

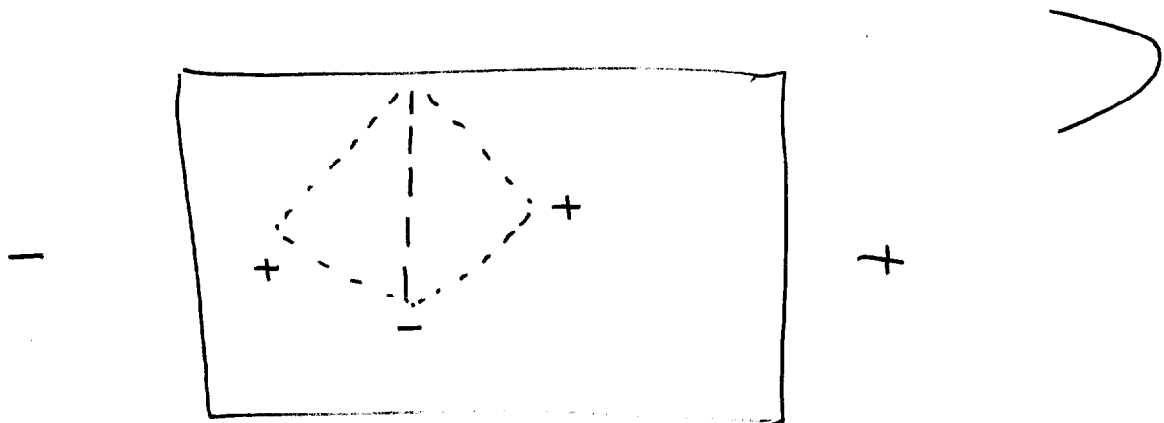
<< 1945

Research Ideas.

1. Implant an egg or embryos of frog in abdomen or onto omentum. Determine influence of 'old' environment on growth of embryonic material, and vice versa. 1. Look for rates of development of the embryos, and possible development of teratomata.
2. Tissue culture of egg cell: in Arbacia, using Ca free seawater if necessary to induce separation of blastomeres, attempt to induce indefinite proliferation of the egg cell by using various media. This should be without differentiation or organization.
3. Culture of grasshopper testes--- what induces the spermatogonium to start its cycle of spermatocyte and differentiation into spermatid. Can sperm-formation be induced in vitro? Egg?? Serious practical and bio-philosophical consequences: the perpetuation of the germ without the Soma.

The influence of respiratory inhibitors on the mitotic figure. Look for a material in which the mitotic function is pronounced, and which is amenable to respiratory and chemical analysis. Even the old one of cyanide on onion root tip would be interesting. Work out a case here even now??? Why bother to introduce the complication of colchicine.

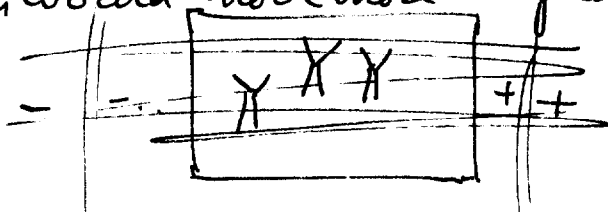
Repeat the electrodynamic experiments of McClendon etc, and determine whether the roots really remain alive. One presumptive test might be the assumption of the c-figure on subsequent treatment with colchicine. Reversibility is the important factor. If there is a fairly simple electrical polarization in the root tip it should be demonstrated as follows



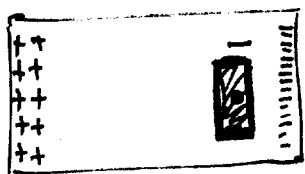
and there is a rigid framework.

If charges balance, there should be no movement of the figure as a whole. But catch the chromosomes themselves - how?

Used colicinized cells. v.g. The presence of smaller mobile ions which would move more rapidly would polarize and neutralize the imposed field.



What does the anodic movement of chromosomes mean? That they bear a net charge.



A significant

a considerable permeability

movement also implies of the cell walls to <sup>all</sup> the small, mobile ions in the cells. The electrical apparatus should not be difficult to construct. Exact impedance measurements would be difficult. A high capacitance term implies a low permeability to ions. Still the exps. have to be repeated.

