

## REVIEW

Liya Shen · Alexander Figurov · Bai Lu

**Recent progress in studies of neurotrophic factors and their clinical implications**

Received: 12 November 1996 / Accepted: 26 March 1997

**Abstract** Neurotrophic factors are endogenous soluble proteins that regulate long-term survival and differentiation of neurons of the peripheral and central nervous systems. These factors play an important role in the structural integrity of the nervous system, and therefore are good candidates as therapeutic agents for neurodegenera-

tive diseases. However, recent studies have revealed some unexpected, novel roles of neurotrophic factors. Of particular significance is the discovery of the new functions of brain-derived neurotrophic factor (BDNF) and glia-derived neurotrophic factor (GDNF). Physiological experiments indicate that BDNF may serve as regulatory factors for synaptic transmission as well as for learning and memory. Gene targeting studies demonstrate that GDNF may be essential for development of the enteric nervous system (ENS) and kidney organogenesis. These results not only provide new insights into our understanding of the function of neurotrophic factors but may also have significant implications in the therapeutic usages of neurotrophic factors.

**Key words** Brain-derived neurotrophic factor · trk receptor · Long-term potentiation · Glia-derived neurotrophic factor · c-ret tyrosine kinase · Dopaminergic neurons · Kidney organogenesis · Enteric nervous system · Gene knockout · Hirschsprung's disease

**Abbreviations** *ALS* Amyotrophic lateral sclerosis · *BDNF* Brain-derived neurotrophic factor · *CNTF* Ciliary neurotrophic factor · *ENS* Enteric nervous system · *GDNF* Glia-derived neurotrophic factor · *LIF* Leukemia inhibitory factor · *LTP* Long-term potentiation · *NGF* Nerve growth factor · *NT* Neurotrophin



**LIYA SHEN** received her Ph.D. degree in Molecular Biology from Columbia Univ. She is presently a National Research Council (NRC) fellow in the Laboratory of Mammalian Genes and Development, NICHD, NIH. Her research interests are: the use of gene targeting technology to study mammalian development, and establishment of molecular etiology of human diseases with mouse models.



**BAI LU** received his Ph.D. degree in Neurobiology from Cornell Univ. Medical College. He is presently the Chief of Unit on Synapse Development and Plasticity, National Institute of Child Health and Human Development (NICHD), NIH. His major research interests include: the role of neurotrophic factors in synapse development and plasticity, and characterization of new molecules involved in synaptogenesis.

L. Shen  
Laboratory of Mammalian Gene and Development, NICHD,  
National Institutes of Health, Bethesda, MD 28920-4480, USA

A. Figurov · Bai Lu (✉)  
Laboratory of Developmental Neurobiology, NICHD,  
National Institutes of Health, Bethesda, MD 28920-4480, USA

**Introduction**

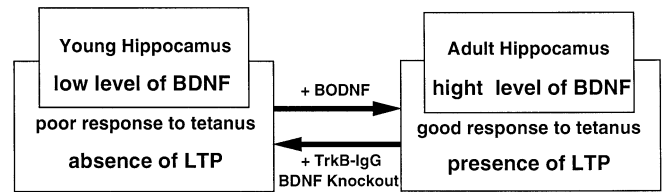
According to the classical definition, neurotrophic factors are endogenous soluble proteins that regulate the long-term survival and differentiation of neurons [1]. It is now generally accepted that they are signaling molecules important for both the development and the maintenance of structural integrity within the peripheral and central nervous systems. Extensive studies have been carried out, and significant progress has been made in the past decade. Many potent neurotrophic factors have been discovered. Their receptors have been identified, and great ef-

forts have been made to elucidate their signal transduction mechanisms. The expression and cellular distribution of these factors and their respective receptors in the nervous system have been extensively investigated, and these studies have provide clues for the cell populations that synthesize or respond to particular kinds of neurotrophic factors. Several approaches have been employed to determine the biological function of neurotrophic factors. In many cases new neurotrophic factors were identified using cultures of a specific population of neurons.

Since the microenvironment of neurons in culture is rigorously controlled, it is easy to test the factor's ability to enhance neuronal survival and differentiation and to protect neuronal death due to toxic agents or insults. The function of the neurotrophic factor can also be examined in animals *in vivo*. The factor is delivered to specific regions of the nervous system to determine its ability to rescue naturally occurring cell death (apoptosis) or to protect against experimentally induced lesions or damage. In recent years the new technology of gene targeting in mice has been used to study the function of neurotrophic factors [2]. In these studies the gene for a particular neurotrophic factor or its receptor has been mutated through homologous recombination in embryonic stem cells, and the consequence of the gene ablation is manifested in the homozygote offspring carrying the mutated alleles in the animal's genome. The mutant mice are studied using physiological, biochemical, morphological, and behavioral approaches. Thus these knockout mice are excellent animal models to study the function of neurotrophic factors.

Several classes of neurotrophic factors have been identified. The prototype is called neurotrophin, which is a family of structurally related secretory proteins that are widely expressed in neurons and their target cells [3, 4]. To date five members have been identified. They are called nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), and neurotrophin (NT) 3, 4/5, and 6. These are signaling molecules that function by binding to their respective receptors on the cell surface. NGF binds to TrkA; BDNF and NT-4/5 to TrkB, and NT-3 to TrkC. A second class of neurotrophic factors is the ciliary neurotrophic factor (CNTF) family, which includes CNTF, leukemia inhibitory factor (LIF), interleukin (IL) 6 and 11, and cardiotrophin 1 [5, 6]. Their receptors consist of several components, a common subunit for all members of CNTF family, and other subunits unique for their respective factors [7].

