

Building Sustainable Alliances

April 1-3, 2001

JSU Liberal Arts Building Jackson State University Jackson, MS 39217







Expanding Opportunities in Oceanic and Atmospheric Sciences

Building Sustainable Alliances

April 1-3, 2001

JSU Liberal Arts Building Jackson State University Jackson, MS 39217

Dedicated to the Memory

of

Dr. Nancy Foster

1941 - 2000

88

A champion and leader who embodied and fully embraced the vision that by expanding opportunities, NOAA could become more diverse and inclusive.

Nancy, "Expanding Opportunities" is your epitaph.

Conference Steering Committee

Dr. James H. Arrington Vice President, Academic Affairs South Carolina State University

Ms. Jocelyn Bennett-Martin EEO/MSI Manager NOAA/National Ocean Service Silver Spring

Dr. Brian L. Bingham Minorities in Marine Science Undergraduate Program Shannon Point Marine Center,WA

Dr. Raymond R. Carthy Assistant Unit Leader - Wildlife University of Florida CFWRU Gainesville

> Dr. Paulinus Chigbu Department of Biology Jackson State University

Ms. Essie Coleman-Duffie Fishery Management Specialist NOAA Fisheries, Miami

Dr. Clayton B. Cook Senior Scientist Harbor Branch Oceanographic Institution, Ft. Pierce FL

Dr. Paul J. Croft Meteorology Program Coordinator Physics Dep't./Atmospheric Sci. Jackson State University

> Dr. Benjamin E. Cuker Associate Professor Hampton University, VA

Dr. Mack Felton, Jr., Professor Dillard University, New Orleans

Dr. Judith F. Freeman Chief, NOAA Chesapeake Bay Program Office, Annapolis, MD

Mr. Jose Garcia Meteorologist In Charge NOAA, National Weather Service, Amarillo, TX

Dr. Matthew R. Gilligan, Director Marine Science Laboratory Savannah State University Dr. Mark Hardy Interim Assoc. Vice President for Academic Affairs and Professor of Biology Jackson State University

> Dr. Emorcia V. Hill Senior Analyst Abt Associates, Inc. Cambridge, MA

Ms. Natalie Huff Program Manager for EEO and Diversity NOAA, NMFS, Silver Spring MD

Dr. Steven G. Hughes Assistant Unit Leader Maryland Co-op. Fish and Wildlife Princess Anne

Dr. Ambrose Jearld, Jr. **Steering Committee Chair** Research Planning and Coordination NOAA, NMFS/NEFSC Woods Hole, MA

Dr. Darryl J. Keith, Oceanographer U.S. Environmental Protection Agency Narragansett, RI

Dr. Margaret S. Leinen Ass't. Director for GeoSciences National Science Foundation Washington, D.C.

Dr. Diana I. Marinez, Dean College of Science & Technology Texas A&M University-Corpus Christi

Dr. Livingston Marshall, Assoc. Prof. Department of Biological Sciences Morgan State University, Baltimore

> Dr. Abdul K. Mohamed Dean, School of Science & Technology Jackson State University

Dr. Mildred D. Ofosu, Director Office of Sponsored Programs Assistant Provost for Research Delaware State University Dr. Joseph M. Okoh Associate Professor and Chairman Department of Natural Sciences University of Maryland Eastern Shore

> Dr. N. Joyce Payne, Director NASULGC/ Office for the Advancement of Public Black Colleges Washington, D.C.

Mr. Marlin O. Perkins GOES DSC Program Manager NOAA, NESDIS, Suitland, MD

Dr. Larry Robinson Professor/Director Environmental Sciences Institute Florida A&M University, Tallahassee

Dr. Carlos Robles Professor/Director, CEA-CREST California State University, LA

Dr. John Snow, Dean Director, Weather Center Programs University of Oklahoma, Norman

Dr. Denise Stephenson-Hawk Chairman The Stephenson Group, Atlanta

Dr. Sheila Stiles, Research Geneticist NOAA, NMFS/NEFSC, Milford, CT

Mr. Robert H. Stockman, Director Strategic Management Staff OFA/NOAA Washington, D.C.

Mr. Shelton Swanier, Director Office of Strategic Initiatives School of Science & Technology Jackson State University

Dr. Edward Thomas, Jr. Assistant Professor, Physics Auburn University

Mr. Benjamin Watkins Operations Planning Officer National Climatic Data Center NOAA/NESDIS, Asheville, NC

Dr. William L. White, Associate Dean School of Science and Technology Jackson State University

Conference Sponsors and Co-Sponsors



National Marine Fisheries Service

National Weather Service

National Ocean Service

Office of Oceanic and Atmospheric Research

National Environmental Satellite, Data, and Information Service

and

Jackson State University

National Association of State Universities and Land-Grant Colleges

National Association of Marine Laboratories

American Society of Limnology and Oceanography

National Association For Equal Opportunity in Higher Education

Hon. Thad Cochran is the three-term senior United States Senator from Mississippi, first elected to the Senate in 1978. He chairs Appropriations and National Security Subcommittees in the Senate, and he is a member of the Agriculture and Rules Committees. He serves on the Board of Visitors of the U.S. Naval Academy, and the Board of Regents of the Smithsonian Institution. He authored of the National Missile Defense Act of 1999, as well as key provisions of several farm bills and conservation initiatives. Before serving in Congress, Sen. Cochran practiced law in Jackson, and he was an officer in the U.S. Navy. He was born in Pontotoc, Mississippi, was educated in the public schools of the state, and earned his college and law degrees from the University of Mississippi. He studied international law for a year under a Rotary Foundation Fellowship at Trinity College Dublin, Ireland, and he has been awarded four honorary degrees from other colleges and universities

DeLois (Dee) A. Cutter is a native of El Dorado, Arkansas and a graduate of Grambling State University. Ms. Cutter has held a number of executive management positions with major corporations such as Control Data Corporation and Progressive Insurances Companies, and presently owns her own business, Tal-Cut Company, providing personnel, information technology, engineering, and telecommunications services to clients. During nine years as Executive Vice-President, Ms. Cutter transformed the company from a contract programming service into a "high tech" government provider. As President for the last decade, Ms. Cutter has led to company to nation recognition in the small- business arena of high technology. In 1999, Ms. Cutter accepted the federal Small Business Administration's Award of Excellence for the services provided by Tal-Cut as a prime contractor to the General Services Administration. Ms. Cutter has served on numerous boards, and received many awards and citations for community service, including those of the Black Business Association and the National Association of Small Disadvantaged Businesses. Ms. Cutter avows the keys to success are persistence, determination, and quality work.

Darrell Jay Grimes, Ph.D. is dean of the College of Marine Sciences and professor of coastal sciences at the University of Southern Mississippi, and directs the University's Gulf Coast Research Laboratory. He serves on the National Research Council's Ocean Studies Board, is cochair of NASULGC's Board on Oceans and Atmosphere and president of the U.S. Federation of Culture Collections. He received his doctorate in microbiology from Colorado State University in 1971, after attending Drake University. As a scientist and program manager at the U.S. Department of Energy, Dr. Grimes worked in the Subsurface Science and Ocean Margins Program, was program manager of the Microbial Genome Program, and co-manager of the Natural and Accelerated Bioremediation Research Program. Grimes has published and presented extensively in the fields of microbial ecology, marine microbiology, and bacterial systematics; serves as a consultant and expert witness in areas pertaining to water-borne disease. Dr. Grimes is a member of the American Society for Microbiology, the American Association for the Advancement of Science, Sigma Xi, the U.S. Federation for Culture Collections. and is a charter member of the American Elasmobranch Society.

Mayor Harvey Johnson, Jr. is the first African-American mayor of Jackson, Mississippi, first elected in June 1997. He was born in the river city of Vicksburg, Mississippi, where he attended public school and graduated from Temple High School. He received his bachelor's degree from Tennessee State University and his master's degree the University of Cincinnati, both in political science. Early in his career, Mayor Johnson founded and served as executive director of the Mississippi Institute for Small Towns, a nonprofit agency, assisting towns in the Mississippi Delta with housing, community development, and infrastructure needs. He was on the faculty at Jackson State University, where he taught political science and directed the Center for Technology Transfer. He was a Captain in the United States Air Force, has served on numerous governmental and community boards and commissions, and serves as a mentor at Rowan Middle School. Mayor Johnson is a member of the U.S. Conference of Mayors, the National Conference of Black Mayors, and the National Conference of Democratic Mayors, the Alpha Phi Alpha Fraternity, Inc., and is a charter member of 100 Black Men of Jackson, Inc.

Dr. Ronald Mason, Jr. is the president of Jackson State University. He earned his undergraduate and law degrees from Columbia University, and is also a graduate of the Harvard University Institute of Educational Management. Prior to his appointment at JSU, Dr. Mason founded and served as Executive Director of the Tulane-Xavier Center in New Orleans, where he coordinated the two universities' extensive involvement in public housing, economic development, and public education. In 1996, he was tapped by the U.S. Secretary of Housing and Urban Development to oversee recovery of the Housing Authority of New Orleans. During his 18 years at Tulane, Dr. Mason served as Senior Vice President and General Counsel, and Vice President for Finance and Operations. He established the Tulane-Xavier-Loyola-Dillard Martin Luther King Week for Peace and the Amistad Research Center, one of the nation's largest collections of original documents and art on the experience of minorities in the United States. Dr. Mason has written, presented and conducted research extensively on issues related to urban development, diversity, and multiculturalism. He is involved in numerous public service activities, including membership on academic and community boards and commissions. He is a recipient of the Mayors Medal of Honor from the City of New Orleans, and the Martin Luther King Lifetime Achievement Award from Dillard-Loyola-Tulane and Xavier Universities.

Dr. N. Joyce Payne is Director of the Office for the Advancement of Public Black Colleges, an information and advocacy office that represents 35 of the nation's largest and most prestigious historically Black public colleges and universities. The office is part of the National Association of State Universities and Land-Grant Colleges in cooperation with the American Association of State Colleges and Universities. She is a noted authority on women's issues in relation to higher education and labor force participation, on which she has published and spoken extensively. Dr. Payne formerly served as President of Global Systems, Inc. and as a senior staff member under the Carter administration with the President's Advisory Committee for Women: President's National Advisory Council on Women's Educational Programs, and the White House Conference on Families. She is a founding member of the D.C. Chapter of the Coalition of 100 Black Women, Inc. and a member of Zeta Phi Beta Sorority. She served on the Southern Education Foundation's Task Force on Desegregation, and the W.K. Kellogg Foundation's Task Force on African-American Boys and Men. In 1987, she created the nationally recognized Thurgood Marshall Scholarship Fund, designed exclusively for exceptional students at public Black colleges and universities. An alumna of the District of Columbia Teachers Colleges, where she received the Bachelor of Science degree, Dr. Payne earned master's and doctoral degrees from Atlanta University. She has received awards from the Ford Foundation, the Spenser Foundation, a Black History Award from the U.S. Air Force, and serves on the distinguished Foreign Service Performance Evaluation Board, U.S. Department of State.

Hon. Bennie G. Thompson began his political career more than 32 years ago, winning his first elected office as Alderman in his hometown of Bolton, MS. He subsequently served as Mayor of Bolton and Supervisor of Hinds County District Two, was elected to the U.S. House of Representatives in a special election in 1993, and represents the state's Second Congressional District. He presently serves on the Committee on Budget and the House Committee on Agriculture. He was educated in public schools of Hinds County and received his bachelor's degree in political science from Tougaloo College, and a master's of science degree from Jackson State University. He has completed extensive coursework toward a doctorate in public administration at the University of Southern Mississippi. A lifelong activitist for civil rights, has worked as a grassroots volunteer, a labor organizer, and community leader as well as a politician. He was one of the original plaintiffs in the Ayers case, a landmark decision addressing disparities in funding between historically Black colleges and white colleges. He has been honored for his accomplishments by the Housing Assistance Council and the National Black Nurses Foundation; he received an honorary Doctor of Law degree from Claflin College in Orangeburg, SC.

AGENDA - 1

AGENDA

Expanding Opportunities in Oceanic and Atmospheric Sciences Building Sustainable Alliances

April 1-3, 2001 Jackson State University Jackson, MS

SUNDAY April 1, 2001

1:00 pm	Registration	Quality Inn Hotel & Suites
6:00 pm - 8:00 pm	Pre-Conference Reception Remarks by The Honorable Bennie G. Thomps United States House of Representatives	Presidential Hall son,

MONDAY April 2, 2001 JSU Liberal Art Building

7:45 am - 8:15 am	Registration	Lobby
	PLENARY SESSION	Room 266 (overflow, Room 146)
8:15 am - 8:45 am	Opening & Dedication Dr. Ambrose Jearld, Jr, Chief Research Planning a tional Marine Fisheries Service/NOAA, Conference	and Coordination Staff, Na-
	Greetings and Welcome Dr. Ronald Mason, Jr., President, Jackson State U The Honorable Harvey Johnson, Jr., Mayor of Jack The Honorable Thad Cochran, United States Sena Rear Admiral Evelyn Fields, Office of Marine and Aviation Operations, NOAA	kson, Mississippi
8:45 am - 9:15 am	 Charge to the Conference Rear Admiral Evelyn Fields, Office of Marine and Aviation Operations, NOAA Dr. N. Joyce Payne, Director, Office for the Advance Colleges, National Association of State Universities Grant Colleges (NASULGC) 	

9:15 am - 10:30 am	Perspectives from NOAA's Leadership - Progress and Impediments: Building Sustainable Alliances
Moderators:	 Dr. Denise Stephenson-Hawk, Chairman The Stephenson Group and Member, NOAA Science Advisory Board Dr. William White, Associate Dean, School of Science and Technology, Jackson State University
Panelists:	 Mr. John Oliver, Deputy Assistant Administrator, NOAA National Marine Fisheries Service Ms. Louisa Koch, Deputy Assistant Administrator, NOAA Office of Oceanic and Atmospheric Research, Captain Ted Lillestolen, Deputy Assistant Administrator, NOAA National Ocean Service Ms. Mary Glackin, Deputy Assistant Administrator, NOAA National Environmental Satellite, Data, and Information Service Dr. Edward Johnson, Director of Strategic Planning and Policy, NOAA National Weather Service
10:30 am -11:00 am	COFFEE BREAK
11:00 am - 12:30 pm	FIRST CONCURRENT WORKSHOPS Models that Work and Lesson Learned
Moderators :	Academia Room 146 Panel A - Achieving Educational and Occupational Goals
	Dr. Edward Thomas, Auburn University Ms. Nikola Garber, National Sea Grant College Program, NOAA
Panelists:	 Dr. William Bonner, University Center for Atmospheric Research Dr. Bradford Brown, NMFS/NOAA Dr. Gleyn Bledsoe, Land Grant and Outreach, Northwest Indian College Dr. Marin Robinson, Northern Arizona University Ms. Letise T. Houser, Brown University Dr. Mark Hardy, Jackson State University
	Private Sector Room 262
Moderators:	Panel A - Building Mutually Beneficial Partnerships Ms. Pamela Neal, Sarkeys Energy Center, Oklahoma University Mr. Robert Stockman, Office of Finance & Administration, NOAA
Panelists:	 Mr. Robert Stockman, Once of Pinance & Administration, NOAA Ms. Jennifer Carfagno, Weather Channel Ms. Cathy Fore, Oak Ridge MEITP, Department of Energy Ms. DeLois Cutter, Tal-Cut Inc. Dr. Edward Johnson, NWS/NOAA Mr. Edwin Mitchell, J-Tek, Inc. Ms. DeJonnette Grantham-King, Advance Environmental Consultants, Inc.
Moderators :	Public SectorRoom 263Panel A - Increasing Access to Public ResourcesMr. Benjamin Watkins, NESDIS/NOAAMs. Rita Presley, Jackson State University

AGENDA - 3

Panelists:	Dr. Jewell Prendeville, National Science Foundation Dr. Robert Menzer, National Center for Environmental Resear Ms. Jacqueline Rousseau, USEC/NOAA Mr. James Harrington, NASA Mr. Anthony Overton, University of Maryland Eastern Shore Ms. Jan Kucklick, NOS/NOAA Dr. Teferi Tsegaye, Alabama A&M University Ms. Julie Marcy, U.S. Army Corps of Engineers	ch, EPA
	Student	Room 266
	Panel A - Creating Paths to Success in Marine and	
Moderators:	Atmospheric Sciences Mr. Jose Garcia, NWS/NOAA	
moderators.	Dr. Dionne Hoskins, Savannah State University	
Panelists:	Dr. Michael Fogarty, NMFS/NOAA	
	Mr. Ricardo Lopez, California State University Dr. Pace Wilber, NOS/NOAA	
	Ms. Andrea Sealy, Pennsylvania State University	
	Dr. Randal Mandock, Clark Atlanta University	
	Mr. M. Brandon Jones, Delaware University	
12:30 pm - 2:00 pm	LUNCH	
	Jacob L. Reddix Campus Union, General Purpose Room	
Moderator:	Dr. Roy DeBerry, Vice President for External Affairs,	
	Jackson State University	
Keynote :	Ms. DeLois A. Cutter, President, Tal-Cut Company	
2:00 pm	Opening of Business, Technology, and Career Trade Fair (Remains open through Tuesday)	
	Lee E. Williams Athletics & Assembly Center	
0.00		D
2:30 pm - 4:00 pm	SECOND CONCURRENT WORKSHOPS Models that Work and Lessons Learned	Room 146
	Academia	
	Panel B - Investing in Building Capacity	
Moderators:	Dr. Larry Robinson, Florida A&M Ms. Margaret McBride, Chesapeake Bay Program, NOAA	
Panelists:	Dr. Joseph Okoh, University of Maryland Eastern Shore	
	Dr. Abdul Mohamed, Jackson State University	
	Dr. Michael Sissenwine, NMFS/NOAA, Northeast Fisheries ScienceCenter	
	Mr. Kurt Shinkle, National Geodetic Survey/NOAA	
	Dr. Arthur Allen, University of Maryland Eastern Shore	
	Dr. Robert Stewart, Texas A&M	

Moderators: Panelists:	Private Sector Panel B - Converging Business and Academic Practices Dr. Mildred Ofosu, Delaware State University Dr. Fred Thurberg, NMFS/NOAA Dr. Barry Costa-Pierce, Mississippi/Alabama Sea Grant Mr. Clinton Twilley, Concurrent Technologies Corp. Mr. Eddie Hanebut, Digital Quest, Inc. Mr. George B. Brooks, Jr., University of Arizona Dr. Richard Gragg, Florida A&M Mr. Kelton Clark, Morgan State University	Room 262
Moderators: Panelists:	Public Sector Panel B - Sustaining Linkages for Social and Economic Development Dr. James Arrington, South Carolina State University Mr. William Parker, NWS/NOAA Mr. Ken Davidson, NESDIS/NOAA Ms. Michele McCoy, EASC/NOAA Dr. Barbara Ousby, Mississippi Department of Economic and Community Development Dr. Mack Felton, Dillard University Dr. Darryll Keith, EPA Mr. Jamison Hawkins, NWS/NOAA	Room 263
	Student Panel B - Becoming a Professional in Marine and Atmospheric Sciences	Room 266
Moderators:	Dr. Brian Bingham, Western Washington University Mr. Alonzo Hamilton, NMFS/NOAA	
Panelists:	Mr. Alon 20 Hamilton, Hum On VOAA Ms. Pamela Lestenkoff, University of Alaska Ms. Noelani Puniwai, Washington State University Ms. Shawn Arellano, Woods Hole Oceanographic Institution Mr. Ashton Robinson-Cook, Jackson State University Ms. Wanda Jones, University of Florida-Gainesville Mr. Anthony Overton, University of Maryland Eastern Shore Ms. Essie Coleman-Duffie, Southeast Fisheries Science Cent Mr. Lawrence C. Tynes, Sr., Eastern Administrative Support Ce	
4:15 pm- 5:30 pm	POSTER PRESENTATIONS Student, Faculty, Public, and Private Sector Posters (Will remain on exhibit until 1:00 pm on Tuesday) Lee Williams Athletics & Assembly Center	
Conveners of s	tudent participants: Dr. Paulinus Chigbu, Jackson State University	
Convener of NC	Dr. Paul Croft, Jackson State University DAA participants: Dr. Sheila Stiles, NMFS/NOAA	
6:00 pm - 8:00 pm	CONFERENCE BANQUET Jacob L. Reddix Campus Union, General Purpose Room	

AGENDA - 5

Moderator:	Dr. Willie Brown, Vice President for Technology,
	Jackson State University
Introducer:	Dr. Joyce Payne, Director, Office for the Advancement
	of Public Black Colleges, NASULGC
Keynote :	Dr. Ronald Mason, Jr., President
	Jackson State University

Tuesday April 3, 2001 JSU Liberal Arts Building

7:45 am - 8:30 am	Registration and Continental Breakfast	Lobby
8:30 am - 10:00 am	PLENARY SESSION Untapped Resources and Opportunities for Building Sustainable Alliances (overflow	Room 266 Room 146)
Moderator:	Dr. Livingston Marshall, Associate Professor Morgan State University	
Keynote:	Dr. Margaret Leinen, Assistant Director for GeoSciences, National Science Foundation	
Panelists:	 Ms. Cathy Fore, Manager MEITP, Department of Energy Mr. Justin Ahanhanzo, Consultant, UNESCO/IOC Dr. Russell Schneider, Chief Science Support, Storm Prediction Center, NOAA Mr. Eddie Hanebut, President, Digital Quest, Inc. Dr. Robert Shepard, Science and Engineering Alliance Ms. Senayt Asseta, Program Coordinator, American Association for the Advancement of Science Mr. Roy Pemberton, Graduate Student, Virginia Institute of Marine Science 	
10:00 am - 10:30 am	COFFEE BREAK	
10:30 am - 12:00 pm	WORKING GROUPS Looking Ahead and Innovations	
Facilitators:	National Marine Fisheries Service Dr. Bradford Brown, NOAA Dr. Matthew Gilligan, Savannah State University	Room 266
Facilitators:	Office of Oceanic and Atmospheric Research Ms. Ann Georgilas, NOAA Dr. Livingston Marshall, Morgan State University	Room 262
Facilitators:	National Weather Service Mr. Jose Garcia, NOAA Dr. Paul Croft, Jackson State University	Room 263

Facilitators:	National Ocean Service Ms. Jocelyn Martin, NOAA Dr. George Burbanck, Hampton University	Room 146
Facilitators:	National Environmental Satellite, Data, and Information Service Mr. Benjamin Watkins, NOAA Dr. Abdul Mohamed, Jackson State University	Room 254
Facilitators:	Office of Finance and Administration Dr. Robert Stockman, NOAA Dr. Mark Hardy, Jackson State University	Room 264
12:00 pm - 1:30 pm	LUNCH Jacob L. Reddix Campus Union, General Purpose Ro	om
Moderator: Keynote:	Dr. Felix Okojie, Vice President for Research and Stra Initiatives, Jackson State University Dr. Darrell Jay Grimes, Dean, Institute of Marine Sciences The University of Southern Mississippi Best Posters Awards Special Recognitions	tegic
2:00 pm - 4:00 pm	PLENARY SESSION	Room 266
Moderators :	Closing Ms. Carmella Davis Watkins, National Environmental Satellite, Data, and Information Service, NOAA Dr. Emorcia Hill, Abt Associates, Inc.	verflow Room 146)
	Concluding Remarks: Dr. Ambrose Jearld, NMFS/NOAA, Conference Chair	
4:00 pm	ТЕА	To be announced

MONDAY April 2, 2001

9:15 am - 10:30am Room 266

Perspectives from NOAA's Leadership - Progress and Impediments: Building Sustainable Alliances

Moderators:	Dr. Denise Stephenson-Hawk
	Dr. William White

Panelists:Mr. John OliverMs. Mary GlackinMs. Louisa KochDr. Edward JohnsonCaptain Ted Lillestolen

- What is the representation of minorities in oceanic/marine, environmental, and atmospheric (OME&A) fields in academia (students and faculty) and the workforce?
- What has been the trend over the last five years in the production of minority students, faculty, and professionals in these fields?
- What joint strategies should NOAA, academia, and the private sector pursue to "dramatically" increase minority access to employment and educational opportunities in the OME&A fields?
- Which of your organization's policies and practices have proven successful or unsuccessful in the production of minorities in these fields?
- How does your organization measure its success in increasing minority participation in these professions?
- What types of cutting edge technologies are being pursued by your agency which offer opportunities for Minority Serving Institutions (MSIs) to partner with the federal sector?
- How does working with MSIs impact your scientific programs, financially, technically, and socially?
- If you left this agency tomorrow, what would you want your legacy to be with respect to MSIs?

FIRST CONCURRENT WORKSHOPS Models that Work and Lesson Learned

11:00 am - 12:30pm Room 146

Academia Panel A – Achieving Educational and Occupational Goals

Moderators:	Dr. Edward Thomas Ms. Nikola Garber	
Panelists:	Dr. William Bonner	Dr. Marin Robinson
	Dr. Bradford Brown	Ms. Letise T. Houser
	Dr. Gleyn Bledsoe	Dr. Mark Hardy

Questions to be Addressed During this Session:

- Where do the oceanic/marine, environmental, and atmospheric (OME&A) fields fit in your institution's overall mission?
- What are the major components of your institution's long range plan and how does this plan support the OME&A fields?
- What are some of your institution's current model programs (academic, research, occupational) in these fields? Which do you consider the most successful?
- How adequately does your institution's infrastructure support activities/programs in these fields? What improvements, if any, are needed?
- How successful has your institution been in advancing students to graduate studies, internally and externally? What occupations have your graduates pursued in the OME&A fields?

Room 262

Private Sector Panel A - Building Mutually Beneficial Partnerships

Moderators:	Ms. Pamela Neal Mr. Robert Stockman	
Panelists:	Ms. Jennifer Carfagno	Dr. Edward Johnson
	Ms. Cathy Fore	Mr. Edwin Mitchell.
	Ms. DeLois Cutter	Ms. DeJonnette Grantham-King

Questions to be Addressed During this Session:

- What are some models of your successful partnerships? What are the key elements that drive its success? What are some of the pros and cons of partnerships?
- What are the varieties of partnership arrangements? How do they differ? Does each require different strategies to sustain them?
- What are some of the ways to add value to partnerships to ensure its success, e.g. technology, equipment, and facilities?
- How do you assess your organization's readiness to create partnerships?
- What factors are essential to managing and maintaining effective, productive partnerships?
- How can effective, productive partnerships be created in the oceanic/marine, environmental, and atmospheric (OME&A) fields? What roles should academia, public, and private sectors play in building these partnerships?

Room 263

Public Sector Panel A - Increasing Access to Public Resources

Moderators :	Mr. Benjamin Watkins Ms. Rita Presley	
Panelists:	Dr. Jewell Prendeville	Ms. Jan Kucklick
	Dr. Robert Menzer	Dr. Teferi Tsegaye
	Ms. Jacqueline Rousseau	Ms. Julie Marcy
	Mr. James Harrington	Mr. Anthony Overto

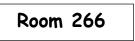
Questions to be Addressed During this Session:

• What are your agency's major initiatives, in the oceanic/marine, environmental, and atmospheric (OME&A) fields for which academic institutions can apply? And other relevant initiatives?

Mr. Anthony Overton

- Is there targeted support for, e.g., HBCUs or other MSIs? Which of your initiatives explicitly encourage collaboration with these institutions?
- Has your agency participated in research programs that bring researchers from NOAA laboratories and faculty/students from HBCUs/MSIs together (e.g., summer or semester research opportunities)?

- How have these types of programs worked? What criteria are needed to select faculty to participate in these programs?
- What does it take to be fully competitive for extramural funding? How does an academic institution remain competitive over the long haul?
- Where do you get your funding? How stable is the funding? How is your agency's funding allocated?
- Where and what are the primary mechanisms through which your agency allocates funding to academic institutions?
- What are your new programs? And, what programs are on the horizon? How does your agency develop request for proposals (RFP)?



Student Panel A - Creating Paths to Success in Marine and Atmospheric Sciences

Moderators:	Mr. Jose Garcia Dr. Dionne Hoskins
Panelists:	Dr. Michael Fogarty Mr. Ricardo Lopez Dr. Pace Wilber

Ms. Andrea Sealy Dr. Randal Mandock Mr. M. Brandon Jones

- What are the various transitional points in one's academic career (high school to college; college to graduate school; graduate school to workforce)?
- What skills are vital to success at each transition? How do I acquire these skills?
- How do I assess my skills and competencies along the way? How do you develop life_long learning skills?
- What opportunities are available for scholarships and training in marine and atmospheric sciences?
- What are examples of successful (and unsuccessful) strategies in advancing one's academic career?
- What are some of the new emerging areas in these fields? How do I prepare to take full advantage of them?

