Detection of VEGF in Formalin-Fixed, Paraffin-Embedded Mouse Tissue

Reagent and Antibody Information

1X Wash Buffer
3% Hydrogen Peroxide
1% BSA Diluent
1X Citrate Buffer
DAB Chromagen
Hematoxylin

Blocking Serum: Normal Donkey Serum
Jackson Immunoresearch Laboratories, Inc.
West Grove, PA 19390
www.jacksonimmuno.com
1-800-367-5296
Catalog # 017-000-001

Avidin / Biotin Blocking Kit Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # SP-2001

Primary Antibody: Rabbit Anti-VEGF Polyclonal Antibody (A-120)
Santa Cruz Biotechnology
Santa Cruz, CA 95060

www.scbt.com
1-800-457-3801
Catalog # sc-152

Negative Control Serum: Normal Rabbit Serum Jackson Immunoresearch Laboratories, Inc. West Grove, PA 19390 www.jacksonimmuno.com 1-800-367-5296 Catalog # 011-000-001

Secondary Antibody: Biotin-Conjugated Donkey Anti-Rabbit IgG Jackson Immunoresearch Laboratories, Inc. West Grove, PA 19390

www.jacksonimmuno.com
1-800-367-5296
Catalog # 711-065-152

<u>Label Complex: Vectastain Elite ABC Kit (Standard)</u>

Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # PK-6100

Staining Procedure

Positive Control Tissue: Kidney (tubules) and islets of the pancreas

Stain Localization: Cytoplasmic

1. Deparaffinize and hydrate slides through the following solutions:

Solution	Repetitions	Time
Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Wash Buffer	2 times	5 minutes

- 2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.
- 3. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.

4.	Heat-Induced Epitope Retrieval Using The Microwave			
	Place a full rack of slides into a Tissue Tek® container with 200 ml of 1X citrate buffer			
	(Insert blank slides into any empty slots in the rack to ensure even heating of slides)			
	Microwave for 5 minutes at power level 5.			

Cool for 1 minute. (Add more citrate buffer, if necessary.)

Microwave again for 5 minutes at power level 5. *Temperature Before Cooling Slides*Cool 20 minutes at room temperature.

	Rinse the slides in 2 changes of distilled water for 3 minutes each time.				
5.	5. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.				
	Block with 10% Normal Donkey Serum for 20 minutes at room temperature. Lot # Date Reconstituted				
	DO NOT RINSE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.				
7.	Avidin / Biotin Blocking Kit				
	Lot # Exp. Date New Kit: yes / no				
	Apply avidin block for 15 minutes at room temperature.				

Quick rinse in 1X Wash Buffer.

Apply biotin block for 15 minutes at room temperature.

DO NOT RINSE SECTIONS WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY. ONLY WIPE EXCESS BLOCK.

	at a 1:50 dilution, and incubate for 1 hour at room temperature.
Lot #	Exp. Date
For negative control slid	es, dilute the protein concentration of the normal rabbit serum to match that of
C	Take a 1:50 dilution from this normalized serum, and apply to the slides.
Incubate for 1 hour at ro	** *
Lot #	Date Reconstituted
9. Rinse the slides in 2 cha	nges of 1X Wash Buffer for 5 minutes each time.
10. Apply the depleay enti	robbit secondary antibody at a 1.500 dilution, and incubate for 20 minutes at
room temperature.	rabbit secondary antibody at a 1:500 dilution, and incubate for 30 minutes at
I of #	Date Reconstituted
Lot II	Bute Reconstituted
11. Rinse the slides in 2 ch	anges of 1X Wash Buffer for 5 minutes each time.
12. Apply the label comple	ex from the Standard Elite Kit, and incubate for 30 minutes at room
temperature. (Prepare	at least 30 minutes prior to use.)
Exp. Date	New Kit: yes / no
13. Rinse the slides in 2 ch	anges of 1X Wash Buffer for 5 minutes each time.
14 Apply the DAR chrom	agen and incubate in the dark for 6 minutes at room temperature.
(Add 1 drop of DAB po	
	Exp. Date New Kit: yes / no
15. Rinse the slides in tap	water 3 minutes.
Counterstain with Harr	is Hematoxylin for 20 seconds.
17. Rinse the slides in tap	votor until votor is along
17. Killse tile shues ili tap	vater until water is clear.
18. Gently agitate slides in	1X Wash Buffer until they turn blue.
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19. Dehydrate through the	following solutions:

Solution	Repetitions	Time
95% Ethanol	1 time	3 minutes
100% Ethanol	3 times	3 minutes
Xylene	2 times	5 minutes