Recently a novel molecule called glia cell line-derived growth factor (GDNF) was discovered for its potent effect on the survival of mesencephalic dopaminergic neurons [8]. GDNF is a distant member of transforming growth factor  $\beta$  superfamily. However, unlike other members in the family, GDNF acts on receptor tyrosine kinase rather than serine/threonine kinases. The receptors for GDNF, similar to the CNTF family molecules, are comprised of multiple components. These include a signaling component called c-ret, an orphan receptor tyrosine kinase [9, 10], and a high-affinity ligand binding



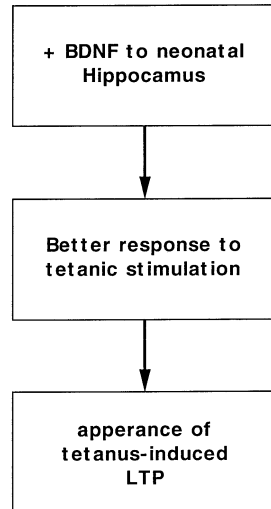
**Fig. 1** Role of BDNF in tetanus-induced LTP during hippocampal development

component, GDNFR- $\alpha$  [11, 12]. Additional members of the GDNF gene family and GDNFR- $\alpha$  are being cloned and characterized.

Since chronic neurodegenerative diseases and acute nervous system injury result in structural damage, there has also been a great deal of interest in the use of neurotrophic factors as therapeutic agents for diseases of the nervous system [13]. Substantial evidence indicates that neurotrophic factors can protect and even restore impaired functions resulting from trauma, aging, toxic agents, and genetically linked neurodegenerative disorders. Clinical trials are now in progress to assess the therapeutic efficacy of neurotrophic factors in a variety of neurodegenerative diseases. These clinical studies are based primarily on functional studies in animals. For example, in both rodents and monkeys GDNF has been shown to enhance the survival and protect against the death of dopaminergic neurons in substantia nigra neurons which project to the striatum [14–16]. These neurons are involved in motor function and are degenerated in patients with Parkinson's disease. GDNF therefore is an attractive candidate drug for Parkinson's disease, and clinical trials are underway. Thus the discovery of new functions for a neurotrophic factor may lead to new clinical use of the factor. On the other hand, new findings in animal studies may also provide insights into the potential side effects.

Despite rapid advances in research on neurotrophic factors functional studies have concentrated largely on their roles in neuronal survival and differentiation [1, 17]. However, the recurring observation that the expression of many neurotrophic factors in the central nervous system is rapidly enhanced by neuronal activity [18] suggests a new role for these factors in activity-dependent processes, such as synaptic development and plasticity [19]. Indeed, a number of recent experiments support this hypothesis. Moreover, expression of the receptors for the neurotrophic factors in several nonneuronal tissues raises the possibility that these factors may have a role outside the nervous system. These studies have challenged the classical view of neurotrophic factors, which defines neurotrophic factors as secretory proteins that regulate survival and differentiation of neurons in the central and peripheral nervous systems. This review focuses on the novel and unexpected functions of the neurotrophic factors BDNF and GDNF. These functions cannot be explained by the traditional concept of neurotrophic factors.

**Fig. 2** Mechanism by which BDNF regulates tetanus-induced LTP in developing hippocampus



### Role of BDNF in learning and memory

Substantial evidence indicates that the expression of neurotrophin genes in neurons can be regulated by neuronal activity. Activity-dependent regulation of NGF and BDNF mRNAs have been observed in hippocampal neurons in culture [20–23] and in the visual cortex in vivo [24]. Moreover, the levels of NGF and BDNF mRNA increase rapidly in response to kindling as well as recurrent limbic seizures and kainate-induced seizures [25–28]. A recent study demonstrated that in BDNF mutant mice the development but not maintenance of kindling is suppressed [29] (but see [30]). This result implies that BDNF facilitates kindling epileptogenesis in the hippocampus by enhancing synaptic transmission. For a long time it has been speculated that the rapid increase in neurotrophin gene expression may play a role in modulating synaptic transmission or synaptic plasticity [17, 31]. The first direct demonstration of neurotrophic regulation of synaptic activity was the acute effect of neurotrophins on synaptic transmission at the neuromuscular junction [32].

It has been reported that within a few minutes of exposure to the neurotrophins BDNF or NT-3 there is a dramatic increase in the frequency of spontaneous synaptic currents and in the amplitude of evoked synaptic currents. Detailed analyses indicate that this effect is a result of the enhancement of transmitter release, most likely due to increased intracellular calcium concentrations [33, 34]. The acute effects of neurotrophins on neuronal activity and synaptic transmission have also been observed in developing CNS neurons in culture by a number of laboratories [35–38]. These experiments have established that (a) the expression of neurotrophin genes can be regulated by neuronal activity, and (b) neurotrophic factors are capable of acutely enhancing synaptic transmission. The physiological significance of these findings has not been well established. A positive feedback hypothesis has been put forward: neuronal impulse

activity enhances the production and secretion of neurotrophins, which in turn serve as retrograde messengers to potentiate neuronal activity and synaptic efficacy [35].

This reciprocal interaction hypothesis strongly suggests a role for neurotrophins in activity-dependent processes such as learning and memory [35]. The best cellular model for learning and memory is the long-term potentiation (LTP) of synaptic efficacy in the hippocampus [39, 40]. The hippocampus is a brain structure known to be involved in the initial retention of information, because damage to this region interferes with the formation of new memory while old memories are not affected. Hippocampal LTP is induced by a brief tetanic stimulation, and the increase in synaptic efficacy can last for hours or even days. The LTP process can be divided into an initial “induction” stage and a later “maintenance” stage [39]. In both the CA1 region and the dentate gyrus of hippocampus the induction of LTP involves a sustained, tetanus-induced depolarization which causes a large influx of  $\text{Ca}^{2+}$  through NMDA-type glutamate receptors and  $\text{Ca}^{2+}$  channels. Subsequent biochemical processes triggered by this  $\text{Ca}^{2+}$  influx have been a subject of intensive studies.

