

Evidence of Detrimental Effects of Environmental Contaminants on Growth and Reproductive Physiology of White Sturgeon in Impounded Areas of the Columbia River

Grant W. Feist,¹ Molly A.H. Webb,¹ Deke T. Gundersen,² Eugene P. Foster,³ Carl B. Schreck,^{1,4,5} Alec G. Maule,⁶ and Martin S. Fitzpatrick³

¹Department of Fisheries and Wildlife, Oregon State University, Corvallis, Oregon, USA; ²Environmental Science Program, Pacific University, Forest Grove, Oregon, USA; ³Oregon Department of Environmental Quality, Portland, Oregon, USA; ⁴Oregon Cooperative Fish and Wildlife Research Unit, Oregon State University, Corvallis, Oregon, USA; ⁵Biological Resources Division, U.S. Geological Survey, Corvallis, Oregon, USA; ⁶Biological Resources Division, U.S. Geological Survey, Columbia River Research Laboratory, Cook, Washington, USA

This study sought to determine whether wild white sturgeon from the Columbia River (Oregon) were exhibiting signs of reproductive endocrine disruption. Fish were sampled in the free-flowing portion of the river (where the population is experiencing reproductive success) and from three reservoirs behind hydroelectric dams (where fish have reduced reproductive success). All of the 18 pesticides and almost all of the 28 polychlorinated biphenyls (PCBs) that were analyzed in livers and gonads were detected in at least some of the tissue samples. Metabolites of *p,p'*-dichlorodiphenyltrichloroethane (DDT) [*p,p'*-dichlorodiphenyldichloroethylene (DDE) and *p,p'*-1,1-dichloro-2,2-bis(4-chlorophenyl)ethane (DDD)] were consistently found at relatively high levels in fish. Some males and immature females showed elevated plasma vitellogenin; however, concentrations were not correlated with any of the pesticides or PCBs analyzed. Negative correlations were found between a number of physiologic parameters and tissue burdens of toxicants. Plasma triglycerides and condition factor were negatively correlated with total DDT (DDD + DDE + DDT), total pesticides (all pesticides detected – total DDT), and PCBs. In males, plasma androgens and gonad size were negatively correlated with total DDT, total pesticides, and PCBs. Fish residing in the reservoir behind the oldest dam had the highest contaminant loads and incidence of gonadal abnormalities, and the lowest triglycerides, condition factor, gonad size, and plasma androgens. These data suggest that endocrine-disrupting chemicals may be accumulating behind dams over time. Overall, results of this study indicate that exposure to environmental contaminants may be affecting both growth and reproductive physiology of sturgeon in some areas of the Columbia River. **Key words:** endocrine-disrupting chemicals, growth, PCBs, pesticides, reproductive physiology, sex steroids, white sturgeon. *Environ Health Perspect* 113:1675–1682 (2005). doi:10.1289/ehp.8072 available via <http://dx.doi.org/> [Online 11 July 2005]

The lower Columbia River supports one of the most productive white sturgeon (*Acipenser transmontanus*) fisheries in North America (DeVore et al. 1995; McCabe and Tracy 1994). Fish trapped behind the dams of the hydroelectric system, however, have reduced reproductive success compared with animals in the free-flowing portion of the river (Beamesderfer et al. 1995). Reduced reproductive fitness of fish in these impounded sections of the river has been attributed to habitat, flow, and temperature, but environmental toxicants could also be playing a role. The long-lived, late-maturing, and benthic lifestyle of sturgeon may make them particularly susceptible to the actions of persistent bioaccumulating pollutants (DeVore et al. 1995).

The Columbia River receives pollution from a variety of sources that include sewage treatment plants, bleached-kraft pulp mills, aluminum smelters, mining operations, and agricultural and urban runoff. Recently, it has been determined that past operation of the hydroelectric facilities has led to contamination of certain areas of the river with polychlorinated biphenyls (PCBs) (URS Corporation 2002). A wide variety of environmental contaminants have been shown to have adverse

effects on reproduction in fishes (Kime 1995; Tyler et al. 1998; Van Der Kraak 1998), and many of these bioaccumulating toxicants have been detected in sediments and fish from the Columbia River [Foster et al. 1999, 2001a, 2001b; U.S. Environmental Protection Agency (EPA) 2002].

This study was designed to examine whether environmental pollutants are having an adverse effect on the reproductive physiology of white sturgeon in the wild and to determine whether fish demonstrate evidence of reproductive endocrine disruption that correlates to specific areas within the river where sturgeon are known to have low reproductive success.

Materials and Methods

Fish sampling. Fish were sampled during the commercial and sport harvest in February through April of 2000 and 2001. Because of state fishing regulations, only fish within a slot limit of 110–137 cm fork length were sampled. This slot limit is set to ensure that mature fish are not removed from the fishery. Fish were sampled from four areas of the Columbia River: the free-flowing portion of the river in the estuary at Astoria, Oregon,

and in reservoirs above Bonneville (river mile 191), The Dalles (river mile 216), and John Day (river mile 292) dams (Figure 1). These dams were constructed in 1938, 1960, and 1971, respectively. A total of 174 fish were sampled, representing 42–45 individuals (19–24 males and 21–23 females) for each location. Length and weight were recorded, and condition factor (CF) was determined. Gonads were removed and weighed, and gonadosomatic index (GSI) was determined. Gonads and livers were collected for both histologic and contaminant analysis. Plasma samples were collected for analysis of 17 β -estradiol (E₂), testosterone (T), 11-ketotestosterone (KT), vitellogenin (Vtg), calcium, and triacylglycerides (TAG). In 2001, pectoral fin spines were collected to determine the age of fish.

All animals were treated in accordance with Oregon State University's Care of Laboratory Animals guidelines (Oregon State University Institutional Animal Care and Use Committee 2005).

Plasma analyses. We extracted the steroids T, KT, and E₂ from plasma following the method of Fitzpatrick et al. (1986). Extraction efficiencies for all steroids were determined by adding tritiated steroids to tubes containing plasma (*n* = 4) during each extraction. This resulted in 12 extraction efficiencies for each steroid. The average extraction efficiencies (ranges) for T, KT, and E₂ were 92.5 (88.8–94.6), 82.5 (81.6–83.0), and 83.4% (79.8–85.5%), respectively. All steroid assay results were corrected for recovery.

Address correspondence to G.W. Feist, Department of Fisheries and Wildlife, 104 Nash Hall, Oregon State University, Corvallis, OR 97331-3803 USA. Telephone: (541) 737-2463. Fax: (541) 737-3590. E-mail: feistg@onid.orst.edu

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We measured plasma concentrations of T, KT, and E₂ by radioimmunoassay (RIA) as described by Sower and Schreck (1982) and modified by Feist et al. (1990). All samples were analyzed in duplicate. The lower limit of detection was 1.25 pg/tube for all assays, except KT (3.12 pg/tube). The intra- and interassay coefficients of variation for all assays were < 5 (*n* = 12) and 10% (*n* = 12), respectively. We validated steroid levels determined by RIA by verifying that serial dilutions were parallel to standard curves.