SECOND CONCURRENT WORKSHOPS Models that Work and Lessons Learned

2:30 pm - 4:00 pm Room 146

Academia Panel B – Investing in Building Capacity

Moderators:	Dr. Larry Robinson,
	Ms. Margaret McBride

Panelists:	Dr. Joseph Okoh	Mr. Kurt Shinkle
	Dr. Abdul Mohamed	Dr. Arthur Allen
	Dr. Michael Sissenwine	Dr. Robert Stewart

- What types of investments has your institution/organization made in building capacity in the oceanic/marine, environmental, and atmospheric (OME&A) fields?
- How would you measure your success? What risks have been involved? What are your major extramural funding sources?
- How do you assess your institution's/organization's current capacity for growth in marine and atmospheric fields? How do you identify the best strategic investment for your institution?
- How does an institution/organization leverage long-term support for capacity building? And, how do you diversify your extramural funding?
- What types and levels of institutional supports are needed to sustain a viable research program in OME&A fields?
- What strategies should universities (majority, HBCUs, and MSIs) use to attract faculty and researchers from traditionally under_represented groups and providing them with the physical, intellectual, and financial infrastructure to grow a successful program in OME&A fields?

Room 262

Private Sector Panel B - Converging Business and Academic Practices

Moderators:	Dr. Mildred Ofosu Dr. Fred Thurberg	
Panelists:	Dr. Barry Costa-Pierce Mr. Clinton Twilley	Mr. George B. Brooks, Jr. Dr. Richard Gragg
	Mr. Eddie Hanebut	Mr. Kelton Clark

Questions to be Addressed During this Session:

- What are business' core values and principles? What are academia's core values and principles?
- How similar or dissimilar are the vocabularies and behaviors (practices) of these two enterprises?
- What are some successful models joint ventures between business and academia? What are the key elements that drive its success? What are some of the pros and cons of these ventures?
- What are some of the short- and long-term research activities within academia that may offer business opportunities and vice versa?
- What relative roles should business, academia, and research laboratories play in determining future areas of research in the oceanic/marine, environmental, and atmospheric (OME&A) fields?

Room 263

Public Sector

Panel B - Sustaining Linkages for Social and Economic Development

Moderators:	Dr. James Arrington Mr. William Parker	
Panelists:	Mr. Ken Davidson Ms. Michele McCoy	Dr. Mack Felton Dr. Darryll Keith
	Dr. Barbara Ousby	Mr. Jamison Hawkins

- What opportunities do government agencies (local, state, and federal) provide for academic institutions to become involved in local economic activity and development? What is the value added by the institutions?
- How do you determine if there is a match between academia and the needs of the local economy?
- How are feasibility studies conducted to identify these matches?
- Are there currently opportunities within your agency for collaborations with HBCUs and MSIs and what is the magnitude of these initiatives?
- What characteristics must an institution possess to be competitive for funding at the local, state, and federal levels?
- How do the three levels of government interact to identify and allocate resources to stimulate socio-economic development in a particular geographic area?
- What current and future activities (if any) do the OME&A researchers at your agency participate in that disseminate knowledge to the local and state communities?
- What research areas can be pursued in academia and at research laboratories (e.g., ecological management, air and water quality management, etc.) that can impact the communities served by HBCUs and MSIs?

Room 266

Student

Panel B - Becoming a Professional in Marine and Atmospheric Sciences

- Moderators: Dr. Brian Bingham Mr. Alonzo Hamilton Panelists: Ms. Pamela Lestenkoff Mr.
 - Ms. Pamela LestenkoffMr. Ashton Robinson-CookMs. Noelani PuniwaiMs. Wanda JonesMs. Shawn ArellanoMr. Anthony OvertonMs. Essie Coleman-DuffieMr. Lawrence C. Tynes, Sr.

- What types of jobs are available in the marine and atmospheric fields? What career options are available with: BS, MS, Ph.D. degrees?
- How do you position yourself for professional success? What are the measures of professional success?

- How do you prepare yourself to be competitive for professional positions in different settings: academia (teaching and research); industry; research laboratory?
- What are some of the new emerging research areas in these fields? What types of skills and competencies do they require?
- What challenges are you likely to face in raising money and writing proposals to pursue your professional activities?
- What is the role of professional organizations in success in these fields?

8:30 am - 10:00 am Room 226 (overflow Room 146)

PLENARY SESSION Untapped Resources and Opportunities for Building Sustainable Alliances

Moderator: Dr. Livingston Marshall

Keynote: Dr. Margaret Leinen

Panelists:Ms. Cathy ForeDr. Robert ShepardMr. Justin AhanhanzoMs. Senayt AssetaDr. Russell SchneiderMr. Roy PembertonMr. Eddie HanebuMr. Roy Pemberton

- What are some key policy/programmatic initiatives currently in effect (or planned) which HBCUs and/or MSIs could benefit from?
- What are some of the creative ways in which organizations/institutions could modify their current policies and practices to provide opportunities for HBCUs and/or MSIs?
- What mechanisms are in place to disseminate information about these new initiatives/ resources? Specifically, how can HBCUs and/or MSIs find out, apply, and receive support from these sources?
- How can HBCUs and/or MSIs posture and promote their institution so that they attract these resources? What incentives does HBCUs and/or MSIs have to offer to gain a competitive advantage in these new funding arenas?

10:30 am - 12:00 pm

WORKING GROUPS Looking Ahead and Innovations

Room 266	
	National Marine Fisheries Service
Facilitators :	Dr. Bradford Brown, NOAA
	Dr. Matthew Gilligan, Savannah State University
Room 262	
	Office of Oceanic and Atmospheric Research
Facilitators:	Ms. Ann Georgilas, NOAA
	Dr. Livingston Marshall, Morgan State University
Room 263	
_	National Weather Service
Facilitators:	Mr. Jose Garcia, NOAA
	Dr. Paul Croft, Jackson State University
Room 146	
	National Ocean Service
Facilitators:	Ms. Jocelyn Martin, NOAA
	Dr. George Burbanck, Hampton University
Room 254	
	National Environmental Satellite, Data,
	and Information Service
Facilitators:	Mr. Benjamin Watkins, NOAA
	Dr. Abdul Mohamed, Jackson State University
Room 264	
-	Office of Finance and Administration
Facilitators:	Dr. Robert Stockman, NOAA
	Dr. Mark Hardy, Jackson State University

- What were some of the key issues related to your field/area discussed during this conference? How would you prioritize these?
- What are the realistic strategies that could be implemented in the short-, medium-, and long-term?
- What types of investments and commitments are needed for these to be successfully implemented?
- What is the ultimate/ideal goal in adopting these strategies? How will these goals be measured, and progress monitored?
- How should responsibilities by (re)distributed among academia, public, and private sectors to maximize the likelihood of accomplishing these objectives?

STUDENT ABSTRACTS FOR POSTER PRESENTATIONS

OCEANIC/MARINE AND FISHERIES SCIENCES

(1)

OCCURRENCE OF SOME FISHES IN VARIOUS HAB-ITATS IN THE VICINITY OF MISSISSIPPI SOUND Sonya D. Barner*, Sara E. LeCroy and Chet F. Rakocinski

University of Southern Mississippi, Institute of Marine Science Gulf Coast Research Laboratory, 703 East Beach Drive, Ocean Springs, MS 39566 ; *Department of Biology, Jackson State University, Jackson, MS 39217

To generate museum records and document biodiversity in Mississippi coastal waters, fishes were collected from various habitats in Mississippi Sound and nearby Gulf of Mexico waters, ranging from Fort Bayou inshore waters to open waters near Horn and Chandaleur Islands. Samples were collected with seines and an otter trawl. Fishes were sorted, identified, catalogued and deposited in the fish museum at Gulf Coast Research Laboratory (GCRL). The GCRL Museum database was searched to provide additional information on the five most abundant species from each habitat in order to compare fish habitat preferences for the most common species collected. Forty species of fish were identified. The following families had the highest number of species: Carangidae (5), Fundulidae (5), Sciaenidae (5), Engraulidae (3) and Sparidae (3), followed by the Clupeidae (2), Gobiidae (2), Mugilidae (2), Synodontidae (2) and Syngnathidae (2). Of the seven species collected at the Fort Bayou station, Menidia beryllina, Anchoa mitchilli, and Lucania parva were the most abundant accounting for about 76% of the total catch. In the offshore areas of Barrier islands, eleven species were collected with Anchoa hepsetus, A. mitchilli and Saurida brasiliensis being the most abundant. Two different distribution patterns were evident based on our collections. Pelagic forage species such as Anchoa mitchilli and Menidia beryllina were widespread, showed no apparent habitat association and were euryhaline, occurring widely throughout the area. The second pattern was shown by species that were associated with one or two habitat types. These included Sauridia brasiliensis, Eutremeus teres, and Stenotomus caprinus, which were collected only at the open water stations, Gobiosoma bosc, from low salinity mud bottoms, Trachinotus carolinus and Menticirrhus americanus from nearshore sand bottoms, and all three species of Fundulus, Oligoplites saurus and Eucinostomus argenteus from high salinity grassbeds.

(2)

Prediction of Shrimp Biomass Fluctuations and Catch Rates Using a Biomass Dynamic Model: A Preliminary Study

Atanasio Brito¹, David Die^{2,4}, Richard Gragg¹, Elijah Johnson¹, Jennifer Cherrier¹, Larry Robinson¹ and Brad Brown³

¹Environmental Sciences Institute, Florida A&M University, Tallahassee Florida and the ²Center for Unified Fisheries Education and Research, ³NOAA Southeast Fisheries Center and the ⁴University of Miami, Rosenstiel School of Marine and Atmospheric Sciences, Miami, Florida

The Mozambigue Sofala Bank (16 20'-21 00') shrimp fishery was over fished and the fishing effort was 30% greater than the effort that would produce the maximum sustainable yield for the year 2000. Recent introductions of powerful shrimping vessels, night time fishing and the constant upgrading of navigation systems such as Global Positioning System and Echo Sounder by commercial fleets have counteracted the management practices of seasonal closures, total allowable catches and limited entries. This work uses a biomass dynamic model to predict monthly biomass fluctuations of penaeid shrimp in Sofala Bank. The model can give managers a predictive capability in estimating monthly biomass flutuactions for the purpose of maintaining a sustainable shrimp fishery. A 9-year series (1991 to 1999) of a commercial fleet shrimping catch and effort statistics, for Farfantepenaeus indicus and Metapenaeus monoceros, were input into the biomass dynamic model. A reasonably good correlation between the estimated catch per unit of effort and the observed monthly catch per unit of effort for both species was observed in the model. The estimated catch per unit of effort, for F. indicus, following the all year recruitment pattern yielded an r-value of 0.7363. The harvestable biomass, from 1991-1999, of F. indicus, oscillated on a monthly basis between 34 and 415 tons. The catchability coefficient for the fleet is estimated to be 0.000085. While the annual growth biomass oscillated between 387 and 1083 tons. The estimated catch per unit of effort, for *M. monoceros*, fits the observed at an r-value equal to 0.5379 for a seven-month recruitment pattern. The model predicts that the harvestable biomass of M. monoceros oscillated between 83 and 783 tons. The catchability coefficient for this species was estimated to be 0.000026. The annual growth biomass oscillated between 593 and 1262 tons. This preliminary work has provided the key elements necessary for a definitive study. Future work will be more comprehensive, using data from all Sofala Bank shrimping fleets and the six shrimp species caught in the past 15 years.

(3)

FISHERY TROPHIC LINKAGES IN THE CHESAPEAKE BAY ECOSYSTEM

Graves, R.H., Morgan State University and USGS Patuxent Wildlife Research Center, Baltimore and Laurel, United States, Marshall, L. S., School of Computer, Mathematics, and Natural Sciences, Baltimore, MD, Keough, J. R., USGS Patuxent Wildlife Research Center, Laurel, MD.

Annually, landings from the Chesapeake Bay ecosystem account for roughly 250,000 - 350,000 metric tonnes of total catch from U.S. fisheries. Collectively, over 50 species of finfish, crustaceans, and bivalves are caught in the Bay. The complexity of the bay ecosystem, along with effects that stem from the steadily increasing human population of its watershed, challenges fishery management. Modern fishery management calls for multi-species models and approaches. Unlike single-species approaches, multi-species approaches treat stocks as part of an ecosystem, taking into account natural mortality, growth effects, and dynamics of the entire food web. In order to develop these approaches, a good understanding of trophic linkages under varying conditions is needed. This study serves to identify trophic linkages through the use of a combination of stomach content and C, N, and S stable isotope analysis. Samples represent important finfish, bivalve, and crab populations in oligohaline and mesohaline regions of the Bay. We seek to identify the trophic roles and connections of and among species and age classes spatially, temporally, and under varying water quality conditions.

(4)

A PRELIMINARY STUDY OF SUBSTRATE PREFER-ENCE BY HATCHERY PRODUCED JUVENILE RED SNAPPER (Lutjanus campechanus)

Jomeka Johnson*, Chigbu, P.* & Ogle, J.**

*Department of Biology, Jackson State University, Jackson, MS 39217; **Institute of Marine Sciences, Gulf Coast Research Laboratory, University of Southern Mississippi, Ocean Springs, MS 39566.

Substrate characteristics influence the distribution of benthic organisms, including fishes. Red snapper larvae settle out to the bottom of the Gulf of Mexico following a planktonic larval existence. A previous laboratory study using circular tanks suggested that 0+ red snapper collected from the wild preferred shell to sand substrate, perhaps because shells provide refuge to juvenile snapper from predation. However, hatchery-produced fish may differ from wild conspecifics in their behavior, for example, in habitat selection. This can increase the vulnerability of hatchery-produced fish or wild conspecifics to predation, and thus can potentially contribute to the failure of fish stock enhancement projects. We examined substrate preference by hatchery-produced 0+ red snapper using two raceways and two circular tanks. Each container was divided into two equal areas, the floor was covered with sand in one area and

clam shells in the other half. Groups of four (n=6) fish < 92.0 mm in size were introduced into the experimental chambers and visually observed at various times from morning to night. On the average, more fish (75%) were observed in the areas of the tanks with sandy substrate than in the areas with shell (25%). Only in one case was there no apparent difference in substrate preference, as about 50% fish were on sand and 50% on shell. These results are inconsistent with a published study that suggested the wild caught juvenile red snapper preferred shell to sandy substrate. Further studies are planned to confirm this observation, to compare substrate selection by hatchery-produced and wild caught snapper, and to assess ontogenetic changes in substrate preference.

(5)

MARINE MAMMAL PROGRAMS AND ACTIVITIES AT THE SOUTHEAST FISHERIES SCIENCE CENTER, PANAMA CITY LABORATORY, FLORIDA

Wanda Jones, University of Florida and National Marine Fisheries Service, Southeast Fisheries Science Center, Panama City Laboratory

The National Oceanic and Atmospheric Administration (NOAA) Fisheries, is mandated by the Marine Mammal Protection Act of 1972 and Amendments of 1994 to manage and conserve certain marine mammal species including all cetaceans, seals and sea lions. In the southeast region, the Southeast Fisheries Science Center (SEFSC) Protected Resources and Biodiversity Branch (PRBD) houses the Marine Mammal Investigation Group that participates in the Marine Mammal Stock Assessment and Marine Mammal Health and Stranding Response Program(s). In compliance with the act mandates, stranding programs, biomonitoring programs, and population surveys have been developed and implemented in the southeast region. The SEFSC programs oversee NMFS research conducted on marine mammal communities in three ecosystems: the Gulf of Mexico Shelf, the South Atlantic Shelf, and the US Caribbean Shelf. A number of cetacean (whale and dolphin) species, pinniped (seals) species, and sirenian (manatees) species inhabit these areas seasonally and year-round. Research conducted by the SEFSC is utilized by NMFS' Office of Protected Resources and Conservation and Recovery Programs in the management and enforcement branches to create, implement, mitigate, and enforce regulations supporting the Marine Mammal Protection Act of 1972.

Several SEFSC field laboratories within the southeast region conduct research and activities in keeping with the goals and mission of the Marine Mammal Programs. The Panama City Laboratory, Panama City Beach, Florida, in cooperation with the University of Florida, conducts marine mammal programs under the PRBD including stock assessment, mortality estimation, health assessment, life history, and behavioral research. Recently, from 1997-2000, Jones (in prep) conducted field research on bottlenose dolphins in Panama City and the surrounding areas. The work focused on stock assessment and the general behavioral ecology of pods and individual dolphins. Information collected included abundance estimates, residency status, habitat use, movement patterns, group associations, and the development of activity budgets for the various pods in northwest Florida marine and estuarine waters. In addition, a photoidentification catalog was created and continues to be updated for dolphins inhabiting the waters surrounding Panama City and neighboring cities, Destin and Port St. Joe. This catalog is a valuable asset in the identification of resident and transient dolphins, for behavioral studies, and for stock assessment goals. It is also useful in determining identification of individuals in unusual mortality events and those examined during the normal course of data collection for the Marine Mammal Health and Stranding Response Program (MMHSRP). Research and activities conducted for the MMHSRP include collaboration with the Southeast Region Stranding Network for marine mammal rescue, rehabilitation, health assessment, and necropsy for samplecollection for the Biomonitoring Program including life history, genetic, morphometric, pathology, virology, and feeding ecology research.

(6)

ABSENCE OF *MONOPYLEPHORUS RUBRONIVEUS* IN FISH FEEDING STUDIES.

Tiffany Lee, Savannah State University, Savannah, GA; David Gillett, University of Charleston, Charleston, SC.

Tidal creeks serve as nursery and refuge habitats for many economically and ecologically important species of fish and crustaceans. During at least one stage of their lives, most of these fish and crustaceans feed upon the benthos living in the creeks. The tubificid oligochaete Monopylephorus rubroniveus is the numerically dominant macrobenthic organism found intertidally in southeastern tidal creeks. Oddly enough, Monopylephorus and similar organisms are rarely found during fish gut-content studies. We have performed a series of experiments in which we fed both oligochaetes and hard-bodied amphipods to a common estuarine fish, Fundulus heteroclitus and measured the state of digestion of each after 2, 15, 30, or 60 minutes in the gut. To evaluate the presence of food, items in the gut were assigned a score, 0- empty, 1-tissue (identified under a compound microscope) and 2- whole, based on the degree of digestion. Amphipods were easily identified to species at all four sampling times. After 2 minutes the oligochaete was still easily identified. At 15, 30, and 60 minutes the oligochaete was only recognizable as setae, if there were any remains left at all. It is our feeling that most fish diets, which are based upon typical gutcontent analysis methods, underestimate the importance of oligochaetes and other soft-bodied organisms while overestimating the importance of hard-

bodied organisms due to their quick digestion time in the gut of the fish.

(7)

ArcView GIS Utilization in Identification and Quantification of Shore Type and Land Use in an Urbanized Subestuary of the Chesapeake Bay

Patrice Longshaw Marine and Environmental Science Undergraduate, Hampton University

The technological advancements that have been made in mapping during the last few years allow for many statistical and database functions to be integrated into one program. One such program is the ArcView GIS program, distributed by Environmental SR Imaging. The Hampton River, a tributary of the James River adjacent to Hampton Roads, at the mouth of the Chesapeake Bay in Virginia, is an ideal location to utilize the capabilities of ArcView GIS to analyze land use and shoreline types. ArcView allows overlaying of digitized maps from various sources such as the City of Hampton. and USGS. Each of these layers, once applied to a basemap, can be shown, or hidden, as appropriate for the particular area being analyzed. This project is in progress, and will provide many opportunities for student research in the future. This will be an ongoing study to better quantify and assess effects of shore types and land usage on the Hampton River ecosystem.

(8)

INVESTIGATIONS OF THE HEALTH OF THE STRIPED BASS IN THE CHESAPEAKE BAY: MYCOBACTERIA INFECTIONS IN STRIPED BASS.

Anthony S. Overton¹, Eric B. May¹, and F.J. Margraf² University of Maryland Eastern Shore, Maryland Cooperative Fish and Wildlife Research Unit 1120 Trigg Hall Princess Anne, Maryland 21853. 2Alaska Cooperative Fish and Wildlife Research Unit 210 Irving I Building P.O. Box 757020 University of Alaska-Fairbanks, AK

Several species of bacteria have been isolated from striped bass in the Chesapeake Bay. Mycobacteria sp. is of particular interest to scientists because if its association with large nodules (granulomas) or ulcers throughout many of their tissues. The resulting disease may be characterized by emaciation, inflammation of the skin, open lesions, and ulceration. We determined the extent of Mycobacteria infection in striped bass and provide evidence of decreased overall condition of fish affected by these bacteria. Almost 50% of the fish showed some sign of an external sore. Granulomas appeared in at least one organ in 53% of the fish in our samples regardless of the presence or absence of external sores. The Gran scores from the spleen and heady kidney were significantly higher than either the heart and liver and appeared to be most closely associated with the presence of external The condition factor was significantly higher for striped bass without sores (0.96) than those with sores (0.82). Condition factors involving all fish clearly shows a trend, with NGNS at 0.99, GNS at 0.92 and GS at 0.81. Slopes of the regression loge weight (g) and loge length (mm) for striped bass with sores was significantly higher than those fish without sores. However

the weight at length were more variable (lower r2) in striped bass with sores than those fish without sores.

(9)

THE HABITAT UTILIZATION OF JUVENILE HAWKS-BILL SEA TURTLES (ERETMOCHELYS IMBRICATA) AT BUCK ISLAND REEF NATIONAL MONUMENT ST. CROIX U.S.VIRGIN ISLANDS

Roy A. Pemberton Jr. and John A. Musick, College of William & Mary, Virginia Institute of Marine Science, Fisheries Science Lab., Gloucester Pt, VA 23062

Juvenile hawksbill, Eretmochelys imbricata, and green, Chelonia mydas, sea turtles are found in waters of Buck Island Reef National Monument, St. Croix U.S. Virgin Islands. Sea turtle surveys were conducted at Buck Island in the summer of 1998 and winter of 1999. Green turtles were found in the southern area of the monument in blocks A through J and block R. These blocks were either adjacent to or near areas of large sea grass beds, which are the predominant foraging areas for green sea turtles. The majority of juvenile hawksbills sighted were along the north side of the monument in blocks J through N. There was a significant difference in the mean number of juvenile hawksbills sighted in blocks with low zoanthid (Zoanthus sociatus) cover and those with high zoanthid cover. More juvenile hawksbills were sighted in blocks with high zoanthid cover. Their habitat utilization around Buck Island reflects their foraging focus on zoanthids.

Radio and sonic telemetry was used to elucidate site philopatry of juvenile hawksbills at Buck Island. The hawksbills exhibited strong site philopatry for the North side of Buck Island. Each turtle monitored had a relatively small home range (2 km²) which it occupied for at least several months. Turtles moved less than 1.09 km over a period of several months. Their ranges appeared to not be limited by the park boundaries but possibly by the distribution of zoanthids in the habitat. Judging from tag recapture studies these animals may be resident in the habitat for periods of at least several years. Strong site philopatry was maintained by one juvenile hawksbill despite the passage of Hurricane Lenny in November of 1999.

(10)

THE ECOLOGICAL IMPORTANCE OF CRINOIDS ON HECETA BANK, OREGON

Puniwai, Noelani and Tissot, Brian N.

Washington State University at Vancouver, 14204 NE Salmon Creek Ave. Vancouver, WA 98686.

Heceta Bank is a commercially important offshore rocky bank with depths of 80-400m. Surveys of Heceta Bank were completed yearly between 1988 and 1990 using the submersible Delta. Transects provided data on physical habitat, and fish and invertebrate diversity and abundance. These transects were revisited this past summer and will be visited again next year as part of a 2 year study. The use of the submersible Delta, as well as the remote operating vehicle ROPOS provided the opportunity to sample a greater variety of habitats on the bank.

The crinoid *Florometra serratissima* is a common invertebrate associated with hard substrate throughout the bank. The goal of this study is to compare the abundance and distribution of past studies to current studies 10 years later. The patterns that emerge from this study can be used to understand the importance of crinoids to the deep-sea benthic community of the Northeast Pacific Ocean and the role these invertebrates, and other benthic organisms, may play in the life cycle of economically important groundfish.

(11)

EFFECTS OF SALINITY ON FOOD CONVERSION AND GROWTH OF JUVENILE RED SNAPPER (Lutjanus campechanus)

*Pamela Riley-Daniels, *Himabindu Remata, Chigbu, P.*, Ogle, J**., & Lotz, J.**

*Department of Biology, Jackson State University, Jackson, MS 39217; **Institute of Marine Sciences, Gulf Coast Research Laboratory, University of Southern Mississippi, Ocean Springs, MS 39566.

Salinity is an important factor affecting growth and distribution of marine organisms. In a recent study, "high value habitat" for juvenile red snapper was defined in the northwestern Gulf of Mexico, as areas with temperature of $24 - 26^{\circ}$ C, dissolved oxygen level > 5 mg/L and salinity of about 35%. Nevertheless, limited information is available on the response of red snapper to different levels of physico-chemical parameters. Defining salinity level for optimal growth of juvenile snapper is invaluable for its culture. It is also useful for predicting the response of snapper to changes in salinity in its natural environment. Juvenile red snapper produced at the University of Southern Mississippi-

Gulf Coast Research Laboratory (USM-GCRL) were transported to JSU marine sciencelaboratory and acclimated to laboratory conditions for one week. Fish were then randomly assigned to 10 gallon aquarium tanks containing 25ppt water. The salinity in the tanks was then gradually adjusted to15ppt, 25ppt or 35ppt over a week period, after which the fish were anaesthetized, measured and weighed. There were three or two fish per tank and four replicates for each salinity treatment. Fish were fed pre-counted, weighed pelleted food, ad libitum, twice daily. After 28 days the fish were measured and weighed. There were no significant differences in the mean relative growth (P = 0.457) and food conversion ratio (P = 0.106) among salinity treatments. Preliminary studies suggest that juvenile red snapper can survive for several days at lower (~7 ppt) and higher (~45 ppt) salinities.

(12)

PREY PREFERENCE AND FUNCTIONAL RESPONSE OF ADULT BLUE CRABS CALLINECTES SAPIDUS TOWARDS TWO PREY SPECIES.

Ariel Settles¹, Anson Hines², Livingston Marshall¹. ¹Morgan State University, 1700 Coldspring Lane, Baltimore, Maryland 21251, ²Smithsonian Environmental Research Center, 647 Contees Wharf Road Edgewater, Maryland USA

Blue crabs are large epi-benthic predators that forage in diverse habitats along the Gulf and Atlantic coasts of the U.S. Our previous research showed that blue crab predation regulates multiple prey abundance and distribution, but this work focused on density-dependent interactions with one prey species at a time to infer regulation of benthic community structure. In this study, we considered prey preference and functional responses of large (140 mm) blue crabs to varying combinations of two prey species simultaneously, juvenile blue crabs 30-70mm and Baltic clams Macoma balthica 20-30 mm, in large lab tanks. Preliminary results indicate that adult crabs preferred clams to juvenile crabs, and addition of a second prey species altered the predator's functional responses to the prey species from type II to type III for juvenile crabs, and from type III to type II for clams. These changes could have major implications for stability of prey populations, since type II responses destabilize while type III stabilize prey populations.

(13)

Pathologic Changes Observed in Pygmy Sperm Whales (*Kogia breviceps*) Stranded Along the Florida Coast 1992-2000

Fred Williams III ^{1,2} and Ruth Ewing ¹

¹ National Oceanic and Atmospheric Administration, National Marine Fisheries Service, Southeast Fisheries Science Center, Miami Laboratory and ²Tuskegee University, College of Veterinary Medicine, Nursing, and Allied Health.

Necropsy and histologic evaluations were performed in 20 pygmy sperm whales (Kogia breviceps) that had been stranded along the Florida coastline. The animals were attended to by the US Southeastern Marine Mammal Stranding Network volunteers as they responded to stranding notices. Their personnel performed the necropsies and collected the tissue samples evaluated in this study. The gross lesions included findings consistent with the cardiomyopathy reported in the species. The microscopic lesions primarily include cardiomyopathies characterized by cardiac atrophy and cardiomyocyte degeneration. These changes were observed with no sex bias, and were commonly accompanied by non-specific changes indicative of cardiovascular collapse or shock. No definitive cause for the lesions was identified. The histiological reports of those cases were reviewed and are presented in a summarized format. This collection of data may be used to further understand the mechanisms behind why these animals strand.

ENVIRONMENTAL SCIENCES

(14)

Cumulative Impact of Photooxidation and Cometabolism on the Degradation of Petroleum Hydrocarbons by Indigenous Nearshore Bacteria

¹Milton Clarke, ¹Xiaoling Ding, ¹Richard Gragg, ¹Jennifer Cherrier and ²Jeff Chanton, ¹Environmental Sciences Institute, Florida A&M University, ²Department of Oceanography, Florida State University

Photooxidation and co-metabolism have been shown to enhance the biodegradation of refractory organic compounds. Petroleum consists of a complex mixture of hydrocarbons, some of which are biologically refractory. A series of 90 day incubations, with bacteria from a nearshore oil-contaminated (NC) site and a nearshore pristine (NP) site were carried out to evaluate the cumulative impact of photooxidation and cometabolism on the biodegradation of petroleum hydrocarbons (PHCs). Crude oil was photooxidized by exposure to natural sunlight for 16 h. A parallel series of incubations were carried out using unphotooxidized oil as the experimental control. Ground pinfish powder, a labile substrate, was used as the co-metabolite. Bacterial respired CO, taken at various time points was used as an indicator for the degree of oil degradation. Our results showed that in NC incubations, the CO₂ concentrations were 5000, 6900, and 11600 ppm for unphotooxidized oil, pinfish, and pinfish plus oil, respectively. Correspondingly, those observed in the NP incubations were 1,500, 14500 and 17800 ppm, respectively. The biodegradation of photooxidized oil was completely inhibited, at both sites, even under the presence of labile pinfish (CO, concentrations were about 500 ppm, which was the same as the background). These findings indicate that bacteria at NC site more efficiently degrade unphotooxidized oil than bacteria at the NP site. Additionally bacteria at the NP site can more efficiently degrade labile substrate than bacteria at the NC site. The time required for reaching the steady-state of CO₂ respiration for oil amended with pinfish (a few days) was much shorter than that of oil alone (about 40 days) at both of these sites. Photoinduced inhibition of CO₂ respiration may be due to the fact that after photomodification, some PHCs are more toxic than their parent compounds. Our gas chromatographic analysis of photooxidized oil also showed that more PHCs dissolved in water, especially smaller fractions, compared to unphotooxidized oil, suggesting that bacteria were exposed to higher PHC concentration in photooxidized oil incubations. Further experiments will investigate the relationship between the extent of PHC photooxidation and PHC photoinduced inhibition or enhancement of bacterial CO₂ respiration.