Substantial evidence indicates that the long-term maintenance of LTP [41, 42], just as long-term memory [43], requires gene transcription and protein synthesis. Which genes are those turned on during LTP? The LTP-inducing tetanic stimulation has been shown to enhance the expression of neurotrophin genes, suggesting that neurotrophins play a role in LTP maintenance. In hippocampal slices the application of tetanus to Schaffer collaterals significantly increases BDNF and NT-3 mRNAs in CA1 neurons [44]. Tetanic stimulation of the perforant path in vivo also elicits an increase of mRNAs for NGF and BDNF in granular neurons of dentate gyrus [45, 46]. It has been speculated that neurotrophins are involved in the maintenance of LTP, either by modulating postsynaptic glutamate receptors or by serving as retrograde messengers that regulate presynaptic transmitter release.

If neurotrophins are truly important for LTP, deletion of neurotrophin genes should impair the LTP process. Indeed, in BDNF knockout mice there is a severe defect in the hippocampal LTP, although brain morphology, basal synaptic transmission, and the behavior of these animals appear to be normal [47, 48]. It appears that hippocampal BDNF must maintain a critical level for the expression of LTP. Adult heterozygotes in which the BDNF gene activity has been reduced to half exhibit the same degree of impairment in LTP as those from homozygous animals. However, in animals younger than P16, homozygotes show more severe impairment than heterozygotes, indicating that younger animals are more susceptible to changes in BDNF levels. The defect in hippocampal LTP can be rescued by introducing exogenous BDNF back into the hippocampus of BDNF knockout mice. LTP is completely restored in hippocampal slices from BDNF knockout mice after prolonged incubation with recombinant BDNF [48] or by infection with BDNF-containing adenovirus [49]. These experiments indicate

that the absence of BDNF per se, rather than cumulative developmental defects, is responsible for impaired LTP in BDNF knockout mice.

In the developing hippocampus the expression of BDNF and its receptor TrkB [50–53] increase in parallel with the ability to undergo LTP [54–56]. Interestingly, the ability to learn and remember also improves progressively throughout childhood. In a recent study BDNF was found to accelerate the appearance of tetanus-induced LTP in the developing hippocampus, suggesting that this neurotrophin promotes memory in young animals [57]. In neonatal hippocampal (p12–13) slices tetanic stimulation induced a typical short-term potentiation (STP) lasting less than 30–40 min. In contrast, the same tetanic stimulation elicited a stable LTP in slices treated with BDNF. The effect of BDNF in the neonatal hippocampus was specific. Neither NGF nor NT-3, which activate TrkA and TrkC receptor tyrosine kinases, respectively, had any effects on LTP development. Moreover, NT-4/5, which acts on the same TrkB receptor, had an effect similar to that of BDNF. The BDNF effect was prevented by pretreatment of the slices for 20–30 min with a TrkB-IgG fusion protein, a specific scavenger for BDNF and NT-4/5 [58]. Thus the BDNF effect was mediated by TrkB receptors. In the adult hippocampus, which normally expresses high levels of endogenous BDNF and TrkB, exogenous BDNF no longer facilitated LTP. Neutralizing BDNF activity by TrkB-IgG, however, significantly inhibited tetanus-induced LTP in the adult hippocampus [57].

These experiments are consistent with the studies using BDNF knockout mice and imply that the level of BDNF in the brain is important for tetanus-induced LTP. Application of exogenous BDNF promotes LTP in neonatal hippocampus where the endogenous BDNF level is low, while mutation of BDNF gene or reduction of BDNF activity by TrkB-IgG has the opposite effect in the adult hippocampus where the endogenous BDNF level is high (Fig. 1). Finally, the mechanism by which BDNF regulates hippocampal LTP has been found to be due to a significant enhancement of hippocampal synapses to respond to tetanic stimulation rather than to an alteration of the LTP triggering mechanism. Hippocampal synapses exhibited a severe synaptic depression when stimulated with repetitive, high-frequency stimulation. In hippocampal slices treated with BDNF, however, synaptic depression was significantly attenuated. These results suggest that the BDNF effect on LTP is mediated by an enhancement of the ability of hippocampal synapses to respond to high-frequency, tetanic stimulation (Fig. 2).

---

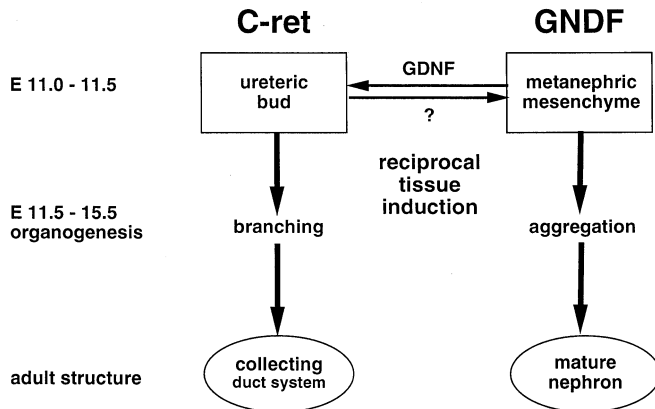
### **Role of GDNF in the development of kidney and enteric nervous system**

GDNF was first discovered for its ability to enhance the survival of midbrain dopaminergic neurons in culture [8]. Subsequent studies demonstrated that GDNF also has potent neurotrophic activities for many peripheral

neuronal populations [59–61]. However, the major focus of the functional studies of GDNF has been on the mid-brain dopaminergic neurons and spinal cord motoneurons, because these neurons are degenerated in patients with neurological disorders such as Parkinson's disease and amyotrophic lateral sclerosis (ALS). The degeneration of the nigrastratial dopaminergic system induced by mechanical lesions and toxic chemicals such as 1-methyl-4-phenylpyridinium and 6-hydroxydopamine is significantly attenuated by GDNF [14, 15, 62–68]. Moreover, a single administration of GDNF has been shown to rescue the motor deficits in a parkinsonian monkey model [16]. Extensive studies also indicate that GDNF prevents motoneuron cell death both during development and after axotomy in the adult [69–72]. These results suggest that GDNF is an important neurotrophic factor for brainstem dopaminergic neurons and spinal cord motoneurons, and raise the exciting possibility that GDNF may be used as a therapeutic agent for ALS and Parkinson's disease.