Vtg was measured by enzyme immunoassay following the methodology of Linares-Casenave et al. (1994) and Heppell and Sullivan (1999). Purified white sturgeon Vtg and antibody were a gift from S. Doroshov (University of California–Davis). The lower limit of detection was 3.9 ng/mL, and the assay was validated by verifying that serial dilutions of samples were parallel to the standard curve. The intra- and interassay coefficients of variation were < 5 (*n* = 72) and 10% (*n* = 72), respectively. We determined calcium and TAG plasma content using diagnostic kits from Sigma (587-A and 334-A; St. Louis, MO).

Histology. Gonad and liver tissue was stored in 10% phosphate-buffered formalin, embedded in paraffin, sectioned at 7 μm, and stained by hematoxylin and eosin (Luna 1968). Slides were examined under a compound scope (Motic Instruments, Inc., Richmond, B.C., Canada) using 10× to 100× objectives. We scored germ cells for stage of development according to the protocol of Van Eenennaam and Doroshov (1998). Stage 1 (differentiation of testis and ovary) and stage 2 (proliferation of spermatogonia and endogenous growth of the oocyte) fish were immature, whereas stage 3–6 males (onset of meiosis through spermiation) and stage 3–7 females (early vitellogenesis through ovulation) were classified as maturing. Each slide (liver and gonad tissue) was examined completely for presence or absence of gross lesions or other abnormalities, followed by semiquantification of macrophage aggregates (MA) in gonad and liver tissue and of eosinophils and lymphocytes in hepatic tissue in a randomly chosen field of view (10×). We formulated an index for semiquantification for the fish captured in the fisheries: 0, no MA or lymphocytes; 1, 1–25% of the tissue contained MA or lymphocytes; 2, 26–50% of the tissue contained MA or lymphocytes; 3, 51–75% of the tissue contained MA or lymphocytes; 4, 75–100% of the tissue contained MA or lymphocytes.

Contaminant analysis. We analyzed a subsample of livers (*n* = 97) and gonads (*n* = 98) for 18 chlorinated pesticides and 28 PCB congeners (Appendix 1). This represented 11–17 males and 10–14 females from each sampling location.

Extraction and cleanup procedures of sturgeon tissues were based on the methods described by Price et al. (1986) and Gundersen et al. (1998). Liver and gonad samples were homogenized using a Brinkmann Polytron tissue homogenizer (Brinkmann Instruments, Inc., Westbury, NY), and a portion was removed for measurement of moisture content. Subsamples of tissue homogenates (~ 5 g) were combined with sodium sulfate (~ 50 g) and ground to a fine powder using a mortar and pestle. Dried tissues were Soxhlet extracted (10 hr) with 170 mL of 1:1 petroleum ether/hexane (vol/vol spectral grade; Sigma-Aldrich, St. Louis, MO). Extracts were concentrated to < 15 mL with a rotary evaporator and

transferred to tared vials, where the remaining solvent was evaporated to dryness using a warm water bath and a stream of pure nitrogen (N₂). The amount of lipid in each sample was determined gravimetrically. Lipid extracts were cleaned using 20 g Florisil-packed glass columns (400 × 19 mm), and PCBs and chlorinated pesticides were eluted with 6% ethyl ether/petroleum ether (vol/vol). PCBs and pesticides were fractionated into two eluates using 5 g silica gel-packed glass columns (10.5 × 300 mm). The first fraction [PCBs and *p,p'*-dichlorodiphenyldichloroethylene (DDE)] was eluted with hexane. The second fraction (chlorinated pesticides) was eluted with benzene.

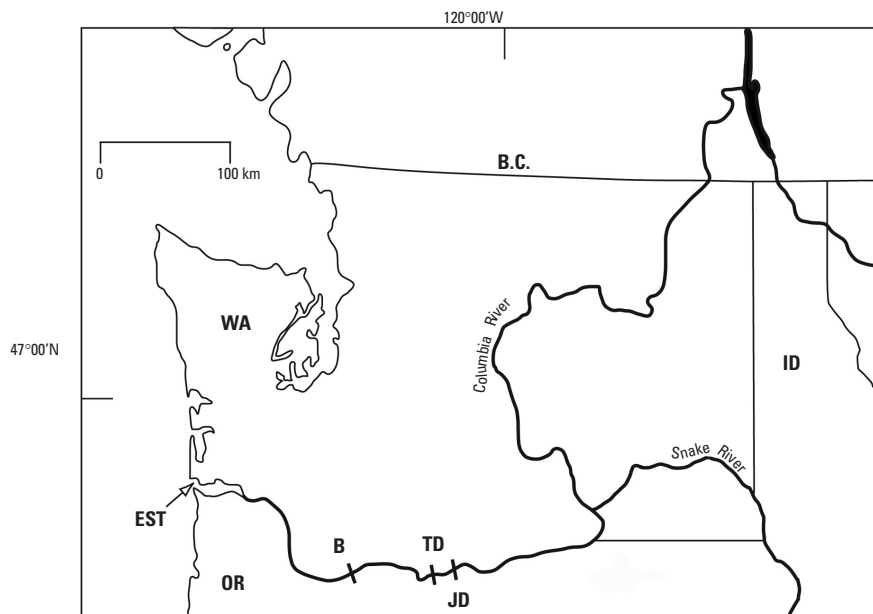


Figure 1. Sample sites for white sturgeon from the Columbia River in the estuary near Astoria, Oregon (EST), and the reservoirs behind Bonneville (B), The Dalles (TD), and John Day (JD) dams. Abbreviations: B.C., British Columbia; ID, Idaho; OR, Oregon; WA, Washington State.

Table 1. Concentration (mean ± SE) of chlorinated pesticides in livers (*n* = 97) and gonads (*n* = 98) of white sturgeon from the Columbia River.

