6/28 (21%) | 10/35 (29%) | 6/28 (21%) |
| First incidence (days) | 422 | 183 | 441 | 284 |
| Life table tests | P=0.255 | P=0.213 | P=0.396 | P=0.175 |
| Logistic regression tests | P=0.341 | P=0.437 | P=0.284 | P=0.288 |
| Cochran-Armitage test | P=0.283 | | | |
| Fisher exact test | | P=0.263 | P=0.263 | P=0.201 |

TABLE B3
Statistical Analysis of Primary Neoplasms in Female Rats in the 2-Year Feed Study of Quercetin
 (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|-------------|-------------|-------------|-------------|
| All Organs: Benign or Malignant Tumors | | | | |
| Overall rates | 49/50 (98%) | 48/50 (96%) | 43/50 (86%) | 45/50 (90%) |
| Adjusted rates | 98.0% | 96.0% | 87.8% | 93.7% |
| Terminal rates | 29/30 (97%) | 26/28 (93%) | 29/35 (83%) | 25/28 (89%) |
| First incidence (days) | 422 | 183 | 441 | 284 |
| Life table tests | P=0.492N | P=0.373 | P=0.059N | P=0.541N |
| Logistic regression tests | P=0.140N | P=0.476N | P=0.035N | P=0.107N |
| Cochran-Armitage test | P=0.138N | | | |
| Fisher exact test | | P=0.500N | P=0.030N | P=0.102N |

(T) Terminal sacrifice

^a Number of tumor-bearing animals/number of animals examined. Denominator is number of animals examined microscopically for adrenal gland, bone marrow, brain, clitoral gland, epididymis, gallbladder (mouse), heart, kidney, larynx, liver, lung, nose, ovary, pancreas, parathyroid gland, pituitary gland, preputial gland, prostate gland, salivary gland, spleen, testes, thyroid gland, and urinary bladder; for other tissues, denominator is number of animals necropsied.

^b Kaplan-Meier estimated tumor incidence at the end of the study after adjustment for intercurrent mortality

^c Observed incidence at terminal kill

^d Beneath the control incidence are the P values associated with the trend test. Beneath the dosed group incidence are the P values corresponding to pairwise comparisons between the controls and that dosed group. The life table analysis regards tumors in animals dying prior to terminal kill as being (directly or indirectly) the cause of death. The logistic regression tests regard these lesions as nonfatal. The Cochran-Armitage and Fisher exact tests compare directly the overall incidence rates. For all tests, a negative trend or a lower incidence in a dose group is indicated by N.

^e Tissue was examined microscopically only when it was observed to be abnormal at necropsy; thus statistical comparisons with the controls are not appropriate.

^f Not applicable; no tumors in animal group.

TABLE B4a
Historical Incidence of Renal Tubule Neoplasms in Untreated Female F344/N Rats^a

| Study | Incidence in Controls | | |
|--|-----------------------|----------------|---------------------------|
| | Adenoma | Adenocarcinoma | Adenoma or Adenocarcinoma |
| Historical Incidence at EG&G Mason Research Institute | | | |
| 4-Hydroxyacetanilide | 0/50 | 0/50 | 0/50 |
| Pentaerythritol tetranitrate | 0/50 | 0/50 | 0/50 |
| Overall Historical Incidence | | | |
| Total | 1/499 (0.2%) | 0/499 (0.0%) | 1/499 (0.2%) |
| Standard deviation | 0.6% | | 0.6% |
| Range | 0%-2% | | 0%-2% |

^a Data as of 17 September 1990

TABLE B4b
Historical Incidence of Oral Cavity Neoplasms in Untreated Female F344/N Rats^a

| Study | Incidence in Controls | | |
|--|---|--------------------------------------|---|
| | Oral Mucosa: Papilloma or Squamous Cell Papilloma | Oral Mucosa: Squamous Cell Carcinoma | Oral Mucosa: Papilloma, Squamous Cell Papilloma, or Carcinoma |
| Historical Incidence at EG&G Mason Research Institute | | | |
| 4-Hydroxyacetanilide | 0/50 | 0/50 | 0/50 |
| Pentaerythritol tetranitrate | 0/50 | 0/50 | 1/50 |
| Total | | | 1/100 (1%) |
| Overall Historical Incidence | | | |
| Total | 3/500 (0.6%) | 0/500 | 4/500 (0.8%) |
| Standard deviation | 1.0% | | 1.0% |
| Range | 0%-2% | | 0%-2% |

^a Data as of 17 September 1990; includes oral mucosa, tongue, pharynx (palate), tooth (gingiva), and lip

TABLE B4c
Historical Incidence of Mammary Gland Fibroadenomas in Untreated Female F344/N Rats^a

| Study | Incidence in Controls |
|--|------------------------------|
| Historical Incidence at EG&G Mason Research Institute | |
| 4-Hydroxyacetanilide | 19/50 |
| Pentaerythritol tetranitrate | 27/50 |
| Total | 46/100 (46.0%) |
| Overall Historical Incidence | |
| Total | 178/500 (35.6%) |
| Standard deviation | 15.0% |
| Range | 8%-56% |

^a Data as of 17 September 1990

TABLE B4d
Historical Incidence of Uterine Stromal Polyps in Untreated Female F344/N Rats^a

| Study | Incidence in Controls |
|--|------------------------------|
| Historical Incidence at EG&G Mason Research Institute | |
| 4-Hydroxyacetanilide | 15/50 |
| Pentaerythritol tetranitrate | 8/50 |
| Total | 23/100 (23.0%) |
| Overall Historical Incidence | |
| Total | 94/500 (19.6%) |
| Standard deviation | 5.4% |
| Range | 12%-30% |

^a Data as of 17 September 1990

TABLE B5
Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Feed Study of Quercetin

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|----------------------------------|----------|-----------|------------|------------|
| Disposition Summary | | | | |
| Animals initially in study | 70 | 70 | 70 | 70 |
| 6-Month interim evaluations | 10 | 10 | 10 | 10 |
| 15-Month interim evaluations | 10 | 10 | 10 | 10 |
| Early deaths | | | | |
| Natural deaths | 1 | 4 | 2 | 3 |
| Moribund | 19 | 18 | 13 | 19 |
| Survivors | | | | |
| Terminal sacrifice | 29 | 28 | 35 | 27 |
| Moribund | | | | 1 |
| Died last week of study | 1 | | | |
| Animals examined microscopically | 50 | 50 | 50 | 50 |
| Alimentary System | | | | |
| Intestine large, cecum | (50) | (11) | (8) | (48) |
| Necrosis, coagulative, acute | | | | 1 (2%) |
| Parasite metazoan | 2 (4%) | | | 4 (8%) |
| Lymphoid tissue, hypoplasia | | 1 (9%) | | |
| Intestine large, colon | (50) | (11) | (7) | (48) |
| Parasite metazoan | 9 (18%) | 1 (9%) | 2 (29%) | 5 (10%) |
| Intestine large, rectum | (48) | (11) | (7) | (47) |
| Parasite metazoan | 5 (10%) | | | 2 (4%) |
| Intestine small, duodenum | (50) | (48) | (50) | (49) |
| Autolysis | | | 1 (2%) | |
| Epithelium, pigmentation | | | | 1 (2%) |
| Intestine small, ileum | (49) | (48) | (49) | (49) |
| Epithelium, pigmentation | | | 19 (39%) | 32 (65%) |
| Serosa, fibrosis | | 1 (2%) | | |
| Intestine small, jejunum | (50) | (47) | (49) | (49) |
| Autolysis | | 1 (2%) | 1 (2%) | |
| Epithelium, pigmentation | | | 3 (6%) | 20 (41%) |
| Liver | (50) | (50) | (50) | (50) |
| Angiectasis | 1 (2%) | 5 (10%) | 1 (2%) | |
| Basophilic focus | 40 (80%) | 38 (76%) | 41 (82%) | 39 (78%) |
| Clear cell focus | 4 (8%) | 4 (8%) | 4 (8%) | 2 (4%) |
| Cyst | | | | 2 (4%) |
| Cyst multilocular | | | 1 (2%) | |
| Cytoplasmic alteration | | 4 (8%) | | 1 (2%) |
| Degeneration | | 1 (2%) | 1 (2%) | |
| Developmental malformation | 3 (6%) | | | 2 (4%) |
| Eosinophilic focus | 3 (6%) | | | 3 (6%) |
| Fatty change | 14 (28%) | 12 (24%) | 12 (24%) | 7 (14%) |
| Fibrosis, focal | | | | 1 (2%) |
| Granuloma | | | 1 (2%) | |
| Hematopoietic cell proliferation | | 1 (2%) | | |
| Hemorrhage | | 1 (2%) | 1 (2%) | |
| Hepatodiaphragmatic nodule | | 3 (6%) | 5 (10%) | |

TABLE B5
Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|----------|-----------|------------|------------|
| Alimentary System (continued) | | | | |
| Liver (continued) | | | | |
| Infarct | | | | 1 (2%) |
| Inflammation, acute | | | 1 (2%) | |
| Inflammation, chronic | 1 (2%) | | 3 (6%) | 3 (6%) |
| Inflammation, chronic active | 2 (4%) | 2 (4%) | 2 (4%) | 7 (14%) |
| Inflammation, granulomatous, chronic | 12 (24%) | 17 (34%) | 10 (20%) | 8 (16%) |
| Mixed cell focus | 4 (8%) | 7 (14%) | 4 (8%) | 1 (2%) |
| Necrosis, coagulative | 4 (8%) | 5 (10%) | 5 (10%) | 8 (16%) |
| Pigmentation | 1 (2%) | 1 (2%) | 1 (2%) | |
| Bile duct, cyst multilocular | | | | 1 (2%) |
| Bile duct, hyperplasia | 17 (34%) | 14 (28%) | 17 (34%) | 15 (30%) |
| Centrilobular, necrosis, coagulative | | | | 1 (2%) |
| Serosa, fibrosis | | 1 (2%) | | |
| Serosa, hemorrhage | | 1 (2%) | | |
| Mesentery | (2) | (6) | (2) | (1) |
| Fibrosis | | 6 (100%) | 1 (50%) | |
| Inflammation, chronic | | 3 (50%) | | |
| Inflammation, chronic active | | 2 (33%) | | |
| Inflammation, granulomatous, chronic | 1 (50%) | 1 (17%) | 1 (50%) | |
| Mineralization | | 1 (17%) | 1 (50%) | |
| Necrosis, coagulative | 2 (100%) | 3 (50%) | | |
| Pigmentation | | | 1 (50%) | |
| Pancreas | (50) | (49) | (50) | (50) |
| Atrophy | 21 (42%) | 21 (43%) | 21 (42%) | 15 (30%) |
| Cytoplasmic alteration | | 2 (4%) | 4 (8%) | 2 (4%) |
| Ectopic liver | | | | 1 (2%) |
| Inflammation, chronic | 17 (34%) | 22 (45%) | 22 (44%) | 17 (34%) |
| Pigmentation | | 1 (2%) | | |
| Artery, hyperplasia | | | 1 (2%) | |
| Artery, perivascular, inflammation, necrotizing, chronic | | 1 (2%) | | |
| Duct, dilatation | | 1 (2%) | | |
| Stomach, forestomach | (49) | (50) | (50) | (47) |
| Acanthosis | 3 (6%) | 3 (6%) | 3 (6%) | 7 (15%) |
| Edema | 1 (2%) | 1 (2%) | | 1 (2%) |
| Erosion | 1 (2%) | | | |
| Hyperkeratosis | 3 (6%) | 2 (4%) | 1 (2%) | 7 (15%) |
| Hyperplasia, basal cell | 4 (8%) | 2 (4%) | 3 (6%) | 5 (11%) |
| Inflammation, acute | | 1 (2%) | | |
| Inflammation, chronic active | 3 (6%) | 3 (6%) | | 2 (4%) |
| Inflammation, necrotizing, chronic active | | | 1 (2%) | |
| Necrosis, coagulative | | 1 (2%) | | |
| Ulcer | 1 (2%) | 1 (2%) | | 2 (4%) |
| Artery, mineralization | | | 1 (2%) | |
| Artery, muscularis, mineralization | | 1 (2%) | | |
| Muscularis, mineralization | | 1 (2%) | 3 (6%) | |
| Submucosa, mineralization | | | 1 (2%) | |

TABLE B5
Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Feed Study of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|----------|-----------|------------|------------|
| Alimentary System (continued) | | | | |
| Stomach, glandular | (50) | (49) | (50) | (50) |
| Cyst epithelial inclusion | | | | 1 (2%) |
| Edema | | 1 (2%) | | |
| Inflammation, chronic active | 3 (6%) | 1 (2%) | | 1 (2%) |
| Ulcer | 1 (2%) | | | |
| Artery, mineralization | | 3 (6%) | 1 (2%) | |
| Epithelium, pigmentation | | | 8 (16%) | 38 (76%) |
| Mucosa, dilatation | 6 (12%) | | 5 (10%) | 4 (8%) |
| Muscularis, developmental malformation | 1 (2%) | | | |
| Muscularis, mineralization | 1 (2%) | 2 (4%) | 6 (12%) | 4 (8%) |
| Tongue | (29) | (43) | (43) | (39) |
| Hemorrhage | | 2 (5%) | 1 (2%) | |
| Inflammation, chronic | 7 (24%) | 7 (16%) | 12 (28%) | 6 (15%) |
| Arteriole, necrosis, fibrinoid | | 2 (5%) | 1 (2%) | |
| Cardiovascular System | | | | |
| Heart | (50) | (13) | (7) | (50) |
| Cardiomyopathy | 47 (94%) | 9 (69%) | 3 (43%) | 47 (94%) |
| Atrium left, thrombus | | | | 1 (2%) |
| Coronary artery, inflammation, chronic active | | | | 1 (2%) |
| Coronary artery, inflammation, necrotizing, chronic active | 1 (2%) | | | |
| Coronary artery, necrosis, fibrinoid | | | | 1 (2%) |
| Epicardium, fibrosis | | | | 1 (2%) |
| Endocrine System | | | | |
| Adrenal gland | (50) | (14) | (13) | (50) |
| Hyperplasia | | | | 1 (2%) |
| Adrenal gland, cortex | (50) | (13) | (13) | (50) |
| Angiectasis | 28 (56%) | 2 (15%) | 2 (15%) | 26 (52%) |
| Atrophy | | 1 (8%) | | |
| Congestion | 1 (2%) | | 1 (8%) | |
| Degeneration, fatty | 1 (2%) | | | |
| Hyperplasia | 16 (32%) | | 2 (15%) | 14 (28%) |
| Necrosis, coagulative | 1 (2%) | | | 1 (2%) |
| Pigmentation | | 1 (8%) | | |
| Thrombus | 1 (2%) | | | |
| Vacuolization cytoplasmic | 28 (56%) | 8 (62%) | 3 (23%) | 24 (48%) |
| Adrenal gland, medulla | (50) | (13) | (12) | (50) |
| Angiectasis | 5 (10%) | | 1 (8%) | |
| Hyperplasia | 3 (6%) | | 1 (8%) | 4 (8%) |
| Necrosis, coagulative | 1 (2%) | | | |
| Islets, pancreatic | (44) | (15) | (8) | (49) |
| Ectopic tissue | 1 (2%) | | | |
| Hyperplasia | | 1 (7%) | | |
| Parathyroid gland | (40) | (39) | (36) | (43) |
| Cyst | 1 (3%) | | | |
| Hyperplasia | 1 (3%) | | 1 (3%) | |