Recent characterization of GDNF null mutants has confirmed the role of GDNF in several ganglia of the peripheral nervous system: there is a significant reduction in the neuronal populations of the petrosal and nodose ganglia (40%), superior cervical ganglia (35%), and dorsal root ganglia (23%) in GDNF-deficient mice [73, 74]. However, the GDNF knockout experiments have generated a few surprises regarding GDNF functions during embryonic development [73–75]. First, the midbrain dopaminergic neurons and noradrenergic neurons of the locus ceruleus appear to be normal in the GDNF knockout mice. Despite the report that GDNF is a potent trophic factor for motoneurons so far discovered [70], there is only a 21% reduction of spinal lumbar motor nuclei [73, 74]. These results suggest that GDNF is not a physiological survival factor for these neurons *in vivo*. Second, the mutant animals do not form permanent kidneys. The homozygote mutants die within 24 h after birth, presumably due to kidney failure. Third, these mice completely lack the enteric nervous system (ENS). Thus the GDNF knockout study has taught us an important lesson: to fully understand the biological function of neurotrophic factors pharmacological experiments should be complemented by molecular genetic studies, and deletion mutations can uncover certain aspects of function which are otherwise difficult to reveal. The GDNF function during normal development in animals, as well as the molecular mechanism of GDNF action at the tissue and cell levels, needs to be understood in much more detail before a rational course of therapy can be charted.

The lack of defects in midbrain dopaminergic neurons in the GDNF knockout mice [73, 74] was unexpected, since this population responds dramatically to the treatment with exogenous GDNF. It is possible that there may be other GDNF-related trophic factors which compensate for the lack of GDNF function in the mutants. Alternatively, GDNF may not be a survival factor for the dopaminergic neurons but can mimic the function of other GDNF-like trophic factors. This is consistent with the



**Fig. 3** Role of GDNF during kidney development

recent discovery of a new member in the GDNF family, *nutrin*. The search for additional GDNF-like molecules is now underway.

The development of the kidney depends on sequential and reciprocal interactions of two primordial tissues, the ureteric bud and the metanephric mesenchyme (Fig. 3) [76]. The ureteric bud induces the metanephric mesenchyme to aggregate and proliferate, leading to the formation of the epithelium of the mature nephron [77]. Meanwhile, the ureteric bud itself is induced by the metanephric mesenchyme to proliferate and branch, ultimately generating the collecting duct system. The role of GDNF in rodent kidney development has in fact been implied by its expression, which coincides with the morphogenesis of the kidney. Metanephric tissues begin to differentiate at embryonic day 11 (E11), and by E13.5 high levels of GDNF expression are detected in condensing metanephric mesenchyme near the tip of ureter bud, the area of the prospective kidney nephrogenic tubules, and maintained through E15.5 (Fig. 3) [78–81]. Soon after epithelization kidney tubules cease to express GDNF. The GDNF knockout studies demonstrate that GDNF is indeed a metanephric mesenchyme-derived factor that induces ureteric bud formation and branching (Fig. 3) [73–75]. Furthermore, the growth or arborization of the ureteric bud of the GDNF mutants is dramatically stimulated by exogenous GDNF in a dose-dependent manner during early stages of kidney morphogenesis [75].

In normal mice *in situ* hybridization experiments have shown abundant expression of GDNF mRNA in the outer mesenchymal layer of the entire developing gastrointestinal tract [73, 80, 81]. GDNF null mice exhibit a dilation of the proximal intestine and a severe occlusion of the pyloric sphincter, suggesting a defect in those innervations that coordinate control of the gastrointestinal motility. Detailed analysis indicate the complete lack of the neural crest derived enteric nervous system, beginning as early as E12.5 [73–75]. Preliminary experiments have found that GDNF enhances the differentiation and fiber outgrowth of the enteric neurons in a dose-dependent manner (unpublished results). These data indicate that GDNF is an important factor for the proper develop-

ment of enteric neurons in the digestive tract. The gastrointestinal defect in the GDNF mutant resembles a human genetic disorder, Hirschsprung's disease (HSCR), named after the Danish physician who first described the disorder in 1887 (for review, see [82]). This disease is characterized by absence of intrinsic enteric ganglion cells in the myenteric and submucosal plexuses of the gut, leading to megacolon formation [83, 84]. Interestingly, some autosomal dominant forms of Hirschsprung's disease have been associated with mutations in the human *RET* locus [85, 86]. Furthermore, mice defective in the *c-ret* gene have phenotypes almost identical to the GDNF mutants. The *c-ret* homozygous mutants do not form the ureteric bud or undergo metanephric development and lack ENS neurons [87].

