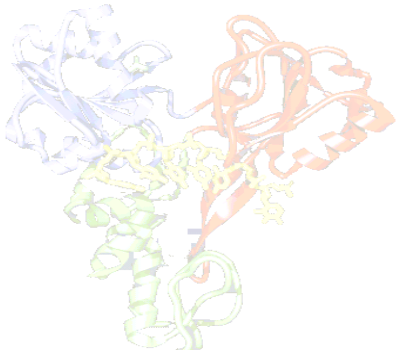




*Life Under Pressure:*  
**Application of Hydrostatic Pressure in Life Sciences**  
*Alexander Lazarev, Ph.D.*



Presented at NCIFCRF on June 10<sup>th</sup>, 2008



## Sample preparation in the Era of “Analytical Arms Race”

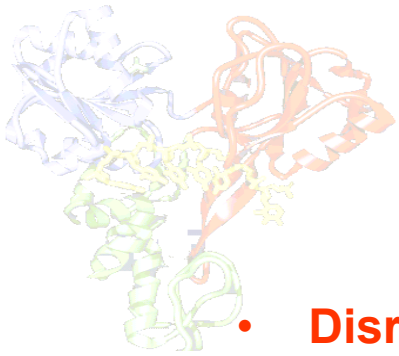


Well-defined experimental goal and well-prepared sample are the foundation of success.

# Conventional cell disruption methods

- Mortar & pestle or Dounce homogenizer (glass on glass)
- Potter-Elvehjem homogenizer (Teflon on glass)
- Enzymatic Digestion
- Polytron shearing homogenizers
- Blenders
- Bead mills
- Sonication
- Repeated freeze/thaw cycles
- French press ( $\leq 2000$  PSI)





## Ideal tissue and cell processor?

- **Disrupts lipid bilayer and molecular complexes**, but not covalent bonds (proteins, DNA, RNA, etc.)
- **Distributes energy** uniformly throughout the sample
- Facilitates **partitioning** of lipids, proteins and nucleic acid
- Does not depend on **aggressive extractions buffers**
- Yet, **compatible** with a wide variety of extraction buffers
- Prevents sample **cross-contamination**
- Keeps samples **enclosed** during the processing
- Provides precise **temperate control**
- Capable of processing **frozen samples** directly
- Processes samples with a **throughput** matching the downstream analysis.
- ...

# Multi-stage extraction approach employing orthogonal methods

Extraction 100 mg tissue: 1200  $\mu$ L of solvent

centrifugation

supernatant

pellet

50  $\mu$ L for protein assay  
250  $\mu$ L for 2DGE  
50  $\mu$ L for SDS PAGE  
200  $\mu$ L for dot blot

Exchange solvent if necessary

no reducing agent  
reduction alkylation  
DTT reduction  
no reduction no detergent ultrafiltration

PRIMARY ANALYSIS

resuspend in appropriate buffer

2<sup>nd</sup> Extraction

centrifugation

pellet

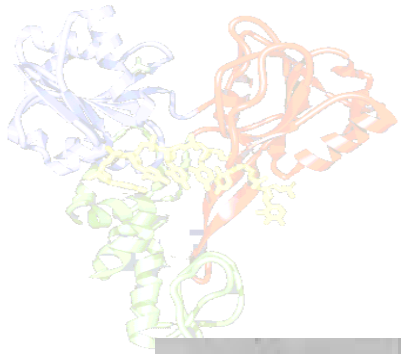
etc.

Supernatant\*

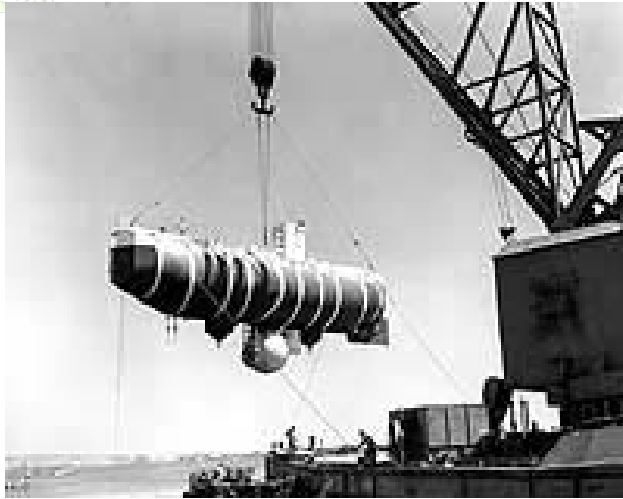
50  $\mu$ L for protein assay  
250  $\mu$ L for 2DGE  
50  $\mu$ L for SDS PAGE  
20  $\mu$ L for dot blot

\* exchange solvent if necessary

SECONDARY ANALYSIS



# Understanding hydrostatic pressure

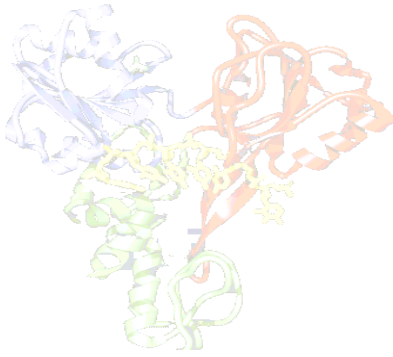


U.S. Navy Bathyscaphe  
***Trieste*** (1958-1963)



Marianas Trench:  
38,713 ft (11,800m) deep  
**16,000 PSI (120MPa)**

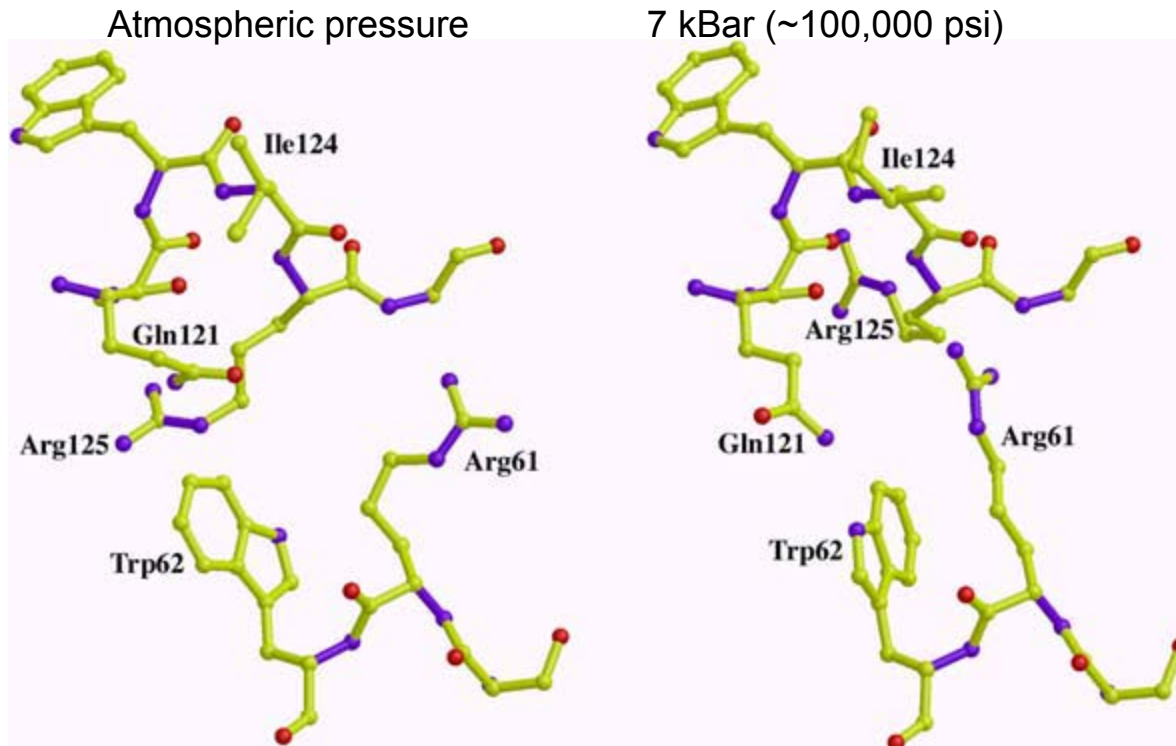
Significant portion of the Global Biosphere is  
subjected to high hydrostatic pressure!



# History of High pressure in Life Sciences

- **1623-1662:** Blaise Pascal – described fundamental concepts of pressure and vacuum.
- **1895:** H. Royer – pressure kills bacteria
- **1899:** B.H. Hite *et al.* – pressure preserves milk
- **1914:** P.w. Bridgman - pressure coagulates egg white
- **1989:** High pressure processing of food products
- **2000:** First International Conference on HPBB

# Protein Crystallography at high pressure



Protein crystals of lysozyme under pressure exhibit more tightly packed structure. The ordering effect of pressure may help to obtain quality crystals of “tough” proteins.



# HPP – High Pressure Food Processing

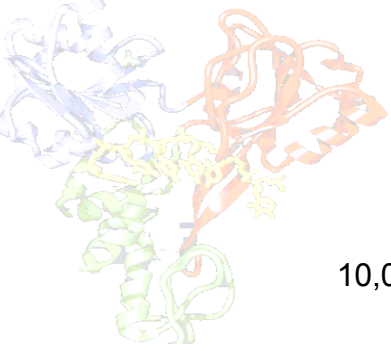
60,000 – 87,000 psi - selective denaturation of proteins



# Commercial Scale Processing, 60,000 – 80,000 psi

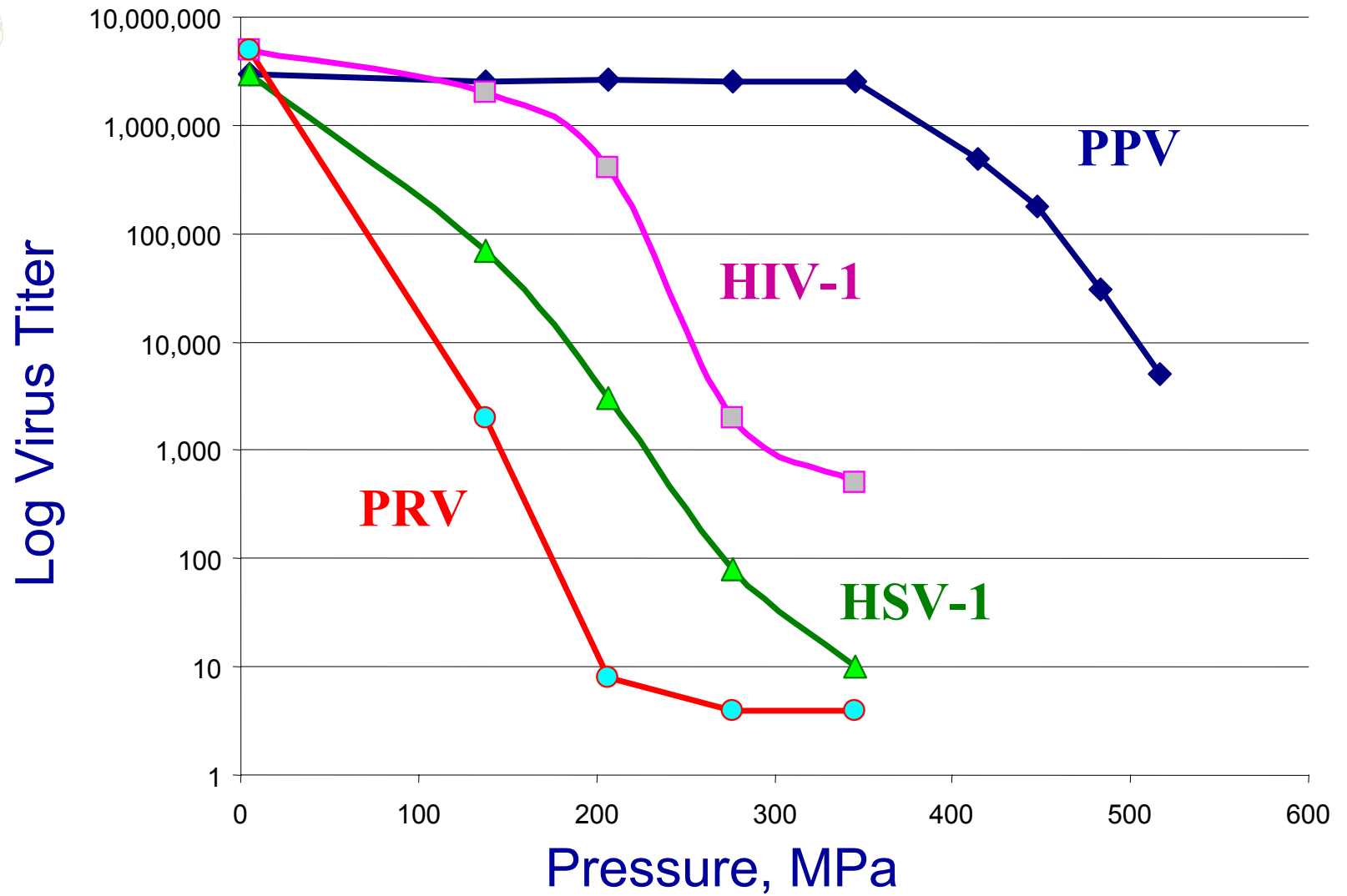
- Inactivates food borne pathogens (listeria, e-coli, etc..) without heat or chemicals
- Extends shelf life
- Pre-packaged product is being processed
- No heat required
- Preserves chemical composition, texture, taste

