

Current Topics in Genome Analysis March 1, 2005



Computational Techniques in Comparative Genomics



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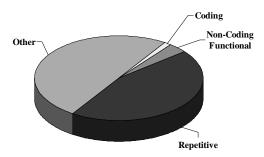


- Fundamental concepts of comparative genomics
- Alignment and visualization tools
 - Pair-wise and multi-species methods
 - Combining with transcription factor binding site data
- Motif Identification
- Comparative genomics resources available at UC Santa
 Cruz -- http://genome.ucsc.edu
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Why Compare Genomic Sequences from Different Species?

Explore evolutionary relationships



Enhanced gene prediction algorithms

Charles Darwin

- Served as naturalist on a British science expedition around the world (1831 -- 1836)
- On the Origin of Species by Means of Natural Selection, or the Preservation of Favoured Races in the Struggle for Life
- The Origin of Species (1859)
 - All species evolved from a single life form
 - "Variation" within a species occurs randomly
 - Natural selection
 - Evolutionary change is gradual

Other Intellectual Foundations

- Darwin (1859)
 - Theories of Evolution
- Mendel (1866) (rediscovered in 1900)
 - Genes are units of heredity
- Avery, McCarty & MacLeod (1944)
 - DNA as the "transforming principle"
- Watson & Crick (1953)
 - Structure of DNA
- Sanger (1977)
 - Methods of sequencing DNA

Rationale

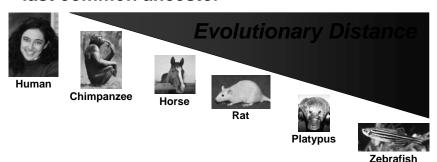
- DNA represents a "blueprint" for structure and physiology of all living things
- All species use DNA
- Mutations in functional DNA are less likely to be tolerated

Comparative Genomics

Find sequences that have diverged less than we expect

These sequences are likely to have a functional role

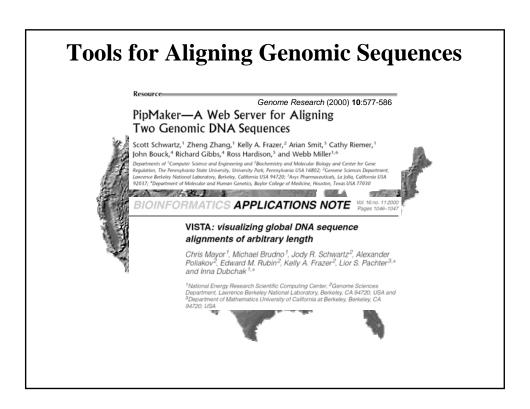
 Our expectation is related to the time since the last common ancestor



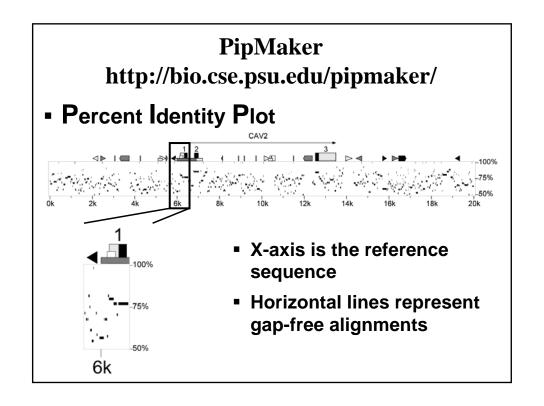
What's in a Name?

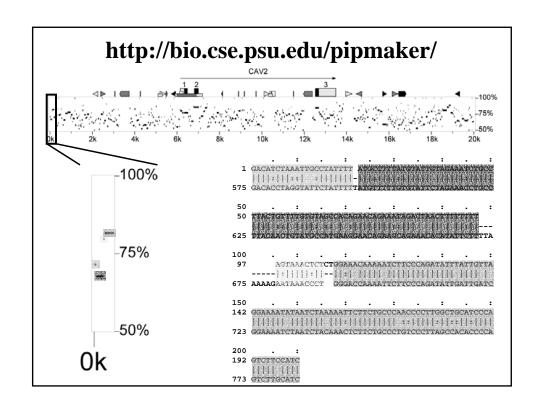
- Highly conserved sequences
- Sequences under purifying selection
- Functionally constrained sequences
- ECOR Evolutionary Conserved Region
 - Variant: ECR
- CNS Conserved Non-coding Sequence
- CNGs Conserved Non-Genic sequence
- MCS Multi-species Conserved Sequence

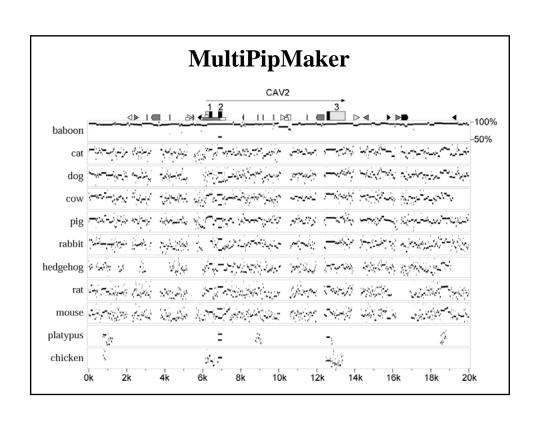
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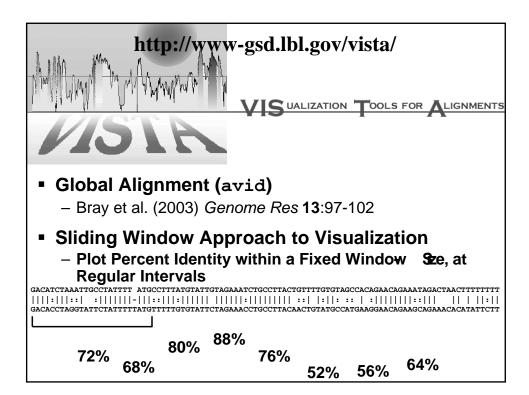


