Detection of CYP2C11 in Formalin-Fixed, Paraffin-Embedded Rat Tissue

Reagent and Antibody Information

1X Wash Buffer
3% Hydrogen Peroxide
1% BSA Diluent
1X Citrate Buffer
DAB Chromagen
Hematoxylin

Blocking Solution: Dakocytomation Protein Block Serum-Free Ready-To-Use

Dakocytomation Corporation Carpinteria CA 93013 www.dakousa.com 1-800-235-5763

Code No. X0909

Avidin / Biotin Blocking Kit

Vector Laboratories, Inc.
Burlingame, CA 94010
www.vectorlabs.com
1-800-227-6666
Catalog # SP-2001

Primary Antibody: Rabbit Polyclonal To Cytochrome P450 2C11 Antibody

Abcam Inc Cambridge, MA 02139 www.abcam.com 1-888-772-2226 Catalog # ab3571

Negative Control Serum: Normal Rabbit Serum Jackson Immunoresearch Laboratories, Inc. West Grove, PA 19390 www.jacksonimmuno.com 1-800-367-5296 Catalog # 011-000-001

Staining Kit: LSAB+ System-HRP
Dakocytomation Corporation
Carpinteria CA 93013
www.dakousa.com
1-800-235-5763
Code No. K0690

Note: This kit includes reagents needed for the secondary antibody (link) and label complex.

Staining Procedure

Positive Control Tissue: Liver (upregulated by treatment) Stain Localization: Centilobular cytoplasmic staining pattern

1. Deparaffinize and hydrate slides through the following solutions:

Solution	Repetitions	Time
Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Wash Buffer	2 times	5 minutes

- 2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.
- 3. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.

4.	Heat-Induced Epitope Retrieval Using The Microwave				
	Place a full rack of slides into a Tissue Tek® container with 200 ml of 1X citrate buffer				
	(Insert blank slides into any empty slots in the rack to ensure even heating of slides)				
	Microwave for 5 minutes at power level 5.				
	Cool for 1 minute. (Add more citrate buffer, if necessary.)				
	Microwave again for 5 minutes at power level 5. Temperature Before Cooling Slides				
	Cool 20 minutes at room temperature.				
	Rinse the slides in 2 changes of distilled water for 3 minutes each time.				
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5. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each time.

Block with the Dako Protein Blocking Reagent for 10 minutes at room temperature.
Lot # Exp Date
DO NOT RINSE THE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.
Avidin / Biotin Blocking Kit
Lot #
Apply avidin block for 15 minutes at room temperature.
Quick rinse in 1X Wash Buffer.
Apply biotin block for 15 minutes at room temperature.
DO NOT RINSE SLIDES WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY.
ONLY WIPE EXCESS BUFFER.
Apply the primary antibody at a 1:100 dilution, and incubate for 1 hour at room temperature.
Date Aliquoted
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For negative control slides, dilute the protein concentration of the normal rabbit serum to match that of

the primary antibody. Make a 1:100 dilution from this normalized serum, and apply to the slides. Incubate for 1 hour at room temperature. Lot # Date Reconstituted
9. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.
LSAB+ Kit Lot # Exp Date
10. Apply the Link (yellow bottle) from the LSAB+ Kit, and incubate for 30 minutes at room temperature.
11. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.
12. Apply the Label (red bottle) from the LSAB+ Kit, and incubate for 30 minutes at room temperature.
13. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.
14. Apply the DAB chromagen, and incubate in the dark for 6 minutes at room temperature. (Add 1 drop of DAB per ml of substrate) Lot # Exp Date New Kit: yes / no
15. Rinse the slides in tap water 3 minutes.
16. Counterstain with Harris Hematoxylin for 20 seconds.
17. Rinse the slides in tap water until water is clear.

19.	Dehy	vdrate	through	the	follo	wing	solutions:

Solution	Repetitions	Time	
95% Ethanol	1 time	3 minutes	
100% Ethanol	3 times	3 minutes	
Xylene	2 times	5 minutes	

18. Gently agitate slides in 1X Wash Buffer until they turn blue.

20. Coverslip

Updated 08/21/06