TABLE B5
Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|----------|-----------|------------|------------|
| Endocrine System (continued) | | | | |
| Pituitary gland | (50) | (49) | (50) | (49) |
| Angiectasis | | | | 1 (2%) |
| Hyperplasia | | | | 2 (4%) |
| Pars distalis, angiectasis | 14 (28%) | 11 (22%) | 12 (24%) | 8 (16%) |
| Pars distalis, cyst | 26 (52%) | 21 (43%) | 23 (46%) | 28 (57%) |
| Pars distalis, cyst, multiple | | | 1 (2%) | 1 (2%) |
| Pars distalis, hemorrhage | 1 (2%) | | | |
| Pars distalis, hyperplasia | 18 (36%) | 23 (47%) | 18 (36%) | 13 (27%) |
| Pars distalis, infiltration cellular, histiocyte | | | 1 (2%) | |
| Pars distalis, pigmentation | 1 (2%) | 3 (6%) | 3 (6%) | |
| Pars intermedia, angiectasis | 5 (10%) | 6 (12%) | 4 (8%) | 6 (12%) |
| Pars intermedia, cyst | | 2 (4%) | 7 (14%) | 6 (12%) |
| Pars intermedia, pigmentation | | | 1 (2%) | |
| Pars nervosa, angiectasis | 1 (2%) | 1 (2%) | | |
| Pars nervosa, cyst | | | 1 (2%) | |
| Thyroid gland | (50) | (43) | (47) | (50) |
| Ultimobranchial cyst | 1 (2%) | | | 3 (6%) |
| C-cell, hyperplasia | 38 (76%) | 37 (86%) | 40 (85%) | 37 (74%) |
| Follicle, cyst | 2 (4%) | | | 4 (8%) |
| Follicular cell, hyperplasia | | 1 (2%) | | |
| General Body System | | | | |
| None | | | | |
| Genital System | | | | |
| Clitoral gland | (14) | (20) | (14) | (12) |
| Abscess | 1 (7%) | 2 (10%) | | 1 (8%) |
| Cyst | 3 (21%) | 2 (10%) | 1 (7%) | |
| Hyperplasia | 2 (14%) | 1 (5%) | | |
| Inflammation, acute | | 1 (5%) | 1 (7%) | |
| Inflammation, chronic | 6 (43%) | 10 (50%) | 4 (29%) | 4 (33%) |
| Inflammation, chronic active | 3 (21%) | 2 (10%) | 3 (21%) | 1 (8%) |
| Ovary | (50) | (17) | (15) | (48) |
| Congestion | | | 1 (7%) | |
| Cyst | 3 (6%) | | | 2 (4%) |
| Periovarian tissue, cyst | 5 (10%) | 3 (18%) | 9 (60%) | 5 (10%) |
| Uterus | (50) | (50) | (50) | (50) |
| Cyst | | | 3 (6%) | |
| Hydrometra | 11 (22%) | 9 (18%) | 5 (10%) | 14 (28%) |
| Inflammation, acute | | | | 1 (2%) |
| Inflammation, chronic | | 2 (4%) | | |
| Inflammation, chronic active | | | 1 (2%) | 1 (2%) |
| Metaplasia, squamous | | | | 1 (2%) |
| Pigmentation | | 1 (2%) | | |
| Cervix, cyst | | 1 (2%) | | |
| Cervix, inflammation, chronic | | 1 (2%) | | |
| Endometrium, hyperplasia | 6 (12%) | 7 (14%) | 8 (16%) | 7 (14%) |
| Lumen, exudate | 2 (4%) | | | |

TABLE B5
Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|--|----------|-----------|------------|------------|
| Hematopoietic System | | | | |
| Lymph node | (48) | (25) | (17) | (49) |
| Deep cervical, hyperplasia, plasma cell | 1 (2%) | | | |
| Lumbar, infiltration cellular, histiocyte | | | | 1 (2%) |
| Lumbar, pigmentation | | | | 1 (2%) |
| Mediastinal, depletion lymphoid | | | | 1 (2%) |
| Mediastinal, hemorrhage | 8 (17%) | 3 (12%) | 3 (18%) | 7 (14%) |
| Mediastinal, infiltration cellular, histiocyte | | | | 1 (2%) |
| Mediastinal, necrosis, coagulative | | | | 1 (2%) |
| Mediastinal, pigmentation | 2 (4%) | 1 (4%) | 1 (6%) | 2 (4%) |
| Pancreatic, ectasia | | | 1 (6%) | 1 (2%) |
| Pancreatic, hemorrhage | 1 (2%) | | | 2 (4%) |
| Pancreatic, infiltration cellular, histiocyte | | | 2 (12%) | 2 (4%) |
| Pancreatic, pigmentation | | | 2 (12%) | 3 (6%) |
| Renal, ectasia | | 2 (8%) | | |
| Renal, hemorrhage | 2 (4%) | 1 (4%) | 2 (12%) | 3 (6%) |
| Renal, infiltration cellular | 1 (2%) | | | |
| Renal, infiltration cellular, histiocyte | 1 (2%) | 3 (12%) | 2 (12%) | 4 (8%) |
| Renal, pigmentation | 1 (2%) | 4 (16%) | 2 (12%) | 3 (6%) |
| Lymph node, mandibular | (46) | (19) | (10) | (46) |
| Congestion | | 1 (5%) | | |
| Ectasia | 11 (24%) | 5 (26%) | | 6 (13%) |
| Hemorrhage | 22 (48%) | 6 (32%) | 3 (30%) | 17 (37%) |
| Hyperplasia, plasma cell | 2 (4%) | | | |
| Infiltration cellular, histiocyte | | 2 (11%) | 1 (10%) | 2 (4%) |
| Necrosis, coagulative | | | 1 (10%) | |
| Pigmentation | 1 (2%) | 2 (11%) | | 2 (4%) |
| Lymph node, mesenteric | (9) | (14) | (12) | (9) |
| Angiectasis | | | 1 (8%) | |
| Ectasia | | | 1 (8%) | 1 (11%) |
| Hemorrhage | 1 (11%) | | 2 (17%) | |
| Infiltration cellular, histiocyte | 8 (89%) | 14 (100%) | 10 (83%) | 7 (78%) |
| Necrosis, coagulative | | | | 1 (11%) |
| Pigmentation | 8 (89%) | 14 (100%) | 8 (67%) | 7 (78%) |
| Thrombus | | | 1 (8%) | |
| Spleen | (50) | (23) | (20) | (50) |
| Depletion lymphoid | 5 (10%) | 5 (22%) | 4 (20%) | 9 (18%) |
| Fibrosis | | 1 (4%) | | |
| Hematopoietic cell proliferation | | 2 (9%) | 1 (5%) | 1 (2%) |
| Hyperplasia, lymphoid | | | 3 (15%) | |
| Inflammation, granulomatous, chronic | | 2 (9%) | 3 (15%) | 1 (2%) |
| Necrosis | | | | 1 (2%) |
| Pigmentation | 1 (2%) | | | |
| Thrombus | 1 (2%) | 1 (4%) | | |
| Capsule, hyperplasia | 1 (2%) | | | |
| Thymus | (8) | (10) | (7) | (9) |
| Depletion lymphoid | 5 (63%) | 7 (70%) | 1 (14%) | 7 (78%) |
| Hemorrhage | | 1 (10%) | 1 (14%) | |
| Necrosis, coagulative | | | | 1 (11%) |

TABLE B5
Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|-----------------------------------|----------|-----------|------------|------------|
| Integumentary System | | | | |
| Mammary gland | (36) | (36) | (24) | (22) |
| Galactocele | | 1 (3%) | 1 (4%) | 1 (5%) |
| Hyperplasia | 13 (36%) | 20 (56%) | 13 (54%) | 13 (59%) |
| Hyperplasia, cystic | 1 (3%) | | | |
| Skin | (11) | (15) | (8) | (11) |
| Inflammation, chronic | | 1 (7%) | | |
| Inflammation, necrotizing, acute | | | 1 (13%) | |
| Subcutaneous tissue, edema | | | | 1 (9%) |
| Musculoskeletal System | | | | |
| None | | | | |
| Nervous System | | | | |
| Brain | (50) | (15) | (8) | (50) |
| Hemorrhage | | | | 2 (4%) |
| Hydrocephalus | 2 (4%) | 1 (7%) | 1 (13%) | 1 (2%) |
| Meninges, hemorrhage | | 2 (13%) | | |
| Spinal cord | (1) | (2) | | (2) |
| Hemorrhage | | 1 (50%) | | 1 (50%) |
| Respiratory System | | | | |
| Lung | (50) | (20) | (20) | (50) |
| Crystals | 1 (2%) | | | |
| Foreign body | | | 1 (5%) | |
| Hemorrhage | 4 (8%) | 3 (15%) | 2 (10%) | 4 (8%) |
| Infiltration cellular, histiocyte | 14 (28%) | 5 (25%) | 3 (15%) | 18 (36%) |
| Inflammation, acute | | | | 1 (2%) |
| Inflammation, chronic | 1 (2%) | | | 1 (2%) |
| Inflammation, chronic active | | | 1 (5%) | 1 (2%) |
| Leukocytosis | | | 1 (5%) | |
| Metaplasia, osseous | 1 (2%) | | | |
| Pigmentation | | | | 1 (2%) |
| Alveolar epithelium, hyperplasia | 2 (4%) | 1 (5%) | 2 (10%) | 1 (2%) |
| Artery, mineralization | 24 (48%) | 3 (15%) | 6 (30%) | 14 (28%) |
| Nose | (7) | (12) | (7) | (10) |
| Congestion | | 1 (8%) | | |
| Capillary, submucosa, thrombus | | | | 1 (10%) |
| Glands, inflammation, acute | | | 1 (14%) | 1 (10%) |
| Lumen, inflammation, acute | | 1 (8%) | | |

TABLE B5
Summary of the Incidence of Nonneoplastic Lesions in Female Rats in the 2-Year Feed Study
of Quercetin (continued)

| | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|----------|-----------|------------|------------|
| Special Senses System | | | | |
| Eye | (6) | (8) | (6) | (5) |
| Cataract | 5 (83%) | 2 (25%) | 3 (50%) | 1 (20%) |
| Synechia | 1 (17%) | 3 (38%) | 3 (50%) | |
| Cornea, fibrosis | | 2 (25%) | | |
| Retina, degeneration | 5 (83%) | 6 (75%) | 4 (67%) | 2 (40%) |
| Sclera, metaplasia, osseous | | | | 1 (20%) |
| Sclera, mineralization | 1 (17%) | | | |
| Harderian gland | | (4) | (2) | (2) |
| Hemorrhage | | 1 (25%) | 1 (50%) | |
| Inflammation, acute | | 1 (25%) | | |
| Inflammation, chronic | | 3 (75%) | 1 (50%) | 1 (50%) |
| Urinary System | | | | |
| Kidney | (49) | (49) | (50) | (50) |
| Autolysis | | 1 (2%) | 1 (2%) | |
| Congestion | | 1 (2%) | | 1 (2%) |
| Inflammation, chronic | | 1 (2%) | 1 (2%) | |
| Nephropathy | 48 (98%) | 48 (98%) | 50 (100%) | 48 (96%) |
| Artery, fibrosis | | 1 (2%) | | |
| Artery, thrombus | | | 1 (2%) | |
| Collecting tubule, mineralization | 1 (2%) | | | |
| Papilla, necrosis, coagulative | | | 1 (2%) | |
| Proximal convoluted renal tubule, degeneration, hyaline | | 1 (2%) | | |
| Proximal convoluted renal tubule, inflammation, acute | | 4 (8%) | | |
| Proximal convoluted renal tubule, pigmentation | | 1 (2%) | 1 (2%) | |
| Renal tubule, hyperplasia | 1 (2%) | 1 (2%) | 3 (6%) | 1 (2%) |
| Renal tubule, hyperplasia, cystic | | | 1 (2%) | |
| Transitional epithelium, hyperplasia | 7 (14%) | 9 (18%) | 5 (10%) | 4 (8%) |
| Urinary bladder | (50) | (49) | (50) | (50) |
| Hemorrhage | | | | 1 (2%) |
| Inflammation, acute | | | | 1 (2%) |
| Inflammation, chronic | | 4 (8%) | | |
| Subserosa, mineralization | 1 (2%) | | | |
| Transitional epithelium, hyperplasia | | | | 2 (4%) |

APPENDIX C

GENETIC TOXICOLOGY

| | |
|---|-----|
| <i>SALMONELLA</i> PROTOCOL | 138 |
| CHINESE HAMSTER OVARY CELL CYTOGENETICS ASSAYS | 138 |
| RESULTS | 139 |
| TABLE C1 Mutagenicity of Quercetin in <i>Salmonella typhimurium</i> | 140 |
| TABLE C2 Induction of Sister Chromatid Exchanges in Chinese Hamster Ovary Cells by Quercetin | 141 |
| TABLE C3 Induction of Chromosomal Aberrations in Chinese Hamster Ovary Cells by Quercetin | 142 |

GENETIC TOXICOLOGY

SALMONELLA PROTOCOL

Testing was performed as reported by Haworth *et al.* (1983) and Zeiger *et al.* (1988). Quercetin was sent to the laboratory as a coded aliquot from Radian Corporation, Austin, TX. It was incubated with the *Salmonella typhimurium* tester strains TA98 and TA100 either in buffer or S9 mix (metabolic activation enzymes and cofactors from Aroclor 1254-induced male Sprague-Dawley rat or Syrian hamster liver) for 20 minutes at 37° C prior to the addition of soft agar supplemented with *l*-histidine and *d*-biotin and subsequent plating on minimal glucose agar plates. Incubation was continued for an additional 48 hours.

Each test consisted of triplicate plates of concurrent positive and negative controls and of at least five doses of quercetin. The high dose was limited by toxicity. Tests were repeated for all negative assays, and all positive assays were retested under the conditions that elicited the positive response.

In this assay, a positive response is defined as a reproducible, dose-related increase in histidine-independent (revertant) colonies in any one strain/activation combination. An equivocal response is defined as an increase in revertants which was not dose related, not reproducible, or of insufficient magnitude to support a determination of mutagenicity. A response was considered negative when no increase in revertant colonies was observed after chemical treatment.

CHINESE HAMSTER OVARY CELL CYTOGENETICS ASSAYS

Testing was performed as reported by Galloway *et al.* (1985, 1987) and is briefly described as follows. Quercetin was sent to the laboratory as a coded aliquot from Radian Corporation, Austin, TX. It was tested in cultured Chinese hamster ovary (CHO) cells for induction of sister chromatid exchanges (SCE) and chromosomal aberrations (Abs) both in the presence and absence of Aroclor 1254-induced male Sprague-Dawley rat liver S9 and cofactor mix. Cultures were handled under gold lights to prevent photolysis of bromodeoxyuridine-substituted DNA. Each test consisted of concurrent solvent and positive controls and of at least three doses of quercetin; the high dose was limited by toxicity.

