

## 8. REFERENCES

- \*Abbas MN, El-Assy NB, Abdel-Maniem SH. 1989. Determination of traces of mercury(II) and phenylmercury by direct polyurethane foam thin-layer spectrophotometry. *Anal Lett* 22(11-12):2575-2585.
- \*Aberg B, Ekman L, Falk R, et al. 1969. Metabolism of methyl mercury ( $^{203}\text{Hg}$ ) compounds in man. *Arch Environ Health* 19:478-484.
- \*ACGIH. 1996. Threshold limit values for chemical substances and physical agents and biological exposure indices for 1996. American Conference of Governmental Industrial Hygienists, Cincinnati, OH.
- \*ADA. 1991. American Dental Association (ADA) News. April 8, 1991, 3.
- \*ADA. 1997. ADA Position Statements. American Dental Association.  
<http://www.ada.org/prac/position/canada.html>.
- \*Adams C, Ziegler D, Lin J. 1983. Mercury intoxication simulating amyotrophic lateral sclerosis. *JAMA* 250:642-643.
- \*Adinolfi M. 1985. The Development of the Human Blood-CSF-Brain Barrier. *Developmental Medicine & Child Neurology* 27:532-537.
- \*Afonso J, deAlvarez R. 1960. Effects of mercury on human gestation. *Am J Obstet Gynecol* 80:145-154.
- \*Agnér E, Jans H. 1978. Mercury poisoning and nephrotic syndrome in two young siblings. *Lancet*:951.
- \*Agrawal R, Chansouria JPN. 1989. Chronic effects of mercuric chloride ingestion on rat adrenocortical function. *Bull Environ Contam Toxicol* 43(3):481-484.
- \*Agrawal YK, Desai TA. 1985. Liquid-liquid extraction, photometric, and atomic absorption spectrophotometric determination of mercury. *Anal Lett* 18:2521-2536.
- \*Aguilar A, Borrell A. 1995. Pollution and Harbour porpoises in the Eastern North Atlantic: A review. *Rep Int Whal Commn* 16:231-242.
- \*Ahmad S, Qureshi IH. 1989. Fast mercury removal from industrial effluent. *J Radioanal Nuclear Chem* 130(2):347-352.
- \*Ahmed R, Duerbeck HW, Stoeppler M, et al. 1988. Gas liquid chromatographic (GLC) analysis of methylmercury in fish and its comparison with total mercury. *Pak J Sci Ind Res* 31(8):535-540.
- Airey D. 1983a. Mercury in human hair due to environment and diet: a review. *Environ Health Perspect* 52:303-316.

---

\*Cited in text

## 8. REFERENCES

- \*Airey D. 1983b. Total mercury concentration in human hair from 13 countries in relation to fish consumption and location. *Sci Total Environ* 31:157-180.
- \*Ajmal M, Mohammad A, Fatima N, et al. 1989. Determination of micro quantities of mercury(II) with preliminary thin-layer chromatographic separation from mercury(I), lead(II), nickel(II), and copper(II) on acid-treated silica gel layers: Recovery of mercury(II) from river waters and industrial waste waters. *Microchem* 39:361-371.
- \*Akagi H, Malm O, Branches F JP, et al. 1995. Human exposure to mercury due to gold mining in the Tapajos river basin, Amazon, Brazil: Speciation of mercury in human hair, blood and urine. *Water Air and Soil Pollution* 80(1-4):85-94.
- \*Akesson I, Schutz A, Attewell R, et al. 1991. Status of mercury and selenium in dental personnel: Impact of amalgam work and own fillings. *Arch Environ Health* 46(2):102-109
- \*Al-Mufti AW, Copplestone JF, Kazanitzis G, et al. 1976. Epidemiology of organomercury poisoning in Iraq: I. Incidence in a defined area and relationship to the eating of contaminated bread. *Bull World Health Organ* 53(suppl):23-36.
- \*Al-Saleem T, Clinical Committee on Mercury Poisoning. 1976. Levels of mercury and pathologic changes in patients with organomercury poisoning. *Bull World Health Organ* 53(suppl):99-104.
- \*Al-Saleh I, Al-Doush I. 1997. Mercury content in skin-lightening creams and potential hazards to the health of Saudi women. *J Toxicol Environ Health* 51(2):123-30.
- \*Al-Shahristani H, Shihab K. 1974. Variation of biological half-time of methylmercury in man. *Arch Environ Health* 18:342-352.
- \*Al-Shahristani J, Shihab KM, Al-Haddad JK. 1976. Mercury in hair as an indicator of total body burden. *Bull World Health Organ (Suppl)* 53:105-112.
- \*Albers J, Cavender G, Levine S, et al. 1982. Asymptomatic sensorimotor polyneuropathy in workers exposed to elemental mercury. *Neurology* 32:1168-1174.
- \*Albers JW, Kallenbach LR, Fine LJ, et al. 1988. Neurological abnormalities associated with remote occupational elemental mercury exposure. *Ann Neurol* 24(5):651-659.
- \*Albrecht J, Szumanska G, Gadamski R, et al. 1994. Changes of activity and ultrastructural localization of alkaline phosphatase in cerebral cortical microvessels of rat after single intraperitoneal administration of mercuric chloride. *Neurotoxicology* 15(4):897-902.
- \*Alcser KH, Birx KA, Fine LJ. 1989. Occupational mercury exposure and male reproductive health. *Am J Ind Med* 15(5):517-529.
- \*Alexidis AN, Rekkas EA, Kourounakis PN. 1994. Influence of mercury and cadmium intoxication on hepatic microsomal CYP2E and subfamilies. *Res Commun Mol Pathol Pharmacol* 85(1):67-72
- \*Ali SF, LeBel CP, Bondy SC. 1992. Reactive oxygen species formation as a biomarker of methylmercury and trimethyltin neurotoxicity. *Neurotoxicology* 13(3):637-648.

## 8. REFERENCES

- \*Allard B, Arsenie I. 1991. Abiotic reduction of mercury by humic substances in aquatic system - an important process for the mercury cycle. *Water Air Soil Pollut* 56:457-464.
- \*Allard M, Stokes PM. 1989. Mercury in crayfish species from thirteen Ontario lakes, Canada in relation to water chemistry and smallmouth bass (*Micropterus dolomieu*) mercury. *Can J Fish Aquat Sci* 46(6):1040-1046.
- \*Altman PK, Dittmer DS. 1974. In: *Biological Handbooks: Biology Data Book, Volume III*, second edition. Bethesda, MD: Federation of American Societies for Experimental Biology, 1987-2008, 2041.
- \*Altman PL, Dittmer DS, ed. 1972. *Biology data book, 2nd ed.*, Bethesda, MD: Federation of American Societies for Experimental Biology, 199-201.
- \*Altmann L, Sveinsson K, Kraemer U, et al. 1998. Visual functions in 6-year-old children in relation to lead and mercury levels. *Neurotoxicol Teratol* 20(1)9-17.
- \*Amin-Zaki L, Elhassani S, Majeed MA, et al. 1974. Intra-uterine methyl mercury poisoning in Iraq. *Pediatrics* 84:587-595.
- \*Amin-Zaki L, Elhassani S, Majeed MA, et al. 1976. Perinatal methylmercury poisoning in Iraq. *Am J Dis Child* 130:1070-1076.
- \*Andersen ME, Krishnan K. 1994. Relating *in vitro* to *in vivo* exposures with physiologically-based tissue dosimetry and tissue response models. In: H. Salem, ed. *Current concepts and approaches on animal test alternatives*. U.S. Army Chemical Research Development and Engineering Center, Aberdeen Proving Ground, Maryland.
- \*Andersen ME, Krishnan K. 1994. Relating *in vitro* to *in vivo* exposures with physiologically-based tissue dosimetry and tissue response models. In: H. Salem, ed. *Animal test alternatives*. U.S. Army Chemical Research Development and Engineering Center, Aberdeen Proving Ground, Maryland.
- \*Andersen ME, MacNaughton MG, Clewell HJ, et al. 1987. Adjusting exposure limits for long and short exposure periods using a physiological pharmacokinetic model. *Am Ind Hyg Assoc J* 48 (4): 335-343.
- \*Andersson A. 1979. Mercury in soils. In: Nriagu JO, ed. *The biogeochemistry of mercury in the environment*. New York, NY: Elsevier/North Holland Biomedical Press, 79-112.
- \*Andersson I, Parkman H, Jernelov A. 1990. The role of sediments as sink or source for environmental contaminants: A case study of mercury and chlorinated organic compounds. *Limnologica* 20(2):347-360.
- \*Andren AW, Nriagu JO. 1979. The global cycle of mercury. In: Nriagu JO, ed. *The biogeochemistry of mercury in the environment*. New York, NY: Elsevier/North Holland Biomedical Press, 1-22.
- \*Andres P. 1984. Brief communications: IgA-IgG disease in the intestine of Brown-Norway rats ingesting mercuric chloride. *Clin Immunol Immunopathol* 30:488-494.
- \*Anneroth G, Ericson T, Johansson I, et al. 1992. Comprehensive medical examination of a group of patients with alleged adverse effects from dental amalgams. *Acta Odontol Scand* 50(2):101-111.

## 8. REFERENCES

- \*Anttila A Sallmen M. 1995. Effects of parental occupational exposure to lead and other metals on spontaneous abortion. [review]. *J Occup Environ Med* 37(8):915-21.
- \*Anwar WA, Gabal MS. 1991. Cytogenetic study in workers occupationally exposed to mercury fulminate. *Mutagenesis* 6(3):189-192.
- \*AOAC. 1984. Official methods of analysis of the Association of Official Analytical Chemists. Arlington, VA: J Assoc Off Anal Chem.
- Aposhian HV, Bruce DC, Alter W, et al. 1992a. Urinary mercury after administration of 2,3-dimercaptopropane-1-sulfonic acid: Correlation with dental amalgam score. *Fed Am Soc Exp Biol J* 6(7):2472-2476.
- \*Aposhian HV, Maiorino RM, Rivera M, et al. 1992b. Human studies with the chelating agents, DMPS and DMSA. *J Toxicol Clin Toxicol* 30(4):505-528.
- \*Arakawa O, Nakahiro M, Narahashi T. 1991. Mercury modulation of GABA-activated chloride channels and non-specific cation channels in rat dorsal root ganglion neurons. *Brain Res* 551:58-63.
- \*Arito H, Takahashi M. 1991. Effect of methylmercury on sleep patterns in the rat. In: Suzuki T, Imura N, Clarkson TW, eds. *Advances in Mercury Toxicology*. New York, NY: Plenum Press, 381-394.
- \*Armbuster G, Gutenmann WH, Lisk DJ. 1988. The effects of six methods of cooking on residues of mercury in striped bass. *Nutr Rep Int* 37(1):123-126.
- \*Armstrong RL, Leach L, Belluscio P, et al. 1963. Behavioral change in the pigeon following inhalation of mercury vapor. *Am Ind Hyg Assoc J* 24:336-375.
- \*Aronow R, Cabbage C, Wisner R, et al. 1990. Mercury exposure from interior latex paint. *Morbidity and Mortality Weekly Report* 39(8):125-126.
- \*Aronsson A-M, Lind B, Nylander M, et al. 1989. Dental amalgam and mercury. *Biol Metals* 2:25-30 (As cited in Weiner and Nylander 1995).
- \*Arvidson B. 1992. Accumulation of inorganic mercury in lower motoneurons of mice. *Neurotoxicol* 13(1):277-280.
- \*Aschner M, Aschner JL. 1990. Mercury neurotoxicity: Mechanisms of blood-brain barrier transport. *Neurosci Biobehav Rev* 14(2):169-176.
- \*Aschner M, Clarkson TW. 1988. Distribution of mercury 203 in pregnant rats and their fetuses following systemic infusions with thiol-containing amino acids and glutathione during late gestation. *Teratology* 38(2):145-155.
- \*Ashe W, Largent E, Dutra F, et al. 1953. Behavior of mercury in the animal organism following inhalation. *Arch Ind Hyg Occup Med* 17:19-43.
- \*ASTER. 1997. ASTER (Assessment Tools for the Evaluation of Risk) ecotoxicity profile. Duluth, MN: Environmental Research Laboratory, U.S. Environmental Protection Agency,

## 8. REFERENCES

- \*Aten J, Veninga A, Bruijn JA, et al. 1992. Antigenic specificities of glomerular-bound autoantibodies in membranous glomerulopathy induced by mercuric chloride. *Clin Immunol Immunopathol* 63:89-102.
- \*Aten J, Veninga A, Deheer E, et al. 1991. Susceptibility to the induction of either autoimmunity or immunosuppression by mercuric-chloride is related to the major histocompatibility complex class-II haplotype. *Eur J Immunol* 21(3):611-616.
- \*Atkinson WS. 1943. A colored reflex from the anterior capsule of the lens which occurs in mercurialism. *Am J Ophthal* 26:685-688.
- \*ATSDR. 1989. Decision guide for identifying substance-specific data needs related to toxicological profiles. Agency for Toxic Substances and Disease Registry, Division of Toxicology, Atlanta, GA.
- \*ATSDR. 1990. Final Report. Technical assistance to the Tennessee Department of Health and Environment. Mercury exposure study Charleston, Tennessee. Atlanta, GA. US Department of Health and Human Services, Public Health Service, Centers for Disease Control, Agency for Toxic Substances and Disease Registry. CDC Centers for Disease Control and Prevention 1995. Mercury Exposure in a residential community- Florida, 1994 *MMWR* 44 (23):436-437.
- \*ATSDR. 1992. Case studies in environmental medicine -mercury toxicity. US Department of Health and Human Services Public Health Service.
- \*ATSDR. 1993. Exposure to hazardous substances and reproductive health. *American Family Physician* 48(8):1441-1448.
- \*ATSDR. 1997. Background: ATSDR and EPA warn the public about continuing patterns of metallic mercury exposure. Department of Health and Human Services. Agency for Toxic Substances and Disease Registry Public Health Service.
- \*ATSDR. 1998. National Alert: A warning about continuing patterns of metallic mercury exposure. Agency for Toxic Substances and Disease Registry, <http://atsdr.atsdr.cdc.gov.8080/alerts/970626.html>.
- ATSDR. [In Press]. Support Report for the Expert Panel Review of the Toxicological Profile for Mercury. Agency for Toxic Substances and Disease Registry Public Health Service.
- AWWA. 1995. Standard methods for the examination of water and wastewater. 3112A, 3112B, 3113A, 3500-Hg A, 3500-Hg B, 3500-Hg C, 3500-Mo A, B, C.
- \*Ayyadurai K, Krishnashamy V. 1988. A study of mercury concentration in nails, hair, and urine of dentists, dental assistants and non-dental personnel. *J Environ Biol* 9(3):281-282.
- \*Bache CA, Gutenmann WH, Rutzke M, et al. 1991. Concentrations of metals in grasses in the vicinity of a municipal refuse incinerator. *Arch Environ Contam Toxicol* 20:538-542.
- \*Baeyens W, Leermakers M. 1989. Determination of metallic mercury and some organomercury compounds using atomic-absorption spectrometry after amalgamation on a gold column. *J Anal At Spectrom* 4(7):635-640.

## 8. REFERENCES

- \*Bagedahl-Strindlund M, Ilie M, Furhoff AK, et al. 1997. A multidisciplinary clinical study of patients suffering from illness associated with mercury release from dental restorations: Psychiatric aspects. *Acta Psychiatr Scand* Dec 96(6):475-82.
- \*Baggett J, Berndt W. 1984. Interaction of potassium dichromate with the nephrotoxins, mercuric chloride and citrinin. *Toxicology* 33:157-169.
- \*Baglan RJ, Brill AB, Schulert A, et al. 1974. *J Environ Research* 8:64-end.
- \*Bagley MP, Schwartz RA, Lambert WC. 1987. Hyperplastic reaction developing within a tattoo: Granulomatous tattoo reaction, probably to mercuric sulfide (cinnabar). *Arch Dermatol* 123(11):1557-1561.
- \*Bakir F, Damluji SF, Amin-Zaki L, et al. 1973. Methylmercury poisoning in Iraq. *Science* 181:230-241.
- \*Bala KV Sridevi K Rao KP. 1993. Inhibition of methyl mercury chloride-induced chromosomal damage by gamma-linolenic acid. *Food Chem Toxicol* 31(6):431-4.
- \*Baldi F. 1988. Mercury pollution in the soil and mosses around a geothermal plant. *Water Air Soil Pollut* 38(1-2):111-119.
- \*Baldi F, Filippelli M. 1991. New method for detecting methylmercury by its enzymic conversion to methane. *Environ Sci Technol* 25(2):302-305.
- \*Ballatori N, Clarkson T. 1982. Developmental changes in the biliary excretion of methylmercury and glutathione. *Science* 216(2):61-63.
- \*Balogh S, Llang L. 1995. Mercury pathways in municipal wastewater treatment plants. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994.* Boston, MA: Kluwer Academic Publishers, 1181-1190.
- \*Baluja G, Hernandez LM, Gonzalez Ma. J, et al. 1982. Presence of organochlorine pesticides, polychlorinated biphenyls and mercury in Spanish human milk samples. *Bull Environ Contam Toxicol* 28:573-577.
- \*Bandyopadhyay S, Das AK. 1989. Determination of mercury in soil by cold vapour AAS after its separation with Aliquat-336. *J Indian Chem Soc* 66(6):427-428.
- \*Bapu C, Purohit RC, Sood PP. 1994. Fluctuation of trace elements during methylmercury toxication and chelation therapy. *Hum Exper Toxicol* 113(12):815-823.
- \*Baranski B, Szymczyk I. 1973. Effects of mercury vapor upon reproductive functions of female white rats. *Med Pr* 24:248. (Polish)
- \*Barber RE. 1978. Inorganic mercury intoxication reminiscent of amyotrophic lateral sclerosis. *J Occup Med* 20:667-669.
- \*Barkay T, Liebert C, Gillman M. 1989. Environmental significance of the potential for mer(TN21)-mediated reduction of Hg<sup>2+</sup> to Hg<sup>0</sup> in natural waters. *Appl Environ Microbiol* 55(5):1196-1202.

## 8. REFERENCES

- \*Barnes DG, Dourson M. 1988. Reference dose (RfD): Description and use in health risk assessments. U.S. Environmental Protection Agency. Regul Toxicol Pharmacol 8:471-486.
- \*Barnes JL, McDowell EM, McNeil JS, et al. 1980. Studies on the pathophysiology of acute renal failure. V. Effect of chronic saline loading on the progression of proximal tubular injury and functional impairment following administration of mercuric chloride in the rat. Vichows Arch B 32:233-260.
- \*Barnett MO, Harris LA, Turner RR, et al. 1997. Formation of mercuric sulfide in soil. Environmental Science Technology 31(11):3037-3043.
- \*Barr, RD, Rees PH, Cordy PE, et al. 1972. Nephrotic syndrome in adult Africans in Nairobi. Brit Med J 2:131-134.
- \*Barr RD, Woodger MB, Rees PH. 1973. Levels of mercury in urine correlated with the use of skin lightening creams. Am J Clin Pathol 59:36-40.
- \*Barregard L, Hogstedt B, Schutz A, et al. 1991. Effects of occupational exposure to mercury vapor on lymphocyte micronuclei. Scand J Work Environ Health 17(4):263-268.
- \*Barregard L, Horvat M, Schutz A. 1994a. No indication of *in vivo* methylation of inorganic mercury in chloralkali workers. Environ Research 67(2):160-167.
- \*Barregard L, Hultberg B, Schutz A, et al. 1988. Enzymuria in workers exposed to inorganic mercury. Int Arch Occup Environ Health 61(1-2):65-69.
- \*Barregard L, Lindstedt G, Schutz A, et al. 1994b. Endocrine function in mercury exposed chloralkali workers. Occup Environ Med 51(8):536-540.
- \*Barregard L, Sallsten G, Jarvholm B. 1990. Mortality and cancer incidence in chloralkali workers exposed to inorganic mercury. Br J Ind Med 47(2):99-104.
- \*Barregard L, Sallsten G, Jarvholm B. 1995. People with high mercury uptake from their own dental amalgam fillings. Occup Environ Med 52:124-128.
- \*Barregard L, Sallsten G, Schutz A, et al. 1992. Kinetics of mercury in blood and urine after brief occupational exposure. Arch Environ Health 47(3):176-184.
- \*Baser ME, Marion D. 1990. A statewide case registry for surveillance of occupational heavy metals absorption. Am J Pub Health 80(2):162-164.
- \*Baxter DC, Frech W. 1989. Determination of mercury by atomic-absorption spectrometry using a platinum-lined graphite furnace for in situ pre-concentration. Anal Chim Acta 225(1):175-183.
- \*Baxter DC, Frech W. 1990. Critical comparison of two standard digestion procedures for the determination of total mercury in natural water samples by cold vapor atomic absorption spectrometry. Anal Chim Acta 236(2):377-384.

## 8. REFERENCES

- \*Becker PR, Mackey EA, Schantz MM et al. 1995. Concentrations of chlorinated hydrocarbons, heavy metals and other elements in tissues banked by the Alaska Marine Mammal Tissue Archival Project. US Dept of commerce, National Institute of Standards and Technology NISTIR 5620.[retrieval in progress]
- \*Beckert WF, Messman JD, Churchwell ME, et al. 1990. Evaluation of SW-846 cold-vapour mercury methods 7470 and 7471. In: Friedman D, ed. Waste testing and quality assurance. Second volume. Philadelphia, PA: American Society for Testing and Material, STP 1062:247-257.
- \*Behmanesh N, Allen DT, Warren JL. 1992. Flow rates and compositions of incinerated waste streams in the United States. J Air Waste Manag Assoc 42(4):437-442.
- \*Bell ZG, Lovejoy HB, Vizona TR. 1973. Mercury exposure evaluations and their correlations with urine mercury excretion: 3. Time-weighted average (TWA) mercury exposures and urine mercury levels. J Occup Med 15:501-508.
- \*Bencko V, Wagner V, Wagnerova M, et al. 1990. Immunological profiles in workers occupationally exposed to inorganic mercury. J Hyg Epidemiol Microbiol Immunol 34(1):9-15.
- \*Bengt-Goran S, Nilsson A, Jonsson E, et al. 1995. Fish consumption and exposure to persistent organochlorine compounds, mercury, selenium and methylamines among Swedish fishermen. Scand J Work Environ Health 21(2):96-105.
- \*Benoit JM, Fitzgerald WF, Damman AWH. 1994. Historic atmospheric mercury deposition in the mid-continental United States as recorded in an ombrotrophic peat bog. In: Watras CJ Huckabee JW eds. Mercury as a global pollutant. Ann Arbor: Lewis Publishers, 187-202.
- \*Berdouses E, Vaidyanathan TK, Dastane A, et al. 1995. Mercury release from dental amalgams: An *in vitro* study under controlled chewing and brushing in an artificial mouth. J Dent Res 74(5):1185-1193.
- \*Bergdahl IA, Schutz A, Hansson G-A. 1995. Automated determination of inorganic mercury in blood after sulfuric acid treatment using cold vapour atomic absorption spectrometry and an inductively heated gold trap. Analyst 120:1205-1209.
- \*Berglund A. 1990. Estimation by a 24-hour study of the daily dose of intra-oral mercury vapor inhaled after release from dental amalgam. J Dent Res 69(10):1646-1651.
- \*Berglund A, Molin M. 1996. Mercury vapor release from dental amalgam in patients with symptoms allegedly caused by amalgam fillings. Eur J Oral Sci 104(1):56-63.
- \*Berglund F, Berlin M, Birke G, et al. 1971. Methyl mercury in fish. A toxicologic-epidemiologic evaluation of risks. Report from an expert group. Nordisk Hygienisk Tidskrift. Supplementum 4. Stockholm.
- \*Berlin M. 1963. Renal uptake, excretion and retention of mercury: Part II. A study in the rabbit during infusion of methyl- and phenylmercuric compounds. Arch Environ Health 6:626-633.
- \*Berlin M. 1986. Mercury. In: Friberg L, Nordberg GR, Vouk VB, eds. Handbook on the toxicology of metals. 2nd ed. New York, NY: Elsevier Press.



## 8. REFERENCES

- \*Berlin M, Blomstrand C, Grand CA, et al. 1975. Tritiated methylmercury in the brain of squirrel monkeys. *Arch Environ Health* 30:591-597.
- \*Berlin M, Fazackerly J, Nordberg G. 1969. The uptake of mercury in the brains of mammals exposed to mercury vapor and mercuric salts. *Arch Environ Health* 18:719-729.
- \*Berlin M, Gibson S. 1963. Renal uptake, excretion and retention of mercury: Part I. A study in the rabbit during infusion of mercuric chloride. *Arch Environ Health* 6:56-63.
- \*Berlin M, Jerksell LG, von Ubisch H. 1966. Uptake and retention of mercury in the mouse brain--a comparison of exposure to mercury vapor and intravenous injection of mercuric salt. *Arch Environ Health* 12:33-42.
- \*Berlin M, Johansson LG. 1964. Mercury in mouse brain after inhalation of mercury vapor and after intravenous injection of mercury salt. *Nature* 204:84-87.
- \*Berlin M, Rylander R. 1964. Increased brain uptake of mercury induced by 2,3-dimercaptopropanol (BAL) in mice exposed to phenylmercuric acetate. *J Pharmacol Exp Ther* 146(2):236-240.
- \*Berlin M, Ullberg S. 1963. Accumulation and retention of mercury in the mouse: Part II. An autoradiographic comparison of phenylmercuric acetate with inorganic mercury. *Arch Environ Health* 6:602-609.
- \*Bernard AM, Collette C, Lauwerys R. 1992. Renal effects of *in utero* exposure to mercuric chloride in rats. *Arch Toxicol* 66(7):508-513.
- \*Bernard AM, Roels HR, Foidart JM, et al. 1987. Search for anti-laminin antibodies in the serum of workers exposed to cadmium, mercury vapor or lead. *Arch Occup Environ Health* 59:303-309.
- \*Berthoud HR, Garman RH, Weiss B. 1976. Food intake, body weight, and brain histopathology in mice following chronic methylmercury treatment. *Toxicol Appl Pharmacol* 36:19-30.
- \*Best CH. 1961. *The physiological basis of medical practice*. Baltimore, 19, 29.
- \*Betti C, Barale R, Pool-Zobel BL. 1993a. Comparative studies on cytotoxic and genotoxic effects of two organic mercury compounds in lymphocytes and gastric mucosa cells of Sprague-Dawley rats. *Environmental and Molecular Mutagenesis* 22(3):172-180.
- \*Betti C, Davini T, Barale R. 1992. Genotoxic activity of methyl mercury chloride and dimethyl mercury in human lymphocytes. *Mutat Res* 281(4):255-260.
- \*Betti C, Davini T, He J, et al. 1993b. Liquid holding effects on methylmercury genotoxicity in human lymphocytes. *Mutat Res* 301(4):267-273.
- \*Beusterien KM, Etzel RA, Agocs MM, et al. 1991. Indoor air mercury concentrations following application of interior latex paint. *Arch Environ Contam Toxicol* 21:62-64.
- \*Biber TUL, Mylle M, Baines AD, et al. 1968. A study of micropuncture and microdissection of acute renal damage in rats. *Am J Med* 44:664-705.

## 8. REFERENCES

- \*Bichler M. 1991. Determination of mercury in mineral samples by employing a high temperature volatilization technique and activation analysis. *J Radioanal Nucl Chem* 154(4):255-263.
- \*Bidstrup P, Bonnell J, Harvey DG, et al. 1951. Chronic mercury poisoning in men repairing direct current meters. *Lancet*:856-861.
- \*Bigazzi PE. 1992. Lessons from animal models: The scope of mercury-induced autoimmunity. *Clin Immunol Immunopathol* 65(2):81-84.
- \*Birke G, Johnels AG, Plantin L-O, et al. 1972. Studies on humans exposed to methylmercury through fish consumption. *Arch Environ Health* 25:77-91.
- \*Birnie SE. 1988. Automated continuous monitoring of inorganic and total mercury in waste water and other waters by flow-injection analysis and cold-vapour atomic-absorption spectrometry. *J Auto Chem* 10(3):140-143.
- \*Biro L, Klein WP. 1967. Unusual complications of mercurial (cinnabar) tattoo: Generalized eczematous eruption following laceration of a tattoo. *Arch Dermatol* 96(2):165-167.
- \*Bjorklund H, Hoffer B, Olson L, et al. 1981. Differential morphological changes in sympathetic nerve fibers elicited by lead, cadmium and mercury. *Environ Res* 26:69-80.
- \*Björkman L, Mottet K, Nylander M, et al. 1995. Selenium concentrations in brain after exposure to methylmercury: Relations between the inorganic mercury fraction and selenium. *Arch Toxicol* 69:228-234.
- \*Björkman L, Sandborgh-Englund G, Ekstrand J. 1997. Mercury in saliva and feces after removal of amalgam fillings. *Toxicol Appl Pharmacol* 144:156-162.
- \*Bjornberg A, Hakanson L, Lundbergh K. 1988. A theory on the mechanisms regulating the bioavailability of mercury in natural waters. *Environ Pollut* 49(1):53-61.
- \*Blakley BR. 1984. Enhancement of urethane-induced adenoma formation in Swiss mice exposed to methylmercury. *Can J Comp Med* 48:299-302.
- \*Blayney MB, Winn JS, Nierenberg DW. 1997. Handling dimethylmercury, *Chemical and Engineering News* 75(19):7.
- \*Bloch P, Shapiro IM. 1986. An x-ray fluorescence technique to measure in situ the heavy metal burdens. *J Occup Med* 28:609-614.
- \*Blondell JM, Knott SM. 1993. Risk analysis for phenylmercuric acetate in indoor latex house paint. *ACS Symposium Series* 522:307-317.
- \*Bloom N. 1989. Determination of picogram levels of methylmercury by aqueous phase ethylation followed by cryogenic gas chromatography with cold vapor atomic fluorescence detection. *Can J Fish Aquat Sci* 46(7):1131-1140.
- \*Bloom NS. 1992. On the chemical form of mercury in edible fish and marine invertebrate tissue. *Can J Fish Aquat Sci* 49(5):1010-1017.

## 8. REFERENCES

- \*Bloom NS. 1993. Sampling and analysis for mercury in environmental media of importance to the natural gas industry. GRI Topical Report Gas Research Institute (8600 West Bryn Maur Avenue, Chicago, IL 60631 USA).
- \*Bloom NS, Effler SW. 1990. Seasonal variability in the mercury speciation of Onodaga Lake (New York). *Water Air Soil Pollut* 53(3-4):251-265.
- \*Bloom NS, Fitzgerald WF. 1988. Determination of volatile mercury species at the picogram level by low temperature gas chromatography with cold-vapor atomic fluorescence detection. *Anal Chim Acta* 208:151-161.
- \*Bluhm RE, Bobbitt RG, Welch LW, et al. 1992a. Elemental mercury vapour toxicity, treatment, and prognosis after acute, intensive exposure in chloralkali plant workers: Part I. History, neuropsychological findings and chelator effects. *Hum Exp Toxicol* 11(3):201-210.
- \*Bluhm RE, Breyer JA, Bobbitt RG, et al. 1992b. Elemental mercury vapour toxicity, treatment, and prognosis after acute, intensive exposure in chloralkali plant workers: Part II. Hyperchloraemia and genitourinary symptoms. *Hum Exp Toxicol* 11(3):211-215.
- \*Blume HP, Brummer G. 1991. Prediction of heavy metal behavior in soil by means of simple field tests. *Ecotoxicol Environ Safety* 22:164-174.
- \*Bodek I, Lyman WJ, Reehl WF, et al. 1988. Mercury. In: *Environmental inorganic chemistry*. New York NY: Pergamon Press.
- \*Bonhomme C, Gladyszczak-Kholer J, Cadou A, et al. 1996. Mercury poisoning by vacuum-cleaner aerosol (1). *Lancet* 347(8994):115.
- \*Boogaard PJ, Houtsma AT, Journee HL, et al. 1996. Effects of exposure to elemental mercury on the nervous system and the kidneys of workers producing natural gas. *Arch Environ Health* 51(2):108-15.
- \*Borgmann U, Whittle DM. 1991. Contaminant concentration trends in Lake Ontario lake trout (*Salvelinus namaycush*). *J Gt Lakes Res* 17(3):368-381.
- \*Borjesson J, Alpsten M, Huang S, et al. 1993. *In vivo* x-ray fluorescence analysis with application to platinum, gold and mercury in man-experiments, improvements, and patient measurements. In: Ellis KJ, Eastman JD eds., *Human body composition*. New York NY: Plenum Press.
- \*Bornhausen M, Musch MR, Greim H. 1980. Operant behavior performance changes in rats after prenatal methylmercury exposure. *Toxicol Appl Pharmacol* 56:305-316.
- \*Boscolo P, Carmignani M, Giuliano G, et al. 1989. Peripheral catecholaminergic mechanisms and baroreflex pathways are involved in vascular and cardiac effects of long-term exposure to inorganic mercury in rats. In: Strano A, Novo S, eds. *Advances in vascular pathology*. Amsterdam: Elsevier Science Publishers, 1061-1066.
- \*Bourgeois M, Doods-Goossens A, Knockaert D, et al. 1986. Mercury intoxication after topical application of a metallic mercury ointment. *Dermatologica* 172:48-51.

## 8. REFERENCES

- \*Bowman C, Mason DW, Pusey CD, et al. 1984. Autoregulation of autoantibody synthesis in mercuric chloride nephritis in the brown Norway rat: A role for T suppressor cells. *Eur J Immunol* 14:464-470.
- \*Bradberry SM, Feldman MA, Braithwaite RA, et al. 1996. Elemental mercury-induced skin granuloma: a case report and review of the literature. *J Toxicol Clin Toxicol* 34 (2):209-16.
- \*Braghioli D, Parenti C, Di Bella M, et al. 1990. Follow-up of methylmercury concentration in brain areas of developing rats exposed during prenatal life using cold-vapor absorption spectrometry. *Boll Chim Farm* 129(7-8):259-262.
- \*Bratel J, Hakeberg M, Jontell M. 1996. Effect of replacement of dental amalgam on oral lichenoid reactions. *J Dent* 24(1-2):41-45.
- \*Bressa G, Cima L, Costa P. 1988. Bioaccumulation of mercury in the mushroom *Pleurotus ostreatus*. *Ecotoxicol Environ Safety* 16(2):85-89.
- \*Bronstein AC, Currance PL. 1988. Emergency care for hazardous materials exposure. Washington, DC: The C.V. Mosby Company, 183-184.
- \*Broomhall J, Kovar IZ. 1986. Environmental pollutants in breast milk. *Rev Environ Health*.
- \*Brosset C, Lord E. 1991. Mercury in precipitation and ambient air: A new scenario. *Water Air Soil Pollut* 56:493-506.
- \*Brown IA. 1954. Chronic mercurialism: A cause of the clinical syndrome of amyotrophic lateral sclerosis. *Arch Neurol Psychiatry* 72:674-681.
- \*Brown TD, Schmidt CE, Radziwon AS. 1993. Comprehensive assessment of toxic emissions from coal-fired power plants. In: Chow W, Connor KK, eds. *Managing hazardous air pollutants - state of the art*. Boca Raton, Florida: Lewis Publishers, 116-125.
- \*Bryan GW, Langston WJ. 1992. Bioavailability, accumulation and effects of heavy-metals in sediments with special reference to United-Kingdom estuaries: A review. *Environmental Pollution* 76(2):89-131.
- \*Bryan SE, Guy AL, Hardy KJ. 1974. Metal constituents of chromatin interaction of mercury *in vivo*. *Biochem* 13:313-319.
- \*Buchet J, Roels H, Bernard A, et al. 1980. Assessment of renal function of workers exposed to inorganic lead, cadmium, or mercury vapor. *J Occup Med* 22:741-750.
- \*Bucknell M, Hunter D, Milton R, et al. 1993. Chronic mercury poisoning. *Br J Ind Med* 50(2):97-106.
- \*Buelke-Sam J, Kimmel CA, Adams, et al. 1985. Collaborative behavioral teratology study: Results. *Neurobehav Toxicol Teratol* 7:591-624.
- \*Bulger RE. 1986. Renal damage caused by heavy metals. *Toxicol Pathol* 14:58-65.

## 8. REFERENCES

- \*Bullock OR. 1997. Langrangian modeling of mercury air emission, transport, and deposition: An analysis of model sensitivity to emissions uncertainty. Atmospheric Sciences Modeling Division Air Resources Laboratory, National Oceanic and Atmospheric Administration, Research Triangle Park, NC.
- \*Bulska E, Emteborg H, Baxter DC, et al. 1992. Speciation of mercury in human whole blood by capillary gas chromatography with a microwave-induced plasma emission detector system following complexometric extraction and butylation. *Analyst* 117(3):657-663.
- \*Buneaux F, Buisine A, Bourdon S, et al. 1992. Continuous-flow quantification of total mercury in whole blood, plasma, erythrocytes and urine by inductively coupled plasma atomic-emission spectroscopy. *J Anal Toxicol* 16(2):99-101.
- \*Burbacher TM, Mohamed MK, Mottett NK. 1988. Methylmercury effects on reproduction and offspring size at birth. *Reprod Toxicol* 1(4):267-278.
- \*Burbacher TM, Monnett C, Grant KS, et al. 1984. Methylmercury exposure and reproductive dysfunction in the nonhuman primate. *Toxicol Appl Pharmacol* 75:18-24.
- \*Burger J, Cooper K, Gochfeld M. 1992. Exposure assessment for heavy metal ingestion from a sport fish in Puerto Rico: Estimating risk for local fishermen. *J Toxicol Environ Health* 36(4):355-365.
- \*Burke J, Hoyer M, Keeler G, et al. 1995. Wet deposition of mercury and ambient mercury concentrations at a site in the Lake Champlain basin. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference held in Whistler, British Columbia, July 10-14, 1994.* Boston, MA: Kluwer Academic Publishers, 353-362.
- \*Burton GV, Meikle AW. 1980. Acute and chronic methylmercury poisoning impairs rat adrenal and testicular function. *J Toxicol Environ Health* 6:597-606.
- \*Bushee DS. 1988. Speciation of mercury using liquid chromatography with detection by inductively coupled plasma mass spectrometry. *Analyst* 113(8):1167-1170.
- \*Buzina R, Suboticane K, Vukusic J, et al. 1989. Effect of industrial pollution on seafood content and dietary intake of total and methylmercury. *Sci Total Environ* 78:45-57.
- \*Cacho J, Castells JE. 1989. Determination of mercury in wine by flameless atomic-absorption spectrophotometry. *At Spectrosc* 10(3):85-88.
- \*Cagiano R, De Salvia MA, Renna G, et al. 1990. Evidence that exposure to methyl mercury during gestation induces behavioral and neurochemical changes in offspring of rats. *Neurotoxicol Teratol* 12(1):23-28.
- \*Callahan MA, Slimak MW, Gabel NW, et al. 1979. Water related environmental fate of 129 priority pollutants, introduction and technical background, metals and inorganics, pesticides and PCBs. Washington, D.C: U.S. Environmental Protection Agency, Office of Water Waste and Management. Document no. EPA 440/4-79-029a., 14-1 -14-15.
- \*Campbell J. 1948. Acute mercurial poisoning by inhalation of metallic vapor in an infant. *Can Med Assoc J* 58:72-75.