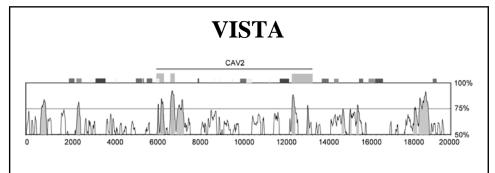
PipMaker vs. VISTA Visualization Alignment Strategy VISTA: avid PipMaker: blastz East Coast – West Coast Lawrence Berkeley National Laboratory



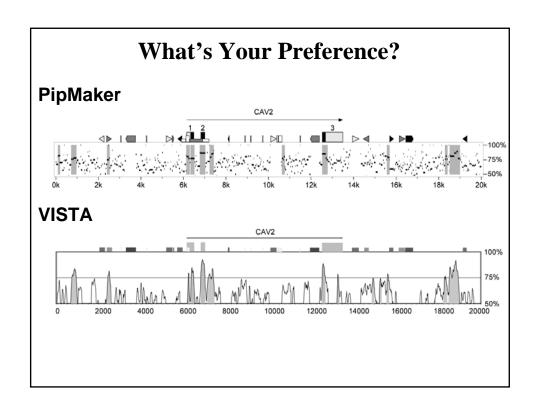








- Percent Identity is plotted from:
 - 100 base windows
 - Moved every 15 bases
- Colored regions meet certain alignment criteria
 - ->100 bp >75% Identity



East & West Coast Unite

http://zpicture.dcode.org/

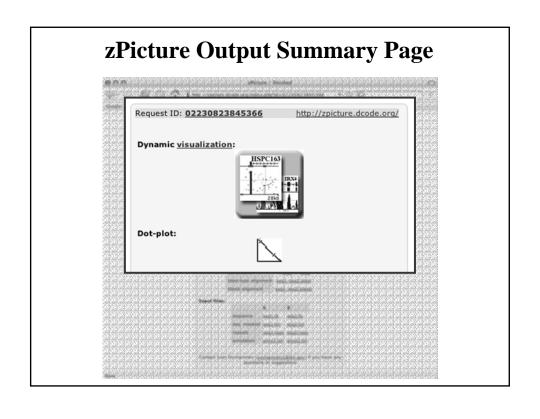
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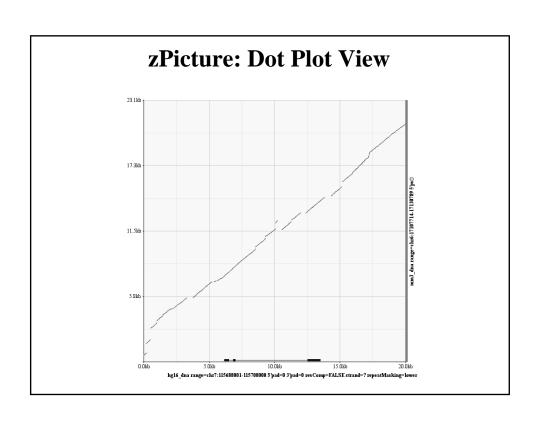
zPicture: Dynamic Alignment and Visualization Tool for Analyzing Conservation Profiles

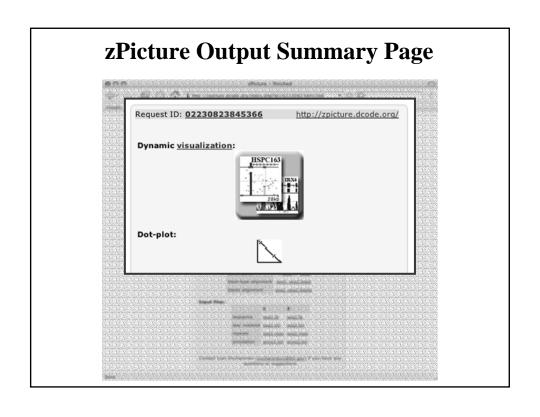
Ivan Ovcharenko,^{1,2} Gabriela G. Loots,² Ross C. Hardison,³ Webb Miller,^{4,5} and Lisa Stubbs^{2,6}

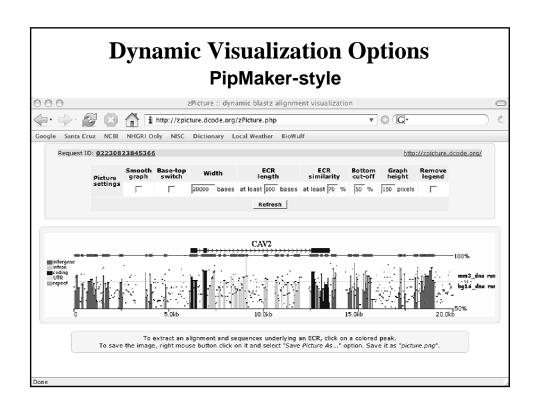
¹Energy, Environment, Biology and Institutional Computing, Lawrence Livermore National Laboratory, Livermore, California 94550, USA; ²Genome Biology Division, Lawrence Livermore National Laboratory, Livermore, California 94550, USA; ³Department of Biochemistry and Molecular Biology, The Pennsylvania State University, University Park, Pennsylvania 16802, USA; ⁴Department of Computer Science and Engineering, The Pennsylvania State University, University Park, Pennsylvania 16802, USA; ⁵Department of Biology, The Pennsylvania State University, University Park, Pennsylvania 16802, USA