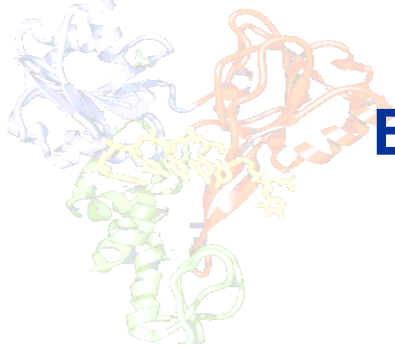




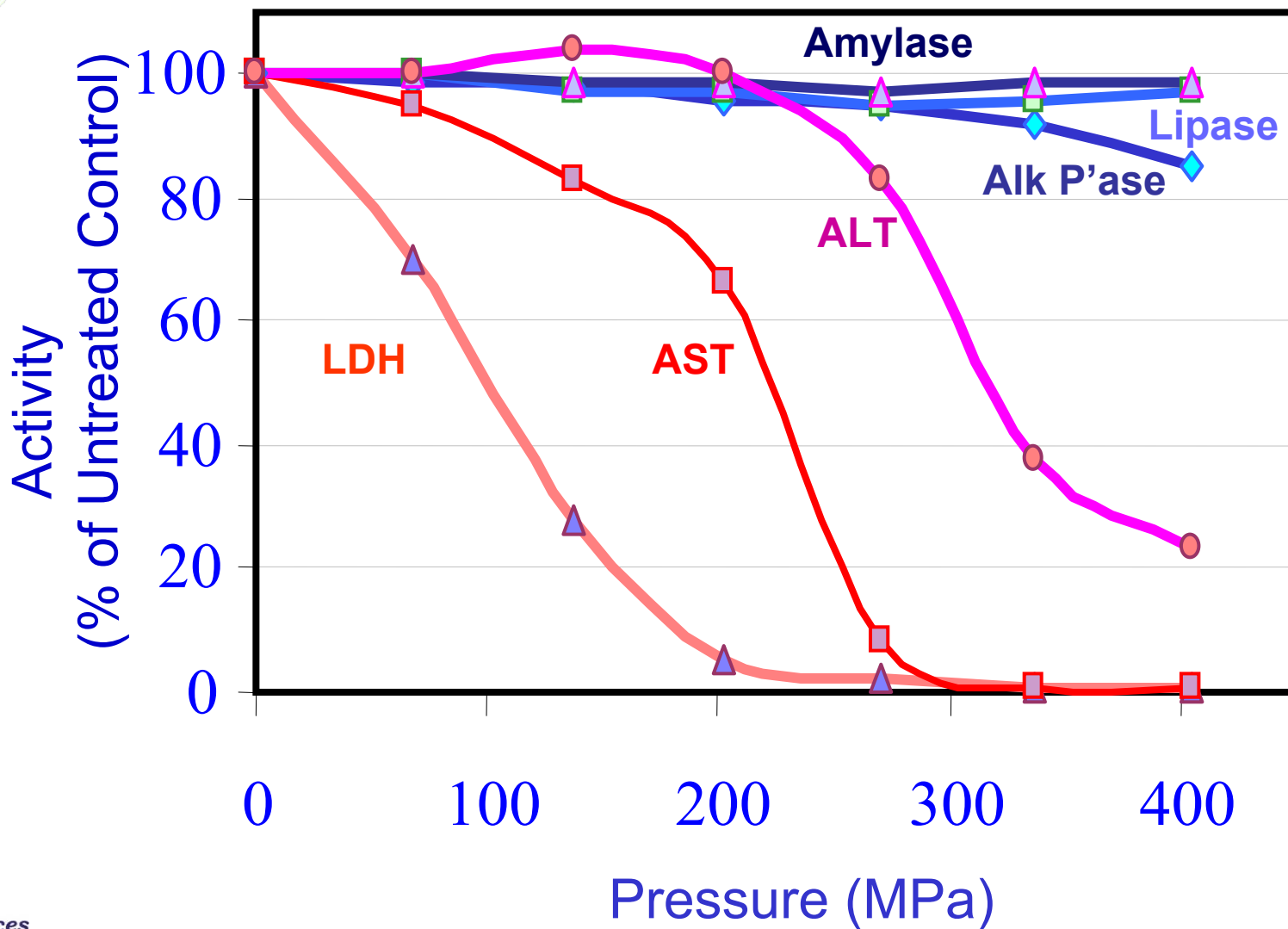
# Viral inactivation by hydrostatic pressure

Note: 240 MPa = 35,000 psi





# Effect of high pressure on enzymatic activity



# Ultra-High Pressure HPLC



**James W. Jorgenson**  
W. R. Kenan, Jr. Professor of  
Chemistry, UNC, Chapel Hill

Ultra High Pressure HPLC  
50,000 psi  
Waters UPLC (15,000 psi)

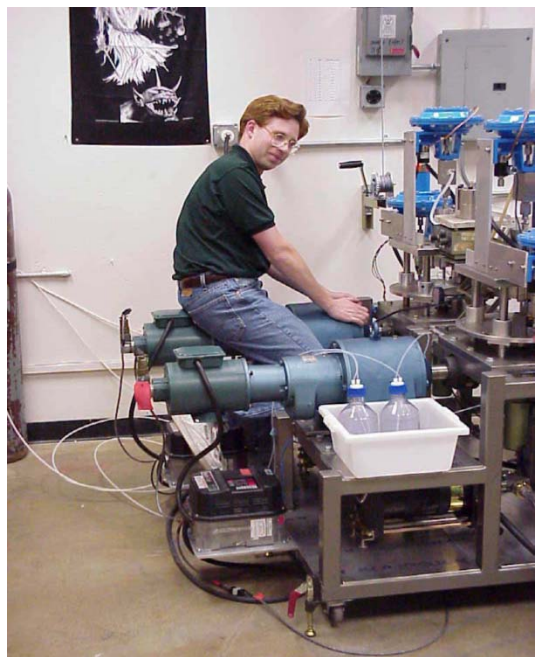
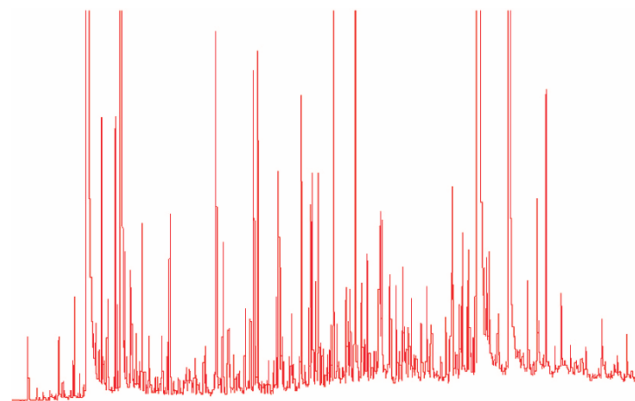
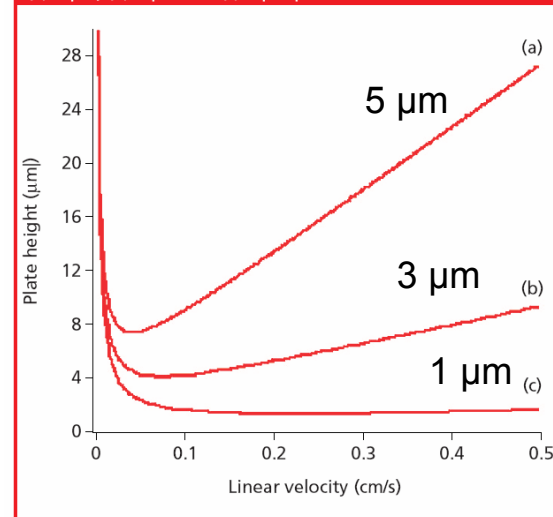
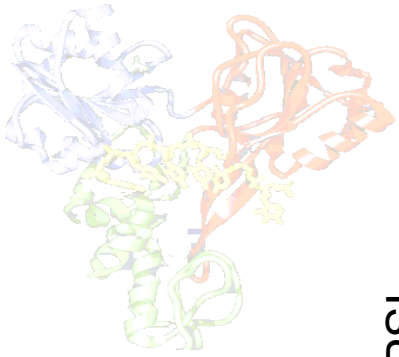


Figure 1: Theoretical performance of columns packed with (a) 5  $\mu\text{m}$ , (b) 3  $\mu\text{m}$  and (c) 1  $\mu\text{m}$  particles.

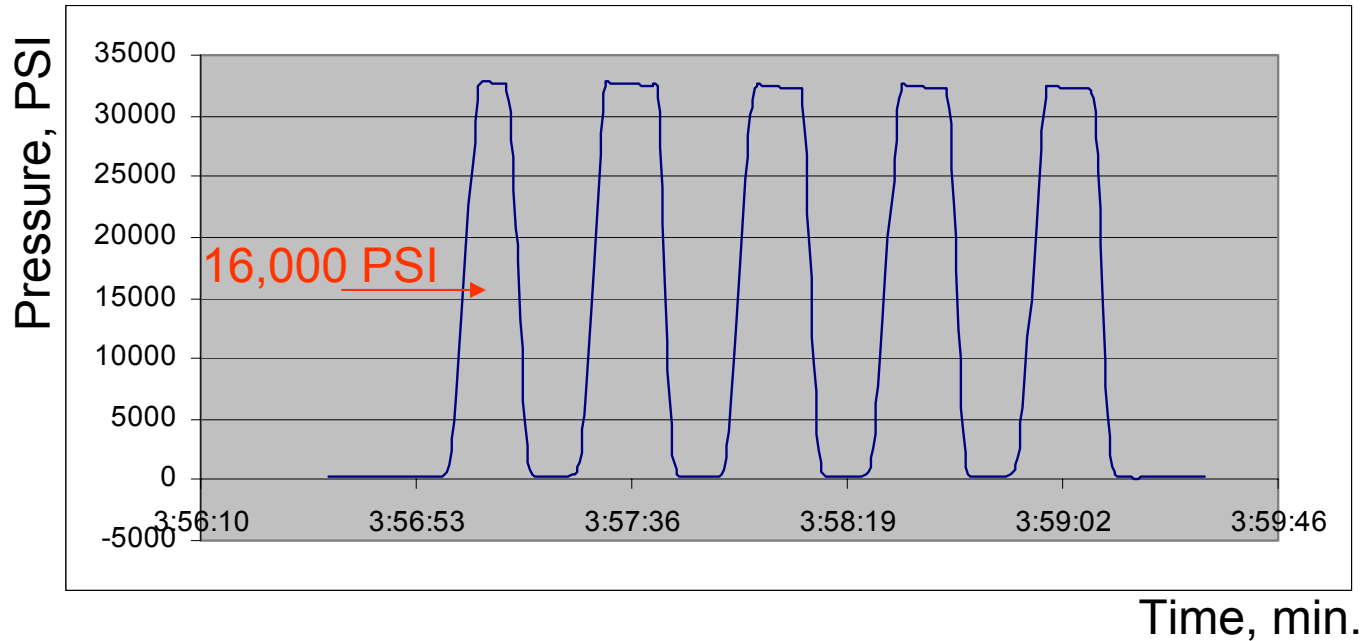


**UHPLC gradient separation of a tryptic digest of bovine serum albumin. A peak capacity of 500 was obtained between 48 and 168 min.**

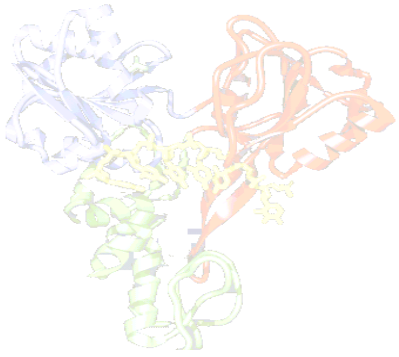


## Pressure Cycling Technology (PCT):

13 US patents  
4 EU patents  
1 AU patent



“Cycles of hydrostatic pressure between ambient and ultra high levels, which allow for the precise control of molecular interactions”



# PCT Sample Preparation System

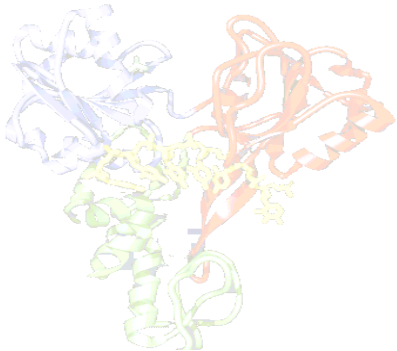


Hydraulic system  
3 samples simultaneously  
Optional temperature control



## Barocycler™ NEP3229

# PCT Sample Preparation System



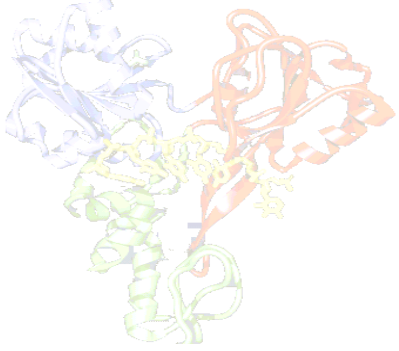
Pneumatic system  
Single sample capacity  
Optional temperature control