In the SCE test without S9, CHO cells were incubated for 26 hours with quercetin in McCoy's 5A medium supplemented with 10% fetal bovine serum, *l*-glutamine (2mM), and antibiotics. Bromodeoxyuridine (BrdU) was added 2 hours after culture initiation. After 26 hours, the medium containing quercetin was removed and replaced with fresh medium containing BrdU and Colcemid, and quercetin incubation was continued for 2 hours. Cells were then harvested by mitotic shake-off, fixed, and stained with Hoechst 33258 and Giemsa. In the SCE test with S9, cells were incubated with quercetin, serum-free medium, and S9 for 2 hours. The medium was then removed and replaced with medium containing BrdU and no quercetin, and incubation proceeded for an additional 25 to 26 hours, with Colcemid present for the final 2 to 3 hours. Harvesting and staining procedures were the same as for cells treated without S9.

In the Abs test without S9, cells were incubated in McCoy's 5A medium with quercetin for 18 hours; Colcemid was added and incubation continued for 2 hours. The cells were then harvested by mitotic shake-off, fixed, and stained with Giemsa. For the Abs test with S9, cells were treated with quercetin and S9 for 2 hours, after which the treatment medium was removed and the cells were incubated for 10 hours in fresh medium, with Colcemid present for the final 2 hours. Cells were harvested in the same manner as for treatment without S9.

Cell cycle delay was anticipated in the Abs test without S9, based on observance of cell cycle progression in the SCE test, and the incubation period prior to cell harvest was therefore extended to allow accumulation of sufficient metaphases for analysis.

Cells were selected for scoring on the basis of good morphology and completeness of karyotype (21 ± 2 chromosomes). All slides were scored blind, and those from a single test were read by the same person. For the SCE test, 25 to 50 second-division metaphase cells were scored for frequency of SCE per cell from each dose; 100 to 200 first-division metaphase cells were scored at each dose for the Abs test. Exceptions were made in each test when a culture showed high levels of damage which allowed fewer cells to provide a representative sample of the whole culture, or which made it difficult to locate scorable cells. Classes of aberrations included simple (breaks and terminal deletions), complex (rearrangements and translocations), and other (pulverized cells, despiralized chromosomes, and cells containing ten or more aberrations).

Statistical analyses were conducted on the slopes of the dose-response curves and on the individual dose points. An SCE frequency 20% above the concurrent solvent control value was chosen as a statistically conservative positive response. The probability of this level of difference occurring by chance at one dose point is less than 0.01; the probability for such a chance occurrence at two dose points is less than 0.001. Abs data are presented as percentage of cells with aberrations. For aberration data, both the dose-response curve and individual dose points were statistically analyzed. A statistically significant ($P \leq 0.05$) difference for one dose point was considered weak evidence for a positive response (+w); significant differences for two or more doses indicated the trial was positive (+) (Galloway *et al.*, 1987).

RESULTS

Exposure to quercetin (0.3 to 1,000 $\mu\text{g}/\text{plate}$) produced a strong, dose-related increase in gene mutations in *Salmonella typhimurium* strains TA100 and TA98 in the presence and in the absence of Aroclor 1254-induced male Sprague-Dawley rat or Syrian hamster liver S9 (Table C1). In cytogenetic tests with CHO cells, quercetin induced marked increases in both SCE and Abs, with and without Aroclor 1254-induced male Sprague-Dawley rat liver S9 (Tables C2 and C3). In the SCE test without S9, positive responses were observed over a dose range of 0.67 to 20 $\mu\text{g}/\text{mL}$ quercetin; with S9, effective doses ranged from 2 to 45 $\mu\text{g}/\text{mL}$. In the Abs test, the trials conducted in the absence of S9 activation employed a delayed harvest protocol to offset quercetin toxicity; positive responses occurred with 10 to 50 $\mu\text{g}/\text{mL}$ quercetin. With S9, standard harvest times were employed and strong increases in aberrations were observed with 25 to 75 $\mu\text{g}/\text{mL}$ quercetin. At the highest dose (75 $\mu\text{g}/\text{mL}$), 100% of the cells scored contained aberrations.

TABLE C1
Mutagenicity of Quercetin in *Salmonella typhimurium*^a

| Strain | Dose ($\mu\text{g}/\text{plate}$) | Revertants/plate ^b | | | | | |
|-------------------------------|--|-------------------------------|----------------|-----------------|----------------|-------------------------------|----------------|
| | | -S9 | | +30% hamster S9 | | +30% rat S9 | |
| | | Trial 1 | Trial 2 | Trial 1 | Trial 2 | Trial 1 | Trial 2 |
| TA100 | 0 | 134 \pm 15.8 | 134 \pm 3.4 | 123 \pm 15.7 | 141 \pm 8.8 | 115 \pm 7.8 | 157 \pm 6.7 |
| | 1 | | | 106 \pm 4.3 | 157 \pm 14.5 | 126 \pm 7.0 | 164 \pm 14.1 |
| | 3 | 153 \pm 9.1 | 156 \pm 4.4 | 132 \pm 7.8 | 172 \pm 10.2 | 127 \pm 11.5 | 142 \pm 5.8 |
| | 10 | | | 296 \pm 17.8 | 271 \pm 6.5 | 361 \pm 8.7 | 306 \pm 29.4 |
| | 33 | 253 \pm 13.6 | 222 \pm 13.6 | 449 \pm 7.2 | 566 \pm 38.7 | 542 \pm 20.5 | 517 \pm 19.8 |
| | 66 | | | | 798 \pm 31.7 | | 613 \pm 32.6 |
| | 100 | | 303 \pm 8.2 | 828 \pm 22.1 | | 798 \pm 32.4 | |
| | 333 | 440 \pm 26.7 | 341 \pm 27.0 | | | | |
| | 666 | 467 \pm 19.2 | 426 \pm 22.0 | | | | |
| | 1,000 | 512 \pm 29.1 ^d | | | | | |
| Trial summary | | Positive | Positive | Positive | Positive | Positive | Positive |
| Positive control ^c | | 402 \pm 23.7 | 466 \pm 13.6 | 654 \pm 45.6 | 553 \pm 54.3 | 615 \pm 41.0 | 498 \pm 9.8 |
| TA98 | 0.0 | 17 \pm 0.9 | 19 \pm 0.6 | 27 \pm 3.7 | 29 \pm 0.7 | 29 \pm 2.0 | 30 \pm 0.7 |
| | 0.3 | | 22 \pm 2.7 | | 26 \pm 3.1 | | 30 \pm 0.6 |
| | 1.0 | | 27 \pm 0.6 | 37 \pm 1.9 | 31 \pm 4.8 | 37 \pm 4.4 | 37 \pm 1.0 |
| | 3.0 | 78 \pm 4.3 | 53 \pm 2.0 | 77 \pm 8.4 | 51 \pm 3.8 | 68 \pm 4.0 | 50 \pm 2.5 |
| | 6.0 | | | | 162 \pm 12.3 | | 199 \pm 9.0 |
| | 10.0 | | 169 \pm 18.8 | 401 \pm 24.3 | 283 \pm 24.2 | 686 \pm 46.4 | 381 \pm 33.0 |
| | 33.0 | 404 \pm 9.8 | 223 \pm 3.8 | 796 \pm 64.7 | | 1053 \pm 5.1 | |
| | 100.0 | | | 916 \pm 63.5 | | 1,116 \pm 60.7 ^d | |
| | 333.0 | 549 \pm 16.3 | | | | | |
| | 666.0 | 576 \pm 39.2 | | | | | |
| 1,000.0 | 671 \pm 28.2 | | | | | | |
| Trial summary | | Positive | Positive | Positive | Positive | Positive | Positive |
| Positive control | | 495 \pm 25.5 | 452 \pm 18.6 | 367 \pm 30.9 | 436 \pm 2.5 | 168 \pm 7.3 | 160 \pm 8.7 |

^a Study performed at SRI, International. The detailed protocol is presented in Haworth *et al.* (1983) with modifications as described by Zeiger *et al.* (1988).

^b Revertants are presented as mean \pm standard error from 3 plates.

^c 2-aminoanthracene was used on both strains in the presence of S9. In the absence of metabolic activation, 4-nitro-*o*-phenylenediamine was tested on TA98, and sodium azide was tested on TA100.

^d Slight toxicity

TABLE C2
Induction of Sister Chromatid Exchanges in Chinese Hamster Ovary Cells by Quercetin^a

| Compound | Dose ($\mu\text{g/mL}$) | Total Cells | No. of Chromo- somes | No. of SCEs | SCEs/ Chromo- somes | SCEs/ Cell | Hrs in BrdU | Relative SCEs/Chromo- some (%) ^b |
|-------------------|------------------------------|----------------|----------------------------|----------------|---------------------------|---------------|----------------|---|
| -S9 | | | | | | | | |
| Trial 1 | | | | | | | | |
| Summary: Positive | | | | | | | | |
| Dimethylsulfoxide | | 50 | 1,050 | 410 | 0.39 | 8.2 | 25.8 | |
| Mitomycin-C | 0.001 | 50 | 1,044 | 663 | 0.63 | 13.3 | 25.8 | 62.64 |
| | 0.010 | 5 | 105 | 181 | 1.72 | 36.2 | 25.8 | 341.47 |
| Quercetin | 0.67 | 50 | 1,044 | 1,041 | 0.99 | 20.8 | 25.8 | 155.36* |
| | 2.00 | 50 | 1,046 | 563 | 0.53 | 11.3 | 25.8 | 37.84* |
| | 6.70 | 5 | 104 | 92 | 0.88 | 18.4 | 25.8 | 126.55* |
| | 20.00 | 50 | 1,046 | 1,087 | 1.03 | 21.7 | 25.8 | 166.14* |
| | | | | | | | | P<0.001 ^c |
| +S9 | | | | | | | | |
| Trial 1 | | | | | | | | |
| Summary: Positive | | | | | | | | |
| Dimethylsulfoxide | | 50 | 1,043 | 404 | 0.38 | 8.1 | 25.8 | |
| Cyclophosphamide | 0.40 | 50 | 1,048 | 613 | 0.58 | 12.3 | 25.8 | 51.01 |
| | 2.00 | 5 | 104 | 169 | 1.62 | 33.8 | 25.8 | 319.53 |
| Quercetin | 2.0 | 50 | 1,048 | 506 | 0.48 | 10.1 | 25.8 | 24.65* |
| | 6.7 | 50 | 1,043 | 587 | 0.56 | 11.7 | 25.8 | 45.30* |
| | 20.0 | 50 | 1,041 | 597 | 0.57 | 11.9 | 25.8 | 48.06* |
| | | | | | | | | P<0.001 |
| Trial 2 | | | | | | | | |
| Summary: Positive | | | | | | | | |
| Dimethylsulfoxide | | 25 | 522 | 180 | 0.34 | 7.2 | 25.3 | |
| Cyclophosphamide | 0.40 | 25 | 521 | 323 | 0.61 | 12.9 | 25.3 | 79.79 |
| | 2.00 | 5 | 103 | 230 | 2.23 | 46.0 | 25.3 | 547.58 |
| Quercetin | 20.0 | 25 | 524 | 272 | 0.51 | 10.9 | 25.3 | 50.54* |
| | 30.0 | 25 | 524 | 308 | 0.58 | 12.3 | 25.3 | 70.46* |
| | 45.0 | 25 | 522 | 414 | 0.79 | 16.6 | 25.3 | 130.00* |
| | | | | | | | | P<0.001 |

* Positive ($\geq 20\%$ increase over solvent control)

^a Study performed at Litton Bionetics, Inc. SCE = sister chromatid exchange; BrdU = bromodeoxyuridine. A detailed description of the SCE protocol is presented by Galloway *et al.* (1985, 1987).

^b Percent increase in SCEs/chromosome of culture exposed to quercetin relative to those of culture exposed to solvent. Values at least 20% above control levels are considered positive.

^c Significance of relative SCEs/chromosome tested by the linear regression trend test vs. log of the dose

TABLE C3
Induction of Chromosomal Aberrations in Chinese Hamster Ovary Cells by Quercetin^a

| -S9 | | | | | +S9 | | | | |
|---|----------------|---------------|--------------|------------------------------|---|----------------|---------------|--------------|------------------------------|
| Dose ($\mu\text{g}/\text{mL}$) | Total Cells | No. of Abs | Abs/ Cell | Percent Cells with Abs | Dose ($\mu\text{g}/\text{mL}$) | Total Cells | No. of Abs | Abs/ Cell | Percent Cells with Abs |
| Trial 1 – Harvest time: 20.2 hours | | | | | Trial 1 – Harvest time: 12.0 hours | | | | |
| Summary: Positive | | | | | Summary: Positive | | | | |
| Dimethylsulfoxide | | | | | Dimethylsulfoxide | | | | |
| | 200 | 2 | 0.01 | 1.0 | | 200 | 5 | 0.03 | 2.5 |
| Mitomycin-C | | | | | Cyclophosphamide | | | | |
| 0.05 | 200 | 95 | 0.48 | 26.5 | 7.5 | 200 | 62 | 0.31 | 14.5 |
| 0.08 | 25 | 38 | 1.52 | 72.0 | 37.5 | 25 | 42 | 1.68 | 56.0 |
| Quercetin | | | | | Quercetin | | | | |
| 7.6 | 200 | 7 | 0.04 | 3.5 | 25.2 | 20 | 58 | 2.90 | 45.0* |
| 10.1 | 200 | 37 | 0.19 | 10.0* | 50.3 | 48 | 27 | 0.56 | 33.3* |
| 25.2 | 200 | 102 | 0.51 | 21.5* | 75.0 | 25 | 171 | 6.84 | 100.0* |
| P < 0.001 ^b | | | | | P < 0.001 | | | | |
| Trial 2 – Harvest time: 19.7 hours | | | | | | | | | |
| Summary: Positive | | | | | | | | | |
| Dimethylsulfoxide | | | | | | | | | |
| | 100 | 1 | 0.01 | 1.0 | | | | | |
| Mitomycin-C | | | | | | | | | |
| 0.05 | 100 | 45 | 0.45 | 30.0 | | | | | |
| 0.08 | 25 | 25 | 1.00 | 60.0 | | | | | |
| Quercetin | | | | | | | | | |
| 25.0 | 100 | 19 | 0.19 | 7.0* | | | | | |
| 37.5 | 100 | 25 | 0.25 | 15.0* | | | | | |
| 50.0 | 100 | 42 | 0.42 | 29.0* | | | | | |
| P < 0.001 | | | | | | | | | |

* Positive ($P \leq 0.05$)

^a Study performed at Litton Bionetics, Inc. Abs = aberrations. A detailed presentation of the technique for detecting chromosomal aberrations is found in Galloway *et al.* (1985, 1987).