The striking similarity between the *c-ret* and GDNF knockouts in both the gastrointestinal and the kidney phenotypes suggests that *c-ret* may mediate GDNF signaling. Indeed, *c-ret*, which was initially discovered as an orphan receptor tyrosine kinase [88], has recently been found to be a critical component of the receptor signaling system for GDNF [9, 10]. Moreover, expression cloning has recently identified GDNFR- $\alpha$  as the ligand binding subunit of the receptor complex [11, 12]. Biochemical evidence revealed that *c-ret* is an integral signal transducing component for the GDNF signaling mechanism. GDNF binds with high affinity to GDNFR- $\alpha$ , which in turn recruits *c-ret* into the ligand-receptor complex and induces tyrosine phosphorylation of *c-ret* [11, 12]. The *c-ret* receptor tyrosine kinase is expressed in cell populations that respond to GDNF, including mid-brain dopaminergic neurons, motoneurons, ureter bud cells, and the enteric ganglioblasts [89]. Identification of the GDNF signaling pathway will facilitate our understanding of the molecular and cellular mechanisms of GDNF action in diverse systems.

### Clinical implications of the novel functions of neurotrophic factors

Recent studies reviewed here indicate that in addition to its classic role in the survival and differentiation of motoneurons and sensory neurons the neurotrophin BDNF may regulate learning and memory, particularly during development. The clinical implications of these findings are twofold. First, one must take this novel BDNF function into consideration when BDNF is considered as a therapeutic agent for the treatment of neurological disorders. For example, a number of studies *in vitro* and *in vivo*, including a variety of lesion and toxin models have indicated that BDNF promotes the survival and differentiation of mesencephalic dopaminergic neurons in substantia nigra. When considering a BDNF therapy for Parkinson's disease, it is important to remember that BDNF may also alter the cognitive functions if delivered into the brain, and the side effect may be more severe in children than adults. Currently the only clinical trial for BDNF is on its effect on the motoneuron disease

ALS. Since BDNF cannot cross the blood-brain barrier, muscle injection of BDNF may not interfere with learning and memory and other cognitive functions. Second, regardless of its mechanism, the effect of BDNF on learning and memory also raises the possibility of activating the BDNF/TrkB system as a potential therapeutic strategy for learning disorders in children and adults.

Since its first discovery GDNF has promised to be a therapeutic agent for the treatment of neurodegenerative diseases, particularly Parkinson's disease and motoneuron diseases such as ALS [90, 91]. In addition to the data in rodents, recent experiments have demonstrated that a single administration of GDNF dramatically corrects the motor deficits in a parkinsonian monkey model [16]. These studies have formed the basis for the initiation of a clinical trial to test the therapeutic role of GDNF in Parkinson's diseases. However, the recent work on GDNF knockout mice has revealed an unexpected function of GDNF in the development of the kidneys and the enteric nervous system. Thus caution must be exercised with the clinical use of GDNF. In motoneuron diseases the side effects of systemic or muscle delivery of GDNF on the overgrowth of kidney and ENS must be tested. For treatment of Parkinson's disease further experiments are required to determine whether CNS delivery of GDNF would cause any side effects in the periphery. On the other hand, since severe defects in kidney and ENS development are observed in mice with GDNF mutation, one can envisage the potential therapeutic uses of GDNF both for defects of renal dysgenesis and for Hirschsprung's disease.

**Acknowledgements** We thank the past and current members of the laboratory for their enthusiasm and support, and Dr. Millicent Dugich-Djordjevic for discussions and comments, and Ms. Mary Mood for assistance in preparing the manuscript.