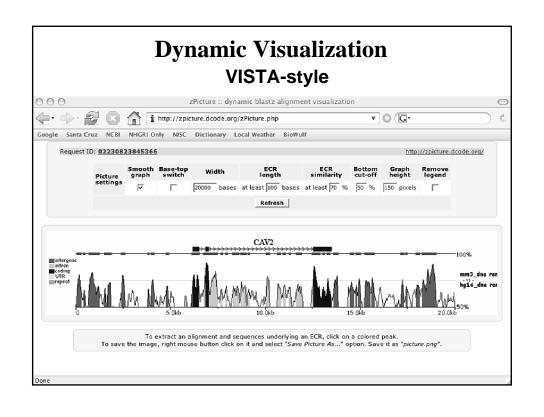
Genome Research, 2004, 14(3):472 7

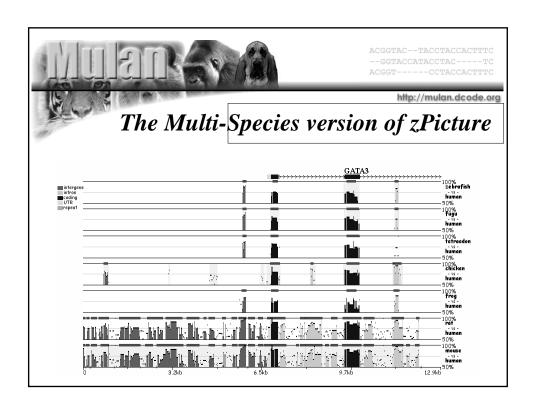






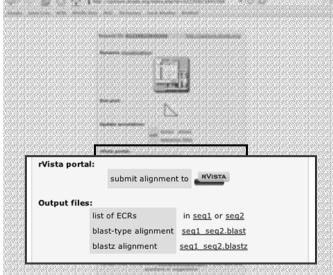






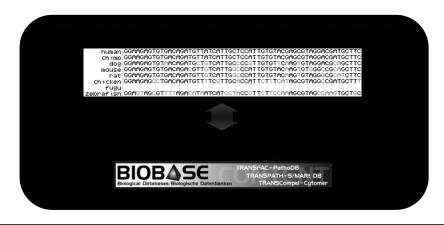
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zPicture Output Summary Page



Are there any transcription factor binding sites in my alignment?





TRANSFAC



http://www.gene-regulation.com/

- A database of:
 - Eukaryotic transcription factors
 - Their genomic binding sites
 - And DNA binding profiles
- Data are collected from published studies
 - Non curated
 - Redundant data

JASPAR: An Alternative to TRANSFAC

Nucleic Acids Research, 2004, Vol. 32, Database issue D91–D94 DOI: 10.1093/nar/gkh012

JASPAR: an open-access database for eukaryotic transcription factor binding profiles

Albin Sandelin, Wynand Alkema, Pär Engström, Wyeth W. Wasserman¹ and Boris Lenhard*

Center for Genomics and Bioinformatics, Karolinska Institutet, Berzelius väg 35, S-17177 Stockholm, Sweden and ¹Centre for Molecular Medicine and Therapeutics, Department of Medical Genetics, University of British Columbia, Vancouver, BC, Canada

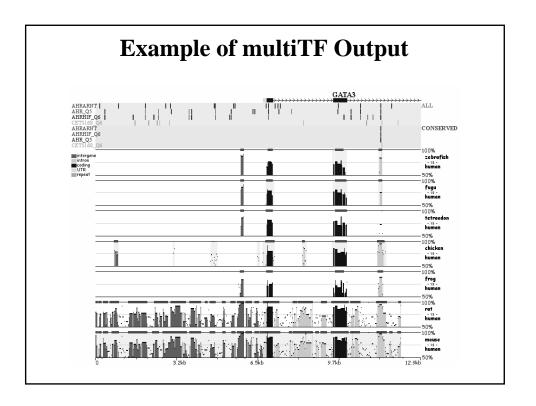
- Differences from TRANSFAC:
 - Manually curated for "high quality" experiments
 - Non redundant collection

http://jaspar.cgb.ki.se/



TRANSFAC Data are inherently "noisy"

- Binding sites are very short6-10 bases in length
- Low complexityOnly 4 "letters" in the DNA alphabet
- Frequently observe binding site by chance
- Conservation can help reduce the noise



Summary of Alignment Tools

- PipMaker (blastz)
- VISTA (avid)
- zPicture and MULAN
- Lagan and mLagan (glocal alignments)
 - http://lagan.stanford.edu/
- rVISTA 2.0
- Box 1 from:

Ureta-Vidal, Ettwiller, and Birney (2003) Comparative Genomics: Genome-Wide Analysis in Metazoan Eukaryotes *Nature Reviews Genetics* **4:** 251-262

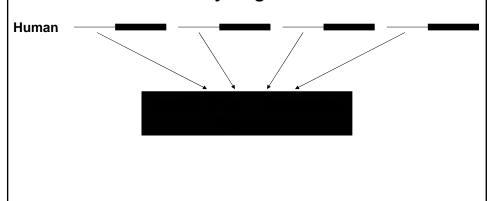
Table 1 from:

Miller, Makova, Nekrutenko, and Hardison (2004) Comparative Genomics *Annual Reviews in Human Genetics* **5**:15-56