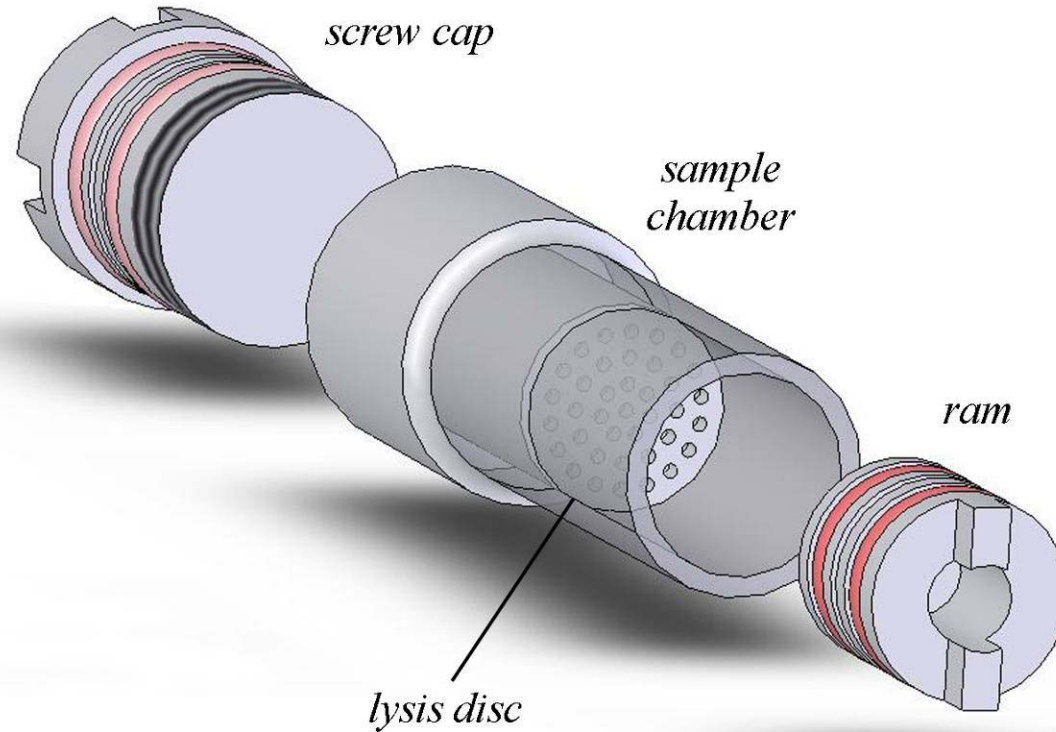
## Barocycler™ NEP2320





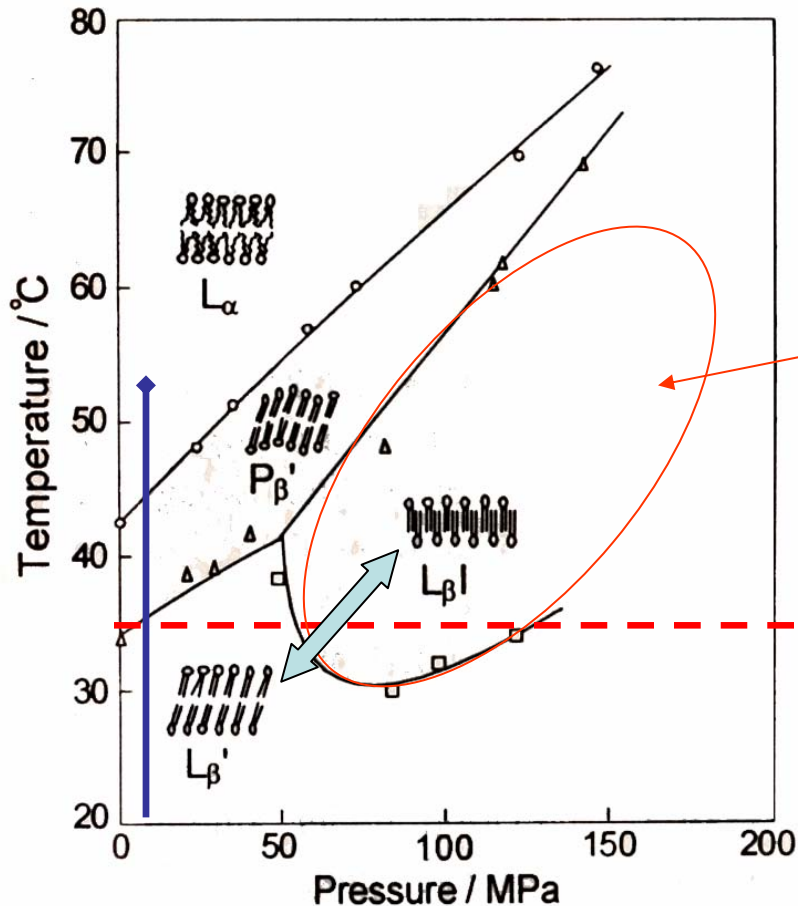


# PULSE™ Tube: disposable sample container



Pressure Used to Lyse Samples for Extraction

# Thermodynamic impact on biological membrane structure



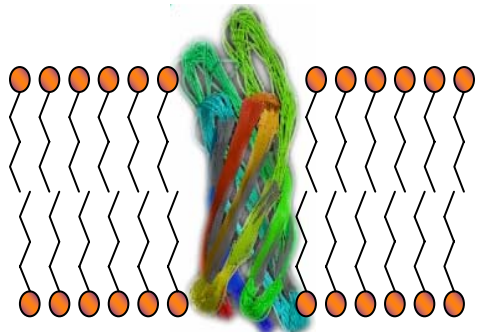
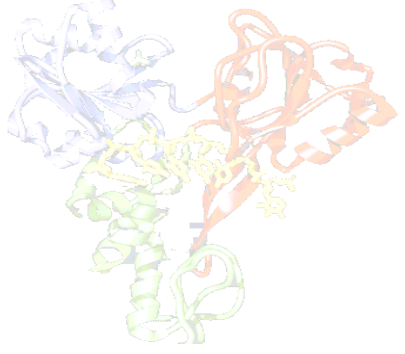
Pressure-induced interdigitation of lipid bilayers in an ester-ester linked HPPC bilayer: HP DSC data.

Interdigitated bilayer

Pressure cycling at 33 °C

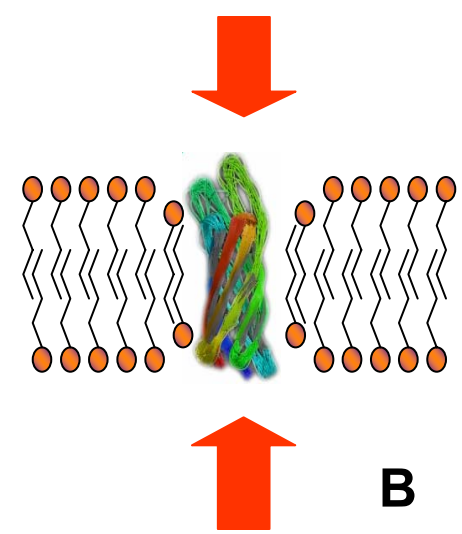
Ichimori H. et al., 1999; in: *Advances in High Pressure Bioscience and Biotechnology*, Horst Ludwig (Ed.), *Proceedings of the Intl. HPBB Conference, Heidelberg, 1998*.

# Pressure cycling acts directly on biological membranes



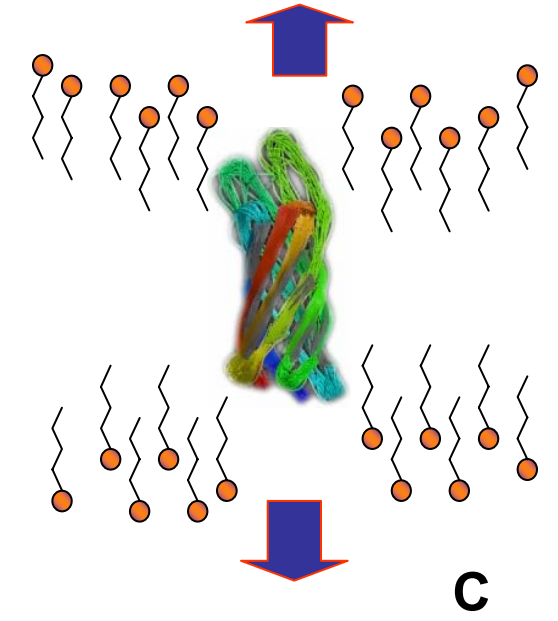
Lipid bilayer  
Membrane Protein

**A**



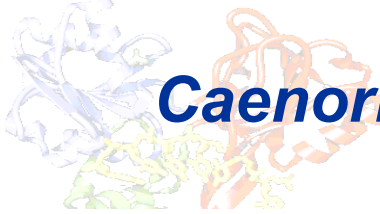
Hydrostatic Pressure Applied  
(Interdigitated bilayer)

**B**

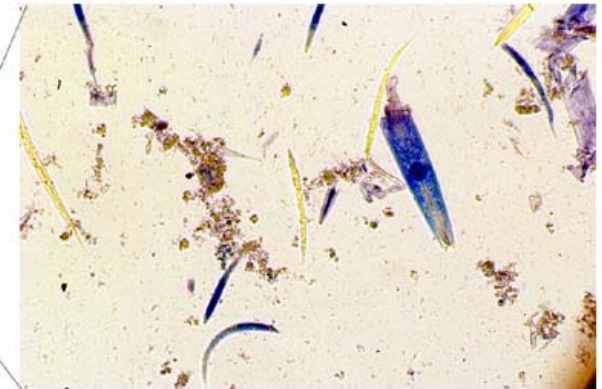
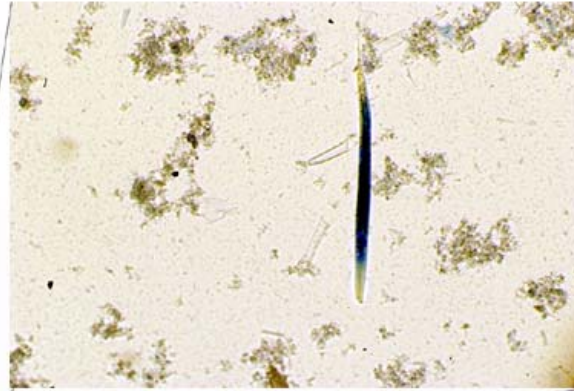
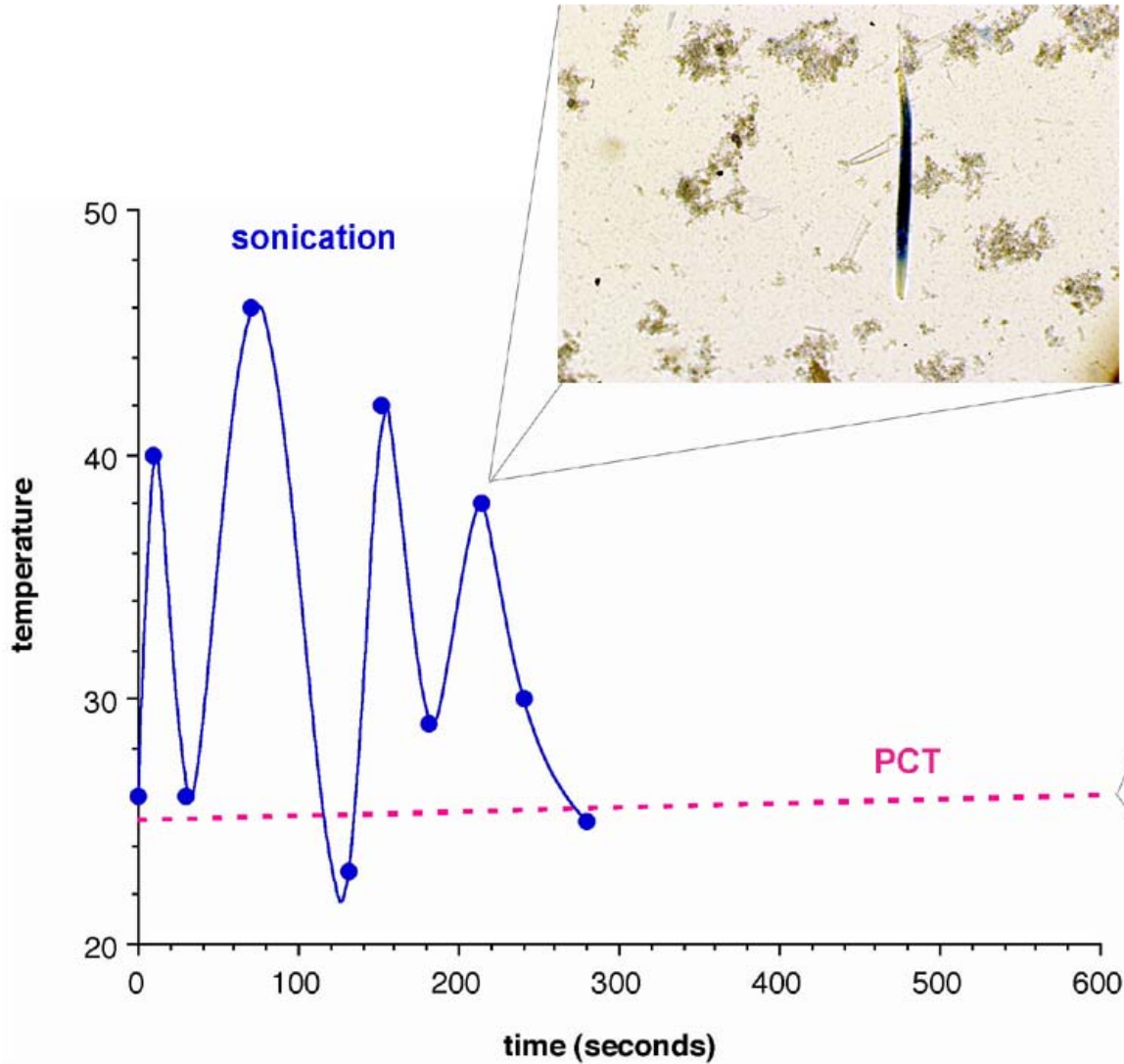