^b Significance of percent cells with aberrations tested by the linear regression trend test vs. log of the dose

APPENDIX D ORGAN WEIGHTS AND ORGAN-WEIGHT-TO-BODY-WEIGHT RATIOS

| | | |
|-----------------|--|------------|
| TABLE D1 | Organ Weights and Organ-Weight-to-Body-Weight Ratios for Rats at the 6-Month Interim Evaluations in the 2-Year Feed Studies of Quercetin | 144 |
| TABLE D2 | Organ Weights and Organ-Weight-to-Body-Weight Ratios for Rats at the 15-Month Interim Evaluations in the 2-Year Feed Studies of Quercetin | 145 |

TABLE D1
Organ Weights and Organ-Weight-to-Body-Weight Ratios for Rats at the 6-Month Interim Evaluations
in the 2-Year Feed Studies of Quercetin^a

| Organ | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|------------------|---------------------------|--------------------------|--------------|---------------|
| Male | | | | |
| n | 9 | 9 | 10 | 10 |
| Necropsy body wt | 416 ± 10 | 417 ± 7 | 397 ± 6 | 401 ± 8 |
| Brain | | | | |
| Absolute | 1.93 ± 0.03 | 1.96 ± 0.02 | 1.72 ± 0.11 | 1.96 ± 0.02 |
| Relative | 4.66 ± 0.11 | 4.69 ± 0.05 | 4.36 ± 0.30 | 4.88 ± 0.07 |
| R. Kidney | | | | |
| Absolute | 1.17 ± 0.05 | 1.22 ± 0.04 ^b | 1.13 ± 0.04 | 1.29 ± 0.02* |
| Relative | 2.79 ± 0.08 | 2.92 ± 0.12 ^b | 2.83 ± 0.09 | 3.23 ± 0.09** |
| Liver | | | | |
| Absolute | 12.87 ± 0.35 ^b | 12.68 ± 0.45 | 12.25 ± 0.34 | 13.66 ± 0.32 |
| Relative | 30.9 ± 0.5 ^b | 30.3 ± 0.8 | 30.8 ± 0.6 | 34.1 ± 0.6** |
| Female | | | | |
| n | 10 | 10 | 10 | 10 |
| Necropsy body wt | 243 ± 5 | 245 ± 6 | 234 ± 4 | 214 ± 5** |
| Brain | | | | |
| Absolute | 1.82 ± 0.03 | 1.83 ± 0.03 | 1.84 ± 0.03 | 1.87 ± 0.02 |
| Relative | 7.48 ± 0.10 | 7.48 ± 0.14 | 7.90 ± 0.13* | 8.74 ± 0.17** |
| R. Kidney | | | | |
| Absolute | 0.68 ± 0.01 | 0.69 ± 0.02 | 0.68 ± 0.01 | 0.66 ± 0.02 |
| Relative | 2.81 ± 0.04 | 2.84 ± 0.05 | 2.89 ± 0.03 | 3.06 ± 0.05** |
| Liver | | | | |
| Absolute | 7.31 ± 0.25 | 7.42 ± 0.20 | 7.50 ± 0.23 | 6.88 ± 0.20 |
| Relative | 30.0 ± 0.7 | 30.3 ± 0.5 | 32.0 ± 0.6* | 32.1 ± 0.4* |

* Significantly different ($P \leq 0.05$) from the control group by Williams' or Dunnett's test

** $P \leq 0.01$

^a Organ weights are given in grams; organ-weight-to-body-weight ratios are given as mg organ weight/g body weight (mean ± standard error).

^b n=10

TABLE D2
Organ Weights and Organ-Weight-to-Body-Weight Ratios for Rats at the 15-Month Interim Evaluations in the 2-Year Feed Studies of Quercetin^a

| Organ | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|------------------|--------------|--------------|--------------|---------------|
| Male | | | | |
| n | 10 | 10 | 10 | 9 |
| Necropsy body wt | 460 ± 12 | 466 ± 8 | 459 ± 14 | 456 ± 15 |
| Brain | | | | |
| Absolute | 2.07 ± 0.03 | 2.06 ± 0.02 | 2.04 ± 0.03 | 2.05 ± 0.03 |
| Relative | 4.53 ± 0.09 | 4.44 ± 0.08 | 4.48 ± 0.14 | 4.53 ± 0.11 |
| R. Kidney | | | | |
| Absolute | 1.44 ± 0.05 | 1.51 ± 0.06 | 1.44 ± 0.04 | 1.59 ± 0.05 |
| Relative | 3.15 ± 0.09 | 3.27 ± 0.17 | 3.16 ± 0.07 | 3.49 ± 0.06* |
| Liver | | | | |
| Absolute | 15.66 ± 0.65 | 15.12 ± 0.63 | 15.23 ± 0.48 | 17.40 ± 0.75 |
| Relative | 34.0 ± 1.0 | 32.4 ± 0.9 | 33.2 ± 0.5 | 38.1 ± 1.0** |
| Female | | | | |
| n | 10 | 10 | 10 | 10 |
| Necropsy body wt | 324 ± 9 | 337 ± 8 | 307 ± 6 | 287 ± 6** |
| Brain | | | | |
| Absolute | 1.90 ± 0.03 | 1.90 ± 0.02 | 1.89 ± 0.02 | 1.90 ± 0.02 |
| Relative | 5.88 ± 0.12 | 5.65 ± 0.13 | 6.20 ± 0.13 | 6.65 ± 0.15** |
| R. Kidney | | | | |
| Absolute | 0.89 ± 0.03 | 0.93 ± 0.02 | 0.87 ± 0.02 | 0.88 ± 0.02 |
| Relative | 2.74 ± 0.06 | 2.77 ± 0.07 | 2.85 ± 0.04 | 3.08 ± 0.09** |
| Liver | | | | |
| Absolute | 9.21 ± 0.21 | 9.44 ± 0.31 | 8.90 ± 0.28 | 9.53 ± 0.34 |
| Relative | 28.5 ± 0.6 | 27.9 ± 0.4 | 29.1 ± 0.8 | 33.2 ± 0.8** |

* Significantly different ($P \leq 0.05$) from the control group by Williams' or Dunnett's test

** $P \leq 0.01$

^a Organ weights are given in grams; organ-weight-to-body-weight ratios are given as mg organ weight/g body weight (mean ± standard error).

APPENDIX E

HEMATOLOGY, CLINICAL CHEMISTRY, AND URINALYSIS RESULTS

| | | |
|-----------------|---|------------|
| TABLE E1 | Hematology, Clinical Chemistry, and Urinalysis Data for Rats at the 6-Month Interim Evaluations in the 2-Year Feed Studies of Quercetin | 148 |
| TABLE E2 | Hematology, Clinical Chemistry, and Urinalysis Data for Rats at the 15-Month Interim Evaluations in the 2-Year Feed Studies of Quercetin | 150 |

TABLE E1
Hematology, Clinical Chemistry, and Urinalysis Data for Rats at the 6-Month Interim Evaluations
in the 2-Year Feed Studies of Quercetin^a

| Analysis | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|-------------------------|-----------------------|--------------------------|---------------|
| Male | | | | |
| n | 10 | 10 | 9 | 10 |
| Hematology | | | | |
| Erythrocytes ($10^6/\mu\text{L}$) | 9.50 ± 0.24 | 9.39 ± 0.25 | 9.57 ± 0.24 | 8.78 ± 0.25* |
| Leukocytes ($10^3/\mu\text{L}$) | 5.77 ± 0.16 | 5.32 ± 0.07* | 4.87 ± 0.17** | 5.11 ± 0.20** |
| Segmented neutrophils ($10^3/\mu\text{L}$) | 1.41 ± 0.12 | 1.34 ± 0.13 | 1.40 ± 0.13 | 1.45 ± 0.14 |
| Lymphocytes ($10^3/\mu\text{L}$) | 4.04 ± 0.13 | 3.63 ± 0.18 | 3.20 ± 0.20** | 3.33 ± 0.15** |
| Monocytes ($10^3/\mu\text{L}$) | 0.28 ± 0.03 | 0.28 ± 0.04 | 0.17 ± 0.03* | 0.21 ± 0.03* |
| Eosinophils ($10^3/\mu\text{L}$) | 0.05 ± 0.02 | 0.06 ± 0.03 | 0.10 ± 0.03 | 0.12 ± 0.03 |
| Nucleated erythrocytes ($10^3/\mu\text{L}$) | 0.02 ± 0.01 | 0.01 ± 0.01 | 0.01 ± 0.01 | 0.00 ± 0.00 |
| Clinical chemistry | | | | |
| BUN (mg/dL) | 12.7 ± 0.9 ^b | 18.6 ± 3.2 | 11.9 ± 0.5 | 10.7 ± 0.3 |
| Creatinine (mg/dL) | 0.70 ± 0.05 | 0.77 ± 0.05 | 0.60 ± 0.03 | 0.57 ± 0.02* |
| Sodium (mEq/L) | 147 ± 1 | 147 ± 1 | 148 ± 1 ^c | 144 ± 0 |
| Potassium (mEq/L) | 3.75 ± 0.08 | 3.83 ± 0.08 | 3.79 ± 0.11 ^c | 3.67 ± 0.07 |
| Chloride (mEq/L) | 108 ± 1 | 109 ± 1 | 109 ± 1 ^c | 106 ± 0 |
| ALT (IU/L) | 72 ± 7 | 63 ± 6 ^b | 67 ± 5 | 53 ± 4* |
| AST (IU/L) | 122 ± 9 | 119 ± 10 ^b | 115 ± 7 | 82 ± 4** |
| SDH (IU/L) | 567 ± 113 ^b | 558 ± 81 ^d | 792 ± 134 | 647 ± 104 |
| Urinalysis | | | | |
| Urinary sodium (mEq/L) | 46 ± 14 | 50 ± 10 | 66 ± 14 ^c | 60 ± 11 |
| Urinary potassium (mEq/L) | 136 ± 21 | 144 ± 23 | 166 ± 21 ^c | 121 ± 17 |
| Urinary chloride (mEq/L) | 91 ± 19 | 105 ± 18 | 121 ± 19 ^c | 98 ± 16 |

TABLE E1
Hematology, Clinical Chemistry, and Urinalysis Data for Rats at the 6-Month Interim Evaluations
in the 2-Year Feed Studies of Quercetin (continued)

| Analysis | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|-----------------------|---------------------------|--------------------------|---------------------------|
| Female | | | | |
| n | 10 | 10 | 10 | 10 |
| Hematology | | | | |
| Erythrocytes ($10^6/\mu\text{L}$) | 8.18 ± 0.22 | 8.75 ± 0.16 | 8.73 ± 0.17 | 8.56 ± 0.19 |
| Leukocytes ($10^3/\mu\text{L}$) | 3.73 ± 0.22 | 4.07 ± 0.22 | 3.80 ± 0.26 | 3.43 ± 0.15 ^b |
| Segmented neutrophils ($10^3/\mu\text{L}$) | 0.78 ± 0.10 | 0.83 ± 0.05 | 0.95 ± 0.07 ^b | 0.74 ± 0.07 ^b |
| Lymphocytes ($10^3/\mu\text{L}$) | 2.80 ± 0.19 | 2.95 ± 0.19 | 2.50 ± 0.15 | 2.60 ± 0.17 |
| Monocytes ($10^3/\mu\text{L}$) | 0.11 ± 0.02 | 0.24 ± 0.03 ^{**} | 0.17 ± 0.02 | 0.13 ± 0.02 ^b |
| Eosinophils ($10^3/\mu\text{L}$) | 0.02 ± 0.01 | 0.05 ± 0.01 | 0.06 ± 0.03 | 0.01 ± 0.01 |
| Nucleated erythrocytes ($10^3/\mu\text{L}$) | 0.01 ± 0.01 | 0.01 ± 0.01 | 0.02 ± 0.01 | 0.01 ± 0.01 |
| Clinical chemistry | | | | |
| BUN (mg/dL) | 17.9 ± 1.3 | 21.7 ± 2.0 | 19.9 ± 1.1 | 21.1 ± 1.0 |
| Creatinine (mg/dL) | 0.54 ± 0.03 | 0.54 ± 0.04 | 0.47 ± 0.03 | 0.42 ± 0.04 ^{**} |
| Sodium (mEq/L) | 143 ± 0 | 144 ± 0 | 144 ± 0 [*] | 144 ± 0 |
| Potassium (mEq/L) | 3.04 ± 0.06 | 3.13 ± 0.10 | 3.21 ± 0.14 | 3.23 ± 0.09 |
| Chloride (mEq/L) | 107 ± 0 | 108 ± 1 | 108 ± 1 | 108 ± 1 |
| ALT (IU/L) | 30 ± 1 ^b | 33 ± 2 | 34 ± 4 ^b | 42 ± 5 |
| AST (IU/L) | 65 ± 2 ^b | 72 ± 3 | 83 ± 7 ^{*b} | 76 ± 5 [*] |
| SDH (IU/L) | 414 ± 23 ^b | 557 ± 62 ^{*b} | 535 ± 85 ^e | 635 ± 76 ^b |
| Urinalysis | | | | |
| Urinary sodium (mEq/L) | 43 ± 6 | 26 ± 2 [*] | 31 ± 4 | 38 ± 4 ^b |
| Urinary potassium (mEq/L) | 99 ± 16 | 61 ± 6 | 110 ± 23 | 123 ± 22 |
| Urinary chloride (mEq/L) | 70 ± 13 | 45 ± 3 | 64 ± 10 | 92 ± 19 |

^{*} Significantly different ($P \leq 0.05$) from the control group by Dunn's or Shirley's test

^{**} $P \leq 0.01$

^a Mean ± standard error. BUN=blood urea nitrogen; ALT=alanine aminotransferase; AST=aspartate aminotransferase; SDH=sorbitol dehydrogenase.