## 8. REFERENCES

- \*Campbell MB, Kanert GA. 1992. High-pressure microwave digestion for the determination of arsenic, antimony, selenium and mercury in oily wastes. *Analyst (London)* 117(2):121-124.
- \*Canady RA, Hanley JE, Susten AS. 1997. ATSDR science panel on the bioavailability of mercury in soils: lessons learned. *Risk Anal* Oct 17 (5):527-32.
- \*Cantoni O, Christie NT, Robison SH, et al. 1984a. Characterization of DNA lesions produced by HgCl<sub>2</sub> in cell culture systems. *Chem Biol Interact* 49:209-224.
- \*Cantoni O, Christie NT, Swann A, et al. 1984b. Mechanism of HgCl<sub>2</sub> cytotoxicity in cultured mammalian cells. *Mol Pharmacol* 26:360-368.
- \*Cantoni O, Costa M. 1983. Correlations of DNA strand breaks and their repair with cell survival following acute exposure to mercury(II) and X-rays. *Mol Pharmacol* 24:84-89.
- \*Cantoni O, Evans RM, Costa M. 1982. Similarity in the acute cytotoxic response of mammalian cells to mercury (II) and X-rays: DNA damage and glutathione depletion. *Biochem Biophys Res Commun* 108:614-619.
- \*Cappon CJ, Smith JC. 1982. Chemical form and distribution of mercury and selenium in edible seafood. *J Anal Toxicol* 6:10-21.
- \*Cardenas A, Roels H, Bernard Am, et al. 1993. Markers of early renal changes induced by industrial pollutants. I. Application to workers exposed to mercury vapour. *Br J Ind Med* 50(1):17-27.
- \*Carmignani M, Boscolo P, Artese L, et al. 1992. Renal mechanisms in the cardiovascular effects of chronic exposure to inorganic mercury in rats. *Br J Ind Med* 49(4):226-232.
- \*Carmignani M, Boscolo P, Preziosi P. 1989. Renal ultrastructural alterations and cardiovascular functional changes in rats exposed to mercuric chloride. *Arch Toxicol (Suppl 13)*:353-356.
- \*Caron GA, Poutala S, Provost TT. 1970. Lymphocyte transformation induced by inorganic and organic mercury. *Int Arch Allergy Appl Immunol* 37:76-87.
- \*Carpi A, Lindberg SE, Prestbo EM, et al. 1998. Methyl mercury contamination and emission to the atmosphere from soil amended with municipal sewage sludge. *J Environ Qual* 26(6):1650-1655.
- \*Carrico LC. 1985. Mercury. In: Mineral facts and problems. Bulletin 675. Washington, DC: U.S. Department of the Interior, Bureau of Mines, 499-508.
- \*Carrillo F, Bonilla M, Camara C. 1986. Determination of mercury in biological samples by a sensitized cold vapor atomic absorption technique. *Microchemical Journal* 33:2-8.
- \*Cassano GB, Armaducci L, Viola PL. 1966. Distribution of mercury in the brain of chronically intoxicated mice (autoradiographic study). *Riv Patol Nerv Ment* 87:214-225.
- \*Cassano GB, Viola PL, Ghetti B, et al. 1969. The distribution of inhaled mercury (Hg<sup>203</sup>) vapors in the brain of rats and mice. *J Neuropathol Exp Neurol* 28:308-320.

## 8. REFERENCES

- \*Castedo M, Pelletier L, Rossert J, et al. 1993. Mercury-induced autoreactive anti-class II T cell line protects from experimental autoimmune encephalomyelitis by the bias of CD8 + Antiergotypic cells in Lewis rats. *J Exp Med* 177:881-889.
- \*Cavalleri A, Belotti L, Gobba F, et al. 1995. Colour vision loss in workers exposed to elemental mercury vapour. *Toxicol Lett* 77:351-356.
- \*Cavanagh JB, Chen FCK. 1971. The effects of methyl-mercury-dicyandiamide on the peripheral nerves and spinal cord of rats. *Acta Neuropathol (Berlin)* 19:208-215.
- \*CDC. 1995. Mercury exposure in a residential community-Florida 1994. Centers For Disease Control. *Morbidity and Mortality Weekly Report* 44 (23):436-443.
- \*CDC/ATSDR. 1990. Biomarkers of organ damage or dysfunction for the renal, hepatobiliary and immune systems. Atlanta, GA: CDC/ATSDR Subcommittee on Biomarkers of Organ Damage and Dysfunction, Centers for Disease Control, Agency for Toxic Substances and Disease Registry. Summary report, August 27, 1990.
- \*Cember H, Gallagher P, Faulkner A. 1968. Distribution of mercury among blood fractions and serum proteins. *Am Ind Hyg Assoc J* 29:233-237.
- \*Cernichiari E, Brewer R, Myers GJ, et al. 1995b. Monitoring methylmercury during pregnancy: Maternal hair predicts fetal brain exposure. *Neurotoxicology* 16 (4):705-10.
- \*Cernichiari E, Toribara TY, Liang L, et al. 1995a. The biological monitoring of mercury in the Seychelles study. *Neurotoxicology* 16(4):613-28.
- \*Chaffin D, Dinman B, Miller J, et al. 1973. An evaluation of the effects of chronic mercury exposure on EMG and psychomotor functions. Washington, DC: U.S. Department of Health and Human Services, National Institute of Occupational Safety and Health. Document no. HSM-099-71-62.
- \*Chambers BJ, Klein NW. 1993. Role of laminin autoantibodies on the embryo toxicity of sera from mercuric chloride treated brown Norway rats. *Reprod Toxicol* (4):333-341.
- \*Chan HM, Satoh M, Zalups RK, et al. 1992. Exogenous metallothionein and renal toxicity of cadmium and mercury in rats. *Toxicology* 76(1):15-26.
- \*Chang L, Hartmann HA. 1972a. Blood-brain barrier dysfunction in experimental mercury intoxication. *Acta Neuropathol (Berlin)* 21:179-184.
- \*Chang L, Hartmann HA. 1972b. Ultrastructural studies of the nervous system after mercury intoxication. *Acta Neuropathol (Berlin)* 20:122-138.
- \*Chang LW. 1983. Protective effects of selenium against methylmercury neurotoxicity: A morphological and biochemical study. *Exp Pathol* 23(3):143-56.
- \*Chang LW, Reuhl KR, Lee GW. 1977. Degenerative changes in the developing nervous system as a result of *in utero* exposure to methylmercury. *Environ Res* 14:414-425.
- \*Chang LW, Sprecher JA. 1976. Degenerative changes in the neonatal kidney following *in utero* exposure to methylmercury. *Environ Res* 11:392-406.

## 8. REFERENCES

- \*Chang LW, Yamaguchi S, Dudley JAW. 1974. Neurological changes in cats following long-term diet of mercury contaminated tuna. *Acta Neuropathol (Berlin)* 27:171-176.
- \*Chang YC, Yeh C, Wang JD. 1995. Subclinical neurotoxicity of mercury vapor revealed by a multimodality evoked potential study of chloralkali workers. *Amer J Ind Med* 27(2):271-279.
- \*Chapman LJ, Sauter SL, Henning RA, et al. 1990. Differences in frequency of finger tremor in otherwise asymptomatic mercury workers. *Br J Ind Med* 47(12):838-843.
- \*Charbonneau S, Munro I, Nera E, et al. 1976. Chronic toxicity of methylmercury in the adult cat. Interim Report. *Toxicology* 5:337-349.
- \*Charleston JS, Body RL, Bolender RP, Mottet NK, Vahter ME, Burbacher TM. 1996. Changes in the number of astrocytes and microglia in the thalamus of the monkey *Macaca fascicularis* following long-term subclinical methylmercury exposure. *Neurotoxicol* 17 (1):127-38.
- \*Charleston JS, Bolender RP, Mottet NK, et al. 1994. Increases in the number of reactive glia in the visual cortex of *Macaca fascicularis* following subclinical long-term methyl mercury exposure. *Toxicol Appl Pharmacol* 129(2):196-206.
- \*Chen W, Margara J, Endoh K, et al. 1990. Comparison of hair mercury concentrations between married couples. *Acta Med Biol* 38(1):45-50.
- \*Cherian L, Gupta VK. 1990. A simple field test for the detection of mercury in polluted water, air and soil samples. *Fresenius J Anal Chem* 336(5):400-402.
- \*Cherian MG, Clarkson TW. 1976. Biochemical changes in rat kidney on exposure to elemental mercury vapor: Effect on biosynthesis of metallothionein. *Chem Biol Interact* 12:109-120.
- \*Cherian MG, Hursh JG, Clarkson TW, et al. 1978. Radioactive mercury distribution in biological fluids and excretion in human subjects after inhalation of mercury vapor. *Arch Environ Health* 33:190-214.
- \*Chien Y-C, Feldman CA, Zohn HK, et al. 1996. Urinary mercury levels before and after amalgam restoration. *Science of the Total Environment* 188:39-47.
- \*Choi CM, Lapham LW, Amin-Zaki L, et al. 1978. Abnormal neuronal migration, deranged cerebral cortical organization and diffuse white matter astrocytosis of human fetal brain: A major effect of methylmercury poisoning *in utero*. *J Neuropathol Exp Neurol* 37:719-732.
- \*Christensen H, Krogh M, Nielsen M. 1937. Acute mercury poisoning in a respiration chamber. *Nature* 139:1026-1027.
- \*Christie NT, Cantoni O, Evans RM, et al. 1984. Use of mammalian DNA repair-deficient mutants to assess the effects of toxic metal compounds on DNA. *Biochem Pharmacol* 33:1661-1670.



## 8. REFERENCES

- \*Christie NT, Cantoni O, Sugiyama M, et al. 1985. Differences in the effects of Hg(II) on DNA repair induced in Chinese hamster ovary cells by ultraviolet or X-rays. *Mol Pharmacol* 29:173-178.
- \*Chu P, Porcella DB. 1995. Mercury stack emissions from U.S. electric utility power plants. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994.* Boston, MA: Kluwer Academic Publishers, 135-144.
- \*Chugh KS, Singhal PC, Uberoi HS. 1978. Rhabdomyolysis and renal failure in acute mercuric chloride poisoning. *Med J Aust* 2:125-126.
- \*Cicmanec JL. 1996. Comparison of four human studies of perinatal exposure to methylmercury for use in risk assessment. *Toxicol* 111(1-3):157-162.
- \*Cimino MC, Tice RR, Liang JC. 1986. Aneuploidy in mammalian somatic cells *in vivo*. *Mutat Res* 167:107-122.
- \*Cinca I, Dumitrescu I, Onaca P, et al. 1979. Accidental ethyl mercury poisoning with nervous system, skeletal muscle, and myocardium injury. *J Neurol Neurosurg and Psychiatry* 43:143-149
- \*Clarkson T, Cox C, Davidson PW, et al. 1998. Mercury in fish (1). *Science* 279/5350:459-460.
- \*Clarkson T, Small H, Norseth T. 1973. Excretion and absorption of methylmercury after polythiol resin treatment. *Arch Environ Health* 26:173-176.
- \*Clarkson TW. 1989. Mercury. *J Am Coll Toxicol* 8(7):1291-1296.
- \*Clarkson TW. 1971. Epidemiological and experimental aspects of lead and mercury contamination. *Food Cosmet Toxicol* 9:229-243.
- \*Clarkson TW. 1972a. Recent advances in toxicology of mercury with emphasis on the alkyl mercurials. *Crit Rev Toxicol* 203-234.
- \*Clarkson TW. 1972b. The pharmacology of mercury compounds. *Ann Rev Pharmacol* 12:375-406.
- \*Clarkson TW. 1978. The metabolism of inhaled mercury vapor in animals and man. Preprints of papers presented at the 176th National Meeting of the American Chemical Society, Division of Environmental Chemistry, September, Miami Beach, Fl. Washington, DC: American Chemical Society, 274-275.
- \*Clarkson TW. 1983. Mercury. *Annu Rev Public Health* 4:375-80.
- \*Clarkson TW. 1990. Human health risks from methylmercury in fish. *Environ Toxicol Chem* 9:957-961.
- \*Clarkson TW. 1992. Mercury: Major issues in environmental health. *Envir Health Persp* 100:31-38.
- \*Clarkson TW. 1995. Environmental contaminants in the food chain. *Am J Clin Nutr* 61(3):682s-686s.

## 8. REFERENCES

- \*Clarkson TW, Amin-Zaki L, Al-Tikriti SK. 1976. An outbreak of methylmercury poisoning due to consumption of contaminated grain. *Fed Proc* 35:2395-2399.
- \*Clarkson TW, Friberg L, Hursh JB, et al. 1988a. The prediction of intake of mercury vapor from amalgams. In: Clarkson TW, Friberg L, Nordberg GF, Sager PR eds. *Biological monitoring of toxic metals*. New York, NY: Plenum Press, 247-264.
- \*Clarkson TW, Gatzky J, Dalton C. 1961. Studies on the equilibration of mercury vapor with blood. Rochester, NY: University of Rochester Atomic Energy Project, Division of Radiation Chemistry and Toxicology.
- \*Clarkson TW, Hursh JB, Sager PR, et al. 1988b. Mercury. In: Clarkson TW, Hursh JB, Sager PR, et al. eds. *Biological monitoring of toxic metals*. New York: Plenum Press, 199-246.
- \*Clarkson TW, Magos L. 1966. Studies on the binding of mercury in tissue homogenates. *Biochem J* 99:62-70.
- \*Clarkson TW, Magos L, Greenwood MR. 1972. The transport of elemental mercury into fetal tissues. *Biol Neonate* 21:239-244.
- \*Clarkson TW, Rothstein A. 1964. The excretion of volatile mercury by rats injected with mercuric salts. *Health Phys* 10:1115-1121.
- \*Clayton GD, Clayton FE. 1981. *Patty's industrial hygiene and toxicology*, 3rd ed. New York, NY: John Wiley and Sons.
- \*Clewell HJ, Gearhart JM, Gentry PR, et al. 1999. Evaluation of the uncertainty in an oral reference dose for methylmercury due to interindividual variability in pharmacokinetics. *Risk analysis*, In Press.
- \*Clewell HJ, Gentry PR, and Shipp AM, et al. 1998. Determination of a site-specific reference dose for methylmercury for fish-eating populations. Peer-reviewed report for the Toxicology Excellence in Risk Assessment (TERA). International Toxicity Estimates for Risk (ITER) Database TERA, Cincinnati, OH, February, 1998. <http://www.tera.org/iter/>
- \*Clewell HJ III, Andersen M. 1985. Risk assessment extrapolations and physiological modeling. *Toxicol Ind Health* 1(4): 111-131.
- \*Cocking D, King ML, Ritchie L, et al. 1994. Earthworm bioaccumulation of mercury from contaminated flood plain soils. In: Watras CJ, Huckabee JW, eds. *Mercury pollution integration and synthesis*, 381-394.
- \*Cole HS, Hitchcock AL, Collins R. 1992. Mercury warning: The fish you catch may be unsafe to eat - A study of mercury contamination in the United States. Washington, D.C.: Clean Water Fund/Clean Water Action.
- \*Concas A, Corda MG, Salis M, et al. 1983. Biochemical changes in the rat cerebellar cortex elicited by chronic treatment with methylmercury. *Toxicol Lett* 18:27-33.
- \*Connelly NA, Brown TL, Knuth BA. 1990. New York State angler survey -1988. New York State Department of Environmental Conservation, Division of Fish and Wildlife, Albany, New York.
- \*Cope WG, Wiener JG, Rada RG. 1990. Mercury accumulation in yellow perch in Wisconsin Seepage Lakes: Relation to lake characteristics. *Environ Toxicol Chem* 9(7):931-940.

## 8. REFERENCES

- \*Cordier S, Deplan F, Mandereau L, et al. 1991. Paternal exposure to mercury and spontaneous abortions. *Br J Ind Med* 48(6):375-381.
- \*Corns WT, Stockwell PB, Jameel M. 1994. Rapid method for the determination of total mercury in urine samples using cold vapour atomic fluorescence spectrometry. *Analyst* 119:2481-2484.
- Cossa D. 1989. A review of the use of *Mytilus* spp. as quantitative indicators of cadmium and mercury contamination in coastal waters. *Oceanol Acta* 12(4):417-432.
- \*Cossa D, Gobeil C, Courau P. 1988. Dissolved mercury behavior in the St. Lawrence Estuary. *Estuarine Coastal Shelf Sci* 26(2):227-230.
- \*Costa M, Christie NT, Cantoni O, et al. 1991. DNA damage by mercury compounds: An overview. In: Suzuki T, Imura N, Clarkson TW, eds. *Advances in mercury toxicology*. New York, NY: Plenum Press, 255-273.
- \*Costanzo RB, Barry EF. 1988. Alternating current plasma detector for selective mercury detection in gas chromatography. *Anal Chem* 60(8):826-829.
- \*Cox C, Clarkson TW, Marsh DO, et al. 1989. Dose-response analysis of infants prenatally exposed to methyl mercury: An application of a single compartment model to single-strand hair analysis. *Environ Res* 49(2):318-332.
- \*CPSC. 1998. Notice of availability of guidance document on hazardous liquid chemicals in children's products. U.S. Consumer Product Safety Commission. *Federal Register*. 63 FR 29182. May 28, 1998.
- \*Cragle D, Hollis D, Qualters J, et al. 1984. A mortality study of men exposed to elemental mercury. *J Occup Med* 26:817-821.
- \*Creason JP et al. 1978a. Human scalp hair: An environmental exposure index for trace elements I. Fifteen trace elements in New York, NY (1971-1972), EPA 600/1-78-037a. US Environmental Protection Agency. Office of Research and Development. Health Effects Research Laboratory, Research Triangle Park, NC.
- \*Creason JP, et al. 1978b. Human scalp hair: an environmental exposure index for trace elements. II. 17 Trace elements in four New Jersey communities (1972). U.S. Environmental Protection Agency, Office of Research and Development. Health Effects Research Laboratory RTP NC. EPA 600/1-78-037b.
- \*Creason JP, et al. 1978c. Human scalp hair: an environmental exposure index for trace elements. III. 17 Trace elements in Birmingham, Alabama, and Charlotte, North Carolina. U.S. Environmental Protection Agency, Office of Research and Development. Health Effects Research Laboratory RTP NC. EPA 600/1-78-037c.
- \*CRITFC. 1994. A fish consumption survey of the Umatilla, Nez Perce, Yakima and Warm Springs tribes of the Columbia River Basin. Columbia River Inter-Tribal Fish Commission. Technical Report 94-3, 1-105.
- \*Crump K, Viren J, Silvers A, et al. 1995. Reanalysis of dose-response data from the Iraqi methylmercury poisoning episode. *Risk Analysis* 1(4):523-532.

## 8. REFERENCES

- \*Crump KS, Kjellstrom T, Shipp A, et al. 1998. Influence of prenatal exposure upon scholastic and psychological test performance: Benchmark analysis of a New Zealand Cohort. *Risk Analysis* 18(6): 701-713.
- \*Civin-Aralar MLA, Furness RW. 1991. Mercury and selenium interaction: A review. *Ecotoxicol Environ Safety* 21(3):348-364.
- \*Czuba M, Mortimer DC. 1980. Stability of methylmercury and inorganic mercury in aquatic plants (*Elodea densa*). *Can J Botany* 58:316-320.
- \*Danielsson BRG, Fredriksson A, Dahlgren L, et al. 1993. Behavioral effects of prenatal metallic mercury inhalation exposure in rats. *Neurotoxicol Teratology* 15:391-396.
- \*Danscher G, Horsted-Bindslev P, Rungby J. 1990. Traces of mercury in organs from primates with amalgam fillings. *Exp Mol Pathol* 52(3):291-299.
- \*Danziger SJ, Possick PA. 1973. Metallic mercury exposure in scientific glassware manufacturing plants. *J Occup Med* 15:15-20.
- \*Daston GP, Rehnberg BF, Hall LL, et al. 1986. Toxicity of mercuric chloride to the developing rat kidney: III. Distribution and elimination of mercury during postnatal maturation. *Toxicol Appl Pharmacol* 85:39-48.
- \*Davidson PW, Myers GJ, Cox C, et al. 1995a. Neurodevelopmental test selection, administration, and performance in the main Seychelles child development study. *Neurotoxicol* 16(4):665-676.
- \*Davidson PW, Myers GJ, Cox C, et al. 1995b. Longitudinal neurodevelopmental study of Seychellois children following *in utero* exposure to methylmercury from maternal fish ingestion: Outcomes at 19 and 29 months. *Neurotoxicol* 16(4):677-688.
- \*Davidson PW, Myers GJ, Cox C, et al. 1998. Effects of prenatal and postnatal methylmercury exposure from fish consumption on neurodevelopment: Outcomes at 66 months of age in the seychelles child development study. *JAMA* 280(8):701-707.
- \*Davis JA, Gunther AJ, O'Connor JM. 1992. Priority pollutant loads from effluent discharges to the San Francisco estuary. *Wat Environ Res* 64:134-140.
- \*Davis LE, Kornfeld M, Mooney HS, et al. 1994. Methylmercury poisoning: Long-term clinical, radiological, toxicological, and pathological studies of an affected family. *Ann of Neurol* 35(6):680-688.
- \*Davis LE, Wands JR, Weiss SA, et al. 1974. Central nervous system intoxication from mercurous chloride laxatives-quantitative, histochemical and ultrastructure studies. *Arch Neurol* 30:428-431.
- \*De Rosa CT, Johnson BL, Fay M, et al. 1996. Public health implications of hazardous waste sites: Findings, assessment and research. *Food Chem Tox* 34:1131-1138.
- \*Dean JG, Bosqui FL, Lanoveite KH. 1972. Removing heavy metals from waste water. *Environ Sci Technol* 6:518-522.

## 8. REFERENCES

- \*DeBont B, Lauwerys R, Govaerts H, et al. 1986. Yellow mercuric oxide ointment and mercury intoxication. *Eur J Pediatr* 145:217-218.
- \*Delio DA, Reuhl KR, Lowndes HE. 1992. Ectopic impulse generation in dorsal root ganglion neurons during methylmercury intoxication: An electrophysiological and morphological study. *Neurotoxicol* 13(3):527-539.
- \*Den Tonkelaar EM, Van Esch GJ, Hofman B, et al. 1974. Mercury and other elements in blood of the Dutch population. In: *Proceedings of an International Symposium on Recent Advances in the Assessment of the Health Effects of Environmental Pollution, Paris, 24-28 June, Luxembourg, Commission of the European Communities, Vol. 2, 1017-1027.*
- \*Dencker L, Danielsson B, Khayat A, et al. 1983. Deposition of metals in the embryo and fetus. In: Clarkson TW, Nordberg GG, Sager PR, eds. *Reproductive and developmental toxicity of metals.* New York, NY: Plenum Press, 607-631.
- \*Department of Public Health (Chicago). 1997. Mercury use in the Hispanic community of Chicago. Office of Hispanic Affairs, Chicago, Ill.
- \*Derobert L, Tara S. 1950. Mercury intoxication in pregnant women. *Ann Med Leg* 30:222-225 (French)
- \*Desjardins RM, Bradbury WC, Seyfried PL. 1988. Effects of metals from mine tailings on the microflora of a marsh treatment system. In: Adams WJ, Chapman GA, Landis WG, eds. *Aquatic toxicology and hazard assessment. 10th Vol.* Philadelphia, PA: American Society for Testing and Materials 10:491-502.
- \*DHHS. 1993. Dental amalgam: A scientific review and recommended public health service strategy for research, education and regulation. Department of Health and Human Services, Public Health Service, Washington, D.C.
- \*DHHS. 1997. Dental Amalgam and Alternative Restorative Materials. An Update Report to the Environmental Health Policy Committee. Working Group on Dental Amalgam. Department of Health and Human Services. Public Health Service. Washington, D.C. October 1997.
- \*Dick AL, Sheppard DS, Patterson JE. 1990. Mercury content of Antarctic surface snow: Initial results. *Atmos Environ Part A* 24A(4):973-978.
- \*Dieter MP, Boorman GA, Jameson CW, et al. 1992. Development of renal toxicity in F344 rats gavaged with mercuric-chloride for 2 weeks, or 2, 4, 6, 15, and 24 months. *J Toxicol Environ Health* 36(4):319-340.
- \*Dieter MP, Luster MI, Boorman GA, et al. 1983. Immunological and biochemical responses in mice treated with mercuric chloride. *Toxicol Appl Pharmacol* 68:218-228.
- \*Discalzi G, Fabbro D, Meliga F, et al. 1993. Effects of occupational exposure to mercury and lead on brainstem auditory evoked potentials. *Int J Psychophysiol* 14(1):21-25.
- \*DOE. 1993. Department of Energy. Code of Federal Regulations. 10 CFR 20.
- \*DOE. 1994a. Department of Energy. Code of Federal Regulations. 10 CFR 30.
- \*DOE. 1994b. Department of Energy. Code of Federal Regulations. 10 CFR 40.
- \*DOE. 1996a. Department of Energy. Code of Federal Regulations. 10 CFR 71.

## 8. REFERENCES

- \*DOE. 1996b. Department of Energy. Code of Federal Regulations. 10 CFR 110.
- \*DOI. 1989. Mercury. In: Minerals yearbook. Washington, DC: U.S. Department of the Interior, Bureau of Mines, 705-708.
- \*Dolar SG, Kenney DR, Chesters G. 1971. Mercury accumulation by *Myriophyllum spicatum* L. *Environ Lett* 69:191-198.
- \*DOT. 1989a. Hazardous materials table. U.S. Department of Transportation. Federal Register 54(185):9501-39505.
- \*DOT. 1989b. Hazardous materials table. U.S. Department of Transportation. Code of Federal Regulations. 49 CFR 172.101.
- \*Drake HJ. 1981. Mercury. In: Mark HF, Othmer DF, Overberger CG, et al. eds. Kirk-Othmer encyclopedia of chemical technology. New York, NY: John Wiley and Sons, Inc., 143-156.
- \*Drasch G, Wanghofer E, Roider G. 1997. Are blood, urine, hair, and muscle valid biomarkers for the internal burden of men with the heavy metals mercury, lead and cadmium? an investigation on 150 deceased. *Trace Elements and Electrocytes* 14(3):116-123.
- \*Druet E, Sapin C, Gunther E, et al. 1977. Mercuric chloride-induced anti-glomerular basement membrane antibodies in the rat: Genetic control. *Eur J Immunol* 7:348-351.
- \*Druet P, Druet E, Potdevin F, et al. 1978. Immune type glomerulonephritis induced by HgCl<sub>2</sub> in the Brown Norway rat. *Ann Immunol* 129C:777-792.
- \*Dubey C, Bellon B, Hirsch F, et al. 1991a. Increased expression of class II major histocompatibility complex molecules on B cells in rats susceptible or resistant to mercury chloride-induced autoimmunity. *Clin Exp Immunol* 86(1):118-123.
- \*Dubey C, Bellon B, Kuhn J, et al. 1991b. Increase of a IA expression on B cells during the course of mercury-induced autoimmune disease in Brown Norway rats. In: Bach PH, et al., eds. Nephrotoxicity: mechanisms, early diagnosis, and therapeutic management. Fourth International Symposium on Nephrotoxicity. New York, NY: Marcel Dekker, Inc., 397-400.
- \*Duhr EF, Pendergrass JC, Slevin JT, et al. 1993. HgEDTA complex inhibits GTP interactions with the e-site of brain beta-tubulin. *Toxicol Appl Pharmacol* 122(2):273-280.
- \*Dunn JD, Clarkson TW. 1980. Does mercury exhalation signal demethylation of methylmercury? *Health Phys* 38:411-414.
- \*Dunn JD, Clarkson TW, Magos L. 1981a. Ethanol reveals novel mercury detoxification step in tissues. *Science* 213:1123-1125.
- \*Dunn JD, Clarkson TW, Magos L. 1981b. Interaction of ethanol and inorganic mercury: Generation of mercury vapor *in vivo*. *J Pharmacol Exp Ther* 216:19-23.

## 8. REFERENCES

- \*Dutczak WJ, Clarkson TW, Ballatori N. 1991. Biliary-hepatic recycling of a xenobiotic gallbladder absorption of methyl mercury. *Am J Physiol* 260(6):G873-G880.
- \*Dvonch JT, Vette AF, Keeler GJ. 1995. An intensive multi-site pilot study investigating atmospheric mercury in Broward County, Florida. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994*. Boston, MA: Kluwer Academic Publishers, 169-178.
- \*Dyall-Smith DJ, Scurry JP. 1990. Mercury pigmentation and high mercury levels from the use of a cosmetic cream. *Med J Aust* 153(7):409-410, 441-415.
- \*Eaton, AD, Clesceri, LS, Greenberg, AE. 1995. Sections 3112 (Metals by cold-vapor atomic absorption spectrometry), 3113 (metals by electrothermal atomic absorption spectrometry) and 3500 (mercury). In: *Standard methods for the examination of water and wastewater, 19th Edition*. American Public Health Association, American Water Works Association, Water Environment Federation, Washington, DC.
- \*Ebert ES, Price PS, Keenan RE. 1996. Estimating exposures to dioxin-like compounds for subsistence anglers in North America. *Organohalogen Compounds* 30:169-173.
- \*Echeverria D, Heyer NJ, Martin MD, et al. 1995. Behavioral effects of low-level exposure to elemental Hg among dentists. *Neurotoxicol Teratol* 1995 17(2):161-8.
- \*Egeland GM, LA Feyk, JP Middaugh. 1998. The use of traditional foods in a healthy diet in Alaska. Alaska Division of Public Health, Anchorage, Alaska.
- \*Egeland GM, Middaugh JP. 1997. Balancing fish consumption benefits with mercury exposure. *Science* 278(5345):1904-5.
- \*Ehrenberg RL, Vogt RL, Smith AB, et al. 1991. Effects of elemental mercury exposure at a thermometer plant. *Am J Ind Med* 19(4):495-507.
- \*Eichholz GG, Petelka MF, Kury RL. 1988. Migration of elemental mercury through soil from simulated burial sites. *Water Res* 22(1):15-20.
- \*Eide R, Wesenberg GR. 1993. Mercury contents of indicators and target organs in rats after long-term, low-level, mercury vapor exposure. *Environ Res* (2):212-222.
- \*Eldefrawi ME, Mansour N, Eldefrawi A. 1977. Interactions of acetylcholine receptors with organic mercury compounds: Membrane toxicity. *Proceedings of the 9th Annual Rochester International Conference on Environmental Toxicity*. *Adv Exp Med Biol* 84:449-463.
- Eley BM. 1997. The future of dental amalgam: A review of the literature. part 2: Mercury exposure in dental practice. *Br Dent J* 182(8):293-7.
- Eley BM, Cox SW. 1993. The release, absorption and possible health effects of mercury from dental amalgam: a review of recent findings. *Brit Dent J* 175:161-168.
- \*Ellenhorn MJ, Barceloux DG. 1988. *Medical toxicology: Diagnosis and treatment of human poisoning*. New York, NY: Elsevier, 1048-1052.
- \*Ellingsen DG, Andersen A, Nordgagen NP, et al. 1993. Incidence of cancer and mortality among workers exposed to mercury vapour in the Norwegian chloralkali industry. *Brit J Ind Med* 50:875-880.

## 8. REFERENCES

- \*Ellingsen DG, Gaarder PI, Kjuus H. 1994. An immunological study of chloralkali workers previously exposed to mercury vapour. *Apmis* 102(3):170-176.
- \*Ellingsen DG, Nordhagen HP, Thomassen Y. 1995. Urinary selenium excretion in workers with low exposure to mercury vapour. *J Appl Toxicol* 15(1):33-36.
- \*Elsner J. 1991. Tactile-kinesthetic system of rats as an animal model for minimal brain dysfunction. *Arch Toxicol* 65(6):465-473.
- \*Emteborg H, Bulska E, Frech W, et al. 1992. Determination of total mercury in human whole blood by electrothermal atomic absorption spectrometry following extraction. *J Anal Atomic Spectrom* 7(2):405-408.
- \*Endo T, Nakaya S, Kimura R. 1989. Factors involved in absorption of organic mercuric compounds from rat small intestine: Comparative study with mercuric chloride in situ. *Pharmacol Toxicol* 65(2):128-135
- \*Endo T, Nakaya S, Kimura R. 1990. Mechanisms of absorption of inorganic mercury from rat small intestine: III. Comparative absorption studies of inorganic mercuric compounds *in vitro*. *Pharmacol Toxicol* 66(5):347-353.
- \*Enestrom S, Hultman P. 1995. Does amalgam affects the immune system: a controversial issue. *Int Arch Allergy Immunol* 106:180-203.
- \*Engle JH, Ferracane JL, Wichmann J, et al. 1992. Quantitation of total mercury vapor released during dental procedures. *Dent Mater* 8(3):176-180.
- \*Engleson G, Herner T. 1952. Alkyl mercury poisoning. *Acta Paediat Scand* 41:289-294.
- \*EPA. 1971a. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 61.01.
- EPA. 1971b. List of hazardous air pollutants. U. S. Environmental Protection Agency. Federal Regulations 36(62):5931.
- \*EPA. 1971c. Water quality criteria data book. Washington, DC: U. S. Environmental Protection Agency.
- \*EPA. 1975a. Emission standards. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 61.52.
- \*EPA. 1975b. National emission standards for hazardous air pollutants, emissions standard. U. S. Environmental Protection Agency. Federal Register 40:48302.
- \*EPA. 1975c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 61, Subpart E.
- \*EPA. 1978a. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 116.4.
- \*EPA. 1978b. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 227.6.



## 8. REFERENCES

- \*EPA. 1980a. Ambient water quality criteria for mercury. Washington, DC: U. S. Environmental Protection Agency, Office of Water Regulations and Standards. Document no. EPA 440/5-80-058.
- EPA. 1980b. Guidelines and methodology used in the preparation of health effect assessment chapters of the consent decree water criteria documents. U. S. Environmental Protection Agency. Federal Register 45:79347.
- EPA. 1980c. Identification and listing of hazardous waste: Appendix VIII. Hazardous constituents. U. S. Environmental Protection Agency. Federal Register 45:33133.
- EPA. 1980d. Identification and listing of hazardous waste: Characteristic of EP toxicity, U.S. Environmental Protection Agency. Federal Register 45:33122.
- \*EPA. 1980e. Identification and listing of hazardous waste, discarded commercial chemical products, off-specification species, container residues, and spill residues thereof, U. S. Environmental Protection Agency. Federal Register 45:33125-33126.
- \*EPA. 1983. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 122, App. D, Table III.
- \*EPA. 1984a. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 125.
- \*EPA. 1984b. Mercury health effects updates: Health issue assessment. Final report. Washington, DC: U.S. Environmental Protection Agency, Office of Health and Environmental Assessment. Document no. EPA 600/8-84-019F.
- \*EPA. 1985a. Designation, reportable quantities and notification: Designation of hazardous substances. U. S. Environmental Protection Agency. Federal Register 50:13490.
- \*EPA. 1985b. Drinking water criteria document for mercury. Washington, DC: U.S. Environmental Protection Agency, Office of Drinking Water.
- \*EPA. 1985c. Health advisory on mercury-final draft. Washington, DC: U. S. Environmental Protection Agency. Office of Drinking Water. ECAO-CIN-025.
- \*EPA. 1986. Test methods for evaluating solid wastewater and wastes: Laboratory manual of physical/chemical methods. Washington, DC: U.S. Environmental Protection Agency, Office of Solid Waste and Emergency Response. Document no. SW-846.
- \*EPA. 1987a. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 372.65.
- \*EPA. 1987b. Emergency planning and notification: The list of extremely hazardous substances and their threshold planning quantities. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 355.
- EPA. 1987c. Health advisory for mercury. Washington, DC: U.S. Environmental Protection Agency, Office of Drinking Water.

## 8. REFERENCES

- EPA. 1987d. List (phase 1) of hazardous constituents for ground-water monitoring. U.S. Environmental Protection Agency. Federal Register 52(131): 25942-25953.
- EPA. 1987e. Maximum concentration of constituents for ground-water protection. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 264.94.
- \*EPA. 1987f. Toxic air pollution/source crosswalk--a screening tool for locating possible sources emitting toxic air pollutants. Research Triangle Park, NC: U.S. Environmental Protection Agency, Office of Air and Radiation, Office of Air Quality Planning and Standards. Document no. EPA 450/4-87-023a.
- \*EPA. 1988. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 148.12.
- \*EPA. 1990a. Interim methods for development of inhalation reference concentrations. U.S. Environmental Protection Agency. EPA/600/8-90/066A.
- \*EPA. 1990b. Pesticide products containing phenylmercuric acetate; Receipts of requests for voluntary cancellation. U.S. Environmental Protection Agency. Fed Reg 55(177):37541-37542.
- \*EPA. 1990c. Standards of performance for volatile organic compounds (VOC) emissions from synthetic organic chemical manufacturing industry (SOCMI) distillation operation. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 60.667.
- \*EPA. 1991a. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 257, App. I.
- \*EPA. 1991b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 258.40.
- \*EPA. 1991c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 265.
- \*EPA. 1991d. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 266.
- \*EPA. 1991e. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 266, App. IX.
- \*EPA. 1992a. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 131.36.
- \*EPA. 1992b. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 141, Subpart F.
- \*EPA. 1992c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261.3.
- \*EPA. 1992d. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261.32.
- \*EPA. 1992e. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 268.45.
- \*EPA. 1992f. National study of chemical residues in fish. Volume I. Environmental Protection Agency.
- \*EPA. 1993a. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261.24.

