





Gag/Pol myr - MA p12 CA NC PR RT IN

Retrovirus	Receptor	Туре	Function
HIV, SIV	CD4 and CXCR4, CCR5, others	TM1, TM7	Immune recognition, G protein-coupled chemokine receptors
E-MLV	CAT-1 (SLC7A1)	TM14	Basic amino acid transport
GALV, 10A1, MLV, FeLV-B, woolly monkey virus	Pit1 (SLC20A1)	TM10-13	Phosphate transport
A-MLV, 10A1, MLV, FeLV-B	Pit2 (SLC20A2)	TM1013	Phosphate transport
BLV	Blvr	TM1	?
ASLV-A	Tva	TM1	LDL receptor-like protein
ASLV-B, ASLV-D, ASLV-E	Tvb	TM1	Fas/NFR-like receptor
MMTV	Mtvr	TM1	?
RD-114, type D SRV, BaEV, HERV-W	RDR (SLC1A5) or RDR2 (SLC1A4)	TM9-10 TM9-10	Neutral amino acid transport Glutamate, neutral amino acid transport
Xenotropic and polytropic MLVs	XPR1	TM8	G protein-cuopled signaling?, transport?
FeLV-C	Flvcr	TM12	Organic anion transporter?
FeLV-T FeLIX and Pit1	(SLC20A1)	Soluble TM10-13	Env-like protein Phosphate transport
JSRV	HYAL2	GPI anchored	Hyaluronidase (weak)





The Vector:







The problem:



Solutions: Split genomes



OR



HIV - an upgrade in retroviral vectors



HIV - vectors production



HIV genome



Helper constructs







RRE Rev Responsive element cPPT central polypurine tract

EF-1 α elongation factor 1α promoter

PRE post-transcriptional regulatory element

SD/SA Splice donor and acceptor sequences

Prom Transcriptional promoter/enhancer element



From: Cockrell et al., Molecular Therapy (2006)



Growth and Amplification

JOURNAL OF VIRCLOGY, Nov. 2000, p. 9868–9877 0022-538X/00/\$04.00+0 Copyright © 2000, American Society for Microbiology. All Rights Reserved. Vol. 74, No. 21

Multiple Blocks to Human Immunodeficiency Virus Type 1 Replication in Rodent Cells

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Received 12 June 2000/Accepted 6 August 2000

Lentiviral vectors are derived from: Primates HIV-1, HIV-2 and SIV Non-primates FIV, EIAV and BIV **Recommendations:**

BL2 for manipulation of generation 3 vectors

ABL1 for housing animals injected with vector

Exceptions:

Animals with human cells that may allow replication of HIV-1 RCL (though non has been detected)

At the discretion of the IBC.

Discussion Questions

The aim of the discussion is to develop some general guidance to assist Institutional Biosafety Committees in conducting risk assessments and determination of appropriate containment for lentiviral vector research.

 What factors should be considered in a biosafety assessment of lentiviral vector research with regard to

- The parental virus the vector is derived from (e.g., HIV-1, FIV, EIAV, etc.)
- Safety modifications to the vector system (e.g., deletion of viral genes, sequence overlap creating potential for recombination, SIN vector, etc.)
- The system used to generate vector (e.g., 2, 3, or 4 plasmid transient transfections, stable packaging cell lines, etc.)
- Pseudotyping (e.g., VSV-G, etc.)

 What factors should be considered in determining containment for different types of manipulations of lentiviral vectors such as

- Vector generation
- Tissue culture
- Small and large animal work
 - Vector administration
 - Housing
- Animals permissive for viral replication
- Large scale production

Discussion Questions

• What consideration should be given to the type of transgene expressed from the vector?

- When should testing for replication competent lentivirus (RCL) be considered?
- What RCL assays may be useful?

• For research in which enhanced BL2 containment is considered appropriate, what types of enhancements to practices would be most useful?