^b n=9

^c n=10

^d n=7

^e n=8

TABLE E2
Hematology, Clinical Chemistry, and Urinalysis Data for Rats at the 15-Month Interim Evaluations
in the 2-Year Feed Studies of Quercetin^a

| Analysis | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|------------------------|-----------------------|-----------------------|-------------|
| Male | | | | |
| n | 10 | 10 | 10 | 10 |
| Hematology | | | | |
| Erythrocytes ($10^6/\mu\text{L}$) | 9.44 ± 0.23 | 9.65 ± 0.21 | 9.65 ± 0.14 | 9.43 ± 0.24 |
| Leukocytes ($10^3/\mu\text{L}$) | 5.08 ± 0.26 | 5.33 ± 0.30 | 4.88 ± 0.21 | 4.99 ± 0.33 |
| Segmented neutrophils ($10^3/\mu\text{L}$) | 2.03 ± 0.24 | 1.67 ± 0.11 | 1.56 ± 0.16 | 1.89 ± 0.19 |
| Lymphocytes ($10^3/\mu\text{L}$) | 2.72 ± 0.19 | 3.38 ± 0.22 | 3.02 ± 0.17 | 2.87 ± 0.18 |
| Monocytes ($10^3/\mu\text{L}$) | 0.24 ± 0.04 | 0.20 ± 0.03 | 0.20 ± 0.03 | 0.19 ± 0.03 |
| Eosinophils ($10^3/\mu\text{L}$) | 0.10 ± 0.02 | 0.08 ± 0.03 | 0.10 ± 0.03 | 0.05 ± 0.02 |
| Nucleated erythrocytes ($10^3/\mu\text{L}$) | 0.03 ± 0.02 | 0.04 ± 0.02 | 0.06 ± 0.02 | 0.03 ± 0.02 |
| Clinical chemistry | | | | |
| BUN (mg/dL) | 17.8 ± 1.0 | 32.4 ± 9.4 | 18.3 ± 1.0 | 17.8 ± 1.4 |
| Creatinine (mg/dL) | 0.49 ± 0.05 | 0.72 ± 0.16 | 0.44 ± 0.02 | 0.58 ± 0.04 |
| Sodium (mEq/L) | 146 ± 0 | 147 ± 1 | 147 ± 0 | 147 ± 0 |
| Potassium (mEq/L) | 3.61 ± 0.08 | 3.72 ± 0.11 | 3.54 ± 0.08 | 3.78 ± 0.06 |
| Chloride (mEq/L) | 110 ± 1 | 108 ± 1 | 109 ± 1 | 107 ± 1* |
| SDH (IU/L) | 816 ± 114 ^b | 621 ± 70 ^b | 708 ± 73 ^b | 345 ± 34** |
| Urinalysis | | | | |
| Urinary sodium (mEq/L) | 54 ± 8 | 57 ± 7 | 63 ± 7 ^b | 38 ± 7 |
| Urinary potassium (mEq/L) | 177 ± 14 | 190 ± 12 | 195 ± 13 | 141 ± 12 |
| Urinary chloride (mEq/L) | 120 ± 12 | 128 ± 9 | 139 ± 9 ^b | 90 ± 10 |

TABLE E2
Hematology, Clinical Chemistry, and Urinalysis Data for Rats at the 15-Month Interim Evaluations
in the 2-Year Feed Studies of Quercetin (continued)

| Analysis | 0 ppm | 1,000 ppm | 10,000 ppm | 40,000 ppm |
|---|----------------------|-------------------------|---------------------|-----------------------|
| Female | | | | |
| n | 10 | 10 | 10 | 10 |
| Hematology | | | | |
| Erythrocytes ($10^6/\mu\text{L}$) | 8.62 ± 0.11 | 8.48 ± 0.12 | 8.56 ± 0.10 | 8.15 ± 0.11** |
| Leukocytes ($10^3/\mu\text{L}$) | 3.12 ± 0.16 | 3.01 ± 0.12 | 3.10 ± 0.17 | 3.33 ± 0.23 |
| Segmented neutrophils ($10^3/\mu\text{L}$) | 1.05 ± 0.06 | 0.89 ± 0.04 | 0.95 ± 0.07 | 0.91 ± 0.10 |
| Lymphocytes ($10^3/\mu\text{L}$) | 1.90 ± 0.11 | 1.93 ± 0.12 | 1.98 ± 0.14 | 2.23 ± 0.15 |
| Monocytes ($10^3/\mu\text{L}$) | 0.13 ± 0.02 | 0.16 ± 0.02 | 0.15 ± 0.02 | 0.16 ± 0.04 |
| Eosinophils ($10^3/\mu\text{L}$) | 0.04 ± 0.01 | 0.03 ± 0.01 | 0.03 ± 0.01 | 0.03 ± 0.01 |
| Nucleated erythrocytes ($10^3/\mu\text{L}$) | 0.04 ± 0.01 | 0.04 ± 0.02 | 0.05 ± 0.01 | 0.04 ± 0.01 |
| Clinical chemistry | | | | |
| BUN (mg/dL) | 16.5 ± 1.2 | 14.0 ± 0.6 ^b | 15.8 ± 0.9 | 17.1 ± 1.7 |
| Creatinine (mg/dL) | 0.55 ± 0.04 | 0.62 ± 0.05 | 0.54 ± 0.02 | 0.59 ± 0.04 |
| Sodium (mEq/L) | 147 ± 0 | 147 ± 1 | 146 ± 1 | 146 ± 0 |
| Potassium (mEq/L) | 3.17 ± 0.07 | 3.32 ± 0.08 | 3.30 ± 0.08 | 3.27 ± 0.09 |
| Chloride (mEq/L) | 110 ± 0 | 111 ± 1 | 111 ± 1 | 111 ± 1 |
| ALT (IU/L) | 29 ± 2 | 27 ± 2 ^b | 29 ± 2 | 33 ± 3 |
| AST (IU/L) | 63 ± 4 | 63 ± 2 ^b | 67 ± 5 | 61 ± 3 |
| SDH (IU/L) | 205 ± 21 | 214 ± 24 | 180 ± 17 | 248 ± 35 |
| Urinalysis | | | | |
| Urinary sodium (mEq/L) | 50 ± 5 ^b | 40 ± 6 | 35 ± 7 ^b | 29 ± 7* |
| Urinary potassium (mEq/L) | 143 ± 7 ^b | 113 ± 5* | 121 ± 7* | 114 ± 13** |
| Urinary chloride (mEq/L) | 110 ± 7 ^b | 90 ± 6* | 87 ± 6* | 78 ± 6** ^b |

* Significantly different ($P \leq 0.05$) from the control group by Dunn's or Shirley's test

** $P \leq 0.01$

^a Mean ± standard error. BUN=blood urea nitrogen; ALT=alanine aminotransferase; AST=aspartate aminotransferase;

^b SDH=sorbitol dehydrogenase.

n=9

APPENDIX F

CHEMICAL CHARACTERIZATION AND DOSE FORMULATION STUDIES

| | |
|--|------------|
| PROCUREMENT AND CHARACTERIZATION | 154 |
| PREPARATION AND ANALYSIS OF DOSE FORMULATIONS | 155 |
| FIGURE F1 Infrared Absorption Spectrum of Quercetin | 156 |
| FIGURE F2 Nuclear Magnetic Resonance Spectrum of Quercetin | 157 |
| TABLE F1 Preparation and Storage of Dose Formulations in the Feed Studies of Quercetin | 158 |
| TABLE F2 Results of Analysis of Dose Formulations in the 2-Year Feed Studies of Quercetin ... | 159 |
| TABLE F3 Results of Referee Analysis of Dose Formulations in the 2-Year Feed Studies of Quercetin | 160 |

CHEMICAL CHARACTERIZATION AND DOSE FORMULATION STUDIES

PROCUREMENT AND CHARACTERIZATION

Quercetin was obtained in two lots from Freeman Industries (Tuckahoe, NY). Lot no. 969-3790-05 (anhydrous form) was used during the first year of the studies and lot no. 969-0483-18BL (dihydrate form) was used during the second year of the studies. Identity, purity and stability analyses were conducted by the analytical chemistry laboratory Midwest Research Institute (MRI) (Kansas City, MO). MRI reports on analyses performed in support of the quercetin studies are on file at the National Institute of Environmental Health Sciences.

The study chemical, a yellow crystalline powder, was identified as quercetin by infrared, ultraviolet/visible, and nuclear magnetic resonance (NMR) spectroscopy. All spectra were consistent with those expected for the structure and with the literature spectra of quercetin, as shown in Figures F1 and F2 (*Sadtler Standard Spectra*).

The purity of both lots was determined by elemental analyses, Karl Fischer water analysis, weight loss on drying, NMR, titration, and chromatographic analyses. Titration of two acid groups was performed in dimethylformamide with 0.1 N tetrabutylammonium hydroxide in methanol:2-propanol (1:9) as the titrant. Thin-layer chromatography was performed with two systems: 1) on MN Polyamide-TLC11 plates with methanol:acetylacetone (60:40), and 2) on silica gel plates with toluene:dioxane:acetic acid:methanol (40:25:20:15). After the plates were sprayed with 2,6-dibromoquinonechloroimide, visualization was accomplished with short wave (254 nm) and long wave (366 nm) ultraviolet light. 2,2',4,4'-Tetrahydroxybenzophenone in absolute ethanol (1 μ L of a 10 mg/mL solution) was used as the reference standard. High-performance liquid chromatography (HPLC) was performed with a μ Bondapak C₁₈ column and a mobile phase mixture of two solvents: A) water with pH adjusted to 2.0 with concentrated phosphoric acid and B) methanol with an equal volume of phosphoric acid as added in solvent A. The ratio of solvents used was 52:48 (A:B), at a flow rate of 1 mL/minute. Ultraviolet detection was at 254 nm.

For the anhydrous form, elemental analyses for carbon and hydrogen showed carbon was low and hydrogen was slightly high. Weight loss on drying indicated the presence of 1% to 3% water. NMR quantification indicated the presence of 2.4% water. Titration of two acid groups indicated a purity of $100.8 \pm 1.1\%$. This method would not necessarily distinguish between quercetin and other non-phenolic acid components or quercetin-like compounds. Thin-layer chromatography indicated a major product spot, a minor spot, and a trace by solvent system 1, and a major spot and two traces by solvent system 2. HPLC indicated three impurities with a combined area of 6.6% relative to the major peak. The largest impurity (6.4% by peak area) was identified as ellagic acid by spectroscopy and mass spectrometry. Quantitation against an ellagic acid standard resulted in an estimate of the impurity level of 2.6% (w/w). The overall purity is estimated at approximately 95% as the anhydrous form.

For the dihydrous form, elemental analyses for carbon and hydrogen showed carbon was slightly high, but the value for hydrogen agreed with the theoretical value. Karl Fischer analysis indicated $11.2 \pm 0.5\%$ water, which is consistent with the theoretical value for the dihydrous form. Weight loss on drying indicated the presence of $9.1 \pm 0.1\%$ water. Titration of acid groups indicated a purity of $112.5 \pm 0.4\%$. Thin-layer chromatography indicated a major product spot and two traces by both solvent systems. HPLC indicated three impurities with areas greater than 0.1% relative to the major peak and a combined relative area of 3.5%. The largest peak (3.1% by peak area) was identified as ellagic acid by spectroscopy and mass spectrometry. Quantitation against an ellagic acid standard

resulted in an estimate of the impurity level of 1.1% (w/w). The overall purity is estimated at approximately 98% as the dihydrate.

Stability studies performed by HPLC with the system described above but with a flow rate of 2.0 mL/minute and with acetanilide added as an internal standard indicated that quercetin, when stored protected from light and under a nitrogen headspace, was stable as a bulk chemical for 2 weeks at temperatures up to 60° C. During the 2-year studies, the stability of the bulk chemical was monitored by the study laboratory using HPLC, with the system described above and with a flow rate of 1.0 mL/minute, and infrared spectral analysis; no degradation of the study material was seen throughout the studies.

PREPARATION AND ANALYSIS OF DOSE FORMULATIONS

The dose formulations were prepared layering a premix, prepared by grinding equal amounts of quercetin and feed with a mortar and pestle, with the remainder of the feed in a blender, (Patterson-Kelley Twin Shell with intensifier bar) and mixing for 15 minutes (Table F1). Studies were conducted by the analytical chemistry laboratory to determine homogeneity and stability of the dosed feed preparations. For homogeneity analyses, the formulations were extracted with methanol:acetic acid (99:1) and the absorbance of the samples was measured versus methanol by ultraviolet spectroscopy at 370 nm. Concentrations were calculated using a standard curve. For the stability studies, a methanol:hydrochloric acid (99.5:0.5) solution was used for extraction and the extract injected into an HPLC system equipped with a μ Bondapak C₁₈ column and a 254 nm detector. The mobile phase was a mixture of two solvents: A) 1.2 mL phosphoric acid and 800 mL water, with pH approximately 2, and B) 1.2 mL phosphoric acid and 800 mL methanol. The ratio of solvents used was 40:60 (A:B) at a flow rate of 2 mL/minute. Visible detection was at 254 nm.

Quercetin at the 10,000 ppm dose level mixed in rodent feed (NIH-07 Rat and Mouse Ration) produced a homogeneous blend and was found to be stable when stored at temperatures up to 25° C. There was a 3% loss of chemical in feed stored 2 weeks at 45° C.

Periodic analyses of the dose formulations of quercetin were conducted at the study laboratory and at the analytical chemistry laboratory using ultraviolet spectroscopy. During the 2-year studies, the dose formulations were analyzed at least once every 8 weeks. All formulations were within the specified 10% of the target concentrations. Results of the dose formulation analyses studies are presented in Table F2. Results of periodic referee analysis performed by the analytical chemistry laboratory indicated good agreement with the results obtained by the study laboratory (Table F3).

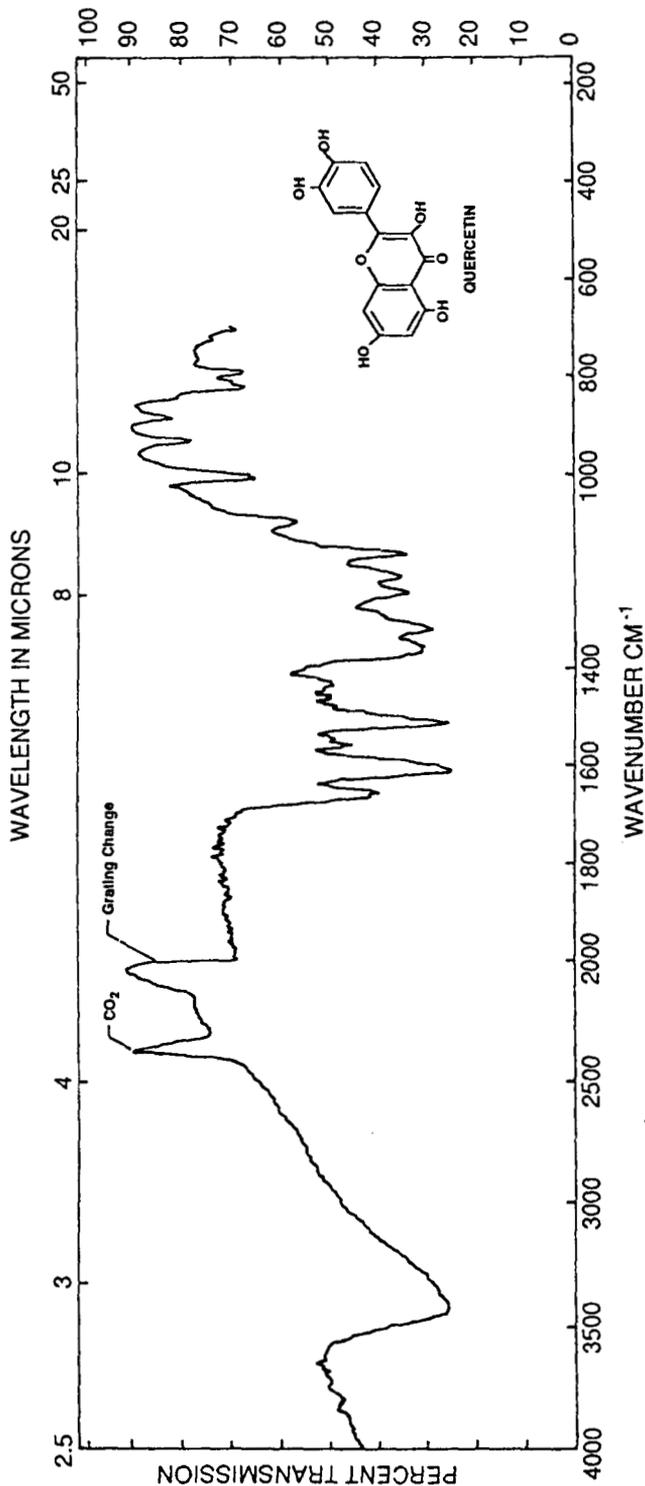


FIGURE F1
Infrared Absorption Spectrum of Quercetin

| | | |
|----------------------|---------------------------------------|-------------------|
| Instrument: Beckman | Speed: 200 cm ¹ /min (out) | Analyst: R. Grese |
| VSE: _____ | Gain: 10 x 3.87 | Date: 5/22/80 |
| Spectrum: 029N | Period: 2 | |
| Sample: Quercetin | Ordinate Scale: 0-100%T | |
| Lot No.: 969-3790-05 | Trimmer comb in reference beam | |
| Batch No.: 01 | | |