## 8. REFERENCES

- \*EPA. 1993b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 266, App. VII.
- \*EPA. 1994a. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 63, Subpart D.
- \*EPA. 1994b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261.33.
- \*EPA. 1994c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261., App. VIII.
- \*EPA. 1994d. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 266, App. XIII.
- \*EPA. 1994e. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 268.42.
- \*EPA. 1994f. Method 7470A. Mercury in Liquid Waste (Manual Cold-VaporTechnique) Test Methods for Evaluating Solid Waste. Office of Solid Waste, U. S. Environmental Protection Agency.
- \*EPA. 1994g. Method 7471A. Mercury in Solid or Semisolid Waste (Manual Cold-Vapor Technique) Test Methods for Evaluating Solid Waste. Office of Solid Waste, U. S. Environmental Protection Agency.
- \*EPA. 1995a. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 60, Subpart Cb
- \*EPA. 1995b. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 60, Subpart Eb.
- \*EPA. 1995c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 117.3.
- \*EPA. 1995d. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 132.
- \*EPA. 1995e. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 192.04 and App. I.
- \*EPA. 1995f. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 264.94.
- \*EPA. 1995g. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 261, App. VII.
- \*EPA. 1995h. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 264, App. IX.
- \*EPA. 1995i. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 302.4.
- \*EPA. 1995j. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 355, App. A.

## 8. REFERENCES

- \*EPA. 1995k. Guidance for assessing chemical contaminant data for use in fish advisories. Volume 1: Fish sampling and analysis. Second Edition. Office of Science and Technology Office of Water, U.S. Environmental Protection Agency.
- \*EPA . 1996a. Mercury study report to Congress - Volume I: Executive summary. U.S. Environmental Protection Agency. EPA-452/R-96-001a.
- \*EPA. 1996b. Mercury study report to Congress Volume II: An inventory of anthropogenic mercury emissions in the United States. U.S. Environmental Protection Agency. EPA452/R-96-001b.
- \*EPA. 1996c. Mercury study report to Congress Volume III: An assessment of exposure from anthropogenic mercury emissions in the United States. U.S. Environmental Protection Agency. EPA452/R-96-001c.
- \*EPA. 1996d. Mercury study report to Congress Volume IV: Health effects of mercury and mercury compounds. U.S. Environmental Protection Agency. EPA452/R-96-001d.
- \*EPA. 1996e. Mercury study report to Congress Volume VI: Characterization of human health and wildlife risks from anthropogenic mercury emissions in the United States. U.S. Environmental Protection Agency. EPA452/R-96-001f.
- \*EPA. 1996f. Toxic chemical release inventory reporting - form R and instructions. U.S. Environmental Protection Agency. 745-K 96 001.
- \*EPA. 1996g. Drinking Water Regulations and Health Advisories. U.S. Environmental Protection Agency. October 1996.
- \*EPA. 1996h. U. S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 51.166.
- \*EPA. 1996i. National listing of fish and wildlife consumption advisories for mercury. U. S. Environmental Protection Agency.
- \*EPA. 1997a. Locating and estimating air emissions from sources of mercury and mercury compounds. Office of Air Quality Planning and Standards and Office of Air and Radiation. U.S. Environmental Protection Agency Research Triangle Park, NC. EPA-454/R-97-012.
- \*EPA. 1997b. The national survey of mercury concentrations in fish database survey 1990-1995. Draft Report. September 1997. Prepared for the US Environmental Protection Agency. Standard and Applied Sciences Division under EPA contract no. 68-C40051 (Tables only).
- \*EPA. 1997c. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 268.40. (62 FR 7502).
- \*EPA. 1998a. Fact sheet update: Listing of fish and wildlife advisories. Office of Water. Washington, D.C. EPA823-F-98-009.
- \*EPA. 1998b. Listing of fish and wildlife advisories - 1997. U.S. Environmental Protection Agency Office of Water. Washington, DC.

## 8. REFERENCES

- \*Erfurth EM, Schutz A, Nilsson A, et al. 1990. Normal pituitary hormone response to thyrotropin and gonadotropin releasing hormones in subjects exposed to elemental mercury vapour. *Brit J Ind Med* 47:639-644.
- \*Ernst E, Lauritsen JG. 1991. Effect of organic and inorganic mercury on human sperm motility. *Pharmacol Toxicol* 69(6):440-444.
- \*Espinoza EO, Mann M-J, Bleasdel B. 1996. Toxic metals in selected traditional Chinese medicinals. *J Forensic Sciences* 41(3):453-456.
- \*Espinoza EO, Mann MJ, Bleasdel B. 1995. Arsenic and mercury in traditional Chinese herbal balls. *The New England Journal of Medicine* 333(12):803-804.
- \*Evans HL, Garman R, Weiss B. 1977. Methylmercury: Exposure duration and regional distribution as determinants of neurotoxicity in nonhuman primates. *Toxicol Appl Pharmacol* 41:15-33.
- \*Evans MS, Noguchi GE, Rice CP. 1991. The biomagnification of polychlorinated biphenyls, toxaphene, and DDT compounds in a Lake Michigan offshore food web. *Arch Environ Contam Toxicol* 20:87-93.
- \*Evans O, McKee GD. 1988. Determination of mercury(II) and organomercury compounds by reversed-phase liquid chromatography with reductive electrochemical detection. *Analyst* 113(2):243-246.
- \*Facemire C, Augspurger T, Bateman D, et al. 1995. Impacts of mercury contamination in the southeastern United States. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994.* Boston, MA: Kluwer Academic Publishers, 923-926.
- \*Fagala GE, Wigg CL. 1992. Psychiatric manifestations of mercury poisoning. *J Am Acad Child Adolesc Psychiatry* 31(2):306-311.
- \*Fairey R, Taberski K, Lamerdin S, et al. 1997. Organochlorines and other environmental contaminants in muscle tissues of sportfish collected from San Francisco Bay. *Marine Pollution Bulletin* 34(12):1058-1071.
- \*Falk SA, Klein R, Haseman JK, et al. 1974. Acute methylmercury intoxication and ototoxicity in guinea pigs. *Arch Pathol Lab Med* 97:297-305.
- \*Faria A, Freitas CD. 1992. Systemic contact dermatitis due to mercury. *Contact Dermatitis* 27(2):110-111.
- \*Farris FF, Dedrick RL, Allen PV, et al. 1993. Physiological model for the pharmacokinetics of methyl mercury in the growing rat. *Toxicol Appl Pharmacol* 119:74-90.
- \*Fawer RF, DeRibaupierre Y, Guillemin M, et al. 1983. Measurement of hand tremor induced by industrial exposure to metallic mercury. *Br J Ind Med* 40:204-208.
- \*FDA. 1974. Food and Drug Administration. Department of Health and Human Services. Code of Federal Regulations. 21 CFR 700.13
- FDA. 1980. Processed grain. Compliance Policy Guides. U.S. Food and Drug Administration. 7104.05.

## 8. REFERENCES

- FDA. 1982a. Bottled water: Quality standards. Food and Drug Administration. Federal Register 46(157):41037.
- FDA. 1982b. Quality standards for food with no identity standards: Bottled water. Food and Drug Administration. Code of Federal Regulation. 21 CFR 103.35.
- FDA. 1984. Fish and seafood. Compliance Policy Guides. U.S. Food and Drug Administration. 7108.07.
- FDA. 1989. Quality standards for food with no identity standards: Bottled water. Food and Drug Administration. Code of Federal Regulations. 21 CFR 103.35.
- \*FDA. 1994. Action levels for poisonous or deleterious substances in human food and animal feed. Action levels for poisonous or deleterious substances in human food and animal feed.
- \*FDA. 1995. Food and Drug Administration. Department of Health and Human Services. Code of Federal Regulations. 21 CFR 165.
- \*FDA. 1998. Mercury in fish: cause for concern? U.S. Food and Drug Administration. FDA Consumer (September 1994) updated 2/26/96. <http://vm.cfsan.fda.gov/~dms/mercury.html> .
- \*FEDRIP. 1998. FEDRIP Literature Search (References and Abstracts) for Mercury. Federal Research in Progress. Dialog Information Service.
- \*Fehling C, Abdulla M, Brun A, et al. 1975. Methylmercury poisoning in the rat: A combined neurological, chemical, and histopathological study. *Toxicol Appl Pharmacol* 33:27-37.
- \*Filippelli M. 1987. Determination of trace amounts of organic and inorganic mercury in biological materials by graphite furnace atomic absorption spectrometry and organic mercury speciation by gas chromatography. *Anal Chem* 59:116-118.
- \*Fiore BJ, Anderson HA, Hanrahan LP, et al. 1989. Sport fish consumption and body burden levels of chlorinated hydrocarbons: A study of Wisconsin anglers. *Arch Environ Health* 44 (2) 82-88.
- \*Fishbein L. 1991. Indoor environments: The role of metals. In: Merian E, ed. *Metals and their compounds in the environment*. Weinheim, Fed Rep Germany: VCH 287-309.
- \*Fiskesjo G. 1979. Two organic mercury compounds tested for mutagenicity in mammalian cells by use of the cell line V 79-4. *Hereditas* 90:103-110.
- Fitzgerald WF. 1979. Distribution of mercury in natural waters. In: Nriagu JO, ed. *The biogeochemistry of mercury in the environment*. New York, NY: Elsevier/North Holland Biomedical Press, 161-174.
- \*Fitzgerald WF, Mason RP, Vandal GM. 1991. Atmospheric cycling and air-water exchange of mercury over midcontinental lacustrine regions. *Water Air Soil Pollut* 56:745-767.
- \*Fitzhugh OG, Nelson AA, Laug EP, et al. 1950. Chronic oral toxicities of mercuric-phenyl and mercuric salts. *Arch Ind Hyg Occup Med* 2:433-442.

## 8. REFERENCES

- \*Fleming LE, Watkins S, Kaderman, R et al. 1995. Mercury exposure in humans through food consumption from the Everglades in Florida. *Water Air Soil Pollut* 80:41-48.
- \*Florentine MJ, Sanfilippo DJ. 1991. Elemental mercury poisoning. *Clin Pharm* 10:213-221.
- \*Foman SJ. 1966. Body composition of the infant (Part I: The male reference infant). In: Falkner F, editor. *Human Development*. Philadelphia, PA: WB Saunders, pp. 239-246.
- \*Foman, SJ, Haschke, F, Ziegler, EE, and Nelson, SE. 1982. Body composition of reference children from birth to age 10 years. *Amer J Clin Nutr* 35:1169-1175.
- \*Foulds D, Copeland K, Franks R. 1987. Mercury poisoning and acrodynia. *Am J Dis Children* 141:124-125.
- \*Foulkes EC, Bergman D. 1993. Inorganic mercury absorption in mature and immature rat jejunum: Transcellular and intercellular pathways *in vivo* and in everted sacs. *Toxicol Appl Pharmacol* 120:89-95.
- \*Fowler BA. 1972. Ultrastructural evidence for neuropathy induced by long-term exposure to small amounts of methylmercury. *Science* 175:780-781.
- \*Fowler BA, Woods JS. 1977. The transplacental toxicity of methylmercury to fetal rat liver mitochondria. *Lab Invest* 36:122-130.
- \*Fowler SW. 1990. Critical review of selected heavy metal and chlorinated hydrocarbon concentrations in marine environment. *Mar Environ Res* 29:1-64.
- \*Franchi E, Loprieno G, Ballardini M, et al. 1994. Cytogenetic monitoring of fishermen with environmental mercury exposure. *Mutat Res* 320:23-29.
- \*Fredriksson A, Dahlgren L, Danielsson B, et al. 1992. Behavioral effects of neonatal metallic mercury exposure in rats. *Toxicology* 74(2-3):151-160.
- \*Fredriksson A, Dencker L, Archer T, et al. 1996. Prenatal coexposure to metallic mercury vapour and methylmercury produce interactive behavioural changes in adult rats. *Neurotoxicol Teratol* 18(2):129-134.
- \*Friberg L, Hammarstrom S, Nystrom A. 1953. Kidney injury after chronic exposure to inorganic mercury. *Arch Ind Hyg Occup Med* 8:149-153.
- \*Friberg L, Nordberg F. 1973. Inorganic mercury-a toxicological and epidemiological appraisal. In: Miller MW, Clarkson TW, eds. *Mercury, mercurials and mercaptans*. Springfield, IL: Charles C. Thomas, 5-22.
- \*Friberg L, Vostal J, eds. 1972. *Mercury in the environment: A toxicological and epidemiological appraisal*. Cleveland, OH: CRC Press.
- \*Friese KH, Roschig M, Wuenschel G, et al. 1990. A new calibration method for the determination of trace amounts of mercury in air and biological materials. *Fresenius J Anal Chem* 337(8):860-866.

## 8. REFERENCES

- \*FSTRAC. 1995. Summary of state and federal drinking water standards and guidelines. U.S. Environmental Protection Agency. Chemical Communications Subcommittee, Federal State Toxicology and Regulatory Alliance Committee.
- \*Fujita M, Takabatake E. 1977. Mercury levels in human maternal and neonatal blood, hair and milk. *Bull Environ Contam Toxicol* 18(2):205-207.
- \*Fukino H, Hirai M, Ideura K, et al. 1992. Effect of the administration of mercuric chloride on zinc deficiency in rats. *J Food Hyg Soc Jpn* 33(1):31-38.
- \*Fukuda K. 1971. Metallic mercury induced tremor in rabbits and mercury content of the central nervous system. *Br J Ind Med* 28:308-311.
- \*Fung YK, Andrew GM, Rack P, et al. 1995. Determination of blood mercury concentrations in Alzheimer's patients. *Clin Toxicol* 33(3):243-247.
- \*Futatsuka M, Kitano T, Nagano M, et al. 1992. An epidemiological study with risk analysis of liver diseases in the general population living in a methyl mercury polluted area. *J Epidemiol Community Health* 46:237-240.
- \*Fuyuta M, Fujimoto T, Hirata S. 1978. Embryotoxic effects of methylmercuric chloride administered to mice and rats during organogenesis. *Teratology* 18:353-366.
- \*Fuyuta M, Fujimoto T, Kiyofuji E. 1979. Teratogenic effects of a single oral administration of methylmercuric chloride in mice. *Acta Anat* 104:356-362.
- \*Gage JC. 1961. The distribution and excretion of inhaled mercury vapour. *Br J Ind Med* 18:287-294.
- \*Gage JC. 1964. Distribution and excretion of methyl and diphenyl mercury salts. *Br J Ind Med* 21:197-202.
- \*Gage JC. 1973. The metabolism of methoxyethylmercury and phenylmercury in the rat. In: Miller MW, Clarkson TW, eds. *Mercury, mercurials and mercaptans*. Springfield, IL: Charles C Thomas, 346-354.
- \*Galal-Gorchev H. 1993. Dietary intake, levels in food and estimated intake of lead, cadmium, and mercury. *Food Additives and Contaminants* 10(1):115-128.
- \*Gale T, Ferm V. 1971. Embryopathic effects of mercuric salts. *Life Sci* 10:1341-1347.
- \*Gale TF. 1974. Embryopathic effects of different routes of administration of mercuric acetate on the hamster. *Environ Res* 8:207-213.
- \*Gallagher PJ, Lee RL. 1980. Role of biotransformation in organic mercury neurotoxicity. *Toxicol* 15:129-134.
- \*Galster WA. 1976. Mercury in Alaskan Eskimo mothers and infants. *Environ Health Perspectives* 15:135-140.



## 8. REFERENCES

- \*Ganser AL, Kirschner DA. 1985. The interaction of mercurials with myelin: Comparison of *in vitro* and *in vivo* effects. *Neurotoxicol* 6:63-78.
- \*Ganther HE. 1980. Interactions of vitamin E and selenium with mercury and silver. *Acad Sci* 355:212-226.
- \*Ganther HS, Goudie C, Sunde ML, et al. 1972. Selenium: Relation to decreased toxicity of methylmercury added to diets containing tuna. *Science* 175:1122-1124.
- \*Garnier R, Fuster J, Conso F, et al. 1981. Acute mercury vapor poisoning. *Toxicol Environ Res* 3:77-86. (French)
- \*Gavis J, Ferguson JF. 1972. The cycling of mercury through the environment. *Water Res* 6:986-1008.
- \*Gear CW. 1971. Numerical initial value problems in ordinary differential equations. Englewood Cliffs, NJ: Prentice-Hall.
- \*Gearhart JM, Clewell HJ III, Crump KS et al. 1995. Pharmacokinetic dose estimates of mercury in children and dose-response curves of performance tests in a large epidemiological study. *Water Air Soil Pollut* 80:49-58.
- \*Geffner ME and Sandler A. 1980. Oral metallic mercury: A folk medicine for gastroenteritis. *Clin Pediatr* 435-437.
- \*Gentry PR, Gearhart JM, Allen BC et al. 1998. Investigation of the potential impact of benchmark dose and pharmacokinetic modeling in noncancer risk assessment, II. Investigation of impact on MRLs for methylmercury, manganese, cadmium, perchloroethylene, chloroform, and metallic mercury vapor. ICF Kaiser Report to ATSDR. KS Crump Group, ICF Kaiser, Ruston, LA. September, 1998.
- \*George L, Scott FE, Cole D, et al. 1996. The mercury emergency and Hamilton school children: A follow-up analysis. *Can J Public Health*. 4:224-6.
- \*Gerhardsson L, Brune DK. 1989. Mercury in dentistry. In: Brune DK, Edling C, eds. Occupational hazards in the health professions. Boca Raton, FL: CRC Press, Inc., 307-321.
- \*Germani MS, Zoller WH. 1988. Vapor-phase concentrations of arsenic, selenium bromine, iodine and mercury in the stack of a coal-fired power plant. *Environ Sci Technol* 22(9):1079-1085.
- \*Ghosh AK, Sen S, Sharma A, et al. 1991. Effect of chlorophyllin on mercuric chloride-induced clastogenicity in mice. *Food Chem Toxicol* 29(11):777-779.
- \*Ghosh N, Bhattacharya S. 1992. Thyrotoxicity of the chlorides of cadmium and mercury in rabbit. *Biomed Environ Sci* 5(3):236-240.
- \*Gilbert SG, Rice DC, Burbacher TM. 1996. Fixed interval/fixed ratio performance in adult monkeys exposed *in utero* to methylmercury. *Neurotoxicol Teratol* 18(5):539-46.
- \*Gill GA, Bruland KW. 1990. Mercury speciation in surface freshwater systems in California and other areas. *Environ Sci Technol* 24(9):1392-1400.

## 8. REFERENCES

- \*Gill GA, Guentzel JL, Landing WM, et al. 1995. Total gaseous mercury measurements in Florida: the FAMS project (1992-1994). In: Porcella DB, Wheatley B, eds. Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994. Boston, MA: Kluwer Academic Publishers, 235-244.
- Gilmour CC, Bloom NS. 1995. A case study of mercury and methylmercury dynamics in a Hg-contaminated municipal wastewater treatment plant. In: Porcella DB, Huckabee JW, eds. Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994. Boston, MA: Kluwer Academic Publishers, 799-803.
- \*Gilmour CC, Henry EA. 1991. Mercury methylation in aquatic systems affected by acid deposition. *Environmental Pollution* 71(2-4):131-169.
- \*Girard M, Dumont C. 1995. Exposure of James Bay Cree to methylmercury during pregnancy for the years 1983-91. In: Porcella DB, Wheatley B, eds. Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994. Boston, MA: Kluwer Academic Publishers, 13-19.
- \*Girardi G, Elias MM. 1991. Effectiveness of n-acetylcysteine in protecting against mercuric chloride-induced nephrotoxicity. *Toxicol* 67(2):155-164.
- \*Glass GE, Sorensen JA, Schmidt KW, et al. 1990. New source identification of mercury contamination in the Great Lakes. *Environ Sci Technol* 24(7):1059-1069.
- \*Glass GE, Sorenson JA, Schmidt KW, et al. 1991. Mercury deposition and sources for the upper Great-Lakes region. *Water Air Soil Pollut* 56:235-249.
- \*Glasser H, Chang DPY, Hickman DC. 1991. An analysis of biomedical waste incineration. *J Air Waste Manag Assoc* 41:1180-1188.
- \*Gleason MN, Gosselin RE, Hodge HC. 1957. *Clinical toxicology of commercial products*. Baltimore, MD: Williams and Wilkins Co, 154.
- \*Goering PL, Fisher BR, Chaudhary PP, et al. 1992. Relationship between stress protein induction in rat kidney by mercuric chloride and nephrotoxicity. *Toxicol Appl Pharmacol* 113(2):184-191.
- \*Goh CL, Ng SK. 1988. Occupational allergic contact dermatitis from metallic mercury. *Contact Dermatitis* 19(3):232-233.
- \*Goldfrank LR, Bresnitz EA, Howland MA, et al. 1990. Mercury. In: Goldfrank LR, Flomenbaum NE, Lewin NA, et al., eds. *Goldfrank's toxicologic emergencies*, 4th ed. Norwalk, CT: Appleton and Lange, 641-648.
- \*Goldin A, Bigelow C, Veneman PLM. 1992. Concentrations of metals in ash from municipal solid waste combustors. *Chemosphere* 24:271-280.
- \*Goldman M, Blackburn P. 1979. The effect of mercuric chloride on thyroid function in the rat. *Toxicol Appl Pharmacol* 48:49-55.

## 8. REFERENCES

- \*Goldman M, Druet P, Gleichmann E. 1991. Th2 cells in systemic autoimmunity: Insights from allogenic diseases and chemically-induced autoimmunity. *Immunol Today* 12:223-227.
- \*Goldwater LJ, Stopford W. 1977. Mercury. In: Lenihan J, Fletcher WW, eds. *Environment and man*. Vol. 6. New York, NY: Academic Press, 38-63.
- \*Golimowski J, Gustavsson I. 1983. Determination of mercury in freeze-dried muscle samples of pike, cod, and perch using an ASV-technique. *Sci Total Environ* 31:89-98.
- \*Gore I, Harding SM. 1987. Sinker lung: Acute metallic mercury poisoning associated with the making of fishing weights. *Ala J Med Sci* 24:267-269.
- \*Gossel TA, Bricker JD. 1984. *Principles of clinical toxicology*. New York, NY: Raven Press, 175-187.
- \*Gotelli CA, Astolfi E, Cox C, et al. 1985. Early biochemical effects of an organic mercury fungicide on infants: "Dose makes the poison." *Science* 277:638-640.
- \*Goto M, Kumagai S, Ishii D. 1988. Continuous micro flow analysis system for monitoring total mercury at sub-ppb level in waste water. *Anal Sci* 4(1):87-90.
- \*Grabo TN. 1997. Unknown toxic exposures - arts and crafts materials. *AAOHN (American Association of Occupational Health Nurses) Journal* 45(3):124-130.
- \*Granato TC, Pietz RI, Gschwind J, et al. 1995. Mercury in soils and crops from fields receiving high cumulative sewage sludge applications: Validation of U.S. EPA's risk assessment for human ingestion. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994*. Boston, MA: Kluwer Academic Publishers, 1119-1127.
- \*Grandjean P, Guldager B, Larsen IB, et al. 1997a. Placebo response in environmental disease. Chelation therapy of patients with symptoms attributed to amalgam fillings. *J Occup Environ Med* 39(8):707-14.
- \*Grandjean P, Weihe J, Needham LL, et al. 1995a. Relation of a seafood diet to mercury, selenium, arsenic, and polychlorinated biphenyl and other organochlorine concentrations in human milk. *Environmental Research* 71/1:29-38.
- \*Grandjean P, Weihe J, Needham LL, et al. 1996. Relation of a seafood diet to mercury, selenium, arsenic, and polychlorinated biphenyl and other organochlorine concentrations in human milk. *Environ Res* 71/1:29-38.
- \*Grandjean P, Weihe P, Jorgensen PJ, et al. 1992. Impact of maternal seafood diet on fetal exposure to mercury, selenium, and lead. *Arch Environ Health* 47(3):185-195.
- \*Grandjean P, Weihe P, Nielsen JB. 1994. Methylmercury: significance of intrauterine and postnatal exposures. [review] [38 refs]. *Clinical Chemistry* 40(7 Pt 2):1395-400.
- \*Grandjean P, Weihe P, White RF. 1995b. Milestone development in infants exposed to methylmercury from human milk. *Neurotoxicology* 16(1):27-33.

## 8. REFERENCES

- \*Grandjean P, Weihe P, White RF, et al. 1997b. Cognitive deficit in 7-year-old children with prenatal exposure to methylmercury. *Neurotoxicol Teratol* 19(6):417-428.
- \*Grandjean P, Weihe P, White RF, et al. 1998. Cognitive performance of children prenatally exposed to "safe" levels of methylmercury. *Environ Res* 77(2):165-172.
- \*Grant-Webster K, Burbacher T, Mottet NK. 1992. Puberal growth retardation in primates: A latent effect of *in utero* exposure to methylmercury. *Toxicol* 12:310.
- \*Gray DG. 1995. A physiologically based pharmacokinetic model for methyl mercury in the pregnant rat and fetus. *Toxicol Appl Pharmacol* 132:91-102.
- \*Grayson M, ed. 1983. Kirk-Othmer encyclopedia of chemical technology; 3rd ed. New York, NY: John Wiley & Sons, 145.
- \*Greenwood MR, Clarkson TW, Magos L. 1972. Transfer of metallic mercury into the fetus. *Experientia* 28:1455-1456.
- \*Grieb TM, Driscoll CT, Gloss SP, et al. 1990. Factors affecting mercury accumulation in fish in the upper Michigan peninsula. *Environ Toxicol Chem* 9:919-930.
- \*Griem P, Scholz E, Turfeld M, et al. 1997. Strain differences in tissue concentrations of mercury in inbred mice treated with mercuric chloride. *Toxicol Appl Pharmacol* 144:163-170.
- \*Grupp DJ, Everitt DA, Bath JB, et al. 1989. Use of a transportable XRF spectrometer for on-site analysis of mercury in soils. *Am Environ Lab* 1(2):32-40.
- \*Gstraunthaler G, Pfaller W, Kotanko P. 1983. Glutathione depletion and *in vitro* lipid peroxidation in mercury or maleate-induced acute renal failure. *Biochem Pharmacol* 32:2969-2972.
- \*Guentzel JL, Landing WM, Gill GA, et al. 1995. Atmospheric deposition of mercury in Florida: the FAMS project (1992-1994). In: Porcella DB, Huckabee JW, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994.* Boston, MA: Kluwer Academic Publishers, 393-402.
- \*Guidetti P, Giacobazzi A, Zanoli P, et al. 1992. Prenatal exposure of rats to methylmercury: Increased sensitivity of the GABA-benzodiazepine receptor functions. *Metal compounds in environment and life: Interrelation between chemistry and biology*, 4:365-371.
- \*Gunderson EL. 1988. FDA total diet study, April 1982-April 1984, dietary intakes of pesticides, selected elements, and other chemicals. *J Assoc Off Anal Chem* 71(6):1200-1209.
- \*Gunderson VM, Grant-Webster KS, Burbacher TM, et al. 1988. Visual recognition memory deficits in methylmercury-exposed *Macaca fascicularis* infants. *Neurotoxicol Teratol* 10(4):373-379.
- \*Gutenmann WH, Ebel JG Jr, Kuntz HT, et al. 1992. Residues of p,p'-DDE and mercury in lake trout as a function of age. *Arch Environ Contam Toxicol* 22:452-455.

## 8. REFERENCES

- \*Gutenmann WH, Lisk DJ. 1991. Higher average mercury concentration in fish fillets after skinning and fat removal. *J Food Safety* 11(2):99-103.
- \*Guzelian PS, Henry CJ, Olin SS. 1992. Similarities and differences between children and adults: Implications for risk assessment. International Life Sciences Institute Press, Washington, D.C.
- \*Hac E, Krechniak J. 1993. Mercury concentrations in hair exposed *in vitro* to mercury vapor. *Biol Trace Element Res* 39(2-3):109-115.
- \*Haddad J, Stenberg E. 1963. Bronchitis due to acute mercury inhalation: Report of two cases. *Am Rev Respir Dis* 88:543-545.
- \*Haddad LM, Winchester JF. 1990. Clinical management: Poisoning and drug overdose. Second edition. Philadelphia, PA: W.B. Saunders, Co., 1005-1009
- \*Halbach S. 1994. Amalgam tooth fillings and man's mercury burden. *Human Exper Toxicol* 13:496-501.
- \*Halbach S. 1995. Mercury exposure from dental amalgam fillings. Institute of Toxicology, GSF-Research Center for Environment and Health, D-85758-Oberschleissheim.
- \*Halbach S, Clarkson TW. 1978. Enzymatic oxidation of mercury vapor by erythrocytes. *Biochem Biophys Acta* 523:522-531.
- \*Hall BD, Bodaly RA, Fudge RJP, et al. 1997. Food as the dominant pathway of methylmercury uptake by fish. *Water Air Soil Pollut* 100(1-2):13-24.
- \*Hallee TJ. 1969. Diffuse lung disease caused by inhalation of mercury vapor. *Am Rev Respir Dis* 99:430-436.
- \*Handley J, Todd D, Burrows D. 1993. Mercury allergy in a contact dermatitis clinic in Northern Ireland. *Contact Dermatitis* 29:258-261.
- \*Hanninen H. 1982. Behavior effects of occupational exposure to mercury and lead. *Acta Neurol Scand* 66:167-175.
- \*Hansen JC. 1988. Has selenium a beneficial role in human exposure to inorganic mercury? *Med Hypotheses* 25(1):45-53.
- \*Hansen JC. 1991. Mercury and selenium concentrations in Greenlandic mother-infant blood samples. In: Dillon HK, Ho MJ, eds. *Biological monitoring of exposure to chemicals: Metals*. New York, NY: John Wiley and Sons, 11-25.
- \*Hansen JC, Danscher G. 1995. Quantitative and qualitative distribution of mercury in organs from arctic sledgedogs: An atomic absorption spectrophotometric and histochemical study of tissue samples from natural long-termed high dietary organic mercury-exposed dogs from Thule, Greenland. *Pharmacol Toxicol* 77(3):189-195.
- \*Hapke HJ. 1991. Metals accumulation in the food chain and load of feed and food. In: Merian E, ed. *Metals and their compounds in the environment*. Weinheim, Fed Rep Germany: VCH, 469-479.

## 8. REFERENCES

- \*Harada H. 1978. Congenital Minamata disease: Intrauterine methylmercury poisoning. *Teratology* 18:285-288.
- \*Haraldsson C, Westerlund C, Ohman P. 1989. Determination of mercury in natural samples at the sub-nanogram level using inductively coupled plasma/mass spectrometry after reduction to elemental mercury. *Anal Chim Acta* 221(1):77-84.
- \*Harding L, Goyette D. 1989. Metals in northeast Pacific coastal sediments and fish, shrimp, and prawn tissues. *Mar Pollut Bull* 13:217-218.
- \*Harnly M, Seidel S, Rojas P, et al. 1997. Biological monitoring for mercury within a community with soil and fish contamination. *Environ Health Perspect* 105(4):424-9.
- \*Harrison SE, Klaverkamp JF. 1990. Metal contamination in liver and muscle of northern pike (*Esox lucius*) and white sucker (*Catostomus commersoni*) from lakes near the smelter at Flin Flon, Michigan. *Environ Toxicol Chem* 9:941-956.
- \*Harsh JB, Doner HE. 1981. Characterization of mercury in a river wash soil. *J Environ Qual* 10(3):333-337.
- \*Hartman, DE. 1995. *Neuropsychological toxicology: Identification and assessment of human neurotoxic syndromes* (second edition). New York, NY: Plenum Press, 132-133.
- Harvery T, Mahaffey KR, Velazquez S. 1995. Holistic risk assessment: An emerging process for environmental decisions. *Reg Toxicol Pharmacol* 22:110-117.
- Hattis D, Erdreich L, Ballew M. 1987. Human variability in susceptibility to toxic chemicals - a preliminary analysis of pharmacokinetic data from normal volunteers. *Risk Analysis* 7:415-426.
- \*Haxton J, Lindsay DG, Hilsop JS, et al. 1979. Duplicate diet study on fishing communities in the United Kingdom: Mercury exposure in a "critical group." *Environ Res* 18:351-368.
- \*Hayes LC, Rodenbeck SE. 1992. Developing a public-health assessment: Impact of a mercury-contaminated discharge to surface-water. *J Environ Health* 55(2):16-18.
- \*HazDat. 1998. Database. Agency for Toxic Substances and Disease Registry (ATSDR), Atlanta, GA.
- \*Health Canada. 1997. Health Canada Position Statement on Dental Amalgam. Sept 15, 1997. [Http://www.hc-sc.gc.ca/main/drugs/zmfiles/english/issues/amalgam\\_position.html](http://www.hc-sc.gc.ca/main/drugs/zmfiles/english/issues/amalgam_position.html).
- \*Hefflin BJ, Etzel RA, Agocs, MM, et al. 1993. Mercury exposure from exterior latex paint. *Appl Occup Environ Hyg* 8:886-870.
- \*Hellou J, Fancey LL, Payne JF. 1992. Concentrations of twenty-four elements in bluefin tuna, *Thunnus thynnus*, from the Northwest Atlantic. *Chemosphere* 24(2):211-218.
- \*Henderson R, Shotwell HP, Krause LA. 1974. Analyses for total, ionic and elemental mercury in urine as a basis for biological standard. *Am Ind Hyg Assoc J* 38:576-580.

## 8. REFERENCES

- \*Hill W. 1943. A report on two deaths from exposure to the fumes of a di-ethyl mercury. *Can J Pub Health* 34:158-160.
- \*Hirano M, Mitsumori K, Maita K, et al. 1986. Further carcinogenicity study on methylmercury chloride in ICR mice. *Jap J Vet Sci* 48(1):127-135.
- \*Hirsch F, Couderc J, Sapin C, et al. 1982. Polyclonal effect of HgCl<sub>2</sub> in the rat, its possible role in an experimental autoimmune disease. *Eur J Immunol* 12:620-625.
- \*Hirsch F, Kuhn J, Ventura M, et al. 1986. Autoimmunity induced by HgCl<sub>2</sub> in Brown-Norway rats--part I. Production of monoclonal antibodies. *J Immunol* 136:3272-3276.
- \*Hirszel P, Michaelson JH, Dodge K, et al. 1985. Mercury-induced autoimmune glomerulonephritis in inbred rats--part II. Immunohistopathology, histopathology and effects of prostaglandin administration. *Surv Synth Pathol Res* 4:412-422.
- \*Hoet P, Lison D. 1997. A nonoccupational source of mercury intoxication (2). *Clinical Chemistry* 43(7):1248.
- \*Hoff RM, Strachan WM, Sweet CW, et al. 1996. Atmospheric deposition of toxic chemicals to the Great Lakes: A review of data through 1994. *Atmospheric Environment* 30(20):3505-3527.
- \*Hook O, Lundgren K-D, Swensson A. 1954. On alkyl mercury poisoning. *Acta Med Scand* 150:131-137.
- \*Horvat M. 1996. Mercury analysis and speciation in environmental samples. In: Baeyens PM, Ebinghaus WR, Vasiliev O, eds. 1-31. *Nato Asi Series*
- \*Horvat M, Byrne AR. 1992. Preliminary study of the effects of some physical parameters on the stability of methylmercury in biological samples. *Analyst* 117:665-668.
- \*Horvat M, Lupsina V. 1991. Determination of total mercury in coal fly ash by gold amalgamation cold vapour atomic-absorption spectrometry. *Anal Chim Acta* 243(1):71-79.
- \*Horvat M, Stegnar A, Byrne R, et al. 1988. A study of trace elements in human placenta, blood, and hair from the Yugoslav central Adriatic. In: Braetter P, Schramel P, eds. *Trace elements-analytical chemistry in medicine and biology*. Berlin: W. de Guyter and Co., 243-250.
- \*Houeto P, Sandoukm P, Baud EF, et al. 1994. Elemental mercury vapour toxicity: Treatment and levels in plasma and urine. *Human Exper Toxicol* 13:848-852.
- \*Howard W, Leonard B, Moody W, et al. 1991. Induction of chromosome changes by metal compounds in cultured CHO cells. *Toxicol Lett* 56(1-2):179-186.
- \*HSDB. 1997. Hazardous Substances Data Bank. National Library of Medicine, National Toxicology Program (via TOXNET), Bethesda, MD. May 9, 1997.
- \*Hudson PJ, Vogt RL, Brondum J, et al. 1987. Elemental mercury exposure among children of thermometer plant workers. *Pediatrics* 79:935-938.