# The Mechanism of the Induction by Radiation of Chromosome Aberrations in *Tradescantia*.

DE Lea and DG Calkins, J. Gen. 44: 216-245 (1943)

1. Thesis (Daf). "The primary breaks formed by the radiation is considerably in excess of the total number observed as aberrations of all kinds, and that the majority recombine in the original formation after existing free for a limited period during which interchanges are possible."

a. Assume that radiation is administered in a short dose at high intensity. Then simultaneous production of breaks. Let  $n_0$  = number of initial breaks, and  $n$  those that are free. If the number of reformations is large, the computation of  $n$  from these is accurate. Then  $n = n_0 \cdot f(t)$  where  $f(t)$  does not depend on  $n_0$  since reunion is uninfluenced (relatively) by other breaks.

The rate of interchange formation  $\frac{dI}{dt} = \beta \cdot n^2$ . As in a bimolecular reaction.

$$\therefore I_n = \int \beta n^2 dt = \beta n_0^2 \int f(t)^2 dt \quad (1)$$

Over the entire period,  $I_n = \beta n_0^2$ ; since  $n_0 \propto$  dose,  $I \propto$  dose<sup>2</sup>

We assume that the average time  $\tau$  elapses between breaks and reformations. Considering the rate of reunion as a first order reaction. (which is ill understood anyhow. Assume, however, that  $\frac{n}{n_0} = f(t) = e^{-t/\tau}$ . If the dose is given at  $I$  r per min. the rate of formation of primary breaks is  $kI$

$$\text{then } \frac{dn}{dt} = kI - \frac{n}{\tau}$$

$$n = kI(1 - e^{-t/\tau})$$

$$n = kI(1 - e^{-t/\tau}) e^{-\frac{(t-T)}{\tau}}$$

$$\text{If } I = 0 \text{ at } t = T$$

then in (1)

$$\int \beta n^2 dt = \frac{1}{2} k^2 \beta \tau (IT)^2 G$$

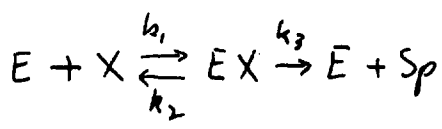
$$G = 2 \left(\frac{\tau}{T}\right)^2 \left[T/\tau - 1 + e^{-T/\tau}\right]$$

for a dose  $IT = D$ ,  $I_n = \text{dose}^2 G \left(\frac{\tau}{T}\right)$

By fitting various data to these curves, the values for  $\tau$  of 3.3 min. (Cambridge) and 4 min (Harvard) are obtained.

Consider:

X = substance combining with  
E = enzyme.



and Sp = products of dissociation, yielding E. If  $k_3 \ll k_2$ , X is a reversible inhibitor.

Consider cases where  $k_3 \rightarrow 0$ .

Then let X = I for inhibitor and X = S for substrate. If v is velocity of substrate breakdown  $v = k_3(ES)$ . If S is in large excess,  $ES \approx E$  and  $v_{max} = k_3 E$ .

In the presence of I,  $v = k_3(E - EI)$ .  $\frac{v}{v_{max}} = 1 - \frac{EI}{E}$  if  $i = \frac{EI}{E}$ ,

$i = \frac{v_{max} - v}{v_{max}}$  = percentage inhibition. If  $k_3 = 0$ ,  $E + I \xrightleftharpoons[k_2]{k_1} EI \xrightleftharpoons[k_1]{k_2} E + I$   $K = \frac{(E - EI)(1 - EI)}{EI}$

$EI = iE$ ,  $I = \frac{K i}{1 - i} + iE$  <sup>3B</sup> since  $iE$  is combined inhibitor,  $\frac{K i}{1 - i} =$  free inhibitor.

For simplicity, express (E) as  $\frac{E}{K} = E' \neq I/K = I'$

$I' = \frac{i}{1 - i} + iE'$  For  $E'$  small,  $I' \approx \frac{i}{1 - i}$  <sup>3A</sup> and inhibition is a function

of (I) only. For  $E'$  large,  $i = \frac{I'}{E'}$  <sup>3C</sup> ... These are defined as zones of enzyme behavior.

