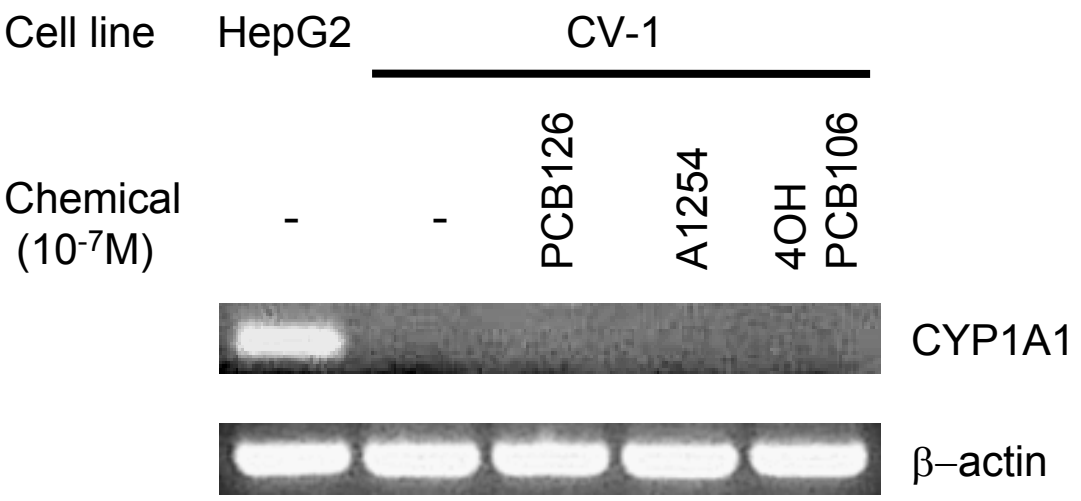
Materials and Methods

Semi-quantitative RT-PCR We used the following primers for RT-PCR: CYP1A1-sense, 5'-ACCTGAACGGTTTTATCACCCC-3'; CYP1A1-antisense, 5'-ATGGTCTCACCGATGCACTTC-3'; β-actin-sense, 5'-GGCGGCACCACCATGTACCCT-3'; β-actin-antisense, 5'-AGGGGCCGGACTCGTCATACT-3'. Total RNA was obtained from HepG2 cells and CV-1 cells using SV Total Isolation System (Promega, Madison, WI) according to the manufacturer's instructions. We obtained the first-strand cDNA using QuantiTect Reverse Transcription Kits according to the manufacturer's protocol (Qiagen, Valencia, CA) and then PCR proceeded using the cDNA as a template and Platinum PCR SuperMix (Invitrogen, Carlsbad, CA). Expressed bands were confirmed by sequencing and the intensity of ethidium bromide-stained bands was analyzed using a Kodak Digital Science EDAS 290 system (Kodak, Norwalk, CT). The relationship between the inverse of band intensity and the number of PCR cycles was linear. The number of PCR cycles determined from the plot was 45 for both CYP1A1 and β-actin.

Figure legend

Supplementary Figure 1S. PCBs did not induce CYP1A1 expression in CV-1 cells.

CV-1 cells were incubated with or without 10^{-7} M PCB126, A1254 or 4OH-PCB106 for 24 h. First-strand cDNA was obtained from total RNAs using reverse transcriptase and oligo-dT primer. Semi-quantitative RT-PCR proceeded using primers indicated in Materials and Methods. Bands were obtained from the increasing phase of PCR products and intensity was analyzed on a Kodak Digital Science EDAS 290 system. Data are representative of experiments that were repeated three times.



Supple. Figure 1S