## 8. REFERENCES

- \*Hueter RE, Fong WG, Henderson G, et al. 1995. Methylmercury concentration in shark muscle by species, size and distribution of sharks in Florida coastal waters. In: Porcella DB, Wheatley B, eds. Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994. Boston, MA: Kluwer Academic Publishers, 893-899.
- Huggins HA. 1983. Mercury - a factor in mental disease? *Oral Health* 73:42-45.
- \*Hughes JA, Annau Z. 1976. Postnatal behavioral effects in mice after prenatal exposure to methylmercury. *Pharmacol Biochem Behav* 4:385-391.
- \*Hughes WL. 1957. A physicochemical rationale for the biological activity of mercury and its compounds. *Ann NY Acad Sci* 65:454-460.
- \*Hultman P, Bell LJ, Enestrom S, et al. 1992. Murine susceptibility to mercury: I. Autoantibody profiles and systemic immune deposits in inbred, congenic, and intra-H-2 recombinant strains. *Clin Immunol Immunopathol* 65(2):98-109.
- \*Hultman P, Enestrom S. 1988. Mercury induced antinuclear antibodies in mice: Characterization and correlation with renal immune complex deposits. *Clin Exp Immunol* 71:269-274.
- \*Hultman P, Enestrom S. 1992. Dose-response studies in murine mercury-induced autoimmunity and immune-complex disease. *Toxicol Appl Pharmacol* 113(2):199-208.
- \*Hultman P, Johansson U. 1991. Strain differences in the effect of mercury on murine cell-mediated immune reactions. *Food Chem Toxicol* 29(9):633-638.
- \*Hultman P, Johansson U, Turley SJ, et al. 1994. Adverse immunological effects and autoimmunity induced by dental amalgam and alloy in mice. *FASEB J* 8(14):1183-1190.
- \*Humes HD, Weinberg JM. 1983. Cellular energetics in acute renal failure. In: Brenner BM, Lazarus JM, eds. Acute renal failure. Philadelphia, PA: W.B. Saunders, 47-98.
- \*Hunter D, Bomford RR, Russell DS. 1940. Poisoning by methyl mercury compounds. *Quart J Med* 9:193-213.
- \*Hurley JP, Watras CJ, Bloom NS. 1991. Mercury cycling in a northern Wisconsin seepage lake - the role of particulate matter in vertical transport. *Water Air Soil Pollut* 56:543-551.
- \*Hursh JB, Clarkson TW, Cherian MG, et al. 1976. Clearance of mercury (Hg-197, Hg-203) vapor inhaled by human subjects. *Arch Environ Health* 31:302-309.
- \*Hursh JB, Clarkson TW, Miles EF, et al. 1989. Percutaneous absorption of mercury vapor by man. *Arch Environ Health* 44:120-127.
- \*Hursh JD, Greenwood MR, Clarkson TW, et al. 1980. The effect of ethanol on the fate of mercury vapor inhaled by man. *J Pharmacol Exp Ther* 214:520-527.
- \*Husztai Z, Balogh I. 1995. Effects of lead and mercury on histamine uptake by glial and endothelial cells. *Pharmacol Toxicol* 76:339-342.



## 8. REFERENCES

- \*IARC 1993. Beryllium, cadmium, mercury, and exposures in the glass manufacturing industry: Evaluation of carcinogenic risks to humans. International Agency For Research On Cancer. Vol 58.
- \*Icard Ph, Pelletier L, Vial M-C, et al. 1993. Evidence for a role of antilaminin-producing B cell clones that escape tolerance in the pathogenesis of HgCl<sub>2</sub>-induced membranous. *Nephrol Dial Transplant* 8:122-127.
- \*Ichinose N, Miyazawa Y. 1989. Simplification of the thermal decomposition process of silver amalgam during the determination of total mercury in tissue samples by flameless atomic absorption. *Fresenius Z Anal Chem* 334(8):740-742.
- \*Ikingura JR, Akagi H. 1996. Monitoring of fish and human exposure to mercury due to gold mining in the Lake Victoria goldfields, Tanzania. *Science of the Total Environment* 191:59-68
- \*Ilback NG. 1991. Effects of methyl mercury exposure on spleen and blood natural-killer (NK) cell-activity in the mouse. *Toxicology* 67(1):117-124.
- \*Ilback NG, Sundberg J, Oskarsson A. 1991. Methyl mercury exposure via placenta and milk impairs natural killer (NK) cell function in newborn rats. *Toxicol Lett* 58(2):149-158.
- \*Imura N, Naganuma A. 1991. Possible mechanism of detoxifying effect of selenium on the toxicity of mercury compounds. In: Suzuki T, Imura N, Clarkson TW, eds. *Advances in mercury toxicology*. New York, NY: Plenum Press, 275-288.
- \*Inasmasu T, Ogo A, Yanagawa M, et al. 1986. Mercury concentration change in human hair after the ingestion of canned tuna fish. *Bull Environ Contam Toxicol* 37:475-481
- \*Inouye M, Kajiwara Y. 1988. Developmental disturbances of the fetal brain in guinea-pigs caused by methylmercury. *Arch Toxicol* 62(1):15-21.
- \*Inouye M, Kajiwara Y. 1990. Placental transfer of methylmercury and mercuric mercury in mice. *Environ Med* 34:169-172.
- \*Inouye M, Murakami U. 1975. Teratogenic effect of orally administered methylmercuric chloride in rats and mice. *Congenital Anomalies* 15:1-9.
- \*Inouye M, Murao K, Kajiwara Y. 1985. Behavioral and neuropathological effects of prenatal methylmercury exposure in mice. *Neurobehav Toxicol Teratol* 7:227-232.
- \*IRIS. 1997. Integrated Risk Information System (IRIS). Online. U.S. Environmental Protection Agency, Office of Health and Environmental Assessment, Environmental Criteria and Assessment Office, Cincinnati, OH May 1997.
- \*Itawi RK, Subramaniam S, Turec. 1990. Determination of selenium, cadmium and mercury in an aquatic environment of Bombay by sub-stoichiometric neutron-activation analysis. *J Radioanal Nucl Chem* 138(1):63-66.
- \*Iverfeldt A, Lindqvist O. 1984. The transfer of mercury at the air/water interface. In: Brutsaert W, and Jirka GH, eds. *Gas transfer at water surfaces*. Reidel Dordrecht.

## 8. REFERENCES

- Iverfeldt A, Lindqvist O. 1986. Atmospheric oxidation of elemental mercury by ozone in the aqueous phase. *Atmos Environ* 20(8):1567-1573.
- Iyer K, Goldgood J, Eberstein A, et al. 1976. Mercury poisoning in a dentist. *Arch Neurol* 33:788-790.
- \*Jackson TA. 1991. Biological and environmental control of mercury accumulation by fish in lakes and reservoirs of northern Manitoba, Canada. *Can J Fish Aquat Sci* 48(12):2449-2470.
- \*Jacobs JM, Carmichael N, Cavanagh JB. 1977. Ultrastructural changes in the nervous system of rabbits poisoned with methylmercury. *Toxicol Appl Pharmacol* 39:249-261.
- \*Jaffe KM, Shurtleff DB, Robertson WO. 1983. Survival after acute mercury vapor poisoning--role of intensive supportive care. *Am J Dis Child* 137:749-751.
- \*Jagiello G, Lin JS. 1973. An assessment of the effects of mercury on the meiosis of mouse ova. *Mutat Res* 17:93-99.
- \*Jalili HA, Abbasi AH. 1961. Poisoning by ethyl mercury toluene sulphonanilide. *Br J Ind Med* 18:303-308.
- \*Janicki K, Dobrowolski J, Drasnicky K. 1987. Correlation between contamination of the rural environment with mercury and occurrence of leukemia in men and cattle. *Chemosphere* 16:253-257.
- \*Jasinski SM. 1993. Mercury. In: Mineral commodity summaries, 1993. Washington, D.C.: US Dept of the Interior, Bureau of Mines, 110-111.
- Jensen AA. 1983. Chemical contaminants in human milk. In: Gunther FA, Gunther JD, eds. *Residue Reviews: Residues of pesticides and other contaminants in the total environment*. Vol. 89. 1-128.
- \*Jenssen O, Ramel C. 1980. The micronucleus test as part of a short-term mutagenicity test program for the prediction of carcinogenicity evaluated by 143 agents tested. *Mutat Res* 75:191-202.
- \*Jiang GB, Ni ZM, Wang SR, et al. 1989. Organic mercury speciation in fish by capillary gas chromatography interfaced with atomic absorption spectrometry. *Fresenius Z Anal Chem* 334(1):27-30.
- \*Jiang Y, Moller G. 1995. *In vitro* effects of HgCl<sub>2</sub> on murine lymphocytes. *J Immunol* 154(7):3138-3146.
- \*Johanson CE. 1980. Permeability and vascularity of the developing brain: Cerebellum vs cerebral cortex. *Brain Research* 190:3-16.
- \*Johnson A, Norton D, Yake B, et al. 1990. Transboundary metal pollution of the Columbia River (Franklin D. Roosevelt Lake). *Bull Environ Contam Toxicol* 45:703-710.
- \*Johnson C. 1999. Elemental mercury use in religious and ethnic practices in Latin American and Caribbean communities in New York City. *Population and Environment* (in press).
- \*Johnson DC, Bramen RS. 1974. Distribution of atmospheric mercury species near the ground. *Environ Sci Technol* 8:1003-1009.

## 8. REFERENCES

- \*Jokstad A, Thomassen Y, Bye E, et al. 1992. Dental amalgam and mercury. *Pharmacol Toxicol (Copenhagen)* 70(4):308-313.
- \*Jones MM. 1991. New developments in therapeutic chelating agents as antidotes for metal poisoning. *CRC Crit Rev Toxicol* 21(3):209-233.
- \*Jonker D, Jones MA, van Bladeren PJ, et al. 1993a. Acute (24 hr) toxicity of a combination of four nephrotoxins in rats compared with the toxicity of the individual compounds. *Food Chemical Toxicol* 31(1):45-52.
- \*Jonker D, Woutersen RA, van Bladeren PJ, et al. 1993b. Subacute (4-wk) oral toxicity of a combination of four nephrotoxins in rats: Comparison with the toxicity of the individual compounds. *Food Chem Toxicol* 31(2):125-136.
- Joselow M, Goldwater L, Alvarez A, et al. 1968a. Absorption and excretion of mercury in man - XV. Occupational exposure among dentists. *Arch Environ Health* 17:39-43.
- \*Joselow MM, Ruiz R, Goldwater L. 1968b. Absorption and excretion of mercury in man: XIV. Salivary excretion of mercury and its relationship to blood and urine. *Arch Environ Health* 17:35-38.
- \*Jugo S. 1976. Retention and distribution of  $^{203}\text{HgCl}_2$  in suckling and adult rats. *Health Phys* 30:240-241.
- \*Kabuto M. 1987. Acute endocrine effects of a single administration of methylmercury chloride (MMC) in rats. *Endocrinol Jpn* 33(5): 683-690.
- \*Kabuto M. 1991. Chronic effects of methylmercury on the urinary excretion of catecholamines and their responses to hypoglycemic stress. *Arch Toxicol* 65(2):164-167.
- \*Kajiwara Y, Inouye M. 1986. Effects of methylmercury and mercuric chloride on preimplantation mouse embryos *in vivo*. *Teratology* 33:231-237.
- \*Kajiwara Y, Inouye M. 1992. Inhibition of implantation caused by methylmercury and mercuric chloride in mouse embryos *in vivo*. *Bull Environ Contam Toxicol* 49(4):541-546.
- \*Kalac P, Burda J, Staskova I. 1991. Concentrations of lead, cadmium, mercury, and copper in mushrooms in the vicinity of a lead smelter. *Sci Tot Environ* 105:109-119.
- \*Kalamegham R, Ash KO. 1992. A simple ICP-MS procedure for the determination of total mercury in whole blood and urine. *J Clin Lab Anal* 6(4):190-193.
- \*Kanematsu N, Hara M, Kada T. 1980. REC assay and mutagenicity studies on metal compounds. *Mutat Res* 77:109-116.
- \*Kanerva L, Komulainen M, Estlander T, et al. 1993. Occupational allergic contact dermatitis from mercury. *Contact Dermatitis* 28(1):26-28.
- \*Kang-Yum E, Oransky SH. 1992. Chinese patent medicine as a potential source of mercury poisoning. *Vet Hum Toxicol* 34(3):235-238.

## 8. REFERENCES

- \*Kanluen S, Gottlieb CA. 1991. A clinical pathologic study of four adult cases of acute mercury inhalation toxicity. *Arch Pathol Lab Med* 115(1):56-60.
- \*Kargacin B, Kostial K. 1991. Methods for decreasing mercury-203 retention in relation to age and route of exposure. In: Suzuki T, Imura N, Clarkson TW, eds. *Advances in mercury toxicology*. New York, NY: Plenum Press, 135-153.
- \*Karpathios T, Zervoudakis A, Thodoridis C, et al. 1991. Mercury vapor poisoning associated with hyperthyroidism in a child. *Acta Paediatr Scand* 80(5):551-552.
- \*Kawahara D, Oshima H, Kosugi H, et al. 1993. Further epidemiologic-study of occupational contact-dermatitis in the dental clinic. *Contact Dermatitis* 28(2):114-115.
- \*Kazantzis G. 1981. Role of cobalt, iron, lead, manganese, mercury, platinum, selenium, and titanium in carcinogenesis. *Environ Health Perspect* 40:143-161.
- \*Kazantzis G, Schiller K, Asscher A, et al. 1962. Albuminuria and the nephrotic syndrome following exposure to mercury and its compounds. *Q J Med* 3:403-419.
- \*Keeler G, Glinsorn G, Pirrone N. 1995. Particulate mercury in the atmosphere: Its significance, transport transformation and sources. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994*. Boston, MA: Kluwer Academic Publishers, 159-168.
- \*Keeler GJ, Hoyner ME, Lamborg CH. 1994. Measurements of atmospheric mercury in the Great Lakes basin. In: Watras CJ, Huckabee JW, eds. *Mercury pollution integration and synthesis*. Boca Raton, Florida: Lewis Publishers, 231-241.
- \*Kelly TJ, Czuczwa JM, Sticksel PR, et al. 1991. Atmospheric and tributary inputs of toxic substances to Lake Erie. *J Gt Lakes Res* 17(4):504-516.
- \*Kerper LE, Ballatori N, Clarkson TW. 1992. Methylmercury transport across the blood-brain barrier by an amino acid carrier. *Am J Physiol* 262(5):R761-R765.
- \*Kerry A, Welbourn PM, Prucha B, et al. 1991. Mercury methylation by sulphate-reducing bacteria from sediments of an acid stressed lake. *Water Air Soil Pollut* 56:565-575.
- \*Kershaw TG, Clarkson TW, Dhahir PH. 1980. The relationship between blood levels and dose of methylmercury in man. *Arch Environ Health* 35:28-36.
- \*Khayat A, Dencker L. 1982. Fetal uptake and distribution of metallic mercury vapor in the mouse - influence of ethanol and aminotriazole. *Int J Biol Res Pregnancy* 3:38-46.
- \*Khera KS, Iversin F, Hierlihy L, et al. 1974. Toxicity of methylmercury in neonatal cats. *Teratology* 10:69-76.
- \*Khera KS, Tabacova SA. 1973. Effects of methylmercuric chloride on the progeny of mice and rats treated before or during gestation. *Food Cosmet Toxicol* 11:245-254.

## 8. REFERENCES

- \*King G. 1954. Acute pneumonitis due to accidental exposure to mercury vapor. *Ariz Med* 11:335.
- \*Kirschbaum BB, Sprinkle FM, Oken DE. 1980. Renal function and mercury level in rats with mercuric chloride nephrotoxicity. *Nephron* 26:28-34.
- \*Kirschner DS, Billau RL, MacDonald TJ. 1988. Fluorescent light tube compaction: Evaluation of employee exposure to airborne mercury. *Appl Ind Hyg* 3(4):129-131.
- \*Kishi R, Hashimoto K, Shimizu S, et al. 1978. Behavioral changes and mercury concentrations in tissues of rats exposed to mercury vapor. *Toxicol Appl Pharmacol* 46:555-566.
- \*Kitagawa K, Nishimoto N. 1989. Thermal vaporizer - capacitively coupled microwave plasma system for trace mercury determination. *J Spectrosc Soc Japan* 38(4):282-287.
- \*Klein R, Herman SP, Bullock BC, et al. 1973. Methylmercury intoxication in rat kidneys. *Arch Pathol Lab Med* 96:83-90.
- \*Knox RC, Canter LW. 1996. Prioritization of ground water contaminants and sources. *Water Air and Soil Pollution* 88(3-4):205-226.
- \*Kohler CC, Heidinger RC, Call T. 1990. Levels of PCBs and trace metals in crab orchard lake sediment, benthos, zooplankton and fish. Report no. HWRICRR-043. Carbandale, IL: Fishery Research Laboratory, University of South Illinois.
- \*Komori M, Nishio K, Kitada M et al. 1990. Fetus-specific expression of a form of cytochrome P-450 in human livers. *Biochemistry* 29:4430-4433.
- \*Kostial K, Kello D, Jugo S, et al. 1978. Influence of age on metal metabolism and toxicity. *Environ Health Perspect* 25:81-86.
- \*Kosuda LL, Greiner DL, Bigazzi PE. 1993. Mercury-induced renal autoimmunity: changes in rt6+ T-lymphocytes of susceptible and resistant rats. *Environ Health Perspectives* 101(2):178-185.
- Kosuda LL, Hosseinzadeh H, Greiner DL, et al. 1994. Role of rt6+ T lymphocytes in mercury-induced renal autoimmunity: experimental manipulations of "susceptible" and "resistant" rats. *J Toxicol Environ Health* 42(3):303-321.
- \*Krabbenhoft DP, Babiarz CL. 1992. The role of groundwater transport in aquatic mercury cycling. *Water Resour Res* 28(12):3119-3128.
- \*Krishnan K, Andersen ME. 1994. Physiologically-based pharmacokinetic modeling in toxicology. In: Wallace Hayes, ed. *Principles and Methods of Toxicology*. 3rd edition. New York, NY:Raven Press Ltd.
- \*Krishnan K, Andersen ME, Clewell HJ III, et al. 1994. Physiologically-based pharmacokinetic modeling of chemical mixtures. In: RSA Yang, ed. *Toxicology of chemical mixtures*. New York, NY: Academic Press.
- \*Kuhnert PM, Kuhnert BR, Erhard P. 1981. Comparison of mercury levels in maternal blood, fetal cord blood and placental tissues. *Am J Obstet Gynecol* 133:209-213.

## 8. REFERENCES

- \*Kuntz WD, Pitkin RM, Bostrom AW, et al. 1982. Maternal and cord blood background levels: A longitudinal surveillance. *Am J Obstet Gynecol* 143:440-443.
- \*Kutsuna M, ed. 1968. Minamata disease: Study group of Minamata disease. Japan: Kumamoto University, 1-4.
- Ladd AC, Zuskin E, Valic R, et al. 1966. Absorption and excretion of mercury in miners. *J Occup Med* 8:122-131.
- Lamm O, Pratt H. 1985. Subclinical effects of exposure to inorganic mercury revealed by somatosensory-evoked potentials. *Eur Neurol* 24:237-243.
- \*Lamperti AA, Printz RH. 1973. Effects of mercuric chloride on the reproductive cycle of the female hamster. *Biology of Reproduction* 8:378-387.
- \*Landi S, Fagioli F, Locatelli C, et al. 1990. Digestion method for the determination of mercury in vegetable matrices by cold vapour atomic-absorption spectrometry. *Analyst* 115(2):173-177.
- \*Langauer-Lewowicka H, Kazibutowska Z. 1989. Multimodality evoked potentials in occupational exposure to metallic mercury vapour. *Pol J Occup Med* 2(2):192-199.
- \*Langlois C, Langis R, Perusse M, et al. 1995. Mercury contamination in northern Quebec environment and wildlife. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant*. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994. Boston, MA: Kluwer Academic Publishers, 1021-1024.
- \*Langolf GD, Chaffin DB, Henderson R, et al. 1978. Evaluation of workers exposed to elemental mercury using quantitative tests. *Am Ind Hyg Assoc J* 39:976-984.
- \*Langworth S, Almkvist O, Soderman E, et al. 1992a. Effects of occupational exposure to mercury vapour on the central nervous system. *Br J Ind Med* 49(8):545-555.
- \*Langworth S, Elinder CG, Sundquist KG, et al. 1992b. Renal and immunological effects of occupational exposure to inorganic mercury. *Br J Ind Med* 49(6):394-401.
- \*Larsson KS. 1995. The dissemination of false data through inadequate citation. *J Int Med* 238:445-450.
- \*Lasora BK, Citternam. 1991. Segmental analysis of mercury in hair in 80 women of Nome, Alaska. OCS Study. MMS 91-0065 NTIS from US Department of Interior Mineral Management Service, Alaska OCS Region.
- \*Laug EP, Kunze FM. 1949. The absorption of phenylmercuric acetate from the vaginal tract of the rat. *J Pharmacol Exp Ther* 95:460-464.
- \*Lauwerys R, Bernard A, Roels H, et al. 1983. Anti-laminin antibodies in workers exposed to mercury vapor. *Toxicol Lett* 17:113-116.
- \*Lauwerys R, Bonnier C, Evard P, et al. 1987. Prenatal and early postnatal intoxication by inorganic mercury resulting from the maternal use of mercury containing soap. *Hum Toxicol* 6:257-260.

## 8. REFERENCES

- \*Lauwerys R, Roels H, Buchet JP, et al. 1977. Non-job related increased urinary excretion of mercury. *Int Arch Occup Environ Health* 39:33-36.
- \*Lauwerys R, Roels H, Genet P, et al. 1985. Fertility of male workers exposed to mercury vapor or to manganese dust: A questionnaire study. *Am J Ind Med* 7:171-176.
- \*LeBel CP, Ali SF, Bondy SC. 1992. Deferoxamine inhibits methyl mercury-induced increases in reactive oxygen species formation in rat brain. *Toxicol Appl Pharmacol* 112(1):161-165.
- \*LeBel CP, Ali SF, McKee M, et al. 1990. Organometal-induced increases in oxygen reactive species: The potential of 2',7'-dichlorofluorescein diacetate as an index of neurotoxic damage. *Toxicol Appl Pharmacol* 104: 17-34.
- \*Lebel J, Mergler D, Lucotte M et al. 1996. Evidence of early nervous system dysfunction in amazonian populations exposed to low-levels of methylmercury. *Neurotoxicol* 17(1):157-168.
- \*LeBihan A, Cabon JY. 1990. Determination of one nanogram per litre levels of mercury in water by electrothermal atomization atomic-absorption spectrometry after solvent extraction. *Talanta* 37(12):1119-1122.
- \*Lecavalier PR, Chu I, Villeneuve D, et al. 1994. Combined effects of mercury and hexachlorobenzene in the rat. *Journal of Environmental Science and Health - Part B: Pesticides, Food Contaminants and Agricultural Wastes* 29(5):951-961.
- \*Lee IP, Dixon RL. 1975. Effects of mercury on spermatogenesis studies by velocity sedimentation cell separation and serial mating. *J Pharmacol Exp Ther* 194:171-181.
- \*Lee JH, Han DH. 1995. Maternal and fetal toxicity of methylmercuric chloride administered to pregnant Fischer 344 rats. *J Toxicol Environ Health* 45(4):415-425.
- \*Lee M, Chan KK-S, Sairenji E, et al. 1979. Effect of sodium selenite on methylmercury-induced cleft palate in the mouse. *Environ Res* 19:39-48.
- \*Lee SH, Jung KH, Lee DH. 1989. Determination of mercury in environmental samples by cold-vapour generation and atomic-absorption spectrometry with a gold-coated graphite furnace. *Talanta* 36(10):999-1003.
- \*Lee YH, Iverfeldt A. 1991. Measurement of methylmercury and mercury in run-off, lake and rain waters. *Water Air Soil Pollut* 56:309-321.
- \*Leeder JS, Kearns GL. 1997. Pharmacogenetics in pediatrics: Implications for practice. *Pediatric Clinics of North America* 44: 55-77.
- \*Lehotzky K, Meszaros I. 1974. Alteration of electroencephalogram and evoked potential in rats induced by organic mercury. *Acta Pharmacol Toxicol* 35:180-184.
- \*Lemus R, Abdelghani AA, Akers TG, et al. 1996. Health risks from exposure to metals in household dusts. *Reviews on Environmental Health* 11(4):179-189.

## 8. REFERENCES

- \*Leroux BG, Leisenring WM, Moolgavkar SH, et al. 1996. A biologically-based dose-response model for developmental toxicology. *Risk Analysis* 16 (4):449-458.
- \*Leung H. 1993. Physiologically-based pharmacokinetic modeling. In: Ballantine B, Marro T, Turner T, eds. *General and Applied Toxicology*. Vol. 1. New York, NY: Stockton Press, 153-164.
- \*Levine SP, Cavender GD, Langolf GD, et al. 1982. Elemental mercury exposure: Peripheral neurotoxicity. *Br J Ind Med* 39:136-139.
- \*Levins P, Adams J, Brenner P, et al. 1979. Sources of toxic pollutants found in influents to sewage treatment plants--part VI. Integrated interpresentation. Washington, DC: U.S. Environmental Protection Agency. Document no. EPA 440/4-81-008.
- \*Lewis RJ. 1993. *Hawley's condensed chemical dictionary*. Twelfth edition.
- \*Lexa J, Stulik K. 1989. Preparation of a gold electrode modified with trioctylphosphine oxide and its application to determination of mercury in the environment. *Talanta* 36(8):843-848.
- \*Leyshon K, Morgan AJ. 1991. An integrated study of the morphological and gross-elemental consequences of methylmercury intoxication in rats, with particular attention on the cerebellum. *Scanning Microsc* 5(3):895-904.
- \*Liang L, Brooks, RJ, et al. 1995. Mercury reactions in the human mouth with dental amalgams. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994*. Boston, MA: Kluwer Academic Publishers, 103-107.
- \*Lilis R, Miller A, Lerman Y. 1985. Acute mercury poisoning with severe chronic pulmonary manifestations. *Chest* 88:306-309.
- \*Lille F, Hazemann P, Garnier R, et al. 1988. Effects of lead and mercury intoxications on evoked potentials. *Clin Toxicol* 26(1-2):103-116.
- \*Lin JL, Lim PS. 1993. Massive oral ingestion of elemental mercury. *J Toxicol Clin Toxicol* 31 (3):487-492.
- \*Lind B, Friberg L, Nylander M. 1988. Preliminary studies on methylmercury biotransformation and clearance in the brain of primates: II. Demethylation of mercury in brain. *J Trace Elem Exp Med* 1(1):49-56.
- \*Lindberg SE. 1984. Emission and deposition of atmospheric mercury. Oak Ridge, Tennessee: Oak Ridge National Laboratory. DE85 006304, 32 pp.
- \*Lindberg SE, Owens JG, Stratton WJ. 1994. Application of throughfall methods to estimate dry deposition of mercury. In: Watras CJ, Huckabee JW, eds. *Mercury pollution integration and synthesis*. Boca Raton, Florida: Lewis Publishers, 261-271.
- \*Lindberg SE, Turner RR, Meyers TP, et al. 1991. Atmospheric concentrations and deposition of mercury to a deciduous forest at Walker Branch Watershed, Tennessee, USA. *Water Air Soil Pollut* 56:577-594.



## 8. REFERENCES

- \*Lindqvist KJ, MaKene WJ, Shaba JK, et al. 1974. Immunofluorescence and electron microscopic studies of kidney biopsies from patients with nephrotic syndrome, possibly induced by skin lightening creams containing mercury. *East Afr Med J* 51:168-169.
- \*Lindqvist O. 1991a. Mercury in the Swedish environment: 9. Mercury in terrestrial ecosystems bioavailability and effects. *Water Air Soil Pollut* 55(1-2):101-108.
- \*Lindqvist O. 1991b. Mercury in the Swedish environment: 4. Emissions of mercury to the atmosphere. *Water, Air, Soil Pollution* 55(1-2):23-32.
- \*Lindqvist O. 1991c. Mercury in the Swedish environment: 6. Transformation and deposition processes. *Water Air Soil Pollut* 55(1-2):49-64.
- Lindqvist O. 1991d. Mercury in the Swedish environment: 7. Regional and global atmospheric budgets. *Water Air Soil Pollut* 55(1-2):65-72.
- \*Lindqvist O. 1991e. Mercury in the Swedish environment: 8. Mercury in terrestrial systems. *Water, Air, Soil Pollution* 55(1-2):73-100.
- \*Lindqvist O. 1994. Atmospheric cycling of mercury: an overview. In: Watras CJ, Huckabee JW, eds. *Mercury pollution integration and synthesis*. Boca Raton, Florida: Lewis Publishers, 181-185.
- \*Lindstedt G, Gottberg I, Holmgren B, et al. 1979. Individual mercury exposure of chloralkali workers and its relation to blood and urinary mercury levels. *Scand J Work Environ Health* 5:59-69.
- \*Lindstrom H, Luthman J, Oskarsson A, et al. 1991. Effects of long-term treatment with methyl mercury on the developing rat brain. *Environ Res* 56(2):158-169.
- Lipfert F, Dephillips M, Saroff L, et al. 1994. Methylmercury health risks to adults from coal combustion. *Neurotoxicol* 15(4):972.
- \*Lipfert FW, Moskowitz PD, Fthenakis V, et al. 1996. Probabilistic assessment of health risks of methylmercury from burning coal. *Neurotoxicol* 17(1):197-212.
- \*Lisk DJ, Gutenmann WH, Rutzke M, et al. 1992a. Composition of toxicants and other constituents in yard or sludge composts from the same community as a function of time-of-waste-collection. *Arch Environ Contam Toxicol* 22(4):380-383.
- \*Lisk DJ, Gutenmann WH, Rutzke M, et al. 1992b. Survey of toxicants and nutrients in composted waste materials. *Arch Environ Contam Toxicol* 22(2):190-194.
- \*Liu KZ, Wu QG, Liu HI. 1990. Application of a Nafion - Schiff-base modified electrode in anodic-stripping voltammetry for the determination of trace amounts of mercury. *Analyst* 115(6):835-837.
- \*Liu Y, Lopez-Avila V, Alcaraz M. 1994. Simultaneous determination of organotin, organolead, and organomercury compounds in environmental samples using capillary gas chromatography with atomic emission detection. *J High Resolution Chromatography* 17:527-536.

## 8. REFERENCES

- Livardjani F, Heimbürger R, Leroy MJF, et al. 1991a Optimization of blood sample mineralization for mercury analysis by cold vapor atomic absorption. *Analysis* 19(7):205-207
- \*Livardjani F, Ledig M, Kopp P, et al. 1991b. Lung and blood superoxide dismutase activity in mercury vapor exposed rats: Effect of N-acetylcysteine treatment. *Toxicology* 66(3):289-295.
- \*Locket S, Nazroo I. 1952. Eye changes following exposure to metallic mercury. *Lancet* 528-530.
- \*Lodenius M, Autio S. 1989. Effects of acidification on the mobilization of cadmium and mercury from soils. *Arch Environ Contam Toxicol* 18(1-2):261-267.
- \*Lodenius M, Tulisalo E. 1984. Environmental mercury contamination around a chlor-alkali plant. *Bull Environ Contam Toxicol* 32:439-444.
- \*Lopez-Gonzalez MA, Gomez MM, Camara C, et al. 1994. On-line microwave oxidation for the determination of organoarsenic compounds by high-performance liquid chromatography-hydride generation atomic absorption spectrometry. *Journal of Analytical Atomic Spectrometry* 9(3):291-295.
- \*Lorscheider FL, Vimy MJ, Summers AO. 1995. Mercury exposure from "silver" tooth fillings: Emerging evidence questions a traditional dental paradigm. *The FASEB Journal* 9(7):504-508.
- \*Lovejoy HB, Bell ZG, Vizena TR. 1974. Mercury exposure evaluations and their correlation with urine mercury excretion. *J Occup Med* 15:590-591.
- \*Lowe TP, May TW, Brumbaugh WG, et al. 1985. National contaminant biomonitoring program: Concentrations of seven elements in freshwater fish, 1978-1981. *Archive of Environmental Contamination and Toxicology*.
- \*Luecke RH, Wosilait WD, Pearce BA, Young JF. 1997. A computer model and program for xenobiotic disposition during pregnancy. *Comput Methods Programs Biomed* 53(3):201-224.
- \*Lund BO, Miller DM, Woods JS. 1991. Mercury-induced hydrogen peroxide production and lipid peroxidation *in vitro* in rat kidney mitochondria. *Biochem Pharmacol* 42, (Suppl) S181-S187.
- \*Lund BO, Miller DM, Woods JS. 1993. Studies on Hg(II)-induced H<sub>2</sub>O<sub>2</sub> formation and oxidative stress *in vivo* and *in vitro* in rat kidney mitochondria. *Biochem Pharmacol* 45(10):2017-2024.
- \*Lund M, Banner W, Clarkson T, Berlin M. 1984. Treatment of acute methylmercury ingestion by hemodialysis with n-acetylcysteine (Mucomyst) infusion and 2,3-dimercaptopropane sulfonate. *J Toxicol Clin Toxicol* 22(1):31-49.
- \*Lundgren KD, Swensson A. 1949. Occupational poisoning by alkyl mercury compounds. *J Indust Hyg Toxicol* 31:190-200.
- \*Lymberi P, Hirsch F, Kuhn J, et al. 1986. Autoimmunity induced by HgCl<sub>2</sub> in Brown-Norway rats--part II. Monoclonal antibodies sharing specificities and idiotypes with mouse natural monoclonal antibodies. *J Immunol* 136:3277-3281.

## 8. REFERENCES

- \*Lytle TF, Lytle JS. 1990. Heavy metals in the eastern oyster, *Crassostrea virginica*, of the Mississippi sound. *Bull Environ Contam Toxicol* 44:142-148.
- \*Mabille V, Roels H, Jacquet P, et al. 1984. Cytogenetic examination of leukocytes of workers exposed to mercury vapor. *Int Arch Occup Environ Health* 53:257-260.
- \*MacDonald JS, Harbison RD. 1977. Methylmercury-induced encephalopathy in mice. *Toxicol Appl Pharmacol* 39:195-205.
- \*MacIntosh DL, Spengler JD, Ozkaynak H, et al. 1996. Dietary exposures to selected metals and pesticides. *Environ Health Perspect* 104(2):202-209.
- \*Mackert JR. 1987. Factors affecting estimation of dental amalgam mercury exposure from measurements of mercury vapour levels in intra-oral and expired air. *J Dent Res* 66:1775-1780.
- \*Magos L. 1967. Mercury-blood interaction and mercury uptake by the brain after vapor exposure. *Environ Res* 1:323-337.
- \*Magos L, Bakir F, Clarkson TW, et al. 1976. Tissue levels of mercury in autopsy specimens of liver and kidney. *Bull WHO* 53:93-96.
- \*Magos L, Brown AW, Sparrow S, et al. 1985. The comparative toxicology of ethyl and methylmercury. *Arch Toxicol* 57:260-267.
- \*Magos L, Butler WH. 1972. Cumulative effects of methylmercury dicyandiamide given orally to rats. *Food Cosmet Toxicol* 10:513-517.
- \*Magos L, Clarkson TW, Hudson AR. 1989. The effects of dose of elemental mercury and first-pass circulation time on exhalation and organ distribution of inorganic mercury in rats. *Biochim Biophys Acta* 991(1):85-89.
- \*Magos L, Clarkson TW, Sparrow S, et al. 1987. Comparison of the protection given by selenite, selenomethionine and biological selenium against the renotoxicity of mercury. *Arch Toxicol* 60:422-426.
- \*Magos L, Halbach S, Clarkson TW. 1978. Role of catalase in the oxidation of mercury vapor. *Biochem Pharmacol* 27:1373-1377.
- \*Magos L, Peristianis GC, Clarkson TW, et al. 1980. The effect of lactation on methylmercury intoxication. *Arch Toxicol* 45:143-148.
- \*Mahanti HS. 1990. Concentration and spectrochemical determination of trace heavy metals in waste water. *Res Indust* 35:124-126.
- \*Mailhes JB. 1983. Methylmercury effects on Syrian hamster metaphase II oocyte chromosomes. *Environ Mutagen* 5:679-686.
- \*Malm O, Branches FJ, Akagi H, et al. 1995. Mercury and methylmercury in fish and human hair from the Tapajos River Basin, Brazil. *Sci Total Environ* 175(2):141-150.

## 8. REFERENCES

- \*Malt UF, Nerdrum P, Oppedal B, et al. 1997. Physical and mental problems attributed to dental amalgam fillings: A descriptive study of 99 self-referred patients compared with 272 controls. *Psychosom Med* 59(1):32-41.
- \*Mannino S, Granata A, Fregapane G. 1990. Determination of mercury in fish muscle by square-wave voltammetry. *Ital J Food Sci* 2(2):97-101.
- \*Mariani G, Benfenati E, Fanelli R, et al. 1992. Incineration of agro-industrial wastes and macro- and micropollutants emission. *Chemosphere* 24:1545-1551.
- \*Marquez M, Silva M, Perez-Bendito D. 1988. Semi-automatic analysis of mercury in pharmaceuticals by catalytic titration. *J Pharm Biomed Anal* 6(3):307-312.
- \*Marsh DO, Clarkson TW, Cox C, et al. 1987. Fetal methylmercury poisoning: Relationship between concentration in single strands of hair and child effects. *Arch Neurol* 44:1017-1022.
- Marsh DO, Clarkson TW, Myers GJ, et al. 1995a. The Seychelles study of fetal methylmercury exposure and child development: introduction. *Neurotoxicol* 16(4):583-596.
- \*Marsh DO, Myers GJ, Clarkson TW, et al. 1980. Fetal methylmercury poisoning: Clinical and toxicological data on 29 cases. *Ann Neurol* 7:348-355.
- \*Marsh DO, Myers GJ, Clarkson TW, et al. 1981. Dose-response relationship for human fetal exposure to methylmercury. *Clin Toxicol* 10:1311-1318.
- \*Marsh DO, Turner MD, Smith JC, et al. 1995b. Fetal methylmercury study in a Peruvian fish-eating population. *Neurotoxicology* 16(4):717-726.
- \*Mason RP, Reinfelder JR, Morel FMM. 1995. Bioaccumulation of mercury and methylmercury. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994.* Boston, MA: Kluwer Academic Publishers, 915-921.
- \*Mason RP, Reinfelder JR, Morel FMM. 1996. Uptake, toxicity and trophic transfer of mercury in a coastal diatom. *Environ Sci Technol* 30:1835-1845.
- \*Massie HR, Greco ME, Vadlamudi L. 1993. The brain-to-liver mercury ratio increases with aging in mice. *Exper Gerontol* 28(2):161-167.
- \*Mateo MD, Forteza R, Cerda V, et al. 1988. Comparative study of a kinetic - thermometric method and the atomic-absorption cold-vapour technique for determination of mercury traces and ultra-traces. *Thermochimica Acta* 128:21-30.
- \*Mathieson PW. 1992. Mercuric chloride-induced autoimmunity. *Autoimmunity (Switzerland)* 13(3):243-247.
- \*Mathieson PW, Stapleton KJ, Oliveira DBG, et al. 1991. Immunoregulation of mercuric chloride-induced autoimmunity in Brown Norway rats: A role for CD8+ T-cells revealed by *in vivo* depletion studies. *Europ J Immunol* 21(9):2105-2109.