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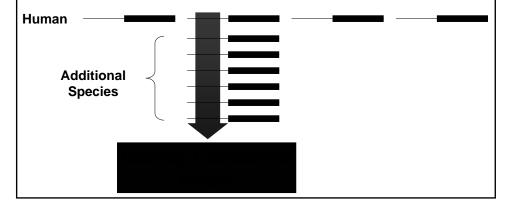
Motif Finding

- Identify Transcription Factor Binding Sites
- What sequences should be searched?
 Coordinately Regulated Genes



Phylogenetic Footprinting

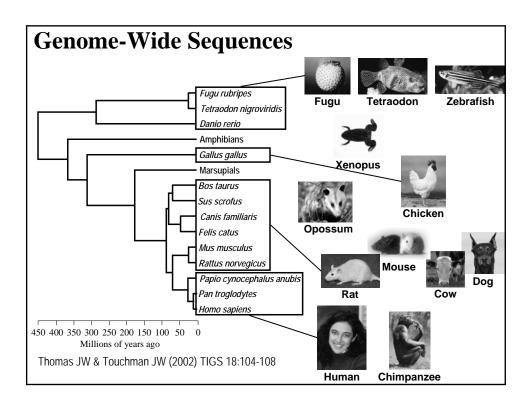
- **FootPrinter** http://bio.cs.washington.edu/software.html
- Takes the phylogeny into account Orthologous Genes



Summary of Phylogenetic Footprinting Tools

- FootPrinter http://bio.cs.washington.edu/software.html
 - Blanchette and Tompa (2003) Nucleic Acids Research **31:**3840–3842
- phyloCon http://oldural.wustl.edu/~twang/PhyloCon/
 - Wang and Stormo (2003) Bioinformatics 19:2369-80
- phyME
 - Sinha, Blanchette, and Tompa (2004) BMC Bioinformatics 28:170
- List of motif finding algorithms:
 - Box 1 of Ureta-Vidal et al. (2003) Nature Reviews Genetics **4:**251-262
- Bayesian Approaches (and home of the Gibbs sampler)
 - http://www.wadsworth.org/resnres/bioinfo/
- Example of motif finding limited by mouse conservation:
 - Wasserman et al. (2000) Nature Genetics 26:225-228

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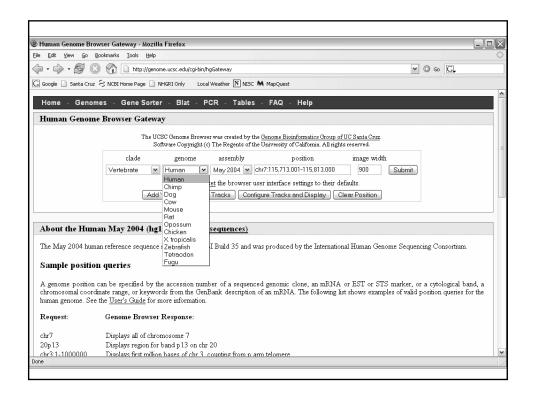
Genome Browsers

UCSC Genome Bioinformatics

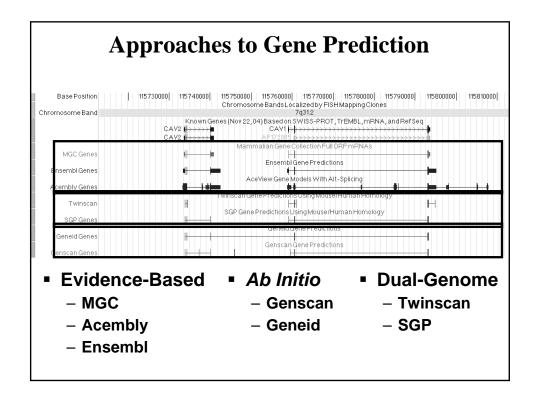
e! project **Ensembl** http://www.ensembl.org



http://www.ncbi.nlm.nih.gov/mapview/



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Additional Gene Prediction Resources

- Fugu BLAT Track at UCSC
- SLAM http://baboon.math.berkeley.edu/~syntenic/slam.html
 - Cawley et al. (2003) Nucleic Acids Research 31:3507-3509
- Exoniphy

Siepel and Haussler. Computational identification of evolutionarily conserved exons. *Proc. 8th Annual Int'l Conf. on Research in Computational Biology*, pp. 177-186, 2004.

http://www.soe.ucsc.edu/~acs/recomb2004.pdf

- Also see genome "test" browser for data
- Box 1 from:
 - Ureta-Vidal et al. (2003) Nature Reviews Genetics 4:251-262

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Chaining Alignments

 Chaining bridges the gulf between large syntenic blocks and base bybase alignments.

The Challenge:

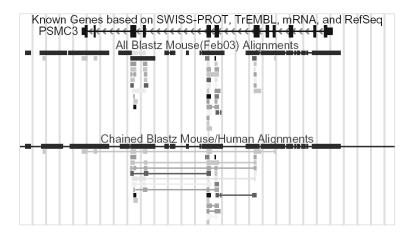
- Local alignments tend to break at transposon insertions, inversions, duplications, etc.
- Global alignments tend to force non homologous bases to align.

The Solution:

 Chaining is a rigorous way of joining together local alignments into larger structures.

