## Detection of CYP2C11 in Formalin-Fixed, Paraffin-Embedded Rat Tissue

## **Reagent and Antibody Information**

1X Wash Buffer 3% Hydrogen Peroxide 1% BSA Diluent 1X Citrate Buffer DAB Chromagen Hematoxylin

Blocking Solution: Dakocytomation Protein Block Serum-Free Ready-To-Use Dakocytomation Corporation Carpinteria CA 93013 www.dakousa.com 1-800-235-5763 Code No. X0909

Avidin / Biotin Blocking Kit Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # SP-2001

Primary Antibody: Rabbit Polyclonal To Cytochrome P450 2C11 Antibody Abcam Inc Cambridge, MA 02139 www.abcam.com 1-888-772-2226 Catalog # ab3571

<u>Negative Control Serum: Normal Rabbit Serum</u> Jackson Immunoresearch Laboratories, Inc. West Grove, PA 19390 <u>www.jacksonimmuno.com</u> 1-800-367-5296 Catalog # 011-000-001

<u>Staining Kit: LSAB+ System-HRP</u> Dakocytomation Corporation Carpinteria CA 93013 <u>www.dakousa.com</u> 1-800-235-5763 Code No. K0690

Note: This kit includes reagents needed for the secondary antibody (link) and label complex.

## **Staining Procedure**

Positive Control Tissue: Liver (upregulated by treatment) Stain Localization: Centilobular cytoplasmic staining pattern

1. Deparaffinize and hydrate slides through the following solutions:

Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Wash Buffer	2 times	5 minutes

- 2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.
- 3. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.
- 4. <u>Heat-Induced Epitope Retrieval Using The Microwave</u> Place a full rack of slides into a Tissue Tek® container with 200 ml of 1X citrate buffer (Insert blank slides into any empty slots in the rack to ensure even heating of slides) Microwave for 5 minutes at power level 5. Cool for 1 minute. (Add more citrate buffer, if necessary.) Microwave again for 5 minutes at power level 5. *Temperature Before Cooling Slides* Cool 20 minutes at room temperature. Rinse the slides in 2 changes of distilled water for 3 minutes each time.
- 5. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each time.
- 6. Block with the Dako Protein Blocking Reagent and incubate for 10 minutes at room temperature. Lot #\_\_\_\_\_ Exp Date\_\_\_\_\_

DO NOT RINSE THE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.

7. <u>Avidin / Biotin Blocking Kit</u> Lot #\_\_\_\_\_ Exp Date\_\_\_\_\_New Kit: yes / no Apply avidin block - 15 minutes at room temperature. Quick rinse in 1X Wash Buffer. Apply biotin block - 15 minutes at room temperature.

DO NOT RINSE SLIDES WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY. ONLY WIPE EXCESS BUFFER.

8. Apply the primary antibody at a 1:100 dilution and incubate for 1 hour at room temperature. Lot #\_\_\_\_\_ Date Aliquoted\_\_\_\_\_

For negative control slides, dilute the protein concentration of the normal rabbit serum to match that of the primary antibody. Make a 1:100 dilution from this normalized serum and apply to the slides.

Incubate for 1 hour at room temperature. Lot #\_\_\_\_\_ Date Reconstituted\_\_\_\_\_

9. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.

LSAB+ Kit Lot # \_\_\_\_\_ Exp Date\_\_\_\_\_

10. Apply the Link (yellow bottle) from the LSAB+ Kit and incubate for 30 minutes at room temperature.

11. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.

- 12. Apply the Label (red bottle) from the LSAB+ Kit and incubate for 30 minutes at room temperature.
- 13. Rinse the slides in 2 changes of 1X Wash Buffer for 5 minutes each.
- 14. Apply the DAB chromagen and incubate in the dark for 6 minutes at room temperature. (Add 1 drop of DAB per ml of substrate) Lot #\_\_\_\_\_ Exp Date\_\_\_\_\_ New Kit: yes / no
- 15. Rinse the slides in tap water 3 minutes.
- 16. Counterstain with Harris Hematoxylin for 30 seconds.
- 17. Rinse the slides in tap water until water is clear.
- 18. Gently agitate slides in 1X Wash Buffer until they turn blue.
- 19. Dehydrate through the following solutions:

95% Ethanol	1 time	3 minutes
100% Ethanol	3 times	3 minutes
Xylene	2 times	5 minutes

20. Coverslip

Updated 08/21/06