## 8. REFERENCES

- \*Matsumoto H, Koya G, Takeuchi T. 1965. Fetal Minamata disease--a neuropathological study of two cases of intrauterine intoxication by a methylmercury compound. *J Neuropathol Exp Neurol* 24:563-574.
- \*Matsumoto N, Spindle A. 1982. Sensitivity of early mouse embryos to methylmercury toxicity. *Toxicol Appl Pharmacol* 64:108-117.
- \*Matsunaga K, Konishi S, Nishimura M. 1979. Possible errors caused prior to measurement of mercury in natural waters with special reference to seawater. *Environ Sci Technol* 13:63-65.
- \*Matsuo N, Takasugi M, Kuroiwa A, et al. 1989. Thymic and splenic alterations in mercuric chloride-induced glomerulopathy. In: Brown SS, Kodama Y, eds. *Toxicology of metals: Clinical and experimental research*. Chichester: Ellis Horwood Limited, 333-334.
- \*Matthes F, Kirschner R, Yow M, et al. 1958. Acute poisoning associated with inhalation of mercury vapor: Report of four cases. *Pediatrics* 22:675-688.
- \*May TW, McKinney GL. 1981. Cadmium, lead, mercury, arsenic, and selenium concentration in freshwater fish, 1976-77-National Pesticide Monitoring Program. *Pesticides Monitoring Journal* 14-38.
- \*Mazo L, Castoldi AF, Coccini T, et al. 1995. Mechanisms of neurotoxicity: Applications to human biomonitoring. *Toxicol Letter* 77:63-72.
- \*McClanahan MA, Bonhomme C, Gladyszczak-Kohler J, et al. 1996. Mercury contamination in the home (7). *Lancet* 347(9007):1044-1045.
- \*McFarland R, Reigel H. 1978. Chronic mercury poisoning from a single brief exposure. *J Occup Med* 20:534-534.
- \*McGregor AJ, Mason HJ. 1991. Occupational mercury vapour exposure and testicular, pituitary and thyroid endocrine function. *Hum Exp Toxicol* 10(3):199-203.
- \*McKeown-Eyssen GE, Ruedy J, Neims A. 1983. Methylmercury exposure in northern Quebec: II. Neurologic findings in children. *Am J Epidemiol* 118:470-479.
- \*Meador JP, Varanasi U, Robisch PA, et al. 1993. Toxic metals in pilot whales (*globicephala melaena*) from strandings in 1986 and 1990 on Cape Cod, Massachusetts. *Can J Fish Aquat Sci* 50:2698-2706.
- \*Meili M. 1991. The coupling of mercury and organic matter in the biogeochemical cycle - towards a mechanistic model for the boreal forest zone. *Water Air Soil Pollut* 56:333-347.
- \*Meili M, Iverfeldt A, Hakanson L. 1991. Mercury in the surface-water of Swedish forest lakes - concentrations, speciation and controlling factors. *Water, Air, Soil Pollution* 56:439-453.
- \*Melkonian R, Baker D. 1988. Risks of industrial mercury exposure in pregnancy. *Obstet Gynecol Surv* 43(11):637-641.
- \*Mengel H, Karlog O. 1980. Studies on the interaction and distribution of selenite, mercuric, methoxyethyl mercuric and methylmercuric chloride in rats. *Acta Pharmacol Toxicol* 90:46:25-31.

## 8. REFERENCES

- \*Merck. 1989. Merck index: an encyclopedia of chemicals, drugs, and biologicals. 11th ed. Budavari S, ed. Rahway NJ: Merck & Co., Inc.
- \*Merwin I, Pruyne PT, Ebel JG, et al. 1994. Persistence, phytotoxicity, and management of arsenic, lead and mercury residues in old orchard soils or New York state. *Chemosphere* 29(6):1361-1367.
- \*Meydani M, Hathcock J. 1984. Effect of dietary methionine on methylmercury and atrazine toxicity. *Drug Nutr Interact* 2:217-233.
- \*Michaelson JH, McCoy JP, Hirzel P, et al. 1985. Mercury-induced autoimmune glomerulonephritis in inbred rats--part I: Kinetics and species specificity of autoimmune responses. *Surv Synth Pathol Res* 4:401-411.
- \*Mierle G. 1990. Aqueous inputs of mercury to precambrian shield lakes in Ontario. *Environ Toxicol Chem* 9:843-851.
- \*Miettinen JK. 1973. Absorption and elimination of dietary (Hg<sup>++</sup>) and methylmercury in man. In: Miller MW, Clarkson TW, eds. *Mercury, mercurial, and mercaptans*. Springfield, IL, C.C. Thomas.
- \*Miettinen JK, Rahola T, Hattula T, et al. 1969. Retention and excretion of <sup>203</sup>Hg- labelled methylmercury in man after oral administration of CH<sub>3</sub><sup>203</sup>Hg biologically incorporated into fish muscle protein - preliminary results. Fifth RIS (Radioactivity in Scandinavia) Symposium, Department of Radiochemistry, University of Helsinki, Stencils, as cited in Berglund et al. 1971.
- \*Miettinen JK, Rahola T, Hattula T, et al. 1971. Elimination of <sup>203</sup>Hg-methylmercury in man. *Ann Clin Res* 3:116-122.
- \*Millar A. 1916. Perchloride of mercury poisoning by absorption from the vagina. *Br Med J* 2:453-454.
- \*Miller CT, Zawidska Z, Nagy E, et al. 1979. Indicators of genetic toxicity in leukocytes and granulocytic precursors after chronic methylmercury ingestion by cats. *Bull Environ Contam Toxicol* 21:296-303.
- \*Miller DM, Woods JS. 1993. Redox activities of mercury-thiol complexes: Implications for mercury-induced porphyria and toxicity. *Chem Biol Interactions* 88:23-35.
- \*Miller J, Chaffin D, Smith R. 1975. Subclinical psychomotor and neuromuscular changes in workers exposed to inorganic mercury. *Am Ind Hyg Assoc J* 36:725-733.
- \*Milne J, Christophers A, De Silva P. 1970. Acute mercurial pneumonitis. *Br J Ind Med* 27:334-338.
- \*Minyard JP, Roberts WE. 1991. Chemical contaminants monitoring--State findings on pesticide residues in foods--1988 and 1989. *J Assoc Off Anal Chem* 74(3):438-452.
- \*Mirtcheva J, Pfeiffer C, Bruijn JA, et al. 1989. Immunological alterations inducible by mercury compounds: H-2A acts as an immune response and H-2E as an immune "suppression" locus for HgCl<sub>2</sub>-induced antinucleolar autoantibodies. *Eur J Immunol* 19:2257-2261.

## 8. REFERENCES

- Mishonova VN, Stepanova PA, Zarudin VV. 1980. Characteristics of the course of pregnancy and births in women with occupational contact with small concentrations of metallic mercury vapors in industrial facilities. *Gig Truda Prof Zabol* 24(2):21-23.
- \*Miskimmin BM. 1991. Effects of natural levels of dissolved organic carbon (DOC) on methyl mercury formation and sediment water partitioning. *Bull Environ Contam Toxicol* 47(5):743-750.
- \*Miskimmin BM, Rudd JWM, Kelly CA. 1992. Influence of dissolved organic carbon, pH, and microbial respiration rates on mercury methylation and demethylation in lake water. *Can J Fish Aquat Sci* 49(1):17-22.
- \*Mitsumori K, Hirano M, Ueda H, et al. 1990. Chronic toxicity and carcinogenicity of methylmercury chloride in B6C3F1 mice. *Fundamental and Applied Toxicology* 14:179-190.
- \*Mitsumori K, Maita K, Saito T, et al. 1981. Carcinogenicity of methylmercury chloride in ICR mice: Preliminary note on renal carcinogenesis. *Cancer Lett* 12:305-310.
- \*Miyakawa T, Murayama E, Sumiyoshi S, et al. 1976. Late changes in human sural nerves in Minamata disease and in nerves of rats with experimental organic mercury poisoning. *Acta Neuropath (Berlin)* 35:131-138.
- \*Miyakawa T, Sumiyoshi S, Deshimaru M. 1974. Late changes in sciatic nerve of rats after a small dose of methyl mercury sulfide. *Acta Neuropath (Berlin)* 30:33-41.
- \*Miyama T, Minowa K, Seki H, et al. 1983. Chronological relationship between neurological signs and electrophysiological changes in rats with methyl mercury poisoning -- special references to selenium poisoning. *Arch Toxicol* 52:173-181.
- \*Mohamed M, Burbacher T, Mottet N. 1987. Effects of methyl mercury on testicular functions in *Macaca fascicularis* monkeys. *Pharmacol Toxicol* 60(1):29-36.
- \*Molin M, Schutz A, Skerfving S, et al. 1991. Mobilized mercury in subjects with varying exposure to elemental mercury-vapor. *Int Arch Occup Environ Health* 63(3):187-192.
- \*Moody JR, Paulsen PF. 1988. Isotope dilution spark-source mass spectrometric determination of total mercury in botanical and biological samples. *Analyst* 113(6):923-927.
- \*Morales-Rubio A, Mena ML, McLeod CW. 1995. Rapid determination of mercury in environmental materials using on-line microwave digestion and atomic fluorescence spectrometry. *Anal Chem Acta* 308:364-370.
- \*Morgan JN, Berry MR, Graves RL. 1997. Effects of commonly used cooking practices on total mercury concentration in fish and their impact on exposure assessments. *J Expo Anal Environ Epidemiol* 7(1):119-33.
- \*Morris G. 1960. Dermatoses from phenylmercuric salts. *Arch Environ Health* 1:53-55.
- \*Morselli L, Zappoli S, Tirabassa T. 1992. Characterization of the effluents from a municipal solid waste incinerator plant and of environmental impact. *Chemosphere* 24:1775-1784.

## 8. REFERENCES

\*Morselli PL, Franco-Morselli R, Bossi L. 1980. Clinical Pharmacokinetics in Newborns and Infants. *Clinical Pharmacokinetics* 5:485-527.

\*Mortimer DC. 1985. Freshwater aquatic macrophytes as heavy metal monitors - the Ottaea River experience. *Environmental Monitoring Assessment* 5:311-323.

Moszczynski P, Bem S, Moszczynski P Jr, et al. 1990a. The indices of immunity and acute phase reaction according to duration of exposure to mercury vapors in men. *Med Pr* 41(3):169-174. (Polish)

\*Moszczynski P, Lisiewicz J, Bartus R, et al. 1990b. The serum immunoglobulins in workers after prolonged occupational exposure to the mercury vapors. *Rev Roum Med Intern* 28(1):25-30.

\*Moszczynski P, Slowinski S, Rutkowski J, et al. 1995. Lymphocytes, T and NK cells, in men occupationally exposed to mercury vapours. *Int J Occup Med Environ Health* 8(1):49-56.

\*Mudroch A, Hill K. 1989. Distribution of mercury in lake St. Clair and the St. Clair river sediments. *Water Pollution Research Journal of Canada* 24:1-21.

\*Muhlendahl KE. 1990. Intoxication from mercury spilled on carpets. *Lancet* 336 (8730):1578.

\*Muir DCG, Wagemann R, Hargrave BT, et al. 1992. Arctic marine ecosystem contamination. *Science of the Total Environment* 122(1-6):75-134.

\*Mumma RO, Raupach DC, Sahadewan K, et al. 1990. National survey of elements and radioactivity in municipal incinerator ashes. *Arch Environ Contam Toxicol* 19:399-404.

\*Mumma RO, Raupach DC, Sahadewan K, et al. 1991. Variation in the elemental composition of municipal refuse incinerator ashes with time of sampling. *Chemosphere* 23:391-395.

\*Munaf E, Takeuchi T, Ishii D, et al. 1991. Continuous monitoring system for total mercury in waste water by cold vapour atomic-absorption spectrometry and continuous-microflow analysis. *Anal Sci* 7(4):605-609.

\*Munthe J, McElroy WJ. 1992. Some aqueous reactions of potential importance in the atmospheric chemistry of mercury. *Atmos Environ Part A Gen Top* 26(4):553-557.

\*Murphy CB, Carleo DJ. 1977. The contribution of mercury and chlorinated organics from urban runoff. *Water Res* 12:531-533.

\*Murphy MJ, Culliford EJ, Parsons V. 1979. A case of poisoning with mercuric chloride. *Resuscitation* 7:35-44.

Myers GJ, Davidson PW. 1998. Prenatal methylmercury exposure and children: neurologic, developmental, and behavioral research. *Environmental Health Perspectives* 106(3):841-847.

\*Myers GJ, Davidson PW, Shamlaye CF, et al. 1997. Effects of prenatal methylmercury exposure from a high fish diet on developmental milestones in the Seychelles child development study. *Neurotoxicol* 18(3):819-29.



## 8. REFERENCES

- \*Naganuma A, Imura N. 1981. Properties of mercury and selenium in a high-molecular weight substance in rabbit tissues formed by simultaneous administration. *Pharmacol Biochem Behav* 15:449-454.
- \*Nakagawa R. 1995. Concentration of mercury in hair of Japanese people. *Chemosphere* 30(1):127-133.
- \*Nakahara T, Kawakami K, Tamotsu W. 1988. Continuous determination of low concentrations of mercury by atomic-emission spectrometry with helium microwave-induced plasma. *Chemistry Express* 3(11):651-654.
- \*Nakai S, Machida I. 1973. Genetic effect of organic mercury on yeast. *Mutat Res* 21:348.
- \*Nakamura I, Hosokawa K, Tamra H, et al. 1977. Reduced mercury excretion with feces in germfree mice after oral administration of methylmercury chloride. *Bull Environ Contam Toxicol* 17:528-533.
- \*Naleway C, Chou HN, Muller T, et al. 1991. On-site screening for urinary Hg concentrations and correlation with glomerular and renal tubular function. *J Public Health Dent* 51(1):12-17.
- \*Naleway C, Muller T, Sakaguchi R, et al. 1985. Urinary mercury levels in U.S. dentists, 1975-1983: Review of health assessment program. *J Am Dent Assoc* 111:37-42.
- NAS. 1977. An assessment of mercury in the environment: Scientific and technical assessments of environmental pollutants. Washington, DC: National Academy of Sciences, National Research Council. PB83-111625.
- \*NAS/NRC. 1989. Biologic markers in reproductive toxicology. National Academy of Sciences/National Research Council. Washington, DC: National Academy Press, 15-35.
- \*NAS/NRC. 1989. Biological markers in reproductive toxicology. National Research Council. Board of Environmental Studies and Toxicology. Committee on Biological Markers, pp. 15-35.
- \*Natajara S. 1988. Determination of parts-per-trillion levels of mercury with low-power microwave-induced argon-plasma emission spectrometry. *Atom Spectrosc* 9(2):59-62.
- \*Nater EA, Grigal DF. 1992. Regional trends in mercury distribution across the Great Lakes states, north central USA. *Nature (London)* 358(6382):139-141.
- \*NATICH. 1992. National air toxics information clearinghouse. Report on state, local, and EPA air toxics activities. U.S. Environmental Protection Agency, Office of Air Quality Planning and Standards, Research Triangle Park, NC. September 1992.
- \*National Research Council (NRC). 1993. Pesticides in the diets of infants and children. Washington DC: National Academy Press.
- \*Navarro M, Lopez MC, Lopez H, et al. 1992. Microwave dissolution for the determination of mercury in fish by cold vapour atomic absorption spectrometry. *Anal Chim Acta* 257(1):155-158.
- \*NESCAUM. 1998. Northeast states and eastern Canadian provinces - mercury study - a framework for action. Northeast States for Coordinated Air Use Management. Boston, MA.

## 8. REFERENCES

- \*Newland MC, Warfvinge K, Berlin M. 1996. Behavioral consequences of *in utero* exposure to mercury vapor: alterations in lever-press durations and learning in squirrel monkeys. *Toxicol Appl Pharmacol* 139(2):374-386.
- \*NFPA. 1994. Fire protection guide and handbook. National Fire Protection Association, Boston, MA.
- \*Ngim CH, Foo SC, Boey KW, et al. 1992. Chronic neurobehavioural effects of elemental mercury in dentists. *Br J Ind Med* 49(11):782-790.
- \*Ngim CH, Foo SC, Phoon WO. 1988. Atomic absorption spectrophotometric determination of mercury in undigested biological samples. *Ind Health* 26(3):173-178.
- \*NHT. 1971. Methyl mercury in fish. A toxicologic-epidemiologic evaluation of risks. Report from an expert group. *Nordisk Hygienisk Tidskrift. Supplementum* 4. Stockholm, pp. 87-88.
- \*Nichols SJ, Manny BA, Schloesser DW, et al. 1991. Heavy metal contamination of sediments in the upper connecting channels of the Great Lakes. *Hydrobiologia* 219:307-316.
- \*Nielsen JB. 1992. Toxicokinetics of mercuric-chloride and methylmercuric chloride in mice. *J Toxicol Environ Health* 37(1):85-122.
- \*Nielsen JB, Andersen HR, Andersen O, et al. 1991. Mercuric chloride-induced kidney damage in mice: Time course and effect of dose. *J Toxicol Environ Health* 34(4):469-483.
- \*Nielsen JB, Andersen O. 1990. Disposition and retention of mercuric chloride in mice after oral and parenteral administration--1990. *J Toxicol Environ Health* 30(3):167-180.
- \*Nielsen JB, Andersen O. 1991a. Methyl mercuric chloride toxicokinetics in mice: I. Effects of strain, sex, route of administration and dose. *Pharmacol Toxicol* 68(3):201-207.
- \*Nielsen JB, Andersen O. 1991b. Methyl mercuric chloride toxicokinetics in mice: II. Sexual differences in whole-body retention and deposition in blood, hair, skin, muscles and fat. *Pharmacol Toxicol* 68(3):208-211.
- \*Nielsen JB, Andersen O. 1992. Time dependent disposition of mercury after oral dosage. *Metal compounds in environment and life: Interrelationship between chemistry and biology* 4:341-348.
- Nielsen JB, Andersen O, Grandjean P. 1994. Evaluation of mercury in hair, blood and muscle as biomarkers for methylmercury exposure in male and female mice. *Archives of Toxicology* 68(5):317-21.
- \*Nielsen-Kudsk F. 1965. The influence of ethyl alcohol on the absorption of methylmercury vapor from the lungs of man. *Acta Pharmacol Toxicol* 23:263-274.
- \*Nielsen-Kudsk F. 1973. Biological oxidation of elemental mercury. In: Miller MW, Clarkson TW, eds. *Mercury, mercurials and mercaptans*. Springfield, IL: Charles C Thomas, 355.
- \*Nierenberg DW, Nordgren RE, Chang MB, et al. 1998. Delayed cerebellar disease and death after accidental exposure to dimethylmercury. *N Engl J Med* (June 4, 1998) 338(23): 1672-1676.

## 8. REFERENCES

- \*Nieschmidt AK, Kim ND. 1997. Effects of mercury release from amalgam dental restorations during cremation on soil mercury levels of three new zealand crematoria. *Bull Environ Contam Toxicol* 58(5):744-51.
- \*NIOSH. 1973. Criteria for recommended standard: Occupational exposure to inorganic mercury. Rockville, MD: National Institute for Occupational Safety and Health. NIOSH-TR-044-73.
- \*NIOSH. 1984a. Manual of analytical methods. Vol. 1. U.S. Department of Health and Human Services, Division of Physical Sciences and Engineering, National Institute for Occupational Safety and Health, Cincinnati, OH.
- \*NIOSH. 1984b. National occupational exposure survey (1980-83) database. Cincinnati, OH: Department of Health and Human Services, National Institute for Occupational Safety and Health.
- \*NIOSH. 1990. NIOSH pocket guide to chemicals hazards. Washington, DC: U.S. Department of Health and Human Service, Center for Disease Control, National Institute for Occupational Safety and Health, Division of Standard Development and Technology Transfer. NIOSH publication no. 90-117.
- \*NIOSH. 1992. NIOSH recommendations for occupational safety and health--compendium of policy documents and statements. National Institute for Occupational Safety and Health. Department of Health and Human Services. Publication No. 92-100. Cincinnati, Ohio.
- \*NIOSH. 1994. Method 6009, Issue 2, Mercury. NIOSH Manual of Analytical Methods (NMAM), 4th Edition. 1994 U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, National Institute for Occupational Safety and Health.
- \*Nishikido N, Furuyashiki K, Naganuma A, et al. 1987. Maternal selenium deficiency enhances the fetolethal toxicity of methyl mercury. *Toxicol Appl Pharmacol* 88:322-328.
- \*Nishino M, Morita H, Shimomura S. 1986. Mercury levels in dentists' hair. *J Int Assoc Dent Child* 17:9-12.
- \*NOAA. 1987. National Oceanic and Atmospheric Administration. The national status and trends program for marine environmental quality. NOAA Technical Memorandum NOS OMA 38. Rockville, Maryland.
- \*NOAA. 1990. National Oceanic and Atmospheric Administration. The potential for biological effects of sediment-sorbed contaminants tested in the national status and trends program. NOAA Technical Memorandum NOS OMA 52. Seattle, Washington.
- \*Nobmann ED Byers T Lanier AP et al. 1992. The diet of Alaska native adults: 1987-1988. *Am J Clin Nutr* 55:1024-1032.
- \*Nobunaga T, Satoh H, Suzuki T. 1979. Effects of sodium selenite on methylmercury embryotoxicity and teratogenicity in mice. *Toxicol Appl Pharmacol* 47:79-88.
- \*Nolen GA, Buchler EV, Geil RG, et al. 1972. Effects of trisodium nitrotriacetate on cadmium and methylmercury toxicity and teratogenicity in rats. *Toxicol Appl Pharmacol* 23:222-237.

## 8. REFERENCES

- \*Nordberg GF, Brune D, Gerhardsson L, et al. 1992. The ICOH and IUPAC international programme for establishing reference values of metals. *Sci Total Environ* 120(1-2):17-21.
- \*Nordberg GF, ed. 1976. *Effects and dose-response of toxic metals*. New York, NY: Elsevier/North Holland Biomedical Press:24-32.
- \*Nordlind K, Liden S. 1992. Patch test reactions to metal salts in patients with oral mucosal lesions associated with amalgam restorations. *Contact Dermatitis* 27(3):157-160.
- \*Norseth T, Clarkson TW. 1970. Studies on the biotransformation of Hg-203-labeled methylmercury chloride. *Arch Environ Health* 21:717-727.
- \*Norseth T, Clarkson TW. 1971. Intestinal transport of Hg-203-labeled methylmercury chloride: Role of biotransformation in rats. *Arch Environ Health* 22:568-577.
- NREPC. 1986. *Natural Resources and Environmental Protection Cabinet. Department for Environmental Protection, Division of Pollution*. Frankfort, KY. 401 KAR 63:022.
- \*Nriagu JO. 1989. A global assessment of natural sources of atmospheric trace metals. *Nature* 338:47-49.
- \*Nriagu JO, Pacyna JM. 1988. Quantitative assessment of worldwide contamination of air, water, and soils by trace metals. *Nature* 333:134-139.
- \*NTP. 1993. *Toxicology and carcinogenesis studies of mercuric chloride (CAS no. 7487-94-7) in F344/N rats and B6C3F1 mice (gavage studies)*. National Toxicology Program, U.S. Department of Health and Human Services, Public Health Service, National Institutes of Health, Research Triangle Park, NC. NTP TR 408. NIH publication no. 91-3139.
- \*NTP. 1997. *Chemical repository (Radian Corporation august 29, 1991) phenylmercuric acetate*. [http://ntp-db.niehs.nih.gov/NTP\\_Chem\\_H&S/NTP\\_Chem6/Radian62-38-4.txt](http://ntp-db.niehs.nih.gov/NTP_Chem_H&S/NTP_Chem6/Radian62-38-4.txt)
- \*Nublein F, Feicht EA, Schulte-Hostede S, et al. 1995. Exposure analysis of the inhabitants living in the neighbourhood of a mercury-contaminated industrial site. *Chemosphere* 30(12):2241-2248.
- \*Nylander M, Friberg L, Eggleston D, et al. 1989. Mercury accumulation in tissues from dental staff and controls in relation to exposure. *Swed Dent J* 13(6):235-243.
- \*Nylander M, Weiner J. 1991. Mercury and selenium concentrations and their interrelations in organs from dental staff and the general-population. *Br J Ind Med* 48(11):729-734.
- \*O'Connor RP, Ehler CN. 1991. Results from the NOAA national status and trends program on distribution and effects of chemical contamination in the coastal and estuarine United States. *Environ Monit Assess* 17:33-49.
- \*Oberly TJ, Piper CE, McDonald DS. 1982. Mutagenicity of metal salts in the L5178Y mouse lymphoma assay. *J Toxicol Environ Health* 9:367-376.
- \*Ochel M, Vohr HW, Pfeiffer C, et al. 1991. IL4 is required for the IgE and IgG1 increase and IgG1 autoantibody formation in mice treated with mercuric chloride. *J Immunol* 146:3006-3011.

## 8. REFERENCES

- \*Odukoya OO. 1990. Modification of Bethge's open-system apparatus for the determination of mercury in biological materials. *Intern J Environ Anal Chem* 39(4):323-327.
- \*Ohi G, Nishigaki HS, Tamura Y, et al. 1980. The protective potency of marine animal meat against the neurotoxicity of methylmercury: Its relationship with the organ distribution of mercury and selenium in the rat. *Food Cosmet Toxicol* 18:139-145.
- \*Oliveira EM, Vassallo DV. 1992. Effects of mercury on the contractility of isolated rat cardiac muscle. *Braz J Med Biol Res* 25(10):1037-1040.
- \*Oliveira EM, Vassallo DV, Sarkis JJ, et al. 1994. Mercury effects on the contractile activity of isolated heart muscle. *Toxicol Appl Pharmacol* 128(1):86-91.
- \*Olson BH, Cayless SM, Ford S, et al. 1991. Toxic element contamination and the occurrence of mercury-resistant bacteria in mercury contaminated soil, sediments, and sludges. *Arch Environ Contam Toxicol* 20(2):226-233.
- \*Olson FC, Massaro EJ. 1977. Pharmacodynamics of methylmercury in the murine maternal/embryo fetal unit. *Toxicol Appl Pharmacol* 39:263-273.
- \*Olson K, Boush GM. 1975. Decreased learning capacity in rats exposed prenatally and postnatally to low doses of mercury. *Bull Environ Contam Toxicol* 13:73-79.
- \*Orloff KG, Ulirsch G, Wilder L, et al. 1997. Human exposure to elemental mercury in a contaminated residential building. *Arch Environ Health* 52(3):169-72.
- \*OSHA. 1974. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1910.1000.
- OSHA. 1987. Access to employee exposure and medical records. U.S. Department of Labor. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1910.20.
- OSHA. 1988. Access to employee exposure and medical records. U.S. Department of Labor. Occupational Safety and Health Administration. *Fed Reg* 53(189): 30163-30164.
- OSHA. 1989a. Toxic and hazardous substances. U.S. Department of Labor. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1910.1000.
- OSHA. 1989b. Toxic and hazardous substances. U.S. Department of Labor. Occupational Safety and Health Administration. *Fed Reg* 54(12): 2920-2960.
- \*Oskarsson A, Ohlin B, Ohlander EM, et al. 1990. Mercury levels in hair from people eating large quantities of Swedish freshwater fish. *Food Addit Contam* 7(4):555-562.
- \*Oskarsson A, Schutz A, Skerfving S, et al. 1996. Total and inorganic mercury in breast milk and blood in relation to fish consumption and amalgam fillings in lactating women. *Arch Environ Health*. 51(3): 234-241.

## 8. REFERENCES

- \*Osol A, ed. 1980. Remington's pharmaceutical sciences. 16th ed. Easton, PA: Mack Publishing Co., 1106.
- \*Ostlund K. 1969. Studies on the metabolism of methyl mercury in mice. *Acta Pharmacol Toxicol (Suppl. 1)* 27:5-132.
- \*OTA. 1990. Neurotoxicology: Identifying and controlling poisons of the nervous system. Office of Technology Assessment, Washington, DC. OTA-BA-438.
- Outridge PM, Noller BN. 1991. Accumulation of toxic trace elements by freshwater vascular plants. *Rev Environ Contam Tox* 121:1-63.
- \*Owen GM, Brozek J. 1966. Influence of age, sex, and nutrition on body composition during childhood and adolescence. In: Falkner F, ed. *Human Development*. Philadelphia, PA: Saunders, pp. 222-238.
- Padberg S, Burow M, Stoeppler M. 1993. Methyl mercury determination in environmental and biological reference and other materials by quality control with certified reference materials (crms). *Fresenius' J Anal Chem* 346: 686-688.
- \*Paisano, E.L. 1995. The American Indian, Eskimo, and Aleut population. [www.cwnsus.gov/prod/1/pop/profile/95](http://www.cwnsus.gov/prod/1/pop/profile/95).
- \*Pambor M, Timmel A. 1989. Mercury dermatitis. *Contact Dermatitis* 20(2):157.
- \*Parizek J, Ostadolva I. 1967. The protective effect of small amounts of selenite in sublimate intoxication. *Experientia* 23:142.
- \*Parks JW, Curry C, Romani D, et al. 1991. Young northern pike, yellow perch and crayfish as bioindicators in a mercury contaminated watercourse. *Environ Monit Assess* 16:39-73.
- \*Passow H, Rothstein A, Clarkson T. 1961. The general pharmacology of the heavy metals. *Pharmacol Rev* 13:185-224.
- \*Patterson JE, Weissberg BG, Dennison PJ. 1985. Mercury in human breath from dental amalgams. *Bull Environ Contam Toxicol* 34:459-468.
- \*Paudyn A, Van Loon JC. 1986. Determination of organic forms of mercury and arsenic in water and atmospheric samples by gas chromatography-atomic absorption. *Fresenius Z Anal Chem* 325:369-376.
- \*Paustenbach DJ, Bruce GM, Chrostowski P. 1997. Current views on the oral bioavailability of inorganic mercury in soil: Implications for health risk assessments. *Risk Anal* 17(5):533-44.
- \*Pedersen GA, Mortensen GK, Larsen EH. 1994. Beverages as a source of toxic trace element intake. *Food Additives and Contaminants* 11(3):351-363.
- \*Pelletier L, Hirsch F, Rossert J, et al. 1987. Experimental mercury-induced glomerulonephritis. *Springer Sem Immunopathol* 9:359-369.

## 8. REFERENCES

- \*Pelletier L, Pasquier R, Hirsch F, et al. 1986. Autoreactive T cells in mercury-induced autoimmune disease: *in vitro* demonstration. *J Immunol* 137:2548-2554
- \*Pelletier L, Pasquier R, Rossert J, et al. 1988. Autoreactive T cells in mercury-induced autoimmunity: Ability to induce the autoimmune disease. *J Immunol* 140:750-754.
- \*Pelletier L, Rossert J, Pasquier R, et al. 1990. Role of CD8+ cells in mercury-induced autoimmunity or immunosuppression in the rat. *Scand J Immunol* 31:65-74.
- \*Perharic L, Shaw D, Colbridge M, et al. 1994. Toxicological problems resulting from exposure to traditional remedies and food supplements. *Durg Safety* 11(6):284-294.
- \*Perlingeiro RC, Queiroz ML. 1995. Measurement of the respiratory burst and chemotaxis in polymorphonuclear leukocytes from mercury-exposed workers. *Human Exper Toxicol* 14(3):281-286.
- \*Pesce AJ, Hanenson I, Sethi K. 1977. B2 microglobulinuria in a patient with nephrotoxicity secondary to mercuric chloride ingestion. *Clin Toxicol* 11:309-315.
- Pesch H-J, Bloss S, Schubert J, et al. 1992. The mercury cadmium and lead content of cigarette tobacco: Comparative analytical-statistical studies in 1987 and 1991 employing Zeeman-AAS. *Fresenius' J Anal Chem* 343(1):152-153.
- \*Petruccioli L, Turillazzi PG. 1991. Effect of methylmercury on acetylcholinesterase and serum cholinesterase activity in monkeys (*Macaca fascicularis*.) *Bull Environ Contam Toxicol* 46(5):769-773.
- \*Pfab R, Muckter H, Roeder G, et al. 1996. Clinical course of severe poisoning with thiomersal. *J Toxicol Clin Toxicol* 34(4):453-460.
- \*Phelps RW, Clarkson TW, Kershaw TG, et al. 1980. Interrelationships of blood and hair mercury concentrations in a North American population exposed to methylmercury. *Arch Environ Health* 35:161-168.
- \*PHS. 1995. Dental amalgam: A scientific review and recommended Public Health Service strategy for research, education and regulation. Public Health Service. NTIS PB95-160941.
- \*Piikivi L. 1989. Cardiovascular reflexes and low long-term exposure to mercury vapour. *Int Arch Occup Environ Health* 61(6):391-395.
- \*Piikivi L, Hanninen H. 1989. Subjective symptoms and psychological performance of chlor-alkali workers. *Scand J Work Environ Health* 15(1):69-74.
- \*Piikivi L, Hanninen H, Martelin T, et al. 1984. Psychological performance and long term exposure to mercury vapors. *Scand J Work Environ Health* 10:35-41.
- \*Piikivi L, Ruokonen A. 1989. Renal function and long-term low mercury vapor exposure. *Arch Environ Health* 44(3):146-149.
- \*Piikivi L, Tolonen U. 1989. EEG findings in chlor-alkali workers subjected to low long term exposure to mercury vapour. *Br J Ind Med* 46(6):370-375.

## 8. REFERENCES

- Pilgrim W. 1995. Ecosystem aspects of mercury pollution. *Mercury in Aquatic Ecosystems*. 64-77.
- \*Pineau A, Piron M, Boiteau HL, et al. 1990. Determination of total mercury in human hair samples by cold vapor atomic absorption spectrometry. *J Anal Toxicol* 14(4):235-238.
- \*Ping L, Dasgupta PK. 1989. Determination of total mercury in water and urine by a gold film sensor following Fenton's reagent digestion. *Anal Chem* 61(11):1230-1235.
- \*Ping L, Dasgupta PK. 1990. Determination of urinary mercury with an automated micro batch analyzer. *Anal Chem* 62(1):85-88.
- \*Piotrowski J, Trojanowska B, Wisniewska-Knypl JM, et al. 1973. Further investigations on binding and release of mercury in the rat. In: Miller MW, Clarkson TW, eds. *Mercury, mercurials and mercaptans*. Springfield, IL: Charles C Thomas, 247.
- \*Piotrowski JK, Szymanska JA, Skrzypinska-Gawrysiak M, et al. 1992. Intestinal absorption of inorganic mercury in rat. *Pharmacol Toxicol (Copenhagen)* 70(1):53-55.
- \*Pirrone N, Keeler GJ, Nriagu JO, et al. 1996. Historical trends of airborne trace metals in Detroit from 1971 to 1992. *Water Air Soil Pollut* 88(1-2):145-165.
- \*Pitkin RM, Bahns JA, Filer LJ, et al. 1976. Mercury in human maternal and cord blood, placenta, and milk. *Proceedings of the Society for Experimental Biology and Medicine* 151:565-567.
- \*Poma K, Kirsch-Volders M, Susanne C. 1981. Mutagenicity study of mice given mercuric chloride. *J Appl Toxicol* 1:314-316.
- \*Ponce RA, Bloom NS. 1991. Effect of pH on the bioaccumulation of low level, dissolved methylmercury by rainbow trout (*Oncorhynchus mykiss*). *Water Air Soil Pollut* 56:631-640.
- \*Popescu HI, Negru L, Lancranjan I. 1979. Chromosome aberrations induced by occupational exposure to mercury. *Arch Environ Health* 34:461-463.
- \*Porcella DB. 1994. Mercury in the environment: Biogeochemistry. In: Watras CJ, Huckabee JW, eds. *Mercury Pollution Integration and Synthesis*. Boca Raton, Florida: Lewis Publishers, 3-19.
- \*Post EM, Yang MG, King JA, et al. 1973. Behavioral changes of young rats force-fed methylmercury chloride. *Proc Soc Exp Biol Med* 143:1113-1116.
- \*Powell MJ, Quan ESK, Boomer DW, et al. 1992. Inductively coupled plasma mass spectrometry with direct injection nebulization for mercury analysis of drinking water. *Anal Chem* 64(19):2253-2257.
- \*Prem AS, Vachhrajani KD, Bose M, et al. 1992. Action of mercuric chloride during one cycle of seminiferous epithelium in the rat. *Bull Environ Contam Toxicol* 48(6):865-868.
- \*Presley BJ, Taylor RJ, Boothe PN. 1990. Trace metals in gulf of Mexico oysters. *Sci Total Environ* 97/98:551-593.