Pesticide	Liver		Gonad	
	D	Lipid (μg/g)	D	Lipid (μg/g)
Aldrin	2	0.002 ± 0.002	5	0.011 ± 0.006
α-BHC	19	0.039 ± 0.009	26	0.023 ± 0.005
β-BHC	14	0.115 ± 0.046	11	0.023 ± 0.005
γ-BHC	8	0.024 ± 0.011	21	0.047 ± 0.014
δ-BHC	9	0.019 ± 0.007	15	0.154 ± 0.127
<i>p,p'</i> -DDD	86	1.863 ± 0.544	93	1.619 ± 0.400
<i>p,p'</i> -DDE	97	18.40 ± 7.313	98	10.60 ± 2.086
<i>p,p'</i> -DDT	28	0.274 ± 0.103	41	0.259 ± 0.073
Dieldrin	16	0.134 ± 0.045	15	0.031 ± 0.009
Endrin	10	0.114 ± 0.060	11	0.022 ± 0.007
Endrin aldehyde	16	0.108 ± 0.062	13	0.064 ± 0.032
Endrine ketone	8	0.038 ± 0.165	2	0.010 ± 0.007
Endosulfan I	34	0.161 ± 0.044	45	0.133 ± 0.025
Endosulfan II	9	0.108 ± 0.051	14	0.087 ± 0.047
Endosulfan sulfate	3	0.005 ± 0.003	8	0.008 ± 0.003
Heptachlor	8	0.018 ± 0.008	13	0.037 ± 0.019
Heptachlor epoxide	15	0.081 ± 0.031	25	0.074 ± 0.024
<i>p,p'</i> -Methoxychlor	14	0.112 ± 0.044	5	0.027 ± 0.017

Abbreviations: BHC, benzene hexachloride; D, number of detections.

We analyzed the cleaned fractions using a Varian CP-3800 gas chromatograph (Varian, Inc., Walnut Creek, CA) equipped with a ^{63}Ni electron capture detector, a CP-8200 AutoSampler, a Star Chromatography Workstation (version 5; Varian Inc.), and an SPB-608 fused silica capillary column (30 mm \times 0.25 mm \times 0.25 μm film thickness; Supelco, Bellefonte, PA). Gas chromatographic parameters used were as follows: carrier gas, helium (1.5 mL/min); makeup gas, nitrogen; detector temperature, 300°C; injector temperature, 290°C; and oven temperature,

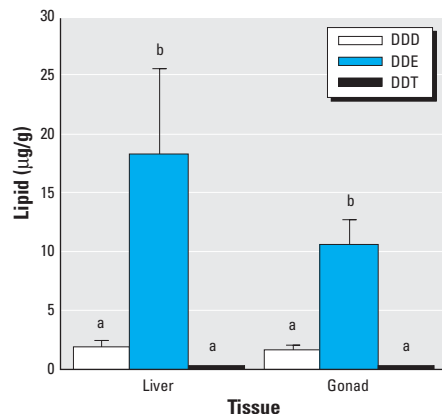


Figure 2. Mean concentrations (\pm SE) of DDT and its metabolites in livers ($n = 97$) and gonads ($n = 98$) of white sturgeon from all sample areas combined. Means with different letters indicate a significant difference within a tissue (ANOVA, $p < 0.05$).

Table 2. Concentration (mean \pm SE) of PCBs in livers ($n = 97$) and gonads ($n = 98$) of white sturgeon from the Columbia River.

Pesticide (IUPAC no.)	Liver		Gonad	
	D	Lipid ($\mu\text{g/g}$)	D	Lipid ($\mu\text{g/g}$)
28	3	0.020 \pm 0.011	0	
44	6	0.055 \pm 0.042	4	0.004 \pm 0.002
52	3	0.038 \pm 0.024	3	0.024 \pm 0.105
60	19	0.125 \pm 0.033	11	0.163 \pm 0.129
66	8	0.131 \pm 0.066	2	0.025 \pm 0.020
74	2	0.008 \pm 0.006	4	0.037 \pm 0.022
87	1	0.006 \pm 0.006	2	0.008 \pm 0.006
99	12	0.101 \pm 0.036	12	0.077 \pm 0.041
101	28	0.238 \pm 0.088	24	0.217 \pm 0.131
105	14	0.135 \pm 0.051	9	0.033 \pm 0.016
110/77	12	0.060 \pm 0.019	17	0.128 \pm 0.050
118	9	0.054 \pm 0.020	10	0.152 \pm 0.085
126	6	0.035 \pm 0.016	5	0.024 \pm 0.018
128	1	0.007 \pm 0.007	6	0.043 \pm 0.031
138	28	0.258 \pm 0.071	28	0.233 \pm 0.072
151	4	0.025 \pm 0.015	7	0.032 \pm 0.014
153	18	0.264 \pm 0.101	20	0.157 \pm 0.062
156	6	0.035 \pm 0.018	7	0.013 \pm 0.006
169	2	0.007 \pm 0.005	0	
170	3	0.006 \pm 0.003	3	0.003 \pm 0.001
180	3	0.030 \pm 0.026	3	0.001 \pm 0.001
183	9	0.042 \pm 0.015	13	0.029 \pm 0.010
187	20	0.163 \pm 0.047	21	0.113 \pm 0.032
194	4	0.018 \pm 0.009	1	0.001 \pm 0.001
199	10	0.022 \pm 0.007	10	0.065 \pm 0.030
203/170	10	0.043 \pm 0.017	10	0.016 \pm 0.008

Abbreviations: D, number of detections; IUPAC, International Union of Pure and Applied Chemistry.

150°C (4 min) to 290°C (10 min) at 8°C/min. Organochlorine pesticides were quantified from individually resolved peak areas with corresponding peak areas of external standards (Supelco). We used individual PCB congeners purchased from AccuStandard (New Haven, CT) to make external standards containing the 28 selected PCB congeners.

Quality assurance measures included the analysis of reagent blanks, duplicates, and matrix spike samples. Percent recoveries of PCB congeners and organochlorine pesticides in matrix spikes were between 90 and 110%; therefore, sample extracts were not corrected for percent recovery. Detection limits for individual PCB congeners and chlorinated pesticides were 0.01 $\mu\text{g/g}$ wet weight. The State of Oregon Environmental Quality Laboratories and Applied Research, Organic Laboratory section (Portland, OR), analyzed two tissue homogenates for chlorinated pesticides (interlaboratory comparison). The relative percent difference of organochlorine pesticide concentrations reported by the two laboratories in the two samples differed by an average of $< 17\%$.

Aging of fish. Ages of fish sampled in 2001 were determined by pectoral fin spine analysis following the procedures described by Beamesderfer et al. (1989). Two independent determinations were conducted at the Oregon Department of Fish and Wildlife (Clackamas, OR) and at University of California–Davis (Davis, CA). Of the fish, 27% had identical age assignments by the different readers, 45% were aged within 1 year, 22% within 2 years, 2% within 3 years, and 4% > 5 years. We averaged ages of fish that were not in agreement between the two determinations.

Western blot analysis. Hepatic microsomes were prepared by differential centrifugation according to Carpenter et al. (1990) and stored at -80°C until use. Briefly, livers were minced in ice-cold buffer (0.1 M Tris-acetate, pH 7.4; 0.1 M KCl; 1 mM EDTA; 20 μM butylated hydroxytoluene; and 1 mM phenylmethylsulfonylfluoride) and homogenized in 4 volumes of the same buffer. The homogenate was centrifuged at $10,000 \times g$ for 30 min, and the resulting supernatant was centrifuged at $100,000 \times g$ for 90 min. The microsomal pellet was resuspended in buffer (0.1 M phosphate buffer, pH 7.25; 20% glycerol; and 1 mM EDTA). Microsomes were stored at -80°C until use.