(15)

CHELATE – ASSISTED PHYTOEXTRACTION OF LEAD FROM CONTAMINATED SOILS USING WHEAT (*TRITICUM AESTIVUM* L.)

B. Crudup, R. Warren, M.F.T. Begonia, and G.B. Begonia, Jackson State University, Jackson, MS 39217

Preliminary studies indicated that wheat (Triticum aestivum L.) can tolerate and accumulate significant amounts of lead (Pb) in its shoots when grown in Pbamended sand. To further evaluate the potential of wheat for phytoextraction, a study was conducted to determine whether the timing of ethylenediaminetetraacetic acid (EDTA) application and acetic acid amendment can further enhance the shoot uptake of Pb. Two seeds were planted in each 150 ml super cell containing top soil and peat (2:1, v:v) amended with various levels of Pb and EDTA. Results revealed that wheat plants can tolerate toxic Pb concentrations as evidenced by the non-significant differences in shoot and root biomass among treatments. An exception to this general observation was the root inhibition of plants grown in 2000 ppm Pb that was amended with EDTA and acetic acid one week before harvest. Generally, root and shoot Pb uptake increased with increasing concentrations of soil-applied Pb. When no EDTA was added to the growth medium, the majority of the Pb was retained in the roots. However, when EDTA was applied one week before harvest, there was an enhanced Pb translocation to the shoots. Application of acetic acid in conjunction with EDTA led to a tremendous increase in shoot Pb uptake.

(16)

EFFECTS OF CHELATES ON THE BIOACCUMULA-TION OF LEAD FROM A CONTAMINATED SOIL US-ING COFFEEWEED (SESBANIA EXALTATA RAF.)

E. Dillon, O. Okuyiga-Ezem, T. Williams, M.F.T. Begonia, and G.B. Begonia, Jackson State University, Jackson, MS 39217

This study was conducted at the Jackson State University greenhouse to evaluate whether the addition of a synthetic chelate, ethylenediaminetetraacetic acid (EDTA), can further enhance the effectiveness of coffeeweed (Sesbania exaltata Raf.) as a phytoextraction species. Coffeeweed seeds were grown in various concentrations of lead (Pb) and EDTA mixed with a growing medium composed of sieved soil, peat, and sand (4:2:1, v:v:v). After six weeks of growth, the plants were harvested and their Pb contents were extracted using a modified nitric acid-hydrogen peroxide digestion technique. Lead concentrations were quantified by atomic absorption spectrophotometry. Results showed that coffeeweed can tolerate soil Pb levels up to 2000 ppm and that EDTA can significantly enhance the uptake of Pb by both the roots and the shoots. The results further demonstrated that pre-planting application of EDTA caused a greater Pb uptake by the plant compared to the untreated control and plants that were amended with EDTA one week before harvest.

(17)

Cytotonic Effects of Pentachlorophenol in Primary Hepatocyte Cultures Obtained from Channel Catfish (Ictalurus punctatus)

Waneene C. Dorsey, Paul B. Tchounwou, and Ali B. Ishaque, Environmental Toxicology Research Laboratory, NIH-Center for Environmental Health School of Science and Technology, Jackson State University, 1400 Lynch Street, Box 18540, Jackson, MS 39217, USA

Organochlorine residues are ubiquitous in the environment and have been found worldwide in soil, water, and air samples. Of particular interest are species of organochlorine compounds found in pesticides, because of their efficient biocidal activity against insects and microorganisms. An example of such pesticides is Pentachlorophenol (PCP), an organochlorine compound that has been used worldwide for many industrial and domestic applications. In this research, we investigated the acute toxicity of PCP to fish hepatocytes. The cytotoxicity of PCP was determined in vitro, using cultured primary hepatocytes from male channel catfish. The viability of cells, exposed for 2 days to PCP, was assessed by the flurorescein diacetate (FDA) method, using a Fluoroscan microplate reader. Study results indicated a linear relationship between cell survival and PCP toxicity. Upon 48 hrs of exposure, the NOAEL, LOAEL, LC₅₀ of PCP were 62.5 + 10.3, 125.0 + 15.2, and 2000.0 + 9.6 ppm, respectively. Based on these preliminary results, further studies are underway to assess the estrogenic potency of PCP, as well as its ability to transcriptionally induce stress genes in liver cells.

(18)

Cellular Response (Cytotoxicity and p53 Expression) to Arsenic and Atrazine by HepG2 Cell Lines

Reshuna Y. Durden, Paul B. Tchounwou, Barbara A. Wilson, and Ali B. Ishague

Environmental Toxicology Research Laboratory, NIH-Center for Environmental Health School of Science and Technology, Jackson State University, 1400 Lynch Street, Box 18540, Jackson, MS 39217, USA

Atrazine and Arsenic-containing compounds are among the leading chemicals that are used in America for agricultural purposes. Atrazine is a man-made herbicide used for the control of broadleaf and grassy weeds. It is persistent in the environment and contaminates ground waters and surface waters that are sources for human consumption and recreation. Lifetime exposure to Atrazine, at levels above 0.003 mg/L, has the potential of causing cancer. Arsenic is a carcinogen not only for skin, but also for internal organs such as the lung and bladder. Health effects associated with arsenic exposure include diabetes, cardiovascular disease, hearing loss, and neurological and neurobehavioral effects. The maximum contaminant level for arsenic in drinking water is 50 ug/L. The goals of this project were: (1) to determine the toxicity of arsenic trioxide and atrazine using the lactate dehydrogenase (LDH) assay, and (2)

to determine the cellular response mechanism of Arsenic and Atrazine in human hepatic carcinoma cell (HepG2) lines. To conduct this experiment, HepG2 cells were seeded at 1x10⁶ cells/mL and exposed to the chemicals for 48 hours. LDH analysis was used to determine the lethal concentration at which 50-percent of the cell population (LC₅₀) dies. Total protein concentration was determined using the Bradford Assay, and Western Blot analysis evaluated p53 cellular protein expression. The results indicated that Atrazine alone was nontoxic to HepG2 cells in the concentration range tested. The LC_{50} value for arsenic trioxide was shown to be 12 ppm. The tumor suppressor gene product p53, which is a critical mediator of the cellular response to DNA damage, was expressed after treatment of HepG2 cells with 12ppm of Arsenic.

(19)

The Influence of Nitrogenous compounds on Growth and Sexuality of Three Microalgal Species

Eric J. Fisher*, I. O. Farah, M. G. Hardy and H. S. Yang, Jackson State University, Jackson, MS 39217

Total growth, sexuality and utilization of various nitrogenous compounds by microalgae in 21-day batch culture media (Lewin's FW1 medium at 25°C within an illuminated growth chamber) were studied using microbiological, spectrophotometrical, microscopical and statistical techniques. Microalgae exhibit luxuriant growth in eutrophic ponds and lakes by metabolizing abundant nitrogen and phosphorous found in these areas. These abundant nutrients facilitate primary productivity and are reassembled in algal cells tobecome part of their biomass. Nitrogen is one of the most important inorganic/ organic nutrients utilized by microorganisms. This microalgal ability may, however, produce an excellent source of quality nitrogen for various uses. Our objectives are (1) to determine if microalgae will grow and transform poor quality nitrogen into quality biomass by their response to different nitrogenous sources in the growth medium and (2) to determine if primary products accumulate in microalgae exposed to various concentrations of nitrogen and the influence on algal sexuality. Three microalgae; Chlorella pyrenoidosa, Chlamydomonas reihardtii and Chlorococcum echinozygotum were inoculated in media containing twenty different nitrogenous compounds (including various amino acids) at 1, 10, 100, 1,000 and 10,000 ppm. Results showed statistically significant differences (p<0.01) among treatments. For the above algal species, mean and SD for total growth ranges were respectively as follows: 0.099 0.001 - 1.07 0.1, 0.071 0.03 - 1.13 0.07, 0.12 0.016 - 1.642 0.02 in OD units/ml. Mean sexuality (cell %/ml) ranges were respectively as follows: 10.8 7.75 - 69.6 3.10, 8.0 2.43 - 81.20 2.71, 3.33 0.11 - 67.00 2.00. All three species showed significant growth at higher concentrations (>1,000 ppm). Urea, asparagine, citrulline and uric acid showed the highest accumulation of organic material in algal cells. An inversely proportional relationship was found between zygotic cell counts and growth.

Therefore, it was concluded that the concentrations to be used for the best primary productivity and sexuality is at least 1,000 ppm. This study points to the potential of using microalgae to transform low quality and cheap nitrogen into biomass, a condition that could be exploited to improve plant cell culture productivity under aquatic conditions.

(20)

Axenic lines of *Spartina alterniflora* from Surface Sterilized Seeds.

Alisa G. Harkins¹, Trenis D. Palmer¹, Marc E. Frisher² and Chandra I. Franklin¹.

¹Marine Environmental Science and Biotechnology Research Center, Savannah State University, P.O. Box 20228, Savannah, GA 31410. ²Skidaway Institute of Oceanography, 10 Ocean Science Circle Savannah, GA 31411

The smooth cordgrass (Spartina alterniflora) is a dominant plant species in saltmarshes of southeastern United States, and it can account for up to 90% of biomass production. In anthropogenically impacted saltmarshes, interactions between S. alterniflora and sediment microbial communities appear to influence the uptake, transformation, and ultimate removal of contaminants including hydrocarbons, heavy metals and xenobiotics. Axenic (microbe-free) lines of S. alterniflora are needed for conducting experiments to assess the individual roles of S. alterniflora and microbes. Under natural conditions, it is not feasible to conduct such experiments, because they cannot be physically separated. In this report, the methodology to produce axenic lines of S. alterniflora from seeds surface-sterilized using chlorine fumes is described. Due to the microbe-rich habitat of S. alterniflora, its seed surface appears to be heavily infested with fungal and bacterial population. Surface sterilization of seeds using conventional methods did not yield satisfactory results. Therefore, an alternate method using chlorine fumes generated by mixing commercial bleach (Clorox) and glacial acetic acid was used. Seeds were exposed to chlorine fumes for 10-90 min (in 5-10 min increments) and subsequently germinated on sterile nutrient medium (100 seeds/treatment). Ten days later, data on number of seeds showing microbial growth (contamination) were collected. Seeds exposed for 10-30 min showed 5-10% contamination. Seeds exposed for 35 min and above showed no contamination. As expected, prolonged exposure to chlorine fumes reduced seed viability. Ten minute exposure reduced seed viability by 13%, whereas 90 min exposure resulted in 60% reduction of seed viability. At 35 min, 29% reduction of seed viability was observed. Results from these studies indicate that 35 min exposure to chlorine fumes is optimal for surface sterilizing S. alterniflora seeds. Viable surface-sterilized seeds germinated into normal plants with well-established root and shoot systems with no signs of microbial growth in the nutrient medium.

(21)

BIOCONCENTRATABLE POLYCYCLIC AROMATIC HYDROCARBONS & QUINONE PHOTOPRODUCTS IN EVERGLADES CANALS C-11 AND C-111.

Haynes, S. ^{1,2}; Gragg, R.D.¹; Orazio, C.E.²; Lebo, J.A.²; R. ²; Frederic Essien¹, Larry Robinson¹, and Maurice Eddington³. Florida A & M University, ¹Environmental Sciences Institute, and the ³Department of Chemistry, Tallahassee, Florida; ²USGS, Columbia Environmental Research Center, Columbia, Missouri USA

Urban runoff is a major source of polycyclic aromatic hydrocarbon (PAH) contamination in the Everglades. Solar ultraviolet radiation is a component of the Everglades temperate climate that has the potential to photo-modify some PAHs. Growing evidence suggests that the real hazards of PAHs to aquatic life may be due to the toxic effects of photooxidized PAH photoproducts formed by PAH absorption of ultra-violet radiation in sunlight. PAHs are difficult to measure by conventional means of sampling and a means of sampling and measuring ultra-trace levels of PAH photoproducts in the aquatic environment has not been reported. In this investigation samples were collected from 2 sites in the Everglades: Canals C-11/S-9 and C-111/S-177. Bioavailable PAHs and guinone photoproducts in the water column were sequestered using a semipermeable membrane device (SPMD). These devices mimic the bioconcentration process of lipophilic compounds by aquatic organisms. After a 28-day sampling period, the SPMDs were retrieved, dialyzed and analyzed by GC-FID and HPLC/UV. A new method for measuring guinone photoproducts was developed without any post-dialysis cleanup by direct HPLC/UV analysis. The GC and HPLC methods, in conjunction with SPMD sampling, enable PAHs in the Everglades to be measured at levels as low as 10ng/ mL. The limit of quantitation for the photoproducts is estimated to be in the g/mL range by this method. Naphthalene, fluoranthene, and chrysene were detected in SPMDs at both sites. No guinone photoproducts were detected. In addition to the PAH related compounds, the SPMD samples were also screened for organochlorine pesticides (OCP). The highest concentration levels of OCP detected from both sites were transchlordane and cis-chlordane with concentrations ranging between 11.5-12.2 and 8.9-9.8 ng/mL, respectively. Endosulfan was highest in canal C-111/S-177 with a concentration of 348 ng/mL. Other pesticides detected in SPMD included DDT, DDE and dieldrin.

(22)

PHYTOREMEDIATION OF LEAD – CONTAMINATED SOILS USING TALL FESCUE (*FESTUCA ARUNDINA-CEA* L.).

M. Ighoavodha, A. Butler, M.F.T. Begonia, and G.B. Begonia, Jackson State University, Jackson, MS 39217

Tall fescue (*Festuca arundinacea* cv. 'Spirit') had been found to tolerate and accumulate substantial amounts of lead (Pb) in its shoots when grown in Pb

amended sand. To further evaluate the suitability of tall fescue for phytoextraction, a study was conducted to determine whether the timing of ethylenediaminetetraacetic acid (EDTA) application and acetic acid amendment can further enhance the shoot uptake of Pb. Seeds were planted in 1.9 L plastic pots containing top soil, peat, and sand (4:2:1, v:v:v) amended with various levels of Pb and EDTA. Generally, root and shoot growth of tall fescue were not significantly affected at the two lower Pb levels, but were inhibited at the two higher Pb treatments with preplanting EDTA amendments. EDTA significantly increased shoot Pb uptake. However, there was no difference in Pb uptake whether EDTA was applied before planting or a week before harvest. Addition of acetic acid to the growing medium one week before harvest, further enhanced shoot and root Pb uptake only in plants grown at 1000 ppm Pb.

(23)

A STUDY OF THE EFFECT OF ULTRA VIOLET LIGHT ON RESIDENTS IN MISSISSIPPI.

Amalee Jayasinghe and Pao-Chiang Yuan, Department of Technology, School of Science and Technology, Jackson State University, Jackson, MS 39217.

The depletion of the ozone layer has increased the amount of harmful ultra violet rays reaching the earth. This results in damage to human health, plants and the ecosystem. Studies have shown that UV radiation causes skin cancer known as melanoma, suppresses the immune system and causes premature wrinkling. The object of this study was to determine whether UV radiation had any effect in the number of cases of skin cancer reported in Mississippi. Health statistics were obtained from the Department of Health along with measurements of UV radiation using ultra violet sunsor scales. Based on data obtained, we can conclude that UV radiation does cause an increase in skin cancer. This study will prove helpful in educating residents about the risks and ways to prevent skin cancers.

(24)

Transient Expression of the Reporter Gene BETA glucuronidase in Spartina Alterniflora Explants

Whitney H. Palefsky, Alisa G. Harkins and Chandra I. Franklin. Marine Environmental Science and Biotechnology Research Center, Savannah State University, P.O. Box 20228, Savannah, GA 31410.

The smooth cordgrass (*Spartina alterniflora*) is a dominant plant species in saltmarshes of southeastern United States, and it can account for up to 90% of biomass production. In anthropogenicallyimpacted saltmarshes, interactions between *S. alterniflora* and sediment microbial communities appear to influence the uptake, transformation, and ultimate removal of contaminants including hydrocarbons, heavy metals and xenobiotics. A gene transfer system is being developed in an effort to utilize transgenic technology to produce genetically engineered *S. alterniflora* that can be used as a biosen-

sor for heavy metal contamination in saltmarshes. Published protocols for genetic transformation or regeneration of S. alterniflora are not currently available. Therefore, studies in this laboratory focus on developing a gene transfer system and a compatible tissue culture system for regenerating transgenic S. alterniflora. As Agrobacterium mediated DNA transfer is not practical with grasses, particle bombardment mediated DNA delivery is being used. In this report, results from studies to optimize DNA delivery to fleshy young shoot explants of S. alterniflora are presented. The efficiency of DNA delivery was assessed based on transient expression of the reporter gene beta-glucuronidase (qus) as measured by the number of gus expressing units [(GEU) - blue spots resulting from the GUS histochemical assay]. Particle bombardment parameters such as helium pressure, vacuum level in the particle delivery chamber, particle travel distance, and the plasmid concentration were optimized for efficient DNA delivery. Obviously, increasing the plasmid concentration resulted in higher GEUs, whereas physical parameters controlling the velocity of particles greatly influenced the DNA delivery even at higher plasmid concentrations. Results from this study indicate that 1550 psi helium pressure, 28 inches of Hg chamber vacuum, 85 mm of particle travel distance and a plasmid concentration >1)g/delivery are optimal for DNA transfer to S. alterniflora explants.

(25)

TEMPORAL PATTERNS IN THE ABUNDANCE OF FISHES IN THE NEARSHORE AREAS OF THE ROSS BARNETT RESERVOIR

*Raymond Portis, *Crockett, R., *Chigbu, P., **Holman, T. & **Bull, L.

*Department of Biology, Jackson State University, Jackson, MS 39217; **Institute of Marine Sciences, Gulf Coast Research Laboratory, University of Southern Mississippi, Ocean Springs, MS 39566.

Information on spatio-temporal patterns in the abundance of prey and predators is important for assessing the extent of their spatial overlap and therefore the degree of predation pressure exerted on the prey. We conducted day time electroshocking in the Ross Barnett reservoir to assess spatial and temporal patterns in the abundance and size structure of fishes in the nearshore areas. Electroshocking was carried out, approximately bimonthly, from July 1999 to July 2000 in four areas of the reservoir. In each area we made three consecutive fifteen minute collections. All fish were placed on ice for later processing in the lab., or preserved immediately after collection in 10% formalin. In the lab. the fishes were identified, counted, measured and weighed. Eighteen (18) species of fish were captured. Threadfin shad (Dorosoma petenense) and gizzard shad (Dorosoma cepedianum),), important prey of largemouth bass (Micropterus salmoides) were the most abundant. Other abundant fishes were silversides and bluegill sunfish (Lepomis macrochirus). Abundances of threadfin and gizzard shad, and that of their major predator, largemouth

bass increased in the nearshore areas of the reservoir from summer to fall.

(26)

Effects of Cadminum on the Bioluminescense of *Vibrio fischeri*, and the Growth and Oxygen Uptake of a Heterogenous Population of Microorganisms Lamar Reed and Paul B. Tchounwou

Environmental Toxicology Research Laboratory, NIH Center for Environmental Health, School of Science and Technology, Jackson State University, 1400 J.R. Lynch St., P.O.Box 18540, Jackson, MS.

Bacterial In vitro assays such as the Microtox Assay have become an attractive alternative to traditional and costly fish and invertebrate methods for toxicological screening. In this research, we tested the toxic effects of cadmium against a marine bacterium (Vibrio fischeri), and a heterogeneous population of bacterium derived from the Pearl River in Jackson, Mississippi. Using the level of bioluminescence (Vibrio fisheri), and the kinetic of dissolved oxygen uptake and growth (mixed bacterial population) as measures of toxicity, cadmium concentrations effecting 50% reduction in these parameters (EC₅₀) were determined as toxic end-points. The activity quotients were also computed to determine the degrees of toxicity. Bacterial bioluminescence was assessed using the Microtox Assay. Optical density (measure of growth) and oxygen uptake were measured over an extended period of time (20 hours). EC₅₀ values of 0.79+0.12, 4.50+0.04 and 5.00+0.42 mg/L were recorded for bioluminescence, growth and oxygen uptake, respectively. As expected, these results indicated that *V.fischeri* was about one order of magnitude (8.3 times) more sensitive to cadmium toxicity than the mixed population of Pearl River microorganisms. Reductions in bioluminescence, growth, and oxygen uptake were directly correlated to cadmium concentration, with toxic levels rangingfrom slightly toxic in lower concentrations, to extremely toxic in higher ones. Upon 20 hours of exposure, a strong correlation (r²=0.98) was found between the times required to produce 50% reduction in dissolved oxygen concentrations (T50s), and the concentrations of cadmium, indicating a time-response relationship with respect to cadmium toxicity.

(27)

Cytotoxicity of Arsenic Trioxide and MSMA to Human Liver Carcinoma Cells: Comparative Study of Two Bioassays

Tanya Robinson, Paul B. Tchounwou, Barbara A. Wilson, Ali B. Ishague

Environmental Toxicology Research Laboratory, NIH-Center for Environmental Health School of Science and Technology, Jackson State University, 1400 Lynch Street, Box 18540, Jackson, MS 39217, USA

Arsenic is the 20th most abundant element. It is released to the environment as a result of natural phenomena. Several adverse health effects have been associated with the exposure of Arsenic and its toxicity depends on the chemical species involved. Cytotoxicity is the study of effect of acute (high) concentration of chemicals on living cells. Diphenyltetrazolium Bromide (MTT) and Fluorescein Diacetate (FDA) assays are among the most widely used methods to test the cytotoxicity of various environmental compounds. The objectives of this research were to find the concentration of Arsenic needed to kill 50% (LC $_{50}$) of HepG2 cells (cytotoxicity), to compare MTT and FDA methods used in evaluating cytotoxicity of Arsenic, and to compare the toxicity of Arsenic Trioxide (inorganic form) and MSMA (organic form). HepG2 cells were grown with DMEM medium in a CO₂ incubator until maximum growth, after which cells were washed and trypsinized. Next, fresh medium was added and centrifuged at 3,000 rpm for 10 minutes.180-ul of cells at a density of 300,000 cells/mL were seeded into each well of two 96-well tissue culture plates. Next, 20ul of serial dilution of Arsenic (0.2,0.4,0.8,1.6,3.1,6.3,12.5,25,50 and 100ppm) and MSMA (2, 4, 8, 16, 31, 63, 125, 250, 500 and 1000ppm) were added columnwise. The FDA assay was conducted by washing the plates once and adding 100ul of 10ppm Flourescein Diacetate solution columnwise to each well and placing the plates in the CO₂ incubator for 30-60 minutes before reading the fluorescence with the Spectrofluorometer. The MTT assay was conducted by adding 50ul of the 5,000ppm MTT solution to each well columnwise and placing the plates in a CO₂ incubator for 30 minutes. The medium was carefully removed and 200ul of DMSO was added to each well. The plates were incubated for 10 minutes in a CO₂ incubator and optical density was read at 405nm. The results showed there were no significant differences between the FDA and MTT assays based on theLC₅₀ values obtained from both assays. It is concluded that Arsenic Trioxide (inorganic) is more toxic than MSMA (organic Arsenic). The two methods gave comparable results and the difference in LC₅₀ between the two bioassays for Arsenic was not significant.

(28)

CYTOTOXICITY AND PROLIFERATION STUDIES WITH SODIUM ARSENITE IN SKIN DERMAL FIBRO-BLASTS, THP-1, AND JURKAT CELLS

Dwayne J. Sutton¹, Paul B. Tchounwou¹, and Hari H.P. Cohly^{2.} ¹Molecular Toxicology Research Laboratory, NIH-Center for Environmental Health, School of Science and Technology, Jackson State University, 1400 Lynch Street, Box 18540, Jackson; MS 39217; ²Department of Plastic Surgery, University of Mississippi Medical Center, Jackson, MS 39216

Ingestion of arsenic affects primarily the skin by causing keratotic lesions that may lead to cancer. Contamination of well water with arsenic leached from underground sediments leads to the formation of these precancerous skin lesions due to arsenic ingestion. It has been proposed that the mechanism underlying the unique properties of arsenic involves changes in mito-

genic signaling. Based on the hypothesis that arsenic exposure results in the modulation of both positive and negative regulators of cell proliferation, this study examined the acute cytotoxicity and proliferation of dermal fibroblasts, monocytes (THP-1) and T cells (Jurkat) in the presence of different concentrations of arsenic. Cytotoxicity was assessed by incubating THP-1 and JKT cells in RPMI 1640 and dermal fibroblasts in Iscove's DMEM with 10% fetal bovine serum, 1% streptomycin and penicillin for 96 hrs in 96-well microtiter plates, at 37°C in a 5% CO, incubator. Cell proliferation in 96-well plates was determined in cultured cells exposed for 24, 48, and 72 hours, using the MTT assay. The concentrations of sodium arsenite effecting 50% reduction in cell viability were found to range from 25 ppm to 50 ppm for all the cell-lines tested. The peak proliferation for Jurkat cells was 6 ppm and that for THP-1 and dermal fibroblasts was 1.5 ppm. These results show that arsenic affects proliferation at different concentrations for different cells.

(29)

Physico-chemical and Bacteriological Assesswment of the Water Quality in the Ross Barnett Reservoir in Central Mississippi

Paul B. Tchounwou, and Mike Warren

Environmental Toxicology Research Laboratory, NIH-Center for Environmental Health, School of Science and Technology, Jackson State University, 1400 Lynch Street, Box 18540, Jackson, Mississippi 30217, USA.

Ross Barnett Reservoir (RBR) serves as a source of water supply for the city of Jackson, Mississippi. It also constitutes an important site for recreational activities for many Mississippi counties' residents. A broad spectrum of illnesses have been associated with watercontact activities such as bathing, and swimming in recreational waters. In this research, we assessed the bacteriological quality of water in the RBR, and compared the bacteria levels to recommended criteria for public health protection. From 06/12/99 to 10/02/99, weekly water samples were collected from two different sites in the reservoir, and tested for a number of bacteriological parameters including: heterotrophic plate counts (HPC), total coliforms (TC), fecal coliforms (FC), and fecal streptococci (FS). Collected samples were also tested for basic physical and chemical characteristics of water. Study results indicated that the mean concentrations of bacteria at the two sampling sites were $5.6 \times 10^6 + 1.5 \times 10^6$, 4.5×10^4 + 5.3×10^4 , 5.0×10^2 + 1.6×10^2 , and 9.1×10^1 + 7.3x101 colony forming units per 100 mL of water for HPC, TC, FC, and FS, respectively. The values of physical and chemical parameters were at acceptable levels. However, bacterial densities often exceeded federal/state guidelines by several orders of magnitude, raising considerable public health concerns.

(30)

nfluence of Plant Growth Regulator Levels and Nitrogen Content on Callus Induction from *Spartina alternaflora* Explants

Laurie M. Thomas, Donna E. McDowell, Kashandra L. Jackson, Archie R. Hayes, Whitney H. Palefsky and Chandra I. Franklin.

Marine Environmental Science and Biotechnology Research Center, Savannah State University, P.O. Box 20228, Savannah, GA 31410.

In salt marshes of southeastern United States, smooth cordgrass (Spartina alterniflora) is a dominant plant species, and it can account for up to 90% of biomass production. Transgenic technology is being developed to genetically engineer S. alterniflora for use as a biosensor for heavy metal contamination in saltmarshes. An efficient tissue culture/regeneration system is needed for developing the transgenic technology. Callus induction is the initial response of explants cultured in vitro. In this report, results from studies to optimize plant growth regulator levels as well as nitrogen source and content of the tissue culture medium for callus induction are presented. Fleshy young shoot explants of S. alterniflora were cultured on medium containing various levels of a cytokinin [6-benzylaminopurine (BAP)] and an auxin [2,4-Dichlorophenoxy acetic acid (2,4-D)]. Similar explants were also cultured on medium containing organic or inorganic nitrogen source. Fleshy young shoots cultured on medium containing high levels of BAP (>5 iM) and 2,4-D (>20 iM) showed typical callus induction patterns. High levels of inorganic nitrogen content (20 mM ammonium 39.4 mM nitrate ions) in the culture medium caused necrosis and rapid death of explants, whereas explants cultured on medium containing low levels of inorganic nitrogen (7 mM ammonium and 28 mM nitrate ions) or an organic nitrogen source showed sustained growth. Results from these experiments indicate that organic nitrogen or low levels of inorganic nitrogen and high levels of a cytokinin and an auxin promote callus induction from young shoot explants of S. alterniflora.

