Phospho-p70 S6 Kinase (Thr389) (108D2) Rabbit mAb

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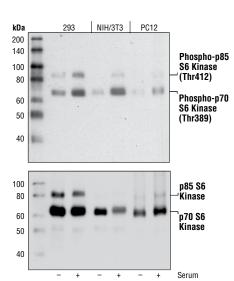
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Background: p70 S6 kinase is a mitogen activated Ser/Thr protein kinase that is required for cell growth and G1 cell cycle progression (1,2), p70 S6 kinase phosphorylates the S6 protein of the 40S ribosomal subunit and is involved in translational control of 5' oligopyrimidine tract mRNAs (1). A second isoform, p85 S6 kinase, is derived from the same gene and is identical to p70 S6 kinase except for 23 extra residues at the amino-terminus, which encode a nuclear localizing signal (1). Both isoforms lie on a mitogen activated signaling pathway downstream of phosphoinositide-3 kinase (PI-3K) and the target of rapamycin, FRAP/mTOR, a pathway distinct from the Ras/MAP kinase cascade (1). The activity of p70 S6 kinase is controlled by multiple phosphorylation events located within the catalytic, linker and pseudosubstrate domains (1). Phosphorylation of Thr229 in the catalytic domain and Thr389 in the linker domain are most critical for kinase function (1). Phosphorylation of Thr389, however, most closely correlates with p70 kinase activity in vivo (3). Prior phosphorylation of Thr389 is required for the action of phosphoinositide 3-dependent protein kinase 1 (PDK1) on Thr229 (4.5). Phosphorylation of this site is stimulated by growth factors such as insulin, EGF and FGF, as well as by serum and some G-proteincoupled receptor ligands, and is blocked by wortmannin, LY294002 (PI-3K inhibitor) and rapamycin (FRAP/mTOR inhibitor) (1,6,7). Ser411, Thr421 and Ser424 lie within a Ser-Pro-rich region located in the pseudosubstrate region (1). Phosphorylation at these sites is thought to activate p70 S6 kinase via relief of pseudosubstrate suppression (1,2). Another LY294002 and rapamycin sensitive phosphorylation site, Ser371, is an in vitro substrate for mTOR and correlates well with the activity of a partially rapamycin resistant mutant p70 S6 kinase (8).

Specificity/Sensitivity: Phospho-p70 S6 Kinase (Thr389) (108D2) Rabbit mAb detects endogenous levels of p70 S6 kinase only when phosphorylated at Thr389. This antibody also detects p85 S6 kinase when phosphorylated at the analogous site (Thr412) and possibly S6KII phosphorylated at Thr401.

Source/Purification: Monoclonal antibodies are produced by immunizing rabbits with a synthetic phospho-peptide (KLH-coupled) corresponding to residues surrounding Thr389 of human p70 S6 kinase.



Western blot analysis of extracts from serum starved or serum treated (20%) 293, NIH/3T3, and PC12 cells, using Phosphop70 S6 Kinase (Thr389) (108D2) Rabbit mAb (upper), or p70 S6 Kinase (49D7) rabbit mAb #2708 (lower).

Background References:

- (1) Pullen, N. and Thomas, G. (1997) FEBS Lett. 410, 78-82
- (2) Dunfer, A. et al. (1999) Exp. Cell Res. 253, 100-109.
- (3) Weng, Q.P. et al. (1998) J. Biol. Chem. 273, 16621-16629
- (4) Pullen, N. et al. (1998) Science 279, 707-710.
- (5) Alessi, D. et al. (1998) Curr. Biol. 8, 69-81.
- (6) Polakiewicz, R.D. et al. (1998) J. Biol. Chem. 273, 23534-23541.
- (7) Finger, D.C. et al. (2002) Genes Dev. 16, 1472-1487.
- (8) Saitoh, M. et al. (2002) J. Biol. Chem. 277, 20104-20112.

Storage: Supplied in 10 mM sodium HEPES (pH 7.5), 150 mM NaCl, 100 µg/ml BSA and 50% glycerol. Store at -20°C. Do not aliquot the antibody.

Recommended Antibody Dilutions:

Western Blotting

Companion Products:

PhosphoPlus® p70 S6 Kinase (Thr389, Thr421/Ser424) Antibody Kit #9430

Phospho-p70 S6 Kinase (Ser371) Antibody #9208

Phospho-p70 S6 Kinase (Thr389) Antibody #9205

Phospho-p70 S6 Kinase (Thr389) (1A5) Mouse mAb #9206 Phospho-Drosophila p70 S6 Kinase (Thr398) Antibody

Phospho-p70 S6 Kinase (Thr421/Ser424) Antibody #9204

p70 S6 Kinase Antibody #9202

p70 S6 Kinase (49D7) Rabbit mAb #2708

p70 S6 Kinase Control Cell Extracts #9203

Phototope®-HRP Western Blot Detection System, Anti-rabbit IgG, HRP-linked Antibody #7071

Anti-rabbit IgG, HRP-linked Antibody #7074

Prestained Protein Marker, Broad Range (Premixed Format)

Biotinylated Protein Ladder Detection Pack #7727

LumiGLO® Reagent and Peroxide #7003

IMPORTANT: For Western blots, incubate membrane with diluted antibody in 5% w/v BSA, 1X TBS, 0.1% Tween-20 at 4°C with gentle shaking, overnight.