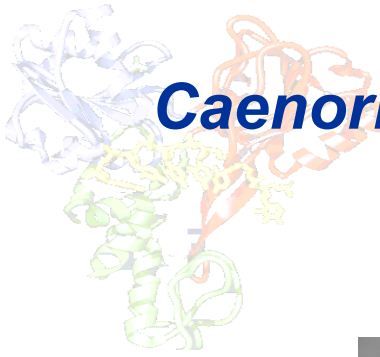
Hydrostatic Pressure Rapidly Released

**C**



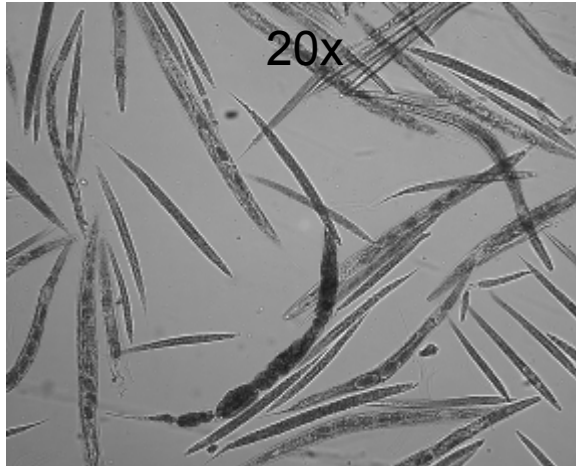
# *Caenorhabditis elegans* extraction, temperature profile



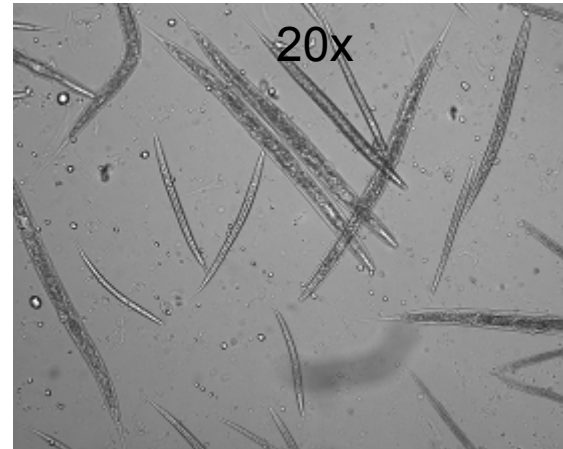


# Caenorhabditis elegans extraction by various methods

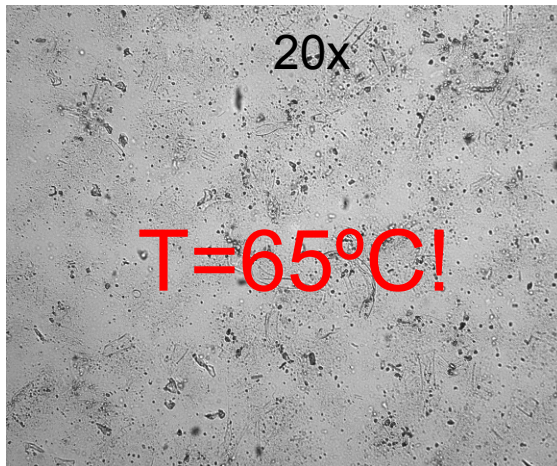
Freeze-thaw



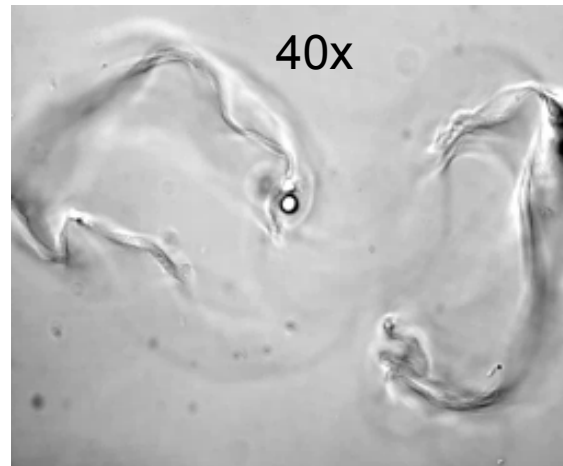
Bead Beater, 4x20s

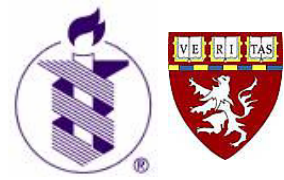
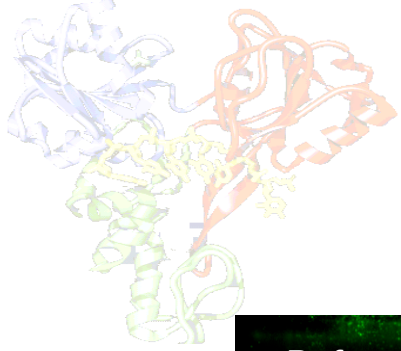


Sonication 3x20s

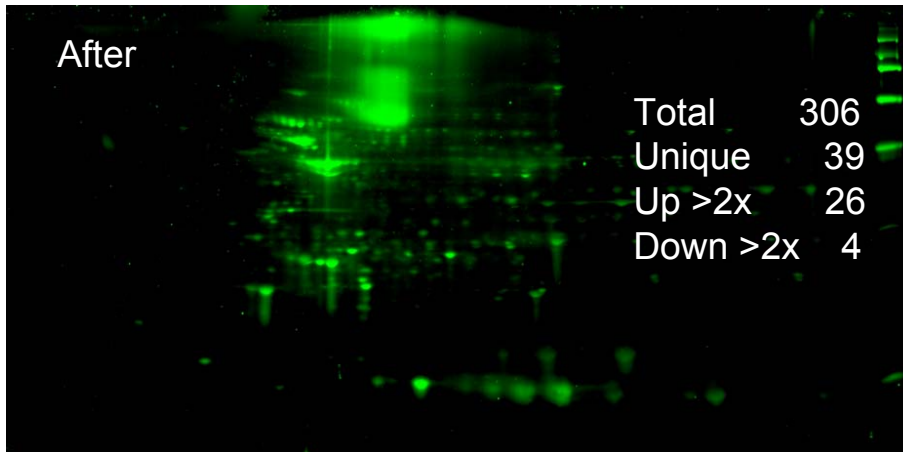
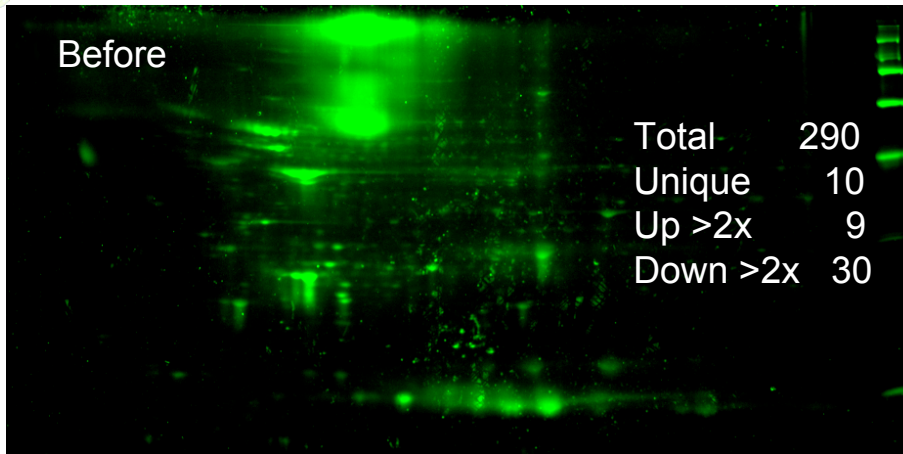


PCT – 5 cycles



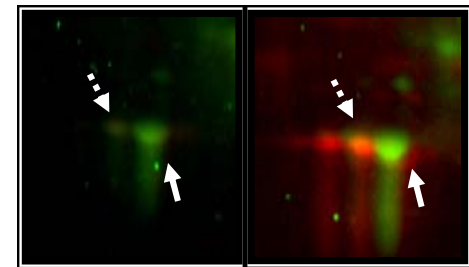


# Human atrium samples before and after cardiac surgery



2D: 3-10 pH gradient, 8-20% SDS-PAGE

- cardioplegia and cardiopulmonary bypass were used
- representative blots form one patient
- 100 mg** of atrial tissue
- SYPRO Ruby and ProQ-Diamond stains