(31)

PM-2.5 Monitoring in Flagstaff During the Prescribed Burn and Residental Wood Burning Seasons

Amanda Thompson and Marin Robinson, Northern Arizona University, Flagstaff, Arizona.

In 1997, the EPA passed a law to regulate fine particulate matter (PM-2.5), in addition to coarse particulate (PM-10), targeting particles with aerodynamic diameters 2.5 m. These regulations were promulgated due to concerns that fine particles can become lodged in the lungs and thereby contribute to pulmonary disease. In addition to health concerns, PM-2.5 contributes to visibility impairment and climate change, since small particles scatter visible light and increase albedo in cloudfree portions of the atmosphere. Major sources of PM-2.5 include fuel combustion, agricultural burning, and forest fires (natural or prescribed). Concomitant with the new PM-2.5 standards has been an expressed desire by Federal, tribal, and State land managers to use prescribed fire as a method for restoring wildland ecosystems. The fire-suppression policy of the 20th century (1910-1990) has left national forests in an unhealthy state, characterized by dense undergrowth and heavy fuel loads, increasing the likelihood of catastrophic wildfire. The potential benefits of prescribed fire are great, yet the smoke generated by such fires may put public health and welfare at risk. Thus the need to speciate chemical components of wood smoke, and identify those components with greatest toxicity, is an issue of primary importance.

To address this concern, we initiated a pilot project to collect PM-2.5 from wood smoke in Flagstaff, AZ. Flagstaff is located in the Coconino National Forest, the largest continuous ponderosa pine forest in the United States. Ongoing and proposed ecological restoration initiatives for this forest include the combined use of mechanical thinning, pile burns, and broadcast burns. Four portable air monitors (AirMetrics) were located at selected sites in Flagstaff to collect 24-hour samples of PM-2.5 during the prescribed burn season (Summer-Fall 2000) and residential wood burning season (February-March 2001). Monitors were operated as saturation samplers, to determine if the EPA 24-hour standard (65 g/ m³) had been exceeded. Monitoring results and plans for future PM-2.5 speciation studies during prescribed burns of the Coconino National Forest will be reported.

(32A)

EFFECTS OF SALINITY ON DEVELOPMENT OF HYLA REGILLA EMBRYOS.

Ezra Yohannes, Shaw University, Raleigh, NC and Vandergon, Davis, Brewster, Plank, Martin and Kats, Natural Sciences Division, Pepperdine University, Malibu, CA.

Chemical contamination in fresh water streams and lakes may have detrimental affects on amphibian species. The affect of salinity (0-7 ppt) on embryonic development, growth and survival in the Pacific tree frog *Hyla regilla* was examined. *Hyla regilla* embryos exposed to < 4 ppt showed normal developmental rates and survivability, whereas embryos exposed to 5 ppt salt concentration showed decreased survival, slower development and abnormal growth. Embryos exposed to 6 or 7 ppt showed slow growth, abnormalities and did not survive to hatching.

(32B)

IDENTIFICATION OF THE PHOTO-DEGRADATION PRODUCTS OF TNT INDUCED BY RIBOFLAVIN WITH GC/MS AND HPLC/MS/MS

Kui Zeng, Department of Chemistry, University of Science and Technology of China, Hefei, Anhui 230026, P.R. China; Huey-Min Hwang, Department of Chemistry, Jackson State University, Jackson, MS 39217; Hua Cui, Hongtao Yu, Department of Biology, Jackson State University, Jackson, MS 39217

2,4,6-Trinitrotoluene (TNT), a toxic explosive chemical, is also a suspected carcinogen. Therefore, it is of our interest to remove TNT from contaminated environment. The effect of riboflavin, an environmentally friendly sensitizer, on the photochemical fate of TNT in a freshwater environment was explored . It was found that riboflavin significantly enhanced the photo degradation rate of TNT in natural water environment. However, the degradation of TNT does not necessarily mean a removal of its deleterious characteristics; some of the degradation products of TNT such as 2-amino-4,6-dinitrotoluene (2ADNT) and 4-amino-2,6-dinitrotoluene (4ADNT) have been shown to have toxic and mutagenic effects on certain biological species. In this work, liquid chromatography/mass spectrometry and gas chromatography/ mass spectrometry were used for the identification of the degradation products of riboflavin-induced TNT degradation. 1,3,5-trinitrobenzene, 2,4,6-trinitrotoluene, 4amino-2,6-dinitrotoluene, 3,5-dinitroaniline, 2-nitroso-4,6dinitrotoluene and 2-amino-4,6-dinitrotoluene were identified in the studied system. In the presence of riboflavin, two new intermediates with max. absorption wavelength at 230 nm were found, demonstrating that transformation of TNT in the presence of riboflavin undergoes different pathways. Furthermore, the pathways for the TNT degradation are proposed.

ATMOSPHERIC SCIENCES

(33)

A STUDY OF TORNADO FORMATION AND ASSOCI-ATED TOPOGRAPHY IN ARKANSAS

Ashton Robinson Cook, Jackson State University, Jackson, Mississippi 39217

A study was performed on severe thunderstorm activity and tornado formation in Arkansas in relation to topography. On March 1, 1997, January 21, 1999, and February 24, 2001 mid latitude cyclones affected Arkansas by spawning tornado outbreaks. Some areas were affected multiple times by these outbreaks as tornadoes hit these areas. The study indicated that topographical difference plays a helpful role in the formation and development of tornadoes which increases the frequency of tornadoes occurring within the areas of topographical differences in Arkansas. More specifically, tornadoes can be focused in these areas where the traits of the landscape change from Ozarks and Ouachita Mountains to the north and west to the flat lands and delta regions of the south and east. The study also illustrates that there is a correlation between this "tornado alley" and the frequency and strength by which these tornadoes exhibit as they propagate through these areas.

(34)

FAST-RESPONSE ENVIRONMENTAL TEMPERA-TURE SENSOR CALIBRATIONS

Quentin Cowans, Thomas Smith, and Randal L. N. Mandock

Morehouse College, Earth System Science Program, Clark Atlanta University, 223 James P. Brawley Dr., SW., Mail Stop 241, Atlanta, GA 30314.

A number of methods have been devised over the years to measure the fluxes of sensible and latent heat near the air/land interface. Indirect methods include profile, Bowen-ratio, Priestley-Taylor, Penman-Monteith, bulktransfer, gradient, structure function, and others. The direct method of surface flux measurement is referred to as eddy correlation, where fluxes are determined as covariances of the perturbations of meteorological variables. In the eddy correlation method, vertical kinematic sensible heat flux is simply <w'T'>, and vertical kinematic latent heat flux is <w'g'>, where w' represents the vertical component of the perturbation wind vector, T' is the perturbation dry bulb temperature, q' is the perturbation specific humidity, and <> represents an averaging operation. The Earth System Science Program at Clark Atlanta University (CAU) has begun to calibrate fast-response wind, temperature and humidity sensors for later deployment in eddy correlation measurements of evaporation, momentum, buoyancy, and sensible heat flux. Initial test data for calibration spectra, averages, and flux measurements are reported. These measurements were made on the roof of the Research Center for Science and Technology at CAU.

(35)

DETERMINATION OF THE OPTICAL EFFECTS OF ATMOSPHERIC AEROSOLS USING SUN PHOTOM-ETERS

Adwoa K. Gyekye

Clark Atlanta University, Department of Physics

The Clark Atlanta University (CAU) Earth System Science program operates an Atmospheric Optics Observatory on the roof of the Research Center for Science and Technology. This observing facility provides a platform for instruments used to monitor the amount of solar energy reaching Earth's surface throughout the day. An Ascension Technology RSP (Rotating Shadowband Pyranometer) measures the global, direct, and diffuse radiation at the surface for visible wavelengths. In addition, a NOAA automated weather station operated at the observing site provides meteorological data on atmospheric pressure, temperature, humidity, wind speed, and wind direction. The ultimate goal of this project is to be able to model the effects of atmospheric haze (aerosol particles) and water vapor variations on the total amount of energy from the sun that reaches the surface. To improve our ability to interpret data obtained by the RSP and the meteorological instruments, CAU is developing a simple, inexpensive atmospheric haze sensor that uses an LED (light-emitting diode) as a narrowband detector. Six different LED wavelengths are available, and data from such a device operated throughout the day at multiple wavelengths can be used to determine the fraction of incident solar energy that has been lost due to the total amount of haze and water vapor in

the atmosphere. Sun photometers have been built and operated for each of the different LED wavelengths. The sun photometer measurements are analyzed by plotting the natural logarithm of the solar radiances measured with the photometer versus the secant of the zenith angle of the sun at the time of each observation (Langley plots). The optical thickness of the atmosphere is obtained by measuring the slope of the plotted line. Results obtained with the CAU sun photometers will be presented.

(36)

SODAR OBSERVATIONS OF THE ATLANTA URBAN BOUNDARY LAYER

Ilomechina, Sochi and Randal L. N. Mandock Clark Atlanta University, Earth System Science Program, 223 James P. Brawley Dr., SW. Mail Stop 241, Atlanta, GA 30314

An Aerovironment 300C monostatic sodar was installed on the roof of the Research Center for Science and Technology at Clark Atlanta University in summer 2000. The sodar is located about one mile west of downtown Atlanta, 22 meters above ground level, in a medium height and density urban environment.

The sodar is currently operating in the facsimile mode, but existing software is under revision to enable it to produce Doppler vertical wind speed and variance measurements early in 2001. Facsimile records showing meteorological phenomona such as fog and low cloud layers, frontal passage, rise and descent of the convective boundary layer, and nocturnal structure at the top of the boundary layer will be described and compared with meteorological data from our rooftop weather station.

(37)

INTER-COMPARISON OF REBS Q*7.1, PDS7.1, & THRDS7.1 RADIOMETERS'S PERFORMANCE CHAR-ACTERISTICS

Dwayne C. Joseph & J. Quentin Cowans Clark Atlanta University, 223 James P. Brawley Drive, Atlanta, GA 30314

Radiometers are instruments used for detecting and measuring the intensity of radiant energy. With the use of net radiometer, total hemispherical radiometer, and a pyranometer these instruments will assist in studying the radiative balance within the atmospheric boundary layer in Atlanta. Furthermore, this research will take into account the role clouds and surface albedo partake in comprising the radiative balance.

(38)

AN INTERCOMPARISON OF TWO ULTRASONIC AN-EMOMETERS

Pierson, Bryant R. and Randal L. Mandock Clark Atlanta University, 304 Lakeside Village Dr., Atlanta, GA 30317

A low cost R.M. Young Ultrasonic anemometer was intercompared with a more expensive Solent Research

Ultrasonic anemometer. Mean and standard deviations, comparability, and bias of wind speed, wind direction, the vertical wind component, and the speed of sound were calculated. Also presented in this research are response characteristics of the sensors, computed accuracy, momentum and buoyancy flux, and virtual temperature.

(39)

GAS TRAPPING IN VAPOR DEPOSITED WATER-ICE FILMS (90-145 K)

Brian Raymond and Marin Robinson (Northern Arizona University, Flagstaff, Arizona)

Water-ice is abundant throughout the solar system and can influence both atmospheric chemistry and composition. One way in which ice influences composition is by trapping and transporting gases, such as with comets. As comets form, the amount of gas trapped depends, in part, on the phase of the condensing ice (amorphous or crystalline). As a comet approaches the Sun, amorphous ice crystallizes, releasing some of the trapped volatiles. Eventually the comet sublimes, releasing the rest. The purpose of this study was to investigate this process with CO₂, CH₄, CO, and N₂. In a high vacuum chamber, a thin film (375 nm) of amorphous (90-110 K) or crystalline (110-145 K) ice was co-deposited with a selected gas (10:1 gas: water) on a pre-cooled Al substrate. The film was annealed (1K/min) through the phase change (147-153 K) and sublimation of the ice film (162-183 K). The phase change was monitored using grazing-angle Fourier Transform Infrared-Reflection Absorption Spectroscopy, while the gas release was detected using integrated flux to a mass spectrometer. Results show that all four gases were trapped by ice. CO₂, and to some extent CH₄, were released during the phase change and sublimation; N, and CO were released only during sublimation. Amorphous ice trapped more CO, than crystalline ice, but roughly equal amounts of N_2 , CH_4 and CO.

(40)

Diagnosing and Predicting Extreme Precipitation Events in Southeast Brazil

Aisha C. Reed and Brant Liebmann. NOAA Climate Diagnostic Center

Extreme daily precipitation events that occurred in Sao Paulo, Brazil from 1979-1995 during the austral summer are examined to document the atmospheric conditions associated with their occurrences. In this study, events are studied in which 14% and 30% events of the average seasonal total rainfall fell in one day at, at least one station. Mean composites and anomalies are obtained for the 500-millibar heights, to make synoptic patterns, and outgoing longwave radiation (OLR), to observe convection, for the day of and days preceding and following the event. There is a stationary trough on the days preceding and the day of the event, and the OLR indicates that convection is in the area at least one day before the event and begins to dissipate the day after an event.

(41)

RAINFALL ESTIMATES OVER WEST AND CENTRAL AFRICA FROM TRMM PR, TRMM MERGED, CMAP, GPCP AND SSM/I: A COMPARISON OF THE INTER-ANNUAL VARIABILITY FOR 1998 AND 1999.

Andrea Sealy and Gregory Jenkins.

Department of Meteorology, Pennsylvania State University, 503 Walker Building, University Park, PA 16802.

The accurate measurement of tropical rainfall is essential to understanding interannual variability, climate change, the hydrologic cycle and its link to the general circulation. There are numerous methods for estimating tropical precipitation including rainguages, the Special Sensor Microwave Imager (SSM/I) satellite, Outgoing Longwave Radiation (OLR) measurements and, more recently, the Tropical Rainfall Measuring Mission (TRMM) satellite which carries as one of its precipitation measuring instruments the first space-borne precipitation radar (TRMM PR). These data sources differ in their rainfall estimates over the tropics due to differences in horizontal resolution as well as spatial and temporal sampling. In this study I have compared the interannual variations in rainfall estimates for 1998 and 1999 during the West and Central Africa wet seasons from TRMM PR, TRMM Merged, CPC Merged Analysis of Precipitation (CMAP), Global Precipitation Climatology Project (GPCP) and SSM/I data. Previous results for West Africa have shown that the TRMM PR and TRMM Merged rainfall estimates differ in the intensity and occurrence of maximum rainfall during June-August 1998. The results from this study point to differences in the amount of rainfall estimated by the different datasets and the occurrence of maximum rainfall during the rainy season. The results also indicate differences in interannual variability of the rainfall estimates from the five datasets.

(42)

A STUDY OF PREDICTIVE MODELS FOR FORECAST-ING TROPICAL CYCLONE AND HURRICANE ACTIV-ITY OVER THE GULF OF MEXICO

Jamese Dacia Sims and R. Suseela Reddy Department of Physics, Atmospheric and General Sciences, Jackson State University, Jackson, MS

Predictive models including a) a regression model and b) Hurricane Predictive Index (HPI) have been developed for predicting the origin and evolution of tropical cyclones and hurricanes over the Gulf of Mexico. These models describe the air-sea interactions and associated tropical cyclone and hurricane activity using NOAA GOES satellite data and data from buoys in the Gulf of Mexico. These models were tested for 1999 (August 18-25) Hurricane Bret, that was formed and developed in the Gulf of Mexico. The models simulated and predicted the air sea interactions and associated tropical cyclone/hurricane activity. (43)

CHARACTERIZATION OF FORTY-TWO YEARS OF RAINFALL DATA FOR EARLY COUNTY, GA.

Monesa Watts, Jackson State University, Jackson, MS and Donald Perkey, Global Hydrology and Climaate Center, Huntsville, AL.

Knowledge of water supply and demand associated with rural and agricultural communities may be greatly beneficial with regard to periods described by water deficit. One of the most important parameters associated with water deficit is the amount of rainfall. The purpose of this study was to determine the frequency of occurence and intervals between rainfall amounts along with calculating the mean and standard deviation for Blakely, GA located in Early County. Daily precipitation totals along with the climatology for the region were obtained from the Southeast Regional Climate Center through the University of Georgia's Website. Excel was used to evaluate all collected data and to estimate the mean rainfall amount; the frequency of precipitation amounts were found to be greatest in March and decrease throughout the following months with a slight increase in July over the forty-two year period. Regarding intervals between daily rainfall, there were greater occurrences of consecutive days without rainfall than consecutive days with rainfall. Data Analysis of spectral distribution of precipitation, frequency of precipitation events, and intervals between precipitation days led to a characterization of precipitation for Early County, GA.

(44)

Some Aspects of Air-Sea Interactions and Associated Tropical Cyclones over the Gulf of Mexico Using Satellite and Buoy Data

Ashley Zachary and R. Reedy, Dept. of Physics, Atmospheric Sciences and General Science, Jackson State University, MS 39217

Under the NASA FAR program, a study has been undertaken for investigating some aspects of air-sea interactions and associated tropical cyclones over the Gulf of Mexico using satellite data. In the present study, we investigated the air-sea interactions for Hurricane Gordon and the related hurricane activity for the year 2000. NOAA satellite GOES imagery and buoy data from NDBC for sea surface temperature and other meteorological variables were used in the to calculate heat, momentum, and moisture fluxes during the formation and development of hurricanes. The study suggested that a) sea surface temperature was maximum (29), b) a 3-5 day oscillation in the heat flux was noticed before the 3 days of the hurricane formation, and c) a strong correlation was noticed between heat fluxes and the sea level pressure.

ABSTRACTS FROM RESEARCHERS AND RESEARCH ORGANIZATIONS

OCEANIC/MARINE AND FISHERIES SCIENCES

(45)

ATLANTIC COD AND HADDOCK EGGS ON GEORG-ES BANK, 1995 TO 1997; INTERANNUAL VARIATION IN DISTRIBUTION, ABUNDANCE AND MORTALITY RATES.

Peter Berrien and John Sibunka

NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, James J. Howard Marine Sciences Laboratory, Highlands, NJ 07732

GLOBEC Broadscale Surveys on Georges Bank were implemented to assess bank-wide features on a monthly basis during the first half of the calendar year. These surveys provide information on distribution and abundance and on biological rates such as mortality/ survival and growth of planktonic life stages of target species. This bank-wide approach provides some of the 'big picture' examination of potential factors in the determination of recruitment rates, as compared to the more focused approach of 'process studies'. Analysis of egg collections of the two targeted fish species, Atlantic cod and haddock, provides the starting point in a 'life table' assessment. It is assumed that egg production rate parameters can be linked with those of larvae, and with the adult stocks that produced them. Additionally, because eggs are passive constituents of the plankton their fate can be strongly influenced by circulation patterns on the Bank.

(46)

A TRIBAL TRAINING AND DEMONSTRATION PROJECT ON SHELLFISH AQUACULTURE: A NORTHWEST INDIAN COLLEGE AND WASHINGTON STATE UNIVERSITY COLLABORATION.

Gleyn E. Bledsoe

Northwest Indian College, 2522 Kwina Rd., Bellingham, WA 98226

The Northwest Indian College (NWIC), in collaboration with Washington State University (WSU), established a Tribal Shellfish Extension Training and Demonstration Facility at the campus of NWIC. The facility is used to train conventional NWIC students and Native American's who desire to establish their own shellfish farms or to work in shellfish aquaculture. It is administered as part of the NWIC/WSU Cooperative Extension Program serving the tribal and non-native communities within the states of Washington, Oregon, and Alaska. The facility also provides Washington State University with a marine aquaculture facility that is available to their Cooperative Extension and academic programs. Previously, WSU did not have such a facility.

An integral part of the project was the development of a shellfish demonstration area in which students can physically construct and experience several methods of shellfish aquaculture. Another function of the demonstration area is to conduct research into new methods of husbandry that improve shellfish growth, permit shellfish grow out in bottoms that are not otherwise suitable for such, and to reduce adverse impact to the environment. An example of this was the construction and maintenance of a modified post and longline system to grow Pacific Oysters (Crassostrea gigas) in an area plaqued by severe wind/tidal damage that previously prevented successful grow out. In addition, the area was the site of a sensitive eelgrass bed. The successful results of the project and information pertaining to the facility in general are presented.

(47)

À PRELIMINARY ANALYSIS OF GROWTH DATA OF JUVENILE LOGGERHEAD (CARETTA CARETTA) SEA TURTLES FROM NORTH CAROLINA, U.S.A. Joanne Braun-McNeill, NOAA, National Marine Fisheries Service, Southeast Fisheries Science Center, Beaufort, North Carolina 28516; Sheryan P. Epperly, NOAA, National Marine Fisheries Service, Southeast Fisheries Science Center, Miami, Florida 33149; Larisa Avens, NOAA, National Marine Fisheries Service, Southeast Fisheries Science Center, Beaufort, North Carolina 28516; and Samuel Sadove Long Island University, Southampton College, Southampton, New York 11968

Determining the age at which sea turtles mature is vital to understanding sea turtle demography, thereby facilitating conservation management practices. However, as sea turtles cannot be aged based on external characteristics, it is only possible to estimate the age at maturity using indirect methods such as measuring the growth rates of wild, recaptured turtles. Previous studies have documented a higher rate of growth for wild, juvenile loggerhead (Caretta caretta) sea turtles in the southern Bahamas and Florida than in Virginia. The National Marine Fisheries Service sea turtle program in Beaufort, North Carolina has been conducting mark-recapture studies in Core and Pamlico Sounds, North Carolina since 1988, and has tagged a total of 978 loggerhead (Caretta caretta) sea turtles, most of which were incidentally captured in commercial pound nets. Data from the New York State Marine Turtle Project were also included in the analysis. Straight-line measurements of carapace length (notch-tip) taken for each turtle ranged from 41.4-103.0cm. External Inconel Style 681 tags applied to both rear flippers and internal passive integrated transponder (PIT) tags injected into the left front flipper enabled identification of recaptured turtles. Growth rates (Carapace length initial - Carapace length at recapture / Years at large) were assigned to a 10cm size class based on the mean of the initial and recapture length and averaged. Multiple t-tests were used to calculate differences in mean growth rates among the recaptured loggerheads from the different regions of the

western Atlantic. To date, 29 loggerheads have been recaptured after being at large for 11 months or more, allowing for measurable growth. Similar to Virginia, North Carolina and New York had slower growth rates compared to Florida and the Bahamas although these differences were not always significant. Differences in growth rate have been attributed to the seasonal availability of food as well as the costs associated with seasonal migrations. Growth rates may also vary depending upon the genetic composition of the respective populations. This may have important ramifications for the management of the declining northern nesting population of loggerhead sea turtles.

(48)

LARVAL HATCH FECUNDITY FOR A STOCK OF FRESHWATER PRAWNS RAISED IN ARIZONA. George B. Brooks, Jr.,

Adjunct Assistant Professor, Wildlife and Fisheries Resources, University of Arizona, Tucson, AZ

Successful prawn hatchery management requires a knowledge of the number of brood stock required to produce a defined number of postlarvae. In *Macrobrachium rosenbergii*, Larval Hatch Fecundity (LHF) is defined as the number of larvae released from the egg mass following incubation. In the literature there are two reported values for average LHF, 1,000 larvae per gram of female prawn weight and approximately 400 larvae per gram of female prawn weight. The research detailed in this paper explores the LHF for a stock of *M. rosenbergii* commonly available in the United States. The results suggest a LHF value close to 400 for this particular stock of prawns. This particular LHF value could be the result of a number of factors including environmental, genetic differences and differences in rearing technique.

(49)

AQUACULTURE AS A TOOL TO TEACH SCIENCE IN THE GILA RIVER INDIAN COMMUNITY

George B. Brooks, Jr. and Joyce Baldwin, Department of Land and Water Resources, Gila River Indian Community, P.O. Box C, Sacaton, Arizona 85247

An environmental/science education program designed to provide a fundamental understanding of biological and environmental science for K-12 students has been initiated in the Gila River Indian Community (GRIC) near Phoenix Arizona. As part of this program, aquaculture was used as a method for teaching the scientific method, cell biology, mathematics, development and ecology. This 9 school program was designed to prepare O'otham (Pima) and Pee Posh (Maricopa) students for careers in natural resource management or as preparation to enter college. The program included gaining experience in tilapia egg incubation and tilapia growth studies.

The breakthrough for the concept occurred in 1997 when students from Estrella school on the Community, took a 4^{th} place with a tilapia culture project at the Inter-

national Science fair. In 1999, students from Vechij Himdag MashchamakuD (New Pathways to Learning) alternative high and middle school on the Gila River Indian Community, competed in the 46th annual Central Arizona Regional Science and Engineering Fair and took home three awards. Most recently a pilot scale commercial fish farm at the GRIC Juvenile Detention an Rehabilitation facility has been added to the program.

(50)

AN OVERVIEW OF THE TRIP INTERVIEW PROGRAM Pamela Brown-Eyo, NOAA, National Marine Fisheries Service, Southeast Fisheries Science Center, 75 Virginia Beach Drive, Miami, Florida 33149

NOAA- Fisheries, Southeast Fisheries Science Center maintains several fisheries statistical data collection programs. These programs collect information that is used to manage the marine fish stocks of the South Atlantic Ocean. One of those commercial fisheries databases is the Trip Interview Program (TIP). The TIP program provides details such as size frequency for species landed. TIP sampling is conducted throughout the southeast states and Puerto Rico and the US Virgin Island. In each area a TIP Scientist monitors the local marine fishing activity. The Scientist interviews the fishermen and collects fish measurements and biological samples on individual fish from the fishing trip. These interviews provide specific information, such as, area fished, hours fished, species landed, the pounds landed, the length of the fish or the width of the crabs, the species sex and the otoliths for age determination. This data is used to evaluate the growth and health of the fish populations. Since 1985, the TIP program has interviewed 49,891commercial fishing trips, collected data on 543 species of marine fish and shellfish, measured the length of 2,993,933 individual species, and removed 98,380 otoliths for aging studies. The TIP program continues to provide detailed data for the management of the Nation's marine resources.

(51)

PRELIMINARY INVESTIGATIONS OF CRAB PREDA-TION ON BAY SCALLOPS

Joseph Choromanski and Sheila Stiles

NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, Milford Laboratory, Milford, CT 06460

In the course of bay scallop aquaculture research conducted at the National Marine Fisheries Service Laboratory in Milford, CT, excess scallops were donated to Connecticut municipal shellfish commissions for free-planting in area waters. The practice of free-planting or tossing seed scallops (10 - 40 mm) directly into the water has come under scrutiny because of the observable decreasing return in the number of adults (>60 mm) caught by recreational fisherman in the towns that have such policies. Field studies of bay scallops have suggested a variety of causes for population fluctuations

including habitat loss, genetic inbreeding depression, and predation. It is generally known that crab predation can be a major factor in survival and growth of bay scallops for reseeding or stock enhancement efforts, especially in sites devoid of eel grass which can serve as a refuge for small scallops.

To evaluate crab predation on scallops, an experimental study was conducted with green crabs (Carcinus maenas) in the laboratory. Six treatment aquaria with 10° C flowing seawater were established with a single crab and 10 scallops. A seventh aquarium, with 10 scallops and no crab to check for non-predator related mortality, was used as a control. Four separate trials were run with scallops in each of the following size classes: 10, 20, 30 and 40 mm shell height. Trials were duplicated with a second set of crabs and scallops. Observations were made at 1, 6, 20, and 48 hours for each study. Results indicated that larger-sized scallops had higher survival rates, indicating some degree of refuge from predation by green crabs. In addition, damage to scallop shells was manifested in a characteristic appearance which could be used in identifying mortality by crab predation in the field.

A smaller scale project involved a comparison of bay scallop predation by Asian shore crabs (Hemigrapsus sanguineus) versus similar-sized green crabs. Six containers were set up for each crab species with fifteen scallops and one crab per container. The shell height of scallops ranged from 6-8 mm and the carapace width of crabs ranged from 18.0 to 25.3 mm. Observations were made after 48 hours when the experiment was terminated. The most notable difference was the number of scallops eaten by the male and female crabs. Generally, male crabs ate all of the scallops in their containers, while the female crabs ate very few to none. This difference could be attributed to the larger size of the claw of the male crabs. There was a slight difference in the number of scallops eaten by the green crabs versus Asian shore crabs. These observations can be used for planning purposes when attempting to enhance or replenish scallop populations.