Curves are given showing the ranges where, for given fractional error, eq. 3A or 3C may be used within zone C, inhibition has no effect, in Zone A only amount of inhibition. The inhibition effect is investigated in a simple algebraic manner. There is apparently a close confirmation in the exp. results

$I' - iI' = i$   $i + iI' = I'$   $i = \frac{I'}{1 + I'}$  so this approximation is valid for large  $i$  yes. i + I'

1. In *A. cepa*, *P. sativum*, *V. faba*, after irradiation there is a peak about 3 hours after irradiation, with a gradual return to normal. - <sup>recovery</sup> <sup>(how desirable?)</sup> <sup>factorial analysis?</sup>
  2. The % normal anaphases are single exponential functions of the x-ray dose. The slopes are identical for 120, 180 and 400  $\times 10^3$  volts.  $\therefore$  The ion-pair rather than the quantum is significant. I.E. it is not the energy of the quantum, or its products that is significant, but the number.  
[Compare response to fluorescence? This is essentially all that is stated]
  3. Curves for the inhibition of mitosis as a function of time after irradiation parallel those for chromosome abnormalities. "Mitosis is inhibited in or at the end of the resting stage since division stages disappear and there is no immediate killing of cells. Therefore cells in anaphase at the peak of inhibition and abnormality curves were in early prophase at irradiation."
  4. Inhibition of mitosis does not depend on chromosome length. (Source?)
  5. Slope of ~~inhibition~~ "survival" curve varies with total metaphase chromosome length. JH 27:459 (1936)  
As *Vicia & Pisum*, ratio of slopes is 2.5, of lengths 2.3 <sup>with curve</sup>
  6.  $\therefore$  Cross-section area of sensitive volume is same.
  7. Sensitive volume is of order of size of  $\times \times 10^{-21}$  cm. and diam:  $10 \times 10^{-8}$
- See JGP 19: 179-198 (1935)

1. Hit-Theory, or Target Hypothesis:

a gene-mutation is produced by ionizing radiation whenever some primary phenomenon, a hit, occurs within a certain region of space, the target.

2. Ionizations normally distributed are considered the primary phys. phen. Assuming random distribution of ionizations, the probability of producing an ionization within the target, and therefore the frequency of mutations is proportional to total ionizations. Therefore the number  $N$  of mutations in  $N_0$  *Drosophila* specimens is; by the "dosage"  $D$ .

$$N = k N_0 D$$

b. If  $D$  is measured as the ionization produced per unit volume,  $N$  should be independent of  $V$  and of time distribution.

3. Freq. of ionizations within target is given by  $V_{\text{target}} \cdot D$  where  $D \propto \frac{\text{ionizations}}{\text{unit volume}}$ . From experimentally determined  $k$ , therefore, target size can be estimated, and has been found to be on a single mutation, as  $1 \mu\text{m}^3 \approx 1000$  atoms.

4. Where distance of successive ionizations is less than the target distance two ionizations occur in the same target, but yield only one mutation. This is called saturation effect. In a few cases this has been found (or claimed!) with neutrons, a saturation effect for sex-linked lethals is found.

The total volume of targets can be found from 3. and from 4. the size. The total number is thus determinable. For sex, this was ca 1860 sperms.

If each gene is capable of mutation, this represents number of genes.

The targets thus found are of same order of magnitude as those for individual visible mutations. Lea's estimate is ratio of frequency of all sex-linked lethals to that of individual mutations.

"The broadest principle of hit theory is that

$N = k N_0 D$  is determined inherently by the biophysical process.

That this has meaning requires: (1) all or none primary phenomenon. (2) genetic material of different sperms is identical (3) the law is valid down to low dosage and long time.  $k$  then is the probability that unit dose mutates a particular sperm.  $k$  is manifestly the effective cross-section or volume. Let unit be  $5 \times 10^{-13}$  cc. = 1 ionization/cc.

The hit theory assumes that  $k$  is the actual volume. It probably may represent also a spatial and a physical probability factor.

Latter is quantum effectivity.

Bibliography

- See J Gen 39:181-182 (1940)
- Muller J Gen 40:1-66 (1940)
- Fano CSH IX (1941)
- Fuiles + Demere PNAS 23:320-7 (1937)

Thers  $\frac{1}{\tau} = \rho V_{rms}$ .

The sat. effect may lead to values of  $\frac{1}{2}$

### Nature of hit!

- Absorption or scattering of a quantum and liberation of e
- Passage of sec. electron across target
- Production of ionization by a sec. electron in a target.