We measured the putative white sturgeon hepatic cytochrome P450 3A (CYP3A) enzyme in microsomes by Western blotting using a polyclonal antibody generated against rainbow trout LMC5 (3A27). Microsomal CYP3A protein was measured using Western immunoblot techniques according to Towbin et al. (1979) with modifications. Briefly, sodium dodecyl sulfate polyacrylamide gel

electrophoresis (SDS-PAGE) was performed using 8% polyacrylamide precast minigels. We prepared membranes according to the manufacturers recommendations, and proteins were transferred to membranes followed by incubation with rabbit anti-trout antibody (a generous gift from D. Buhler). Membranes were rinsed with phosphate-buffered saline–Tween and incubated with horseradish peroxidase-conjugated secondary antibodies (anti-rabbit) for detection of oxidized luminol (Amersham Biosciences, Piscataway, NJ). The chemiluminescent signal was captured on film (Hyperfilm ECL, Amersham Biosciences), and films were scanned for quantification.

Statistics. We conducted all mean comparisons between physiologic parameters, tissue contaminant load, river location, and sex of fish using a one-way analysis of variance (ANOVA) with a Bonferroni procedure. All correlations between tissue contaminant load and physiologic parameters were conducted using reciprocal-Y regression. We performed all analyses using Statview software (Abacus Concepts, Inc., Berkeley, CA), and the accepted level of significance for all tests was $p < 0.05$.

Results

All 18 of the chlorinated pesticides examined in tissues from wild fish were detected in at least some of the samples (Table 1). We consistently found relatively high levels of metabolites of *p,p'*-dichlorodiphenyl-trichloroethane (DDT) [DDE and *p,p'*-1,1-dichloro-2,2-bis(4-chlorophenyl)ethane (DDD)] in fish. Concentrations of DDE were always greater than those of DDD and DDT in both livers and gonads (Figure 2). We found no differences in toxicant levels between tissues. Of the 28 PCB congeners examined, 26 were detected in at least some of the samples (Table 2).

Total DDT (DDD + DDE + DDT), total pesticides (all pesticides detected – total DDT), and PCBs (total of all detected) were significantly higher in livers and gonads of fish from Bonneville Reservoir compared with other locations (Figure 3). Fish from the Bonneville Reservoir had significantly lower TAG plasma concentrations and GSI than two of the other locations (Figure 4). Fish from Bonneville also had significantly lower calcium plasma concentrations and CF compared with all other locations.

We found a negative correlation between plasma TAG and total DDT, pesticides, and PCBs in livers (Table 3). To varying degrees, this was also true for TAG compared with contaminants in gonads and for CF compared with contaminants in livers and gonads. Although we observed significant relationships, r^2 values indicated that a large amount of variation was present within the data.

Plasma concentrations of T were higher in males than in females at all sample locations except Bonneville (Figure 5). Males from the estuary had significantly higher levels of KT than did females, but this was not observed at other locations. Males from the estuary had significantly higher plasma T and KT than did males in the Bonneville and John Day reservoirs. Plasma concentrations of E₂ were very low in all fish examined (Table 4). We observed no differences between either sex or location.

Plasma Vtg was at or very near the detection limit of the assay for all fish sampled in the estuary and Bonneville (Figure 5). Some males and immature females from The Dalles and John Day reservoirs had detectable levels of Vtg. Males from John Day had significantly higher concentrations of Vtg than did fish from all other locations. Females from The Dalles had concentrations of Vtg that were nearly significant compared with females from the estuary (*p* = 0.060) as well as compared with females from Bonneville (*p* = 0.058). There was no correlation between plasma Vtg and any of the pesticides or PCBs that were monitored.

Gonadal histology revealed a total of 82 females, 73 males, and 3 hermaphrodites from the 2 years of sampling. Sixteen gonad samples contained only adipose tissue and no gonial cells. Of the females, 81 were immature (all stage 2 except for 3 stage 1 females), and 1 was a maturing female (stage 3; early vitellogenesis). Of the males, 66 were immature (all stage 2), 1 was in stage 3 of gonadal development (onset of meiosis), and 6 were in stage 5 of development (spermiation). No maturing fish were captured in Bonneville Reservoir. All of the maturing males had significantly higher levels of plasma androgens (T, 92.2 ± 20.9; KT, 84.0 ± 16.4 ng/mL) than did immature males (T, 5.1 ± 1.1; KT, 4.3 ± 1.0 ng/mL). All 3 of the hermaphroditic fish had predominately female ovotestes. Two of the 3 fish were captured in Bonneville Reservoir, and the other was from the estuary. Several fish showed irregular ovarian plasma membranes and

intrusion of muscle into the ovary. MAs were found in both female and male gonadal tissue and were most often found to contain melanin.

Liver histology revealed a high incidence of MA and lymphocytes. However, no pattern was discernible with regard to contaminant level. We found a very high incidence of MA

and/or lymphocytes in liver samples from 11 fish; of these, 7 were from the Bonneville Reservoir, 2 were from the estuary, and 1 each were from The Dalles and John Day reservoirs.

We found a negative correlation between plasma T and total DDT, pesticides, and PCBs in livers of male white sturgeon