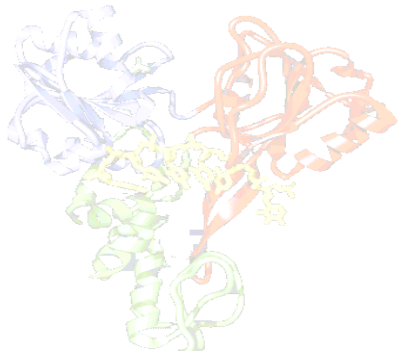
Pre-CP/CPB

Post-CP/CPB

MLC2a Protein

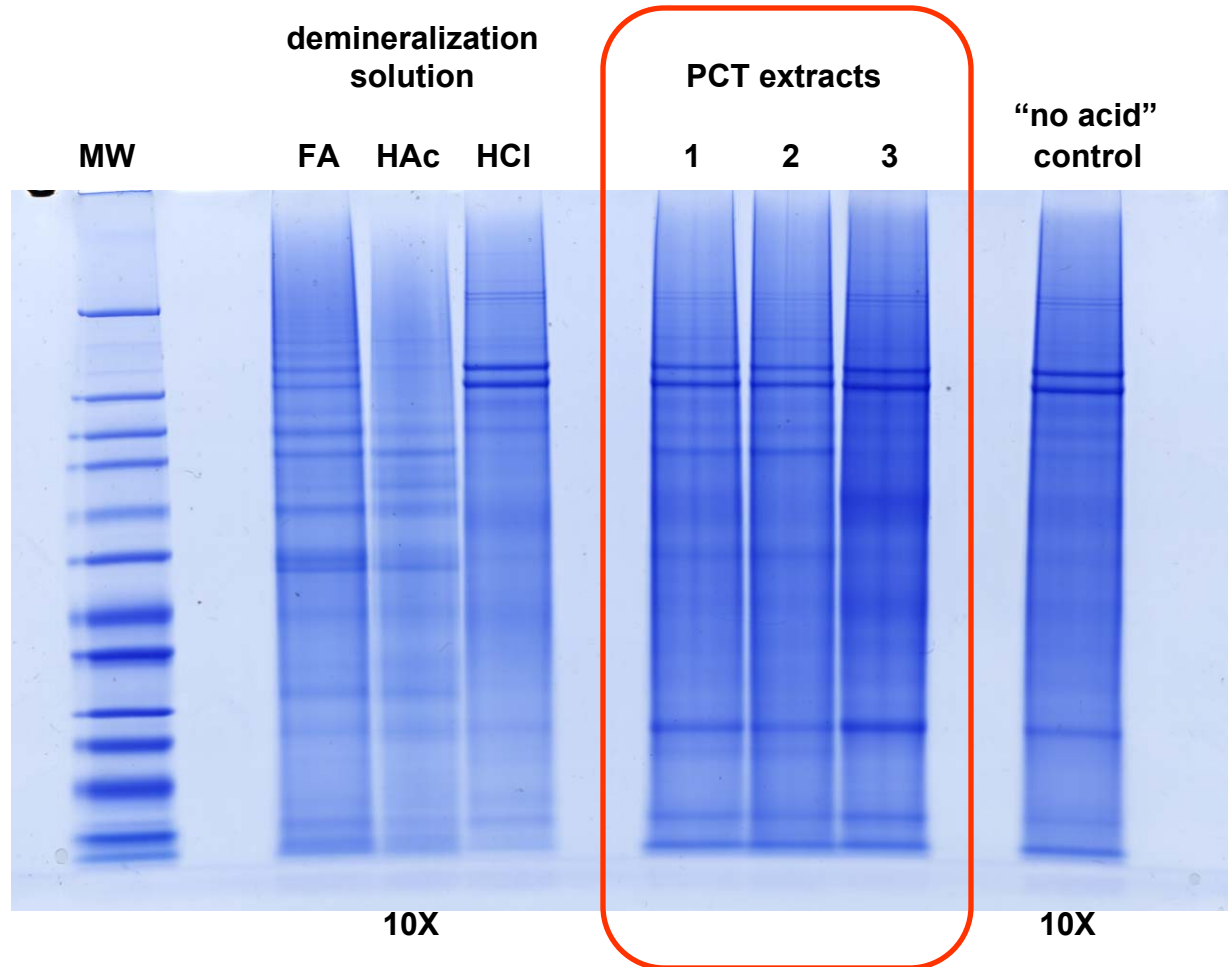
Phosphorylated MLC2a protein

R. Clements et al, AHA 2007 poster presentation

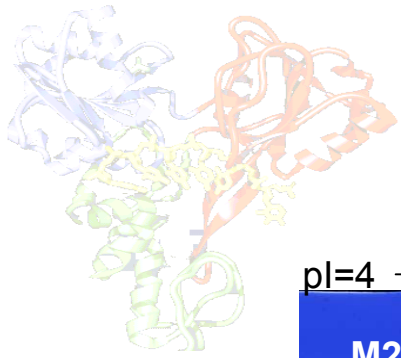


# Protein extraction from cortical bone

PCT releases proteins from bone **without de-mineralization**

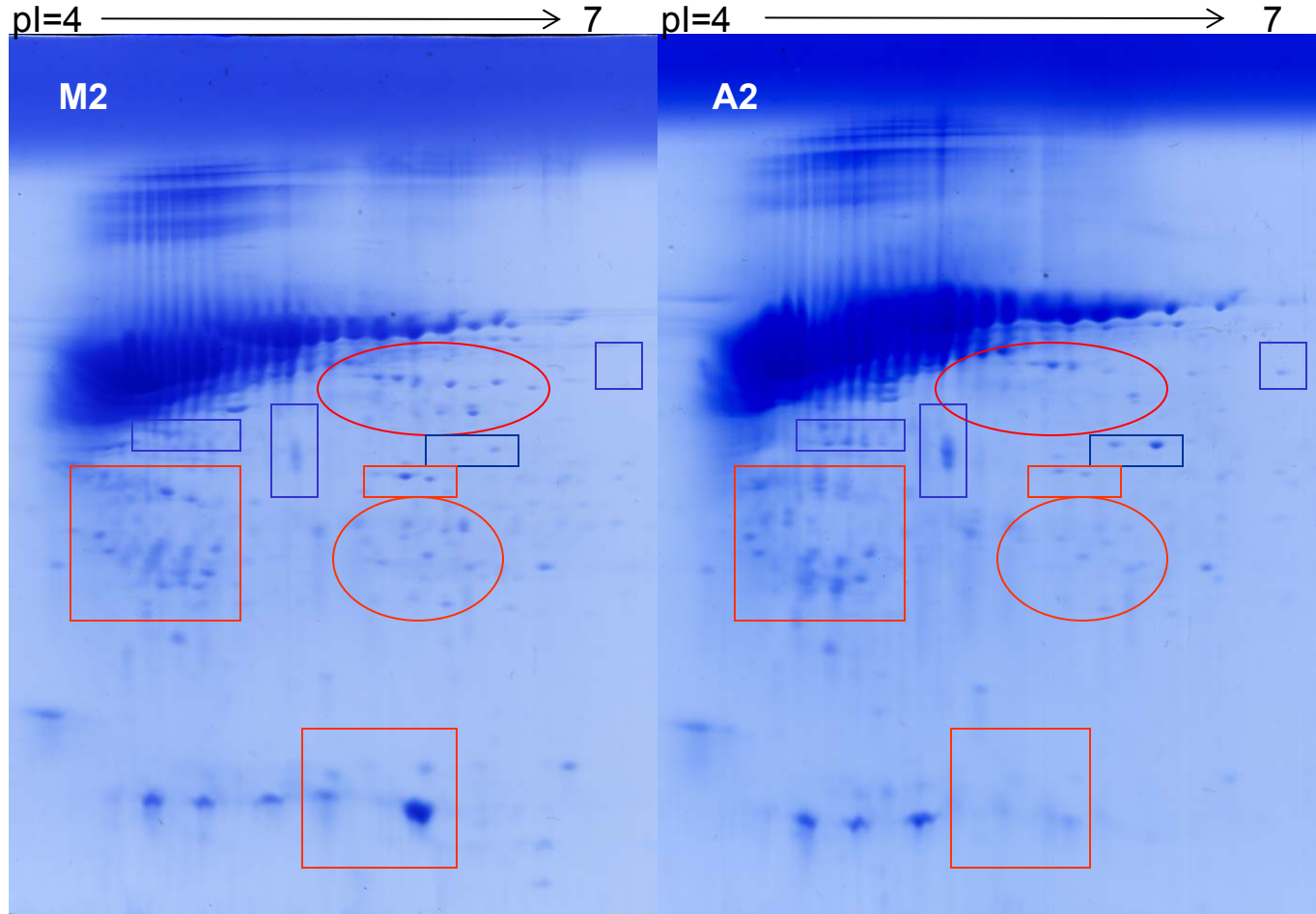


1DGE of aged ostrich bone following acid demineralization, PCT, and Norgen column for removal of Ca and PO<sub>4</sub>



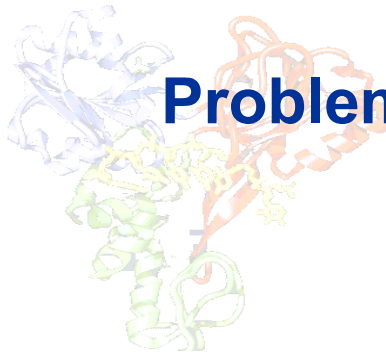
# Human *Stratum corneum* proteome

skin samples of two healthy volunteers



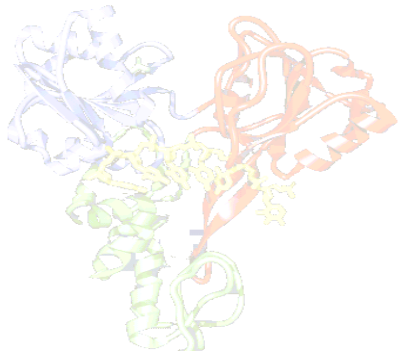
Cells consecutively collected on 10 1" adhesive disks from the same location and processed in two PULSE Tubes in IEF Buffer (7M urea, 2M thiourea, 4% CHAPS)





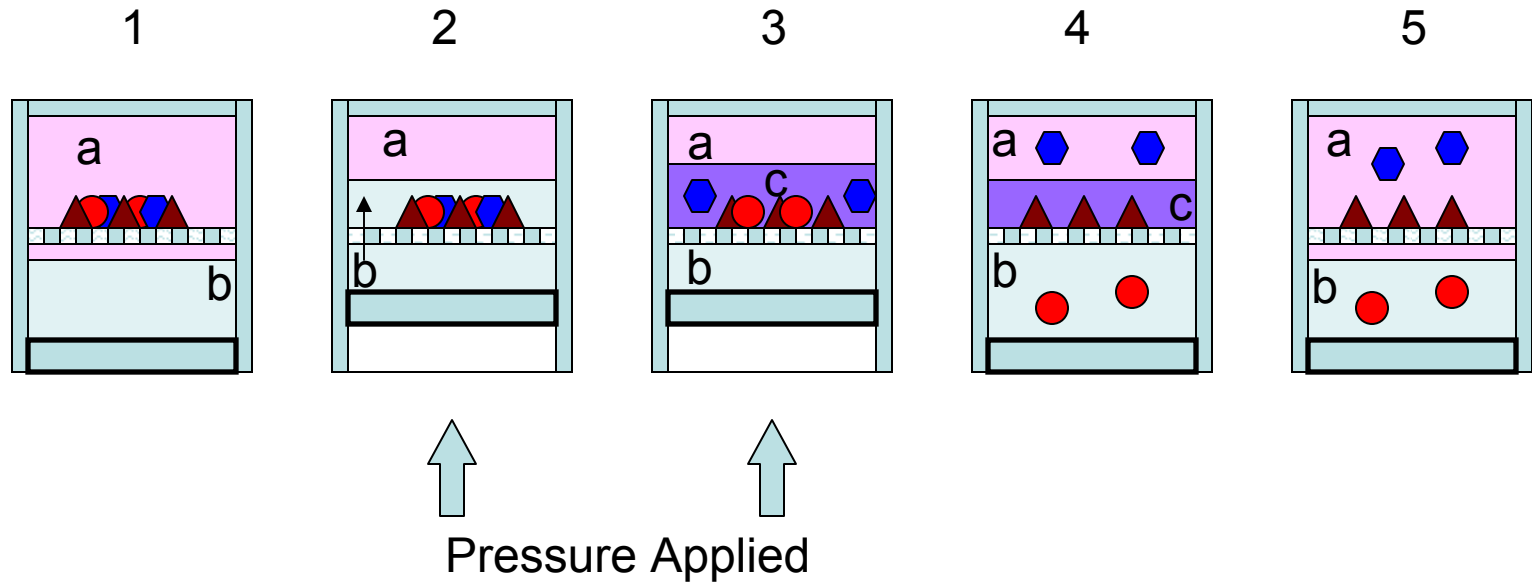
## Problems with traditional methods of protein extraction from sample with high lipid content

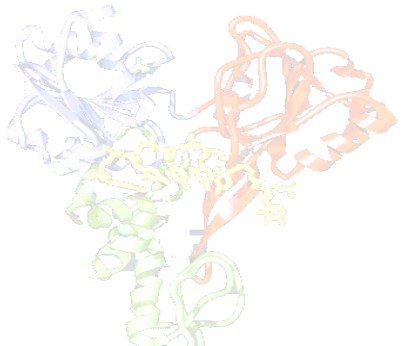
- Proteomic investigations of adipose tissue may lead to discovery of new biomarkers of diabetes, obesity, other metabolic disorders and cancer.
- Adipocytes may contain up to 70% lipids by weight
- Small amount of detergent (1-5%) is sequestered into micelles
- Membrane proteins are captured by micelles or remaining lipid phase
- Cavitation and shearing homogenizers promote formation of emulsion
- Dounce homogenizers, bead beaters: sample loss on the surfaces



# PCT-mediated liquid-liquid extraction

*(patent pending)*





# ProteoSolve<sub>LRS</sub>

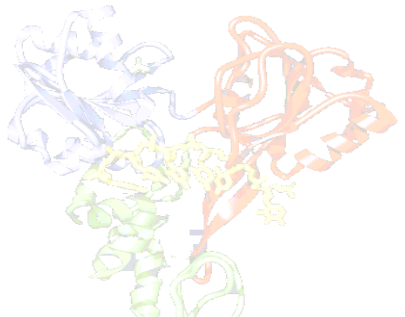
*PCT Dependent Detergent-Free Extraction of Proteins from Lipid-Rich Samples*