## 8. REFERENCES

- \*Prouvost-Danon A, Abadie A, Sapin C, et al. 1981. Induction of IgE synthesis and potentiation of anti-ovalbumin IgE antibody response by HgCl<sub>2</sub> in the rat. *J Immunol* 126:699-702.
- \*Pusey CD, Bowman C, Morgan A, et al. 1990. Kinetics and pathogenicity of autoantibodies induced by mercuric chloride in the brown Norway rat. *Clin Exp Immunol* 81:76-82.
- Qasim FJ, Mathieson PW, Thiru S, et al. 1994. Use of methyl prednisolone and antioxidants in mercuric chloride-induced experimental vasculitis. *Clin Exp Immunol* 98(1):66-70.
- \*Qasim FJ, Mathieson PW, Thiru S, et al. 1995. Cyclosporin A exacerbates mercuric chloride induced vasculitis in the Brown Norway rat. *Laboratory Investigations* 72(2):183-190.
- \*Rada RG, Wiener JG, Winfrey MR, et al. 1989. Recent increases in atmospheric deposition of mercury to north-central Wisconsin lakes inferred from sediment analyses. *Arch Environ Contam Toxicol* 18(1-2):175-181.
- \*Rahola T, Hattula, T, Korolainen A, et al. 1973. Elimination of free and protein-bound ionic mercury 203Hg<sup>2+</sup> in man. *Ann Clin Res* 5:214-219.
- \*Rajanna B, Chetty CS, Rajanna S, et al. 1995. Modulation of protein kinase c by heavy metals. *Toxicology Letters* 81(2-3):197-203.
- \*Raman B, Shinde VM. 1990. Extraction, separation and spectrophotometric determination of cadmium and mercury using triphenylphosphine oxide and its application to environmental samples. *Analyst* 115:93-98.
- \*Ramelow GJ, Webre CL, Mueller CS, et al. 1989. Variations of heavy metals and arsenic in fish and other organisms from the Calcasieu river and lake, Louisiana. *Arch Environ Contam Toxicol* 18:804-818.
- \*Rana SVS, Boora PR. 1992. Antiperoxidative mechanisms offered by selenium against liver injury caused by cadmium and mercury in rat. *Bull Environ Contam Toxicol* 48(1):120-124.
- \*Rastogi SC. 1992. Cadmium, chromium, lead, and mercury residues in finger-paints and make-up paints. *Bull Environ Contam Toxicol* 48(2):289-294.
- \*Rastogi SC, Pritzl G. 1996. Migration of some toxic metals from crayons and water colors. *Bull Environ Contam Toxicol* 56(4):527-33.
- \*Rathje AO, Marcero DH, Dattilo D. 1974. Personal monitoring technique for mercury vapor in air and determination. *Am Ind Hyg Assoc J* 35:571-575.
- \*Reed A, Herlofson BB. 1994. Inhibitory effects of HgCL<sub>2</sub> on excitation secretion coupling at the motor nerve terminal and excitation contraction coupling in the muscle cell. *Cellular Mol Neuro* 14(6):623-636.
- \*Reese RG. 1990. Mercury. In: Minerals yearbook. Washington, D.C.: US Dept of the Interior, Bureau of Mines, 743-747.
- \*Reese RG. 1991. Mercury. In: Mineral commodity summaries, 1991. Washington, DC: U.S. Department of the Interior, Bureau of Mines, 102-103.

## 8. REFERENCES

- \*Regnell O, Tunlid A. 1991. Laboratory study of chemical speciation of mercury in lake sediment and water under aerobic and anaerobic conditions. *Appl Environ Microbiol* 57(3):789-795.
- \*Reif JS, Tsongas TA, Anger WK. 1993. Two-stage evaluation of exposure to mercury and biomarkers of neurotoxicity at a hazardous waste site. *J Toxicol Environ Health* 40:413-422.
- Renwick AG. 1991. Safety factors and establishment of acceptable daily intakes. *Food Addit Contam* 8:135-49
- Renwick AG. 1993. Data-derived safety factors for the evaluation of food additives and environmental contaminants. *Food Addit Contam* 10:275-305.
- \*Reuhl KR, Chang LW, Townsend JW. 1981a. Pathological effects of *in utero* methylmercury exposure on the cerebellum of the Golden hamster. I. Early effects upon the neonatal cerebellar cortex. *Environ Res* 26:281-306.
- \*Reuhl KR, Chang LW, Townsend JW. 1981b. Pathological effects of *in utero* methylmercury exposure on the cerebellum of the Golden hamster. II. Residual effects on the adult cerebellum. *Environ Res* 26:307-327.
- \*Revis NW, Osborne TR, Holdsworth G, et al. 1989. Distribution of mercury species in soil from a mercury-contaminated site. *Water Air Soil Pollut* 45(1-2):105-114.
- \*Revis NW, Osborne TR, Holdsworth G, et al. 1990. Mercury in soil: A method for assessing acceptable limits. *Arch Environ Contam Toxicol* 19:221-226.
- \*Ribeyre F. 1991. Experimental ecosystems - comparative study of two methods of contamination of the water column by mercury compounds in relation to bioaccumulation of the metal by rooted macrophytes (*ludwigia-natans*). *Environ Technol* 12(6):503-518.
- \*Ribeyre F, Boudou A, Maurybrachet R. 1991. Multicompartment ecotoxicological models to study mercury bioaccumulation and transfer in fresh water systems. *Water, Air, Soil Pollution* 56:641-652.
- \*Rice DC. 1989a. Blood mercury concentrations following methyl mercury exposure in adult and infant monkeys. *Environ Res* 49(1):115-126.
- \*Rice DC. 1989b. Brain and tissue levels of mercury after chronic methylmercury exposure in the monkey. *J Toxicol Environ Health* 27(2):189-198.
- \*Rice DC. 1989c. Delayed neurotoxicity in monkeys exposed developmentally to methylmercury. *Neurotoxicology* 10(4):645-650.
- \*Rice DC. 1992. Effects of pre-plus postnatal exposure to methylmercury in the monkey on fixed interval and discrimination reversal performance. *Neurotoxicology* 13(2):443-452.
- \*Rice DC. 1996a. Evidence for delayed neurotoxicity produced by methylmercury. *Neurotoxicology* 17(3-4):583-596.

## 8. REFERENCES

- \*Rice DC. 1996b. Sensory and cognitive effects of developmental methylmercury exposure in monkeys, and a comparison to effects in rodents. *Neurotoxicology* 17(1):139-154.
- \*Rice DC, Gilbert SG. 1982. Early chronic low-level methylmercury poisoning in monkeys impairs spatial vision. *Science* 216:759-761.
- \*Rice DC, Gilbert SG. 1990. Effects of developmental exposure to methyl mercury on spatial and temporal visual function in monkeys. *Toxicol Appl Pharmacol* 102(1):151-163.
- \*Rice DC, Gilbert SG. 1992. Exposure to methyl mercury from birth to adulthood impairs high-frequency hearing in monkeys. *Toxicol Appl Pharmacol* 115(1):6-10.
- \*Rice DC, Krewski D, Collins BT, et al. 1989. Pharmacokinetics of methylmercury in the blood of monkeys (*macaca fascicularis*). *Fundam Appl Toxicol* 12(1):23-33.
- \*Richardson GM. 1995. Assessment of mercury exposure and risks from dental amalgam. Medical Devices Bureau, Environmental Health Directorate, Health Canada.  
[Http://www.hc-sc.gc.ca/main/drugs/zmfiles/english/issues/mercury\\_exposure.html](http://www.hc-sc.gc.ca/main/drugs/zmfiles/english/issues/mercury_exposure.html).
- \*Richardson GM, Mitchell M, Coad S et al. 1995. Exposure to mercury in Canada: A multimedia analysis. *Water Air Soil Pollut J* 80:21-30.
- \*Rieber M, Harris DP. 1994. Mercury pollution: The impact of U. S. government stockpile releases. In: Watras CJ, Huckabee JW, eds. *Mercury pollution integration and synthesis*. Boca Raton, Florida: Lewis Publishers, 615-620.
- \*Riisgard HU, Hansen S. 1990. Biomagnification of mercury in a marine grazing food-chain: Algal cells *phaeodactylum tricornutum*, mussels *Mytilus edulis* and flounders *platichthys flesus* studied by means of a stepwise-reduction-CVAA method. *Mar Ecol Prog Ser* 62(3):259-270.
- \*Risher JF and De Rosa CT. 1997. The precision, uses, and limitations of public health guidance values. *Human Ecol. Risk Assmt.* 3(5):681-700.
- \*Robinson KG, Shuman MS. 1989. Determination of mercury in surface waters using an optimized cold-vapour spectrophotometric technique. *Intern J Environ Anal Chem* 36:111-123.
- \*Roed A Herlofson BB. 1994. Inhibitory effects of hgcl<sub>2</sub> on excitation-secretion coupling at the motor nerve terminal and excitation-contraction coupling in the muscle cell. *Cellular & Molecular Neurobiology* 14(6):623-36.
- \*Roels H, Abdeladim S, Ceulemans E, et al. 1987. Relationships between the concentrations of mercury in air and in blood or urine in workers exposed to mercury vapour. *Ind Occup Hyg* 31(2):135-145.
- \*Roels HA, Lauwerys R, Buchet JP, et al. 1982. Comparison of renal function and psychomotor performance in workers exposed. *Int Arch Occup Environ Health* 50:77-93.
- \*Roman-Franco AA, Turiello M, Albini B, et al. 1978. Anti-basement membrane antibodies and antigen-antibody complexes in rabbits injected with mercuric chloride. *Clin Immunol Immunopathol* 9:464-481.

## 8. REFERENCES

- \*Rosenman KD, Valciukas JA, Glickman L, et al. 1986. Sensitive indicators of inorganic mercury toxicity. *Arch Environ Health* 41:208-215.
- \*Rossert J, Pelletier L, Pasquier R, et al. 1988. Autoreactive T cells in mercury-induced autoimmunity: demonstration by limiting dilution analysis. *Eur J Immunol* 18:1761-1766.
- \*Rothstein A, Hayes AL. 1960. The metabolism of mercury in the rat studied by isotope techniques. *J Pharmacol Exp Ther* 130:166-176.
- \*Rothstein A, Hayes AL. 1964. The turnover of mercury in rats exposed repeatedly to inhalation of vapor. *Health Phys* 10:1099-1113.
- \*Rowens B, Guerrero-Betancourt D, Gottlieb CA, et al. 1991. Respiratory failure and death following acute inhalation of mercury vapor: A clinical and histologic perspective. *Chest* 99(1):185-190.
- \*Rowland AS, Baird DD, Weinberg CR, et al. 1994. The effect of occupational exposure to mercury vapour on the fertility of female dental assistants [see comments]. *Occup Environ Med* 51(1):28-34.
- \*Rowland I, Davies M, Evans J. 1980. Tissue content of mercury in rats given methylmercury chloride orally: Influence of intestinal flora. *Arch Environ Health* 35:155-160.
- \*Rozalski M, Wierzbicki R. 1983. Effect of mercuric chloride on cultured rat fibroblasts: Survival, protein biosynthesis and binding of mercury to chromatin. *Biochem Pharmacol* 32:2124-2126.
- \*RTECS. 1997. Registry of Toxic Effects of Chemical Substances (RTECS). National Institute for Occupational Safety and Health (NIOSH). Computer database online.
- \*RTECS. 1998. Registry of Toxic Effects of Chemical Substances (RTECS). National Institute for Occupational Safety and Health (NIOSH). Computer database online.
- Rudd WM. 1995. Sources of methyl mercury to freshwater ecosystems: a review. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994.* Boston, MA: Kluwer Academic Publishers, 697-713.
- \*Rule JH, Iwashchenko MS. 1998. Mercury concentrations in soils adjacent to a former chlor-alkali plant. *Journal Environmental Quality* 27:31-37.
- \*Rumbeiha WK, Gentry PA, Bhatnagar MK. 1992. The effects of administering methylmercury in combination with ethanol in the rat. *Vet Hum Toxicol* 34(1):21-25.
- \*Ryan DM, Sin YM, Wong MK. 1991. Uptake, distribution and immunotoxicological effects of mercury in mice. *Environ Monit Assess* 19(1-3):507-517.
- \*Sager PR, Aschner M, Rodier PM. 1984. Persistent, differential alterations in developing cerebellar cortex of male and female mice after methylmercury exposure. *Dev Brain Res* 12:1-11.
- \*Sager PR, Doherty RA, Olmsted JB. 1983. Interaction of methylmercury with microtubules in cultured cells and *in vitro*. *Exp Cell Res* 146:127-137.

## 8. REFERENCES

- \*Sager PR, Doherty RA, Rodier PM. 1982. Effects of methylmercury on developing mouse cerebellar cortex. *Exp Neurol* 77:179-183.
- \*Sahaphong S, Trump BF. 1971. Studies of cellular injury in isolated kidney tubules of the flounder. *Am J Pathol* 63:277-290.
- \*Sakamoto M, Nakano A, Kinjo Y, et al. 1991. Present mercury levels in red blood cells of nearby inhabitants about 30 years after the outbreak of Minamata disease. *Ecotoxicol Environ Safety* 22:58-66.
- \*Sallsten G, Barregard L, Achutz A, et al. 1993. Decrease in mercury concentration in blood after long term exposure: A kinetic study of chloralkali workers. *Brit J Ind Med* 50:814-821.
- \*Sallsten G, Barregard L, Schutz A. 1994. Clearance half life of mercury in urine after the cessation of long term occupational exposure: influence of a chelating agent (DMPS) on excretion of mercury in urine. *Occup Environ Med* 51(5):337-342.
- \*Sallsten G, Thoren J, Barregard L, et al. 1996. Long-term use of nicotine chewing gum and mercury exposure from dental amalgam fillings. *J Dent Res* 75(1):594-8.
- \*Salonen JT, Seppanen K, Nyyssonen K, et al. 1995. Intake of mercury from fish, lipid peroxidation and the risk of myocardial infarction and coronary, cardiovascular and any death in eastern Finnish men. *Circulation* 91 (3):645-655.
- \*Salvaterra P, Lown B, Morganti J, et al. 1973. Alterations in neurochemical and behavioral parameters in the mouse induced by low levels of methylmercury. *Acta Pharmacol Toxicol* 33:177-190.
- \*Samudralwar DL, Garg AN. 1996. Minor and trace elemental determination in the Indian herbal and other medicinal preparations. *Biol Trace Elem Res* 54(2):113-21.
- \*Samuels ER, Heick HMC, McLaine PN, et al. 1982. A case of accidental inorganic mercury poisoning. *J Anal Toxicol* 6:120-122.
- \*Sandborgh-Englund G, Elinder C-G, Johanson G, et al. 1998. The absorption, blood levels, and excretion of mercury after a single dose of mercury vapor in humans. *Toxicol Appl Pharmacol* 150: 146-153.
- \*Sandborgh-Englund G, Nygren AT. 1996. No evidence of renal toxicity from amalgam fillings. *Amer J Physiol* 271(4):R941-R945.
- \*Sapin C, Hirsch F, Delaporte J-P, et al. 1984. Polyclonal IgE increase after HgCl<sub>2</sub> injections in BN and LEW rats: A genetic analysis. *Immunogenetics* 20:227-236.
- \*Sarafian T, Verity MA. 1991. Oxidative mechanisms underlying methyl mercury neurotoxicity. *Int J Dev Neurol* 9(2):147-153.
- \*Sarafian TA. 1993. Methyl mercury increases intracellular Ca<sup>2+</sup> and inositol phosphate levels in cultured cerebellar granule neurons. *J Neurochem* 61(2):648-657.
- \*Sasso FS, Ferraiuolo R, Garetano G, et al. 1996. Mercury exposure among residents of a building formerly used for industrial purposes--New Jersey, 1995. *JAMA* 276(1):17-8.

## 8. REFERENCES

- \*Sato K, Sada K. 1992. Effects of emissions from a coal-fired power plant on surface soil trace element concentrations. *Atmos Environ Pt 26A*:325-331.
- \*Sau P, Solivan G, Johnson FB. 1991. Cutaneous reaction from a broken thermometer. *J Am Acad Dermatol 25(5 Pt 2)*:915-919.
- \*Schaller KH, Triebig G, Schiele R, et al. 1991. Biological monitoring and health surveillance of workers exposed to mercury. In: Dillon HK, Ho MJ, eds. *Biological monitoring of exposure to chemicals: Metals*. New York, NY: John Wiley and Sons, 3-9.
- \*Schamberg J, Kolmer J, Raiziss G. 1918. Experimental studies of the mode of absorption of mercury when applied by injection. *JAMA 70*:142.
- \*Schintu M, Kauri T, Kudo A. 1989. Inorganic and methyl mercury in inland waters. *Water Res 23(6)*:699-704.
- \*Schionning JD, Poulsen EH, Moller-Madsen B, et al. 1991. Ultrastructural localization of mercury in rat dorsal root ganglia after exposure to mercury vapor. In: Graumann W, ed. *Progress in histochemistry and cytochemistry*, vol. 23, No. 1-4: Histo- and cytochemistry as a tool in environmental toxicology. XXXII Symposium of the International Association of Histochemists, Gargellen, Austria, September 26-29, 1990. New York, NY: Gustav Fischer Verlag, 249-255.
- \*Schmitt CJ, Brumbaugh WG Jr. 1990. National contaminant biomonitoring program: Concentrations of arsenic, cadmium, copper, lead, mercury, selenium, and zinc in U.S. freshwater fish, 1976-1984. *Arch Environ Contam Toxicol 19*:731-747.
- \*Schoof RA, Nielsen JB. 1997. Evaluation of methods for assessing the oral bioavailability of inorganic mercury in soil. *Risk Anal 17(5)*:545-55.
- \*Schroeder WH, Fanaki FJ. 1988. Field measurements of water-air exchange of mercury in freshwater systems. *Environ Technol Lett 9(5)*:369-374.
- \*Schroeder WH, Yarwood G, Niki H. 1991. Transformation processes involving mercury species in the atmosphere - results from a literature survey. *Water, Air, Soil Pollution 56*:653-666.
- \*Schuckmann F. 1979. Study of preclinical changes in workers exposed to inorganic mercury in chloralkali plants. *Int Arch Occup Environ Health 44*:193-200.
- \*Schuster E. 1991. The behavior of mercury in the soil with special emphasis on complexation and adsorption process - a review of the literature. *Water, Air, Soil Pollution 56*:667-680.
- \*Schwartz JG, Snider TE, Montiel MM. 1992. Toxicity of a family from vacuumed mercury. *Am J Emerg Med 10(3)*:258-261.
- \*Schwarz S, Husstedt IW, Bertram HP, et al. 1996. Amyotrophic lateral sclerosis after accidental injection of mercury (7). *Journal of Neurology Neurosurgery and Psychiatry 60/6*:698.

## 8. REFERENCES

- \*Sedman RM, Polisini JM, Esparza JR. 1994. The evaluation of stack metal emissions from hazardous waste incinerators: assessing human exposure through noninhalation pathways. *Environ Health Perspect* 102(2):105-112.
- \*Sekowski JW, Malkas LH, Wei Y, et al. 1997. Mercuric ion inhibits the activity and fidelity of the human cell DNA synthetase. *Toxicol Appl Pharmacol* 145:268-276.
- \*Sengar CBS, Kumar A, Hasan MZ, et al. 1990. Comparative studies on extraction of mercury from suspended particulate matter. *Ind J Environ Health* 32(1):1-4.
- \*Setchell BP, Waites GMH. 1975. The blood testis barrier. In: Creep RO, Astwood EB, eds., executive ed. Geiger SR. *Handbook of physiology: Endocrinology* (chapter 6). American Physiological Society, Washington DC.
- \*Sexton D, Powell K, Liddle J, et al. 1976. A nonoccupational outbreak of inorganic mercury vapor poisoning. *Arch Environ Health* 33:186-191.
- \*Shapiro IM, Sumner AJ, Spitz LK, et al. 1982. Neurophysiological and neuropsychological function in mercury exposed dentists. *Lancet* 1:1147-1150.
- \*Sharma RL, Singh HG. 1989. Derivative spectrophotometric determination of mercury(II) with PAN (1-(2-pyridylazo)-2-naphthol) in the aqueous phase. *Talanta* 36(4):457-461.
- \*Sharma RP, Aldous CN, Farr CH. 1982. Methylmercury induced alterations in brain amine synthesis in rats. *Toxicol Lett* 13:195-201.
- \*Shaw BP, Dash S, Panigrahi AK. 1991. Effect of methyl mercuric chloride treatment on haematological characteristics and erythrocyte morphology of Swiss mice. *Environmental Pollution* 73(1):43-52.
- \*Shaw BP, Sahu A, Panigrahy AK. 1986. Mercury in plants, soil, and water from a caustic chlorine industry. *Bull Environ Contam Toxicol* 36:299-305.
- \*Shenker BJ, Berthold P, Rooney C, et al. 1993. Immunotoxic effects of mercuric compounds on human lymphocytes and monocytes: III. Alterations in B-cell function and viability. *Immunopharmacol Immunotoxicol* 15(1):87-112.
- \*Sherlock J, Hislop J, Newton D, et al. 1984. Elevation of mercury in human blood from controlled chronic ingestion of methylmercury in fish. *Human Toxicol* 3:117-131.
- \*Sherlock JC, Lindsay DG, Evans WH, et al. 1982. Duplication diet study on mercury intake by fish consumers in the United Kingdom. *Arch Environ Health* 37(5):271-278.
- \*Sherlock JC, Quinn M. 1988. Underestimation of dose-response relationship with particular reference to the relationship between the dietary intake of mercury and its concentration in blood. *Hum Toxicol* 7:129-132.
- \*Shimada H, Fukudome S, Kiyozumi M, et al. 1993. Further study of effects of chelating agents on excretion of inorganic mercury in rats. *Toxicology (Ireland)* 77(1-2):157-169.

## 8. REFERENCES

- \*Shkinev VM, Gomolitskii VN, Spivakov BY, et al. 1989. Determination of trace heavy metals in waters by atomic-absorption spectrometry after pre-concentration by liquid-phase polymer-based retention. *Talanta* 36(8):861-863.
- \*Shofstahl JH, Hardy JK. 1990. Method for the determination of the priority pollutant metals by HPLC. *J Chromatog Sci* 28(5):225-229.
- \*Siblerud RL. 1990. The relationship between mercury from dental amalgam and the cardiovascular system. *Sci Total Environ* 99(1-2):23-36.
- Siblerud RL. 1992. A comparison of mental health of multiple sclerosis patients with silver/mercury dental fillings and those with fillings removed. *Psychol Rep* 70(3):1139-1151.
- Siblerud RL and Kienholz E. 1994. Evidence that mercury from silver dental fillings may be an etiological factor in multiple sclerosis. *Sci Total Environ* 142(3):191-205.
- \*Siblerud RL, Kienholz E. 1997. Evidence that mercury from silver dental fillings may be an etiological factor in reduced nerve conduction velocity in multiple sclerosis patients. *Journal of Orthomolecular Medicine* 12(3):169-172.
- \*Sidle WC. 1993. Naturally occurring mercury contamination in a pristine environment? *Environ Geology* 21:42-50.
- \*Sikorski R, Juskiewicz T, Paszkowski T, et al. 1987. Women in dental surgeries: Reproductive hazards in occupational exposure to metallic mercury. *Int Arch Occup Environ Health* 59:551-557.
- \*Silberberg I, Prutkin L, Leider M. 1969. Electron microscopic studies of transepidermal absorption of mercury. *Arch Environ Health* 19:7-14.
- \*Sin YM, Lim YF, Wong MK. 1983. Uptake and distribution of mercury in mice from ingesting soluble and insoluble mercury compounds. *Bull Environ Contam Toxicol* 31(5):605-612.
- \*Sin YM, Teh WF. 1992. Effect of long-term uptake of mercuric sulphide on thyroid hormones and glutathione in mice. *Bull Environ Contam Toxicol* 49(6):847-854.
- \*Sin YM, Teh WF, Wong MK, et al. 1990. Effect of mercury on glutathione and thyroid hormones. *Bull Environ Contam Toxicol* 44(4):616-622.
- \*Singh HG, Kumar B, Sharma RL, et al. 1989. Direct spectrophotometric determination of trace amounts of mercury(II) in aqueous media as its dithizonate complex in the presence of a neutral surfactant. *Analyst* 114(7):853-855.
- \*Skare I. 1995. Mass balance and systemic uptake of mercury released from dental amalgams. *Water Air and Soil Pollution* 80:59-67.
- \*Skare I, Bergstroem T, Engqvist A, et al. 1990. Mercury exposure of different origins among dentists and dental nurses. *Scand J Work Environ Health* 16:340-347.



## 8. REFERENCES

- \*Skare I, Engqvist A. 1994. Human exposure to mercury and silver released from dental amalgam restorations. *Arch Environ Health* 49(5):384-394.
- \*Skerfving S. 1974. Methylmercury exposure, mercury levels in blood and hair, and health status in Swedes consuming contaminated fish. *Toxicology* 2:3-23.
- \*Skerfving S. 1978. Interaction between selenium and methylmercury. *Environ Health Perspect* 25:57-65.
- \*Skerfving S. 1988. Mercury in women exposed to methylmercury through fish consumption, and in their newborn babies and breast milk. *Bull Environ Contam Toxicol* 41(4):475-482 .
- \*Skerfving S, Hansson K, Lindsten J. 1970. Chromosome breakage in humans exposed to methylmercury through fish consumption. *Arch Environ Health* 21:133-139.
- \*Skerfving S, Hansson K, Mangs C, et al. 1974. Methylmercury-induced chromosome damage in man. *Environ Res* 7:83-98.
- \*Skoglund A, Egelrud T. 1991. Hypersensitivity reactions to dental materials in patients with lichenoid oral mucosal lesions and in patients with burning mouth syndrome. *Scand J Dent Res* 99(4):320-328.
- \*Smith PJ, Langolf GD, Goldberg J. 1983. Effects of occupational exposure to elemental mercury on short term memory. *Br J Ind Med* 40:413-419.
- \*Smith RG, Vorwald AJ, Patel LS, et al. 1970. Effects of exposure to mercury in the manufacture of chlorine. *Am Ind Hyg Assoc J* 31:687-700.
- \*Smith TG, Armstrong AJ. 1975. Mercury in seals, terrestrial carnivores, and principle food items of the Inuit, from Holman, N.W.T. - interim report. *Journal Fisheries Research Board of Canada* 32 (6):795-801.
- \*Snapp KR, Boyer DB, Peterson LC, Svare CW. 1989. The contribution of dental amalgam to mercury in blood. *J Dent Res* 68:780-785
- \*Snodgrass W, Sullivan JB, Rumack BH, et al. 1981. Mercury poisoning from home gold ore processing: Use of penicillamine and dimercaprol. *JAMA* 246:1929-1931.
- \*Snyder R, Seelinger D. 1976. Methylmercury poisoning: Clinical follow-up and sensory nerve conduction studies. *J Neurol Neurosurg Psychiatry* 39:701-704.
- \*Snyder WS, Cook MT, Karhausen LR. et al. 1981. International Commission of Radiological Protection. No. 23: Report of the task group on reference man. New York, NY: Pergamon Press.
- \*Solecki R, Hothorn L, Holzweissig M, et al. 1991. Computerised analysis of pathological findings in longterm trials with phenylmercuric acetate in rats. *Arch Toxicol (Supp 14)*:100-103.
- \*Somjen SG, Herman S, Klein R, et al. 1973. The uptake of methylmercury (<sup>203</sup>Hg) in different tissues related to its neurotoxic effects. *J Pharmacol Exp Ther* 187:602-611.
- \*Soni JP, Singhania RU, Bansal A, et al. 1992. Acute mercury vapor poisoning. *Indian Pediatr* 29(3):365-368.

## 8. REFERENCES

- \*Soria ML, Sanz P, Martinez D, et al. 1992. Total mercury and methylmercury in hair maternal and umbilical blood and placenta from women in the Seville area. *Bull Environ Contam Toxicol* 48(4):494-501.
- \*Southworth GR, Peterson MJ, Turner RR. 1994. Changes in concentrations of selenium and mercury in largemouth bass following elimination of fly ash discharge to a quarry. *Chemosphere* 29(1):71-79.
- \*Spencer JH, Voigt AF. 1968. Thermodynamics of the solution of mercury metal: I. Tracer determination. *J Phys Chem* 72:464-470.
- \*Spyker JM, Smithberg M. 1972. Effects of methylmercury on prenatal development in mice. *Teratology* 5:181-190.
- \*Spyker JM, Sparber SB, Goldberg AM. 1972. Subtle consequences of methyl mercury exposure: Behavioral deviations in offspring of treated mothers. *Science* 177:621-623.
- \*SRI. 1996. Stanford Research Institute International. Directory of chemical producers: United States of America. Menlo Park, Ca: SRI International.
- \*Stein ED, Cohen Y, Winer AM. 1996. Environmental distribution and transformation of mercury compounds. *Crit Rev Environ Sci Technol* 26(1):1-43.
- Stern AH. 1993. Re-evaluation of the reference dose for methylmercury and assessment of current exposure levels. *Risk Analysis* 13(3):355-364.
- \*Stewart W, Guirgis H, Sanderson J, et al. 1977. Urinary mercury excretion and proteinuria in pathology laboratory staff. *Br J Ind Med* 34:26-31.
- \*Stockwell PB, Rabl P, Paffrath M. 1991. Monitoring elemental mercury in an urban environment. *Process Control Qual* 1(4):293-298.
- \*Stokinger, H. 1981. Patty's industrial hygiene and toxicology. In: Clayton GD, Clayton FE, eds. 3rd ed, vol. IIA. New York, NY: John Wiley & Sons, 1769-1792.
- \*Stoltenburg-Didinger G, Markwort S. 1990. Prenatal methylmercury exposure results in dendritic spine dysgenesis in rats. *Neurotoxicol Teratol* 12(6):573-576.
- \*Stonard MD, Chater BR, Duffield DP, et al. 1983. An evaluation of renal function in workers occupationally exposed to mercury vapor. *Int Arch Occup Environ Health* 52:177-189.
- \*Stopford W, Bundy SD, Goldwater LJ, et al. 1978. Microenvironmental exposure to mercury vapor. *Am Ind Hyg Assoc J* 39:378-384.
- \*Storm DL. 1994. Chemical monitoring of California's public drinking water sources: Public exposures and health impacts. In: Wang RGM. *Water contamination and health*. New York: Marcel Dekker, Inc., 67-124.
- \*Stoz F, Aicham P, Janovic S, et al. 1995. [Is a generalized amalgam ban justified? Studies of mothers and their newborn infants]. *Zeitschrift Fur Geburtshilfe Und Perinatologie* (1):35-41. (German)

## 8. REFERENCES

- \*Stutz DR, Janusz. 1988. Hazardous materials injuries: A handbook for pre-hospital care. Second edition. Beltsville, MD: Bradford Communications Corporation, 323-332.
- \*Su M, Okita GT. 1976. Embryocidal and teratogenic effects of methylmercury in mice. *Toxicol Appl Pharmacol* 38:207-216.
- \*Suda I, Eto K, Tokunaga H, et al. 1989. Different histochemical findings in the brain produced by mercuric chloride and methyl mercury chloride in rats. *Neurotoxicology* 10(1):113-125.
- \*Suda I, Hirayama K. 1992. Degradation of methyl- and ethylmercury into inorganic mercury by hydroxyl radical produced from rat liver microsomes. *Arch Toxicol* 66(6):398-402.
- \*Suda I, Takahashi H. 1992. Degradation of methyl and ethyl mercury into inorganic mercury by other reactive oxygen species besides hydroxyl radical. *Arch Toxicol* 66(1):34-39.
- \*Suda I, Totoki S, Takahashi H. 1991. Degradation of methyl and ethyl mercury into inorganic mercury by oxygen free radical-producing systems: Involvement of hydroxyl radical. *Arch Toxicol* 65(2):129-134.
- \*Sue Y-J. 1994. Mercury. In: Goldfrank LR, Flomenbaum NE, Lewin NA eds. *Goldfrank's toxicologic emergencies*, Fifth Edition. Norwalk, Connecticut: Appleton and Lange, 1051-1062.
- \*Sundberg J, Jonsson S, Karlsson MO, et al. 1998. Kinetics of methylmercury and inorganic mercury in lactating and nonlactating mice. *Toxicology and Applied Pharmacology* 51:319-329.
- \*Sundberg J, Oskarsson A. 1992. Placental and lactational transfer of mercury from rats exposed to methylmercury in their diet: Speciation of mercury in the offspring. *J Trace Elem Exp Med* 5(1):47-56.
- \*Sunderman FW Sr. 1978. Clinical response to therapeutic agents in poisoning from mercury vapor. *Ann Clin Lab Sci* 8(4):259-269.
- \*Suo Y, Yi F, Huang Y. 1992. Determination of trace mercury in hair, urine and nail by flameless nondispersive atomic fluorescence spectrometry. *Fenxi Huaxue* 20(3):335-338.
- \*Suter KE. 1975. Studies on the dominant-lethal and fertility effects of the heavy metal compounds methylmercuric hydroxide, mercuric chloride and cadmium chloride in male and female mice. *Mutat Res* 30:365-374.
- \*Suzuki T, Hongo T, Matsuo N, et al. 1992. An acute mercuric mercury poisoning: Chemical speciation of hair mercury shows a peak of inorganic mercury value. *Hum Exp Toxicol* 11(1):53-57.
- \*Suzuki T, Hongo T, Yoshinaga J, et al. 1993. The hair-organ relationship in mercury concentration in contemporary Japanese. *Arch Environ Health* 48(4):221-229.
- \*Swain EB, Engstrom DR, Brigham ME, et al. 1992. Increasing rates of atmospheric mercury deposition in midcontinental North America. *Science* 257(5071):784-787.
- \*Sweet CW, Vermette SJ. 1993. Sources of toxic trace elements in urban air in Illinois. *Environmental Science and Technology*. 27:2502-2510.

## 8. REFERENCES

- \*Syversen T. 1977. Effects of methylmercury on *in vivo* protein synthesis in isolated cerebral and cerebellar neurons. *Neuropathol Appl Neurobiol* 3:225-236.
- \*Takahata N, Hayashi H, Watanabe B, et al. 1970. Accumulation of mercury in the brains of two autopsy cases with chronic inorganic mercury poisoning. *Folia Psychiatr Neurol Jpn* 24:59-69.
- \*Takeuchi T, Eto K, Tokunaga H. 1989. Mercury level and histochemical distribution in a human brain with Minamata disease following a long-term clinical course of 26 years. *Neurotoxicology* 10(4):651-658.
- \*Tamashiro H, Akagi H, Arakaki M, et al. 1984. Causes of death in Minamata disease: Analysis of death certificates. *Int Arch Occup Environ Health* 54:135-146.
- \*Tamashiro H, Arakaki M, Akagi H, et al. 1986. Effects of ethanol on methyl mercury toxicity in rats. *J Tox Environ Health* 18:595-605.
- \*Tanaka T, Naganuma A, Kobayashi K, et al. 1991. An explanation for strain and sex differences in renal uptake of methylmercury in mice. *Toxicology* 69(3):317-329.
- \*Tanaka T, Naganuma A, Miura N, et al. 1992. Role of testosterone in gamma-glutamyl transpeptidase-dependent renal methylmercury uptake in mice. *Toxicol Appl Pharmacol* 112(1):58-63.
- \*Task Group on Metal Accumulation. 1973. Accumulation of toxic metals with special reference to their absorption, excretion, and biological half-times. *Environ Physiol Biochem* 3:65-107.
- \*Taskaev E, Penev I, Kinova L. 1988. Radiochemical determination of mercury, arsenic, cadmium, zinc and copper in biological materials. *J Radioanal Nucl Chem* 120(1):83-88.
- \*Taueg C, Sanfilippo DJ, Rowens B, et al. 1992. Acute and chronic poisoning from residential exposures to elemental mercury. *J Toxicol Clin Toxicol* 30(1):63-67.
- \*Taylor NB. 1961. A text in applied physiology, 7th ed., The Williams and Wilkins Company, 19 and 29.
- \*Teisinger J, Fiserova-Bergerova V. 1965. Pulmonary retention and excretion of mercury vapors in man. *Ind Med Surg* 34:580.
- \*Tejning S. 1967. Mercury contents in blood corpuscles, blood plasma and hair in persons who had for long periods a high consumption of freshwater fish from Lake Vaner. Report 67 08 31 from Department of Occupational Medicine, University Hospital, S-221 85 Lund, Stencils. Swedish, as cited in Berglund et al. 1971.
- \*Temmerman E, Vandecasteele C, Vermeir G, et al. 1990. Sensitive determination of gaseous mercury in air by cold-vapour atomic-fluorescence spectrometry after amalgamation. *Anal Chim Acta* 236(2):371-376.
- \*Teng C, Brennan J. 1959. Acute mercury vapor poisoning: A report of four cases with radiographic and pathologic correlation. *Radiology* 73:354-361.
- \*Tennant R, Johnston H, Wells J. 1961. Acute bilateral pneumonitis associated with the inhalation of mercury vapor: A report of five cases. *Conn Med* 25:106-109.

## 8. REFERENCES

- \*Thomas D, Fisher H, Hall L, et al. 1982. Effects of age and sex on retention of mercury by methyl mercury-treated rats. *Toxicol Appl Pharmacol* 62:445-454.
- \*Thorp JM Jr, Boyette D, Watson WJ, et al. 1992. Elemental mercury exposure in early pregnancy. *Obstet Gynecol* 79(5 Pt 2):874-876.
- \*Thuvander A Sundberg J Oskarsson A. 1996. Immunomodulating effects after perinatal exposure to methylmercury in mice. *Toxicology*;114(2):163-75.
- \*Tichenor BA, Guo Z. 1991. Small-chamber determinations of the emission rates of mercury from latex paints. Report ISS EPA/600/D-91/155.
- \*Tollefson L, Cordle F. 1986. Methylmercury in fish: A review of residue levels, fish consumption and regulatory action in the United States. *Environ Health Perspect* 68:203-8.
- \*Toribara TY, Clarkson TW, Nierenberg DW. 1997. More on working with dimethylmercury. *Chemical and Engineering News*. 75(24):3,6,11,12.
- \*Toro EC, Das HA, Fardy JJ. 1994. Toxic Heavy metals and other trace elements in foodstuffs from 12 different countries. In: Schrauzer GN., *Biological Trace Element Research*. Humana Press Inc.
- \*Torresani C, Caprari E, Manara GC. 1993. Contact urticaria syndrome due to phenylmercuric acetate. *Contact Dermatitis* 29(5):282-3.
- \*Travis CC, Blaylock BP. 1992. Validation of terrestrial food-chain model. *J Expos Anal Environ Epidemiol* 2(2):221-239.
- \*TRI91. 1993. Toxic Chemical Release Inventory. National Library of Medicine, National Toxicology Information Program, Bethesda, MD.
- \*TRI94. 1996. Toxic Chemical Release Inventory. National Library of Medicine, National Toxicology Information Program, Bethesda, MD.
- \*TRI96. 1998. Toxic Chemical Release Inventory. National Library of Medicine, National Toxicology Information Program, Bethesda, MD.
- \*Troen P, Kaufman SA, Katz KH. 1951. Mercuric bichloride poisoning. *N Engl J Med* 244:459-463.
- \*Trotter RT. 1985. Greta and Azarcon : A survey of episodic lead poisoning from a folk remedy. *Human Organization* 44(1) 64 -72
- \*Tsubaki 1971b. In: *Special Symposium on Mercury in Man's Environment*. Ottawa, February 15-16, p. 131, as cited in WHO 1976.
- \*Tsubaki T. 1971a. Personal communication. Letter (Dnr F 2395/68 Hg 87/69) to S. Skerfving, Oct. 4, 1969. Letter to M. Berlin, May 28, 1969, as cited in Berglund et al. 1971.
- \*Tsubaki T, Takahashi H. 1986. *Recent advances in Minamata disease studies*. Tokyo, Japan: Kodansha, Ltd.