## References

1. Barde Y (1989) Trophic factors and neuronal survival. *Neuron* 2:1525–1534
2. Snider WD (1994) Functions of neurotrophins during nervous system development: what the knockouts are teaching us. *Cell* 77:627–636
3. Korsching S (1993) The neurotrophic factor concept: a reexamination. *J Neurosci* 13:2739–2748
4. Gotz R, Koster R, Winkler C, Raulf F, Lottspeich F, Scharl M, Thoenen H (1994) Neurotrophin-6 is a new member of the nerve growth factor family. *Nature* 372:266–269
5. Sendtner M, Carroll P, Holtmann B, Hughes RA, Thoenen H (1994) Ciliary neurotrophic factor. *J Neurobiol* 25:1436–1453
6. Pennica D, Arce V, Swanson TA, Vejsada R, Pollock RA, Armanini M, Dudley K (1996) Cardiotrophin-1, a cytokine present in embryonic muscle, supports long-term survival of spinal motoneurons. *Neuron* 17:63–74
7. Stahl N, Yancopoulos GD (1994) The tripartite CNTF receptor complex: activation and signaling involves components shared with other cytokines. *J Neurobiol* 25:1454–1466
8. Lin LF, Doherty DH, Lile JD, Bektess S, Collins F (1993) GDNF: a glial cell line-derived neurotrophic factor for mid-brain dopaminergic neurons. *Science* 260:1130–1132
9. Durbec P, Marcos GC, Kilkenny C, Grigoriou M, Wartiovaara K, Suvanto P, Smith D (1996) GDNF signalling through the Ret receptor tyrosine kinase. *Nature* 381:789–793
10. Trupp M, Arenas E, Fainzilber M, Nilsson AS, Sieber BA, Grigoriou M, Kilkenny C (1996) Functional receptor for GDNF encoded by the c-ret proto-oncogene. *Nature* 381:785–788
11. Treanor JJ, Goodman L, de SF, Stone DM, Poulsen KT, Beck CD, Gray C (1996) Characterization of a multicomponent receptor for GDNF. *Nature* 382:80–83
12. Jing S, Wen D, Yu Y, Holst PL, Luo Y, Fang M, Tamir R (1996) GDNF-induced activation of the ret protein tyrosine kinase is mediated by GDNFR-alpha, a novel receptor for GDNF. *Cell* 85:1113–1124
13. Hefti F (1994) Neurotrophic factor therapy for nervous system degenerative diseases. *J Neurobiol* 25:1418–1435
14. Beck KD, Valverde J, Alexi T, Poulsen K, Moffat B, Vandlen RA, Rosenthal A (1995) Mesencephalic dopaminergic neurons protected by GDNF from axotomy-induced degeneration in the adult brain. *Nature* 373:339–341
15. Tomac A, Lindqvist E, Lin LF, Ogren SO, Young D, Hoffer BJ, Olson L (1995) Protection and repair of the nigrostriatal dopaminergic system by GDNF in vivo. *Nature* 373:335–339
16. Gash DM, Zhang Z, Ovadia A, Cass WA, Yi A, Simmerman L, Russell D (1996) Functional recovery in parkinsonian monkeys treated with GDNF. *Nature* 380:252–255
17. Thoenen H (1991) The changing scene of neurotrophic factors. *Trends Neurosci* 14:165–170
18. Isackson PJ (1995) Trophic factor response to neuronal stimuli or injury. *Curr Opin Neurobiol* 5:350–357
19. Thoenen H (1995) Neurotrophins and neuronal plasticity. *Science* 270:593–596
20. Zafra F, Hengerer B, Leibrock J, Thoenen H, Lindholm D (1990) Activity-dependent regulation of BDNF and NGF mRNA in the rat hippocampus is mediated by non-NMDA glutamate receptors. *EMBO J* 9:3545–3550
21. Lu B, Yokoyama M, Dreyfus CF, Black IB (1991) Depolarizing stimuli regulate NGF mRNA expression in cultured hippocampal neurons. *Proc Natl Acad Sci USA* 88:6289–6292
22. Zafra F, Castren E, Thoenen H, Lindholm D (1991) Interplay between glutamate and  $\gamma$ -aminobutyric acid transmitter systems in the physiological regulation of brain-derived neurotrophic factor and nerve growth factor synthesis in hippocampal neurons. *Proc Natl Acad Sci USA* 88:10037–10041
23. Zafra F, Lindholm D, Castren E, Hartikka J, Thoenen H (1992) Regulation of Brain-derived neurotrophic factor and nerve growth factor mRNA in primary cultures of hippocampal neurons and astrocytes. *J Neurosci* 12:4793–4799
24. Castren E, Zafra F, Theonen H, Lindholm D (1992) Light regulates expression of brain-derived neurotrophic factor mRNA in the rat visual cortex. *Proc Natl Acad Sci USA* 89:9444–9448
25. Gall CM, Isackson PJ (1989) Limbic seizures increase neuronal production of messenger RNA for nerve growth factor. *Science* 245:758–761
26. Isackson PJ, Huntsman MM, Murray KD, Gall CM (1991) BDNF mRNA expression is increased in adult rat forebrain after limbic seizures: temporal patterns of induction distinct from NGF. *Neuron* 6:937–948
27. Ernfors P, Bengzon J, Kokaia Z, Persson H, Lindvall O (1991) Increased levels of messenger RNAs for neurotrophic factors in the brain during kindling epileptogenesis. *Neuron* 7:165–176
28. Dugich-Djordjevic MM, Tocco G, Willoughby DA, Najm I, Pasinetti G, Thompson R, Baudry M (1992) BDNF mRNA expression in the developing rat brain following kainic acid-induced seizure activity. *Neuron* 8:1127–1138
29. Kokaia M, Ernfors P, Kokaia Z, Elmer E, Jaenisch R, Lindvall O (1995) Suppressed epileptogenesis in BDNF mutant mice. *Exp Neurol* 133:215–24
30. Larmet Y, Reibel S, Carnahan J, Nawa H, Marescaux C, Depaulis A (1995) Protective effects of brain-derived neurotrophic factor on the development of hippocampal kindling in the rat. *Neuroreport* 6:1937–41
31. Lo DC (1992) Signal transduction and regulation of neurotrophins. *Curr Opin Neurobiol* 2:336–340

