The Need for the Review and Understanding of SELDI/MALDI Data Prior to Analysis (Analyzer Beware)

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WINDOW INTO DISEASES Plasma Serum Urine Effusions Mucous Saliva Fecal Matter **Tissue (PAP; Urinary Sediment) Bile CSF Sweat**



Illustration of SELDI Time-Of-Flight (TOF) Mass Spectrometry. (Modified with permission from Ciphergen Biosystems, Inc.)



Contents of Duct Including Products of Dying Cells and Living Cells e.g., PSA



Products of Dying Cancer Cells Collect in Interstitial Space and are Absorbed into Vascular and Lymphatic Vessels



Venous and Iymphatic fluids containing tumor and tissue-reaction products and their metabolites

tissue

reaction

tumor 🕡

kidney

Urine with tumor and tissuereaction products and metabolites

liver

ONCOFETAL TUMOR ANTIGENS AND METABOLITES







JUNK ➡Statistician➡ JUNKBioinformatician

SELDI ANALYSIS **PROBLEMATIC ISSUES IN ANALYSIS Experimental Design** Patient Sample **Protein Chip Spectra Analytical Approach**

Experimental Design Selecting Cases and Controls Collecting and Processing Samples Performing Assays Without Bias Selecting Optimal Approach to Analysis **Avoiding Over-Analysis**



PATIENT

Groups Comparable Sites Racial/Ethnic Balance Homeostatic Balance No Bias



PATIENT Usually Comparing Disease vs. Control or Disease A vs. Condition B vs. Normal



PATIENT

Control Definition

Is Disease Absent? Is There Bias In The Controls?



Туре Collection Processing Storage Transfer

SAMPLE



SAMPLE

Type Serum versus Plasma Sensitivity- Can Products of Tumors Be Detected

LNCAP CELL LYSATES USING WCX2 ARRAYS







Processing

Time from Collection toFreezing (Too Restrictive?)For Consistent Results, RoboticProcessing Is Required



Processing

Removal of Proteins Present in Large Concentrations (e.g., Albumin) May Also Remove Peptides Being Carried by Removed Proteins



SAMPLE

Storage Length Temperature





Aliquot of Sample (#20) Stored at

Aliquot of Sample (#20) Transferred from -80C and Stored for 3 Months

Aliquot of Sample (#20) Stored at -20C for 5 Months

Aliquot of Sample (#20) Stored at -20C for 7 Months

Aliquot of Sample (#20) Stored at -20C for 8 Months

Second Aliquot of Sample (#20) Stored at -20C for 8

Second Aliquot of Sample (#20) Stored ONLY at -80C for Ten **Additional Months**



Transfer

Freeze-Thaw Cycles Quantity (Triplicates and Repeat-300 mcl)

Decrease in Protein Intensity in Human Serum due to repetitive Freeze-Thaws



Decrease in Protein Intensity in Human Serum (#6) due to repetitive Freeze-Thaws



Number of Freeze-Thaws

Figure1: The graph above shows the decrease in protein intensities in three different proteins in one human serum sample. The legend indicates the molecular weight of each protein. The specific intensity values are displayed above each of the bars.



TABLE 1:

	Old Designation	Current Chip	Biochemical Action of Surface Chemistry
	IMAC3	IMAC30 (with	Bivalent metals can be attached to the chip. Proteins
		hydrophobic	that bind to these divalent metals (e.g., Cu ⁺²) are bound
		barrier)	by the chip.
	WCX2	Same	This is a weak cation exchange chip. It contains
•		(CM10 mimics	negatively charged (anionic) carboxylate groups that
		WCX2 but does	will bind proteins with positively charged areas
		not replace it)	containing high numbers of lysine, arginine, and/or
			histidine amino acids.
	H4	Same (C16	The chip contains multiple chains of 16 methylene
		contains 16 CH ₃)	groups. This binds molecules that are hydrophobic.
	SAX2	Q10 (with	Strong anion exchanger which is composed of
		hydrophobic	quaternary ammonium groups that are charged
		barrier)	positively. This chip will bind proteins/peptides with
			regions rich in acidic groups, especially regions of
			peptides high in aspartic and/or glutamic amino acids.
	NP1 and NP2	NP20	General protein binding surface with binding of
			hydrophilic proteins.
	PS1 and PS2	PS10 / PS20	Chip designed to bind capture molecules of choices e.g.,
			antibodies, receptors, and nuclei acid binding proteins
			PS-1 (carbonyl diimidazole groups), PS-2 (epoxy
			groups). Also the PS-2 has a hydrophobic coating.
	SENDID		Incorporates EAM into chip.

QC Sample Spectra at Different Biochips B



Α

Protein Chip

 High Concentration Proteins May Block Binding of Low Concentration Proteins: 10,000 Ci of 5500 D Protein vs. 10 Ci of 7500 D Protein both with Same Binding Characteristics



All Proteins/Peptides Bound to Chip May Not Be Released/Ionized.

QC Sample Spectra at Multi-times Reading



Spectrum

5500

6000

 "Directed" and "Non-Directed" Approaches to Begin Spectral Analysis
Directed= Peak at 5500 Same As Peak at 5507 Based on Resolution +0.2%

5500

6000

Sectrum

Primary Peaks- Disease Has Unique or Larger Peak than Non-Disease; Thus, the Disease Produces a Molecular Product.

 Secondary Peak-Disease Causes a Decrease in Molecular Species Normally Present via Change in Metabolism or Excretion and/or Shutdown in Production

Spectrum

 Components of Spectra at Molecular Weights of Less Than 20,000 May Represent Metabolites of Proteins/Peptides Rather Than Intact Proteins/Peptides



Sectrum

Peaks May Not Provide Independent Information: For Example the Peak at 5500 D May Be A Metabolite of the Peak at 7500



Spectrum A High Concentration Protein May Prevent Identification of Low Concentration Protein: 1200 Ci of 5500 D Protein vs. 100 Ci of 5510 D Protein Even with Different Binding Characteristics to Same Chip



Spectrum All Areas of the Spectrum Are Not The Same Molecular Weights of Less Than 2000 No Standards; Noise; Contamination Weights of Greater Than 50,000 Proteins of High Concentration

SELDI ANALYSIS Spectrum

How Variable Is The Peak Location and/or Amplitude When the Same Sample Is Run On the Same Day on the Same Machine? On the Next Day on the Same Machine? On a Different Machine?



 Eastern Virginia Medical School; UAB; U of Texas San Antonio; U of Pittsburgh Medical Center; Johns Hopkins Medical Center; Uniformed Health Services

All Were Able To Standardize Their Machines and To Obtain Comparable Data on 14 Cancer and 14 Non-Cancer Cases