## 8. REFERENCES

- \*Tsuzuki Y. 1981. Effect of chronic methylmercury exposure on activities of neurotransmitter enzymes in rat cerebellum. *Toxicol Appl Pharmacol* 60:379-381.
- \*Tubbs R, Gordon D, Gephardt N, et al. 1982. Membranous glomerulonephritis associated with industrial mercury exposure--study of pathogenic mechanisms. *Am J Clin Pathol* 77:409-413.
- \*Tunnessen WW, McMahon KJ, Baser M. 1987. Acrodynia: Exposure to mercury from fluorescent light bulbs. *Pediatrics* 79:786-789.
- \*Turner CJ, Bhatnagar MK, Yamashiro S. 1981. Ethanol potentiation of methyl mercury toxicity: A preliminary report. *J Tox Environ Health* 7:665-668.
- \*Turner MD, Smith JC, Kilpper RW, et al. 1975. Absorption of natural methylmercury (MeHg) from fish. *Clin Res* 23:2.
- \*Turner RR, Bogle MA. 1993. Ambient air monitoring for mercury around an industrial complex. In: Chow W, Connor KK, eds. *Managing hazardous air pollutants state of the art*. Boca Raton, Florida: Lewis Publishers, 162-172.
- \*Turner RR, Bogle MA, Heidel LL, et al. 1992. Mercury in ambient air at the Oak Ridge Y-12 Plant, July 1986 through December 1990. *Govt Reports Announcements and Index (GRA&I) Issue 02*.
- \*U.S. Congress. 1986. Superfund amendments and reauthorization act of 1986. Title III Emergency Planning and Community Right-to-know. Ninety-ninth Congress of the United States of America.
- \*U.S. Congress. 1990. Clean air act amendments. Title III, Hazardous Air Pollutants, Section 112, Hazardous Air Pollutants as Amended October 26, 1990. One Hundred and First Congress of the United States of America, 2nd Session Report 101-952.
- \*Urano T, Iwasaki A, Himeno S, et al. 1990. Absorption of methylmercury compounds from rat intestine. *Toxicol Lett* 50(2-3):159-164.
- \*USGS 1997. Mercury. United States Geological Survey.
- \*Uzzell BP, Oler J. 1986. Chronic low-level mercury exposure and neurophysiological functioning. *J Clin Exp Neuropsychol* 8:581-593.
- \*Vachhrajani KD, Chowdhury AR, Dutta KK. 1992. Testicular toxicity of methylmercury: Analysis of cellular distribution pattern at different stages of the seminiferous epithelium. *Reprod Toxicol* 6(4):355-361.
- \*Vahter M, Mottet NK, Friberg L, et al. 1994. Speciation of mercury in the primate blood and brain following long-term exposure to methyl mercury. *Toxicol Appl Pharmacol* 124:221-229.
- \*Van Delft W, Vos G. 1988. Comparison of digestion procedures for the determination of mercury in soils by cold-vapour atomic absorption spectrometry. *Anal Chim Acta* 209(1-2):147-156.

## 8. REFERENCES

- \*Van der Meide PH, De Labie MC, Botman CA, et al. 1993. Mercuric chloride down-regulates T cell interferon-gamma production in Brown Norway but not in Lewis rats: Role of glutathione. *Eur J Immunol* 23(3):675-681.
- \*Vandal GM, Mason RP, Fitzgerald WF. 1991. Cycling of volatile mercury in temperate lakes. *Water, Air, Soil Pollution* 56:791-803.
- \*Veien NK. 1990. Stomatitis and systemic dermatitis from mercury in amalgam dental restorations. *Dermatol Clin* 8(1):157-160.
- \*Verberk M, Salle H, Kemper C. 1986. Tremor in workers with low exposure to metallic mercury. *Am Ind Hyg Assoc J* 47:559-562.
- \*Verity MA, Sarafian T. 1991. Role of oxidative injury in the pathogenesis of methylmercury neurotoxicity. In: Suzuki T, Imura N, Clarkson TW, eds. *Advances in mercury toxicology*. New York, NY: Plenum Press, 209-222
- \*Vermeir G, Vandecasteele C, Dams R. 1989. Microwave dissolution for the determination of mercury in biological samples. *Anal Chim Acta* 220(1):257-261.
- \*Vermeir G, Vandecasteele C, Dams R. 1991a. Atomic fluorescence spectrometry combined with reduction aeration for the determination of mercury in biological samples. *Anal Chim Acta* 242(2):203-208.
- \*Vermeir G, Vandecasteele C, Dams R. 1991b. Atomic fluorescence spectrometry for the determination of mercury in biological samples. In: Aitio A, ed. *Trace elements in health and disease, International Symposium, Espoo, Finland, June 5-8, 1990*. Boca Raton, FL: CRC Press Inc, 29-36.
- \*Vermeir G, Vandecasteele C, Temmerman E, et al. 1988. Determination of mercury in biological materials by CV (cold-vapour) AAS after wet digestion. *Mikrochim Acta* 3:305-313.
- \*Verschaeve L, Kirsch-Volders M, Susanne C, et al. 1976. Genetic damage induced by occupationally low mercury exposure. *Environ Res* 12:306-316.
- \*Verschaeve L, Tassignon J-P, Lefevre M, et al. 1979. Cytogenic investigation on leukocytes of workers exposed to metallic mercury. *Environ Mutagen* 1:259-268.
- \*Verschoor MA, Herber R FM, Zielhuis RL. 1988. Urinary mercury levels and early changes in kidney function in dentists and dental assistants. *Community Dent Oral Epidemiol* 16(3):148-152.
- \*Verschuere K. 1983. *Handbook of environmental data on organic chemicals*. 2nd ed. New York, NY: Van Nostrand Reinhold Co., 991.
- \*Verschuuren HG, Kroes R, Den Tonkelaar EM, et al. 1976. Toxicity of methylmercury chloride in rats. III. Long-term toxicity study. *Toxicol* 6:107-123.
- \*Vesterberg O. 1991. Automatic method for quantitation of mercury in blood, plasma and urine. *J Biochem Biophys Methods* 23(3):227-236.

## 8. REFERENCES

- \*Vieira I, Sonnier M, Cresteil T. 1996. Developmental expression of CYP2E1 in the human liver: hypermethylation control of gene expression during the neonatal period. *European Journal of Biochemistry* 238:476-483.
- \*Vignani R, Milanese C, Di Simplicio P. 1992. Disruption of cytoskeleton by methylmercury in cultured CHO cells. *Toxicol in Vitro* 6(1):61-70.
- \*Vimy MJ, Lorscheider FL. 1985. Serial measurement of intra-oral air mercury: Estimation of daily dose from dental amalgam. *J Dent Res* 64:1072-1075. (As cited in Weiner and Nylander 1995).
- \*Vogel DG, Margolis RL, Mottet NK. 1985. The effects of methyl mercury binding to microtubules. *Toxicol Appl Pharmacol* 80:473-486.
- \*Vogel DG, Margolis RL, Mottet NK. 1989. Analysis of methyl mercury binding sites on tubulin subunits and microtubules. *Pharmacol Toxicol* 64(2):196-201.
- \*Votaw AL, Zey J. 1991. Vacuuming a mercury-contaminated dental office may be hazardous to your health. *The Dental Assistant* January/February: 27-29.
- \*Vroom FQ, Greer M. 1972. Mercury vapor intoxication. *Brain* 95:305-318.
- \*WAC. 1988. Wisconsin Administrative Code. Control of hazardous pollutants. Chapter NR 455. Department of Natural Resources.
- \*Wada O, Toyokawa K, Suzuki T, et al. 1969. Response to a low concentration of mercury vapor: Relation to human porphyrin metabolism. *Arch Environ Health* 19:485-488.
- \*Wagemann R, Lockhart WL, Welch H, et al. 1995. Arctic marine mammals as integrators and indicators of mercury in the arctic. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant. Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994.* Boston, MA: Kluwer Academic Publishers, 683-693.
- \*Wakita Y. 1987. Hypertension induced by methylmercury in rats. *Toxicol Appl Pharmacol* 89:144-147.
- \*Walker BL, Cooper CD. 1992. Air pollution emission factors for medical waste incinerators. *J Air Waste Manag Assoc* 42(6):784-791.
- \*Wands JR, Weiss SW, Yardley JH, et al. 1974. Chronic inorganic mercury poisoning due to laxative abuse--a clinical and ultrastructural study. *Am J Med* 57:92-101.
- \*Wang JS, Huang PM, Liaw WK, et al. 1991. Kinetics of the desorption of mercury from selected fresh water sediments as influenced by chloride. *Water, Air, Soil Pollution* 56:533-542.
- \*Wankhade HK, Garg AN. 1989. Neutron-activation analysis of coal dust particulates and atmospheric pollution. *Indian J Environ Health* 31(2):125-130.
- \*Warfvinge K. 1995. Mercury distribution in the mouse brain after mercury vapour exposure. *Int J Exp Path* 76:29-35.



## 8. REFERENCES

- \*Warfvinge K, Hansson H, Hultman P. 1995. Systemic autoimmunity due to mercury vapor exposure in genetically susceptible mice: Dose-responses studies. *Toxicol Appl Pharm* 132:299-309.
- \*Warfvinge K, Hua J, Berlin M. 1992. Mercury distribution in the rat brain after mercury vapor exposure. *Toxicol Appl Pharmacol* 117(1):46-52.
- \*Warkany J, Hubbard DM. 1953. Acrodynia and mercury. *J Pediat* 42:365-386.
- \*Warren CJ, Dudas MJ. 1992. Acidification adjacent to an elemental sulfur stockpile: II. Trace element redistribution. *Can J Soil Sci* 72(2):127-134.
- \*Watras CJ, Bloom NS. 1992. Mercury and methylmercury in individual zooplankton: Implications for bioaccumulation. *Limnol Oceanogr* 37:1313-1318.
- \*WDNR. 1987. Surface water quality criteria for toxic substances, 1987. Wisconsin Department of Natural Resources. Order of the State of Wisconsin Natural Resources Board repealing, renumbering, renumbering and amending, amending, repealing and recreating, and creating rules. Section 27, Chapter NR 105.
- \*Weast RC, ed. 1988. *CRC Handbook of Chemistry and Physics*. 69th ed. Boca Raton, FL: CRC Press Inc., B-106.
- Webb M, Holt D. 1982. Endogenous metal binding proteins in relation to the differences in absorption and distribution of mercury in newborn and adult rats. *Arch Toxicol* 49:237-245
- \*Weigert P. 1991. Metal loads of food of vegetable origin including mushrooms. In: Merian E, ed. *Metals and their compounds in the environment*. VCH: Weinheim, Fed Rep Ger, 449-468.
- \*Weihe P, Grandjean P, Debes F, et al. 1996. Health implications for Faroe Islanders of heavy metals and PCBs from pilot whales. *The Science of the Total Environment* 186:141-148.
- \*Weiner JA, Nylander M. 1995. An estimation of the uptake of mercury from amalgam fillings based on urinary excretion of mercury in Swedish subjects. *Sci Total Environ* 168(3):255-65.
- \*Weiner JA, Nylander M, Berglund F. 1990. Does mercury from amalgam restorations constitute a health hazard? *Sci Total Environ* 99(1-2):1-22.
- \*Weiss G, ed. 1986. *Hazardous chemicals data book*. 2nd ed. New Jersey: Noyes Data Corp, 650-662.
- \*Weiss SH, Wands JR, Yardley JH. 1973. Demonstration by electron defraction of black mercuric sulfide (b-HgS) in a case of "melanosis coli and black kidneys" caused by chronic inorganic mercury poisoning. *Lab Invest* 401-402.
- \*Welsh SO. 1979. The protective effect of vitamin E and N, n-diphenyl-p-phenylenediamine against methylmercury toxicity in the rat. *J Nutr* 109:1673-1681.
- \*Wendroff AP. 1990. Domestic mercury pollution. *Nature* 347:623.

## 8. REFERENCES

- \*Wendroff AP. 1991. Bringing attention to mercury threat. Society for Applied Anthropology newsletter. 2(1):3-5.
- Wendroff AP. 1995. Magico-religious mercury use and cultural sensitivity. *AJPH* 85(3):409-410.
- \*West I, Lim J. 1968. Mercury poisoning among workers in California mercury mills--preliminary report. *J Occup Med* 10:697-701.
- \*West JR, Smith HW, Chasis H. 1948. Glomerular filtration rate, effective renal blood flow, and maximal tubular excretory capacity in infancy. *J. of Pediatrics* 32a: 10-18.
- \*Wheatley B, Paradis S. 1995a. Exposure of Canadian aboriginal peoples to methylmercury. In: Porcella DB, Wheatley B, eds. *Mercury as a global pollutant . Proceedings of the Third International Conference Whistler, British Columbia, July 10-14, 1994.* Boston, MA: Kluwer Academic Publishers, 3-11.
- \*Wheatley B, Paradis S. 1995b. Exposure of Canadian Aboriginal people to methylmercury. *Water, Air, Soil Pollution* 80:3-11.
- \*White RF, Feldman RG, Moss MB, et al. 1993. Magnetic resonance imaging (MRI), neurobehavioral testing, and toxic encephalopathy: Two cases. *Environ Res* 61:117-123.
- \*WHO. 1976. *Environmental Health Criteria: Mercury.* Geneva, Switzerland: World Health Organization, 121.
- \*WHO. 1984. *Guidelines for drinking water quality. Volume 1: Recommendations.* World Health Organization.
- WHO. 1989. *Mercury - environmental aspects. Vol. 86.* Geneva, Switzerland: World Health Organization, International Programme on Chemical Safety.
- \*WHO. 1990. *Methyl mercury. Vol. 101.* Geneva, Switzerland: World Health Organization, International Programme on Chemical Safety.
- \*WHO. 1991. *Inorganic mercury. Vol. 118.* Geneva, Switzerland: World Health Organization, International Programme on Chemical Safety, 168.
- \*Widdowson EM, Dickerson JWT. 1964. Chapter 17: Chemical composition of the body. In: *Mineral metabolism: an advanced treatise volume II - the elements part A* (editors: C.L. Comar and Felix Bronner), Academic Press, New York.
- \*Wiener JG, Fitzgerald WF, Watras CJ, et al. 1990. Partitioning and bioavailability of mercury in an experimentally acidified Wisconsin Lake. In: *Symposium on Metal Chemistry and Bioavailability in Acid Waters Ninth Annual Meeting of the Society of Environmental Toxicology and Chemistry, Arlington, VA, November 16, 1988.* *Environ Toxicol Chem* 9(7):909-918.
- \*Wilhelm M, Idel H. 1996. Hair analysis in environmental medicine. *Zentralblatt Fuer Hygiene Und Umweltmedizin* 198(6):485-501.

## 8. REFERENCES

- \*Willes RF, Truelove JF, Nera EA. 1978. Neurotoxic response of infant monkeys to methylmercury. *Toxicol* 9:125-135.
- \*Willett KL, Turner RR, Beauchamp JJ. 1992. Effect of chemical form of mercury on the performance of dosed soils. *Hazardous Waste and Hazardous Materials* 9(3):275-288.
- \*Williams CH, Arscott LD, Shulz GE. 1982. Amino acid sequence homology between pig heart lipoamide dehydrogenase and human erythrocyte glutathione reductase. *Proc Natl Acad Sci USA* 79:2199-2201.
- \*Williams MV, Winters T, Waddel KS. 1987. *In vivo* effects of mercury (II) on deoxyuridine triphosphate nucleotidohydrolase, DNA polymerase (alpha, beta), and uracil-DNA glycosylase activities in cultured human cells: Relationship to DNA damage, DNA repair, and cytotoxicity. *Mol Pharmacol* 31:200-207.
- \*Williamson AM, Teo R, Sanderson J. 1982. Occupational mercury exposure and its consequences for behavior. *Int Arch Occup Environ Health* 50:273-286.
- \*Williston SH. 1968. Mercury in the atmosphere. *J Geophys Res* 73:7051-7055.
- \*Wilson BL, Mitchell DL. 1991. Trace metal study of sediment samples near electrical generating facility. *J Environ Sci Health*. A26(4):493-509.
- \*Winger PV, Schultz DP, Johnson WW. 1990. Environmental contaminant concentrations in biota from the lower Savannah river, Georgia and South Carolina. *Arch Environ Contam Toxicol* 19:101-117.
- \*Winship KA. 1985. Toxicity of mercury and its inorganic salts. *Adverse Drug React Acute Poisoning Review* 4(3):129-160.
- \*Wolfe RJ, Walker RJ. 1987. Subsistence economies in Alaska: Productivity, geography, and development impacts. *Arctic Anthropology* 24(2):56-81.
- \*Wolff MS. 1983. Occupationally derived chemicals in breast milk. *Am J Ind Med* 4:259-281.
- \*Wong PK. 1988. Mutagenicity of heavy metals. *Bull Environ Contam Toxicol* 40(4):597-603.
- \*Wood JM. 1974. Biological cycles for toxic elements in the environment. *Science* 183:1049-1052.
- \*Woods JS. 1996. Altered porphyrin metabolism as a biomarker of mercury exposure and toxicity. *Can J Physiol Pharmacol* 74(2):210-215.
- \*Woods JS, Bowers MA, Davis HA. 1991. Urinary porphyrin profiles as biomarkers of trace-metal exposure and toxicity: Studies on urinary porphyrin excretion patterns in rats during prolonged exposure to methyl mercury. *Toxicol Appl Pharmacol* 110(3):464-476.
- \*Wren C. 1992. Relationship of mercury levels in sportfish with lake sediment and water quality variables. Toronto: Ontario Environmental Research Program. Govt Reports Announcements and Index (GRA&I) Issue 08.
- \*Wright N, Yeoman WB, Carter GF. 1980. Massive oral ingestion of elemental mercury without poisoning [letter]. *Lancet* 1(8161):206.

## 8. REFERENCES

- \*Wulf HC, Kromann N, Kousgaard N, et al. 1986. Sister chromatid exchange (SCE) in Greenlandic Eskimos: Dose-response relationship between SCE and seal diet, smoking, and blood cadmium and mercury concentrations. *Sci Total Environ* 48:81-94.
- \*Yamada H, Miyahara T, Kozuka H, et al. 1993. Potentiating effects of organomercuries on clastogen-induced chromosome aberrations in cultured Chinese hamster cells. *Mutat Res* 290(2):281-291.
- \*Yamaguchi S, Matsumoto H, Hoshide M, et al. 1971. Occurrence of alkylmercury compound in caustic soda factory. *Arch Environ Health* 23:196-201.
- \*Yamaguchi S, Nunotani H. 1974. Transplacental transport of mercurials in rats at the subclinical dose level. *Environ Physiol Biochem* 4:7-15.
- \*Yan D, Zhang J, Schwedt G. 1989. [Ion-chromatographic trace analysis of mercury, cadmium, and zinc by post-column derivatization with a water-soluble porphyrin.] *Fresenius Z Anal Chem* 334(6):507-510.
- \*Yanagisawa M, Ida K, Kitagawa K. 1989. Direct determination of mercury in rat tissues by atomic-absorption spectrometry with a separative column atomizer. *Anal Sci* 5(6):765-766.
- \*Yang MG, Wang JHC, Garcia JD, et al. 1973. Mammary transfer of 203 Hg from mothers to brains of nursing rats. *Proc Soc Exp Biol Med* 142:723-727.
- \*Yang Y-J, Huang C-C, Shih T-S, et al. 1994. Chronic elemental mercury intoxication: clinical and field studies in lampsocket manufactures. *Occup Environ Med* 51(4):267-270.
- \*Yannai S, Berdicevsky I, Duek L. 1991. Transformations of inorganic mercury by *Candida albicans* and *Saccharomyces cerevisiae*. *Appl Environ Microbiology* 57(1):245-247.
- \*Yannai S, Sachs KM. 1993. Absorption and accumulation of cadmium, lead and mercury from foods by rats. *Food Chem Toxicol* 31(5):351-355.
- \*Yasuda Y, Datu AR, Hirata S, et al. 1985. Characteristics of growth and palatal shelf development in ICR mice after exposure to methylmercury. *Teratology* 32:273-286.
- Yasutake A, Adachi T, Hirayama K, et al. 1991a. Integrity of the blood-brain barrier system against methylmercury acute toxicity. *Eisei Kagaku* 37(5):355-362.
- \*Yasutake A, Hirayama Y, Inouye M. 1991b. Sex differences of nephrotoxicity by methylmercury in mice. In: Bach PH, et al., eds. *Nephrotoxicity: Mechanisms, early diagnosis, and therapeutic management*. Fourth International Symposium on Nephrotoxicity, Guilford, England, UK, 1989. New York, NY: Marcel Dekker, Inc., 389-396.
- \*Yeates KO, Mortensen ME. 1994. Acute and chronic neuropsychological consequences of mercury vapor poisoning in two early adolescents. *J Clin Exper Neuropsychology* 16(2):209-222.
- \*Yeoh TS, Lee AS, Lee HS. 1986. Absorption of mercuric sulphide following oral administration in mice. *Toxicology* 41(1):107-111.

## 8. REFERENCES

- \*Yeoh TS, Lee HS, Lee AS. 1989. Gastrointestinal absorption of mercury following oral administration of cinnabar in a traditional Chinese medicine. *Asia Pac J Pharmacol* 4(2):69-73.
- \*Yess NJ. 1993. U.S. Food and Drug Administration survey of methyl mercury in canned tuna. *J AOAC Int* 76(1):36-38.
- \*Yip RK, Chang LW. 1981. Vulnerability of dorsal route neurons and fibers toward methylmercury toxicity: A morphological evaluation. *Environ Res* 26:152-167.
- \*Yoshida M. 1985. Relation of mercury exposure to elemental mercury levels in the urine and blood. *Scand J Work Environ Health* 11:33-37.
- \*Yoshida M, Satoh H, Aoyama H, et al. 1989. Distribution of mercury in neonatal guinea pigs after exposure to mercury vapor. *Bull Environ Contam Toxicol* 43(5):697-704.
- \*Yoshida M, Satoh H, Kishimoto T. 1992. Exposure to mercury via breast milk in suckling offspring of maternal guinea pigs exposed to mercury vapor after parturition. *J Toxicol Environ Health* 35(2):135-139.
- \*Yoshida M, Satoh H, Kojima S, et al. 1990. Retention and distribution of mercury in organs of neonatal guinea pigs after *in utero* exposure to mercury vapor. *J Trace Elem Exp Med* 3(3):219-226.
- \*Yoshida M, Satoh H, Kojima S, et al. 1991. Metallothionein concentrations and organ retention of mercury in the liver and kidney of the neonatal guinea pig after exposure to mercury vapor. *Tohoku J Exp Med* 164(1):13-22.
- \*Yoshida M, Watanabe C, Satoh H, et al. 1994. Milk transfer and tissue uptake of mercury in suckling offspring after exposure of lactating maternal guinea pigs to inorganic or methylmercury. *Arch Toxicol* 68:174-178.
- \*Yoshida M, Yamamura Y. 1982. Elemental mercury in urine from workers exposed to mercury vapor. *Int Arch Occup Environ Health* 51:99-104.
- \*Yuan Y, Atchison WD. 1994. Comparative effects of inorganic divalent mercury, methylmercury and phenylmercury on membrane excitability and synaptic transmission of cal neurons in hippocampal slices of the rat. *Neurotoxicology* 15(2):403-411.
- \*Zalups RK. 1993. Influence of 2,3-dimercaptopropane-1-sulfonate (dmpps) and meso-2,3-dimercaptosuccinic acid (dmsa) on the renal disposition of mercury in normal and uninephrectomized rats exposed to inorganic mercury. *J Pharmacol Exper Thera* 267(2):791-800.
- \*Zalups RK, Cherian MG. 1992. Renal metallothionein metabolism after a reduction of renal mass: II. Effects of zinc pretreatment on the renal toxicity and intrarenal accumulation of inorganic mercury. *Toxicology* 71(1-2):103-117.
- Zalups RK, Cherian MG, Barfuss DW. 1993. Mercury-metallothionein and the renal accumulation and handling of mercury. *Toxicol* 83(1-3):61-78.
- \*Zalups RK, Lash LH. 1994. Advance in understanding the renal transport and toxicity of mercury. *J Toxicol Environ Health* 42:1-44.

## 8. REFERENCES

- \*Zanoli P, Truzzi C, Veneri C, et al. 1994. Methyl mercury during late gestation affects temporarily the development of cortical muscarinic receptors in rat offspring. *Pharmacol Toxicol* 75:261-264.
- \*Zasukhina GD, Vasilyeva IM, Sdirkova NI, et al. 1983. Mutagenic effect of thallium and mercury salts on rodent cells with different repair activities. *Mutat Res* 124:163-173.
- \*Zayas LH, Ozuah PO. 1996. Mercury use in Espiritismo: A survey of botanicas. *American Journal of Public Health* 86(1):111-112.
- \*Zelikoff JT, Bertin JE, Burbacher TM, et al. 1995. Health risks associated with prenatal metal exposure. *Fund Appl Toxicol* 25: 161-170.
- \*Zhuang G, Wang Y, Zhi M, et al. 1989. Determination of arsenic, cadmium, mercury, copper and zinc in biological samples by radiochemical neutron-activation analysis. *J Radioanal Nucl Chem* 129(2):459-464.
- \*Ziegler EE, Edwards BB, Jensen RL et al. 1978. Absorption and retention of lead by infants. *Pediatr Res* 12:29-34.
- \*Zillioux EJ, Porcella DB, Benoit JM. 1993. Mercury cycling and effects in fresh water wetland ecosystems. *Environ Tox Chem* 12:2245-2264.
- \*Zirschky J. 1990. Employee transported contaminant releases. *Hazardous Waste and Hazardous Materials* 7(2):201-209.
- \*Zirschky J, Witherell L. 1987. Cleanup of mercury contamination of thermometer workers' homes. *Am Ind Hyg Assoc Journal* 48(1):81-84.

## 9. GLOSSARY

**Absorption**—The taking up of liquids by solids, or of gases by solids or liquids.

**Acute Exposure**—Exposure to a chemical for a duration of 14 days or less, as specified in the Toxicological Profiles.

**Adsorption**—The adhesion in an extremely thin layer of molecules (as of gases, solutes, or liquids) to the surfaces of solid bodies or liquids with which they are in contact.

**Adsorption Coefficient ( $K_{oc}$ )**—The ratio of the amount of a chemical adsorbed per unit weight of organic carbon in the soil or sediment to the concentration of the chemical in solution at equilibrium.

**Adsorption Ratio (Kd)**—The amount of a chemical adsorbed by a sediment or soil (i.e., the solid phase) divided by the amount of chemical in the solution phase, which is in equilibrium with the solid phase, at a fixed solid/solution ratio. It is generally expressed in micrograms of chemical sorbed per gram of soil or sediment.

**Benchmark Dose (BMD)**—is usually defined as the lower confidence limit on the dose that produces a specified magnitude of changes in a specified adverse response. For example, a  $BMD_{10}$  would be the dose at the 95% lower confidence limit on a 10% response, and the benchmark response (BMR) would be 10%. The BMD is determined by modeling the dose response curve in the region of the dose response relationship where biologically observable data are feasible.

**Benchmark Dose Model**—is a statistical dose-response model applied to either experimental toxicological or epidemiological data to calculate a BMD.

**Bioconcentration Factor (BCF)**—The quotient of the concentration of a chemical in aquatic organisms at a specific time or during a discrete time period of exposure divided by the concentration in the surrounding water at the same time or during the same period.

**Biomarkers**—are broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility.

**Cancer Effect Level (CEL)**—The lowest dose of chemical in a study, or group of studies, that produces significant increases in the incidence of cancer (or tumors) between the exposed population and its appropriate control.

**Carcinogen**—A chemical capable of inducing cancer.

**Case-Control Study**—A type of epidemiological study which examines the relationship between a particular outcome (disease or condition) and a variety of potential causative agents (such as toxic chemicals). In a case-controlled study, a group of people with a specified and well-defined outcome is identified and compared to a similar group of people without outcome.

**Case Report**—describes a single individual with a particular disease or exposure. These may suggest some potential topics for scientific research but are not actual research studies.

## 9. GLOSSARY

**Case Series**—describes the experience of a small number of individuals with the same disease or exposure. These may suggest potential topics for scientific research but are not actual research studies.

**Ceiling Value**—A concentration of a substance that should not be exceeded, even instantaneously.

**Chronic Exposure**—Exposure to a chemical for 365 days or more, as specified in the Toxicological Profiles.

**Cohort Study**—A type of epidemiological study of a specific group or groups of people who have had a common insult (e.g., exposure to an agent suspected of causing disease or a common disease) and are followed forward from exposure to outcome. At least one exposed group is compared to one unexposed group.

**Cross-sectional Study**—A type of epidemiological study of a group or groups which examines the relationship between exposure and outcome to a chemical or to chemicals at one point in time.

**Data Needs**—substance-specific informational needs that if met would reduce the uncertainties of human health assessment.

**Developmental Toxicity**—The occurrence of adverse effects on the developing organism that may result from exposure to a chemical prior to conception (either parent), during prenatal development, or postnatally to the time of sexual maturation. Adverse developmental effects may be detected at any point in the life span of the organism.

**Dose-Response Relationship**—the quantitative relationship between the amount of exposure to a toxicant and the incidence of the adverse effects.

**Embryotoxicity and Fetotoxicity**—Any toxic effect on the conceptus as a result of prenatal exposure to a chemical; the distinguishing feature between the two terms is the stage of development during which the insult occurs. The terms, as used here, include malformations and variations, altered growth, and in utero death.

**Environmental Protection Agency (EPA) Health Advisory**—An estimate of acceptable drinking water levels for a chemical substance based on health effects information. A health advisory is not a legally enforceable federal standard, but serves as technical guidance to assist federal, state, and local officials.

**Epidemiology**—refers to the investigation of factors that determine the frequency and distribution of disease or other health-related conditions within a defined human population during a specified period.

**Genotoxicity**—a specific adverse effect on the genome of living cells that, upon the duplication of affected cells, can be expressed as a mutagenic, clastogenic or carcinogenic event because of specific alteration of the molecular structure of the genome.

**Half-life**—a measure of rate for the time required to eliminate one half of a quantity of a chemical from the body or environmental media.

**Immediately Dangerous to Life or Health (IDLH)**—The maximum environmental concentration of a contaminant from which one could escape within 30 minutes without any escape-impairing symptoms or irreversible health effects.



## 9. GLOSSARY

**Incidence**—The ratio of individuals in a population who develop a specified condition to the total number of individuals in that population who could have developed that condition in a specified time period.

**Intermediate Exposure**—Exposure to a chemical for a duration of 15-364 days, as specified in the Toxicological Profiles.

**Immunological Effects**—are functional changes in the immune response.

**Immunologic Toxicity**—The occurrence of adverse effects on the immune system that may result from exposure to environmental agents such as chemicals.

**In Vitro**—Isolated from the living organism and artificially maintained, as in a test tube.

**In Vivo**—Occurring within the living organism.

**Lethal Concentration<sub>(LO)</sub> (LC<sub>LO</sub>)**—The lowest concentration of a chemical in air which has been reported to have caused death in humans or animals.

**Lethal Concentration<sub>(50)</sub> (LC<sub>50</sub>)**—A calculated concentration of a chemical in air to which exposure for a specific length of time is expected to cause death in 50% of a defined experimental animal population.

**Lethal Dose<sub>(LO)</sub> (LD<sub>LO</sub>)**—The lowest dose of a chemical introduced by a route other than inhalation that has been reported to have caused death in humans or animals.

**Lethal Dose<sub>(50)</sub> (LD<sub>50</sub>)**—The dose of a chemical which has been calculated to cause death in 50% of a defined experimental animal population.

**Lethal Time<sub>(50)</sub> (LT<sub>50</sub>)**—A calculated period of time within which a specific concentration of a chemical is expected to cause death in 50% of a defined experimental animal population.

**Lowest-Observed-Adverse-Effect Level (LOAEL)**—The lowest exposure level of chemical in a study, or group of studies, that produces statistically or biologically significant increases in frequency or severity of adverse effects between the exposed population and its appropriate control.

**Lymphoreticular Effects**—represent morphological effects involving lymphatic tissues such as the lymph nodes, spleen, and thymus.

**Malformations**—Permanent structural changes that may adversely affect survival, development, or function.

**Minimal Risk Level (MRL)** —An estimate of daily human exposure to a hazardous substance that is likely to be without an appreciable risk of adverse noncancer health effects over a specified route and duration of exposure.

**Modifying Factor (MF)**—A value (greater than zero) that is applied to the derivation of a minimal risk level (MRL) to reflect additional concerns about the database that are not covered by the uncertainty factors. The default value for a MF is 1.

## 9. GLOSSARY

**Morbidity**—State of being diseased; morbidity rate is the incidence or prevalence of disease in a specific population.

**Mortality**—Death; mortality rate is a measure of the number of deaths in a population during a specified interval of time.

**Mutagen**—A substance that causes mutations. A mutation is a change in the DNA sequence of a cell's DNA. Mutations can lead to birth defects, miscarriages, or cancer.

**Necropsy**—The gross examination of the organs and tissues of a dead body to determine the cause of death or pathological conditions.

**Neurotoxicity**—The occurrence of adverse effects on the nervous system following exposure to a chemical.

**No-Observed-Adverse-Effect Level (NOAEL)**—The dose of a chemical at which there were no statistically or biologically significant increases in frequency or severity of adverse effects seen between the exposed population and its appropriate control. Effects may be produced at this dose, but they are not considered to be adverse.

**Octanol-Water Partition Coefficient ( $K_{ow}$ )**—The equilibrium ratio of the concentrations of a chemical in *n*-octanol and water, in dilute solution.

**Odds Ratio**—a means of measuring the association between an exposure (such as toxic substances and a disease or condition) which represents the best estimate of relative risk (risk as a ratio of the incidence among subjects exposed to a particular risk factor divided by the incidence among subjects who were not exposed to the risk factor). An odds ratio of greater than 1 is considered to indicate greater risk of disease in the exposed group compared to the unexposed.

**Organophosphate or Organophosphorus Compound**—a phosphorus containing organic compound and especially a pesticide that acts by inhibiting cholinesterase.

**Permissible Exposure Limit (PEL)**—An Occupational Safety and Health Administration (OSHA) allowable exposure level in workplace air averaged over an 8-hour shift of a 40 hour workweek.

**Pesticide**—general classification of chemicals specifically developed and produced for use in the control of agricultural and public health pests.

**Pharmacokinetics**—is the science of quantitatively predicting the fate (disposition) of an exogenous substance in an organism. Utilizing computational techniques, it provides the means of studying the absorption, distribution, metabolism and excretion of chemicals by the body.

**Pharmacokinetic Model**—is a set of equations that can be used to describe the time course of a parent chemical or metabolite in an animal system. There are two types of pharmacokinetic models: data-based and physiologically based. A data-based model divides the animal system into a series of compartments which, in general, do not represent real, identifiable anatomic regions of the body whereby the physiologically based model compartments represent real anatomic regions of the body.

## 9. GLOSSARY

**Physiologically Based Pharmacodynamic (PBPD) Model**—is a type of physiologically based dose-response model which quantitatively describes the relationship between target tissue dose and toxic end points. These models advance the importance of physiologically based models in that they clearly describe the biological effect (response) produced by the system following exposure to an exogenous substance.

**Physiologically Based Pharmacokinetic (PBPK) Model**—is comprised of a series of compartments representing organs or tissue groups with realistic weights and blood flows. These models require a variety of physiological information: tissue volumes, blood flow rates to tissues, cardiac output, alveolar ventilation rates and, possibly membrane permeabilities. The models also utilize biochemical information such as air/blood partition coefficients, and metabolic parameters. PBPK models are also called biologically based tissue dosimetry models.

**Prevalence**—The number of cases of a disease or condition in a population at one point in time.

**Prospective Study**—a type of cohort study in which the pertinent observations are made on events occurring after the start of the study. A group is followed over time.

$q_1^*$ —The upper-bound estimate of the low-dose slope of the dose-response curve as determined by the multistage procedure. The  $q_1^*$  can be used to calculate an estimate of carcinogenic potency, the incremental excess cancer risk per unit of exposure (usually  $\mu\text{g/L}$  for water,  $\text{mg/kg/day}$  for food, and  $\mu\text{g/m}^3$  for air).

**Recommended Exposure Limit (REL)**—A National Institute for Occupational Safety and Health (NIOSH) time-weighted average (TWA) concentrations for up to a 10-hour workday during a 40-hour workweek.

**Reference Concentration (RfC)**—An estimate (with uncertainty spanning perhaps an order of magnitude) of a continuous inhalation exposure to the human population (including sensitive subgroups) that is likely to be without an appreciable risk of deleterious noncancer health effects during a lifetime. The inhalation reference concentration is for continuous inhalation exposures and is appropriately expressed in units of  $\text{mg/m}^3$  or ppm.

**Reference Dose (RfD)**—An estimate (with uncertainty spanning perhaps an order of magnitude) of the daily exposure of the human population to a potential hazard that is likely to be without risk of deleterious effects during a lifetime. The RfD is operationally derived from the No-Observed-Adverse-Effect Level (NOAEL- from animal and human studies) by a consistent application of uncertainty factors that reflect various types of data used to estimate RfDs and an additional modifying factor, which is based on a professional judgment of the entire database on the chemical. The RfDs are not applicable to nonthreshold effects such as cancer.