(52)

POPULATION ENHANCEMENT EFFORTS FOR THE BAY SCALLOP, ARGOPECTEN IRRADIANS, IN THE NIANTIC RIVER ESTUARY, CONNECTICUT, USA. Ronald Goldberg, Jose Pereira and Paul Clark, NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, Milford Laboratory, Milford, CT 06460; Bernard Faber, David Porter, Lorenz Rinek, Barbara Kanter, Eric Kanter and Walter Lord, Waterford East-Lyme Shellfish Commission, 5 Rope Ferry Road, Waterford CT 06385

The Niantic River supports presently a small population of bay scallops, *Argopecten irradians*, that is harvested recreationally. Numbers of bay scallops have fluctuated greatly with a peak level estimated as high as 20 million animals in the 1940s (Marshall, 1960). Three potential enhancement strategies were evaluated: 1)

collection of natural spatfall, 2) direct re-seeding, and 3) over-wintering of hatchery-reared stock for creation of spawner sanctuaries. Assessment of natural spatfall in 1997 indicated that peak spawning occurred in late July and that spat were dispersed widely, however, too few spat were available for enhancement activities. In direct re-seeding experiments, time of planting and the inferred predation intensity were major factors affecting survival, while planting density had no significant effect. The Waterford East- Lyme Shellfish Commission (WELSCO) held 26,000 bay scallops in suspension culture during the 1998 – 1999 winter, of which, 60 - 80 % survived and spawned during the summer of 1999 within mobile spawner sanctuaries (cages suspended on long-lines). This effort is being repeated during 1999 - 2000. An annual recreational harvest survey has been initiated to assess enhancement activities. The pro-active approach of WELSCO in using aquacultural methods for enhancement of bay scallop populations is appropriate when natural recruitment is poor and habitat and environmental conditions are not limiting.

(53)

A CONTRIBUTION TO THE EARLY LIFE HISTORY OF DEEP-SEA SMELTS (FAMILY:BATHYLAGIDAE) IN THE GULF OF MEXICO.

Alonzo N. Hamilton, Jr. and Joanne Lyczkowski-Shultz NOAA, National Marine Fisheries Service, Southeast Fisheries Science Center, Mississippi Laboratories, Pascagoula Facility, 3209 Frederic St., P.O. Drawer 1207, Pascagoula, MS 39568-1207

Five species in a single genus of deep-sea smelts occur in the central western Atlantic and among these the larvae of only three species have been described; Bathylagus bericoides, B. longirostris, and B. compsus Southeast Area Monitoring and Assessment Program (SEAMAP) ichthyoplankton collections are taken in the deep ocean habitat of these fishes within the Gulf of Mexico only during April to June with infrequent observations during winter months. Under current SEAMAP protocols bathylagid larvae are initially identified to the family level by scientists at the Polish Sorting and Identification Center in Szczecin, Poland. A recent synthesis of existing literature on the Bathylagidae uncovered numerous questions concerning their nomenclature, species validity and larval identification. Reexamination of archived bathylagid larvae from Gulfwide collections has allowed us to describe the distribution of the larvae of these fishes and to investigate their occurrence relative to oceanographic conditions and features in the deep Gulf.

(54)

CONTINUING THE LEGACY OF MARINE EDUCATION AND RESEARCH PARTNERSHIP: THE SAVANNAH STATE UNIVERSITY/ NATIONAL OCEANOGRAPHIC AND ATMOSPHERIC ADMINISTRATION COOPERA-TIVE MARINE EDUCATION AND RESEARCH PRO-GRAM: (SSU/NOAA CMER).

Dionne L. Hoskins

NOAA, National Marine Fisheries Service, Southeast Fisheries Science Center, Savannah State University, Savannah, Georgia 31404

The Cooperative Marine Education and Research (CMER) Program was established on February 6, 1995 when a Memorandum of Understanding was signed between Savannah State University (SSU) and the National Oceanographic and Atmospheric Administration (NOAA). The relationship between NOAA and SSU is a longstanding one. NOAA has provided substantial guidance and support to Savannah State University for almost 30 years, beginning in the early 1970's when the university first began offering marine science courses and in 1979 when SSU formalized a Bachelor of Science degree in Marine Biology. NOAA has continued to provide vessels, dive training, student support and capacity-building assistance to Savannah State University while Savannah State University has offered research, education products, and information to NOAA through formal grants, contracts, and Memoranda of Understanding since 1981. The relationship between NOAA and SSU has now been expanded to provide for even more active cooperation in the advancement, organization, and operation or marine research, education, in-service training, and demonstration programs. In its first year, the CMER program has supported several coordinated research and education programs funded by Sea Grant and the Department of Education. These include internship opportunities that expose students to marine aquaculture and workshops in statistical analysis, research presentation and website design, and marinerelated geographical information system (GIS) applications.

(55)

NOAA'S CENTER FOR COASTAL FISHERIES AND HABITAT RESEARCH: A JOINT NOAA FISHERIES AND NATIONAL OCEAN SERVICE FACILITY

Donald E. Hoss

101 Pivers Island Road; Beaufort, NC 28516

Founded in 1899, the laboratory has served under various government agencies since opening. Currently there are 75 +employees, plus port samplers, post-doctorate researchers, graduate and undergraduate students who staff the laboratory. Research topics are broad in scope and support NOAA's goals to build sustainable fisheries, sustain healthy coasts, and recover protected species.

Organizational structure of the Center consists of one research branch with 9 teams conducting research on environmental and fishery questions. Applied Spatial Ecology and Habitat Characterization, Coastal Ecosystem Research and Restoration, Fisheries Oceanography and Ecology, Plankton Ecology and Physiology, and South Florida Fish Ecology and Contaminants teams address environmental issues related to ecology of living marine resources in various habitats. Research

projects involve ecological processes in salt marshes and seagrass beds and the fish and shellfish that utilize them in locations ranging from Virginia to south Florida to Louisiana, Texas, and California coasts. State of the art ecological, chemical, biochemical, and satellite imagery methodologies are used in the course of these studies. Fisheries focused investigations are conducted by the Fish Biology, Population Dynamics and Cetacean and Sea Turtle teams. The research is oriented toward understanding environmental and fishery factors that control the abundances of important fishery species, sea turtles, and marine mammals. The reef fish community of the SEU.S. continental shelf and menhaden resources of the Atlantic and Gulf of Mexico receive major attention by these teams. Population model development and evaluations of stock dynamics for fishery management in coastal fisheries as well as highly migratory species are conducted. Sea turtle studies focus on biology and dynamics of loggerhead, green, and Kemp's ridley in the NC coastal waters. Cetacean studies deal with all coastal marine mammals and with a concentration on the coastal bottlenosed dolphin stocks. The Information Technology and Spatial Analysis team provides statistical assistance, satellite imagery, and spatial data that are used by other research groups within the Laboratory, and they make the satellite data available to academic institutions and the private sector.

Researchers for these teams provide scientific and management advice to Fishery Management Councils along the Atlantic and Gulf coasts and to habitat managers in NOAA and other Federal and state agencies. Frequently staff are called upon to provide scientific information in instances where there has been environmental damage, and scientific advice is needed on the impacts as well as potential approaches to mitigate for impact and restore habitats to improve both damaged habitats and fisheries.

(56)

OVERVIEW OF FOOD WEB DYNAMICS IN THE NORTHWEST ATLANTIC: DETECTING CHANGES IN KEY PROCESSES AND PARAMETERS IN A MULTI-SPECIES CONTEXT.

Jason S. Link, Frank P. Almeida, Cheryl G. Milliken, and Lance P. Garrison. Food Web Dynamics Program, NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, 166 Water St., Woods Hole, MA 02543

The fish component of the northwest Atlantic continental shelf ecosystem has undergone well-documented changes over the past three decades. Particularly, gadids, flatfish, and other demersals are less abundant whereas smaller pelagics and elasmobranchs are more abundant than historical levels. In addition to abundance, we monitor several diet parameters including percent diet composition, percent prey frequency, total stomach contents, predator size, and prey size to estimate and assess major processes and rates for key fish in this ecosystem. Some of the major processes and rates we examine from these parameters are 1) trophic links and interaction strengths for multi-species models, 2) food web statistics, 3) M vs. F, 4) the partitioning of energy, biomass, and yield, 5) recruitment bottlenecks, 6) essential fish habitat, and 7) consumption, predation, competition and production rates. Our time series extends for 25 years, contains more than 100 predators and 1200 prey items, ranges from Cape Hatteras to Nova Scotia, covers over 83,700 square nautical miles, and spans the period and locale of intense perturbation on the fish community of this ecosystem. Changes in these key processes and rates are not trivial to detect at these spatial and temporal scales, but if observed, as they have been in this ecosystem, allow quantification of the effects of over-exploitation and more importantly provide an assessment of the potential for recovery of economically important fish stocks. This example demonstrates the value and utility of maintaining long-term, broad-scale monitoring efforts.

(57)

IS SIROLPIDIUM ZOOPHTHORUM THE ANIMAL EAT-ER ITS NAME SUGGESTS? NEW EVIDENCE OF PAR-ASITISM.

Christopher Martin

NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, Milford Laboratory, Milford, CT 06460

A phycomycetous fungus has been observed repeatedly in larvae of the bay scallop, *Argopecten irradians,* at the Milford Laboratory. This microorganism has been tentatively identified as *Sirolpidium zoophthorum* Vishniac, first observed at this laboratory by V.L. Loosanoff almost fifty years ago. The morphology and development of the fungus have been previously described. While apparently enzootic in scallop cultures at our laboratory, *S. zoophthorum* has not been tied directly to mass mortalities of this species. Loosanoff regarded it as parasitic in the bay scallop and in the larvae and juveniles of other bivalves. However, his evidence was largely circumstantial, i.e., conspicuous presence of "infected" larvae in cultures suffering high mortality.

Using pure cultures of *S. zoophthorum*, freshly isolated from affected scallop larvae, it has been possible to demonstrate that this fungus is one likely cause of observed mortality. Exposure of 72-hour scallop larvae to suspensions of recently emerged zoospores resulted in approximately 70% mortality in 4 days. Fungal thalli were detected in up to 88% of dead larvae. Untreated controls remained unaffected. *S. zoophthorum* was successfully reisolated from parasitized larvae, thus satisfying Koch's Postulates.

(58)

EFFECT OF HARMFUL ALGAE ON MARINE RE-SOURCES IN THE GULF OF MEXICO

Steve L. Morton, Kate Schaffer, Tina Mikulski, Steven Eaker, Mark Busman, Brad Mitchell, Nikki Wiggens and Peter Moeller

Marine Biotoxin Program, National Ocean Service/ NOAA, Center for Coastal Environmental Health and Biomolecular Research, 219 Ft. Johnson Rd., Charleston, SC 29412.

A number of species of microalgae commonly observed from the Gulf of Mexico produce bioactive compounds that have profound effects on marine resources. These biotoxins lead to mortality events of marine resources and human diseases after consumption of contaminated shellfish or finfish. The 1995 and 1996 manatee mortality events and the 1999 dolphin mortality event will be used as examples of biotoxins leading to loss of marine resources. Human diseases caused by toxic algae known to be found along the Gulf Coast include Neurotoxic Shellfish Poisoning (NSP) and Ciguatera Fish Poisoning (CFP). The cause of the mortality events and human intoxications will be discussed. Current research conducted by the Marine Biotoxin Program on new potential toxic algal species will also be presented.

(59)

LABORATORY CULTURE OF LARVAL TAUTOG: RECENT UPDATES AND CHANGES.

Dean M. Perry, David A. Nelson and Robin S. Katersky NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, Milford Laboratory, Milford, CT 06460

Adult field-captured tautog, Tautoga onitis, were spawned in the laboratory. Larvae were cultured according to standard laboratory procedures developed at the Milford laboratory using a recirculating system containing six 1140 L conical rearing tanks. During the summer of 2000, the following changes were made to our protocol which increased larval survival. These changes included increasing live feed density in the rearing system, extending the duration of rotifer feeding, and decreasing the initial larval stocking density. Larvae were fed an axverage of 9 x 10⁶ rotifers per day, which is double the amount fed in previous years. They were also fed rotifers for a 25d period compared to 14d in prior years. With this increased larval survival we are currently monitoring growth rates of these juveniles under laboratory conditions. From December 14, 2000 to February 2, 2001, total length (mm) and biomass were measured and recorded biweekly. Specific growth rate was 0.30 mm d⁻¹ and biomass increased at an average of 0.04 g d⁻¹.

(60)

AN ANALYSIS OF SIMILARITY COEFFICIENTS AND TAXONOMIC CLUSTERING METHODS FOR IDENTIFICATION OF BACTERIAL FISH PATHOGENS,

Steven C. Pitchford and Richard A. Robohm NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, Milford, Connecticut 06460

Reliable classification of bacterial pathogens often poses a problem for those investigating the cause of

fish disease. In the absence of sufficient DNA probes for fish pathogens, the most reliable identification method is numerical taxonomy based on bacterial phenotypic traits. A number of mathematical formulas exist for first producing similarity coefficients and then clustering organisms for construction of taxonomic trees. A systematic comparison of these approaches has not been published for cross family classification of bacteria.

We report here an evaluation of programs for similarity coefficients and clustering techniques that are available in the NTSYS-PC software package from Exeter Software (Setauket, NY). We tested 96 permutations of pairs of similarity coefficients and clustering techniques for their ability to produce correct taxonomic trees from representative bacteria (primarily fish pathogens) from six taxonomic families. Reliability was determined by looking for correct phylogenetic groupings after the programs were applied to a database of phenotypic characteristics for 28 known pathogenic species that we compiled from the literature and 30 that we isolated and identified through a consensus of five taxonomic schemes. A minimum of 40 and a maximum of 86 traits were compared between organisms depending upon the available data. A set of 10 semi-quantitative measures were established to evaluate the resulting taxonomic trees.

The most accurate permutation proved to be Kulczynski no. 2 (K2) association coefficient in combination with the Complete Linkage method. Use of this combination in conjunction with a sufficient database will help in classifying bacteria when family and genus are unknown.

(61)

DISTRIBUTION OF BDELLOVIBRIO-LIKE BACTERIA IN ALASKA'S EASTERN BERING SEA (EBS) SHELF DeLois M. Powell, Department of Natural and Physical Sciences, Shaw University, Raleigh, NC; Henry N. Williams, Ruby Singh and Andy Sneider, University of Maryland at Baltimore; and J. Frank Morado and Lisa Appesland, NOAA, National Marine Fisheries Service, Alaska Fisheries Science Center/RACE, Seattle, WA

Scientists around the world are currently involved in ongoing studies to investigate the ecology of a unique predator in the marine environment, the endoparasitic bacterium, Bdellovibrio and to note its interaction with other organisms in the marine ecosystem. Previous studies about the distribution of Bdellovibrio in Alaskan waters did not exist. This project intended to determine if Bdellovibrio or Bdellovibrio-like bacteria occur in these waters and to add isolates to the collection that is currently being characterized and compared from other parts of the world. A Nissin bottle dropped from the deck of the NOAA trawl Fishing Vessel (F/V), Aldebaran collected subsurface water (from approximately 1-meter depth) of the Eastern Bering Sea (EBS) during July 1-23, 1999. Water, mud and fish and shellfish gut samples were collected with the intent to survey the diverse habitats of the EBS and to focus on near- and offshore areas in

the Bristol Bay. A total of 30 samples were enriched for detection of Bdellovibrio-like bacteria (addition of Vibrio parahemolytica) and analyzed in the NOAA laboratory, using a modified agar overlay technique. Seven of 19 samples tested were positive for Bdellovibrio-like bacteria. Thirty-six mud-gut samples were collected from EBS and 5 mud-gut samples were collected from Larrabee State Park. Of 15 samples assayed, one proved positive for Bdellovibrio-like bacteria. Two additional mud samples collected at H184 and H195 (haul designation number) were positive for Bdellovibrio-like bacteria. A modification of the membrane filter technique used to analyze water for fecal coliforms was employed to survey water samples for the presence of possible hosts. Preliminary colonial characterization of bacterial isolates indicated a diverse population of potential host organisms.

(62)

THE TRAP POND PROGRAM: A SERVICE LEARN-ING PARTNERSHIP BETWEEN COLLEGES AND PARKS.

Michael A. Reiter, Department of Agriculture and Natural Resources, 1200 N. DuPont Hwy., Dover, DE 19901-2277; Jenna Luckenbaugh, Trap Pond State Park, RD2, Box 331, Laurel, DE 19956

The Trap Pond Program (TPP) is a service-based teaching and research relationship between the Department of Agriculture and Natural Resources of Delaware State University and Trap Pond State Park, DE. The park provides expertise and opens its lands for research and teaching programs in freshwater, wetland, terrestrial, and coastal ecology/environmental sciences, as well as interdisciplinary resource management, and in return the department works on topics relating to the management needs of the park. The university gains an extensive field site containing a wide variety of aquatic and terrestrial communities and a working relationship with the state's resource personnel. Students gain the ability to work on issues of present-day concern and access to the expertise that exists in the state's resource management agencies. At the same time, the TPP benefits the park directly and the State of Delaware indirectly by providing inexpensive manpower to study freshwater, terrestrial, and coastal questions impacting the park and its mission. The program also benefits the park and the state by encouraging students to develop an interest in the sciences while providing future employees and citizens trained in local environmental issues. The TPP is intended to be a model program for other state parks and institutions of higher learning (particularly small colleges and regional universities) both within Delaware and beyond.

(63)

SELECTION STUDIES ON GROWTH AND SURVIVAL OF BAY SCALLOPS (ARGOPECTEN IRRADIANS) FROM LONG ISLAND SOUND

Sheila Stiles, Joseph Choromanski, and Christopher Cooper

USDOC, NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, Milford, CT 06460

Selection studies are underway to investigate genetic responses for improving growth and survival of bay scallops (*Argopecten irradians*) from Long Island Sound through breeding. Previous findings from early responses to selection for growth indicated variation among several lines, with some modest gains. Several reasons were given for the range in early responses, including stage and age of scallops at the time of selection, the selection differential, inbreeding, density and other culture factors.

Subsequent measurements on the growth and survival of scallops from the same lines at a later time in development demonstrated some, but less variability in the responses among the lines. Responses in the progeny are best measured at the same stage or age as when the parents were selected. Measurements also provided more definitive information that density-dependence was a critical factor in evaluating responses. Preliminary results from breeding the F₁ generation suggest that interaction with survival may be a more important factor in selecting scallop populations than selecting for growth alone. For example, mortality was observed among the smallest and the largest scallops of some lines, thus the mean size of the survivors in the groups or sublines was similar. This could be connected with genotype-related mortality which was suggested from genetic population analyses employing allozymes. Progeny of the next generation currently in early phases of growth should further elucidate responses to selection.

(64)

POPULATION CHARACTERISTICS OF THE U.S. SOUTH ATLANTIC RED PORGY STOCK

Douglas S. Vaughan

NOAA-CCFHR, National Marine Fisheries Service, 101 Pivers Island Road, Beaufort, NC 28516

The age structure and status of the U.S. south Atlantic stock of red porgy is examined, using recorded and estimated landings and size frequencies of fish from commercial, recreational, and headboat fisheries from 1972-1997. Estimates of annual, age-specific population numbers and fishing mortality rates (F) were obtained by application of a calibrated virtual population analyses (VPA), using fishery-independent data from MARMAP hook-and-line and trap gears in the calibration procedure. Fishing mortality rates (F) increased from 0.05 in 1974 to 1.34 in 1997 for fully recruited ages (assumed 4+ throughout for comparative purposes) with M = 0.28, while spawning potential ratios declined from 90% to 32% based on mature female biomass and from 89% to 17% based on total mature (male and female) biomass. Recruits to age 1 declined from a peak of 7.6 million age-1 red porgy in 1973 to 12,000 age-1 red porgy in 1997; while total spawning stock (mature) biomass declined from a peak of 11,700 t in 1978 to 323 t in 1997. Long-term and severely declining recruitment to age 1, headboat CPUE, and MARMAP Survey CPUE raise concerns about overfishing. Generally static SPR has been at or below the South Atlantic Fishery Management Council's criteria for overfishing (SPR = 30%) since 1981. Keeping in mind the difference between thresholds and targets, it would appear that reducing F to a level at or below that equivalent to 40% static SPR is necessary for rebuilding the U.S. south Atlantic red porgy stock.

(65)

GROWTH AND SURVIVAL OF BAY SCALLOPS, AR-GOPECTEN IRRADIANS IRRADIANS, FED TETRA-SELMIS CHUI BY TWO METHODS

James C. Widman Jr. and David Veilleux

NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, Milford Laboratory, Milford, CT 06460

Tetraselmis chui was grown using two different methods, carboy vs GRAMPS (Greenhouse Algal Mass Production System) and fed to bay scallops, Argopecten irradians irradians - growth and survival were monitored. Algae grown by the traditional Milford carboy method utilize artificial light and artificial seawater with the addition of various nutrients, trace elements and vitamins. GRAMPS-grown algae utilize sunlight and natural seawater from Milford CT harbor, which is UV treated and then enriched with a commercially available F/2 media. Current results indicate a slight growth advantage when using algae grown in the carboys. Increases in mean shell height ranged from 2.6 -3.1 mm for scallops fed with GRAMPS-grown algae, while scallops fed carboy-grown algae increased 3.6 - 4.1 mm. Most surprising was the decrease in survival when scallops were fed Tetraselmis grown using the GRAMPS method. Survival of scallops on the GRAMPS-grown Tetraselmis averaged 64%, and scallops fed carboy Tetraselmis averaged 89%. Additional research is needed to determine the cause of the growth and survival discrepancies observed between the two growing methods.

(66)

PUTTING OUR TECHNOLOGY TO WORK ON THE (FISH AND SHELLFISH) FARM

Gary H. Wikfors, Barry C. Smith, Jennifer H. Alix, and Mark S. Dixon, NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, Milford Laboratory, Milford, CT 06460

As part of our technology-transfer activities, we have organized an annual training session titled, "Growing Microalgal Feeds for Marine Aquaculture -- The Milford Workshop." Microalgae are cultured as feeds for molluscan shellfish (scallops, clams, and oysters) and as part of the live-food chain for marine finfish. Practical knowledge and skills required to culture microalgae successfully are diverse, ranging from microbiology to ecology, chemistry, engineering, physics, and hard-knocks lessons. The purpose of this annual Workshop is to organize and condense the diverse, practical information and skills needed to culture microalgae in a hatchery setting into a three-

day crash-course integrating both lecture and hands-on laboratory sessions. The Workshop is offered at no cost, as part of the Milford Laboratory's ongoing efforts to support development of US aquaculture, with first preference given to commercial growers and extension personnel. Enrollment is limited to 16. The 2000 class included participants from commercial shellfish farms, state extension programs, academic research laboratories, and Native American Tribal groups on both Atlantic and Pacific coasts of the US.

(67)

ENHANCING COASTAL MANAGEMENT THROUGH GEOSPATIAL ANALYSES.

Pace Wilber

NOAA Coastal Services Center, 2234 Hobson Avenue, Charleston, SC 29405

Working with partners from academia, non-governmental organizations, and state/local governments, the NOAA Coastal Services Center fosters the application of geographic information systems (GIS) and remote sensing in coastal management. Our remote sensing activities work with data streams from satellite, airborne, and in-situ sources to identify new or under-utilized remote sensing activities, such as LIDAR and acoustic sensors, that could assist coastal managers. GIS activities cover a broad range and include "rescuing" geospatial data from non-digital formats and customizing GIS applications to more effectively integrate remote sensing and other data to aid decisions in the coastal and marine environment. Distributing tools for developing metadata, providing training in development of metadata, and hosting applications that allow simultaneous searching of multiple geospatial archives fosters access to geospatial data and technologies. Through fellowships, internships, cooperative agreements, and traditional classroom settings, the Center also provides handon training in the use of these technologies and their practical application to coastal management issues.

ENVIRONMENTAL SCIENCES

(68)

EFFECTIVE MOLECULAR MECHANISMS TO IDEN-TIFY MICROORGANISMS RESISTANT TO TOXIC METALS IN THE ENVIRONMENT

Raul Cuero, John Williams, Nnana Mogongwa, Felicia King, Alonso Sadberry, and Ashley Chatham,

Prairie View A&M University, PO BOX 685, Prairie View, Texas 77446

Although many metals including Zn^{+2} , Cu^{+2} , Fe^{+2} and others, are essential to microbial metabolism, all are potentially toxic. However, microorganisms such as bacteria and fungi can develop mechanisms to accumulate these metals, thus making the microorganisms and/or

their metabolic products ideal for metal recovery and/or environmental protection. Metals affect microbial metabolisms, enzymatic activity, membrane transport, and nucleic acid synthesis. Therefore, the aim of the present investigation, was to use fungal molecular structures and products such as RNA molecule and enzyme chitosanase, respectively, in relation to microbial growth, under different concentrations of toxic metals, thus identifying strains resistant to toxic metals that can be used for further decontamination of these toxic metals in the environment. Different fungal isolates and a bacteria strain were tested during the study. However, Aspergillus flavus, and Fusarium graminearum, were mainly used in every experiment. The fungal isolates were sequentially grown in agar media, then inoculated in sterile soil media containing sterile ground corn, and finally transferred to broth media. All the media contained 0-5 ppm concentrations of toxic metal ,including Zn⁺², Cu⁺², Fe⁺². Some cultures were amended with the natural polysaccharide chitosan to induce chitosanase production. The toxic metal-amended fungal cultures were incubated in replicates at room temperature. Agar and broth media were incubated for 3-6 days, while soil cultures were incubated 3-12 months. Soil cultures were always used as the primary inoculum for the sequential inoculation of the agar and broth media. Soil and liquid cultures were subjected to analysis for fungal growth; only liquid cultures were used for RNA yield (Cuero et al., 2000), while chitosanase production was assessed in chitosanamended agar, for which a qualitative assay is reported based on an arbitrary scale (- = little enzyme production; ++ = medium enzyme production; +++ = abundant enzyme production; and ++++ = very abundant enzyme production) (Cuero, 1996). The results show that both copper and zinc either in single or combined treatment stimulated fungal growth 10 times (up to 0.45 gr. biomass) in liquid cultures. Similarly, fungal populations increased 4-12 times in soils treated with the metal ions (up to 1x10⁶ CFU) as compared to non-treated fungal cultures, after 3 months. Stimulation of fungal growth corresponded with an increase in RNA yield and concentration, depending upon the fungal species. Growth and RNA production by A. flavus were stimulated more by zinc, while F. graminearum was stimulated more by copper. RNA and growth of the bacteria Bacillus spp., were markedly increased by zinc. Chitosanase production was also higher in fungal cultures amended with chitosan and toxic metals. The present results show an effective molecular approach to use RNA and/or enzymes to identify microorganisms with potential for removal of toxic metals from the environment.

(69)

PETROLEUM HYDROCARBONS IN SEDIMENTS AND RIBBED MUSSELS FROM RESTORED, UNRE-STORED, AND REFERENCE MARSHES IN THE ARTHUR KILL, NEW YORK / NEW JERSEY, USA. Ashok D. Deshpande, Bruce W. Dockum, and Amy M. Tesolin NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center. James J. Howard Marine Sciences Laboratory at Sandy Hook, 74 Magruder Road, Highlands, New Jersey 07732

An underwater pipeline ruptured during January 1990, discharging about 567,000 gallons of No. 2 fuel oil into the Arthur Kill. The City of New York Parks and Recreation Department restored 2.6 hectares of the impacted marsh with Spartina alterniflora. We collected sediment and ribbed mussel samples from restored, unrestored, and reference sites over two collection periods to assess the functional effectiveness of the restoration efforts with respect to total petroleum hydrocarbons (TPHs). TPHs in sediments varied within and between sites, and within sections of the individual cores, with generally higher concentrations found in the deeper sections. TPHs in surface sediments ranged from less than the method detection limit (MDL, 181 mg/g wet weight) to 490 mg/g wet weight for a restored site, less than MDL to 7890 mg/g wet weight for an unrestored site, and less than MDL for a reference site. TPHs in ribbed mussels also varied within and between sites, with a range of less than MDL (54 mg/g wet weight) to 540 mg/g wet weight. There was no correlation between TPHs in mussels and TPHs in sediments. Another spill of about 50,000 gallons of No. 2 fuel oil occurred at one of the reference sites shortly after the last collection period. We collected additional samples of sediment and ribbed mussels at this site to study the petroleum hydrocarbon patterns before and after the spill. Hydrocarbon fingerprints, double ratio plots, and principal component analyses were used to examine if sediments and ribbed mussels were contaminated with the spilled oil.

(70)

HIGH PERFORMANCE UV (HPUV) INACTIVATION OF E. COLI.

Kurt A. Garrett, Cheryl M. Davis and De Lois M. Powell Department of Natural and Physical Sciences, Shaw University, Raleigh, NC

The use of ultraviolet light for inactivation of microorganisms is well documented in the literature. Ultraviolet light used for inactivation is sometimes called germicidal UV or UV254 (the specific wavelength known to have the most lethal effect on bacteria, fungi and viruses). While this band of invisible light is part of the sun's spectral energy, artificial sources of germicidal UV come from manufacturers such as Sylvannia and General Electric. The greatest users of germicidal UV lamps are wastewater treatment facilities seeking alternative methods of disinfection that reduce the need for chlorine and chlorine derivatives. The disinfection mechanism generally points to the disruption of the microorganism's DNA. In turn, the disruption of microbial DNA prevents replication and can cause 'death'. Early UV light inactivation research was set back when conventional ultraviolet lamps performed below expectations. This failure was traced to lamp design. While researchers sought

intense high-energy germicidal UV light, manufacturers designed low-energy UV lamps, primarily used for wastewater disinfection. In research typical inactivation times, using conventional germicidal UV lamps, range from 20 minutes to several days for virus and similar times for bacteria. Recently, we tested a new lamp system, High Performance Ultraviolet Light (HPUV; CSMO, Inc.) and found it to reduce 10⁶ colonies of herpes simplex virus (HSV) in three seconds. We are proposing use of the HPUV for inactivation of E. coli. Our studies will measure inactivation initiation, inactivation optimum and hyper-inactivation. HPUV is expected to result in guicker inactivation times for E. coli than conventional UV lamps. This data will allow us to construct a response curve for E. coli when using HPUV inactivation. Future research will use these data to determine inactivation conditions that preserve important protein triggers of the cellular immune response.