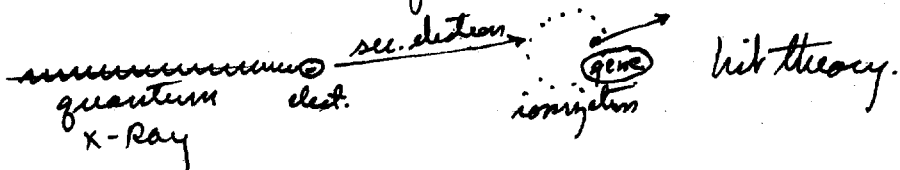
In case (a) the unit of D. is 1 sec. electron/cc

(b) flow of 1 secondary e. / sq. cm

(c) prod. of 1 ionization/cc. = submultiple of r.

Only (c) is independent of the wavelength, as found in biol. exp. and in chemical exp. It can be considered as a working model, but does not necessarily represent the actual hit.

As  $N = N_0 k D$ , k is the efficiency of a unit of radiation. "The biological action depends only on the total number of ions produced by all secondary electrons."



Each ionization is an independent cause. Here "excitation" must be considered as well as ionization. Also may also distinguish the effects of tertiary electrons.

Concentration effect: cooperative effect when single ionizations are ineffective

Saturation effect: density of ionization tracks. When ion density is small, the effectiveness of particle flow or energy delivered normally per unit path. The slow onset of saturation may be ascribed to the heterogeneity of the particles produced.



# Bacterial genetics

date: \_\_\_\_\_  
pres 45 of 6

1. Segregation after X-irradiation. Compare mutant frequency  $\bar{c}$  plate counts as a function of time. [Correlate  $\bar{c}$  cytological studies, particularly  $\bar{c}$  treatments such as colchicine, acenaphthene, etc. ?? Hyp. Vegetative bacterial cells are binucleate; haploid segregating by cell division at each fission. Problem: effective duplicity in the nuclei at time of irradiation. Problem: selection during growth.
2. Statistical correlation in reverse mutation.
3. Sexual recombination genetically studied.
4. Induction of mutations by radioactive labelled atoms in various compounds.
5. Localization of the radio-isotope: Inverse material containing relatively high activity of radioisotope in a fluorescent solution (e.g. eosin), and watch for scintillation. Test  $\bar{c}$  a  $PO_4$  ppt. (Look up radiographic technique.) Also, in considering mutations, isolate those cells which have had a discharge. Study lethality of discharge by direct observation.

1/20/46

~~The Wilder~~  
Lab.Plasmagenic Inheritance

The plasmagene concept refers to the production by nuclear genes of independently self-reproducing units which are topographically and physiologically independent of the nuclei from which they are derived. For the study of plasmagenes, it is necessary to find situations in which (1) nuclei of particular genotype can be separated from cytoplasm that has been under their influence and (2) the plasmagenes are in some stage of the organism's life cycle eliminated. These conditions have been met with in two conditions: in the maturation of male gametes of <sup>higher</sup> plants and animals, cytoplasmic elements are largely eliminated, while they are retained in the maturation of the egg; the comparison of the progeny of reciprocal crosses allows this study. In another instance, reported by Lindgren + Spiegelman, plasmagenes have been studied in yeasts. Here, genes are separated from plasmagenes by hybridization, and isolation of segregants. The plasmagene is eliminated by virtue of its instability in the absence of substrate (~~glucose~~ melibiose). These investigators have shown that a plasmagene for the production of melibiose-glycosylase is initiated by a nuclear gene, but is perpetuated in its absence so long as melibiose is provided. The plasmagene is, however, unstable in the absence of melibiose, and disappears. Its reinitiation depends on the presence of the nuclear gene.

The formation of heterocaryons in *Neurospora* provides material for the study of plasmagenesis. The diffusion of nuclear products across

nuclear membranes is well shown by the growth on minimal media of heterocaryons between different ~~the~~ biochemical mutants. It is not now known what these products are: plasmagens, enzymes, or enzyme products. ~~While~~ it is possible to segregate nuclei by isolation of (presumably) uninucleate microconidia; however, the failure by this procedure to obtain evidence favorable to the plasmagen hypothesis can readily be interpreted on the basis of an elimination of plasmagens during sporulation (analogous to sperm maturation).