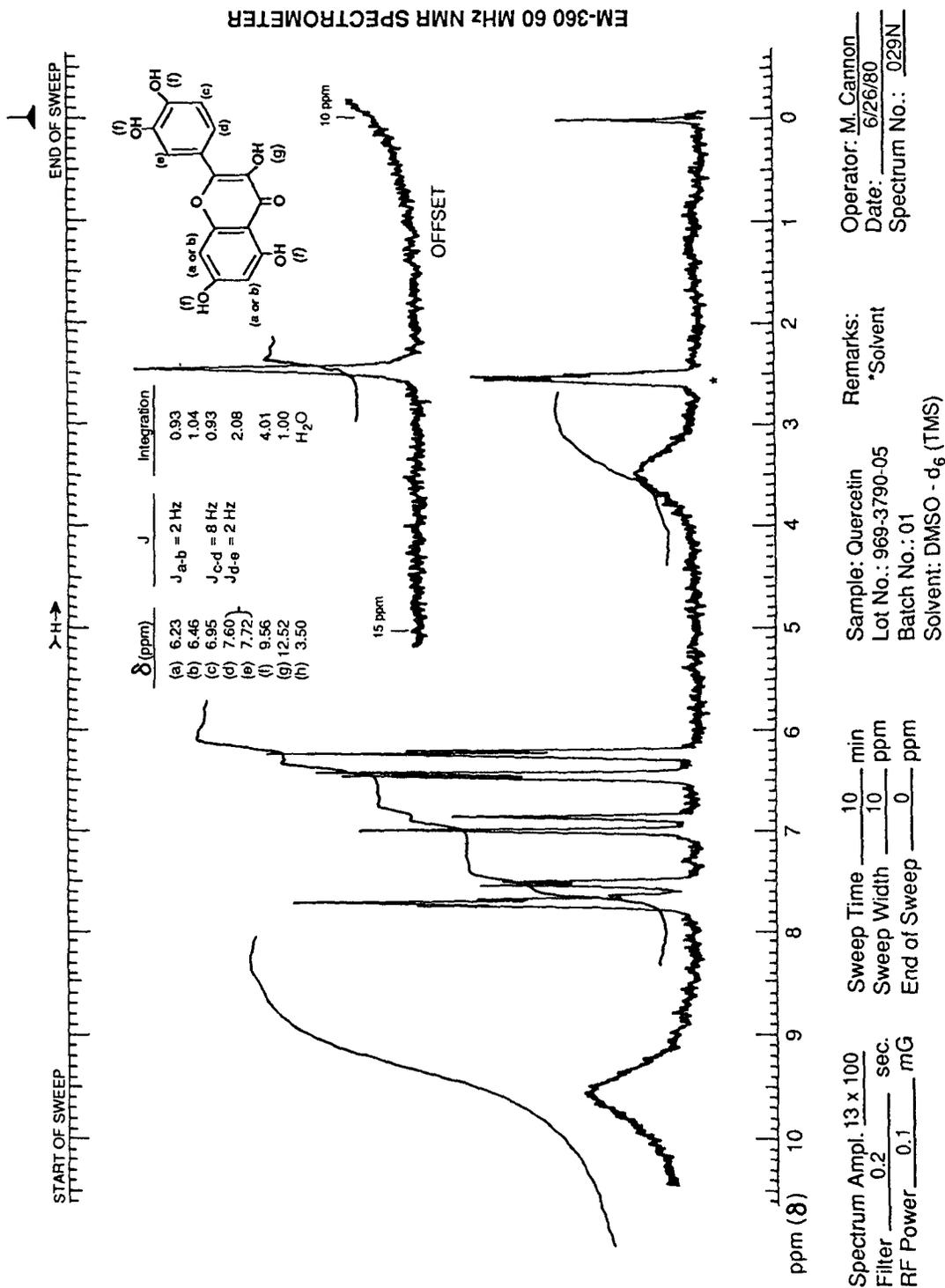


FIGURE F2
 Nuclear Magnetic Resonance Spectrum of Quercetin

TABLE F1
Preparation and Storage of Dose Formulations in the Feed Studies of Quercetin

Preparation

Dose formulations prepared weekly. Chemical-feed premix prepared by grinding quercetin and feed with mortar and pestle; premix and remaining feed layered in a blender with intensifier bar and mixed for 15 minutes.

Chemical Lot Number

969-3790-05

969-0483-18BL

Maximum Storage Time

Two weeks

Storage Conditions

Cold room at approximately 4° C, in opaque plastic bags

TABLE F2
Results of Analysis of Dose Formulations in the 2-Year Feed Studies of Quercetin

| Date Prepared | Date Analyzed | Target Concentration (ppm) | Determined Concentration ^a (ppm) | % Difference from Target |
|------------------|------------------|----------------------------|---|--------------------------|
| 17 June 1982 | 18 June 1982 | 1,000 | 970 ^b | -3 |
| 17 June 1982 | 21 June 1982 | 1,000 | 952 ^c | -5 |
| | | 1,000 | 980 ^d | -2 |
| | | 10,000 | 9,820 | -2 |
| | | 40,000 | 38,900 ^b | -3 |
| | | 40,000 | 39,100 ^c | -2 |
| | | 40,000 | 40,000 ^d | 0 |
| 17 August 1982 | 18 August 1982 | 1,000 | 980 | -2 |
| 17 August 1982 | 19 August 1982 | 10,000 | 9,970 | 0 |
| | | 40,000 | 39,900 | 0 |
| 9 November 1982 | 17 November 1982 | 1,000 | 980 | -2 |
| 9 November 1982 | 18 November 1982 | 10,000 | 9,980 | 0 |
| | | 40,000 | 40,200 | 0 |
| 7 December 1982 | 8 December 1982 | 1,000 | 990 | -1 |
| 7 December 1982 | 9 December 1982 | 10,000 | 10,000 | 0 |
| | | 40,000 | 40,500 | +1 |
| 1 March 1983 | 2 March 1983 | 1,000 | 990 | -1 |
| | | 10,000 | 9,900 | -1 |
| | | 40,000 | 39,800 | -1 |
| 5 April 1983 | 7 April 1983 | 1,000 | 980 | -2 |
| | | 10,000 | 10,200 | +2 |
| | | 40,000 | 39,200 | -2 |
| 31 May 1983 | 2 June 1983 | 1,000 | 960 | -4 |
| | | 10,000 | 10,500 | +5 |
| | | 40,000 | 41,600 | +4 |
| 19 July 1983 | 20 July 1983 | 1,000 | 1,000 | 0 |
| 19 July 1983 | 21 July 1983 | 10,000 | 9,900 | -1 |
| | | 40,000 | 40,000 | 0 |
| 2 September 1983 | 6 September 1983 | 1,000 | 970 | -3 |
| | | 10,000 | 9,950 | -1 |
| | | 40,000 | 39,800 | -1 |
| 13 December 1983 | 14 December 1983 | 1,000 | 980 | -2 |
| 13 December 1983 | 15 December 1983 | 10,000 | 10,100 | +1 |
| | | 40,000 | 39,400 | -2 |

TABLE F2
Results of Analysis of Dose Formulations in the 2-Year Feed Studies of Quercetin (continued)

| Date Prepared | Date Analyzed | Target Concentration (ppm) | Determined Concentration (ppm) | % Difference from Target |
|------------------|------------------|----------------------------|--------------------------------|--------------------------|
| 14 February 1984 | 15 February 1984 | 1,000 | 1,000 | 0 |
| | | 10,000 | 10,400 | +4 |
| | | 40,000 | 39,700 | -1 |
| 13 March 1984 | 15 March 1984 | 1,000 | 960 | -4 |
| | | 10,000 | 9,900 | -1 |
| | | 40,000 | 38,700 | -3 |
| 15 May 1984 | 17 May 1984 | 1,000 | 970 | -3 |
| | | 10,000 | 10,050 | +1 |
| | | 40,000 | 38,900 | -3 |

^a Results of duplicate analyses

^b Sample selection from top left zone of PK Blender

^c Sample selection from top right zone of PK Blender

^d Sample selection from bottom of PK Blender

TABLE F3
Results of Referee Analysis of Dose Formulations in the 2-Year Feed Studies of Quercetin

| Date Mixed | Target Concentration (ppm) | Determined Concentration (ppm) | |
|------------------|----------------------------|--------------------------------|---------------------------------|
| | | Study Laboratory ^a | Referee Laboratory ^b |
| 17 June 1982 | 1,000 | 970 | 1,020 |
| 7 December 1982 | 10,000 | 10,000 | 9,980 |
| 31 May 1983 | 40,000 | 41,600 | 40,500 |
| 13 December 1983 | 10,000 | 10,100 | 9,560 |

^a Results of duplicate analysis

^b Results of triplicate analysis

APPENDIX G
FEED AND COMPOUND CONSUMPTION
IN THE 2-YEAR FEED STUDIES

| | | |
|-----------------|---|------------|
| TABLE G1 | Feed and Compound Consumption by Male Rats in the 2-Year Feed Study of Quercetin | 162 |
| TABLE G2 | Feed and Compound Consumption by Female Rats in the 2-Year Feed Study of Quercetin | 163 |

TABLE G1
Feed and Compound Consumption by Male Rats in the 2-Year Feed Study of Quercetin

| Week | 0 ppm | | 1,000 ppm | | | 10,000 ppm | | | 40,000 ppm | | |
|---------------|-------------------|-----------------|-----------|-----------------|-----------------------|------------|-----------------|----------|------------|-----------------|----------|
| | Feed ^a | Body Weight (g) | Feed | Body Weight (g) | Dose/Day ^b | Feed | Body Weight (g) | Dose/Day | Feed | Body Weight (g) | Dose/Day |
| 1 | 20.0 | 162 | 19.8 | 160 | 124 | 19.8 | 165 | 1,198 | 18.7 | 167 | 4,495 |
| 2 | 19.0 | 195 | 17.9 | 196 | 91 | 18.0 | 203 | 889 | 16.7 | 198 | 3,387 |
| 3 | 17.7 | 225 | 18.6 | 230 | 81 | 18.5 | 233 | 794 | 18.0 | 228 | 3,145 |
| 4 | 16.6 | 253 | 16.8 | 255 | 66 | 17.5 | 257 | 680 | 19.2 | 252 | 3,047 |
| 5 | 17.7 | 271 | 17.4 | 272 | 64 | 17.6 | 272 | 648 | 18.4 | 259 | 2,837 |
| 8 | 19.0 | 310 | 18.7 | 304 | 62 | 19.3 | 312 | 619 | 19.6 | 309 | 2,529 |
| 9 | 18.8 | 322 | 18.3 | 321 | 57 | 18.1 | 325 | 559 | 18.5 | 320 | 2,310 |
| 12 | 19.7 | 340 | 21.0 | 337 | 62 | 19.1 | 334 | 572 | 19.9 | 328 | 2,425 |
| 13 | 18.1 | 354 | 18.1 | 350 | 52 | 22.7 | 343 | 663 | 22.8 | 343 | 2,656 |
| 17 | 17.6 | 376 | 18.0 | 375 | 48 | 19.9 | 373 | 535 | 17.8 | 364 | 1,954 |
| 21 | 17.8 | 399 | 17.4 | 399 | 44 | 18.7 | 395 | 474 | 17.5 | 382 | 1,833 |
| 25 | 20.8 | 416 | 24.6 | 412 | 60 | 25.1 | 409 | 613 | 23.7 | 393 | 2,408 |
| 29 | 17.7 | 434 | 17.3 | 438 | 40 | 17.6 | 430 | 410 | 17.5 | 413 | 1,696 |
| 30 | 23.4 | 445 | 22.4 | 438 | 51 | 23.1 | 435 | 532 | 22.6 | 409 | 2,210 |
| 33 | 18.6 | 456 | 18.7 | 456 | 41 | 19.1 | 448 | 426 | 18.1 | 426 | 1,701 |
| 37 | 19.1 | 457 | 19.1 | 460 | 41 | 20.6 | 458 | 450 | 21.0 | 432 | 1,946 |
| 41 | 18.7 | 464 | 18.6 | 466 | 40 | 20.0 | 464 | 431 | 21.3 | 439 | 1,943 |
| 45 | 21.1 | 469 | 18.5 | 470 | 39 | 18.3 | 464 | 395 | 19.7 | 442 | 1,784 |
| 49 | 17.7 | 481 | 18.6 | 486 | 38 | 19.3 | 482 | 400 | 21.1 | 453 | 1,864 |
| 53 | 24.0 | 484 | 21.2 | 487 | 44 | 21.1 | 481 | 438 | 22.5 | 453 | 1,989 |
| 57 | 24.6 | 487 | 24.2 | 491 | 49 | 27.1 | 488 | 555 | 27.2 | 460 | 2,369 |
| 61 | 18.2 | 478 | 18.5 | 487 | 38 | 19.3 | 484 | 399 | 19.8 | 457 | 1,733 |
| 65 | 17.4 | 485 | 17.6 | 491 | 36 | 18.2 | 483 | 378 | 18.6 | 453 | 1,639 |
| 68 | 19.0 | 486 | 18.8 | 493 | 38 | 17.7 | 490 | 361 | 18.7 | 458 | 1,631 |
| 73 | 22.4 | 492 | 21.0 | 497 | 42 | 21.8 | 491 | 444 | 22.0 | 458 | 1,921 |
| 81 | 17.5 | 492 | 18.6 | 492 | 38 | 18.3 | 483 | 379 | 20.2 | 451 | 1,792 |
| 85 | 21.2 | 485 | 20.8 | 488 | 43 | 20.1 | 480 | 418 | 22.6 | 444 | 2,037 |
| 89 | 17.2 | 476 | 16.7 | 477 | 35 | 17.0 | 473 | 359 | 19.0 | 436 | 1,742 |
| 93 | 17.9 | 473 | 18.9 | 482 | 39 | 17.0 | 465 | 366 | 19.6 | 426 | 1,835 |
| 97 | 18.5 | 479 | 19.2 | 485 | 40 | 17.4 | 450 | 386 | 20.6 | 418 | 1,972 |
| 101 | 10.9 | 447 | 11.8 | 451 | 26 | 10.5 | 427 | 247 | 12.2 | 402 | 1,213 |
| 104 | 24.8 | 464 | 24.6 | 451 | 55 | 23.1 | 440 | 525 | 24.8 | 403 | 2,463 |
| Weeks 1-13: | | | | | | | | | | | |
| Mean | 18.5 | 270 | 18.5 | 270 | 73 | 19.0 | 272 | 736 | 19.1 | 267 | 2,981 |
| Weeks 14-52: | | | | | | | | | | | |
| Mean | 19.2 | 440 | 19.3 | 440 | 44 | 20.2 | 436 | 467 | 20.0 | 415 | 1,934 |
| Weeks 53-104: | | | | | | | | | | | |
| Mean | 19.5 | 479 | 19.4 | 483 | 40 | 19.1 | 472 | 404 | 20.6 | 440 | 1,872 |