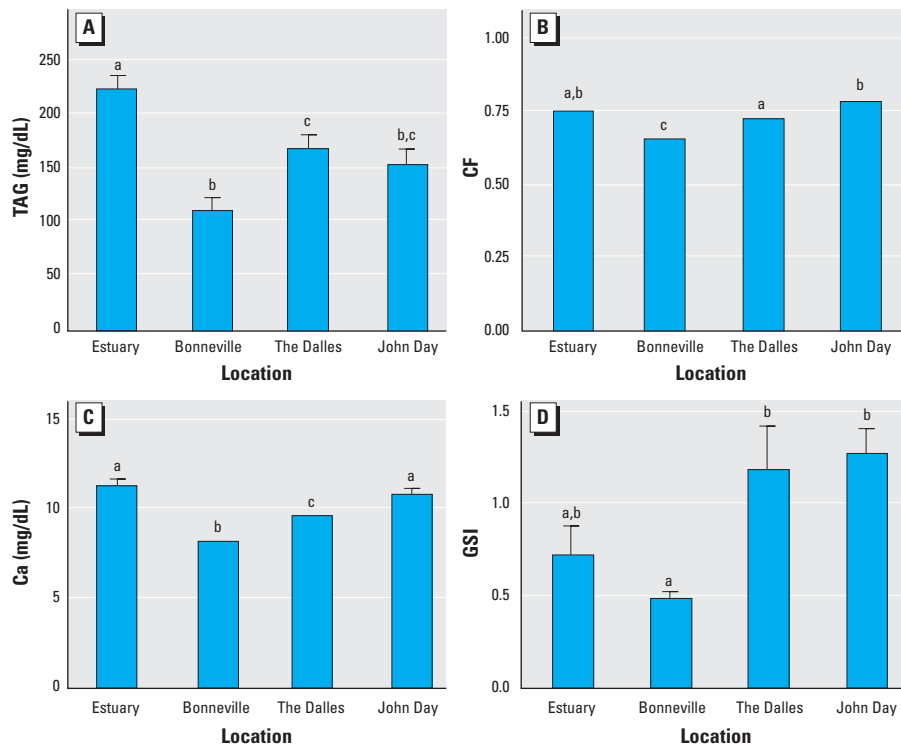


Figure 4. Mean plasma concentrations (± SE) of TAG (A), CF (B), calcium (C), and GSI (D) in white sturgeon from four locations on the Columbia River. Each bar represents a sample size of 42–45. Means with different letters indicate a significant difference between locations (ANOVA, *p* < 0.05).

Table 3. Regression analyses of TAG and CF versus various contaminants in livers and gonads of Columbia River white sturgeon.

Contaminant	Liver				Gonad			
	TAG		CF		TAG		CF	
	<i>r</i> ²	<i>p</i> -Value	<i>r</i> ²	<i>p</i> -Value	<i>r</i> ²	<i>p</i> -Value	<i>r</i> ²	<i>p</i> -Value
Total DDT	0.60	< 0.001	0.08	< 0.005	0.20	< 0.001	0.11	< 0.001
Total pesticides	0.48	< 0.001	0.15	< 0.001	0.04	< 0.050	0.18	< 0.001
Total PCBs	0.60	< 0.001	0.11	< 0.002	0.10	< 0.002	0.07	< 0.008

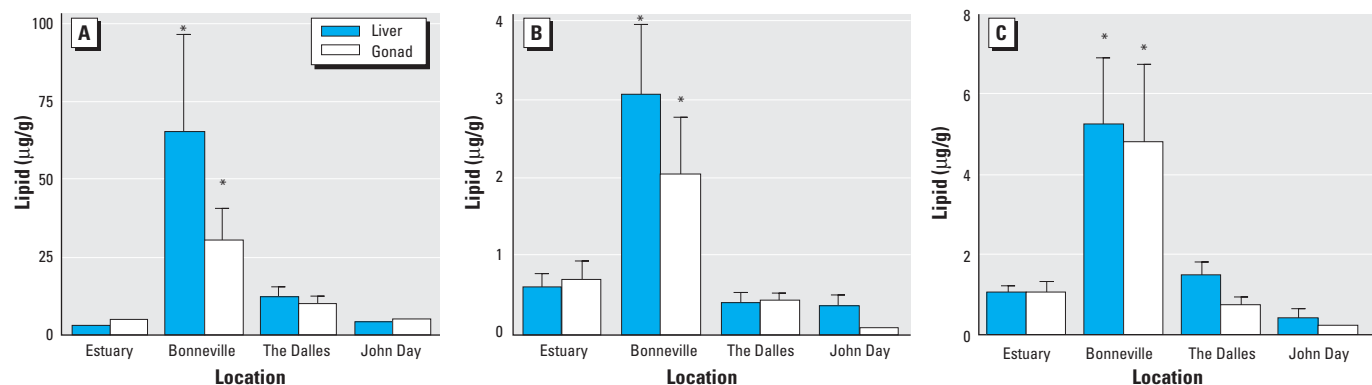


Figure 3. Concentrations (mean ± SE) of total DDT (A; DDD + DDE + DDT), total pesticides (B; all pesticides detected – total DDT), and PCBs (C; total of all detected) in livers and gonads of white sturgeon from four locations on the Columbia River. Each bar represents a sample size of 22–28.

*Statistically different from other locations (ANOVA, *p* < 0.05).

(Figure 6). We also observed these relationships for contaminants in gonads (Figure 7). To varying degrees, this was also true for plasma KT and GSI compared with contaminants in gonads and livers (Table 5).

Spermatogonia proliferation (stage 2) in white sturgeon is associated with increased

circulating androgen concentrations regardless of age or size (Feist et al. 2004). In immature wild white sturgeon, T concentrations > 4 ng/mL may be used to differentiate stage 2 males from stage 1 males and immature females (Webb et al. 2002). All 66 immature males in our study were in stage 2 of gonadal

development, yet 47 (71.2%) had plasma T concentrations that were < 4 ng/mL. Of the 48 stage 2 males that were analyzed for toxicants, 31 had levels of T < 4 ng/mL. In addition, no males with liver contaminant levels > 9.5 ppm (total DDT), > 5.6 ppm (total pesticides), or > 2.8 ppm (PCBs) had plasma T concentrations > 4 ng/mL (Figure 6). Where this was observed, concentrations of toxicants in gonads were 11.6, 3.7, and 2.5 ppm, for total DDT, total pesticides, and PCBs, respectively (Figure 7).

Age determination of fish by pectoral fin spine analysis in 2001 revealed that sturgeon from Bonneville (18.3 ± 1.0 years; range, 14–27) and John Day (17.4 ± 0.4 years; range, 14–20) were significantly older than those sampled in The Dalles (14.8 ± 0.5 years; range, 10–19). Bonneville fish were also significantly older than estuary fish (14.6 ± 1.0 years; range, 10–17).

To investigate the possibility that DDE reduces plasma androgens by increasing steroid metabolism and excretion via up-regulation of liver cytochrome P450 isozymes, we conducted a preliminary and purely qualitative Western blot analysis to measure the putative CYP3A in microsomes. In trout this enzyme is responsible for hydroxylating steroids as a first step for metabolism and excretion (Lee et al. 2001). A Western blot for this isozyme is shown in Figure 8. Male sturgeon with higher liver content of DDE showed increased immunoreactivity for CYP3A.