- A** Dissolution Reagent
- B** Partitioning Reagent
- C** Precipitation Reagent



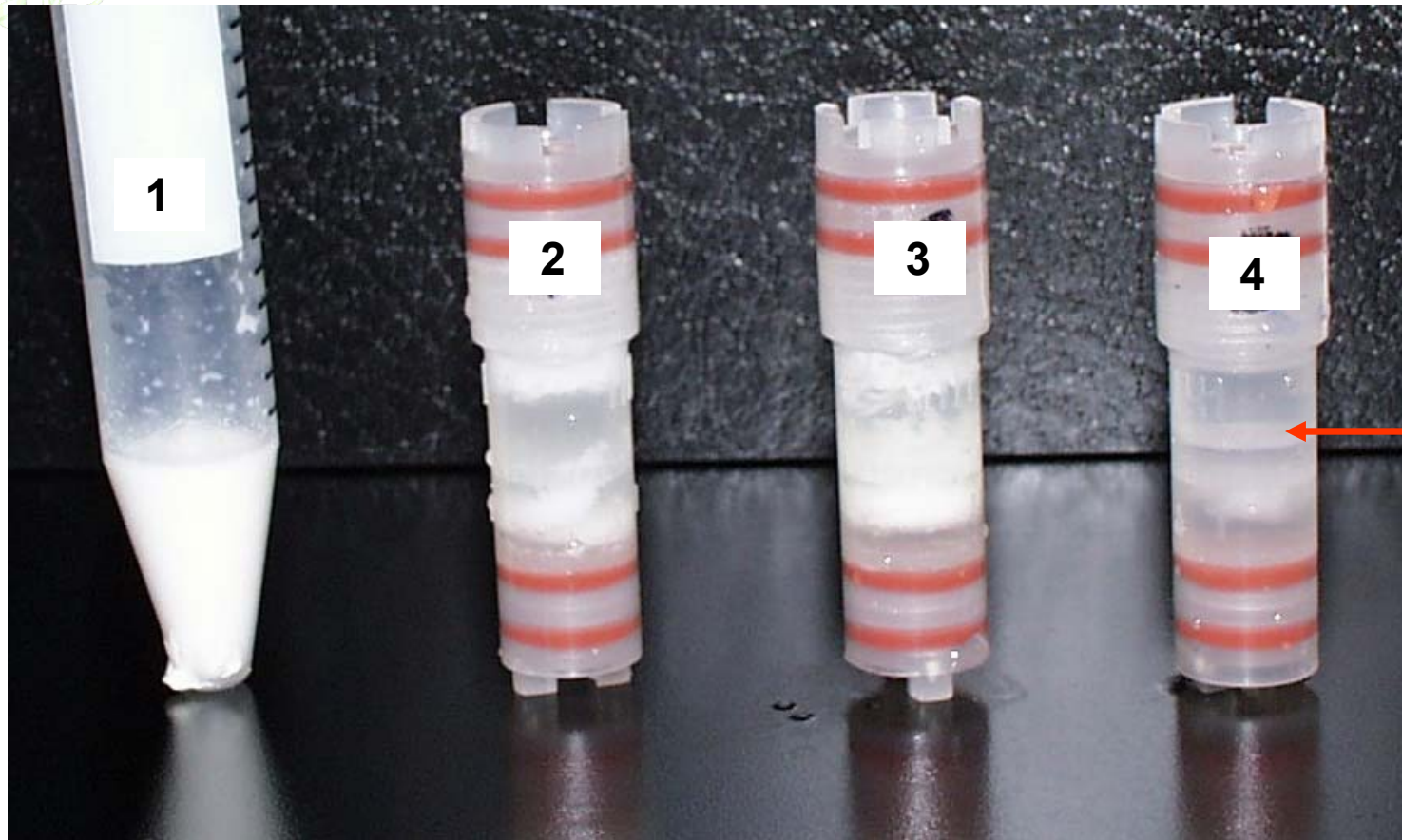
12 PULSE Tubes™

**2007 Frost and Sullivan  
Innovative Product Award**



# ProteoSolve<sub>LRS</sub> kit performance

200 mg of porcine adipose tissue per tube



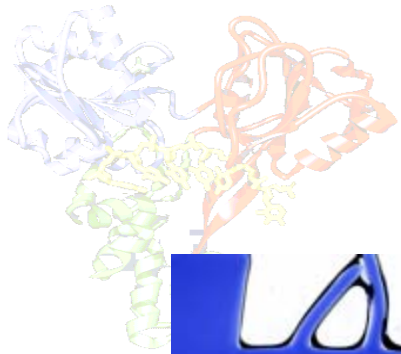
Two liquid phases,  
Clearly visible interface,  
Nearly complete dissolution

Sonication  
2% SDS

PBS

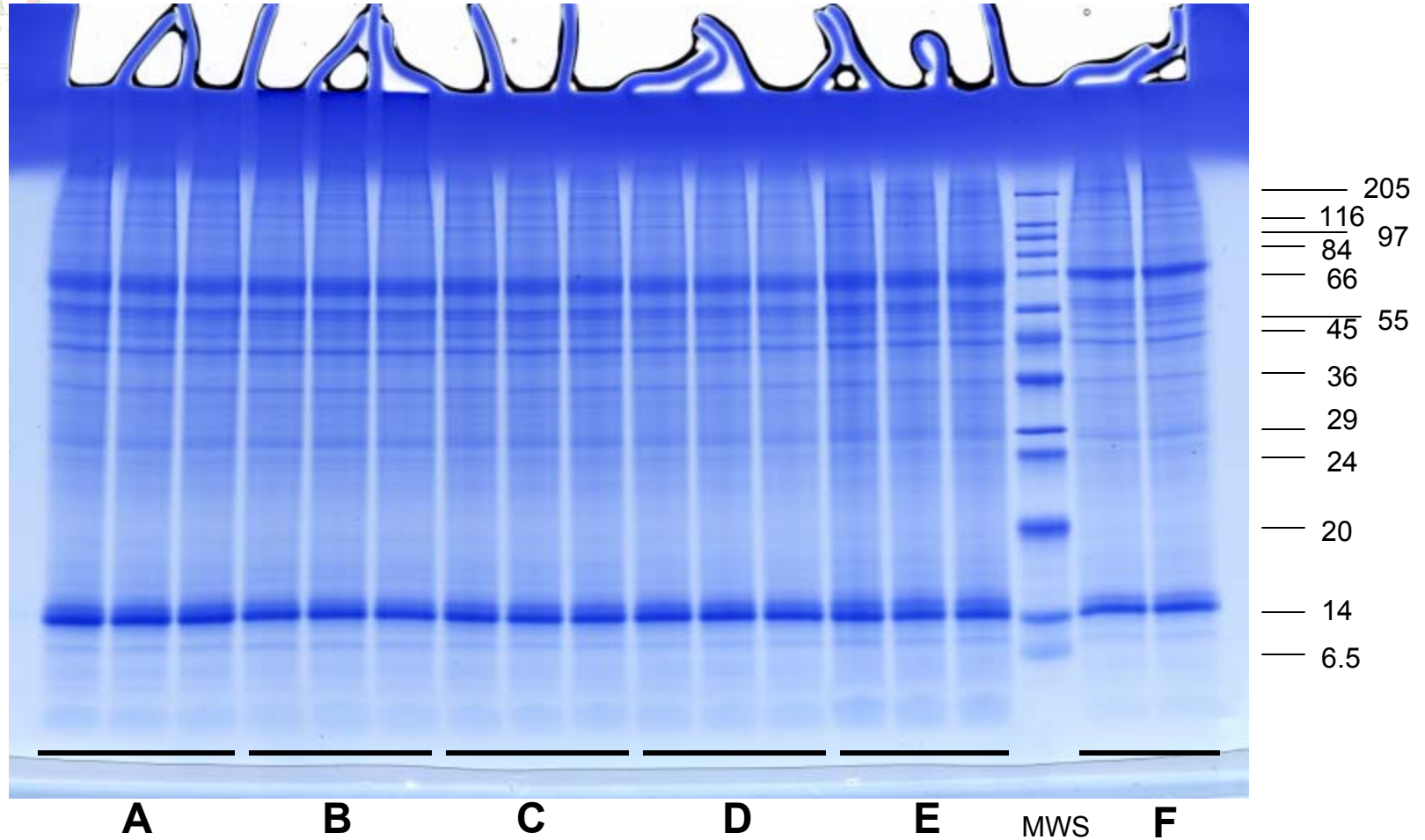
2% SDS

ProteoSolve<sub>LRS</sub>

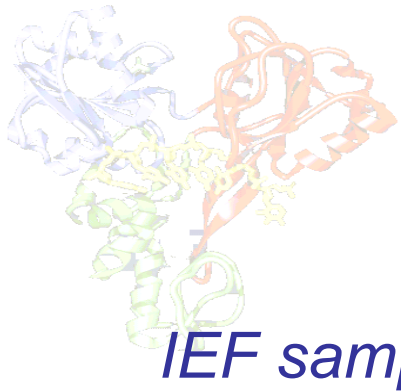


# ProteoSolve<sub>LRS</sub> kit reproducibility

Murine Abdominal Fat Pad Protein Extraction

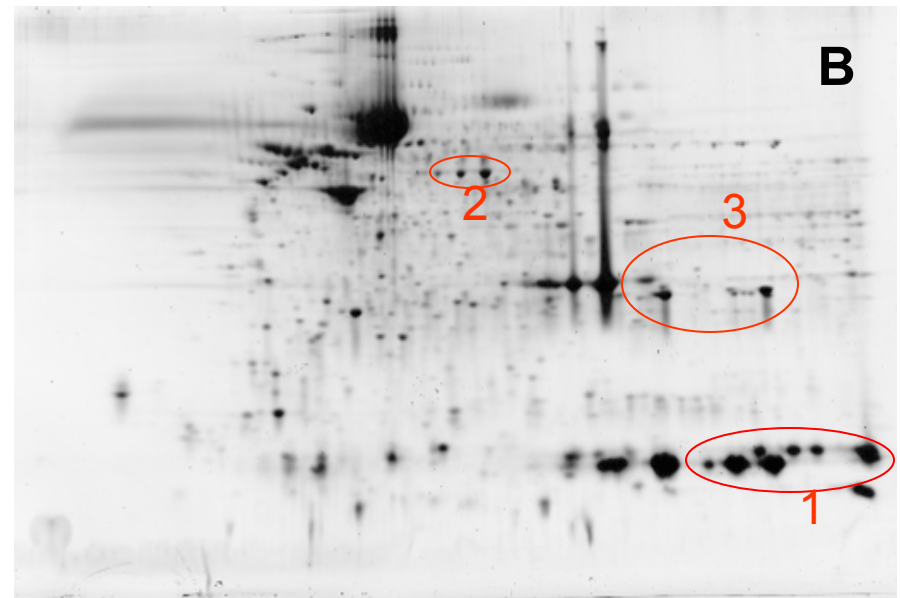
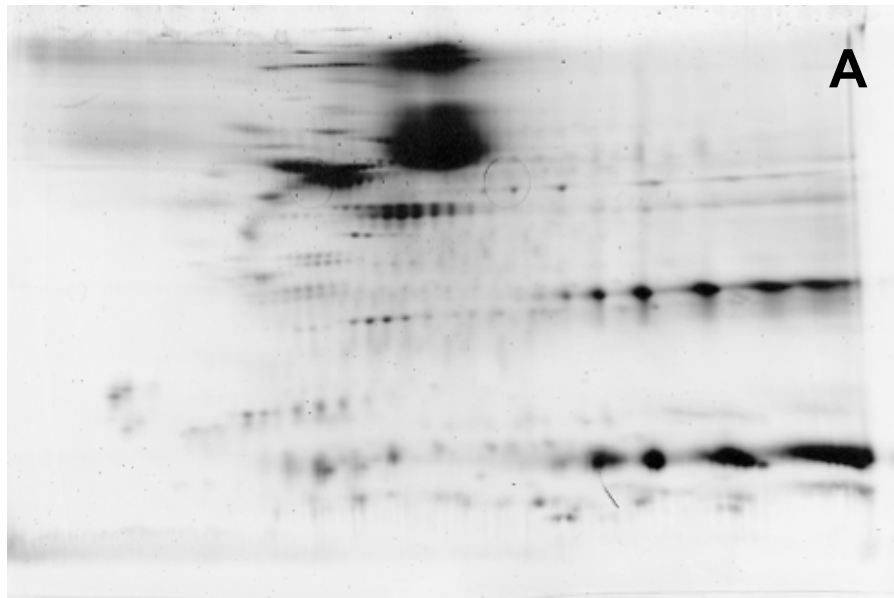


Six 200 mg pieces of tissue (A-F above) were processed in parallel, and loaded in triplicate onto 8-16% gradient SDS-PAGE gel.

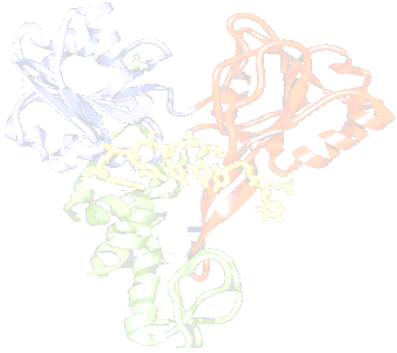


## Extraction of protein from adipose tissue

**ProteoSolve**<sub>LRS</sub>



Proteins from the identical samples of murine adipose tissue extracted by PCT using a traditional detergent-based buffer (9M urea, 4% CHAPS, pH=8) *versus* a detergent-free ProteoSolve LRS kit based on a combination of organic solvents [B].



# Proposed *ProteoSolve*<sup>SB</sup> workflow

Place sample into PULSE Tube. Add Reagent A +/- Reagent B.  
Vortex 10-20 seconds

Place into Barocycler and run program

Remove from Barocycler and Vortex 10-20 seconds.

Transfer entire contents of PULSE tube to centrifuge tube and spin at ~12,000g for 10 min.

**Nucleic Acids**

Solid material contains RNA, DNA  
and some DNA-associated proteins.

Dissolve pellet material in appropriate  
reagent for DNA or RNA isolation.

Vortex sample and proceed with  
isolation according to  
manufacturer's protocol. No liquid  
nitrogen grinding or other tissue  
disruption is necessary.

**Lipids**

Upper non-polar phase  
contains extracted lipids.