^a Grams of feed consumed per animal per day

^b Milligrams of quercetin consumed per day per kilogram of body weight

TABLE G2
Feed and Compound Consumption by Female Rats in the 2-Year Feed Study of Quercetin

| Week | 0 ppm | | 1,000 ppm | | | 10,000 ppm | | | 40,000 ppm | | |
|---------------|-------------------|-----------------|-----------|-----------------|-----------------------|------------|-----------------|----------|------------|-----------------|----------|
| | Feed ^a | Body Weight (g) | Feed | Body Weight (g) | Dose/Day ^b | Feed | Body Weight (g) | Dose/Day | Feed | Body Weight (g) | Dose/Day |
| 1 | 12.3 | 138 | 12.9 | 141 | 91 | 12.3 | 139 | 886 | 11.3 | 141 | 3,225 |
| 2 | 11.7 | 153 | 12.6 | 155 | 81 | 11.6 | 152 | 769 | 12.1 | 152 | 3,171 |
| 5 | 12.6 | 177 | 12.6 | 178 | 71 | 12.3 | 177 | 692 | 12.4 | 176 | 2,813 |
| 6 | 12.4 | 187 | 13.1 | 186 | 70 | 12.2 | 183 | 668 | 12.6 | 181 | 2,771 |
| 7 | 12.7 | 191 | 13.0 | 193 | 67 | 12.4 | 189 | 656 | 12.1 | 186 | 2,599 |
| 8 | 13.6 | 199 | 12.9 | 199 | 65 | 12.4 | 194 | 642 | 11.6 | 190 | 2,445 |
| 12 | 14.4 | 215 | 14.4 | 214 | 67 | 12.5 | 202 | 619 | 12.9 | 195 | 2,636 |
| 13 | 14.6 | 215 | 14.7 | 219 | 67 | 14.4 | 207 | 700 | 13.8 | 192 | 2,885 |
| 17 | 15.0 | 225 | 14.7 | 226 | 65 | 13.5 | 217 | 621 | 12.2 | 203 | 2,409 |
| 21 | 13.8 | 233 | 13.0 | 233 | 56 | 12.8 | 222 | 579 | 11.7 | 209 | 2,242 |
| 25 | 13.4 | 244 | 13.4 | 246 | 55 | 13.0 | 232 | 561 | 11.5 | 220 | 2,092 |
| 29 | 15.5 | 255 | 14.3 | 257 | 55 | 13.1 | 237 | 554 | 12.2 | 220 | 2,207 |
| 33 | 12.9 | 257 | 12.5 | 263 | 48 | 11.8 | 242 | 488 | 11.6 | 225 | 2,059 |
| 37 | 14.2 | 268 | 15.1 | 276 | 55 | 13.2 | 252 | 524 | 12.2 | 231 | 2,113 |
| 41 | 14.0 | 279 | 15.3 | 288 | 53 | 13.7 | 261 | 526 | 13.1 | 239 | 2,200 |
| 45 | 14.4 | 292 | 15.5 | 299 | 52 | 13.7 | 273 | 502 | 13.4 | 246 | 2,185 |
| 49 | 14.8 | 301 | 14.6 | 305 | 48 | 13.2 | 279 | 474 | 12.5 | 248 | 2,027 |
| 53 | 14.6 | 311 | 15.3 | 317 | 48 | 13.4 | 290 | 460 | 13.4 | 256 | 2,098 |
| 57 | 15.3 | 319 | 15.5 | 329 | 47 | 14.1 | 299 | 471 | 14.8 | 265 | 2,244 |
| 61 | 14.5 | 327 | 16.1 | 337 | 48 | 14.4 | 310 | 465 | 15.7 | 277 | 2,274 |
| 65 | 15.4 | 336 | 15.2 | 344 | 44 | 14.8 | 320 | 463 | 14.8 | 285 | 2,074 |
| 69 | 14.6 | 343 | 15.6 | 349 | 45 | 13.7 | 331 | 416 | 15.3 | 291 | 2,100 |
| 73 | 14.9 | 350 | 14.9 | 355 | 42 | 14.3 | 335 | 426 | 14.2 | 296 | 1,928 |
| 77 | 15.3 | 355 | 17.0 | 364 | 47 | 15.5 | 340 | 455 | 15.4 | 303 | 2,031 |
| 81 | 15.0 | 362 | 14.6 | 368 | 40 | 13.9 | 345 | 405 | 14.7 | 308 | 1,913 |
| 85 | 15.5 | 365 | 15.5 | 367 | 42 | 14.9 | 348 | 429 | 15.4 | 311 | 1,983 |
| 89 | 15.5 | 369 | 14.8 | 371 | 40 | 15.2 | 352 | 431 | 14.4 | 314 | 1,837 |
| 93 | 16.7 | 369 | 18.0 | 376 | 48 | 16.1 | 355 | 453 | 17.9 | 318 | 2,249 |
| 97 | 10.6 | 360 | 9.9 | 367 | 27 | 8.2 | 340 | 242 | 10.1 | 312 | 1,295 |
| 101 | 11.4 | 365 | 11.5 | 368 | 31 | 12.3 | 351 | 350 | 12.0 | 317 | 1,511 |
| 104 | 11.5 | 357 | 12.1 | 360 | 34 | 12.5 | 349 | 359 | 12.2 | 311 | 1,564 |
| Weeks 1-13: | | | | | | | | | | | |
| Mean | 13.1 | 184 | 13.3 | 186 | 72 | 12.5 | 180 | 704 | 12.3 | 177 | 2,818 |
| Weeks 14-52: | | | | | | | | | | | |
| Mean | 14.2 | 261 | 14.3 | 266 | 54 | 13.1 | 246 | 537 | 12.3 | 227 | 2,170 |
| Weeks 53-104: | | | | | | | | | | | |
| Mean | 14.4 | 349 | 14.7 | 355 | 42 | 13.8 | 333 | 416 | 14.3 | 297 | 1,936 |

^a Grams of feed consumed per animal per day

^b Milligrams of quercetin consumed per day per kilogram of body weight

APPENDIX H
INGREDIENTS, NUTRIENT COMPOSITION,
AND CONTAMINANT LEVELS
IN NIH-07 RAT AND MOUSE RATION

| | | |
|-----------------|---|------------|
| TABLE H1 | Ingredients of NIH-07 Rat and Mouse Ration | 166 |
| TABLE H2 | Vitamins and Minerals in NIH-07 Rat and Mouse Ration | 166 |
| TABLE H3 | Nutrient Composition of NIH-07 Rat and Mouse Ration | 167 |
| TABLE H4 | Contaminant Levels in NIH-07 Rat and Mouse Ration | 168 |

TABLE H1
Ingredients of NIH-07 Rat and Mouse Ration^a

| Ingredients ^b | Percent by Weight |
|--|-------------------|
| Ground #2 yellow shelled corn | 24.50 |
| Ground hard winter wheat | 23.00 |
| Soybean meal (49% protein) | 12.00 |
| Fish meal (60% protein) | 10.00 |
| Wheat middlings | 10.00 |
| Dried skim milk | 5.00 |
| Alfalfa meal (dehydrated, 17% protein) | 4.00 |
| Corn gluten meal (60% protein) | 3.00 |
| Soy oil | 2.50 |
| Dried brewer's yeast | 2.00 |
| Dry molasses | 1.50 |
| Dicalcium phosphate | 1.25 |
| Ground limestone | 0.50 |
| Salt | 0.50 |
| Premixes (vitamin and mineral) | 0.25 |

^a NCI, 1976; NIH, 1978

^b Ingredients ground to pass through a U.S. Standard Screen No. 16 before being mixed

TABLE H2
Vitamins and Minerals in NIH-07 Rat and Mouse Ration^a

| | Amount | Source |
|---|---------------|---|
| Vitamins | | |
| A | 5,500,000 IU | Stabilized vitamin A palmitate or acetate |
| D ₃ | 4,600,000 IU | D-activated animal sterol |
| K ₃ | 2.8 g | Menadione |
| <i>d</i> - α -Tocopheryl acetate | 20,000 IU | |
| Choline | 560.0 g | Choline chloride |
| Folic acid | 2.2 g | |
| Niacin | 30.0 g | |
| <i>d</i> -Pantothenic acid | 18.0 g | <i>d</i> -Calcium pantothenate |
| Riboflavin | 3.4 g | |
| Thiamine | 10.0 g | Thiamine mononitrate |
| B ₁₂ | 4,000 μ g | |
| Pyridoxine | 1.7 g | Pyridoxine hydrochloride |
| Biotin | 140.0 mg | <i>d</i> -Biotin |
| Minerals | | |
| Iron | 120.0 g | Iron sulfate |
| Manganese | 60.0 g | Manganous oxide |
| Zinc | 16.0 g | Zinc oxide |
| Copper | 4.0 g | Copper sulfate |
| Iodine | 1.4 g | Calcium iodate |
| Cobalt | 0.4 g | Cobalt carbonate |

^a Per ton (2,000 lb) of finished product

TABLE H3
Nutrient Composition of NIH-07 Rat and Mouse Ration

| Nutrients | Mean \pm Standard Deviation | Range | Number of Samples |
|--|-------------------------------|--------------|-------------------|
| Protein (% by weight) | 22.95 \pm 1.19 | 21.2-25.9 | 26 |
| Crude fat (% by weight) | 5.08 \pm 0.46 | 4.2-5.8 | 26 |
| Crude fiber (% by weight) | 3.50 \pm 0.60 | 2.8-4.5 | 26 |
| Ash (% by weight) | 6.66 \pm 0.21 | 6.3-7.1 | 26 |
| Amino Acids (% of total diet) | | | |
| Arginine | 1.320 \pm 0.072 | 1.310-1.390 | 5 |
| Cystine | 0.319 \pm 0.088 | 0.218-0.400 | 5 |
| Glycine | 1.146 \pm 0.063 | 1.060-1.210 | 5 |
| Histidine | 0.571 \pm 0.026 | 0.531-0.603 | 5 |
| Isoleucine | 0.914 \pm 0.030 | 0.881-0.944 | 5 |
| Leucine | 1.946 \pm 0.056 | 1.850-1.990 | 5 |
| Lysine | 1.280 \pm 0.067 | 1.200-1.370 | 5 |
| Methionine | 0.436 \pm 0.165 | 0.306-0.699 | 5 |
| Phenylalanine | 0.938 \pm 0.158 | 0.665-1.050 | 5 |
| Threonine | 0.855 \pm 0.035 | 0.824-0.898 | 5 |
| Tryptophan | 0.277 \pm 0.221 | 0.156-0.671 | 5 |
| Tyrosine | 0.618 \pm 0.086 | 0.564-0.769 | 5 |
| Valine | 1.108 \pm 0.043 | 1.050-1.170 | 5 |
| Essential Fatty Acids (% of total diet) | | | |
| Linoleic | 2.290 \pm 0.313 | 1.830-2.520 | 5 |
| Linolenic | 0.258 \pm 0.040 | 0.210-0.308 | 5 |
| Vitamins | | | |
| Vitamin A (IU/kg) | 11,565 \pm 4,265 | 4,200-22,000 | 26 |
| Vitamin D (IU/kg) | 4,450 \pm 1,382 | 3,000-6,300 | 4 |
| α -Tocopherol (ppm) | 43.58 \pm 6.92 | 31.1-48.0 | 5 |
| Thiamine (ppm) | 18.46 \pm 3.89 | 12.0-31.0 | 26 |
| Riboflavin (ppm) | 7.6 \pm 0.85 | 6.10-8.20 | 5 |
| Niacin (ppm) | 97.8 \pm 31.68 | 65.0-150.0 | 5 |
| Pantothenic acid (ppm) | 30.06 \pm 4.31 | 23.0-34.0 | 5 |
| Pyridoxine (ppm) | 7.68 \pm 1.31 | 5.60-8.80 | 5 |
| Folic acid (ppm) | 2.62 \pm 0.89 | 1.80-3.70 | 5 |
| Biotin (ppm) | 0.254 \pm 0.053 | 0.19-0.32 | 5 |
| Vitamin B ₁₂ (ppb) | 24.21 \pm 12.66 | 10.6-38.0 | 5 |
| Choline (ppm) | 3,122 \pm 416.8 | 2,400-3,430 | 5 |
| Minerals | | | |
| Calcium (%) | 1.26 \pm 0.10 | 1.04-1.43 | 26 |
| Phosphorus (%) | 0.96 \pm 0.05 | 0.90-1.10 | 26 |
| Potassium (%) | 0.900 \pm 0.098 | 0.772-0.971 | 3 |
| Chloride (%) | 0.513 \pm 0.114 | 0.380-0.635 | 5 |
| Sodium (%) | 0.323 \pm 0.043 | 0.258-0.371 | 5 |
| Magnesium (%) | 0.167 \pm 0.012 | 0.151-0.181 | 5 |
| Sulfur (%) | 0.304 \pm 0.064 | 0.268-0.420 | 5 |
| Iron (ppm) | 410.3 \pm 94.04 | 262.0-523.0 | 5 |
| Manganese (ppm) | 90.29 \pm 7.15 | 81.70-99.40 | 5 |
| Zinc (ppm) | 52.78 \pm 4.94 | 46.10-58.20 | 5 |
| Copper (ppm) | 10.72 \pm 2.76 | 8.09-15.39 | 5 |
| Iodine (ppm) | 2.95 \pm 1.05 | 1.52-3.82 | 4 |
| Chromium (ppm) | 1.85 \pm 0.25 | 1.44-2.09 | 5 |
| Cobalt (ppm) | 0.681 \pm 0.14 | 0.490-0.780 | 4 |