32. Lohof AM, Ip NY, Poo MM (1993) Potentiation of developing neuromuscular synapses by the neurotrophins NT-3 and BDNF. *Nature* 363:350–353
33. Stoop R, Poo MM (1995) Potentiation of transmitter release by ciliary neurotrophic factor requires somatic signaling. *Science* 267:695–699
34. Stoop R, Poo MM (1996) Synaptic modulation by neurotrophic factors: differential and synergistic effects of brain-derived neurotrophic factor and ciliary neurotrophic factor. *J Neurosci* 16:3256–3264
35. Kim HG, Wang T, Olafsson P, Lu B (1994) Neurotrophin 3 potentiates neuronal activity and inhibits  $\gamma$ -aminobutyrate synaptic transmission in cortical neurons. *Proc Natl Acad Sci USA* 91:12341–12345
36. Knipper M, Leung LS, Zhao D, Rylett RJ (1994) Short-term modulation of glutamatergic synapses in adult rat hippocampus by NGF. *Neuroreport* 5:2433–2436
37. Lessmann V, Gottmann K, Heumann R (1994) BDNF and NT-4/5 enhance glutamatergic synaptic transmission in cultured hippocampal neurones. *Neuroreport* 6:21–25
38. Levine ES, Dreyfus CF, Black IB, Plummer MR (1995) Brain-derived neurotrophic factor rapidly enhances synaptic transmission in hippocampal neurons via postsynaptic tyrosine kinase receptors. *Proc Natl Acad Sci USA* 92:8074–8077
39. Bliss TVP, Collingridge GL (1993) A synaptic model of memory: long-term potentiation in the hippocampus. *Nature* 361:31–39
40. Malenka RC, Nicoll RA (1993) NMDA-receptor-dependent synaptic plasticity: multiple forms and mechanisms. *Trends Neurosci* 16:521–527
41. Frey U, Krug M, Reymann KG, Mathies H (1984) Anisomycin, an inhibitor of protein synthesis, blocks late phases of LTP-phenomena in the CA-region in vitro. *Brain Res* 452:57–65
42. Nguyen PT, Abel T, Kendal ER (1994) Requirement of a critical period of transcription for induction of a later phase of LTP. *Science* 265:1104–1107
43. Davies H, Squire L (1984) Protein synthesis and memory: a review. *Psychol Bull* 96:518–599
44. Patterson S, Grover LM, Schwartzkroin PA, Bothwell M (1992) Neurotrophin expression in rat hippocampal slices: a stimulus paradigm inducing LTP in CA1 evokes increases in BDNF and NT-3 mRNAs. *Neuron* 9:1081–1088
45. Castren E, Pitkanen M, Sirvio J, Parsadanian A, Lindholm D, Thoenen H, Riekkinen PJ (1993) The induction of LTP increases BDNF and NGF mRNA but decreases NT-3 mRNA in the dentate gyrus. *Neuroreport* 4:895–898
46. Bramham CR, Southard T, Sarvey J, Herkenham M, Brady LS (1996) Unilateral LTP triggers bilateral increases in hippocampal neurotrophin and trk receptor mRNA expression in behaving rats: evidence for interhemispheric communication. *J Comp Neurol* 368:371–382
47. Korte M, Carroll P, Wolf E, Brem G, Thoenen H, Bonhoeffer T (1995) Hippocampal long-term potentiation is impaired in mice lacking brain-derived neurotrophic factor. *Proc Natl Acad Sci USA* 92:8856–8860
48. Patterson SL, Abel T, Deuel TA, Martin KC, Rose JC, Kandel ER (1996) Recombinant BDNF rescues deficits in basal synaptic transmission and hippocampal LTP in BDNF knockout mice. *Neuron* 16:1137–1145
49. Korte M, Griesbeck O, Gravel C, Carroll P, V. Staiger, Thoenen H, Bonhoeffer T (1996) Virus-mediated gene transfer into hippocampal CA1 region restores long-term potentiation in brain-derived neurotrophic factor mutant mice. *Proc Natl Acad Sci USA* 93:12547–12552
50. Maisonpierre PC, Belluscio L, Friedman B, Alderson RF, Wigand SJ, Furth ME, Lindsay RM (1990) NT-3, BDNF, and NGF in the developing rat nervous system: Parallel as well as reciprocal patterns of expression. *Neuron* 5:501–509
51. Friedman WJ, Olson L, Persson H (1991) Cells that express brain-derived neurotrophic factor mRNA in the developing postnatal rat brain. *Eur J Neurosci* 3:688–697
52. Ringstedt T, Kagercrantz H, Persson H (1993) Expression of members of the trk family in the developing postnatal rat brain. *Dev Brain Res* 72:119–131
53. Dugich-Djordjevic MM, Ohsawa F, Hefti F (1993) Transient elevation in catalytic trkB mRNA during postnatal development of the rat brain. *NeuroReport* 4:1091–1094
54. Harris KM, Teyler TJ (1984) Developmental onset of long-term potentiation in area CA1 of the rat hippocampus. *J Physiol* 346:27–48
55. Muller D, Oliver M, Lynch G (1989) Developmental changes in synaptic properties in hippocampus of neonatal rats. *Dev Brain Res* 49:105–114
56. Jackson PS, Suppes T, Harris KM (1993) Stereotypical changes in the pattern and duration of long-term potentiation expressed at postnatal day 11 and 15 in the rat hippocampus. *J Neurophysiol* 70:1412–1419
57. Figurov A, Pozzo ML, Olafsson P, Wang T, Lu B (1996) Regulation of synaptic responses to high-frequency stimulation and LTP by neurotrophins in the hippocampus. *Nature* 381:706–709
58. Shelton DL, Sutherland J, Gripp J, Camerato T, Armanini MP, Phillips HS, Carroll K (1995) Human trks: molecular cloning, tissue distribution, and expression of extracellular domain immunoadhesins. *J Neurosci* 15:477–491
59. Ebendal T, Tomac A, Hoffer BJ, Olson L (1995) Glial cell line-derived neurotrophic factor stimulates fiber formation and survival in cultured neurons from peripheral autonomic ganglia. *J Neurosci Res* 40:276–284
60. Trupp M, Ryden M, Jornvall H, Funakoshi H, Timmusk T, Arenas E, Ibanez CF (1995) Peripheral expression and biological activities of GDNF, a new neurotrophic factor for avian and mammalian peripheral neurons. *J Cell Biol* 130:137–148
61. Buj-Bello A, Buchman VL, Horton A, Rosenthal A, Davies AM (1995) GDNF is an age-specific survival factor for sensory and autonomic neurons. *Neuron* 15:821–828
62. Hoffer BJ, Hoffman A, Bowenkamp K, Huettl P, Hudson J, Martin D, Lin LF (1994) Glial cell line-derived neurotrophic factor reverses toxin-induced injury to midbrain dopaminergic neurons in vivo. *Neurosci Lett* 182:107–111
63. Bowenkamp KE, Hoffman AF, Gerhardt GA, Henry MA, Bidle PT, Hoffer BJ, Granholm AC (1995) Glial cell line-derived neurotrophic factor supports survival of injured midbrain dopaminergic neurons. *J Comp Neurol* 355:479–489
64. Kriegstein K, Suter CC, Fischer WH, Unsicker K (1995) TGF-beta superfamily members promote survival of midbrain dopaminergic neurons and protect them against MPP+ toxicity. *EMBO J* 14:736–742
65. Kearns CM, Gash DM (1995) GDNF protects nigral dopamine neurons against 6-hydroxydopamine in vivo. *Brain Res* 672:104–111
66. Lindner MD, Winn SR, Baetge EE, Hammang JP, Gentile FT, Doherty E, McDermott PE (1995) Implantation of encapsulated catecholamine and GDNF-producing cells in rats with unilateral dopamine depletions and parkinsonian symptoms. *Exp Neurol* 132:62–76
67. Sauer H, Rosenblad C, Bjorklund A (1995) Glial cell line-derived neurotrophic factor but not transforming growth factor beta 3 prevents delayed degeneration of nigral dopaminergic neurons following striatal 6-hydroxydopamine lesion. *Proc Natl Acad Sci USA* 92:8935–8939
68. Hou JG, Lin LF, Mytilineou C (1996) Glial cell line-derived neurotrophic factor exerts neurotrophic effects on dopaminergic neurons in vitro and promotes their survival and regrowth after damage by 1-methyl-4-phenylpyridinium. *J Neurochem* 66:74–82
69. Zurn AD, Baetge EE, Hammang JP, Tan SA, Aebischer P (1994) Glial cell line-derived neurotrophic factor (GDNF), a new neurotrophic factor for motoneurons. *Neuroreport* 6:113–118
70. Henderson CE, Phillips HS, Pollock RA, Davies AM, Lemeulle C, Armanini M, Simmons L (1994) GDNF: a potent survival factor for motoneurons present in peripheral nerve and muscle. *Science* 266:1062–1064