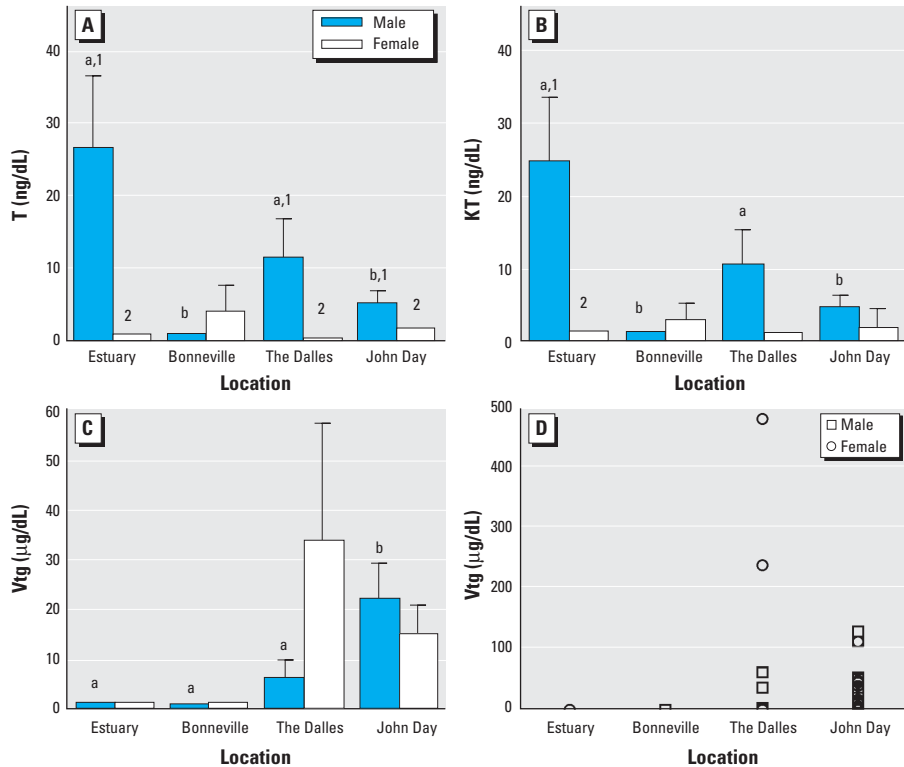


Figure 5. Mean plasma concentrations (± SE) of T (A), KT (B), and Vtg (C) and individual Vtg concentrations (D) in male and immature female white sturgeon from four locations on the Columbia River. Each bar represents a sample size of 19–24 (A–C). Means with different letters or numbers indicate a significant difference between locations or between sexes within a location, respectively (ANOVA, $p < 0.05$).

Table 4. Concentration (mean ± SE) of plasma E_2 (ng/mL) in male ($n = 19–24$) and female ($n = 21–23$) white sturgeon at four locations from the Columbia River.

	Estuary	Bonneville	The Dalles	John Day
Female	0.09 ± 0.02	0.11 ± 0.03	0.13 ± 0.02	0.28 ± 0.05
Male	0.16 ± 0.03	0.07 ± 0.01	0.14 ± 0.03	0.38 ± 0.10

Discussion

The life history of white sturgeon may make them particularly susceptible to the actions of persistent bioaccumulating pollutants. These fish are bottom dwellers and feed on benthic prey items that are closely associated with sediments containing hydrophobic pollutants. Sturgeon can live for > 100 years, and females mature between 16 and 35 years of age (DeVore et al. 1995). Thus, toxicants may

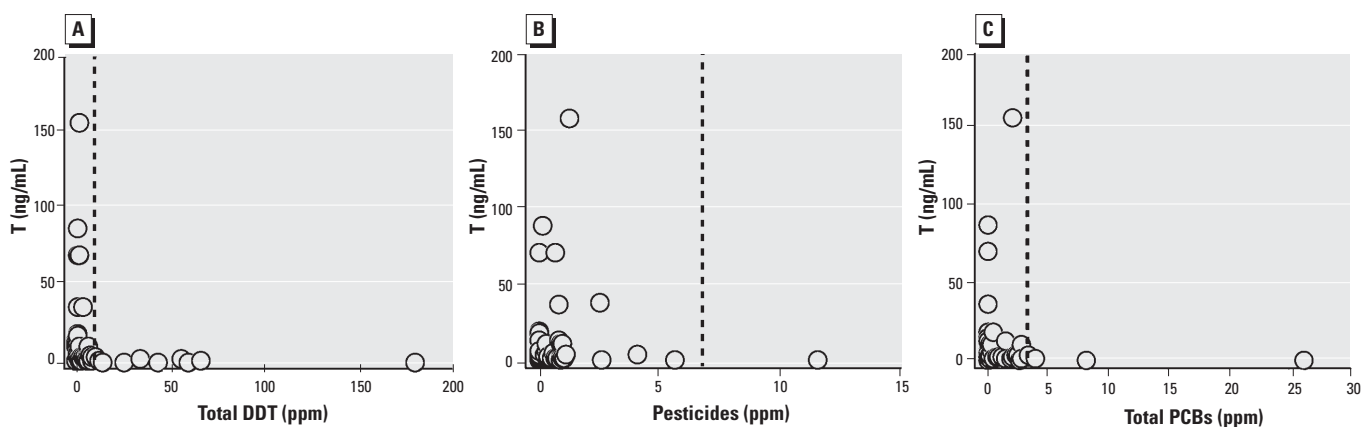


Figure 6. Individual plasma T versus total DDT (A), total pesticides (B), or total PCB (C) concentrations in livers of male white sturgeon. Reciprocal-Y regression: $p < 0.001$ and $r^2 = 0.79$ for DDT, $p < 0.001$ and $r^2 = 0.56$ for pesticides, and $p < 0.001$ and $r^2 = 0.80$ for PCBs. All males with toxicant levels higher than those denoted by the vertical dashed line have < 4 ng/mL T.

accumulate and have deleterious effects over a long period of time before the fish reach a stage when they are able to reproduce. A recent study in the Columbia River found that sturgeon contained the highest body burdens of contaminants out of 12 species of fish examined (U.S. EPA 2002). Levels of toxicants seen in the present study were comparable with those found by the U.S. EPA and also comparable with levels previously reported by our laboratory (Foster et al. 2001a, 2001b).

Fish trapped behind the oldest of the dams examined (Bonneville) had the highest contaminant loads and the lowest CF, gonad size, and plasma androgens and triglycerides. These fish also had the highest incidence of gonadal abnormalities. This suggests that endocrine-disrupting chemicals (EDCs) may be accumulating behind dams over time.

It has recently been determined that past operation of the dam at Bonneville has resulted in areas within the reservoir that have very high levels of PCBs (URS Corporation 2002). In our study, Bonneville fish were older than fish from two of the other sampling locations. Fish from this reservoir also grow slower, and females mature at a later age than other locations (Beamesderfer et al. 1995). Thus, these fish may be exposed to higher levels of contaminants and for longer periods of time than comparably sized fish from other areas of the river. Food availability may be the main cause for reduced growth in Bonneville fish, but effects of toxicants cannot be ruled out. The negative correlations found between plasma triglycerides and CF with tissue burdens of pesticides and PCBs add strength to this possibility.