Applications Kev: W—Western IP—Immunoprecipitation

IHC—Immunohistochemistry IC—Immunocytochemistry

IF-Immunofluorescence Mi-mink C-chicken X-Xenopus

F—Flow cytometry E—FLISA Z—zebra fish All—all species expected

D-DFI FIA®



Western Immunoblotting Protocol (Primary Ab Incubation In BSA)

For Western blots, incubate membrane with diluted antibody in 5% w/v BSA, 1X TBS, 0.1% Tween-20 at 4°C with gentle shaking, overnight.

A Solutions and Reagents

NOTE: Prepare solutions with Milli-Q or equivalently purified water.

- 1. 1X Phosphate Buffered Saline (PBS)
- 1X SDS Sample Buffer: 62.5 mM Tris-HCI (pH 6.8 at 25°C), 2% w/v SDS, 10% glycerol, 50 mM DTT, 0.01% w/v bromophenol blue or phenol red
- 3. Transfer Buffer: 25 mM Tris base, 0.2 M glycine, 20% methanol (pH 8.5)
- 10X Tris Buffered Saline (TBS): To prepare 1 liter of 10X TBS: 24.2 g Tris base, 80 g NaCl; adjust pH to 7.6 with HCl (use at 1X).
- **5.** Nonfat Dry Milk (weight to volume [w/v])
- Blocking Buffer: 1X TBS, 0.1% Tween-20 with 5% w/v nonfat dry milk; for 150 ml, add 15 ml 10X TBS to 135 ml water, mix. Add 7.5 g nonfat dry milk and mix well. While stirring, add 0.15 ml Tween-20 (100%).
- 7. Wash Buffer: 1X TBS, 0.1% Tween-20 (TBS/T)
- 8. Bovine Serum Albumin (BSA)
- Primary Antibody Dilution Buffer: 1X TBS, 0.1% Tween-20 with 5% BSA; for 20 ml, add 2 ml 10X TBS to 18 ml water, mix. Add 1.0 g BSA and mix well. While stirring, add 20 µl Tween-20 (100%).
- 10. Phototope®-HRP Western Blot Detection System #7071: Includes biotinylated protein ladder, secondary anti-rabbit (#7074) antibody conjugated to horseradish peroxidase (HRP), anti-biotin antibody conjugated to HRP, LumiGLO® chemiluminescent reagent and peroxide.
- 11. Prestained Protein Marker, Broad Range (Premixed Format) #7720
- **12.** Biotinylated Protein Ladder Detection Pack #7727
- Blotting Membrane: This protocol has been optimized for nitrocellulose membranes, which CST recommends. PVDF membranes may also be used.

B Protein Blotting

A general protocol for sample preparation is described below.

- 1. Treat cells by adding fresh media containing regulator for desired time.
- 2. Aspirate media from cultures; wash cells with 1X PBS; aspirate.
- 3. Lyse cells by adding 1X SDS sample buffer (100 µl per well of 6-well plate or 500 µl per plate of 10 cm diameter plate). Immediately scrape the cells off the plate and transfer the extract to a microcentrifuge tube. Keep on ice.
- 4. Sonicate for 10–15 seconds to shear DNA and reduce sample viscosity.
- **5.** Heat a 20 μ l sample to 95–100°C for 5 minutes; cool on ice.
- 6. Microcentrifuge for 5 minutes.
- 7. Load 20 µl onto SDS-PAGE gel (10 cm x 10 cm).

NOTE: CST recommends loading prestained molecular weight markers (#7720, 10 μ I/lane) to verify electrotransfer and biotinylated protein ladder (#7727, 10 μ I/lane) to determine molecular weights.

8. Electrotransfer to nitrocellulose or PVDF membrane.

C Membrane Blocking and Antibody Incubations

NOTE: Volumes are for 10 cm x 10 cm (100 cm²) of membrane; for different sized membranes, adjust volumes accordingly.

- (Optional) After transfer, wash nitrocellulose membrane with 25 ml TBS for 5 minutes at room temperature.
- 2. Incubate membrane in 25 ml of blocking buffer for 1 hour at room temperature.
- 3. Wash three times for 5 minutes each with 15 ml of TBS/T.
- Incubate membrane and primary antibody (at the appropriate dilution) in 10 ml primary antibody dilution buffer with gentle agitation <u>overnight</u> at 4°C.
- 5. Wash three times for 5 minutes each with 15 ml of TBS/T.
- 6. Incubate membrane with HRP-conjugated secondary antibody (1:2000) and HRP-conjugated anti-biotin antibody (1:1000) to detect biotinylated protein markers in 10 ml of blocking buffer with gentle agitation for 1 hour at room temperature.
- 7. Wash three times for 5 minutes each with 15 ml of TBS/T.

D Detection of Proteins

 Incubate membrane with 10 ml LumiGL0® (0.5 ml 20X LumiGL0®, 0.5 ml 20X Peroxide and 9.0 ml Milli-Q water) with gentle agitation for 1 minute at room temperature.

NOTE: LumiGLO® substrate can be further diluted if signal response is too fast.

Drain membrane of excess developing solution (do not let dry), wrap in plastic wrap and expose to x-ray film. An initial 10-second exposure should indicate the proper exposure time.

NOTE: Due to the kinetics of the detection reaction, signal is most intense immediately following LumiGLO® incubation and declines over the following 2 hours.