(71)

RESTORATION AND ASSESSMENT OF URBAN SALT MARSH HABITAT DAMAGED BY A SEVERE OIL SPILL

David Packer and Joe Vitaliano, NOAA, National Marine Fisheries Service, Northeast Fisheries Science Center, James J. Howard Marine Sciences Laboratory, Highlands, NJ 07732; and Carl Alderson, Salt Marsh Restoration Team, Natural Resources Group, New York City Parks, 200 Nevada Ave., Staten Island, NY 10306

Few scientific studies have focused on restored salt marshes (restored because of a severe environmental impact) as opposed to created or constructed marshes (created in response to mitigation). In 1990, a 576,000 gallon oil spill seriously damaged marshes of the Arthur Kill, the strait separating Staten Island, NY from NJ. The Salt Marsh Restoration Team of NYC Parks implemented a multi-year restoration and monitoring project to restore marshes directly impacted by the 1990 spill. To date, restoration activities included the successful reintroduction of over 9 acres of Arthur Kill-propagated salt marsh cordgrass, Spartina alterniflora. SMRT has been monitoring several parameters in oiled marshes that were replanted and oiled marshes left for natural recovery, including Spartina biomass/density, ribbed mussel (Geukensia demissa) density, fish abundance/diversity, frequency/duration of feeding of wading birds, and sediment total petroleum hydrocarbons (TPH) in replanted and unplanted sites. In 1996 the National Marine Fisheries Service?s James J. Howard Marine Sciences Laboratory extended the study by characterizing and assessing the structure and function of replanted, unplanted, and reference marshes. Studies included sediment chemistry, contaminant analyses, macrobenthic surveys, and stomach content analyses of mummichogs (Fundulus heteroclitus). Results are presented from the two investigations. A quantitative assessment of these marshes may allows us to evaluate our ability to restore this habitat?s functional attributes, and identify indicators of habitat and living resource health and recovery within a heavily urbanized and degraded estuary.

(72) THE EVOLUTION OF MTBE AS A POLLUTANT

Pao-Chiang Yuan, Doris McPherson and Youguo Xu, Hazardous Materials Management Program, Department of Technology, School of Science and Technology, Jackson State University, Jackson, Mississippi

We have learned a lot of lessons from the past. The famous materials we've used in the past are asbestos, PCB (polychlorinated biphenyl) transformer oils, freon, and lead in paint and also in gasoline. They were wonderful useful materials that served their purposes for us until we found they have harmful health problems. Most of them cause chronic diseases and irreversible harm. Some materials have been used for thousands of years, such as asbestos, until two to three decades ago, when we stopped using it in this country, it is still being used by other countries. For most of these toxic materials, their substitutes have been found and they have been totally exclude from the United States market. MTBE is used as a fuel additive mixed into gasoline starting from about a decade ago, due to the Clean Air Act Amendment (CAAA) that required improvement in the air quality in some parts of the country, until scientist found MTBE in our water and ground water supply. This is a case of our efforts to control air pollution now threatening water quality. MTBE is a possible human carcinogen according to the US EPA (United States Environmental Protection Agency).

ATMOSPHERIC SCIENCES

(73)

HPVCI - DISTANCE LEARNING

Paul Croft, Rafael Mahecha, R. Suseela Reddy, and Pat Fitzpatrick, Meteorology Department, Jackson State University, Jackson, MS

As a part of the High Performance Visualization Center Initiative (HPVCI) at Jackson State University, the JSU Meteorology Program (JSU MP) is developing a Distance Learning Initiative involving mesoscale modeling and products (MM5 and COAMPS) for college and professional users. Through computer visualizations of mesoscale model data and results, and their presentation and use in the classroom, both students and professionals will have the opportunity to learn and examine the nature of numerical weather prediction models. Visualization products will be available to the broader atmospheric and minority communities for education and training, particularly in an electronic classroom. Assisted directly by several undergraduates, three websites, including a developmental site(http://www.angelfire.com/ ms2/hpvci/index.html), the JSU Meteorology Program's homepage (http://weather.jsums.edu), and a research platform site http://betsy.jsums.edu/~hafner/oper5.html) are in development to address the needs of users. Ultimately all three sites will be linked and/or merged. Numerical model guidance and their output will be used to

create class materials. Both models (MM5 and CO-AMPS), run in real-time, and will be used to explain the reasons for weather modeling, the need to do so, and the methodological approaches used. A study through beta testing is planned. Various target audiences will be selected to consider general and specific skills and knowledge that different users may, or may not have, to define their goals when visiting the websites. Users with various academic backgrounds, and levels of meteorological skills will be introduced to the sites, asked questions about it (e.g., how easy is it to navigate the site? Is labeling clear?), and asked for comments. Before final release, the sites should have a layout, information, products, and labels that are clear and easy to use (without losing technical quality) for all audiences. Suggested improvements to the sites include further explanation of products, less crowded maps, and «make-your-own» overlays. Upon completion the Jackson State University Meteorology Program's HPVCI websites will serve as a clearinghouse for model output and products.

(74)

A NEW U.S. CLIMATE ATLAS

Marc S. Plantico, Lynn A. Goss, Liz Love-Brotak, Gregory Hammer, National Climatic Data Center, Scientific Services Division, Asheville, NC 28801; Christopher Daly, Spatial Climate Analysis Service, Oregon State University, Corvallis, OR 97333; George Taylor, Oregon Climate Service, Oregon State University, Corvallis, OR 97333

The National Climatic Data Center (NCDC) has developed a new, electronic Climate Atlas of the United States which is available on CD-ROM. The purpose of this Atlas is to depict the climate of the United States in terms of the distribution and variation of major climatic elements. The Climate Atlas is intended to meet the needs for climatic information from commercial, industrial, agricultural, research, and educational institutions, as well as from the general public. The new atlas replaces the previous Climatic Atlas of the United States which was published in 1968. Data used for the old publication were from the period 1931-1960 and the nearly 200 maps were hand contoured. The new atlas used innovative technology and easy to use Geographic Information Systems (GIS) to objectively generate many of the maps. The analytical model, PRISM (Parameter-elevation Regressions on Independent Slopes Model) which was developed at the Oregon Climate Service (OCS) at Oregon State University, was used to generate nearly 350 different map products. An additional 350 maps were generated at NCDC. Most atlas products are derived from the 1961-1990 period of record. This poster illustrates some of the map products that are contained in the new atlas.

(75)

SOME ASPECTS OF AIR-SEA INTERACTIONS, HUR-RICANE PREDICTIVE MODELS AND VARIABILITY OF TROPICAL CYCLONES

R. Suseela Reddy, Jackson State University, Jackson, MS, and Richard L. Miller, Earth System Science, NASA Stennis Space Center

Under the NASA research grants, we present the work established on (a) the air-sea interactions and associated tropical cyclones during their formation and evolution over the Gulf of Mexico, (b) the development of predictive models for hurricane activity, and (c) the variability of tropical cyclones over the North Atlantic. The studies suggested that, (a) a 3-5 day oscillation was noticed in the heat and momentum fluxes during the formation and evolution of hurricane activity, (b) Hurricane Predictive Index (HPI) was used for hurricane prediction, and (c) a long-term ~ 50-year oscillation was noticed in the tropical cyclone activity over the North Atlantic Ocean.

(76)

MESOSCALE CIRCULATION OVER THE COMPLEX PHYSIOGRAPHY INLOUISIANA-MISSISSIPPI (LA-MS) COASTAL REGION.

R. Suseela Reddy, Paul Croft and Pat Fitzpatrick, Jackson State University, Jackson, MS

Mesoscale air flow in coastal regions is determined by land-sea temperature contrast (land-sea breeze) and by the shape of a coastline. In addition to topography, shape of coastline has an effect on mesoscale wind flow, and thus, on various meteorological variables including cloudiness and air pollution. We present in this study an understanding of the mesoscale circulation in LA-MS coastal region and in particular the combined effect of the lake and shape of the coastline. The Penn State/ NCAR Mesoscale Model (MM5 version 3.1) was used with several options including multiple nesting and several convective and boundary layer parameterizations, as a modeling tool to investigate mesoscale wind flow patterns in the study area. Two cases of mesoscale circulation during the summer of 1999 were selected for numerical simulations (24 July and 17 August). The study suggested that the MM5 model is suitable for numerical studies of sea-breeze circulations and it is capable to simulate diurnal variations of wind patterns and convective cloudiness. Results will be presented and discussed.

(77)

TECHNOLOGICAL ADVANCES AT THE LOWER MIS-SISSIPPI RIVER FORECAST CENTER USING GIS TECHNOLOGY

Keith Stellman , Amanda Roberts, and Dave Reed NOAA, National Weather Service, Lower Mississippi River Forecast Center, 62300 Airport Road, Slidell, LA 70460

The National Weather Service (NWS) is responsible for supporting the nation's welfare and economy by

issuing river and flood forecasts and warnings. To do this, the NWS has 13 River Forecast Centers (RFC) in the United States. The Lower Mississippi River Forecast Center (LMRFC), located in Slidell, LA, is responsible for issuing routine river and flood forecasts at over 170 locations daily and over 30 locations when rivers are expected to exceed flood stage at these locations. The NWS uses hydrologic computer models on stateof-the-art technology to prepare these forecasts.

The LMRFC collects, processes, and analyses a large amount of data. To keep its users informed, the LMRFC prepares a significant number of products using ArcView. These products are used internally at LM-RFC and within the NWS and may be posted to the LMRFC webpage. Products prepared using ArcView include: precipitation estimates from the NWS WSR-88D Radar for time periods ranging from hourly to yearly; contoured observed precipitation for time periods ranging from daily to yearly; monthly normal precipitation maps and departure from normal; forecasted precipitation in 6-hour increments for 24 hours in the future including gridded model output; and maps displaying the status of river forecast locations to name a few. We will demonstrate this technology and how it is integrated into LMRFC products and services.

(78)

REGIONAL CLIMATE MODELING IN THE SOUTH-EASTERN UNITED STATES

Loren D. White

Jackson State University, Jackson, MS

Besides their application to long-term large-scale issues such as global warming, numerical «climate models» are increasingly being developed to study and predict anomalous conditions on seasonal and interannual timescales. A complex regional climate model developed at Florida State University (the FSU Nested Regional Spectral Model) has been used at Jackson State University to examine issues of climate predictability and forcing over the Southeastern U.S. Most emphasis has been on the interrelationship between surface forcing from the ocean (e.g. El Nino) and from the land surface (e.g. soil moisture). Sensitivity of model results to specifics of the domain choice was also investigated in one set of winter season experiments. A unique approach to «ensemble» model forecasts has been explored by variation in the details of the initial soil moisture and temperature conditions. This technique provides a useful means of determining the degree to which model forecast uncertainties over specific regions may be attributable to uncertainties in the initial conditions of the land surface. Finally, experiments which vary the number of vegetation types modeled within the domain will be discussed. Especially at high horizontal resolutions, the question of how many vegetation types «need» to be recognized by a climate model is currently unknown and can most likely be resolved by many numerical experiments of this type.

Ashley Abraham Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Sunnie Aburime Clark Atlanta University Associate Professor 223 James P. Brawley Dr., SW Atlanta, GA 30314 Telephone: (404)880-6776 Fax: (404)880-8267 Email: saburime@cau.edu

John S. Adams Texas A&M University Field Research Specialist 6300 Ocean Drive Corpus Christi, TX 78412 Telephone: (361)825-5759 Fax: (361)825-5704 Email: jadams@cbi.tamucc.edu

Cinyere Agwaramgbo Tougaloo College Student P. O. Box 598 Tougaloo, MS 39174 Telephone: (601)362-4774 Fax: (601)362-6773 Email: ramgbo@bellsouth.net

Lovell Agwaramgbo Tougaloo College Professor of Chemistry 500 W. County Line Road Tougaloo, MS 39174 Telephone: (601)977-7785 Fax: (601)977-6142 Email: lovell.agwaramgbo@tougaloo.edu

Justin Ahanhanzo UNESCO/IOC Consultant, Oceanographer 1 Rue Miollis Paris, France 75015 Telephone: 33-1 45 68 36 41 Fax: 33-1 45 68 58 10/12 Email: j.ahanhanzo@unesco.org

Hinda Ahmed Tougaloo College Student 1043 Lakeshore Drive Grandon, MS 39047 Telephone: (601)992-6749 Fax: (601)977-6142 Email: hinda21@hotmail.com

Frantz Alcindor ETAP Program Program Administrator 316 Washington Avenue Telephone: (304)243-2535 Fax: (304)243-4392 Email: rkeane@nttc.edu Kawana Alexander Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Yoseph Ali Jackson State University Student Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: alibaba555@yahoo.com

Zeyad Ali Jackson State University Research Associate P.O. Box 18839 Jackson, MS 39217 Telephone: 601-979-4058 Fax: 601-979-3716

Arthur Allen University of Maryland Eastern Shore Associate Research Director/Associate Professor Carver Hall, 1126 Princess Anne, MD 21853 Telephone: (401)651-6254 Fax: (401)651-7739 Email: alallen@mail.umes.edu

Sheila Allen Government Geospatial Analyst 6000 McArthur Blvd. Bethesda, MD 30816 Telephone: (301)227-7882 Fax: (301)227-4388

Yolanda Amadeo WALB-TV 1709 Stuart Avenue Albany, GA 31707 Telephone: (229)446-4089 Fax: (229)446-4061 Email: yolandaa@walb.com

Melvin Anderson Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: newsont@hotmail.com

Randel Anderson Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718 Shawn Arellano WHOI Minority Fellow 27B Challenger Drive Woods Hole, MA 02543 Telephone: (508)289-3575 Email: basfimoo@hotmail.com

James H. Arrington South Carolina State University Vice President for Academic Affairs P. O. Box 7307 300 College Street, NE Orangeburg, SC Telephone: (803)536-7180 Fax: (803)533-3775 Email: ZF_Arrington@scsu.edu

Senayt Assefa American Assocation for the Advancement of Science 1200 New York Ave., NW Washington, DC 20005 Telephone: (202)326-7019 Fax: (202)321-9849 Email: sassefa@aaa.org

Julius M. Baham JSU/CSDRA Co-Director P.O. Box 18739 Jackson, MS 39217 Telephone: 601-979-2989 Fax: 601-979-2478 Email: julius@stooges.jsums.edu

Ana Balarezo Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Jennifer Banks Jackson State University System Administrator P.O. Box 18839 Jackson, MS 39217 Telephone: 601-979-4058 Fax: 601-979-3716

Beneia Barnes Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Eugene N. Baskerville, Jr. Shaw University Professor of Biology DNPS 118 East South Street Raleigh, NC 27601 Telephone: (919)546-8216 Fax: (919)546-8258

Gregorio B. Begonia Jackson State University Associate Professor, Interim Chair Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-2586 Fax: (601)979-5853

Maria Begonia Jackson State University Assistant Professor P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3469 Fax: (601)979-5853

Larry Belton Jackson State University Director, Marketing & Recruitment Jackson, MS 39217 Telephone: (601)979-2911 Fax: (601)979-5843

Jocelyn Bennett-Martin National Ocean Service EEO/MSI Program Manager 1305 East-West Hwy SSMC4, Room 13636 Silver Spring, MD Telephone: (301)713-3074 Fax: (301)713-4269 Email: Jocelyn.Martin@noaa.gov

Eugene J. Billiot Texas A&M University Assistant Professor 6300 Ocean Drive Corpus Christi, TX 78412 Telephone: (361)825-2680 Fax: (361)825-2742 Email: ebilliot@falcon.tamucc.edu

Brian Bingham Associate Professor 1900 Shannon Point Road Anacortes, WA 98221 Telephone: (360)293-2188 Fax: (360)293-1083 Email: bingham@cc.wwu.edu

Gleyn E. Bledsoe Northwest Indian College Dean for Land Grant & Outreach 2522 Kwina Road Gellingham, WA 98226 Telephone: (360)676-2772 ext 4310 Fax: (306)738-0136 Email: gleyn@nwic.edu

Reed Bohne NOAA Grays Reef National Marine Sanctuary Manager 10 Ocean Science Circle Savannah, GA 31411 Telephone: (912)5982345 Fax: (912)598-2367 Email: Reed.Bohne@noaa.gov William Bonner UCAR Senior Research Associate 3855 Birchwood Drive Boulder, CO 80304 Telephone: (303)442-2509 Fax: (303)443-7997 Email: bbonner@attglobal.net

Kandis Boyd National Weather Service Hydrologist 10159 East 11th Street Telephone: (918)832-4109 Fax: (918)832-4101 Email: kandis.boyd@noaa.gov

Ronald Bozeman Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Jill M. Brandenberger Texas A&M University Graduate Student 4822 Edith Street Corpus Christi, TX 78411 Telephone: (361)825-3088 Email: jmbrand@cbi.tamucc.edu

Eddie Brassell Lockheed Martin 700 N. Frederick Avenue Gaithersburg, MO 20879 Telephone: (301)240-5829 Fax: (301)240-6176

Atanasio Brito Florida A&M University, ESI ESI Graduate Student 1520 S. Bronough St. Tallahassee, FL 32307 Telephone: 850-599-8193 Fax: 850-561-2248

Kenneth Brookins Jackson State University Student Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3461 Fax: (601)979-5853

George B. Brooks, Jr. Gila River Indian Community Environmental Coordinator 5012 S. 21st Way Phoenix, Arizona 85040 Telephone: 520-562-6706 Fax: 520-562-6796 Email: gbbrooks@gilariver.com

Sandra J. Brooks Immigration and Naturalization Service 425 I Street, NW,Room 1122 Washington, D.C. 20536 Telephone: (202)514-1246 Fax: (202)514-9866 Email: m.kathleen.lane@usdoj.gov

Brad Brown NOAA Fisheries Senior Advisor 75 Virginia Beach Drive Miami, FL 33149 Telephone: 305-361-4284 Fax: 305-361-4219 Email: brad.brown@noaa.gov

Eula M. Brown DOC/NOAA Civil Rights EEO Manager 1305 East/West Highway Silver Spring, MD Telephone: (301)713-0500 x141 Fax: (301)713-0983 Email: eula.m.brown@noaa.gov

Laurita Brown Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: labr2003@stallion.jsums.edu

Nick Brown National Imagery and Mapping Agency 6000 MacArthur Blvd. Bethesda, MD 20816-5003 Telephone: (301)227-7786 Fax: (301)227-4388

Willie Brown Jackson State University VP of Information Technology P.O. Box 17750 Jackson, MS 39217 Telephone: 601-979-4330 Fax: 601-979-4301 Email: wbrown@ccaix.jsums.edu

Ashok Budamgunta Jackson State University System Administrator P.O. Box 18180 Jackson, MS 39217 Telephone: 979-4230 Fax: 979-3310 Email: ashok@tiger.jsums.edu

George P. Burbanck Hampton University Chair, Marine & Environmental Science Department Marine & Environmental Science Department Hampton, VA Telephone: (757)727-5752 Fax: (757)727-5740 Email: george.burbanck@hamptonu.edu

Oiana Cage Jackson State University Research Asst.

P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Robert Cage, Jr. National Data Buoy Center Logistics Management Specialist NDBC, NOAA John Stennis Space Center Stennis Space Cenger, MS 39529 Telephone: (228)688-7431 Fax: (228)688-3153 Email: bob.cage@noaa.gov

Peter T. Capozzoli U. S. Army Cecom Small Business Specialist Angel-SB SADBUO Office Fort Monmouth, NJ 07703 Telephone: (732)532-6999 Fax: (732)532-8732 Email: Peter.Capozzoli@mail1.monmouth.army.mil

Jen Carfagno The Weather Channel Meteorology Data and Product Leader 300 Interstate North Parkway Atlanta, GA 30339 Telephone: 770-226-2086 Fax: 770-226-2951 Email: jcarfagno@weather.com

Sheree Carney Tougaloo College Student Berkshire Cottage 500 W. County Line Road Tougaloo, MS Telephone: (601)978-7509 Email: msree81@hotmail.com

Nikeya Carter Mississippi Valley State University Student Department of Natural Sciences Itta Bena, MS Telephone: (662)254-3377 Fax: (662)254-3668 Email: wahome@mvsu.edu

Milton Chambliss Mississippi Space Commerce Initiative Commercial Operations Manager Building 1103, Room 143 Stennis Space Center, MS 39529 Telephone: (228)688-1936 Fax: (228)688-2629 Email: : mchambliss@spacecommerce.com

Ashley Chatham Prairie View A&M University Student P. O. Box 2881 Prairie View, TX 77446 Telephone: (936)857-2380 Email: ashleychatham@hotmail.com Paulinus Chigbu Jackson State University Director, Marine Science Program Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3461 Fax: (601)979-5853 Email: pchigbu@stallion.jsums.edu

Addie Citchens Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Kelton Clark Morgan State University 7959 Telegraph Road Severn, MD 21144 Telephone: (410)482-2440 Email: Keltonclark@hotmail.umd.edu

Robert Clemmons Mississippi Valley State university Student Dept. of Natural Sciences, 14000 HWY 82 W Itta Bena, MS 38941 Telephone: 662-254-3378 Fax: 662-254-3668

Jai Cole University of Maryland Eastern Shore Graduate Student Carver Hall, Room 1126 Princess Anne, MD 21853 Telephone: (410)651-6254 Fax: (410)651-7739 Email: jecole@mail.umes.edu

Eron Coleman Jackson State University Student Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-8062 Fax: (601)979-5853

Kimberly Coleman Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: kcol2003@stallion.jsums.edu

Novella M. Coleman Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu Willie Coleman Lockheed Martin 700 N. Frederick Avenue Gaithersburg, MO 20879 Telephone: (301)240-5829 Fax: (301)240-6176

Andrea Cook Mississippi Valley State university Student Dept. of Natural Sciences 14000 Hwy 82 W Itta Bena, MS 38941 Telephone: 662-254-3378 Fax: 662-254-3668

Barry Costa-Pierce Mississippi-Alabama Sea Grant Consortium P. O. Box 7000 Ocean Springs, MS 39566-7000 Telephone: (228)875-9368 Fax: (228)875-0528 Email: b.costapierce@usm.edu

Celdrick Course Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Terrence B. Course USDA Technician-Genetics 4254 Richmond Circle Jackson, MS 39209 Telephone: (504)209-6626 Fax: Email: tcourse@srrc.ars.usda.gov

Consuela Y. Cowan DOC/NOAA/NMFS Computer Specialist P. O. Drawer 1207 Pascagoula, MS 39568-1207 Telephone: (228)762-4591 Fax: (228)769-9200 Email: ccowan@triton.pas.nmfs.gov

Quentin Cowans Clark Atlanta University Research student 223 James P. Brawley Dr, Atlanta, GA 30314 Telephone: 404-880-6905 Fax: 404-880-8499 Email: qcowans@msn.com

Tyrus Cox US Army Corps of Engineers - Vicksburg Dist. 4155 Clay Street Vicksburg, MS 39183 Telephone: (601)631-5560 Fax: (601)631-5583 Email: Mike.A.Turner@mvk02.usace.army.mil

Paul Croft Jackson State University Asst. Professor P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-8261 Fax: 601-979-8623 Email: pcroft@stallion.jsums.edu

Bobby Crudup Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Paul Cuero Prairie View A&M University Research Scientist P. O. Box 685 Prairie View, TX 77446 Telephone: (936)857-3012 Fax: (936)857-2325 Email: olimpa@aol.com

De Lois A. Cutter Tal-Cut Company President/Owner One Research Blvd., Ste. 204 Starkville, MS 39759 Telephone: (800)591-0026 Fax: (440)716-3267 Email: talcut@ix.netcom.com

William Daniels Delaware State University Research Associate Professor 1200 N. Dupont Hwy Dover, DE 19901 Telephone: (302)857-6436 Fax: (302)857-6455 Email: wdaniels@dsc.edu

Kenneth Davidson NOAA/NESDIS/NCDC Deputy Director, NCDC 151 Patton Avenue, Room 120 Telephone: (828)271-4848 Fax: (828)271-4246 Email: kenneth.d.davidson@noaa.gov

Charles Davis Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Cheryl Mercedes Davis Shaw University Student 118 East South Street Raleigh, NC 27601 Telephone: (919)546-8595 Fax: (919)546-8258

Andre Debose DOC/NOAA/NMFS Res. Fishery Biologist P. O. Drawer 1207 Pascagoula, MS 39568-1207 Telephone: (228)762-4591 Fax: (228)769-9200 Email: adebose@triton.pas.nmfs.gov

Adella Dexter Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: astroweather@madmail.com

Erica Dillon Jackson State University Student P. O. Box 18149 Jackson, MS 39217 Telephone: (601)979-3473 Fax: (601)979-5853

Sontyna Dixon Jackson State University Meteorology Student Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630

Wanenne Dorsey Jackson State University Environmental Science Ph.D. Student 2005 MLK Drive Grambling, LA 71245 Telephone: (318)618-9648 Fax: (318)274-6041 Email: dorseywc@alpha0.gram.edu

Joyce Douglas Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Carl T. Drake Jackson State University Instructor of Physics Box 17660 Jackson, MS 39217 Telephone: 601-979-3624 Fax: 601-979-3630 Email: cdrake@ccaix.jsums.edu

Essie Coleman Duffie NMFS/Southeast Fisheries Center Fishery Management Specialist 75 Virginia Beach Drive Miami, FL 33149 Telephone: (305)361-4237 Fax: (305)361-4219 Email: Essie.C.Duffie@noaa.gov

Michelle Duncan

Savannah State University Savannah, GA 31411

Patricia Duncan Jackson State University Research Associate Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3461 Fax: (601)979-5853

Reshuna Durden Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Patricia Durrant U. S. Environmental Protection Agency HBCU Coordinator 1200 Pennsylvania Ave., NW Mail Code 1230A Washington, DC 20460 Telephone: (202)564-4738 Fax: (202)501-0756

Chraisma Edwards Clark Atlanta University Research Student 223 James P. Brawely Drive Telephone: 404-880-6776 Fax: 404-880-8499 Email: chrisma_d_e@hotmail.com

Ekhuemuenogiemwen Ekunwe Jackson State University Telecommunication Specialist P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2697 Fax: 601-979-2007

Dan Elliott West Valley Nuclear Services Co. Sr. Human Resources Rep. 10282 Rock Springs Road West Valley, NU 14171 Telephone: (716)942-2812 Fax: (716)942-4096 Email: elliots@wvnsco.com

Sara A. Elliott West Valley Nuclear Services Co. Sr. Human Resources Rep. 10282 Rock Springs Road West Valley, NU 14171 Telephone: (716)942-2812 Fax: (716)942-4096 Email: elliots@wvnsco.com

Tenille A. Epperson Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu Felecia L. Evans Pennsylvania State University Student 716 'Wolf Hall University Park, PA 16802 Telephone: (814)862-7496 Fax. Email: angel12896 99@yahoo.com Ruth Ewing DOC, NOAA Fisheries Veterinary Medical Officer 75 Virginia Beach Dr. Miami, FL 33149 Telephone: 305-361-4233 Fax: 305-365-4102 Email: ruth.ewing@noaa.gov Ibrahim Farah Jackson State University Professor Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3321 Fax: (601)979-5853 Email: John W. Farrington Woods Hole Oceanography Dean MS #31, 360 Woods Hole Woods, Hole MA Telephone: (508)289-2200 Fax: (508)457-2188 Email: jfarrington@whoi.edu Michelle Farver Jackson State University Student Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (t601)979-7012 Fax: (601)979-3630 Email: mfarver@twister.jsums.edu Mack Felton, Jr. **Dillard University** Professor of Biology 7210 Farwood Drive New Orleans, LA 70126 Telephone: (504)241-1626 Fax: (504)241-7457 Email: fnormack@mindpring.com Norma D. Felton University of New Orleans Co-Director/Site Coordinator 7210 Farwood Drive New Orleans, LA 70126 Telephone: (504)241-1626 Fax: (504)241-7457 Email: fnormack@mindspring.com Evelyn J. Fields

U. S. Department of Commerce/NOAA Director, Office of Marine & Aviation Operations 1315 East West Highway, Room 12837 Silver Spring, MD 20910 Telephone: (301)713-1045 Fax: (301)713-1541 Email: evelyn.fields@noaa.gov

Tomas Figueroa, Jr. Pennsylvania State University Student 141 Hamilton Hall Telephone: (814)862-6712 Email: tjf158@psu.edu

Pat Fitzpatrick Jackson State University Assistant Professor of Meteorology PO Box 17660 Jackson, MS 39217 Telephone: 6019793617 Fax: 6019793630 Email: fitz@twister.jsums.edu

Melinda Floyd JSU Department of Urban & Regional Planning Recruiter 3825 Ridgewood Road, Box 23 Jackson, MS 39211 Telephone: (662)432-6853 Fax: (662)432-6862 Email: mfloyd@mail1.jsums.edu

Michael Fogarty NOAA 162 Water Street Woods Hole, MA 02543 Telephone: (508)495-2318 Fax: (508)495-2004 Email: ambrose.jearld@noaa.gov