TABLE H4
Contaminant Levels in NIH-07 Rat and Mouse Ration

| Contaminants | Mean \pm Standard Deviation ^a | Range | Number of Samples |
|---|--|---------------|-------------------|
| Arsenic (ppm) | 0.51 \pm 0.14 | 0.18–0.74 | 26 |
| Cadmium (ppm) | 0.12 \pm 0.04 | 0.10–0.20 | 26 |
| Lead (ppm) | 0.65 \pm 0.52 | 0.27–2.93 | 26 |
| Mercury (ppm) | <0.05 | | 26 |
| Selenium (ppm) | 0.31 \pm 0.06 | 0.21–0.45 | 26 |
| Aflatoxins (ppb) | <5.0 | | 26 |
| Nitrate nitrogen (ppm) ^b | 9.66 \pm 4.49 | 2.50–19.0 | 26 |
| Nitrite nitrogen (ppm) ^b | 1.43 \pm 1.50 | 0.10–6.10 | 26 |
| BHA (ppm) ^c | 4.04 \pm 4.98 | 2.00–20.0 | 26 |
| BHT (ppm) ^c | 2.92 \pm 2.59 | 1.00–13.0 | 26 |
| Aerobic plate count (CFU/g) ^d | 146,527 \pm 143,387 | 6,200–420,000 | 26 |
| Coliform (MPN/g) ^e | 585 \pm 859 | <3.0–2400 | 26 |
| <i>E. coli</i> (MPN/g) ^f | 3.83 \pm 2.68 | <3.00–15.00 | 25 |
| <i>E. coli</i> (MPN/g) | 9.42 \pm 28.79 | <3.00–150.00 | 26 |
| Total nitrosoamines (ppb) ^g | 5.30 \pm 5.98 | 0.80–30.30 | 26 |
| <i>N</i> -Nitrosodimethylamine (ppb) ^g | 4.47 \pm 5.91 | 0.50–30.00 | 26 |
| <i>N</i> -Nitrosopyrrolidine (ppb) ^g | 0.81 \pm 0.65 | 0.30–2.20 | 26 |
| Pesticides (ppm) | | | |
| α -BHC ^h | <0.01 | | 26 |
| β -BHC | <0.02 | | 26 |
| γ -BHC | <0.01 | | 26 |
| δ -BHC | <0.01 | | 26 |
| Heptachlor | <0.01 | | 26 |
| Aldrin | <0.01 | | 26 |
| Heptachlor epoxide | <0.01 | | 26 |
| DDE | <0.01 | | 26 |
| DDD | <0.01 | | 26 |
| DDT | <0.01 | | 26 |
| HCB | <0.01 | | 26 |
| Mirex | <0.01 | | 26 |
| Methoxychlor ⁱ | <0.05 | 0.06 | 26 |
| Dieldrin ⁱ | <0.01 | 0.02 | 26 |
| Endrin | <0.01 | | 26 |
| Telodrin | <0.01 | | 26 |
| Chlordane | <0.05 | | 26 |
| Toxaphene | <0.1 | | 26 |
| Estimated PCBs | <0.2 | | 26 |
| Ronnel | <0.01 | | 26 |
| Ethion | <0.02 | | 26 |
| Trithion | <0.05 | | 26 |
| Diazinon | <0.1 | | 26 |
| Methyl parathion | <0.02 | | 26 |
| Ethyl parathion | <0.02 | | 26 |
| Malathion ^j | 0.15 \pm 0.17 | 0.05–0.81 | 26 |
| Endosulfan I | <0.01 | | 26 |
| Endosulfan II | <0.01 | | 26 |
| Endosulfan sulfate | <0.03 | | 26 |

TABLE H4
Contaminant Levels in NIH-07 Rat and Mouse Ration (continued)

- a For values less than the limit of detection, the detection limit is given for the mean.
- b Sources of contamination: alfalfa, grains, and fish meal
- c Sources of contamination: soy oil and fish meal
- d CFU = colony-forming unit
- e MPN = most probable number
- f Excludes one high value of 150 MPN/g obtained from the lot milled on 26 August 1982.
- g All values were corrected for percent recovery.
- h BHC = hexachlorocyclohexane or benzene hexachloride
- i Value and date of one observation which was above the detection limit is given under the range. All other values were less than the detection limit.
- j Fifteen lots contained more than 0.05 ppm.

APPENDIX I
SENTINEL ANIMAL PROGRAM

METHODS **172**

SENTINEL ANIMAL PROGRAM

METHODS

Rodents used in the Carcinogenesis Program of the National Toxicology Program are produced in optimally clean facilities to eliminate potential pathogens that may affect study results. The Sentinel Animal Program is part of the periodic monitoring of animal health that occurs during the toxicologic evaluation of chemical compounds. Under this program, the disease state of the rodents is monitored via serology on sera from extra (sentinel) animals in the study rooms. These animals are untreated, and these animals and the study animals are subject to identical environmental conditions. The sentinel animals come from the same production source and weanling groups as the animals used for the studies of chemical compounds.

Fifteen F344/N rats of each sex were selected at the time of randomization and allocation of the animals to the various study groups. Five animals of each designated sentinel group were killed at 6, 12, and 18 months on study. Samples for viral screening at 24 months were collected from 10 diet control animals, 5 per sex. The blood from each animal was collected and clotted, and the serum was separated. The serum was cooled on ice and shipped to Microbiological Associates' Comprehensive Animal Diagnostic Service for determination of the antibody titers. The following tests were performed:

Method of Analysis

Hemagglutination Inhibition

PVM (pneumonia virus of mice)

Sendai

KRV (Kilham rat virus)

H-1 (Toolan's H-1 virus)

Time of Analysis

6, 12, 18, and 24 months

ELISA

RCV/SDA (rat corona virus/sialodacryoadenitis virus)

6, 12, 18, and 24 months

All test results for sentinel animals were negative.

NATIONAL TOXICOLOGY PROGRAM TECHNICAL REPORTS
PRINTED AS OF AUGUST 1992

| TR No. | CHEMICAL | TR No. | CHEMICAL |
|--------|---|--------|---|
| 201 | 2,3,7,8-Tetrachlorodibenzo- <i>p</i> -dioxin (Dermal) | 274 | Tris(2-ethylhexyl)phosphate |
| 206 | 1,2-Dibromo-3-chloropropane | 275 | 2-Chloroethanol |
| 207 | Cytembena | 276 | 8-Hydroxyquinoline |
| 208 | FD & C Yellow No. 6 | 277 | Tremolite |
| 209 | 2,3,7,8-Tetrachlorodibenzo- <i>p</i> -dioxin (Gavage) | 278 | 2,6-Xylidine |
| 210 | 1,2-Dibromoethane | 279 | Amosite Asbestos |
| 211 | C.I. Acid Orange 10 | 280 | Crocidolite Asbestos |
| 212 | Di(2-ethylhexyl)adipate | 281 | HC Red No. 3 |
| 213 | Butyl Benzyl Phthalate | 282 | Chlorodibromomethane |
| 214 | Caprolactam | 284 | Diallylphthalate (Rats) |
| 215 | Bisphenol A | 285 | C.I. Basic Red 9 Monohydrochloride |
| 216 | 11-Aminoundecanoic Acid | 287 | Dimethyl Hydrogen Phosphite |
| 217 | Di(2-ethylhexyl)phthalate | 288 | 1,3-Butadiene |
| 219 | 2,6-Dichloro- <i>p</i> -phenylenediamine | 289 | Benzene |
| 220 | C.I. Acid Red 14 | 291 | Isophorone |
| 221 | Locust Bean Gum | 293 | HC Blue No. 2 |
| 222 | C.I. Disperse Yellow 3 | 294 | Chlorinated Trisodium Phosphate |
| 223 | Eugenol | 295 | Chrysotile Asbestos (Rats) |
| 224 | Tara Gum | 296 | Tetrakis(hydroxymethyl) phosphonium Sulfate & Tetrakis(hydroxymethyl) phosphonium Chloride |
| 225 | D & C Red No. 9 | 298 | Dimethyl Morpholinophosphoramidate |
| 226 | C.I. Solvent Yellow 14 | 299 | C.I. Disperse Blue 1 |
| 227 | Gum Arabic | 300 | 3-Chloro-2-methylpropene |
| 228 | Vinylidene Chloride | 301 | <i>o</i> -Phenylphenol |
| 229 | Guar Gum | 303 | 4-Vinylcyclohexene |
| 230 | Agar | 304 | Chlorendic Acid |
| 231 | Stannous Chloride | 305 | Chlorinated Paraffins (C ₂₃ , 43% chlorine) |
| 232 | Pentachloroethane | 306 | Dichloromethane (Methylene Chloride) |
| 233 | 2-Biphenylamine Hydrochloride | 307 | Ephedrine Sulfate |
| 234 | Allyl Isothiocyanate | 308 | Chlorinated Paraffins (C ₁₂ , 60% chlorine) |
| 235 | Zearalenone | 309 | Decabromodiphenyl Oxide |
| 236 | <i>D</i> -Mannitol | 310 | Marine Diesel Fuel and JP-5 Navy Fuel |
| 237 | 1,1,1,2-Tetrachloroethane | 311 | Tetrachloroethylene (Inhalation) |
| 238 | Ziram | 312 | <i>n</i> -Butyl Chloride |
| 239 | Bis(2-chloro-1-methylethyl)ether | 313 | Mirex |
| 240 | Propyl Gallate | 314 | Methyl Methacrylate |
| 242 | Diallyl Phthalate (Mice) | 315 | Oxytetracycline Hydrochloride |
| 243 | Trichloroethylene (Rats and Mice) | 316 | 1-Chloro-2-methylpropene |
| 244 | Polybrominated Biphenyl Mixture | 317 | Chlorpheniramine Maleate |
| 245 | Melamine | 318 | Ampicillin Trihydrate |
| 246 | Chrysotile Asbestos (Hamsters) | 319 | 1,4-Dichlorobenzene |
| 247 | L-Ascorbic Acid | 320 | Rotenone |
| 248 | 4,4'-Methylenedianiline Dihydrochloride | 321 | Bromodichloromethane |
| 249 | Amosite Asbestos (Hamsters) | 322 | Phenylephrine Hydrochloride |
| 250 | Benzyl Acetate | 323 | Dimethyl Methylphosphonate |
| 251 | 2,4- & 2,6-Toluene Diisocyanate | 324 | Boric Acid |
| 252 | Geranyl Acetate | 325 | Pentachloronitrobenzene |
| 253 | Allyl Isovalerate | 326 | Ethylene Oxide |
| 254 | Dichloromethane (Methylene Chloride) | 327 | Xylenes (Mixed) |
| 255 | 1,2-Dichlorobenzene | 328 | Methyl Carbamate |
| 257 | Diglycidyl Resorcinol Ether | 329 | 1,2-Epoxybutane |
| 259 | Ethyl Acrylate | 330 | 4-Hexylresorcinol |
| 261 | Chlorobenzene | 331 | Malonaldehyde, Sodium Salt |
| 263 | 1,2-Dichloropropane | 332 | 2-Mercaptobenzothiazole |
| 266 | Monuron | 333 | <i>N</i> -Phenyl-2-naphthylamine |
| 267 | 1,2-Propylene Oxide | 334 | 2-Amino-5-nitrophenol |
| 269 | Telone II® (1,3-Dichloropropene) | 335 | C.I. Acid Orange 3 |
| 271 | HC Blue No. 1 | 336 | Penicillin VK |
| 272 | Propylene | 337 | Nitrofurazone |
| 273 | Trichloroethylene (Four Rat Strains) | | |

**NATIONAL TOXICOLOGY PROGRAM TECHNICAL REPORTS
PRINTED AS OF AUGUST 1992**

| TR No. | CHEMICAL | TR No. | CHEMICAL |
|--------|---------------------------------------|--------|---|
| 338 | Erythromycin Stearate | 372 | 3,3'-Dimethoxybenzidine Dihydrochloride |
| 339 | 2-Amino-4-nitrophenol | 373 | Succinic Anhydride |
| 340 | Iodinated Glycerol | 374 | Glycidol |
| 341 | Nitrofurantoin | 375 | Vinyl Toluene |
| 342 | Dichlorvos | 376 | Allyl Glycidyl Ether |
| 343 | Benzyl Alcohol | 377 | <i>o</i> -Chlorobenzalmononitrile |
| 344 | Tetracycline Hydrochloride | 378 | Benzaldehyde |
| 345 | Roxarsone | 379 | 2-Chloroacetophenone |
| 346 | Chloroethane | 380 | Epinephrine Hydrochloride |
| 347 | D-Limonene | 381 | <i>d</i> -Carvone |
| 348 | <i>a</i> -Methyldopa Sesquihydrate | 382 | Furfural |
| 349 | Pentachlorophenol | 385 | Methyl Bromide |
| 350 | Tribromomethane | 386 | Tetranitromethane |
| 351 | <i>p</i> -Chloroaniline Hydrochloride | 387 | Amphetamine Sulfate |
| 352 | <i>N</i> -Methylolacrylamide | 388 | Ethylene Thiourea |
| 353 | 2,4-Dichlorophenol | 389 | Sodium Azide |
| 354 | Dimethoxane | 390 | 3,3'-Dimethylbenzidine Dihydrochloride |
| 355 | Diphenhydramine Hydrochloride | 391 | Tris(2-chloroethyl) Phosphate |
| 356 | Furosemide | 392 | Chlorinated Water and Chloraminated Water |
| 357 | Hydrochlorothiazide | 393 | Sodium Fluoride |
| 358 | Ochratoxin A | 395 | Probenecid |
| 359 | 8-Methoxypsoralen | 396 | Monochloroacetic Acid |
| 360 | <i>N,N</i> -Dimethylaniline | 397 | C.I. Direct Blue 15 |
| 361 | Hexachloroethane | 399 | Titanocene Dichloride |
| 362 | 4-Vinyl-1-Cyclohexene Diepoxide | 401 | 2,4-Diaminophenol Dihydrochloride |
| 363 | Bromoethane (Ethyl Bromide) | 403 | Resorcinol |
| 364 | Rhodamine 6G (C.I. Basic Red 1) | 405 | C.I. Acid Red 114 |
| 365 | Pentaerythritol Tetranitrate | 406 | γ -Butyrolactone |
| 366 | Hydroquinone | 407 | C.I. Pigment Red 3 |
| 367 | Phenylbutazone | 410 | Naphthalene |
| 368 | Nalidixic Acid | 412 | 4,4-Diamino-2,2-Stilbenedisulfonic Acid |
| 369 | Alpha-Methylbenzyl Alcohol | 415 | Polysorbate 80 |
| 370 | Benzofuran | 419 | HC Yellow 4 |
| 371 | Toluene | | |

These NTP Technical Reports are available for sale from the National Technical Information Service, U.S. Department of Commerce, 5285 Port Royal Road, Springfield, VA 22161 (703-487-4650). Single copies of this Technical Report are available without charge (and while supplies last) from the Public Health Service, National Toxicology Program, Central Data Management, P.O. Box 12233, MD A0-01, Research Triangle Park, NC 27709

**DEPARTMENT OF
HEALTH & HUMAN SERVICES**

Public Health Service
National Toxicology Program
Central Data Management
P.O. Box 12233, MD A0-01
Research Triangle Park, NC 27709

SPECIAL FOURTH-CLASS MAIL
POSTAGE & FEES PAID
DHHS / NIH
Permit No. G-763

DR. K. ABDO
NTP/NIHHS
P.O. BOX 12233, MD A0-01
RESEARCH TRIANGLE PARK, NC
27709

**NIH Publication No. 92-3140
September 1992**