It sometimes occurs, however, as a statistical accident, that all of the nuclei at the ~~tip~~<sup>hyphal</sup> tip of a heterocaryotic mycelium are of a single type. The fact that this hypha has grown on minimal medium demonstrates that products from the heterologous <sup>have mixed</sup> nuclei. The isolation of such hyphal tips to minimal medium allows conclusions to be drawn as to the nature of the diffusible gene products. [A <sup>conocytic</sup> uninucleate septate organism might be preferable! - See Phycomycus.] If these products are the diffusible end-metabolites (vitamins or amino acids), the ~~the~~ growth of this isolated hypha on minimal medium should be minimal; if a stable enzyme is found there should be linear (not exponential) growth of the hypha, until the enzyme is too diluted to be active, or disintegrates. If plasmagens occur, however, there should be considerable growth.

The only methods now available for determining the homo- or heterocaryotic condition of a hypha ~~is~~ are genetic. Since no genes affecting nuclear characteristics are available, the genetic constitution of the hypha must be determined by labels of old characters.

It is essential, for obvious reasons, <sup>to</sup> ~~to~~ exclude the possibility of plasmagene transmission of the labels. This can be accomplished by passing through a sexual generation, ~~which~~ for characters known to be inherited in Mendelian fashion. Thus if a hypha from an arginine + lysineless heterocaryon is crossed  $\bar{c}$  arginineless of the opposite sex, the absence of growth on minimal from a mass inoculation of ascospores is evidence against the presence of any lysineless <sup>has 50% nuclei either 4/13 arg, or 1/13 arg, or 1/13 arg</sup> nuclei in a parental nucleus. Crossing over of lysineless  $\bar{c}$  arginineless will ~~not~~ yield a wild type nucleus, which can grow on minimal. Failing this, a lysineless nucleus can form a prototrophic heterocaryon  $\bar{c}$  arginineless. A reverse argument applies to lysineless.

This would, however, be a laborious procedure <sup>if each</sup> ~~for the heterocaryon~~ hyphal isolate which grew on minimal had to be so isolated. From Beadle and Tomada's data it is clear that most such hyphal isolates are heterocaryotic. Fortunately, it is possible to ~~then use~~ more facile methods. An independent <sup>sufficient</sup> evidence it is probable that plasmagene are not produced at the albino loci 4637 and 15300. This is indicated by the disappearance of color from heterocaryons between leucineless - 15300 and 4637 heterocaryons when either nucleus is selected for. If the recessive nuclei of a heterocaryon are labelled  $\bar{c}$  these color genes, heterocaryosis is indicated by the formation of colored ~~conidia~~ <sup>conidia</sup>; heterocaryosis by albino conidia. If this is achieved, the ~~conidia~~ conidia should be tested for prototrophesis. If ~~the~~ the conidia

grows minimal, the possibility of genetic modification must be considered. If the conidia do not germinate on minimal, there are either 1) no heterocaryotic nuclei 2) an unusual selective phenomenon preventing the manifestation of them. This can be investigated independently. Another heterocaryon possibility is colored wild type + the albino ~~was~~ - histidine deficient.

Experimental procedure (A). X-4637A. + Y-15300A.

1. Prepare heterocaryon on a minimal slant. Conidia should be colored.
2. Inoculate these conidia on an agar plate (minimal).
3. Isolate hyphal tip and transfer to minimal. (only a few).
4. If conidia are colored, transfer to minimal plates.
5. Isolate numerous hyphal tips and transfer to minimal medium
  - (1) to growth tubes. Atypical rates interesting.
  - (2) slants. Look for white conidia.
6. Test conidia on minimal medium. If they do not grow, test on X, Y, complete.
7. If they grow, cross with
 

(1) X-	X	Y	XY	
(2) Y-	1. +	-	-	Y plasmagones
(3) X-Y.	2. -	+	-	X plasmagones
8. Test progeny in mass
 

	3. +	+	+	<u>still a heterocaryon</u>
	4. +	+	+	<u>transmuted</u>
9. If "still a heterocaryon" or "transmuted", spores must be ~~not~~ isolated from XY cross.

2/13/45.