Hanan Forawi Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Gareth Forde Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Cathy Fore BWXT Y-12 Manager, MEITP P. O. Box 2009 Telephone: (865)576-6838 Fax: (865)576-8750 Email: forecs@y12.doe.gov

Michael Foster University of Maryland Eastern Shore Graduate Teaching Assistant Carver Hall, 1126 Princess Anne, MD 21853 Telephone: (410)651-6254 Fax: (410)651-7739 Email: michaeljfoster@hotmail.com Chandra Franklin Savannah State University Associate Professor P. O. Box 20600 Telephone: (912)691-6201 Fax: (912)691-6201 Email: franklin@tigerpaw.savstate.edu

Taheerah Fulgham Jackson State University Research Assistant 348 N. Flag Chapel Road, #U-8 Jackson, MS Telephone: (601)922-8318 Fax: (601)979-2478 Email: taheerah f2000@yahoo.com

Nikola Garber National Sea Grant College Program Knauss Sea Grant Fellows Program Manager 1315 East - West Highway R/SG, Rm 11718 Silver Spring, MD 20910 Telephone: (301)713-2431 x124 Fax: (301)713-0799 Email: nikola.garber@noaa.gov

Jose Garcia National Weather Service Meteorologist in Charge 1900 English Road Telephone: (806)335-2911 Fax: (806)335-3118 Email: jose.garcia@noaa.gov

Carlos Garza Clark Atlanta University Adjunct Professor 223 James P. Brawley Dr., SW Atlanta, GA 30314 Telephone: (404)880-6903 Fax: (404)880-8499 Email: cgarza@cau.edu

Aisha Gates Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Ann H. Georgilas NOAA/OAR Diversity Program Manager 1315 East West Highway SSMC3/Room 11625 Silver Spring, MD Telephone: (301)713-1495 x122 Fax: (301)713-0163 Email: ann.georgilas@noaa.gov

Eboni Gilbert Alabama A&M University

Matt Gilligan Savannah State University P. O. Box 20467

Savannah, GA 31404 Telephone: (912)356-2809 Fax: (912)356-2808 Email: gillganm@savstate.edu

Mary M. Glackin NOAA/National Environmental Deputy Administrative Assistant 1335 East West Hwy Silver Spring, MD Telephone: (301)713-2010 Fax: (301)713-1249 Email: Mary.Glackin@noaa.gov

Abu Godwin Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Mudlagiri B. B. Goli Mississippi Valley State University Assoc. Prof of Chemistry Dept. of Natural Science & Env. Health 14000 HWY 82 W Itta Bena, MS 38941 Telephone: 662-254-3394 Fax: 662-254-3668 Email: mbgoli@yahoo.com

Richard Gragg Florida A &M University Associate Professor/Director CEEJ 1520 S. Bronough St. Tallahassee, FL 32307 Telephone: 850-599-8193 Fax: 850-561-2248 Email: richard.graffiii@famu.edu

Barbara Graham Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3484 Fax: 601-979-3674 Email: becjegrah@hotmail

Kimberly Grant Alabama A&M University College Student

DeJonnette Grantham-King Advanced Environmental Consultants, Inc. President 5430 Executive Place, Suite 2C Jackson, MS 39206 Telephone: (601)362-1788 Fax: (601)362-3967 Email: adk123@msn.com

Reginald H. Graves, II Morgan State University Student Trainee 11510 American Holly Drive Laurel, MD 20708 Telephone: (301)497-5677 Fax: (301)497-5744 Email: reginald_graves@usgs.gov

Lester Gray NBDC/SAIC Senior Quality Assurance Specialist Building 3205 Stennis Space Center, MS 39529 Telephone: (228)688-2937 Fax: (228)688-3489 Email: les.gray.contractor@noaa.gov

Kentave Greene Jackson State University Research Assistant P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Tamika Greenwood Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Chantain Clevon Greer Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Darrell Grimes University of Southern Mississippi College of Marine Science Hattiesburg, MS

Ronnie Guyton Jackson State University Research Assistant JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: ronnie@twister.jsums.edu

Adwoa Gyekye Clark Atlanta University Research Student 223 James P. Brawley Dr., SW Atlanta, GA 30314 Telephone: (404)880-6905 Fax: (404)880-8499 Email: adj_kg@hotmail.com

Marques Hall Jackson State University Student 732 Woodacre Jackson, MS Telephone: 601-918-2525 Fax: mth274@aol.com Elgenaid Hamadain Jackson State University Assistant Professor of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-2586 Fax: (601)979-5853 Email:

Elsadig Hamed Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Alonzo N. Hamilton, Jr. DOC/NOAA/NMFS/SEFSC Research Fishery Biologist P. O. Drawer 1207 Pascagoula, MS 39568-1207 Telephone: (228)769-4591 x279 Fax: (228)769-9200 Email: adebose@triton.pas.nmfs.gov

Greg Hammer NOA/NESDIS/NCDC Meteorologist 151 Patton Avenue, Room 120 Telephone: (828)271-4263 Fax: (828)271-4328 Email: Gregory.R.Hammer@noaa.gov

Jeremy Hardy Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Alisa Harkins Savannah State University Student P. O. Box 20600 Savannah, GA 31404 Telephone: (912)691-6201 Fax: (912)691-6201 Email: alisaharkins@hotmail.com

Elizabeth Harper Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

James Harrington NASA/GSFC Program Manager Code 933 Greenbelt, MD 20771 Telephone: 301-286-4063 Fax: 301-286-1775 Email: james@muspin.gsfc.nasa.gov

Maggie Harris

Chevron Refinery P. O. Box 1300 Pascagoula, MS 39568 Telephone: (228)938-4600 Fax: (228)938-4366

Elden Hawkes University of Maryland Eastern Shore Graduate Teaching Assistant Carver Hall, 1126 Princess Anne, MD 21853 Telephone: (410)651-6254 Fax: (410)651-7739 Email: Hawkes2@hotmail.com

Cassandra Hawkins Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Jamison S. Hawkins National Weather Service Chief, Plans and Programs Division 1325 East West Hwy Room 15146 Silver Spring, MD 20910 Telephone: (301)713-1975 x135 Fax: (301)713-0173 Email: jamie.hawkins@noaa.gov

Jarrette Hawkins Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Urseline Hawkins Tougaloo College Student 111 Windsor Hills Drive Jackson, MS Telephone: (601)898-0200 Fax: Email: u.hawkins@att.net

Shawana Hawthorne Jackson State University Research Assistant Route 4, Box 273A Kosciusko, MS 39090 Telephone: (601)979-3565 Fax: (601)979-2478 Email: shawana-h@yahoo.com

Rodrick A. Hay California State University Dominguez Hills Associate Professor 100 E. Victoria St. Carson, CA 90747 Telephone: (310)243-3385 Fax: (310)516-4268 Email: rhay@csudh.edu Archie Hayes Savannah State University Student P. O. Box 20600 Savannah, GA 31404 Telephone: (912)691-6201 Fax: (912)691-6201 Email: ricardo73@collegeclub.com

Sekeenia Haynes Florida A & M University Graduate Research Assistant 1520 S. Bronough St. Tallahassee, FL 32307 Telephone: 850-599-8193 Fax: 850-561-2248 Email:

J. Michael Hemsley National Data Buoy Center (NOAA/NWS) Chief, Engineering Branch Building 1100 Stennis Space Center,, MS 39529 Telephone: 228-688-2490 Fax: 228-688-3153 Email: mike.hemsley@noaa.gov

Dagne Hill Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853 Email:

Emorcia V. Hill ABT Associates, Inc. Senior Analyst 55 Wheeler Street Cambridge, MA 02138 Telephone: (617)349-2664 Fax: (617)520-2954 Email: emorcia hill@abtassoc.com

Gerald Hill Texas Southern University Instructor 12307 Valley Hills Drive Missouri City, 77489 Telephone: 713-313-7428 Fax: 713-313-4217 Email: hill_gd@tsu.edu

William T. Hogarth NOAA/NMFS Acting Assistant Administrator for Fisheries 1315 East - West Highway Silver Spring, MD 20910 Telephone: (301)713=2239 Fax: (301)713-1940 Email: bill.hogarth@noaa.gov

Chery G. Horton JobDirect.com Manager, National Diversity Programs 201 Summer Street Stamford, CT 06854 Telephone: (203)943-0925 Fax: (203)327-2261 Email: cherylh@jobdirect.com

Dionne Hoskins DOC/NOAA/NMFS Fishery Ecologist P. O. Box 20228 SSU Savannah, GA 31404 Telephone: (912)352-8925 Fax: (912)351-3571 Email: hoskins@savstate.edu

Letise T. Houser Brown University Student 66-A Fruit Hill Avenue Providence, RI 02909 Telephone: (401)954-0275 Email: Letise_Houser@brown.edu

David Howard Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Frederick Howard Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: frho@twister.jsums.edu

Natalie Huff NMFS/OMI Program Manager for EEO and Diversity 1315 East-West Highway Silver Spring, MD 20910 Telephone: (301)713-1456 Fax: (301)713-1464 Email: Natalie.Huff@noaa.gov

Katessa Hughes Alabama A&M University College Student 54 Manning Road Aliceville, AL 35442 Telephone: (256)851-3362 Fax: (256)851-5905 Email: ladyvelvet98@hotmail.com

Ken Hughes Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Robert Hughes JSU/CSDRA GIS Specialist P.O. Box 18739, JSU Jackson, MS 39217 Telephone: 601-968-8249 Email: rhughes@ccaix.jsums.edu

Sochi Ilomechina Clark Atlanta University Research Student 223 James P. Brawley Dr., SW Atlanta, GA 30314 Telephone: (404)880-6905 Fax: (404)880-8499 Email: sochina@hotmail.com

Ali B. Ishaque Jackson State University Post-Doctoral Research Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-4108 Fax: (601)979-5853

Edet E. Isuk Morgan State University Director 1700 E. Cold Spring Lane Baltimore, MD 21251 Telephone: (443)885-3447 Fax: (410)662-6178 Email: eisuk.moac.morgan.edu

Christi Jackson Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Kashandra Jackson Savannah State University Student P. O. Box 20600 Savannah, GA 31404 Telephone: (912)691-6201 Fax: (912)691-6201 Email: Raquel118@aol.com

Nina Jackson NOAA/NESDIS Education and Diversity Coordinator 1325 East West Highway Room 8244 Silver Spring, MD 20910 Telephone: (301)713-9200 ext. 118 Fax: (301)713)9219 Email: Nina.Jackson@noaa.gov

Shalawn James Delaware State University Student 1200 N. Dupont hwy Dover, DE 19901 Telephone: (302)857-6412 Fax: (302)857-6455 Email: mreiter@dsc.edu

Amalee Jayasinghe Jackson State University Graduate Student P. O. Box 1043 Clinton, MS 39060 Telephone: (601)925-0648 Fax: (601)925-0648 Email: ajayasing@hotmail.com Ambrose Jearld, Jr. NOAA/NMFS/NEFSC Chief, Research Planning and Coordination 162 Water Street Woods Hole, MA 02543 Telephone: (508)495-2318 Fax: (508)495-2258 Email: ambrose.jearld@noaa.gov

Wayne Jearld Morgan State University/P2K Development Officer 5200 Perring Parkway Baltimore, MD 21251 Telephone: (443)885-3621 Fax: (410)319-3843 Email: wjarld@engimorgan.edu

Clement Jedjou Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Gary Jeffress Texas A&M University Professor 6300 Ocean Drive Corpus Christi, TX 78412 Telephone: (361)825-2720 Fax: (361)825-2795 Email: jeffress@tamucc.edu

Tia Jennings Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Donna Johnson NOAA/NMFS/NEFSC Research Fish. Biologist 74 Margruder Road Highlands, NJ 07724 Telephone: (732)872-3092 Fax: (732)872-3088 Email: Donna.Johnson@noaa.gov

Edward R. Johnson National Weather Service Director, Strategic Planning & Policy 1325 East West Highway Silver Spring, MD Telephone: (301)713-0258 x131 Fax: (301)713-1239 Email: Edward.R.Johnson@noaa.gov

Leroy Johnson Alcorn State University Instructor 1000 ASU Dr. #870 Lorman, MS Telephone: 601-877-2303 Fax: 601-827-3989 Email: Ijohnson@lorman.alcorn.edu Lori Johnson Tougaloo College Student 1035 Devonshire Drive Jackson, MS 39206 Telephone: (601)982-5776 Fax: (601)977-6142 Email: Frenchie 90@hotmail.com

Sonji Johnson Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Jeanette X. Jones Alabama A&M University Professor 5006 Inwood Lane Huntsville, AL Telephone: (256)858-4924 Fax: (256)851-5905 Email: aamjxj01@asnaam.aamu.edu

Michael Brandon Jones University of Delaware Ph.D. Student 924 Bowman Drive Lewes, DE 19958 Telephone: (302)645-4036 Fax: (302)645-4007 Email: mbjones@udel.edu

Wanda Jones University of Florida/NMFS Student/Marine Mammal Bio. Tr. NMFS, 3500 Delwood Beach Rd. Panama City, Fl Telephone: (850)234-6541 x226 Fax: (850)235-3559 Email: wanda.jones@noaa.gov

Jenny Jordan Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Dwayne Joseph Clark Atlanta Unversity Graduate student reseracher 223 James P. Brawley, Dr., SW Atlanta, GA 30314 Telephone: 404-880-5904 Fax: 404-880-8499 Email: dwayne_joseph@hotmail.com

Quintin Julius Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu Rezwanul Karim Jackson State University Associate Professor P. O. Box 17660 Jackson, MS 39217 Telephone: 979-7012 Fax: 979-3630 Email: rkarim@ccaix.jsums.edu

Abul B. Kazi Mississippi Valley State University Associate Professor 14000 HWY 82 W 5189 Itta Bena, MS 38941 Telephone: 662-254-3709 Fax: 662-254-3668 Email: kazi@mvsu.edu

Darryl Keith USEPA/Atlantic Ecology Division Oceanographer 27 Tarzwell Drive Narragansett, RI 02882 Telephone: (401)782-3135 Fax: (401)782-3030 Email: keith.darryl@epa.gov

Alex Kessie Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Abu O. A. Khan Jackson State University, Dept of Physics Visiting Assistant Professor 3E-499 Pear Orchard Road Ridgeland, MS 39157 Telephone: (601)-979-8628 Email: ansarik@bellsouth.net

Noreen Khan Texas Southern University Student 15927 Willbriar Lane Telephone: 281-437-3129 Fax: 713-313-4217 Email: noreen_khan@hotmail.com

Latasha Kincaid Tougaloo College Student 56 Acree St. Morton, MS 39117 Telephone: (601)978-1150 Email: latashay@hotmail.com

Jesse F. Kingsberry, III U. S. Environmental Protection Agency 1200 Pennsylvania Ave., NW Washington, DC 20460 Telephone: (202)564-4738 Fax: (202)501-0756

Louisa Koch NOAA/Office of NOAA Research Deputy Assistant Administrator 1315 East West Highway Room 11651 Silver Spring, MD Telephone: (301)713-2458 Fax: (301)713-0163 Email: Louisa.Koch@NOAA.gov

Mikhail Kojine Jackson State University Research Asst. P.O. Box 17910 Jackson, MS 39217 Telephone: 601-979-3723 Fax: 601-979-3723

Beaulah Kondur Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Jan Kucklick NOAA Costal Service Center Outreach Program Manger 2234 South Hobson Ave. Charleston, SC 29401 Telephone: (843)740-1379 Fax: (843)740-1313 Email: jan.kucklick@noaa.gov

Peter Kuntu-Mensah Texas A&M University - Corpus Cristi Assistant Professor 6300 Ocean Drive Cl309 Corpus Cristi, TX Telephone: (316)825-3419 Fax: (361)825-2795 Email: pkuntume@falcon.tamucc.edu

Debora Ladner Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Kevin Lattimore Jackson State University Director, Marketing & Recruitment Jackson, MS 39217

Reginald B. Lawrence NOAA/NESDIS Physical Scientist 8455 Colesville Rd, Suite 1450 Silver Spring, MD Telephone: (301)427-2164 Fax: (301)427-2164 Email: reginald_lawrence@noaa.gov

Tiffany Lee Savannah State University Student P. O. Box 20600 Telephone: (912)691-6201 Fax: (912)691-6201 Email: leet04@hotmail.com Zelma Leflore Jackson State University Lab Coordinator P.O. Box 18839 Jackson, MS 39217 Telephone: 601-979-3451 Fax: 601-979-3716 Email: zleflore@stallion.jsums.edu

Evelyn Leggette Jackson State University Dean, University College P. O. Box 18240 Jackson, MS 39217 Telephone: (601)979-2127 Email: eleggette@ccaix.jsums.edu

Pamela Lestenkof MIMSUP Shannon Point Marine Center Student 1900 Shannon Point Road Anacortes, WA 98221 Telephone: (360)293-2188 Email: pamelalestenkof@hotmail.com

Jerzy Leszczynski Jackson State University Professor of Chemistry P.O. Box 17910 Jackson, MS 39217 Telephone: 601-979-3723 Fax: 601-979-3723

Yongzuo Li Jackson State University Research Associate JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630

Ted I. Lillestolen DOC/NOAA/NOS Acting Assistant Administrator, NOS 1305 East West Highway, Ste. 13632 Silver Spring, MD 20910 Telephone: (301)713-3074 Fax: (301)713-4269 Email: Ted.Lillestone@noaa.gov

Zhi Lin Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Patrice Longshaw Hampton University Student Marine & Environmental Science Department Hampton, VA Telephone: (757)727-5752 Fax: (757)727-5740 Email: patrice.longshaw@hamptonu.edu

Ricardo Lopez California State University -Los Angeles, CEA-CRES Student 5151 State University Drive Los Angeles, CA 90032 Telephone: (323)343-5661 Fax: (323)343-6451 Email: fsrjl@hotmail.com

Howard Loving Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Jesus Loya Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Duanjuan Lu Jackson State University Research Asst. 968 Bellevue PI. Apt. 111 Jackson, MS 39202 Telephone: 601-979-3617 Fax: 601-979-3630 Email: dlu@twister.jsums.edu

Raynette Lyles National Imagery and Mapping Agency 6000 MacArthur Blvd. Bethesda, MD 20816-5003 Telephone: (301)227-7786 Fax: (301)227-4388 Email: biggse@nims_mil

Ally Mack Mississippi Consortium for International Development (MCID) 1225 Robinson Street Jackson, MS 39203 Telephone: (601)979-3370 Fax: (601)979-8657

Natasha Magee Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Rafael Mahecha JSU/HPVCI Technical Asst. 1400 J.R. Lynch Street Jackson, MS 39217 Telephone: 601-979-3617 Fax: 601-979-3630 Email: mahache@twister.jsums.edu Malik B. Malik University of Maryland Eastern Shore Assistant Professor 30398 Pine Street Princess Anne, MD 21853 Telephone: (410)651-6421 Fax: (410)651-7673 Email: mbmalik@mail.umes.edu

Frank Malinky Delaware State University Student 1200 N. Dupont hwy Dover, DE 19901 Telephone: (302)857-6412 Fax: (302)857-6455 Email: mreiter@dsc.edu

Qutaibah M. Malluhi Jackson State University Assoc. Professor P.O. Box 18839 Jackson, MS 39217 Telephone: 601-979-2225 Fax: 601-979-2478 Email: qmalluhi@ccaix.jsums.edu

Randal Mandock Clark Atlanta University Professor 223 James P. Brawley Dr., SW Atlanta, GA 30314 Telephone: (404)880-8964 Fax: (404)880-8499 Email: edmac@cau.edu

Julie Marcy US Army Corps of Engineers - Vicksburg Dist. Chief, Mississippi Recruitment Team 4155 Clay Street Vicksburg, MS 39183 Telephone: (601)631-5560 Fax: (601)631-5583

Maheala Marian Jackson State University Environmental Science Ph.D. Students P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Livingston S. Marshall, Jr. Morgan State University Associate Professor 1700 East Cold Spring Lane Baltimore, MD 21251 Telephone: (443)885-4468 Fax: (410)426-4732 Email: Lmarshall@moac.morgan.edu

Camille Mason Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu Gary C. Matlock NCCOS Acting Director 1305 East West Hwy Silver Spring, MD Telephone: (301)713-3060 Ext. 131 Fax: (301)713-4270 Email: Gary.C.Matlock@noaa.gov

Latrice Maxie National Weather Service Meteorologist 2090 Airport Road New Braunfels, TX 78130 Telephone: (210)657-7356 Email: latrice.maxie@noaa.gov

Amber May Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Eric May University of Maryland Eastern Shore Visiting Assistant Professor Carver Hall, 1126 Princess Anne, MD 21853 Telephone: (410)651-6254 Fax: (410)651-7739 Email: ebmay@mail.umes.edu

Margaret Mary McBride NOAA Chesapeake Bay Fishery Biologist 410 Severn Avenue , Telephone: (410)267-5674 Fax: (410)267-5666 Email: margaret.mcbride@noaa.gov

Travis McCarty Jackson State University Research Asst. P.o. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Michele McCoy NOAA Small Business Specialist/Contract Specialist 2514 Lamp Post Court Chesapeake, VA 23325 Telephone: (757)441-6879 Fax: (757)441-3786 Email: Michele.A.McCoy@noaa.gov

Vera McDaniels Texas Southern University Student 6711 Maczali Drive, Missouri City, TX 77489 Telephone: 281-437-2646 Fax: 713-313-4217 Email: veramcd@hotmail.com

Jennifer McDonald Jackson State University

Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Donna McDowell Savannah State University Student P. O. Box 20600 Savannah, GA 31404 Telephone: (912)691-6201 Fax: (912)691-6201 Email: rosebud003@hotmail.com

Michael McIntosh JSU/CSDRA Research Assistant/Grad. P.O. Box 18739 Jackson, MS 39217 Telephone: 601-979-2989 Fax: 601-979-2478 Email: mackiemac@yahoo.com

Eric McLendon Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Doris McPherson Jackson State University Research Assistant 3640 Dry Grove Road Crystal Springs, MS 39059 Telephone: (601)979-3960 Fax: (601)979-3960 Email: dtmcph@hotmail.com

Robert E. Menzer US EPA Senior Science Advisor 90 Highpoint Drive Gulf Breeze, FL 32561 Telephone: (850)934-7765 Email: menzer.robert@epa.gov

Pam Miller The University of Southern Mississippi Director, Office of Research and Sponsored Program Hattiesburg, MS Telephone: (601)266-4119 Fax: (601)266-4312 Email: pam.miller@usm.edu

Roderick Mitchell Jackson State University Graduate Student Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3860 Fax: (601)979-5853 Email: Mitch4146@yahoo.com

Abdul K. Mohamed

Jackson State University Dean, School of Science & Technology P. O. Box 18750 Jackson, MS 39217 Telephone: (601)979-2153 Fax: (601)979-2058 Email: amohamed@stallion.jsums.edu

Anthony Moncrief Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Ronald Moore Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Tiarra Moore Clark Atlanta University Research student 223 James P. Brawley Dr., SW Box 241 Atlanta, GA Telephone: 404-880-6776 Fax: 404-880-8499 Email: tiarram@hotmail.com

Dennis Morris National Data Buoy Center Building 1100 Stennis Space Center, MS 39529 Telephone: (228)688-1704 Fax: (228)688-3153

Leisa Morrison Howard University Student 752 Fairmont Street, N.W., Washington, DC 20001 Telephone: 202-986-9557 Email: Nickypooh70@hotmail.com

Brenitra L. Mosley Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Guergui Mostovoi Jackson State University Research Associate JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630

Nancy J. Munroe NOAA/NMFSC Bio. Lab. Tech. 166 Water St. Woods, Hole MA Telephone: 508-495-2332 Fax: 508-495-2258 Email: nancy.munroe@noaa.gov

Kinney Murphy Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Helen Mustafa NOAA 162 Water Street Woods Hole, MA 02543 Telephone: (508)495-2318 Fax: (508)495-2004 Email: ambrose.jearld@noaa.gov

Brian Napolian Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Kenneth Ndebele Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Belinda Ndow Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Pamela Neal University of Oklahoma Assistant Dean 100 E. Boyd Street, Room 710 Norman, OK 73019 Telephone: (405)325-3101 Fax: (405)325-3148 Email: pneal@ou.edu

Melvin Newson Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Scott Nichols Mississippi Laboratories Director P. O. Box 1207 Pascagoula, MS 39568-1207

Telephone: (228)762-4591 Fax: (228)769-9200 Email: Scott.Nichols@noaa.gov

Victor Obot Texas Southern University Professor P.O. Box 720122 Houston, TX 77272-0122 Telephone: 713-206-8060 Fax: 713-313-4217 Email: obot@uhibpd.phys.uh.edu

Mildred Huff Ofosu Delaware State University Assistant Provost for Research 1200 N. Dupont Hwy Dover, DE 19901 Telephone: (302)857-6811 Fax: (302)857-6804 Email: mofosu@dsc.edu

Joseph M. Okoh University of Maryland Eastern Shore Chairman, Department of Natural Science Carver Hall, 1126 Princess Anne, MD 21853 Telephone: (410)651-6013 Fax: (410)651-7739 Email: jmokoh@mail.umes.edu

Olufunke Okuyiga-Ezem Jackson State University Student P. O. Box 18149 Jackson, MS 39217 Telephone: (601)979-3473 Fax: (601)979-5853

Peter Olson Florida A&M University, ESI ESI Graduate Student 1520 S. Bronough St. Tallahassee, FL 32307 Telephone: 850-599-8193 Fax: 850-561-2248

Mike Omoregie Jackson State University Chair, Department of Technology Jackson, MS 39217 Telephone: (601)979-2009 Fax: (601)979-4110 Email: omoregie@ccaix.jsums.edu

Barbara R. Ousby Mississippi Development Authority -Energy Division Environmental Scientist P. O. Box 850 Jackson, MS 39205-0850 Telephone: (601)359-6600 Fax: (601)359-6642 Email: bousby@missippi.org

Anthony S. Overton University of Maryland Eastern Shore Graduate Research & Teaching Fellow 1120 Trigg Hall Princess Anne, MD 21853 Telephone: (410)651-7293 Fax: (410)651-7662 Email: asoverton@mail.umes.edu

Lisa Overton University of Maryland Eastern Shore MEES Program Assitant Carver Hall, 1126 Princess Anne, MD 21853 Telephone: (410)651-6254 Fax: (410)651-7739 Email: Iwoverton@mail.umes.edu

Lisa Owens Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Leonard Pace Hampton University Student Marine & Environmental Science Department Hampton, VA Telephone: (757)727-5752 Fax: (757)727-5740 Email: leonard.pace@hamptonu.edu

Trenis Palmer Savannah State University Student P. O. Box 20600 Savannah, GA 31404 Telephone: (912)691-6201 Fax: (912)691-6201 Email: palefsky@tigerpaw.savstate.edu

Bill Parker National Weather Service Meteorologist 5655 Hollywood Avenue 5655 Hollywood Avenue, Telephone: (318)636-7245 Fax: (318)636-9620 Email: Bill.Parker@noaa.gov.