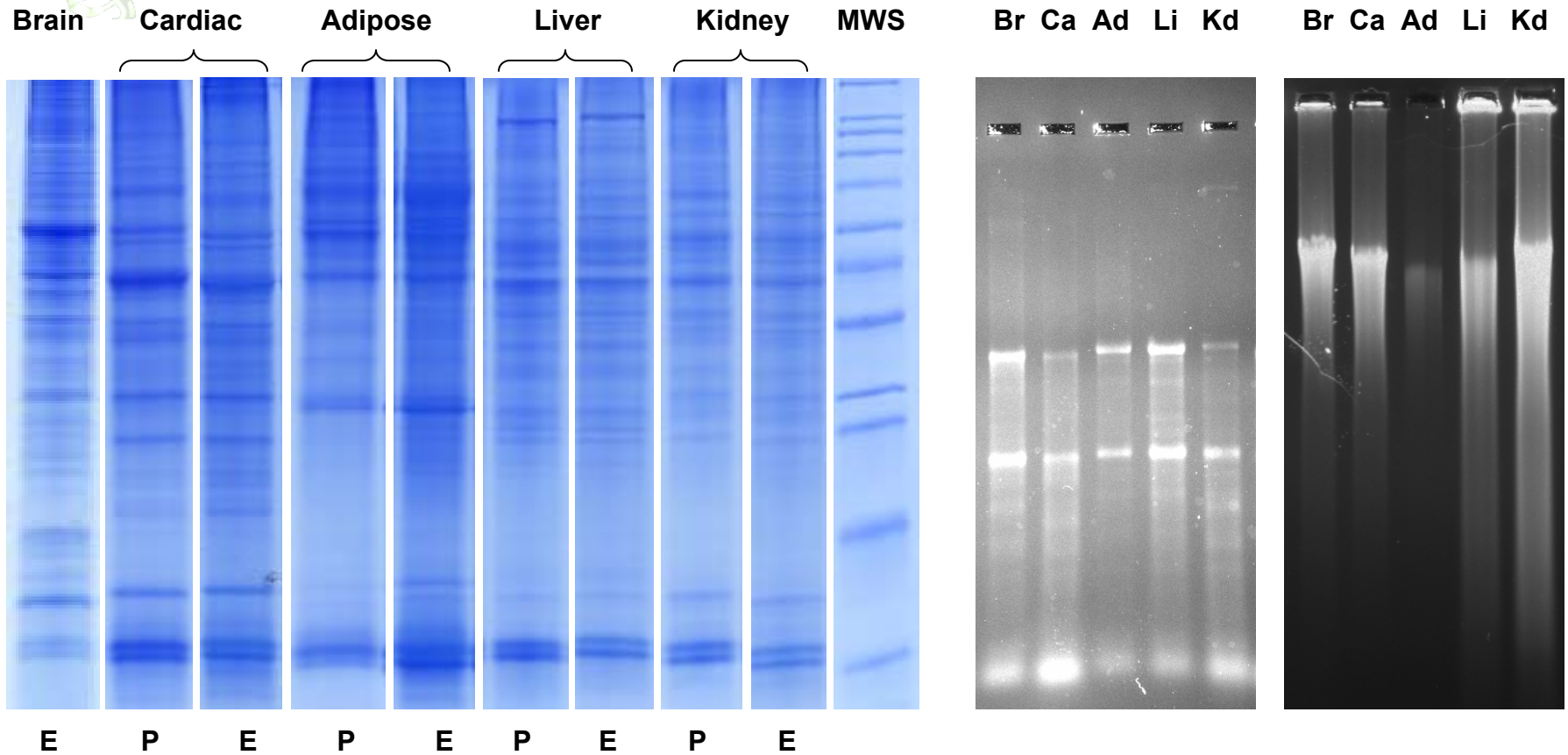
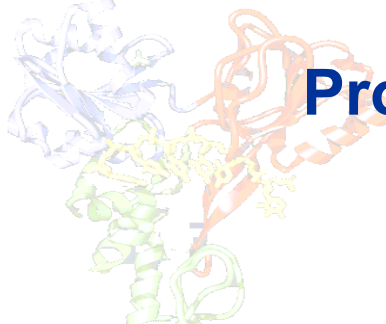
GC-MS,  
LC-MS,  
MALDI/TOF  
TLC, NMR

**Proteins**

Lower liquid phase contains  
dissolved sample proteins.  
Remove solvent by evaporation  
or precipitation.

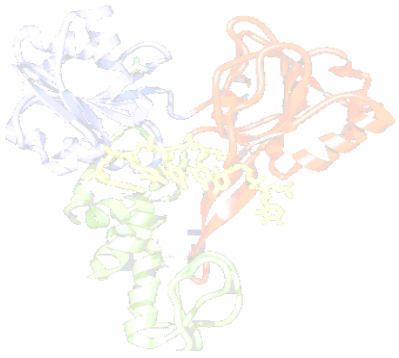
SDS-PAGE, 2D PAGE,  
Western blotting, LC-MS/MS,  
MALDI-TOF, etc...

# Protein, RNA and DNA Recovery From Rat Tissues

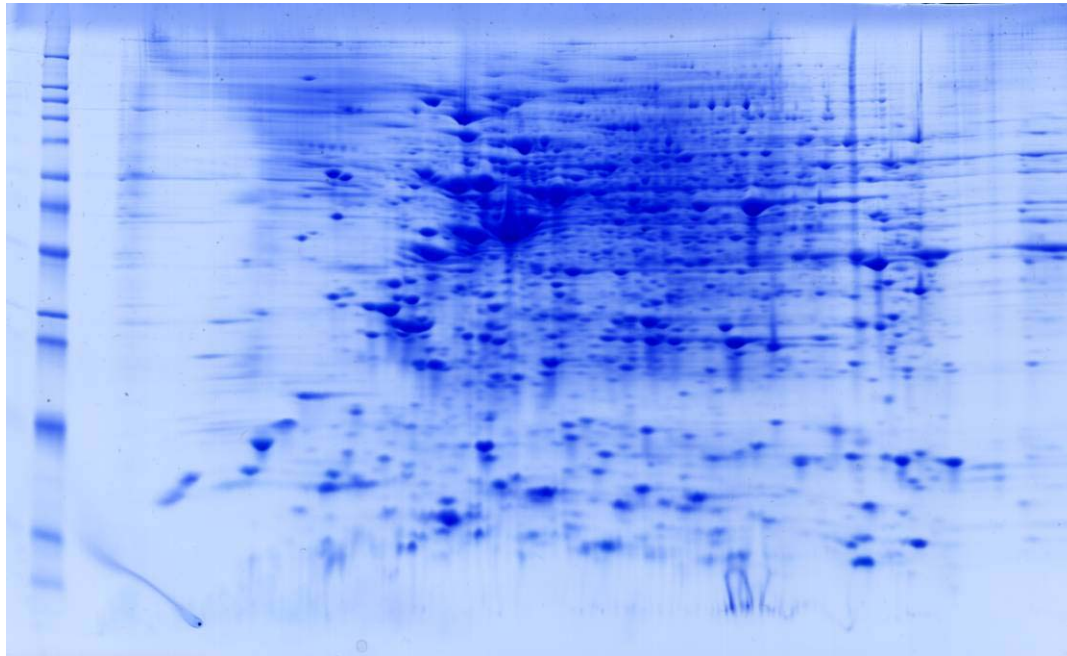


Solvent removed by: E – evaporation; P - precipitation

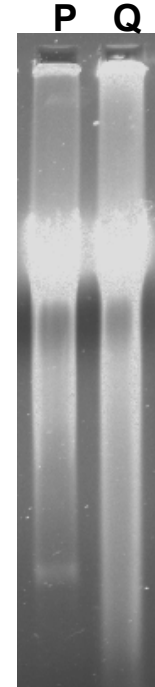




# Protein and DNA Recovery, PC12 cells

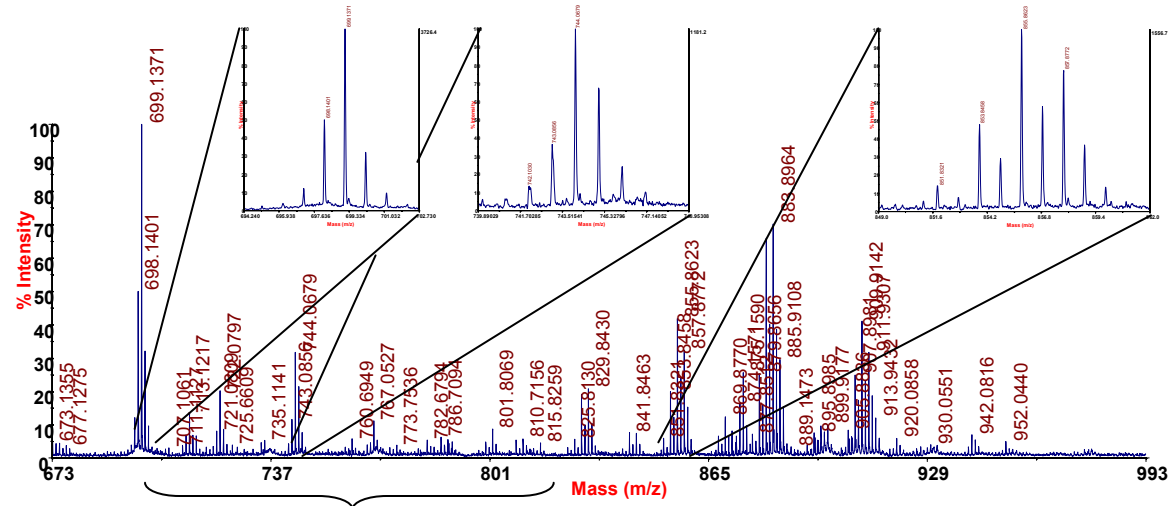
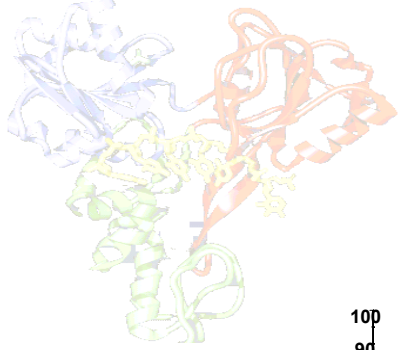