Our laboratory has previously documented a negative correlation between plasma androgens and tissue content of *p,p'*-DDE for Columbia River sturgeon (Foster et al. 2001b). In the present study, we observed negative correlations between both plasma

androgens and GSI of males compared with total DDT, total pesticides, and PCBs. Our sample size for this study was much greater than our previous research, which may explain why these relationships were not seen in the earlier study. *p,p'*-DDE has also been shown to have demasculinizing effects in the guppy (*Poecilia reticulata*) (Baatrup and Junge 2001; Bayley et al. 2002). Our data also suggest that DDT and its metabolites may reach threshold levels in liver and gonad above which the fish are incapable of elevating plasma T concentrations. This may result in the inability of males with high body burdens of contaminants to attain sexual maturity.

We have preliminary evidence that the mechanism of action of plasma androgen reduction by *p,p'*-DDE, or possibly by other pesticides or PCBs, is by increasing steroid metabolism through up-regulation of CYP3A. DDE has been shown to induce this isozyme and increase metabolism of T in mice (Dai et al. 2001). Rainbow trout (*Oncorhynchus mykiss*) injected with DDE, however, showed a decrease in CYP3A-dependent 6 β -hydroxylation of T (Machala et al. 1998). The dose

used for the rainbow trout study was much higher (50 mg/kg) than levels seen in wild fish in our study and may not have simulated the effects of chronic exposure to lower concentrations of DDE.

Our finding that plasma androgens were higher in males than females (except in the Bonneville Reservoir) has been previously documented by our laboratory (Foster et al. 2001a, 2001b). We have used differences in plasma steroids between males and females to develop a model for sexing both immature and maturing wild white sturgeon and for determining sex of cultured fish at an early age (Feist et al. 2004; Webb et al. 2002).

Although banned for use in the United States in 1973, DDT and its metabolites are still being detected in sturgeon at relatively high levels. This indicates that this compound is extremely persistent in the environment. Tissue burdens were always DDE > DDD and DDT, indicating that aerobic degradation of DDT (yielding primarily DDE) is the main metabolic pathway as opposed to anaerobic degradation (yielding primarily DDD) (Spencer et al. 1996). This suggests that the most likely source of DDT metabolites is from agricultural runoff of

Table 5. Regression analyses of KT and GSI versus various contaminants in livers and gonads of male Columbia River white sturgeon.

Contaminant	Liver				Gonad			
	r^2	p -Value	r^2	p -Value	r^2	p -Value	r^2	p -Value
Total DDT	0.08	< 0.050	0.24	< 0.001	0.11	< 0.020	0.21	< 0.001
Total pesticides	NS	NS	0.15	< 0.006	NS	NS	0.22	< 0.001
Total PCBs	0.16	< 0.004	NS	NS	NS	NS	0.10	< 0.030

NS, not significant.

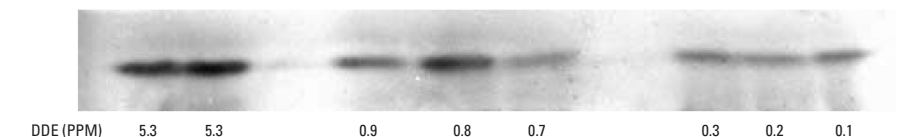


Figure 8. Western blot of CYP3A protein in individual livers of male white sturgeon with varying levels of liver DDE.

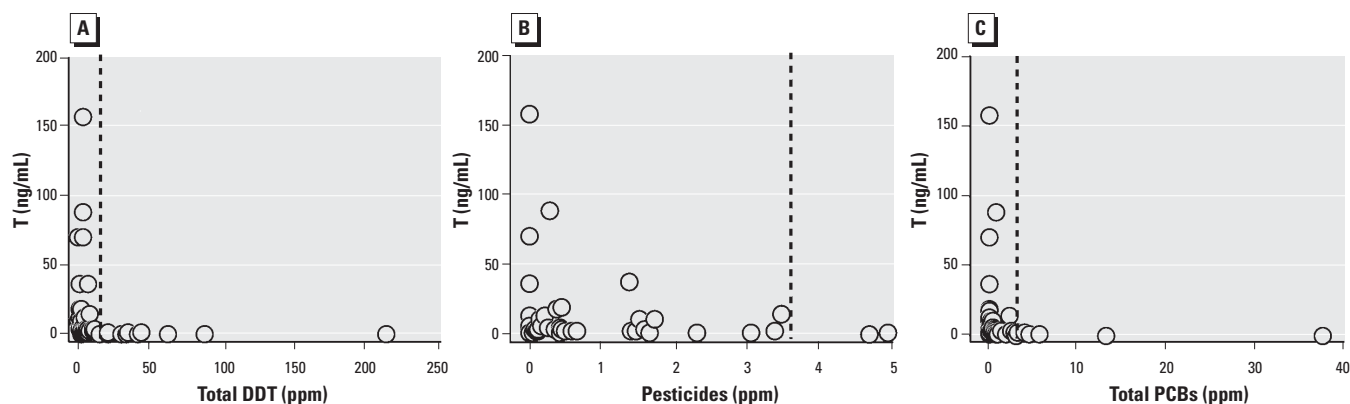


Figure 7. Individual plasma T versus total DDT (A), total pesticides (B), or total PCB (C) concentrations in gonads of male white sturgeon. Reciprocal-Y regression: $p < 0.001$ and $r^2 = 0.85$ for DDT, $p < 0.001$ and $r^2 = 0.31$ for pesticides, and $p < 0.001$ and $r^2 = 0.82$ for PCBs. All males with toxicant levels higher than those denoted by the vertical dashed line have < 4 ng/mL T.

the parent compound as opposed to anaerobic degradation of DDT in sediments.

The type and source of xenoestrogen(s) responsible for elevating plasma Vtg in males and immature females from The Dalles and John Day reservoirs remains uncertain. None of the pesticides or PCBs monitored in this study was correlated with plasma Vtg. Fish exposed in our laboratory to the pesticides (permethrin and pyriproxyfen) or herbicides (atrazine and simazine) that are currently being used in agricultural practices in the Columbia basin did not show increases in plasma Vtg (data not shown). Caged sturgeon, in areas of the river where some wild fish had elevated Vtg, also did not show an increase in this protein (data not shown). This suggests that wild fish either are being exposed to potential EDCs for longer periods of time or are bioaccumulating them through ingestion of prey.

Other candidates for induction of Vtg include the alkylphenols, which have been shown to be weakly estrogenic in fish (Jobling et al. 1996; White et al. 1994). Fish exposed to octylphenol and nonylphenol in our laboratory experienced increased plasma Vtg (data not shown), but we are unable to find a likely source for alkylphenolic compounds in The Dalles and John Day reservoirs. There are many sources of alkylphenols in the estuary and Bonneville Reservoir, yet we found no elevated Vtg in wild sturgeon sampled in this area of the river. The cause of elevated Vtg in wild fish is most likely due to other EDCs or

metabolites of toxicants not yet identified, or combinations of compounds.

