Detection of GDF-15 in Formalin-Fixed, Paraffin-Embedded Human Tissue

Reagent and Antibody Information

1X Wash Buffer
3% Hydrogen Peroxide
1% BSA Diluent
1X Citrate Buffer
DAB Chromagen
Hematoxylin

Blocking Serum: Normal Donkey Serum
Jackson Immunoresearch Laboratories, Inc.
West Grove, PA 19390
www.jacksonimmuno.com
1-800-367-5296
Catalog # 017-000-011

Avidin / Biotin Blocking Kit Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # SP-2001

Primary Antibody: Goat Polyclonal Anti-Human GDF-15 Antibody R&D Systems Minneapolis, MN 55413 www.rndsystems.com 1-800-343-7475 Catalog # AF957

Negative Serum Control: Normal Goat Serum Jackson Immunoresearch Laboratories, Inc. West Grove, PA 19390 www.jacksonimmuno.com 1-800-367-5296 Catalog # 005-000-121

Secondary Antibody: Donkey Anti-Goat IgG (H+L) Biotin-SP-Conjugated Jackson Immunoresearch Laboratories, Inc.
West Grove, PA 19390
www.jacksonimmuno.com
1-800-367-5296
Catalog # 705-065-147

<u>Label Complex: Peroxidase-Conjugated Streptavidin SS Label</u>

Biogenex Laboratories San Ramon, CA 94583 www.biogenex.com 1-800-421-4149 Catalog # HK330-9K

Staining Procedure

Positive Control Tissue: Prostate Carcinoma

Stain Localization: Cytoplasmic

1. Deparaffinize and hydrate slides through the following solutions:

Solution	Repetitions	Time
Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Wash Buffer	2 times	5 minutes

- 2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.
- 3. Rinse slides in 2 changes of 1X Wash Buffer for 5 minutes each.

4. <u>Heat-Induced Epitope Retrieval Using The Decloaker</u> Add 500 ml of distilled water to the pan inside the decloaker. Place a full rack of slides into a Tissue Tek® container with 200 ml of 1X citrate buffer	
Place a full rack of slides into a Tissue Tek® container with 200 ml of 1X citrate buffer	
(Insert blank slides into any empty slots in the rack to ensure even heating of slides)	
Place the container stably inside the pan and decloak for 5 minutes. Maximum Pressure	
Depressurize for 10 minutes.	
Remove pan top and cool for 10 minutes. Temperature Before Cooling Slides	_
Rinse the slides in 2 changes of distilled water for 3 minutes each time.	
5. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each time.	
6. Block with 10% Normal Donkey Serum for 20 minutes at room temperature.	
Lot # Date Reconstituted	

DO NOT RINSE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.

7. Avidin / Biotin	<u>n Blocking Kit</u>			
Lot #	Exp. Date	New Kit:	yes	/ no
Apply avidin b	plock for 15 minutes at room tem	perature.		
Quick rinse in	1X Wash Buffer.			
Apply biotin b	lock for 15 minutes at room temp	nerature		

DO NOT RINSE SECTIONS WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY. ONLY WIPE EXCESS BLOCK.

8. Apply primary antibody at a 1:1500 dilution, and incubate for 1 hour at room temperature. Lot # Exp. Date
For negative control slides, dilute the protein concentration of the normal goat serum to match that of the primary antibody. Make a 1:1500 dilution from this normalized serum, and apply to the slides. Incubate for 1 hour at room temperature. Lot # Date Reconstituted
9. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each time.
10. Apply the donkey anti-goat secondary antibody at a 1:500 dilution, and incubate for 30 minutes at room temperature. Lot # Date Reconstituted
11. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each time.
12. Apply the Streptavidin SS Label, and incubate for 30 minutes at room temperature. Lot # Exp Date
13. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each time.
14. Apply the DAB chromagen, and incubate in the dark for 6 minutes at room temperature. (Add 1 drop of DAB per ml of substrate) Lot # Exp. Date New Kit: yes / no
15. Rinse the slides in tap water 3 minutes.
16. Counterstain with Harris Hematoxylin for 20 seconds.
17. Rinse the slides in tap water until water is clear.
18. Gently agitate slides in 1X Wash Buffer until they turn blue.

Solutions	Repetitions	Time
95% Ethanol	1 time	3 minutes
100% Ethanol	3 times	3 minutes
Xylene	2 times	5 minutes

19. Dehydrate through the following solutions:

20. Coverslip