71. Yan Q, Matheson C, Lopez OT (1995) In vivo neurotrophic effects of GDNF on neonatal and adult facial motor neurons. *Nature* 373:341–344
72. Oppenheim RW, Houenou LJ, Johnson JE, Lin LF, Li L, Lo AC, Newsome AL (1995) Developing motor neurons rescued from programmed and axotomy-induced cell death by GDNF. *Nature* 373:344–346
73. Moore MW, Klein RD, Farinas I, Sauer H, Armanini M, Phillips H, Reichardt LF (1996) Renal and neuronal abnormalities in mice lacking GDNF. *Nature* 382:76–79
74. Sanchez MP, Silos SI, Frisen J, He B, Lira SA, Barbacid M (1996) Renal agenesis and the absence of enteric neurons in mice lacking GDNF. *Nature* 382:70–73
75. Pichel JG, Shen L, Sheng HZ, Granholm AC, Drago J, Grinberg A, Lee EJ (1996) Defects in enteric innervation and kidney development in mice lacking GDNF. *Nature* 382:73–76
76. Saxen L (ed) (1987) *Organogenesis of the kidney*. Cambridge University Press, Cambridge
77. Ekblom P (1989) Developmentally regulated conversion of mesenchyme to epithelium. *FASEB J* 3:2141–2150
78. Suter CC, Unsicker K (1994) GDNF is expressed in two forms in many tissues outside the CNS. *Neuroreport* 5:2486–2488
79. Trupp M, Ryden M, Jornvall H, Funakoshi H, Timmusk T, Arenas E, Ibanez C (1995) Peripheral expression and biological activities of GDNF, a new neurotrophic factor for avian and mammalian peripheral neurons. *J Cell Biol* 130:137–148
80. Hellmich HL, Kos L, Cho ES, Mahon KA, Zimmer A (1996) Embryonic expression of glial cell-line derived neurotrophic factor (GDNF) suggests multiple developmental roles in neural differentiation and epithelial-mesenchymal interactions. *Mech Dev* 54:95–105
81. Suvanto P, Hiltunen JO, Arumae U, Moshnyakov M, Sariola H, Sainio K, Saarma M (1996) Localization of glial cell line-derived neurotrophic factor (GDNF) mRNA in embryonic rat by in situ hybridization. *Eur J Neurosci* 18:101–107
82. Badner JA, Sieber WK, Garver KL, Chakravarti A (1990) A genetic study of Hirschsprung disease. *Am J Hum Genet* 46:568–580
83. Bolande RP (1975) Animal model of human disease. Hirschsprung's disease, aganglionic or hypoganglionic megacolon; animal model: aganglionic megacolon in piebald and spotted mutant mouse strains. *Am J Path* 79:189–192
84. Case D (1986) Hirschsprung's disease: a historical review. *Prog Pediatr Surg* 20:199–214
85. Romeo G, Ronchetto P, Luo Y, Barone V, Seri M, Ceccherini I, Pacini B (1994) Point mutations affecting the tyrosine kinase domain of the RET protooncogene in Hirschsprung's disease. *Nature* 367:377–378
86. Edery P, Lyonnet S, Mulligan L, Palet A, Dew E, Abel L, Holder S (1994) Mutations of the RET proto-oncogene in Hirschsprung's disease. *Nature* 367:378–380
87. Schuchardt A, D'Agati V, Larsson-Blomberg L, Costantini F, Pachnis V (1994) Defects in the kidney and enteric nervous system of mice lacking the tyrosine kinase receptor Ret. *Nature* 367:380–383
88. Takahashi M, Buma Y, Iwamoto T, Inaguma Y, Ikehara H, Hiai H (1988) Cloning and expression of the ret protooncogene encoding a tyrosine kinase with two potential transmembrane domains. *Oncogene* 3:571–578
89. Pachnis V, Mankoo B, Constantini F (1993) Expression of the c-ret proto-oncogene during mouse embryogenesis. *Development* 119:1005–1007
90. Lindsay RM, Altar CA, Cedarbaum JM, Hyman C, Wiegand SJ (1993) The therapeutic potential of neurotrophic factors in the treatment of Parkinson's disease. *Exp Neurol* 124:103–118
91. Thoenen H, Hughes RA, Sendtner M (1993) Trophic support of motoneurons: physiological pathophysiological, and therapeutic implications. *Exp Neurol* 124:47–55