The overall results of this study indicate that exposure to environmental contaminants may be affecting both growth and reproductive physiology of sturgeon in some areas of the Columbia River. Questions remain, however, as to what effects these contaminants have on the ability of sturgeon to successfully reproduce. It is unknown if lowered energy reserves, GSI, and androgens, and elevated Vtg actually inhibit or decrease the ability of sturgeon to mature and spawn. Because of the slot-size limit (fish that are 110–137 cm in fork length), most wild fish sampled in this study were immature. Larger sturgeon that have reached a sufficient size and age to mature must be examined to determine possible deleterious effects of contaminants on reproduction. Different year classes of sturgeon also need to be investigated to determine if toxicants are bioaccumulating as the fish age. Finally, prey items need to be examined for the presence of EDCs to determine if sturgeon are acquiring these compounds from their diet or other sources.

The poor reproductive success of sturgeon in impounded areas of the Columbia River is most likely due to a wide variety of stressors, including food availability, poor spawning habitat, and changes in flow and temperature. Exposure to environmental contaminants may be an additional stressor that is contributing to this reduced reproductive fitness.

Appendix 1. Chlorinated pesticides and PCBs measured in Columbia River white sturgeon livers and gonads.

Chlorinated pesticide	PCB (IUPAC no.)
Aldrin	2,2',5'-Trichlorobiphenyl (18)
α-BHC	2,4,4'-Trichlorobiphenyl (28)
β-BHC	2,2',3,5'-Tetrachlorobiphenyl (44)
γ-BHC	2,2',5,5'-Tetrachlorobiphenyl (52)
δ-BHC	2,3,4,4'-Tetrachlorobiphenyl (60)
p,p'-DDD	2,3',4,4'-Tetrachlorobiphenyl (66)
p,p'-DDE	2,4,4',5'-Tetrachlorobiphenyl (74)
p,p'-DDT	3,3',4,4'-Tetrachlorobiphenyl (77)
Dieldrin	2,2',3,4,5'-Pentachlorobiphenyl (87)
Endrin	2,2',4,4',5'-Pentachlorobiphenyl (99)
Endrin aldehyde	2,2',4,5,5'-Pentachlorobiphenyl (101)
Endrine ketone	2,3,3',4,4'-Pentachlorobiphenyl (105)
Endosulfan I	2,3,3',4',6'-Pentachlorobiphenyl (110)
Endosulfan II	2,3',4,4',5'-Pentachlorobiphenyl (118)
Endosulfan sulfate	3,3',4,4',5'-Pentachlorobiphenyl (126)
Heptachlor	2,2',3,3',4,4'-Hexachlorobiphenyl (128)
Heptachlor epoxide	2,2',3,4,4',5'-Hexachlorobiphenyl (138)
p,p'-Methoxychlor	2,2',3,5,5',6'-Hexachlorobiphenyl (151)
	2,2',4,4',5,5'-Hexachlorobiphenyl (153)
	2,3,3',4,4',5'-Hexachlorobiphenyl (156)
	3,3',4,4',5,5'-Hexachlorobiphenyl (169)
	2,2',3,3',4,4',5'-Heptachlorobiphenyl (170)
	2,2',3,4,4',5,5'-Heptachlorobiphenyl (180)
	2,2',3,4,4',5',6'-Heptachlorobiphenyl (183)
	2,2',3,4',5,5',6'-Heptachlorobiphenyl (187)
	2,2',3,3',4,4',5,5'-Octachlorobiphenyl (194)
	2,2',3,3',4,4',5,5',6'-Octachlorobiphenyl (199)
	2,2',3,4,4',5,5',6'-Octachlorobiphenyl (203)

Abbreviations: BHC, benzene hexachloride; IUPAC, International Union of Pure and Applied Chemistry.

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The American Plastics Council respectfully requests that *EHP* address the misinformation that appeared in these articles and which is available on the *EHP* website.

The author is employed by the American Chemistry Council/American Plastics Council.

Patricia A. Enneking
American Plastics Council
Arlington, Virginia
E-mail: patty_enneking@plastics.org

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Editor's note: The following erratum was published in the January 2006 issue (Environ Health Perspect 114:A21):

In the October articles "Children's Centers Study Kids and Chemicals" [Environ Health Perspect 113:A664–A668 (2005)] and "Are EDCs Blurring Issues of Gender?" [Environ Health Perspect 113:A670–A677 (2005)], photographs and their captions erroneously imply that plastic drink bottles contain ortho-phthalates. Plastic drink bottles sold in the United States are made from polyethylene terephthalate and do not contain ortho-phthalates. Also, at the end of the EDCs article, references are made to plastic wrap and Saran Wrap. For clarification, neither plastic wrap nor Saran Wrap contains ortho-phthalates. EHP regrets these errors.

ERRATA

Azziz-Baumgartner et al. noticed two errors in "Case-Control Study of an Acute Aflatoxicosis Outbreak—Kenya?" [*Environ Health Perspect* 113:1779–1783]. The units in Figure 2 and Table 2 should be nanograms per milligram instead of micrograms per milligram. The errors were introduced when new figures and tables were generated during the final revision of the paper. The authors apologize for these errors.

In the article by Feist et al. [*Environ Health Perspect* 113:1675–1682], the units were incorrect in several figures and tables: "Lipid ($\mu\text{g/g}$)" should be " $\mu\text{g/g}$ lipid" in Tables 1 and 2 and in the y -axes of Figures 2 and 3A–C. Also, on the y -axes in Figure 5A–D, "dL" should be "mL." *EHP* regrets these errors.

The photograph on page A29 of the January 2006 NIEHS News section should have been credited to Jennifer Gorenstein/UTMDACC COEP. The photographs on page A30 should have been credited to Tom Van Biersel/Louisiana Geological Survey (left) and Bryan Parras/UTMB (right). Additionally, Parras's photograph depicts residents of Pointe-aux-Chenes, not LaRose, and includes no COEP staff.

In the Beyond the Bench article in this same section, "COEPs Contribute to Hurricane Relief" [*Environ Health Perspect* 114:A30–A31 (2006)], Peter Thorne was incorrectly identified as director of the University of Iowa COEP; he is in fact director of the University of Iowa Environmental Health Sciences Research Center as well as head of the NIEHS Working Group on Mold, Microbial Agents, and Respiratory Diseases. It was the latter group that "collected air and surface samples from water-damaged homes in New Orleans" as our article stated. Finally, the aid teams that traveled throughout Louisiana included members from the UTMDACC COEP as well as the UTMB COEP.

EHP regrets the errors.

In the January Focus article "In Katrina's Wake" [*Environ Health Perspect* 114:A32–A39 (2005)], Hurricane Katrina was identified as a Category 4 storm, reflecting statements from the National Hurricane Center as of press time. The National Hurricane Center has since reported that Katrina was actually a Category 3 storm at the time of landfall.