Slide (though modified) Courtesy of Jim Kent

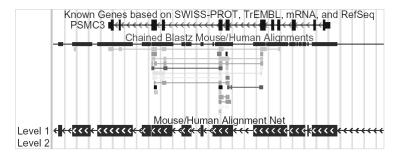
Chains join together related local alignments



Protease Regulatory Subunit 3

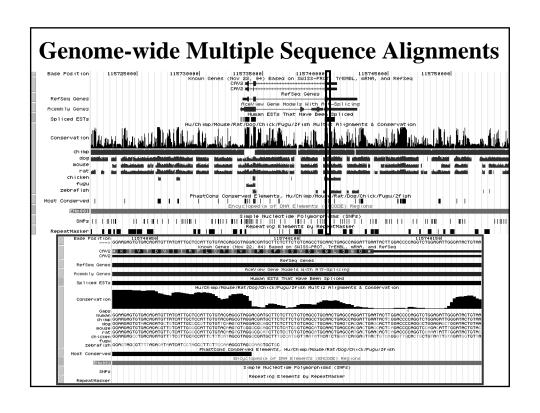
Slide Courtesy of Jim Kent

Net Alignments: Focus on Orthology



- Frequently, there are numerous mouse alignments for any given human region, particularly for coding regions.
- Net finds best mouse match for each human region.

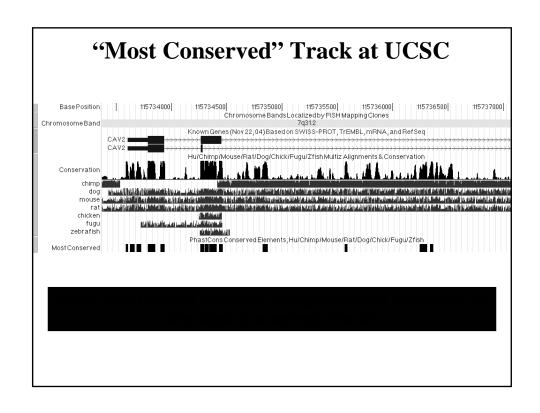
Slide (though modified) Courtesy of Jim Kent

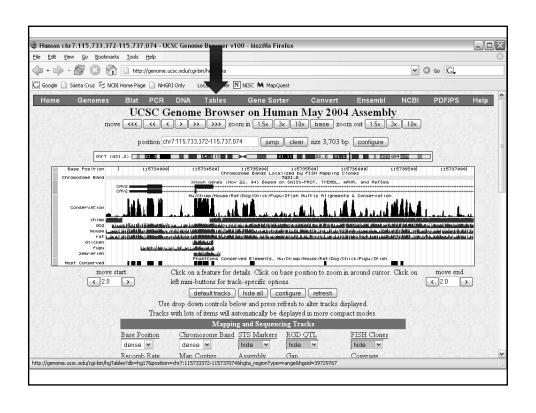


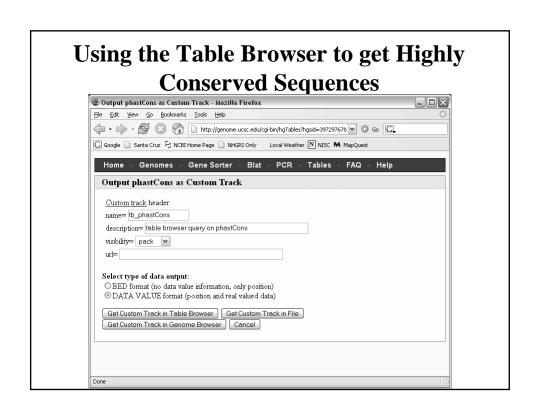
Conservation Score at UCSC

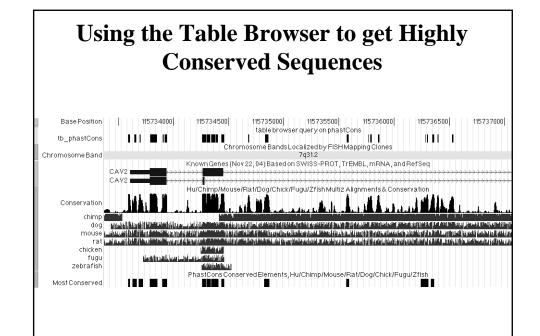
 Displays evolutionary conservation based on a phylogenetic hidden Markov model

- "Most Conserved" track represents highly conserved regions
 - Tuned to cover ~4% of the genome

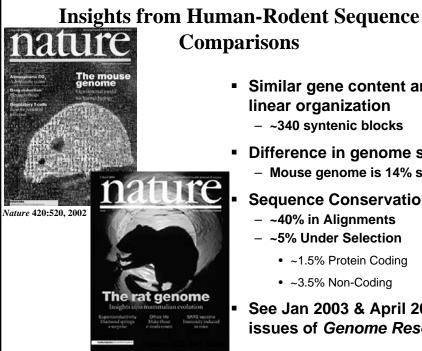








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Comparisons

- Similar gene content and linear organization
 - ~340 syntenic blocks
- Difference in genome size
 - Mouse genome is 14% smaller
- **Sequence Conservation**
 - ~40% in Alignments
 - ~5% Under Selection
 - ~1.5% Protein Coding
 - ~3.5% Non-Coding
- See Jan 2003 & April 2004 issues of Genome Research

Neutral Evolution

- No selective pressure/advantage to keep or change the DNA sequence
- Rate of variation should correlate with:
 - Mutation rate
 - Amount of time since the last common ancestor
- The neutral rate can vary across the genome

