Detection of CYP3A1 in Formalin-Fixed, Paraffin-Embedded Rat Tissue

Reagent and Antibody Information

1X Wash Buffer
3% Hydrogen Peroxide
1% BSA Diluent
1X Citrate Buffer
DAB Chromagen
Hematoxylin

Blocking Solution: Dakocytomation Protein Block Serum-Free Ready-To-Use

Dakocytomation Corporation Carpinteria CA 93013 www.dakousa.com 1-800-235-5763

Code No. X0909

Avidin / Biotin Blocking Kit Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # SP-2001

Primary Antibody: Rabbit Anti-Rat Cytochrome P450 Enzyme CYP3A1

Chemicon International, Inc Temecula, CA 92590 www.chemicon.com 1-800-437-7500 Catalog # AB1253

Negative Control Serum: Normal Rabbit Serum
Jackson Immunoresearch Laboratories, Inc.
West Grove, PA 19390
www.jacksonimmuno.com
1-800-367-5296
Catalog # 011-000-001

Staining Kit: LSAB+ System-HRP
Dakocytomation Corporation
Carpinteria CA 93013
www.dakousa.com
1-800-235-5763

Code No. K0690

Note: This kit includes reagents needed for the secondary antibody (link) and label complex.

Staining Procedure

Positive Control Tissue: Liver (upregulated by treatment)

Stain Localization: Cytoplasmic

1. Deparaffinize and hydrate slides through the following solutions:

Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Wash Buffer	2 times	5 minutes

- 2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.
- 3. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.
- 4. Heat-Induced Epitope Retrieval Using The Microwave

Place a full rack of slides into a Tissue Tek® container with 200 ml of 1X citrate buffer (Insert blank slides into any empty slots in the rack to ensure even heating of slides) Microwave for 5 minutes at power level 5.

Cool for 1 minute. (Add more citrate buffer, if necessary.)

Microwave again for 5 minutes at power level 5. Temperature Before Cooling Slides_____

Cool 20 minutes at room temperature.

Rinse the slides in 2 changes of distilled water for 3 minutes each time.

- 5. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each time.
- 6. Block with the Dako Protein Blocking Reagent and incubate for 10 minutes at room temperature. Lot #_____ Exp Date_____

DO NOT RINSE THE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.

7. Avidin / Biotin	Blocking Kit			
Lot #	Exp Date	New Kit:	yes	/ no
Apply avidin b	lock - 15 minutes at room temp	perature.		
Quick rinse in	1X Wash Buffer.			
Apply biotin bl	lock - 15 minutes at room temp	erature.		

DO NOT RINSE SLIDES WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY. ONLY WIPE EXCESS BUFFER.

8. Apply the primary antibody at a 1:1500 dilution and incubate for 30 minutes at room temperature. Lot #_____ Date Aliquoted_____

For negative control slides, dilute the protein concentration of the normal rabbit serum to match that of the primary antibody. Make a 1:1500 dilution from this normalized serum and apply to the slides. Incubate for 30 minutes at room temperature.

Lot # Date Reconstituted
9. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.
LSAB+ Kit Lot # Exp Date
10. Apply the Link (yellow bottle) from the LSAB+ Kit and incubate for 30 minutes at room temperature
11. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.
12. Apply the Label (red bottle) from the LSAB+ Kit and incubate for 30 minutes at room temperature.
13. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.
14. Apply the DAB chromagen and incubate in the dark for 6 minutes at room temperature. (Add 1 drop of DAB per ml of substrate) Lot # Exp Date New Kit: yes / no
15. Rinse the slides in tap water 3 minutes.
16. Counterstain with Harris Hematoxylin for 30 seconds.
17. Rinse the slides in tap water until water is clear.

19. Dehydrate through the following solutions:

95% Ethanol	1 time	3 minutes
100% Ethanol	3 times	3 minutes
Xylene	2 times	5 minutes

18. Gently agitate slides in 1X Wash Buffer until they turn blue.

20. Coverslip

Updated 02/01/06