This is plenty 1946. Cf. 482

# 1. The detection of nutritional mutants of microorganisms.

The <sup>now</sup> classical method for the detection of nutritional mutants consists in (1) isolating pure lines and (2) testing these individually for their nutritional characteristics. Since in *Neurospora*<sup>(1)</sup> and bacteria<sup>(2)</sup> only a small proportion of the cells are mutants, it is evident that considerable labor must be done in (1) for the collection of a few strains upon which (2) will be successful.

A method has been devised for the detection of some such mutants. The method depends on the fact that the growth of a mutant strain is limited by the concentration of its unique nutrient in the medium. Thus, by supplying distinctly suboptimal (liminal) amounts of the nutrient in an agar pour, plating the mixture of prototrophic and mutant cells into such a plate. The mutant colonies are often detectable as they are much smaller than the wild type colonies in such a plate. They can then be picked individually, transferred to an optimal medium, and its requirements determined. more facile.

The critical problem to be overcome in the application of this method is the elaboration of a satisfactory liminal medium. This has been accomplished on the basis of the simple, if only roughly accurate, assumption, that the needs of an organism are in proportion to their composition. Thus, whole bacteria have been hydrolyzed, and added in such quantity that a single mutant (methionineless) can readily be distinguished from the wild type. It was found that this quantity was adequate for the detection of other, already available, mutants, so that we may induce

that it will be satisfactory for other mutants. It has not yet been applied to X-Rayed material.

The advantages of this method for quantitative studies, over the old one of picking and testing at random are obvious. Several studies detailed below depend on this method.

A second method may be of more general application, but has not been studied, and is suggested purely on an a priori basis. The mixture of mutants + ~~old~~ prototrophs is washed and plated into a minimal agar medium. Only the prototrophs will develop colonies. After 24-36 hours, giving these a uniform "head start". At this time, an optimal (or specific) nutrient supplement is added. As this diffuses through the agar, new colonies will appear, which can be ~~detected~~ detected at the appropriate time by their size and recent appearance. When they are sufficiently grown, they can be picked and tested. Since this method does not require a carefully titrated medium, it should be particularly valuable in the detection of mutants requiring particular substances.

These methods should be adaptable to any organism which can be "plated out", in particular bacteria and yeasts. Slides should be more refractory.

## Analysis of the growth of mutant bacteria

2. The response of a mutant strain to its specific nutrient affords unexampled material for the study of growth, insofar as the "master reaction" controlling its rate is identified. E.g. the utilization of leucine. The analysis of the growth rate - nutrient response curves can therefore be accomplished on a rational basis. The quantitative extent of the intermediary absorption of the nutrient can be studied, as well as the efficiency of its utilization. Temperature and pH responses also lend themselves to study, both in *Neurospora* and bacteria. Sufficient has been done  $\bar{c}$  leucineless *Neurospora* to warrant selection of this organism for more detailed study. [One may also inquire why different mutants vary in their requirements for the same substance.] The lower limit should be the composition of the organism.

This problem abuts into that of the composition of the organism.

A corollary to this study is the development of ordinary microbiologic assay methods. Of more novel interest is the possibility of the use of such organisms, under certain conditions, for the quantitative removal of specific substrates,  $\bar{c}$  applicability in the isotopic analysis of small samples.

Effect of a "final" product on the intermediary synthesis.



5. Genetics of Bacteria. I. Segregation of Radiation-induced Mutations.

There is cytological evidence that the vegetative phase of bacterial cells consists of a bicaupon. That is, a single, (haploid) nucleoid gives rise to two nuclei in each bacterial cell; when the cell divides, a monocupon is obtained, which by nuclear division gives rise to the next generation's bicaupon. An implication of this is that the expression of recessive mutations will be ~~not~~ delayed until one cell division has occurred in order that the mutated nucleus segregate from the prototrophic one. This can be studied by comparing the ~~effects of~~ mutant proportions <sup>in</sup> plating at varying times after irradiation, and correlating  $\bar{c}$  increase in cell number, and cytological changes. Groups of individual colonies can be replated also to show the later appearance of the mutants.

As further extensions, attempts may be made to induce cytological changes  $\bar{c}$  drugs such as acetylthiourea, gelicine, etc., and determine changes, if any, in response to X Ray. Other organisms may be studied by this genetic method.

Ryan, meanwhile, is working on back mutations in multiple mutants (mutual influence of mutations, etc.) studies such as influence of temperature, mutating agents (ethyl isothiocyanate) are best here.