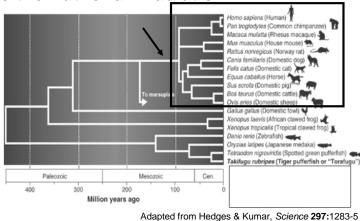
Types of Neutrally Evolving DNA

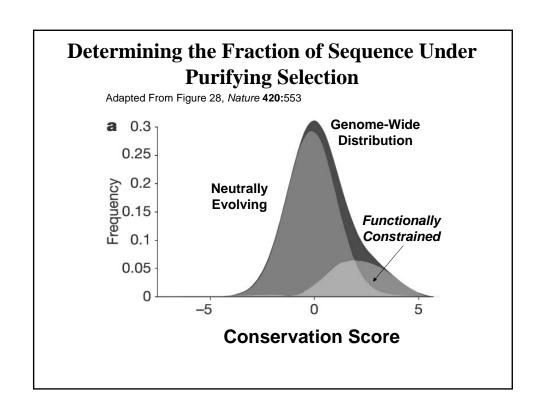
- 4-Fold Degenerate Sites
 - Third position of codons which can be any base and code for the same amino acid

	Second														
First	U	С	Α	G	Last										
U	Phe	Ser	Tyr	Cys	U										
	Phe	Ser	Tyr	Cys	С										
	Leu	Ser	Stop	Stop	Α										
	Leu	Ser	Stop	Trp	G										
С	Leu	Pro	His	Arg	U										
	Leu	Pro	His	Arg	С										
	Leu	Pro	Gln	Arg	Α										
	Leu	Pro	Gln	Arg	G										
Α	lle	Thr	Asn	Ser	U										
	lle	Thr	Asn	Ser	С										
	lle	Thr	Lys	Arg	Α										
	Met	Thr	Lys	Arg	G										
G	Val	Ala	Asp	Gly	U										
	Val	Ala	Asp	Gly	С										
	Val	Ala	Glu	Gly	Α										
	Val	Ala	Glu	Gly	G										

Types of Neutrally Evolving DNA

- Ancestral Repeats
 - Ancient Relics of Transposons Inserted Prior to the Eutherian Radiation





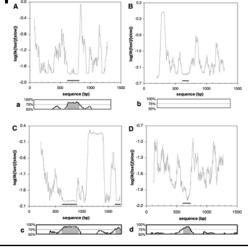


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Phylogenetic Shadowing

Boffelli et al. (2003) Science 299:1391-1394.

 Identifying sequence differences between multiple primate species



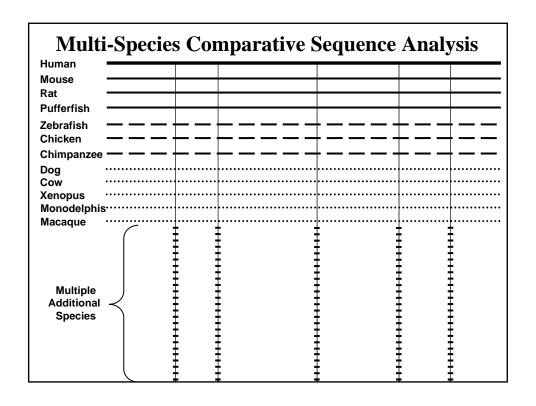
Multi-Species Comparative Sequence Analysis

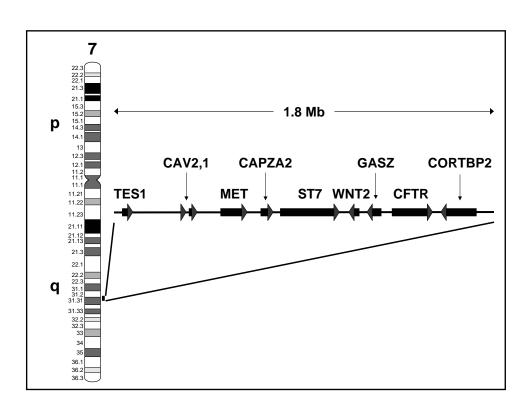
Comparative analyses of multi-species sequences from targeted genomic regions

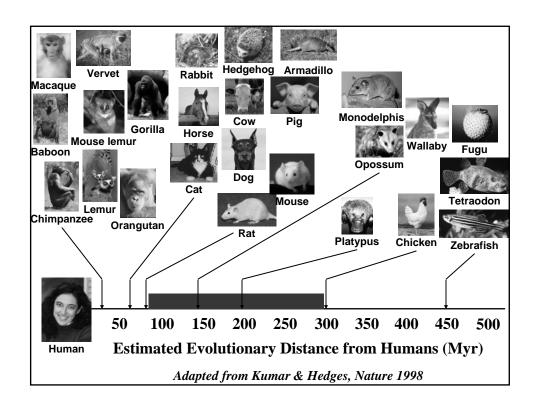
J. W. Thomas¹a, J. W. Touchman¹a², R. W. Blakesley¹a², G. G. Bouffard¹a², S. M. Beckstrom-Sternberg¹a², E. H. Margulies¹, M. Blanchette³, A. C. Siepel³, P. J. Thomas², J. C. McDowell³, B. Maskeri³, N. F. Hansen³, M. S. Schwartz³, R. J. Weber³, W. J. Kent³, D. Karolchika³, T. C. Bruen³, R. Bevan³, D. J. Cuttler⁴, S. Schwartz², L. Elnitski², J. R. Idol¹, A. B. Prasad¹, S.-Q. Lee-Lin¹, V. V. B. Maduro¹, T. J. Summers¹, M. E. Portnoy¹, N. L. Dietricha³, N. Akhtera³, K. Ayelea³, B. Benjamin², K. Cariaga³, C. P. Brinkley³, S. Y. Brooksa³, S. Granite³, X. Guan³, J. Gupta², P. Haghighta³, S.-L. Hoa², M. C. Huanga³, E. Karlinsa³, P. L. Larica³, R. Legaspta³, M. J. Lima³, Q. L. Maduroa³, C. A. Masielloa³, S. D. Mastrian³, J. C. McCloskeya³, R. Pearsona², S. Stantripopa³, E. E. Tiongsona³, J. T. Trana³, C. Tsurgeon³, J. L. Vogt², M. A. Walkera³, K. D. Wetherby³, L. S. Wigginsa³, A. C. Younga³, L.-H. Zhanga³, K. Osoegawaa°, B. Zhua°, B. Zhao°, C. L. Shu°, P. J. De Jonga°, C. E. Lawrencea², A. F. Smita³, A. Chakravarti⁴, D. Hausslera³, P. Green¹°, W. Millera³ & E. D. Green¹¹a²

