## Detection of IgM in Formalin-Fixed, Paraffin-Embedded Rat Tissue

## **Reagent and Antibody Information**

1X Wash Buffer
3% Hydrogen Peroxide
1% BSA Diluent
DAB Chromagen
Hematoxylin

Blocking Serum: Normal Goat Serum
Jackson Immunoresearch Laboratories, Inc.
West Grove, PA 19390
www.jacksonimmuno.com
1-800-367-5296
Catalog # 005-000-121

Avidin / Biotin Blocking Kit Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # SP-2001

Primary Antibody: Goat Anti-Rat IgM Antibody HRP-Conjugated
Bethyl Laboratories, Inc
Montgomery, TX 77356

www.bethyl.com
1-800-338-9579
Catalog # A110-100P

## **Staining Procedure**

Positive Control Tissue: Lymph Node Stain Localization: Cytoplasmic (Secreted)

1. Deparaffinize and hydrate slides through the following solutions:

| Solution       | Repetitions | Time      |
|----------------|-------------|-----------|
| Xylene         | 2 times     | 5 minutes |
| 100% Ethanol   | 2 times     | 3 minutes |
| 95% Ethanol    | 2 times     | 3 minutes |
| 1X Wash Buffer | 2 times     | 5 minutes |

| 2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.                                                                       |
|---------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| 3. Rinse slides in 2 changes of 1X Wash Buffer for 5 minutes each.                                                                                                  |
| 4. Block with 10% Normal Goat Serum for 20 minutes at room temperature.  Lot # Date Reconstituted                                                                   |
| DO NOT RINSE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.                                                                                                               |
| 5. <u>Avidin / Biotin Blocking Kit</u> Lot # Exp. Date New Kit: yes / no                                                                                            |
| Apply avidin block for 15 minutes at room temperature.  Quick rinse in 1X Wash Buffer.  Apply biotin block for 15 minutes at room temperature.                      |
| DO NOT RINSE SECTIONS WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY. ONLY WIPE EXCESS BLOCK.                                                                           |
| 6. Apply primary antibody at a 1:200 dilution, and incubate for overnight at 4°C.  Lot # Exp. Date                                                                  |
| **************************************                                                                                                                              |
| 7. Bring the slides up to room temperature in 1X Wash Buffer for at least 15 minutes.                                                                               |
| 8. Apply the DAB chromagen, and incubate in the dark for 6 minutes at room temperature.  (Add 1 drop of DAB per ml of substrate)  Lot # Exp. Date New Kit: yes / no |
| 9 Rinse the clides in tan water 3 minutes                                                                                                                           |

- 9. Rinse the slides in tap water 3 minutes.
- 10. Counterstain with Harris Hematoxylin for 20 seconds.
- 11. Rinse the slides in tap water until water is clear.

- 12. Gently agitate slides in 1X Wash Buffer until they turn blue.
- 13. Dehydrate through the following solutions:

| Solutions    | Repetitions | Time      |
|--------------|-------------|-----------|
| 95% Ethanol  | 1 time      | 3 minutes |
| 100% Ethanol | 3 times     | 3 minutes |
| Xylene       | 2 times     | 5 minutes |

14. Coverslip

Updated 02/19/09