

## Characterization of an Antiproliferative Factor from Interstitial Cystitis Patients

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Interstitial cystitis (IC) is a debilitating chronic painful bladder disorder from which approximately one million Americans suffer; its etiology is unknown. Abnormalities seen in the bladder of IC patients include petechial hemorrhages, ulcers that extend into the lamina propria, and thinning of the bladder epithelium to 1-2 cell layers. A novel antiproliferative factor (APF) has been described in the urine of 94% of symptomatic patients with IC that was determined to be uniquely produced by the bladder epithelial cells of these patients (1). The purified APF factor can induce several changes in normal bladder epithelial cells, including significantly decreased rates of proliferation (2).

Microcapillary reversed-phase liquid chromatography ( $\mu$ RPLC) coupled online to an ion-trap mass spectrometer (MS) operated in a data-dependent tandem MS mode was used to analyze urine preparations from IC patients. Microcapillary RPLC-MS/MS analysis of a biological isolate from the urine of patients suffering from IC and *de novo* sequencing of the active fraction resulted in the identification of a small O-linked glycopeptide (N-acetylneuramic acid/Galactose/N-acetylgalactosamine-TVPAVVVA) with potent bladder epithelial cell antiproliferative activity. The amino acid sequence of this peptide shares 100% identity with a segment in the sixth trans-membrane domain of frizzled 8 (amino acids 541-549), a receptor for Wnt proteins. Northern blot analysis demonstrates that APF is expressed solely in the bladder epithelium of IC patients with no expression evident in normal human bladder epithelial cells *in vitro*.

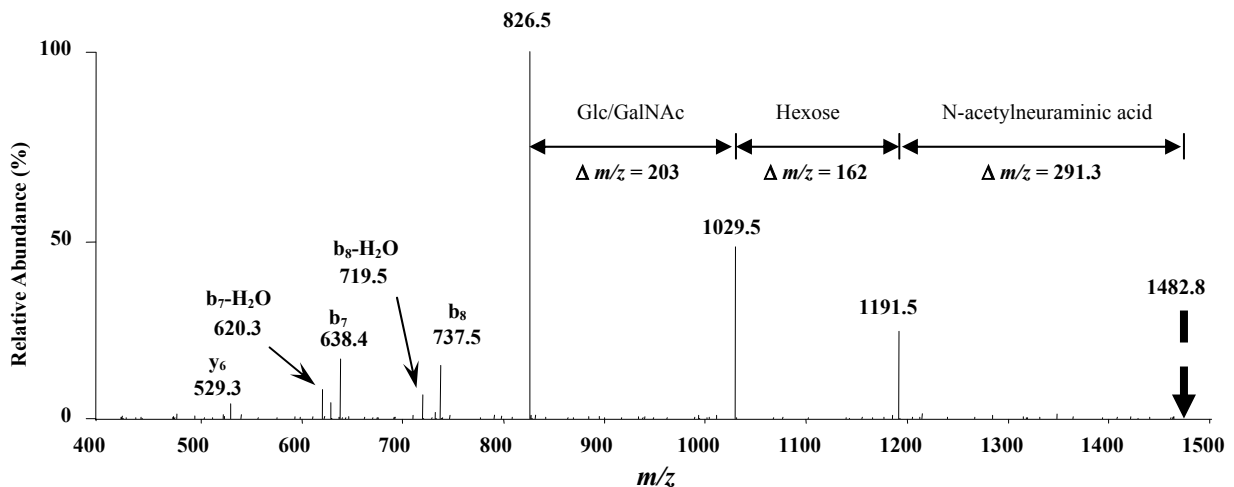


Figure 1. Tandem MS of the molecular ion at  $m/z = 1482.8$ . *De novo* sequence interpretation of this fragment ion spectrum indicates the presence of a small peptide glycosylated with N-acetylneuramic acid-hexose-N-acetyl(glucose or galactose)amine.