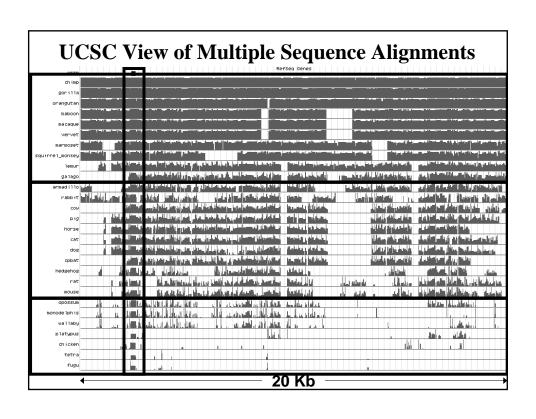
Nature 424:788, 2003

Multi	-{	Sj	p	e	C	ie	S	; (C	0	n	n	p) a	11	.	1	ti	V	e	S	e	q	u	e	n	C	e	: 1	4	r	12	al	ly	/S	si;	S	
Human Mouse Rat Pufferfish																																						
Zebrafish Chicken Chimpanzee Dog Cow Xenopus Monodelphis Macaque				-							•		† 		-	<u>-</u>		-		- - - -	-							•						•	•			



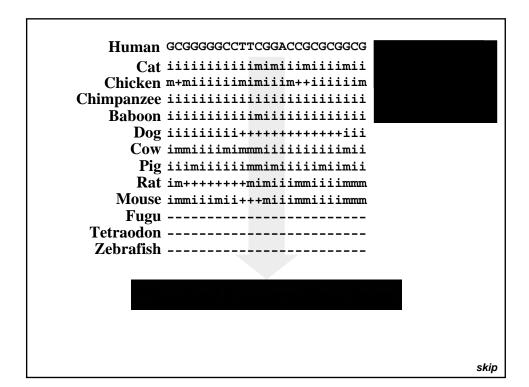


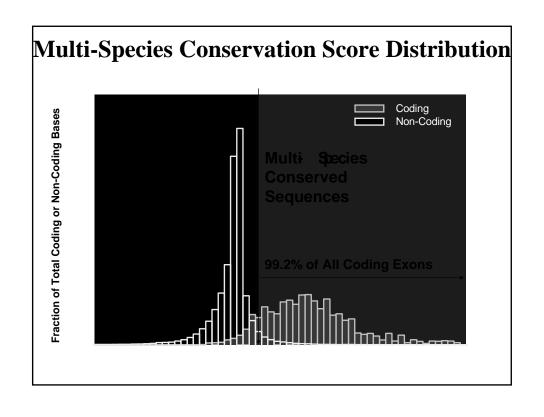


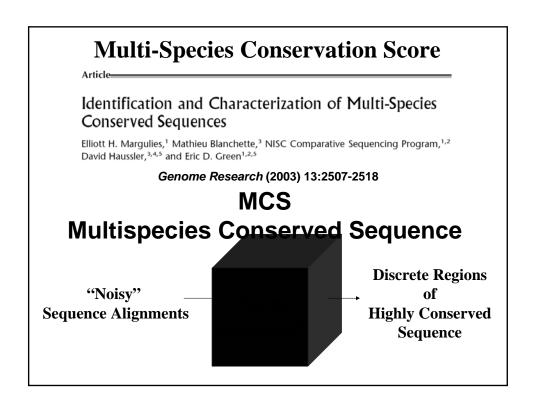


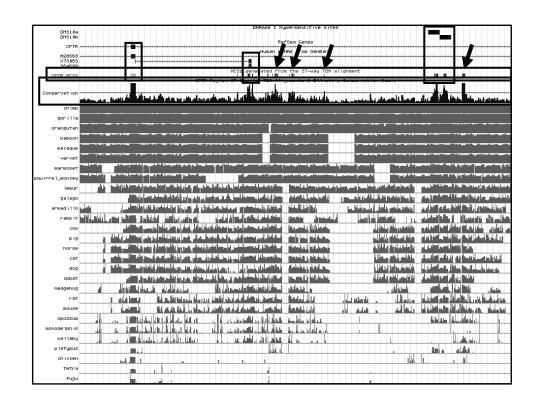
Multi-Species Weighted Conservation Score

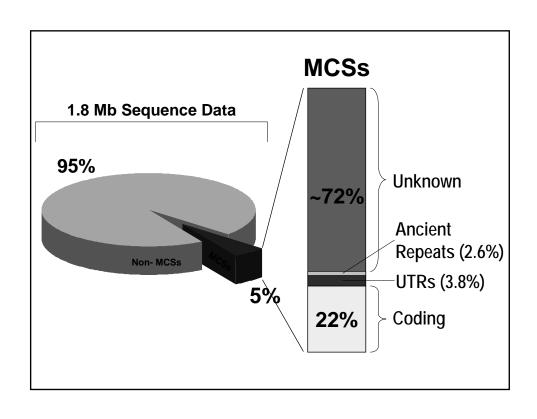
- Takes into Account the Different Divergence Rates of Each Species
 - "A Chicken Alignment Will Contribute More Than a Baboon Alignment"
- Based On the Substitution Rates at Bases under Neutral Selection
 - Calculated from 4-Fold Degenerate Positions