Protein

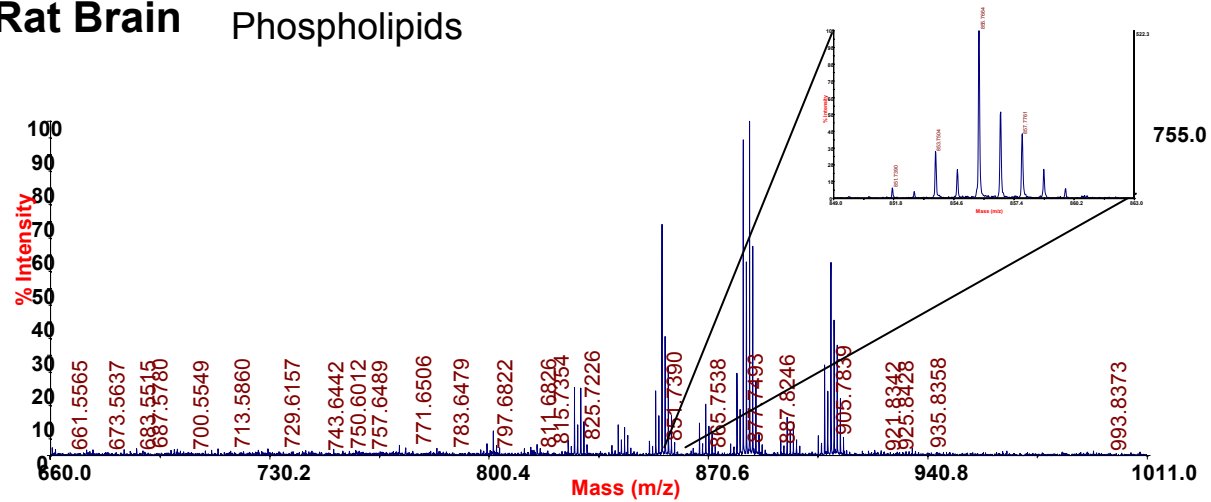


DNA

# Direct Lipid Profiling by MALDI-TOF MS

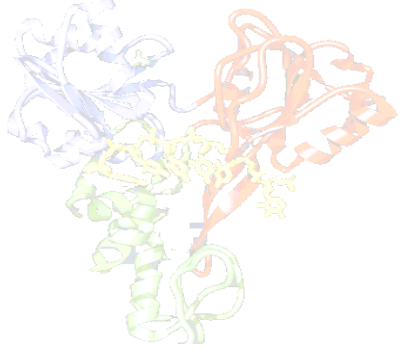


**Rat Brain Phospholipids**

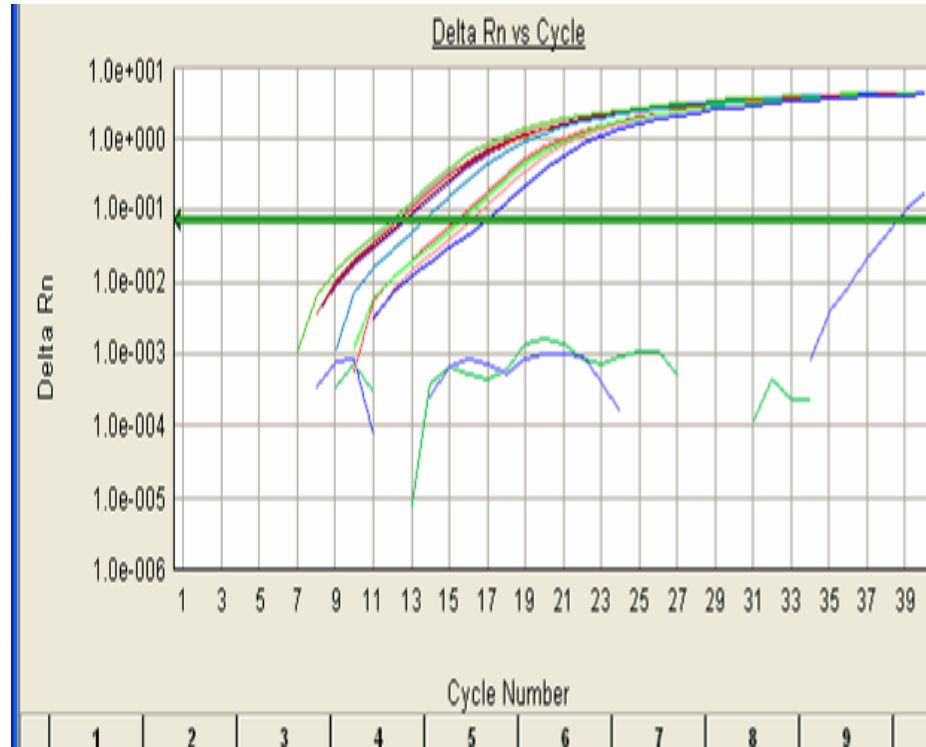


**Beef pericardial fat**

**Triacylglycerides**



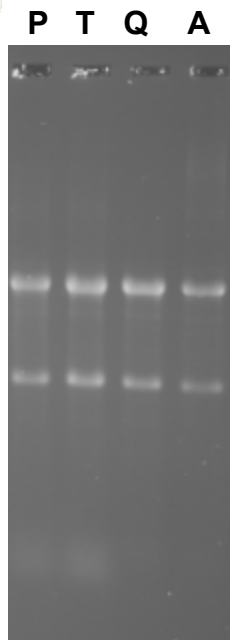
# RNA extracted in ProteoSolve<sub>LRS</sub> is compatible with RT-PCR amplification



RNA recovery from  $10^6$  PC12 cells- Quantification by real-time RT-PCR using primers for rat  $\beta$ -Actin.

# Protein and RNA Recovery, PC12 cells

## Comparison of 4 commercially available kits.



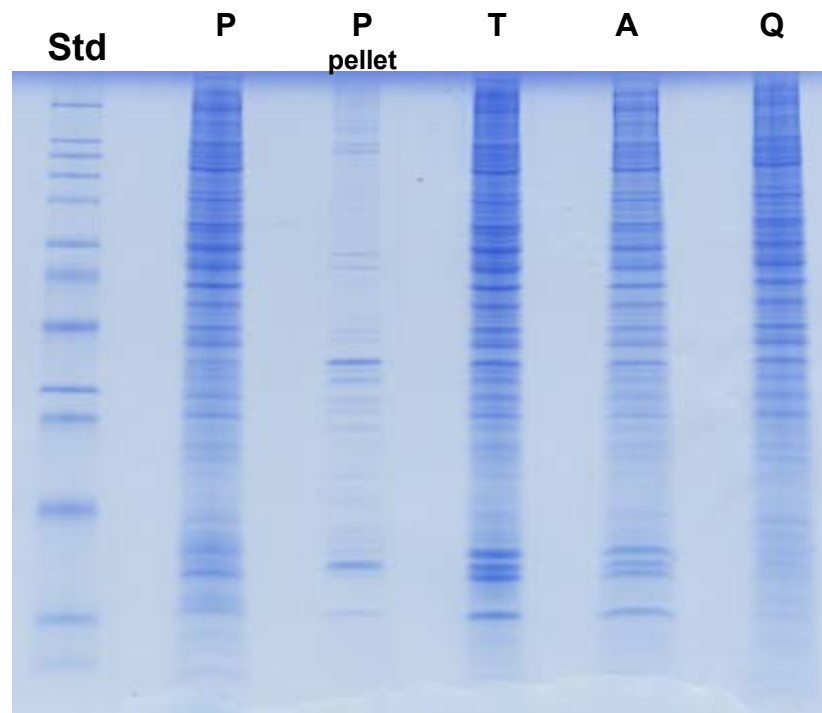
RNA recovery:

**ProteoSolve<sub>LRS</sub>: 11.9 µg total RNA**

Ambion: 5.3 µg total RNA

Qiagen: 11.4 µg total RNA

Trizol: 16.2 µg total RNA



**A:** Ambion PARIS kit

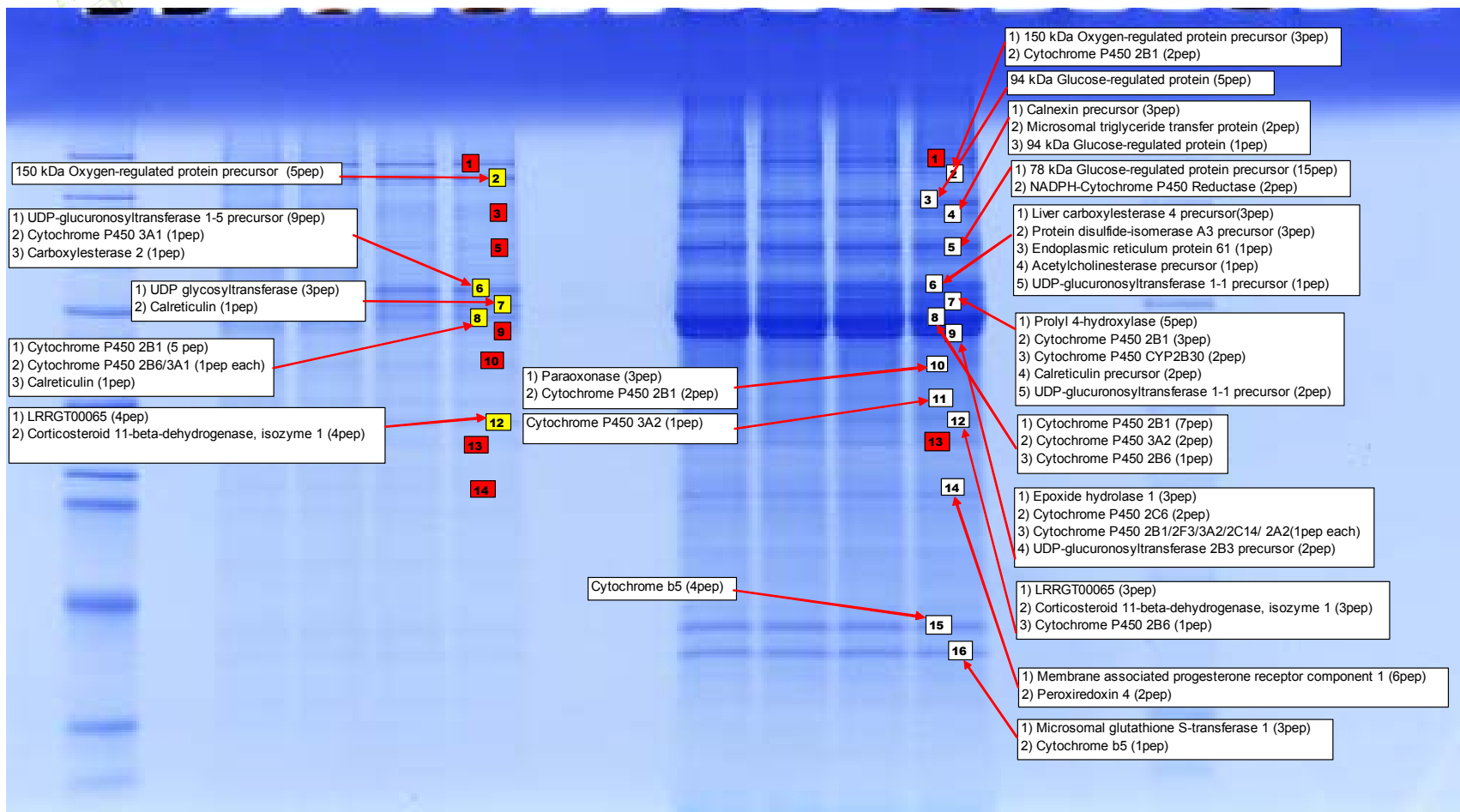
**Q:** Qiagen Allprep kit

**T:** Trizol

**P:** ProteoSolve<sub>LRS</sub>

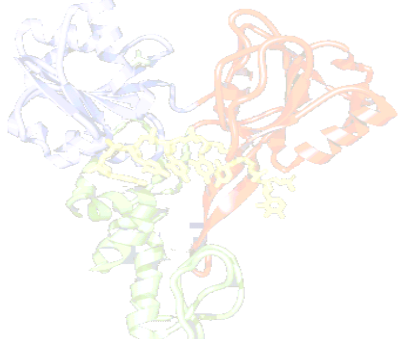
Starting material for each sample was 10<sup>6</sup> PC12 cells.

# Fractionation of Rat Microsomal Proteins using Proteosolve<sub>LRS</sub>

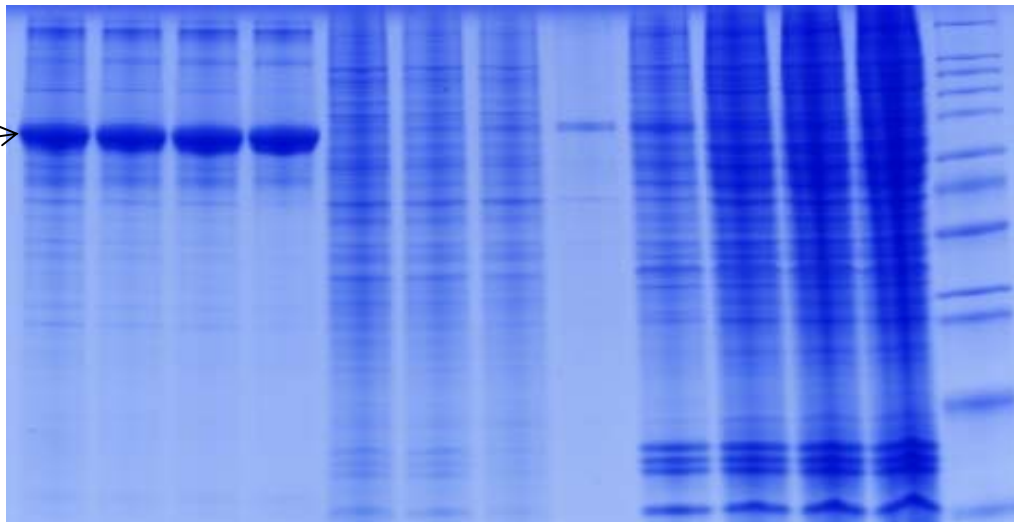


Data courtesy Dr. Michail Alterman, CBER, FDA

# Extraction of Mitochondria from PC12 Cells. Optimization of PCT Conditions.



BSA,  
Buffer  
ingredient



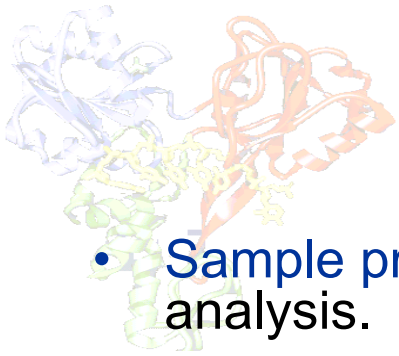
HSP 60

Prohibitin

VDAC

25 15 5 0    25 15 5 0    25 15 5 0  
Cytosolic fraction    Mitochondrial fraction    First fraction

PCT at indicated pressure, 15 cycles (30 sec up, 20 sec down)  
MIB (HEPES, BSA, Sucrose), differential centrifugation



## Conclusions:

- Sample preparation frequently present a **bottleneck** in biomarker analysis.
- Pressure cycling technology is applicable to a **wide variety of applications**, including initial steps of sample preparation for genomics and proteomics.
- PCT should be considered as an **orthogonal extraction technique**, not just homogenization or cell disruption method.
- PCT **acts directly on the lipid bilayers, micelles and emulsions**, promoting phase separation in poorly miscible solvent systems.
- ProteoSolve<sub>LRS</sub> kit combined with PCT allows **nearly complete tissue dissolution and fractionation** of tissue-derived molecules.
- Barocycler system provides several advantages over conventional extraction methods, including **reproducibility, safety, convenience, speed, automation and precise control** over the process.

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