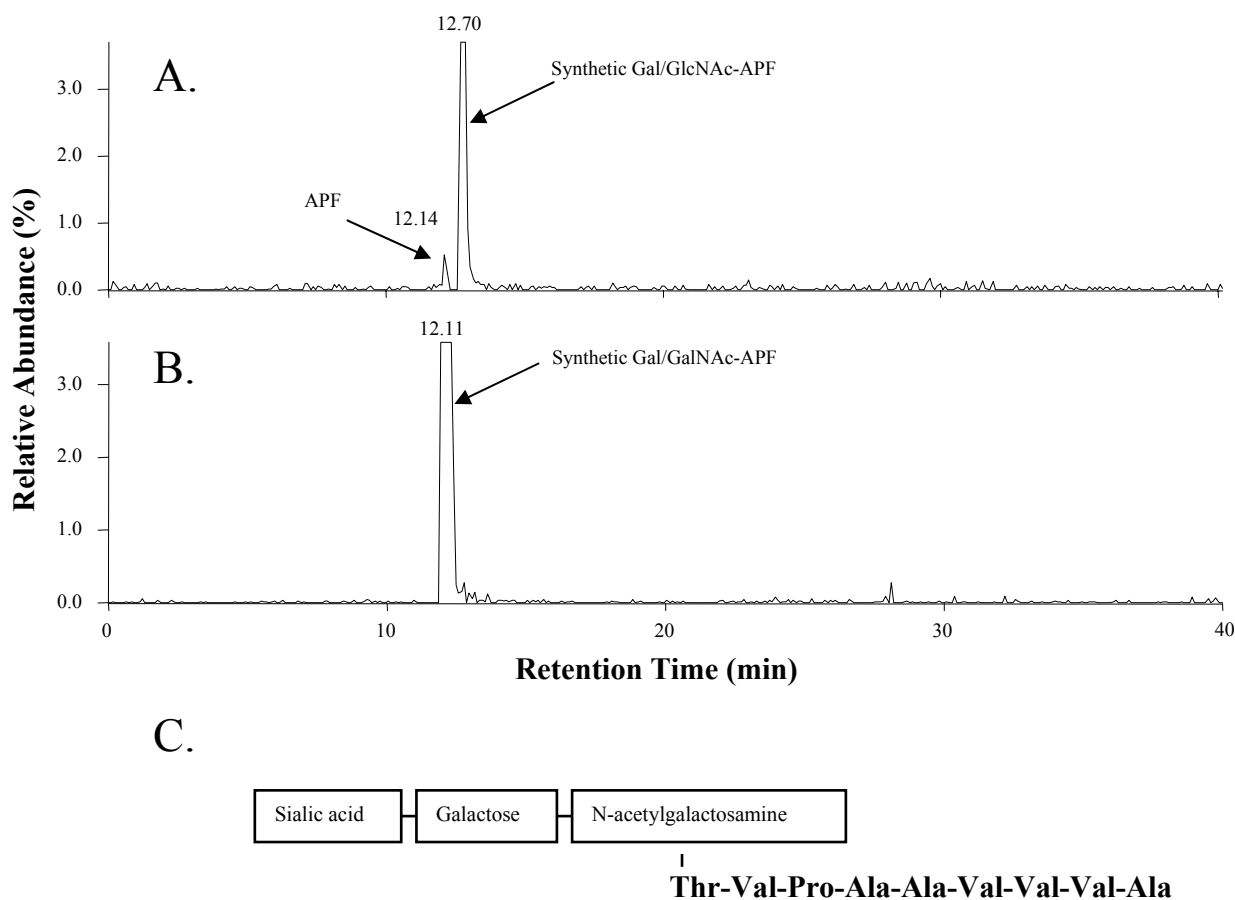


Figure 2. Total synthesis was conducted to generate both putative glycosylated forms of APF: Gal-GlcNAc-TVPAVVVA (A.) and Gal-GalNAcTVPAVVVA (B.). Each of these glycoforms was used to spike an "as isolated" APF preparation that was desialated. The two spiked APF samples were analyzed by  $\mu$ RPLC-MS/MS. The ability to resolve both glycoforms allowed the determination of native APF to be (Sialic acid)-Gal-GalNAc-TVPAVVVA (C.).

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#### References

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