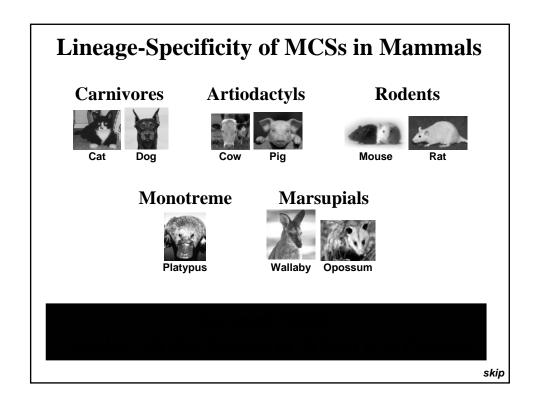


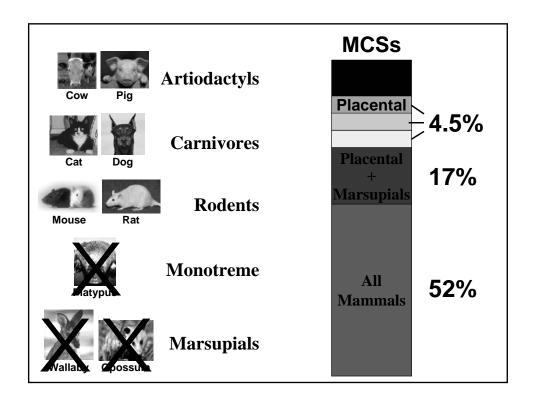


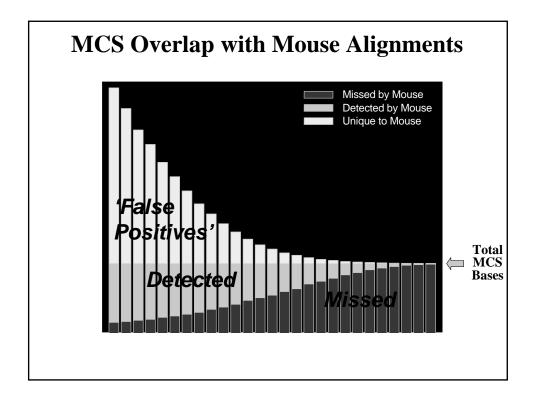






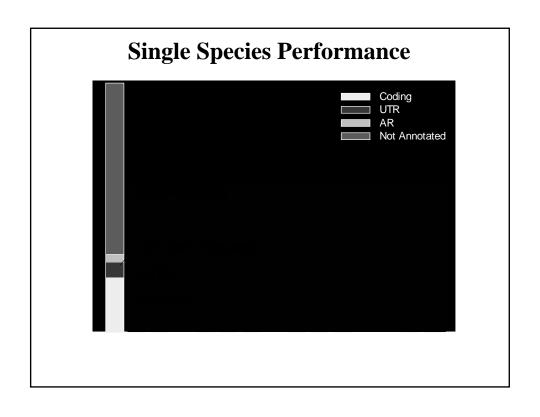


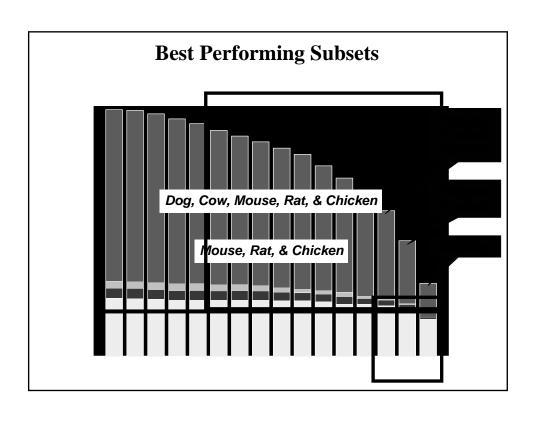


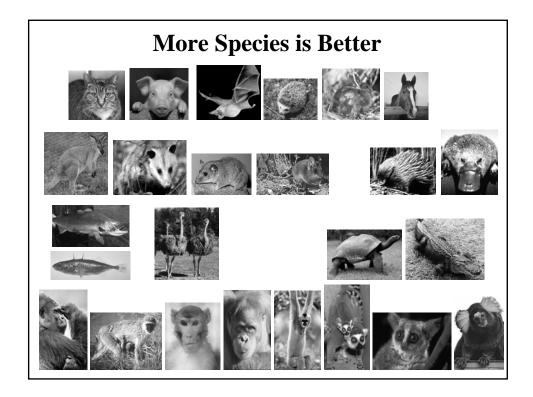


Detection of MCSs with Different Species

- Investigating the Relative Contribution of Different Species' Sequences to MCS Detection using More Quantitative Approaches
- Re-Compute Conservation Score for All* Possible Subsets of Species
- Compare to a 'Reference Set' of MCSs
 - Generated with All Species
 - Surrogates for Conserved Functional Elements







MCS Detection and Sequence Quality

- To date, MCS detection has been with reasonably high-quality sequence
- What quality of sequence is desired for MCS detection — especially provided a set of high-quality reference sequences?

MCS Detection and Sequence Quality

What Tradeoffs are encountered between sequence coverage vs. number of species?



- 1) Re-create 0.5X, 1X, 2X... Read-Coverage Datasets
- 2) Analyze for MCSs
- 3) Compare to "Finished" MCSs