Chuck Patrick Jackson State University Visualization Specialist P.O. Box 18839 Jackson, MS 39217 Telephone: 601-979-5913 Fax: 601-979-3716 Email: chuck_patrick@excite.com

Kevin Patterson Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

N. Joyce Payne NASULGC Director, OAPBC 1307 New York Avenue, N.W., Ste 400 Telephone: (202) 478-6041 Fax: (202)478-6046 Email: jpayne@nasulgc.org

Johnnie Pearson U. S. Environmental Protection Agency Diversity Program Manger NHEERL (MD-85) Telephone: (919)541-0572 Fax: (919)541-2581 Email: pearson.johnnie@epa.gov

Roy A. Pemberton, Jr. Virginia Institute of Marine Science Graduate Student P.O. Box 223 Gloucester Point VA 23062 Telephone: 804-684-7589 Fax: 804-684-7327 Email: rap@vims.edu

Talia M. Pennington Dillard University Student 7210 Farwood Drive New Orleans, LA 70126 Telephone: (504)816-4732 Fax: (504)816-4724 Email: fnormack@mindpring.com

James Perkins Mississippi Consortium for International Development (MCID) Director of Marketing & Program Development 1225 Robinson Street Jackson, MS 39203 Telephone: (601)979-3370 Fax: (601)979-8657 Email: perk199@aol.com

Bryant Pierson Clark Atlanta University Graduate researcher 223 James P. Brawley Dr. SW Atlanta, GA 30314 Telephone: 404-880-6905 Fax: 404-880-8499 Email: br_pierson@bigfoot.com

Taconya Piper University of Maryland Eastern Shore Graduate Teaching Assistant Carver Hall, 1126 Princess Anne, MD 21853 Telephone: (410)651-6254 Fax: (410)651-7739 Email: tdpiper@hotmail.com

Antonio Pitchford Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Arlene Simpson Porter NOAA Grants Management Division MSI Grants Liaison

1325 East West Highway Silver Spring, MD 20910 Telephone: (301)713-0922 ext 152 Fax: (301)713-0947 Email: Arlene.S.Porter@noaa.gov

Raymond Portis Jackson State University Graduate Student Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)362-1168 Fax: (601)979-5853:

Brian Potter USDA Forest Service Research Meteorologist 1407 S. harrison Suite 220 East Lansing, MI 48823 Telephone: (517)355-7740 x28 Fax: (517)355-5121 Email: bpotter@fs.fed.us

Doris Powe Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Alice Powell Alcorn State University Chairperson/Professor 1000 ASU Dr. #870 Lorman, MS Telephone: 601-877-6237 Fax: 601-877-2328 Email: apowell@lorman.alcorn.edu

DeLois M. Powell Shaw University Associate Professor Environmental Science DNPS 118 South Street Raleigh, NC 27601 Telephone: (919)546-8595 Fax: (919)546-8258 Email: powelld@shawu.edu

Nathaniel Powell Mississippi Valley State University Student Dept. of Natural Sciences Itta Bena, MS It38941 Telephone: 662-254-3378 Fax: 662-254-3668

Jewel Prendeville National Science Foundation Staff Associate for Diversity and Education Suite 705, 4201 Wilson Blvd. Arlington, VA 22230 Telephone: (703)292-8500 Fax: (703)292-9042 Email: jprendev@naf.gov Noelani Puniwai Washington State University - Vancouver Graduate Student 14204 NE Salmon Creek Avenue Telephone: (360)600-9899 Email: puniwai@vancouver.wsu.edu

Reginald Quinn Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Monika Rabarison JSU - OIT/HPVCI System Administrator OIT P.O. Box 17750 Jackson, MS 39217 Telephone: 601-979-5933 Fax: 601-979-3716 Email: mrabaris@jsums.edu

James E. Randolph National Marine Fisheries Service Fesearch Fishery Biologist 4700 Avenue U Galveston, TX 77551 Telephone: (409)766-3505 Fax: (409)766-3508 Email: james.randolph@noaa.gov

Brenda K. Rascoe JSU-OIT/HPVCI Admin. Assistant Office of Information Technology P. O. Box 17550 Jackson, MS 39217 Telephone: 601-979-4058 Fax: 601-979-3716 Email: brascoe@ccaix.jsums.edu

Stephen Ratchford University of the Virgin Islands Assistant Professor of Marine Biology UVI 2 John Brewers Bay St. Thomas, VI 00802 Telephone: (340)693-1387 Fax: (340)693-1385 Email: sratchf@uvi.edu

Mac V. Rawson Georgia Sea Grant Director Telephone: (706)542-6009 Fax: (706)542-3652)

Brian Raymond Northern Arizona University Student 106A Paseo del Rio Flagstaff, AZ 86001 Telephone: (520)556-3021 Fax: (520)523-8111 Email: btr@dana.ucc.nau.edu Aisha Reed Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: aisha2002@stallion.jsums.edu

David B. Reed NWS/Lower Mississippi River Forecast Center Hydrometeorologist 62300 Airport Road Telephone: (504)641-4343 Fax: (504)643-1226

Lamar Reed Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Suseela Reedy Jackson State University Asst. Professor P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-3636 Fax: 601-979-8623 Email: rsreddy@ccaix.jsums.edu

Andrew Reggens Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Quinn Reginald Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630

Tim Reid Mississippi-Alabama Sea Grant Consortium Communications Coordinator P. O. Box 7000 Ocean Springs, MS 39566-7000 Telephone: (228)875-9341 Fax: (228)875-0528 Email: tim.reid@usm.edu

Michael Reiter Delaware State University Associate Professor 1200 N. Dupont hwy Dover, DE 19901 Telephone: (302)857-6412 Fax: (302)857-6455 Email: mreiter@dsc.edu

Himabindu Remata Jackson State University Student Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3461 Fax: (601)979-5853

Praveena Remata Jackson State University Student Assistant JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: rpraveena@ccaix.jsums.edu

Umesh Remata Jackson State University Technical Assistant JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: umreddy@ccaix.jsums.edu

Arena Nauchette Richardson South Carolina State University Student 468 Rogers Drives Telephone: (803)533-3865 Email: nauchette@hotmail.com

Joseph P. Richardson Savannah State University Professor P. O. Box 20583 Savannah, GA Telephone: (912)356-2809 Fax: (912)356-2395 Email: richards@savstate.edu

Pamela Riley-Daniels Jackson State University Student Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3461 Fax: (601)979-5853

Larry Robinson Florida A&M University Professor/Director 1520 S. Bronough St. Tallahassee, FL 32307 Telephone: 850-599-3550 Fax: 850-599-8183 Email: larry.robinson@famu.edu

Marin Robinson Northern Arizona University Assistant Professor Department of Chemistry Box 5698 Flagstaff, AZ 86011 Telephone: (520)523-6295 Fax: (520)523-8111 Email: marin.robinson@nau.edu Tanya Robinson Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853

Ashton Robinson-Cook Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: arobinson104@hotmail.com

Christian Rogers Jackson State University Research Technician Department of Chemistry P. O. Box 17910 Jackson, MS 39217 Telephone: (601)979-2831 Email: csr57@hotmail.com

Crystal Rose Dillard University Student Division of Science New Orleans, LA 70126 Telephone: (504)816-4732 Fax: (504)816-4724 Email: fnormack@mindpring.com

Jason A. Rothenberg NOAA/NMFS Program Analyst 9721 Executive Ctr. Telephone: (727)570-5301 Fax: (727)570-5320 Email: jason.rothenberg@noaa.gov

Shayla Roundtree Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Jacqueline Rousseau NOAA 14th & Constitution Ave. N.W. Room 5230 Washington, DC Telephone: 301-713-0325 Fax: 301-713-1974

Julian Saulas U. S. Navy Officer Programs District Manager 2400 Presidents Drive, Suite 250 Montgomery, AL 36176 Telephone: (334)279-8545 Fax: (334)279-7342

Nora Savage Howard University Postdoctoral Research Associate 4th & College Sts, NW, Annex III, Wash DC 20059 Itta Bena, MS 38941 Telephone: (202)806-7887 Fax: (202)462-4053 Email: nsavage@howard.edu

John Schattel NWS-MDL Meteorologist 1325 East West Hwy Silver Spring\, MD 20910 Telephone: (301)713-0056 ext 195 Fax: (301)713-9395 Email: John.Schattel@noaa.gov

Russell Schneider NOAA-NWS-NCEP Storm Prediction Center Chief, Science Support 1313 Halley Circle Norman,, OK 73099 Telephone: 405-579-0704 Fax: 405-579-0700 Email: russell.schneider@noaa.gov

Glenda Schornick National Data Buoy Center Assistant Chief, Resources Branch Building 1100 Stennis Space Center, MS 39529 Telephone: (228)688-1704 Fax: (228)688-3153 Email: Glenda.Schornick@noaa.gov

Andrea Sealy Pennsylvania State University Student 503 Walker Building University Park, PA 16802 Telephone: (814)863-1036 Fax: (814)865-3663 Email: bajan_75@yahoo.com

Lawrence Self Department of Commerce Deputy Director, OCR 145h & Constitution Avenue, NW Washington, DC 20230 Telephone: (202)482-0626 Fax: (202)273-3364 Email: Iself@doc.gov

Ariel Settles Morgan State University Student 1301 Lakeside Avenue Baltimore, MD 21218 Telephone: Fax: Email: asettles77@yahoo.com

Billy Settles Morgan State University Undergraduate Student 1301 Lakeside Ave Telephone: 410-366-5889 Email: bsettles1@hotmail.com Vijaya Shankar Jackson State University Visiting Instructor 4701 Lakeland Drive Apt. 14 B Flowood, MS 39208 Telephone: 601-420-0741 Fax: (601)979-3630 Email: vijayashankararan @hotmail.com

Robert L. Shepard Science and Engineering Alliance Exeuctive Director 1522 K Street, Suite 210 Washington, D.C. 20005 Telephone: Email: Shepard@sea2.org

Kurt D. Shinkle NOAA/NGS State Geodetic Advisor P. O. Box 1850 Stop 83-20 Jackson, MS 39215 Telephone: (601)359-5357 Fax: (601)359-5349 Email: ngs-shinkle@mdot.state.ms.us

Somiya Shukla Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Keita Shumaker Jackson State University Environmental Science Ph.D. Student 6243 Waylawn Drive Jackson, MS 39206 Telephone: (601)979-3586 Email: ketials@hotmail.com

Walid Siddig Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Jamese Sims Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Michael Sissenwine NOAA/NMFS Science Director 166 Water Street Woods Hole, MA 02543 Telephone: (508)495-2233 Fax: (508)495-2232 Email: Michael.Sissenwine@noaa.gov Angela U. Smith Dillard University Student 7210 Farwood Drive New Orleans, LA 70126 Telephone: (504)816-4732 Fax: (504)816-4724 Email: fnormack@mindpring.com

Daryl S. Spencer Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Kasey Spriggs Delaware State University Student 1200 N. Dupont hwy Dover, DE 19901 Telephone: (302)857-6412 Fax: (302)857-6455 Email: mreiter@dsc.edu

Queen E. Spriggs NOAA/NMFS EEO/Diversity Team 1315 East-West Hwy Silver Spring, MD 20910 Telephone: (301)713-2252 ext. 167 Fax: (301)713-2258 Email: queen.spriggs@noaa.gov

Denise Stephenson-Hawk The Stephenson Group Chairman 4000 Colony Square, ite 200 Telephone: (404)870-9060 Fax: (404)870-9005 Email: thestephensongroup.msn.com

Janita Stewart JSU Center for Business Development 931 Highway 80 West Jackson, MS 39204 Telephone: (601)979-2797 Fax: (601)979-2796

Robert Stewart Texas A&M University Professor Oceanography Department Corpus Christi, TX Telephone: (979)845-2995 Email: stewart@ocean.tamu.edu

Rondah Stewart Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: smart_jsu@yahoo.com Shundra Stewart Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: ssram@bellsouth.net

Sheila Stiles DOC/NOAA/NMFS/NEFSC Research Geneticist Milford Lab 212 Rogers Avenue Milford, CT 06460 Telephone: (203)579-7024 Fax: (203)579-7070 Email: sheila.stiles@noaa.gov

Robert H. Stockman NOAA/OFA Director, Strategic Management Staff 6811 Herbert C. Hoover Bldg. Washington, DC 20230 Telephone: (202)482-2461 Fax: (202)482-0295 Email: Bob.Stockman@noaa.gov

Romona J. Strong Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Richard Sullivan Jackson State University Professor & Chair P.O. Box 17910 Jackson, MS 39217 Telephone: 601-979-2171 Fax: 601-979-3730 Email: sullivan@tiger.jsums.edu

Turkesa Sonyette Sullivan Alabama A&M University Student 168 Providence Avenue Aliceville, AL 35442 Telephone: (256)851-3403 Fax: (256)851-5905 Email: turkesas@hotmail.com

Dwayne J. Sutton Jackson State University Environmental Science Ph.D. Student 1125 Raymond Road, Apt. 1C Jackson, MS Telephone: (601)979-4108 Email: sutt2006@stallion.jsums.edu

Shelton Swanier Jackson State University Asst. to the Dean P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718 LaDon Swann Mississippi-Alabama Sea Grant Consortium Associate Director Dauphin Island Sea Lab P. O. Box 369-370 Dauphin Island, AL, AL 36582 Telephone: (334)861-7544 Fax: (334)861-4646 Email: swanndl@auburn.edu

Olayiwola Taylor Clark Atlanta University Graduate Researcher 223 James P. Brawley Dr., SW Atlanta, GA 30314 Telephone: (404)880-8321 Fax: (404)880-8499 Email: Itb66@yahoo.com

Paul B. Tchounwou Jackson State University Professor, Director Environmental Science Ph.D. Department of Biology P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3321 Fax: (601)979-5853 Email: paul@stallion.jsums.edu

Edward Thomas, Jr. Auburn University Assistant Professor 206 Allison Lab Physics Department Auburn University, AL 36849 Telephone: (334)844-4126 Fax: (334)844-4613 Email: physics.auburn.edu

Henry G. Thomas JSU Center for Business Development Director 931 Highway 80 West Jackson, MS 39204 Telephone: (601)979-2797 Fax: (601)979-2796 Email: hthomas@ccaix.jsums.edu

Laurie Thomas Savannah State University Student P. O. Box 20600 Savannah, GA 31404 Telephone: (912)691-6201 Fax: (912)691-6201 Email: LaurieT18@excite.com

Renard Thomas Texas Southern University graduate student 144 Houston St., Angleton, TX 77515 Telephone: 713-515-1710 Fax: 713-313-4217 Email: rthomas@angleton.isd.tenet.edu

Amanda Thompson Northern Arizona University Student 213 W. Elm Avenue Flagstaff, AZ 86001 Telephone: (520)213-5635 Fax: (520)523-8111 Email: ajt2@dana.ucc.nau.edu

Tera Thompson Tougaloo College Student P. O. Box 662 Magnolia, MS 39652 Telephone: (601)978-1136 Email: 843429@tougaloo.edu

Fred Thurberg NOAA 162 Water Street Woods Hole, MA 02543 Telephone: (508)495-2318 Fax: (508)495-2004 Email: ambrose.jearld@noaa.gov

Muneerah Travis Tougaloo College Student 110 Pine Knoll Drive Ridgeland, MS 39157 Telephone: (601)956-7176 Email: munet1@msn.com

Carolyn Trigg US Army Corps of Engineers - Vicksburg Dist. 4155 Clay Street Vicksburg, MS 39183 Telephone: (601)631-5560 Fax: (601)631-5583 Email: Mike.A.Turner@mvk02.usace.army.mil

Paul S. Trotter National Weather Service Meteorologist in Charge 62300 Airport Road Telephone: (504)645-0984 Fax: (504)649-2907 Email: Paul.Trotter@noaa.gov

Donna Turgeon NOAA/National Ocean Service Lead Ecologist N/Sci1 1305 East-West Hwy Silver Spring, MD Telephone: (301)713-3028 x174 Fax: (301)713-4388 Email: donna.turgeon@noaa.gov

Justin Turner Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Michael Turner US Army Corps of Engineers -Vicksburg Dist. Chief, Mississippi Recruitment Team 4155 Clay Street Vicksburg, MS 39183 Telephone: (601)631-5560 Fax: (601)631-5583 Email: Mike.A.Turner@mvk02.usace.army.mil

Clinton Twilley Concurrent Technologies Manager, New Orleans 3520 Gen De Gaulle Suite 4040 New Orleans, LA 70114 Telephone: (504)364-6301 Fax: (504)364-6310 Email: twilley@ctc.com

Cortavius Tyner Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Lawrence C. Tynes, Sr. DOC/NOAA/EASC Human Resources Advisor DOC/EASC, 200 World Trade Center Norfolk, VA 23510-1624 Telephone: (757)441-3638 Fax: (757)441-3563 Email: Lawrence.C.Tynes@noaa.gov

Joseph A. Uravitch NOAA/National Ocean Service Marine Protected Areas Manager 1305 East-West Hwy, (N) Silver Spring, MD 20910 Telephone: (301)713-3155 ext. 195 Fax: (301)713-4367 Email: joseph.uravitch@noaa.gov

Aparna Vaddi Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Gregory Vaughn Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Joseph M. Wahome Mississippi Valley State University Assistant Professor Dept. of Natural Sciences Itta Bena, MS Itta Bena, MS 38941 Telephone: 662-254-3383 Fax: 662-254-3668 Email: wahome@mvsu.edu

Gloria J. Walker National Weather Service EEO Program Manager 1325 East West Hwy, Room 11348 Silver Spring, MD Telephone: (301)713-0692 x198 Fax: (301)713-3374 Email: gloria.walker@noaa.com

Sara Waller California State University Dominguez Hills Assistant Professor 4034 Emeral Street Telephone: (310)793-1354 Fax: Email: viennaravens@socal.rr.com

Tiawana R. Ware Jackson State University Student JSU P. O. Box 17087 Jackson, MS 39217 Telephone: (601)979-2155 Fax: (601)979-2058 Email: swhitfld@stallion.jsums.edu

Mike Warren Jackson State University Environmental Science Ph.D. Student P.O. Box 18540 Jackson, MS 39217 Telephone: 601-979-3321 Fax: 601-979-5853 Email:

Deborah Washington Chevron Refinery P. O. Box 1300 Pascagoula, MS 39568 Telephone: (228)938-4600 Fax: (228)938-4366

Michael Washington National Weather Service Meteorologist 1325 E-W Highway Silver Spring, MD Telephone: 301-713-3379 Fax: 301-713-1239 Email: Michael.J.Washington@noaa.gov

Teresa Washington Jackson State University Environmental Science Ph.D. Student 875 William Blvd, #1716 Ridgeland, MS 39157 Telephone: (601)206-8045 Email: teresawashington@hotmail.com

Benjamin Watkins NOAA/NESDIS Physical Science Administrator 151 Patton Avenue Telephone: (828)271-4384 Fax: (828)271-4246 Email: Benjamin.Watkins@noaa.gov

Carmella Watkins NOAA/NESDIS/NCDC Meteorologist 151 Paton Avenue Telephone: (828)271-4105 Fax: (828)271-4022 Email: carmella.davis.watkins@noaa.gov

Monesa Watts Jackson State University Research Assistant JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: mwatts@stallion.jsums.edu

Michelle Webb NOAA/NESDIS Graduate Scientist 13103 Larchdale Rd., Apt. #3 Laurel, MD 20708 Telephone: (301)457-5244 Fax: (301)457-5199 Email: Michelle.Webb@noaa.gov

Tiffanee Wells Texas A&M University 6515 Ocean Drive Unit 0721 Corpus Christi, TX Telephone: (361)825-4392 Email: tiffanee@cbi.tamucc.edu

Shannon Wheat Hampton University Student Marine & Environmental Science Department Hampton, VA Telephone: (757)727-5752 Fax: (757)727-5740 Email: shannon.wheat@hamptonu.edu

Loren White Jackson State University Visiting Assistant Professor JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: Iwhite@stallion.jsums.edu

William L. White
Jackson State University
Associate Dean, School of Science & Technology
P. O. Box 18750
Jackson, MS 39217
Telephone: (601)979-2153
Fax: (601)979-2058
Email: wwhite@stallion.jsums.edu

Pace Wilber NOAA Coastal Services Center LCR Program Manager 2234 Hobson Avenue Telephone: 843-740-1235 Fax: 843-740-1315 Email: Pace.Wilber@noaa.gov Albert L. Williams Jackson State University Research Assistant 247 Bazinsk Road Vicksburg, MS 39180 Telephone: (601)636-2988 Fax: (601)979-2478 Email: hope1al@hotmail.com

Ammie Williams Mississippi Valley State University Student Department of Natural Sciences Itta Bena, MS Telephone: (662)254-3377 Fax: (662)254-3668 Email: wahome@mvsu.edu

Fred Williams, III Tuskegee University College of Veterinary Medicine Student 2709 Althea Street Telephone: (334)727-7785 Email: sinisterIII@hotmail.com

Joyce Williams Jackson State University Environmental Science Ph.D. Student P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3321 Fax: (601)979-5853

Tiffany Williams Jackson State University Student P. O. Box 18149 Jackson, MS 39217 Telephone: (601)979-3473 Fax: (601)979-5853

Voletta P. Williams Alcorn State University Assistant Professor 1000 ASU Drive #1027 Lorman, MS Telephone: (601)877-6243 Fax: (601)877-3989

Jonathan Wilson Morgan State University Assoicate Professor & Director 1700 E. Coldspring Lane C-108 Baltimore, MD 21251 Telephone: (443)885-3313 Fax: (410)319--3324 Email: jwilson@morgan.edu

Marenda Wilson Dillard University Student 7210 Farwood Drive New Orleans, LA 70126 Telephone: (504)816-4732 Fax: (504)816-4724 Email: fnormack@mindpring.com Yuoguo Xu Jackson State University Graduate Student 948 Bellevue Place, #219 Jackson, MS 39202 Telephone: (601)979-3960 Fax: (601)979-3690 Email: ygxu@hotmail.com

Ilya Yandy Jackson State University Research Asst. P.O. Box 17910 Jackson, MS 39217 Telephone: 601-979-3723 Fax: 601-979-3723 Email: ilya@tiger.jsums.edu

William David Yates Jackson State University Environmental Science Ph.D. Student P. O. Box 18540 Jackson, MS 39217 Telephone: (601)979-3321 Fax: (601)979-5853

Xiadong Yin Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718 Ezra Yohannes Shaw University Student DNPS 118 East South Street Raleigh, NC 27601 Telephone: (919)508-7273 Fax: (919)546-8258 Email:

John Young JSU/CSPRA GIS Specialist P.O. Box 288 Vicksburg, MS 39180 Telephone: 601-968-8249 Email: jyoung@ccaix.jsums.edu

Pao-Chiang Yuan Jackson State University Professor P. O. Box 18480 1400 John R. Lynch Street Jackson, MS 39217 Telephone: (601)979-2489 Fax: (601)979-4110 Email: pyuan@ccaix.jsums.edu

Ashley Zachary Jackson State University Student JSU Department of Physics P. O. Box 17660 Jackson, MS 39217 Telephone: (601)979-7012 Fax: (601)979-3630 Email: ashleyzach@excite.com M. S. Zaman Alcorn State University Asst. Professor 1000 ASU Dr. #870 Lorman, MS Telephone: 601-877-6245 Fax: 601-877-3989 Email: zaman@lorman.alcorn.edu

Freddie Zeigler National Weather Service Meteorologist 62300 Airport Road Telephone: (504)649-0429 Fax: (504)649-2907 Email: freddie.zeigler@noaa.gov

Rui Zhang Jackson State University Research Asst. P.O. Box 18109 Jackson, MS 39217 Telephone: 601-979-2312 Fax: 601-979-2718

Featured Speakers

Dr. Margaret S. Leinen was appointed Assistant Director for Geosciences at the National Science Foundation effective January 10, 2000. In addition to her responsibilities as the Assistant Director, Dr. Leinen is responsible for coordinating environmental science, engineering and education programs within the National Science Foundation (NSF), and for environmental cooperation and collaborations between NSF and other Federal agencies.

Prior to coming to NSF, Dr. Leinen was Dean, Graduate School of Oceanography and Vice Provost for Marine and Environmental Programs at the University of Rhode Island. She was also interim Dean, College of the Environment and Life Sciences. Dr. Leinen spent her entire academic career at the University of Rhode Island, considered one of the country's top institutions for marine studies. During her tenure, she spearheaded the University's efforts to build a cohesive interdisciplinary marine and environmental focus. Dr. Leinen is a well-known researcher in paleooceanography and paleoclimatology. Her work focuses on the history of biogenic sedimentation in the oceans and its relationship to global biogeochemical cycles, and the history of eolian sedimentation in the oceans and its relationship to climate.

Dr. Leinen received her B.S. degree (1969) in Geology from the University of Illinois; M.S. (1975) in Geological Oceanography from Oregon State University; and Ph.D. (1980) in Geological Oceanography from the University of Rhode Island.

She is past president of The Oceanography Society. She served on the Board of Governors of the Joint Oceanographic Institutions, Inc., and the Ocean Research Advisory Council. Dr. Leinen also served as the Vice Chair of the International Geosphere-Biosphere Programme and on the Board on Global Change of the National Research Council/National Academy of Sciences.

POSTER ABSTRACTS

(79)

PESTICIDE INDUCED METHYLATION OF DNA BASE PAIRS

Garetth Forde*1.2, Glake Hill², Latasha Salter³ and Jerzy Lesczynski²

¹Environmental Science Ph.D. Program, School of Science and Technology, ²Computational Center for Molecular Structures and Interactions, Jackson State University, MS 39217, ³Tougaloo College, Tougaloo, MS 39174.

Environmental scientists continue to invest a great deal of time and resources into developing ways to protect DNA from damage causing agents. Chemical agents such as pesticides can be metabolized into products capable of causing DNA mutations. These mutagens can damage DNA in several ways. One common method is through the formation of covalently bound DNA adducts. Our investigation will focus on the methyl adduct. DNA methylation results from the covalent modification of nucleotide bases at nucleophilic sites. In our investigation, we used ab-initio techniques to study the effects of methylation on the O⁶, N³, N⁷ atoms of guanine and the N³ and O² atoms of cytosine. Our primary aim was to characterize the changes in interaction energy and molecular structures of interacting species as a result of methylation. Additionally, we investigated the ability of methylation to stabilize rare tautomers of guanine. These results will provide insight for future investigations aimed at understanding the mechanisms behind methylaton induced mutagenesis.

ADDENDUM - 3

JSU Conference Committee Members

Dr. Abdul K. Mohamed, Dean School of Science & Technology

Mrs. Evola Bates, Chief of Staff Office of the President

> Dr. Paulinus Chigbu Assistant Professor Department of Biology

Chief Rebecca Coleman, Chief Public Safety

Ms. Melvene Coney Office of Development & Alumni Affairs

> Mr. Nathaniel Hoskins Food Services

Mr. Wilbur Smith, Director Food Services

Mrs. Sherrill W. Whitfield Administrative Assistant/Coordinator School of Science & Technology

Sheridan Maiden, Assistant Chief Public Safety

Dr. Gregorio Begonia, Interim Chair Department of Biology

Dr. Rezwanul Karim, Interim Chair Dept. of Physics, Atmospheric & General Sciences

> Dr. Roy DeBerry, Vice President External Affairs

Dr. Thomas Dunaye, Dean School of Allied Health

Ms. Carolyn S. Fletcher Administrative Assistant/Coordinator School of Science & Technology

> Mrs. Jean Franzier, Director Public Information

Dr. James Dixon, Associate Dean School of Allied Health

> Dr. Dorris Gardner, Dean Graduate School

Dr. Glenda Glover, Dean School of Business

Ms. Shonda Allen Administrative Assistant Department of Chemistry

Mr. Wayne Goodwin, Interim Director Facilities Management

> Mrs. Mildred B. Kelley Assist. Dean & Director Universities Center

Dr. Mark G. Hardy Interim Associate Vice President Academic Affairs

Ms. Linda Rush, Senior Recruiter Marketing & Recruitment

Mr. Obra Hackett, Director Career Counseling & Placement Center

Mr. Dowell T. Taylor, Assistant Professor Department of Music

> Dr. Evelyn Leggette, Dean University College

Mrs. Judith Alsobrooks-Meredith W23BC/LPTV

Mr. Harold Loving Office of Information Technology

Dr. Mark Colomb, Director MS Urban Research Center

Dr. Mike Omoregie, Chair Department of Technology

Mrs. Shirley LuAllen, Director Office of Publications

Dr. Gwendolyn Prater, Dean School of Social Work

Mr. Larry McAdoo, Director WJSU

Dr. Dollye M. E. Robinson, Dean School of Liberal Arts

Mrs. Sylvia Watley, Director Student Publications

Dr. Lou Helen Sanders, Dean Jackson State University Libraries

Mr. Julius Baham Center for Spatial Data School of Science & Technology

Mr. Shelton J. Swanier Assistant to the Dean for Strategic Initiatives School of Science & Technology

Mr. Marco McMillian, President SGA

> Dr. Paul Tchounwou Assistant Professor Department of Biology

Mr. Joe Swain, President Graduate School Association

Dr. Evelyn White, Dean School of Education

Dr. Willie Brown, Vice President Information Technology Dr. Samuel White, Dean School of Engineering

Dr. Bettye Graves, Director Data Management & Records

Dr. William L. White, Associate Dean School of Science & Technology

Mrs. Zelma Leflore, Lab Coordinator Information Technology

Dr. Paul Croft Assistant Professor of Meteorology Department of Physics, Atmospheric and General Science

Mrs. Barbara R. Powell Retention/Intervention Coordinator School of Science & Technology

Mr. Curtis Johnson Office of Development & Alumni Affairs

> Mr. Ron Bailey, Grants Writer Office of Sponsored Programs

> Ms. Rita Presley Office of Sponsored Programs

Mr. Larry Belton, Director Marketing & Recruitment

Mr. Johnny Brown Equipment/Facilities Coordinator Department of Athletics Administration

Ms. Maxine Simpson, Coordinator MBRS

Dr. Sam Cobbins, Professor Department of Technology

Mrs. Lori T. Swanier Director, Placement, Alumni & Non-Degree Programs School of Business

Ms. Jennie Griffin Facilities Management

Dr. Vivian Taylor, Professor Department of Curriculum and Instruction

ADDENDUM - 4

NOAA Conference Planning Committee Members

Ms. Eula Brown NOAA, Office of Civil Rights Silver Spring, MD

Ms. Margaret M. McBride NOAA, Chesapeake Bay Program Office Annapolis, MD

> Ms. Ann Georgilas NOAA, OAR Silver Spring, MD

Ms. Helen Mustafa, Program Analyst NOAA, NMFS, Woods Hole, MA

Ms. Natalie Huff Program Manager for EEO and Diversity NOAA, NMFS, Silver Spring, MD Ms. Arlene Simpson Porter Team Leader, Grants Management NOAA, OFA, Silver Spring, MD

Ms. Nina Jackson Education and Diversity Coordinator NOAA, NESDIS, Suitland, MD

> Ms. Queen E. Spriggs NOAA, NMFS, OMI Silver Spring, MD

Dr. Ambrose Jearld, Jr. (Chair) Chief, Research Planning and Coordination NOAA, NMFS, Woods Hole, MA Ms. Gloria Walker EEO Manager NOAA, NWS, Silver Spring, MD

Ms. Jocelyn Bennett-Martin EEO/MSI Manager NOAA, NOS, Silver Spring, MD

Ms. Carmella Davis Watkins National Climatic Data Center Asheville, NC