

# Detection of Insulin in Formalin-Fixed, Paraffin-Embedded Mouse Tissue

## Reagent and Antibody Information

[1X Wash Buffer](#)  
[3% Hydrogen Peroxide](#)  
[1% BSA Diluent](#)  
[DAB Chromagen](#)  
[Hematoxylin](#)

### Staining Kit: M.O.M. Immunodetection Peroxidase Kit

Vector Laboratories, Inc.  
Burlingame, CA 94010  
[www.vectorlabs.com](http://www.vectorlabs.com)  
1-800-227-6666  
Catalog # PK-2200

**Note:** This kit contains all reagents necessary to make the blocking reagent, secondary antibody and label complex.

### Avidin / Biotin Blocking Kit

Vector Laboratories, Inc.  
Burlingame, CA 94010  
[www.vectorlabs.com](http://www.vectorlabs.com)  
1-800-227-6666  
Catalog # SP-2001

### Primary Antibody: Monoclonal Anti-Insulin Antibody

Sigma-Aldrich  
St. Louis, MO  
[www.sigmaaldrich.com](http://www.sigmaaldrich.com)  
1-800-325-3010  
Catalog # I2018

### Negative Control Serum: Normal Mouse Serum

Jackson ImmunoResearch Laboratories, Inc.  
West Grove, PA 19390  
[www.jacksonimmuno.com](http://www.jacksonimmuno.com)  
1-800-367-5296  
Catalog # 015-000-001

## Staining Procedure

Positive Control Tissue: Pancreas (islets of Langerhans)  
Stain Localization: Cytoplasmic

1. Deparaffinize and hydrate slides through the following solutions:

Solution	Repetitions	Time
Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Wash Buffer	2 times	5 minutes

2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.
3. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.

4. Heat-Induced Epitope Retrieval Using The Decloaker

Add 500 ml of distilled water to the pan inside the decloaker.

Place a full rack of slides into a Tissue Tek® container with 200 ml of 0.1 M citrate buffer  
(Insert blank slides into any empty slots in the rack to ensure even heating of slides)

Place the container stably inside the pan and decloak for 5 minutes. *Maximum Pressure* \_\_\_\_\_

Depressurize for 10 minutes.

Remove pan top and cool for 10 minutes. *Temperature Before Cooling Slides* \_\_\_\_\_

Rinse the slides in 2 changes of distilled water for 3 minutes each time.

5. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each time.

M.O.M Peroxidase Kit

Exp. Date \_\_\_\_\_ New Kit: yes / no

6. Apply the blocking reagent from the M.O.M. Kit, and incubate for 1 hour at room temperature.  
(Add 2 drops of the Mouse IgG Blocking Reagent to 2.5 ml of 1X PBS.)

DO NOT RINSE THE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.

7. Avidin / Biotin Blocking Kit

Lot # \_\_\_\_\_ Exp. Date \_\_\_\_\_ New Kit: yes / no

Apply avidin block for 15 minutes at room temperature.

Quick rinse in 1X Wash Buffer.

Apply biotin block for 15 minutes at room temperature.

DO NOT RINSE SLIDES WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY.  
ONLY WIPE EXCESS BUFFER.

**M.O.M. Diluent:** Add 600ul of the Protein Concentrate stock solution to 7.5 ml of 1X PBS. Use this as the diluent for the primary, negative, and secondary antibodies.

8. Apply the primary antibody at a 1:16,000 dilution, and incubate for 5 minutes at room temperature.  
Lot # \_\_\_\_\_ Exp. Date \_\_\_\_\_

For negative control slides, dilute the protein concentration of the normal mouse serum to match that of the primary antibody. Make a 1:16,000 dilution from this normalized serum, and apply to the slides. Incubate for 5 minutes at room temperature.

Lot # \_\_\_\_\_ Date Reconstituted \_\_\_\_\_

9. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.

10. Apply the secondary antibody from the M.O.M. Kit, and incubate for 10 minutes at room temperature. (Add 10ul of the Biotinylated anti-Mouse IgG Reagent to 2.5 ml of the M.O.M. Diluent).

11. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.

12. Apply the label complex from the M.O.M. Kit, and incubate for 5 minutes at room temperature. (Add 2 drops of Reagent A to 2.5 ml of 1X PBS. Mix. Then add 2 drops of Reagent B and mix. Prepare at least 30 minutes prior to use.)

13. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.

14. Apply the DAB chromagen, and incubate in the dark for 6 minutes at room temperature. (Add 1 drop of DAB per ml of substrate)

Lot # \_\_\_\_\_ Exp. Date \_\_\_\_\_ New Kit: yes / no

15. Rinse the slides in tap water 3 minutes.

16. Counterstain with Harris Hematoxylin for 20 seconds.

17. Rinse the slides in tap water until water is clear.

18. Gently agitate slides in 1X Wash Buffer until they turn blue.

19. Dehydrate through the following solutions:

<b>Solution</b>	<b>Repetitions</b>	<b>Time</b>
95% Ethanol	1 time	3 minutes
100% Ethanol	3 times	3 minutes
Xylene	2 times	5 minutes

20. Coverslip

*Updated 10/17/06*