**Reportable Quantity (RQ)**—The quantity of a hazardous substance that is considered reportable under the Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA). Reportable quantities are (1) 1 pound or greater or (2) for selected substances, an amount established by regulation either under CERCLA or under Section 311 of the Clean Water Act. Quantities are measured over a 24-hour period.

## 9. GLOSSARY

**Reproductive Toxicity**—The occurrence of adverse effects on the reproductive system that may result from exposure to a chemical. The toxicity may be directed to the reproductive organs and/or the related endocrine system. The manifestation of such toxicity may be noted as alterations in sexual behavior, fertility, pregnancy outcomes, or modifications in other functions that are dependent on the integrity of this system.

**Retrospective Study**—A type of cohort study based on a group of persons known to have been exposed at some time in the past. Data are collected from routinely recorded events, up to the time the study is undertaken. Retrospective studies are limited to casual factors that can be ascertained from existing records and/or examining survivors of the cohort.

**Risk**—the possibility or chance that some adverse effect will result from a given exposure to a chemical.

**Risk Factor**—An aspect of personal behavior or lifestyle, an environmental exposure, or an inborn or inherited characteristic, that is associated with an increased occurrence of disease or other health-related event or condition.

**Risk Ratio**—The ratio of the risk among persons with specific risk factors compared to the risk among persons without risk factors. A risk ratio greater than 1 indicates greater risk of disease in the exposed group compared to the unexposed.

**Short-Term Exposure Limit (STEL)**—The American Conference of Governmental Industrial Hygienists (ACGIH) maximum concentration to which workers can be exposed for up to 15 min continually. No more than four excursions are allowed per day, and there must be at least 60 min between exposure periods. The daily Threshold Limit Value - Time Weighted Average (TLV-TWA) may not be exceeded.

**Target Organ Toxicity**—This term covers a broad range of adverse effects on target organs or physiological systems (e.g., renal, cardiovascular) extending from those arising through a single limited exposure to those assumed over a lifetime of exposure to a chemical.

**Teratogen**—A chemical that causes structural defects that affect the development of an organism.

**Threshold Limit Value (TLV)**—An American Conference of Governmental Industrial Hygienists (ACGIH) concentration of a substance to which most workers can be exposed without adverse effect. The TLV may be expressed as a Time Weighted Average (TWA), as a Short-Term Exposure Limit (STEL), or as a ceiling limit (CL).

**Time-Weighted Average (TWA)**—An allowable exposure concentration averaged over a normal 8-hour workday or 40-hour workweek.

**Toxic Dose<sub>(50)</sub> (TD<sub>50</sub>)**—A calculated dose of a chemical, introduced by a route other than inhalation, which is expected to cause a specific toxic effect in 50% of a defined experimental animal population.

**Toxicokinetic**—The study of the absorption, distribution and elimination of toxic compounds in the living organism.

## 9. GLOSSARY

**Uncertainty Factor (UF)**—A factor used in operationally deriving the Minimal Risk Level (MRL) or Reference Dose (RfD) or Reference Concentration (RfC) from experimental data. UFs are intended to account for (1) the variation in sensitivity among the members of the human population, (2) the uncertainty in extrapolating animal data to the case of human, (3) the uncertainty in extrapolating from data obtained in a study that is of less than lifetime exposure, and (4) the uncertainty in using Lowest-Observed-Adverse-Effect Level (LOAEL) data rather than No-Observed-Adverse-Effect Level (NOAEL) data. A default for each individual UF is 10; if complete certainty in data exists, a value of one can be used; however a reduced UF of three may be used on a case-by-case basis, three being the approximate logarithmic average of 10 and 1.

**Xenobiotic**—any chemical that is foreign to the biological system.



## APPENDIX A

### ATSDR MINIMAL RISK LEVEL AND WORKSHEETS

The Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) [42 U.S.C. 9601 et seq.], as amended by the Superfund Amendments and Reauthorization Act (SARA) [Pub. L. 99-499], requires that the Agency for Toxic Substances and Disease Registry (ATSDR) develop jointly with the U.S. Environmental Protection Agency (EPA), in order of priority, a list of hazardous substances most commonly found at facilities on the CERCLA National Priorities List (NPL); prepare toxicological profiles for each substance included on the priority list of hazardous substances; and assure the initiation of a research program to fill identified data needs associated with the substances.

The toxicological profiles include an examination, summary, and interpretation of available toxicological information and epidemiologic evaluations of a hazardous substance. During the development of toxicological profiles, Minimal Risk Levels (MRLs) are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration for a given route of exposure. An MRL is an estimate of the daily human exposure to a hazardous substance that is likely to be without appreciable risk of adverse noncancer health effects over a specified duration of exposure. MRLs are based on noncancer health effects only and are not based on a consideration of cancer effects. These substance-specific estimates, which are intended to serve as screening levels, are used by ATSDR health assessors to identify contaminants and potential health effects that may be of concern at hazardous waste sites. It is important to note that MRLs are not intended to define clean-up or action levels.

MRLs are derived for hazardous substances using the no-observed-adverse-effect level/uncertainty factor approach. They are below levels that might cause adverse health effects in the people most sensitive to such chemical-induced effects. MRLs are derived for acute (1–14 days), intermediate (15–364 days), and chronic (365 days and longer) durations and for the oral and inhalation routes of exposure. Currently, MRLs for the dermal route of exposure are not derived because ATSDR has not yet identified a method suitable for this route of exposure. MRLs are generally based on the most sensitive chemical-induced end point considered to be of relevance to humans. Serious health effects (such as irreparable damage to the liver or kidneys, or birth defects) are not used as a basis for establishing MRLs. Exposure to a level above the MRL does not mean that adverse health effects will occur.

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MRLs are intended only to serve as a screening tool to help public health professionals decide where to look more closely. They may also be viewed as a mechanism to identify those hazardous waste sites that are not expected to cause adverse health effects. Most MRLs contain a degree of uncertainty because of the lack of precise toxicological information on the people who might be most sensitive (e.g., infants, elderly, nutritionally or immunologically compromised) to the effects of hazardous substances. ATSDR uses a conservative (i.e., protective) approach to address this uncertainty consistent with the public health principle of prevention. Although human data are preferred, MRLs often must be based on animal studies because relevant human studies are lacking. In the absence of evidence to the contrary, ATSDR assumes that humans are more sensitive to the effects of hazardous substance than animals and that certain persons may be particularly sensitive. Thus, the resulting MRL may be as much as a hundredfold below levels that have been shown to be nontoxic in laboratory animals.

Proposed MRLs undergo a rigorous review process: Health Effects/MRL Workgroup reviews within the Division of Toxicology, expert panel peer reviews, and agencywide MRL Workgroup reviews, with participation from other federal agencies and comments from the public. They are subject to change as new information becomes available concomitant with updating the toxicological profiles. Thus, MRLs in the most recent toxicological profiles supersede previously published levels. For additional information regarding MRLs, please contact the Division of Toxicology, Agency for Toxic Substances and Disease Registry, 1600 Clifton Road, Mailstop E-29, Atlanta, Georgia 30333.



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**MINIMAL RISK LEVEL (MRL) WORKSHEET**

Chemical Name: Mercury (metallic, vapor)  
CAS Number: 7439-97-6  
Date: June 15, 2001  
Profile Status: Final Draft  
Route:  Inhalation  Oral  
Duration:  Acute  Intermediate  Chronic  
Key to figure: 21  
Species: Human

Minimal Risk Level: 0.0002  mg/kg/day  mg/m<sup>3</sup>

Reference: Fawer RF, de Ribaupierre Y, Guillemin MP, et al. 1983. Measurement of hand tremor induced by industrial exposure to metallic mercury. British Journal of Industrial Medicine 40:204-208.

Experimental design. Hand tremors were measured in 26 male workers exposed to metallic mercury and 25 control males working in the same facilities, but not exposed to mercury. Workers had been exposed to mercury through the manufacture of fluorescent tubes, chloralkali, or acetaldehyde. Mercury-exposed workers had a duration of exposure of 15.3±2.6 years, blood mercury of 41.3±3.5 micromoles Hg/L, and urinary mercury of 11.3±1.2 micromoles Hg/mole of creatinine. The mean mercury level measured using personal air monitors was 0.026±0.004 mg/m<sup>3</sup> (3 subjects were exposed to greater than 0.05 mg/m<sup>3</sup>). Hand tremors were measured in the subjects using an accelerometer attached to the dorsum of the hand both at rest and while holding 1,250 grams. The highest peak frequency of the acceleration was determined.

Effects noted in study and corresponding doses: The highest peak frequency of the tremor was greater in exposed men than in controls. The highest peak frequency corresponded significantly to duration of exposure and age. Comparison of tremors using an index of the entire spectrum of the tremor showed no differences between exposed men and controls at rest, but the changes observed between rest and load were higher in the exposed men. These changes correlated with the duration of exposure and biological indices of exposure (blood and mercury levels), but not with age.

Dose and end point used for MRL derivation: 0.026 mg/m<sup>3</sup>; increased frequency of tremors.

NOAEL  LOAEL

Uncertainty and Modifying Factors used in MRL derivation: 30

1  3  10 (for use of a minimal LOAEL)  
 1  3  10 (for extrapolation from animals to humans)  
 1  3  10 (for human variability)

Was a conversion factor used from ppm in food or water to a mg/body weight dose?

If so explain: No.

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Was a conversion used from intermittent to continuous exposure?

If so, explain: Yes. To estimate an equivalent continuous exposure concentration, the average concentration assumed for the 8 hour/day exposures was multiplied by 8/24 and 5/7 ( $0.026 \text{ mg/m}^3 \times 8/24 \text{ hours/day} \times 5/7 \text{ days/week} = 0.0062 \text{ mg/m}^3$ ). Uncertainty factors of 10 for variability in sensitivity to mercury within the human population and 3 for use of a minimal effect LOAEL in MRL derivation were then applied to the calculated  $0.0062 \text{ mg/m}^3$  value, yielding a chronic inhalation MRL of  $0.2 \text{ } \mu\text{g/m}^3$ . It should be noted that this MRL, although based upon an adult working population, is considered also to be sufficiently protective of neurodevelopmental effects in developing embryos/fetuses and children, the most sensitive subgroups for metallic mercury toxicity.

$$\begin{aligned} \text{LOAEL}_{(\text{ADJ})} &= 0.026 \text{ mg/m}^3 \times (8 \text{ hr}/24 \text{ hr}) \times (5 \text{ days}/7 \text{ days}) \\ &= 0.0062 \text{ mg/m}^3 \end{aligned}$$

$$\text{MRL} = \text{LOAEL}_{(\text{ADJ})} \div \text{UF} = 0.0062 \text{ mg/m}^3 \div 30 = 0.0002 \text{ mg/m}^3$$

If an inhalation study in animals, list the conversion factors used in determining human equivalent concentration (HEC): No.

Additional studies or pertinent information which lend support to this MRL: Inhaled metallic mercury is quickly absorbed through the lungs into the blood. Its biologic half-life in humans is approximately 60 days, with the half-life varying with the physiological compartment (e.g., 21 days in the head, versus 64 days in the kidneys; Cherian et al. 1978). Since the duration of exposure does influence the level of mercury in the body, the exposure level reported in the Fawer et al. (1983) occupational study was extrapolated from an 8-hour/day, 40-hour/workweek exposure to a level equivalent to a continuous 24 hour/day, 7 days/week exposure as might be encountered near a hazardous waste site containing metallic mercury.

The ability of long-term, low level exposure to metallic mercury to produce a degradation in neurological performance was also demonstrated in other studies. One such study (Ngim et al. 1992) attributed adverse neurological effects to a lower average level of exposure than did the Fawer et al. (1983) study; however, this study was not used in deriving a chronic inhalation MRL due to uncertainties concerning the study protocol, including methodological and reporting deficiencies. In the Ngim et al. (1992) study, dentists with an average of 5.5 years of exposure to low levels of metallic mercury were reported to have demonstrated impaired performance on several neurobehavioral tests. Exposure levels measured at the time of the study ranged from  $0.0007$  to  $0.042 \text{ mg/m}^3$ , with an average of  $0.014 \text{ mg/m}^3$ . Mean blood mercury levels among the dentists ranged from  $0.6$  to  $57 \text{ } \mu\text{g/L}$ , with a geometric mean of  $9.8 \text{ } \mu\text{g/L}$ . The performance of the dentists on finger tapping (motor speed measure), trail making (visual scanning measure), digit symbol (measure of visuomotor coordination and concentration), digit span, logical memory delayed recall (measure of visual memory), and Bender-Gestalt time (measures visuomotor coordination) were significantly poorer than controls. The exposed dentists also showed higher aggression than did controls. Furthermore, within the group of exposed dentists, significant differences were reported to have been observed between a subgroup with high mercury exposure compared to a subgroup with lower exposure. These exposure severity subgroups were not compared to controls, and average exposure levels for the subgroups were not reported. The design and reporting of this study limit its usefulness in deriving an MRL for metallic mercury. The exposure status of the subjects was known to the investigator during testing, mercury levels were not reported for controls, and methods used to correct for confounders (especially the common use in this population of traditional medicines containing mercury) were not reported. It was also unclear whether the results for the mercury exposure group were inordinately influenced or skewed by the individual dentists with the highest exposures and/or blood levels. These confounding factors precluded the use of the Ngim et al. (1992) study for the derivation of an MRL, but the study does provide support for both the premise that

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low-dose chronic exposure to metallic mercury can result in adverse health sequelae and the chronic inhalation MRL that is based upon the Fawer et al. (1983) study of occupationally exposed individuals.

Other occupational studies further support the ability of metallic mercury to induce neurologic deficits. Several studies have reported significant effects on tremor or cognitive skills among groups exposed occupationally to comparable or slightly higher (up to 0.076 mg/m<sup>3</sup>) levels (Ehrenberg et al. 1991; Piikivi et al. 1984; Roels et al. 1982). Difficulty with heel-to-toe gait was observed in thermometer plant workers subjected to mean personal breathing zone air concentrations of 0.076 mg/m<sup>3</sup> (range of 0.026–0.27 mg/m<sup>3</sup>) (Ehrenberg et al. 1991).

Tremors have also been reported in occupationally exposed workers with urinary mercury concentrations of 50–100 µg/g creatinine, and blood levels of 10–20 µg/L (Roels et al. 1982). By comparison, blood mercury levels in the Fawer et al. (1983) study averaged 41.3 and 16.6 µmol Hg/L for the exposed and control groups, respectively. Urinary mercury levels for the exposed workers in the Fawer et al. (1983) study averaged 11.3 µmol Hg/mol creatinine (about 20 µg/g creatinine), compared with 3.4 µmol/mol creatinine in the controls. In another study (Piikivi et al. 1984), decreases in performance on tests that measured intelligence (similarities) and memory (digit span and visual reproduction) were observed in chloralkali workers exposed for an average of 16.9 years (range, 10–37 years) to low levels of mercury when compared to an age-matched control group. In this study, significant differences from controls were observed on these tests among 16 workers with blood levels ranging from 75 to 344 nmol/L and urine levels ranging from 280 (about 56 µg/L) to 663 nmol/L. Abnormal nerve conduction velocities have also been observed in chloralkali plant workers at a mean urine concentration of 450 µg/L (Levine et al. 1982). These workers also experienced weakness, paresthesias, and muscle cramps. Prolongation of brainstem auditory evoked potentials was observed in workers with urinary mercury levels of 325 µg/g creatinine (Discalzi et al. 1993). Prolonged somatosensory evoked potentials were found in 28 subjects exposed to airborne mercury concentrations of 20–96 mg/m<sup>3</sup> (Langauer-Lewowicka and Kazibutowska 1989).

Agency Contact (Chemical Manager): John Risher

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**MINIMAL RISK LEVEL (MRL) WORKSHEET**

Chemical Name: Mercury inorganic  
CAS Number: 7439-97-6  
Date: June 15, 2001  
Profile Status: Final Draft  
Route:  Inhalation  Oral  
Duration:  Acute  Intermediate  Chronic  
Key to figure: 7  
Species: Rat

Minimal Risk Level: 0.007  mg/kg/day  mg/m<sup>3</sup>

Reference: NTP. 1993. NTP technical report on the toxicology and carcinogenesis studies of mercuric chloride (CAS no. 7487-94-7) in F344/N rats and B6C3F<sub>1</sub> mice (gavage studies). NTP TR408.

Experimental design: Fischer 344 rats (5/sex/group) were administered 0, 0.93, 1.9, 3.7, 7.4, or 14.8 mg Hg/kg/day as mercuric chloride once daily for 14 days, excluding weekends. The mercuric chloride was administered in deionized water via gavage. Body weights were measured and a complete necropsy was performed. Organ weights were obtained for the brain, heart, kidney, liver, lung, and thymus.

Effects noted in study and corresponding doses: The relative and absolute kidney weights were significantly increased for males exposed to at least 1.9 mg Hg/kg/day and for females exposed to at least 3.7 mg Hg/kg/day. An increased incidence of renal tubular necrosis (graded minimal in severity) was observed in 3 of 5 males and 1 of 5 females at the 3.7 mg Hg/kg/day dose level. At 7.4 mg Hg/kg/day, 5/5 males and 3/5 females had minimal-to-mild effects, and at 14.8 mg Hg/kg/day all animals exhibited mild-to-moderate effects.

Dose and end point used for MRL derivation: 0.93 mg Hg/kg/day; no renal effects.

NOAEL  LOAEL

Uncertainty Factors used in MRL derivation: 100

1  3  10 (for use of a LOAEL)  
 1  3  10 (for extrapolation from animals to humans)  
 1  3  10 (for human variability)

Was a conversion factor used from ppm in food or water to a mg/body weight dose?

If so explain: No.

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Was a conversion used from intermittent to continuous exposure?

If so, explain: Yes. To estimate an equivalent continuous exposure concentration, the average concentration was multiplied by 5 days/7 days.

$$\begin{aligned}\text{NOAEL}_{(\text{ADJ})} &= 0.93 \text{ mg/kg/day} \times (5 \text{ days}/7 \text{ days}) \\ &= 0.66 \text{ mg/kg/day}\end{aligned}$$

$$\text{MRL} = \text{NOAEL}_{(\text{ADJ})} \div \text{UF} = 0.66 \text{ mg/kg/day} \div 100 = 0.007 \text{ mg/kg/day}$$

If an inhalation study in animals, list the conversion factors used in determining human equivalent concentration (HEC): None.

Additional studies or pertinent information which lend support to this MRL: Several other studies examining the effects of oral exposure to inorganic mercury salts have also shown renal toxicity in humans as a result of acute oral exposures. Kidney effects (i.e., heavy albuminuria, hypoalbuminemia, edema, and hypercholesterolemia) have been reported after therapeutic administration of inorganic mercury (Kazantzis et al. 1962). Acute renal failure has been observed in a number of case studies in which mercuric chloride has been ingested (Afonso and deAlvarez 1960; Murphy et al. 1979; Samuels et al. 1982). Autopsy of a 35-year-old man who ingested a lethal dose of mercuric chloride and exhibited acute renal failure showed pale and swollen kidneys (Murphy et al. 1979). A case study reported acute renal failure characterized by oliguria, proteinuria, hematuria, and granular casts in a woman who ingested 30 mg mercury/kg as mercuric chloride (Afonso and deAlvarez 1960). Another case study reported a dramatic increase in urinary protein secretion by a patient who ingested a single dose of 15.8 mg mercury/kg as mercuric chloride (assuming a body weight of 70 kg) (Pesce et al. 1977). The authors of the report surmised that the increased excretion of both albumin and  $\beta_2$ -microglobulin were indicative of mercury-induced tubular and glomerular pathology. Acute renal failure that persisted for 10 days was also observed in a 19-month-old child who ingested an unknown amount of powdered mercuric chloride (Samuels et al. 1982). Decreased urine was also observed in a 22-year-old who attempted suicide by ingesting approximately 20 mg mercury/kg (Chugh et al. 1978).

Agency Contact (Chemical Manager): John Risher

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**MINIMAL RISK LEVEL (MRL) WORKSHEET**

Chemical name(s): Mercury (inorganic)  
CAS number(s): 7439-97-6  
Date: June 15, 2001  
Profile status: Final Draft  
Route:  Inhalation  Oral  
Duration:  Acute  Intermediate  Chronic  
Key to figure: 17  
Species: Rat

Minimal Risk Level: 0.002  mg/kg/day  ppm

Reference: NTP. 1993. NTP technical report on the toxicology and carcinogenesis studies of mercuric chloride (CAS no. 7487-94-7) in F344/N rats and B6C3F<sub>1</sub> mice (gavage studies). NTP TR408.

Experimental design: Fischer 344 rats (10/sex/group) were administered 0, 0.23, 0.46, 0.93, 1.9, or 3.7 mg Hg/kg/day as mercuric chloride in deionized water by oral gavage once daily 5 days per week for 26 weeks. Body weights were recorded weekly. Surviving animals were sacrificed and necropsied. Organ weights were determined for the brain, heart, liver, lung, kidney, thymus, and testes. Histopathological examinations were performed.

Effects noted in study and corresponding doses: The relative and absolute kidney weights were significantly increased for dosed males and for females exposed to at least 0.46 mg/kg/day. At the two low-dose groups and the control group, minimal nephropathy was observed in nearly all the males. At 0.93 mg/kg/day level, renal tubule necrosis became more severe (moderate) and was statistically significant and remained at this severity at the higher dose groups. The female rats had a significant increased incidence at the high dose only, and severity was minimal. Nephropathy was characterized by foci of tubular regeneration, thickened tubular basement membrane, and scattered dilated tubules containing hyaline casts. Macroscopic changes included granular kidneys in dosed males. After 4 months of exposure, urinary levels of alkaline phosphatase, aspartate aminotransferase, lactate dehydrogenase, and gamma-glutamyl transferase were significantly elevated in both sexes at 3.7 mg Hg/kg/day, but at 6 months control levels had increased such that enzyme levels in males were no longer statistically significant and only levels of alkaline phosphatase and gamma-glutamyl transferase were significantly elevated in females.

Dose end point used for MRL derivation: 0.23 mg Hg/kg/day; no renal effects  
 NOAEL  LOAEL

Uncertainty and modifying factors used in MRL derivation: 100

1  3  10 (for use of a LOAEL)  
 1  3  10 (for extrapolation from animals to humans)  
 1  3  10 (for human variability)

Was a conversion factor used from ppm in food or water to a mg/body weight dose?  
If so explain: No conversion factor used.

Was a conversion used from intermittent to continuous exposure?

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If so, explain: Yes. The dose was adjusted for a continuous exposure by multiplying the NOAEL (0.23 mg/kg/day) by a conversion factor of 5/7:

$$\begin{aligned}\text{NOAEL}_{(\text{ADJ})} &= 0.23 \text{ mg/kg/day} \times (5 \text{ days}/7 \text{ days}) \\ &= 0.16 \text{ mg/kg/day}\end{aligned}$$

$$\text{MRL} = \text{NOAEL}_{(\text{ADJ})} \div \text{UF} = 0.16 \text{ mg/kg/day} \div 100 = 0.002 \text{ mg/kg/day}$$

If an inhalation study in animals, list conversion factors used in determining human equivalent dose:  
Not applicable.

Additional studies or pertinent information that lend support to this MRL: Renal toxicity has been observed in other intermediate-duration oral studies on rats and mice exposed to inorganic mercury (Carmignani et al. 1992; Jonker et al. 1993a; NTP 1993), as well as case reports on humans ingesting inorganic mercury for acute and chronic durations (Afonso and deAlvarez 1960; Davis et al. 1974; Kang-Yum and Oransky 1992; Nielsen et al. 1991; Pesce et al. 1977).

Agency Contact (Chemical Manager): John Risher

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**MINIMAL RISK LEVEL (MRL) WORKSHEET**

Chemical Name: Methylmercury  
CAS Number: 22967-92-6  
Date: June 15, 2001  
Profile Status: Final Draft  
Route:  Inhalation  Oral  
Duration:  Acute  Intermediate  Chronic  
Key to figure: 88  
Species: Human

Minimal Risk Level: 0.0003  mg/kg/day  mg/m<sup>3</sup>

Reference: Davidson et al. 1998. Effects of prenatal and postnatal methylmercury exposure from fish consumption on neurodevelopment: Outcomes at 66 months of age in the Seychelles Child Development Study. JAMA 280(8):701-707.

Experimental design. This MRL is based on the results of the Seychelles Child Development Study (SCDS), a series of evaluations on a population in the Seychelles Islands. The chronic oral MRL for methylmercury is based upon the Seychelles Child Development Study (SCDS), in which over 700 mother-infant pairs have, to date, been followed and tested from parturition through 66 months of age (Davidson et al. 1998). The SCDS was conducted as a double-blind study and used maternal hair mercury as the index of fetal exposure. Enrollees were recruited by the head nurse/hospital midwife by asking the mothers if they wished to participate in the study when they arrived at the hospital for delivery. The first 779 who did not decline participation became the mothers in the study cohort. Of the initial 779 mothers enrolled in the study at parturition, 740 remained at the predetermined child testing age of 6.5 months, 738 remained in the 19-month cohort, 736 remained at 29 months, and 711 remained for the 66-month neurobehavioral and developmental examinations.

The Seychellois were chosen as a study population for a number of reasons. (1) All fish contain some level of methylmercury (Davidson et al. 1998); and the Seychellois regularly consume a large quantity and variety of ocean fish, with 12 fish meals per week representing a typical methylmercury exposure. (2) The median total mercury concentration in 350 fish sampled from 25 species consumed by the Seychellois was <1 ppm (range, 0.004–0.75 ppm), comparable to that consumed by the U.S. population; thus, the methylmercury levels in the Seychellois population are 10–20 times those in the United States, not because they consume more highly contaminated fish than do Americans, but rather because they consume more fish than the U.S. population. (3) The Seychelles represent a relatively pristine environment, with no local industry for pollution, and are situated more than 1,000 miles from any continent or large population center. (4) The population is highly literate, cooperative, and has minimal immigration and emigration. (5) The Seychellois constitute a generally healthy population, with low maternal alcohol consumption and tobacco use (<2%). (6) In the 66-month study cohort, the mean maternal hair level of total mercury during pregnancy was 6.8 ppm (range, 0.5–26.7 ppm).

Effects noted in study and corresponding doses: The results of the 66-month testing in the SCDS revealed no evidence of adverse effects attributable to chronic ingestion of low levels of methylmercury in fish (Davidson et al. 1998). In this study, developing fetuses were exposed *in utero* through maternal fish ingestion before and during pregnancy (Davidson et al. 1998). Neonates continued to be exposed to maternal mercury during breastfeeding (i.e., some mercury is secreted in breast milk), and methylmercury



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exposure from the regular diet continued after the gradual post-weaning shift to a fish diet. In the 66-month study cohort, the mean maternal hair level of total mercury during pregnancy was 6.8 ppm (range, 0.5–26.7 ppm; n = 711), and the mean child hair level at the 66-month testing interval was 6.5 ppm (range, 0.9–25.8 ppm; n = 708). The 66-month test battery, which was designed to test multiple developmental domains, included as primary measures the following: (1) General Cognitive Index (GCI) of the McCarthy Scales of Children's Abilities (to estimate cognitive ability); (2) the Preschool Language Scale (PLS) total score (to measure both expressive and receptive language ability); (3) the Letter and Word Recognition and (4) Applied Problems subtests of the Woodcock-Johnson (W-J) Tests of Achievement (to measure reading and arithmetic achievement); (5) the Bender-Gestalt test (to measure visual-spatial ability); and (6) the total T score from the Child Behavior Checklist (CBCL) (to measure the child's social and adaptive behavior). Serum sampling revealed no detectable levels of PCBs (detection limit = 0.2 ng/mL).

None of the tests indicated an adverse effect of methylmercury exposure. In contrast, four of the six measures showed better scores in the highest MeHg-exposed groups, compared with lower exposure groups for both prenatal and postnatal exposure (the four test were the (1) General Cognitive Index (GCI) of the McCarthy Scales of Children's Abilities (to estimate cognitive ability); (2) the Preschool Language Scale (PLS) total score (to measure both expressive and receptive language ability); (3) the Letter and Word Recognition and (4) Applied Problems subtests of the Woodcock-Johnson (W-J) Tests of Achievement (to measure reading and arithmetic achievement). While the positive outcomes are not considered to indicate any beneficial effect of methylmercury on neurological development or behavior, they might be more appropriately attributed to the beneficial effects of omega-3 fatty acids or other constituents present in fish tissue, since the methylmercury levels in hair are known to correlate closely with fish intake. The slight decreases in the subjectively reported activity level of boys reported in the 29-month observations were not seen during the 66-month tests. The mean maternal hair level of 15.3 ppm in the group with the highest exposure in the 66-month test cohort is, therefore, considered a NOAEL for SCDS, and is used by ATSDR as the basis for derivation of a chronic oral MRL for methylmercury. A related study (Myers et al. 1997) by the same team of researchers from the University of Rochester examined the Seychellois children for attainment of the same developmental milestones reported to have been delayed in the Iraqi poisoning incident in the early 1970s (Cox et al. 1989) and found no such delays in the Seychellois children exposed *in utero*. Since the children had been exposed *in utero*, they represent the most sensitive subpopulation.

#### Sensitivity of Neurobehavioral Measures /Reliability of Tests Used in Critical Study

The neurobehavioral test battery used in the 66-month Seychelles study was designed to assess multiple developmental domains (Davidson et al. 1998). The tests were considered to be sufficiently sensitive and accurate to detect neurotoxicity in the presence of a number of confounding factors. On-site test administration reliability was assessed by an independent scorer, and mean interclass correlations for interscorer reliability were 0.96–0.97 (Davidson et al. 1998). The sample size was determined to be sufficient to detect a 5.7 point difference on any test with a mean (SD) of 100 (16) between low (0–3 ppm) and high >12 ppm) hair mercury concentration groups for a 2-sided test ( $\alpha = 0.05$  at 80% power).

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Converting blood concentration to daily intake.

The concentration of mercury in the blood may be converted to a daily intake by using the following equation from WHO (1990):

$$C = \frac{f(d)}{b(V)} \cdot \frac{A_D(A_B(d))}{b(V)}$$

Where:

- C = concentration in blood
- f = fraction of the daily intake taken up by the blood
- d = daily dietary intake
- b = elimination constant
- A<sub>D</sub> = percent of mercury intake in diet that is absorbed
- A<sub>B</sub> = percent of the absorbed amount that enters the blood
- V = volume of blood in the body

Hair to Blood Concentration Ratio.

The hair: blood concentration ratio for total mercury is frequently cited as 250. However, a precise basis for this particular value is unclear. Ratios reported in the literature range from 140 to 370, a difference of more than a factor of 2.5 (see Table 2-9). Differences in the location of hair sampled (head versus chest, distance of sample from head or skin) may contribute to differences in observed ratios between studies. For example, as much as a 3-fold seasonal variation in mercury levels was observed in average hair levels for a group of individuals with moderate-to-high fish consumption rates, with yearly highs occurring in the fall and early winter (Phelps et al. 1980; Suzuki et al. 1992). Thus, it is important to obtain hair samples as close to the follicle as possible to obtain an estimate of recent blood levels. Large errors (the direction of which depends on whether samples were taken while blood levels were falling or rising) could result if hair samples are not taken close to the scalp. Several studies did not report the distance to the scalp for the hair samples taken. The high slope reported by Tsubaki (1971a) may have reflected the fact that mercury levels were declining at the time of sampling (Berglund et al. 1971), so the hair levels may reflect earlier, higher blood levels. Hair taken from different parts of the body also may yield different ratios. In 26 subjects with moderate-to-high fish consumption, axillary hair (i.e., from the armpit area) was found to contain an average of 23% less mercury than head hair (Skerfving et al. 1974).

Phelps et al. (1980) obtained multiple blood samples and sequentially analyzed lengths of hair from 339 individuals in Northwestern Ontario. The large sample size and the attention to sampling and analysis with regard to the hair: blood relationship make this study the most appropriate to use for estimating the mercury blood levels of the Seychellois women during pregnancy. The actual ratio Phelps et al. (1980) observed between the total mercury concentration in hair taken close to the scalp and simultaneous blood sampling for this group was 296. To estimate the actual ratio, the authors assumed that blood and hair samples were taken following complete cessation of methylmercury intake. They also assumed a half-life of methylmercury in blood of 52 days and a lag of 4 weeks for appearance of the relevant level in hair at the scalp. Based on these assumptions, they calculated that if the actual hair: blood ratio were 200, they would have observed a ratio of 290 (i.e., essentially equivalent to the observed value of 296). Based on these and other considerations, Phelps et al. (1980) state that the actual ratio is "probably higher than 200, but less than the observed value of 296." As the authors point out, two-thirds of the study population were sampled

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during the falling phase of the seasonal variation and one-third or less in the rising phase. This fact would tend to result in a lower observed ratio; therefore, the actual average value is likely to be >200. Phelps et al. (1980) also provide estimates assuming a 2-week lag for the appearance of the relevant level of mercury in the centimeter of hair nearest the scalp. For a 2-week lag time, an actual ratio of 250 would have resulted in an observed ratio of 301 (again, essentially identical to the observed value of 296). A study of ingestion of a large dose of mercuric chloride in one individual suggests that the lag time is longer than 2 weeks (Suzuki et al. 1992). Hair samples were taken at 41 and 95 days following ingestion of the mercuric chloride. In the 41-day hair sample, a large mercury peak occurred in the centimeter of hair closest to the scalp, with no elevation in mercury in the second centimeter of hair. Head hair grows at a rate of about 1.1 cm a month (Al-Shahristani and Shihab 1974; Cox et al. 1989). If emergence had occurred so that the elevation in mercury could be measured in the first centimeter of hair by 2 weeks after exposure, then by day 41 after exposure the peak should have moved into the second centimeter of hair, at least enough to raise the mercury level slightly in the second centimeter. Because no elevation was seen in the second centimeter of hair at 41 days, it would appear that emergence occurred at a lag of >2 weeks. In the hair sample taken at 95 days, the leading edge of the mercury peak occurred in the third centimeter of hair.

Based on the data presented in Phelps et al. (1980) and the lag time indicated in the individual studied by Suzuki et al. (1992), the actual average value is likely to be somewhere between 200 and 250. Because the data do not allow a more accurate determination of an average ratio, the value 250 is acceptable for the purpose of estimating average blood levels in the Seychellois population. Using 250 rather than a lower number results in a lower MRL. It should be noted that a wide range in hair:blood ratios has been reported for individuals in various studies: 137–342 in Soria et al. (1992), 171–270 in Phelps et al. (1980), and 137–585 in Birke et al. (1972). Therefore, this ratio (250) should not be used as the sole basis for determining levels of exposure and potential effect for individuals.

#### Calculation of dietary intake from blood concentration.

*Fraction of mercury in diet that is absorbed ( $A_D$ ).* Radiolabeled methyl-mercuric nitrate was administered in water to three healthy volunteers (Aberg et al. 1969). The uptake was >95%. Miettinen et al. (1971) incubated fish liver homogenate with radiolabeled MeHgNO<sub>3</sub> to yield a methylmercury proteinate. The proteinate was then fed to fish that were killed after a week, cooked, and fed to volunteers after confirmation of the methylmercury in the fish. Mean uptake exceeded 94%. For the derivation of an MRL, an absorption factor of 0.95 is used.

*Fraction of the absorbed dose that is found in the blood ( $A_B$ ).* The value 0.05 has been used for this parameter in the past (Berglund et al. 1971; WHO 1990). Three studies report observations of the fraction of the absorbed methylmercury dose distributed to blood volume in humans. Kershaw et al. (1980) report an average fraction of 0.059 of the absorbed dose in the total blood volume, based on a study of 5 adult male subjects who ingested methylmercury-contaminated tuna. In a group of 9 male and 6 female volunteers who had received <sup>203</sup>Hg-methylmercury in fish, approximately 10% of the total body burden was present in 1 L of blood in the first few days after exposure, dropping to approximately 5% over the first 100 days (Miettinen et al. 1971). In another study, an average value of 1.14% for the percentage of absorbed dose in 1 kg of blood was derived from subjects who consumed a known amount of methylmercury in fish over a period of 3 months (Sherlock et al. 1984). Average daily intake for the 4 groups observed in the study ranged from 43 to 233 µg/day. The authors report a dose-related effect on the estimated percentage of the absorbed dose in 1 kg of blood, with 1.26% of the absorbed dose in 1 kg of blood at an average daily intake of 43 µg/day and 1.03% of the absorbed dose in 1 kg of blood at an average daily intake of 233 µg/day. The average for all subjects in the study was 1.14%. When individual values for distribution to one kilogram of blood reported in the study are converted into the percentage of the absorbed

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dose in the total blood volume (assuming that blood is 7% of body weight [Best 1961] and using body weights reported for individuals in the study), the average value for  $A_B$  for all individuals is 0.056 (0.057 using the values for percentage in 1 kg normalized for body weight as reported in the study). The average value for  $A_B$  for 6 women as reported in Sherlock et al. (1984) is 0.048 (0.047 using values normalized for body weight). The average for 14 men is 0.059 (0.061 using values normalized for body weight).

The average values for  $A_B$  for all studies ranged from 0.047 to 0.061 (the values for women and men reported in Sherlock et al. [1984]). The data suggest that the average value of  $A_B$  for women may be lower than that for men, and they further suggest that 0.05 may be appropriate for modeling intake in a group of women (Sherlock et al. 1984). Based on these studies, the best estimate of  $A_B$  based on the available data is 0.05. Use of a higher value (i.e., 0.06 instead of 0.05) for this parameter would result in a lower MR, but the sensitive populations are pregnant women and developing fetuses, making the 0.5 value more appropriate for the Seychelles study population.

*Elimination constant (b).* Reported clearance half-times for methylmercury from blood or hair range from 48 to 65 days (Table 2-5). The average elimination constant based on the 6 studies listed in Table 2.5 is 0.014. The average of the individual values for  $b$  reported for 20 volunteers ingesting from 42 to 233  $\mu\text{g}$  Hg/day in fish for 3 months (Sherlock et al. 1984) is also 0.014. Use of the value 0.014 for this parameter, rather than 0.01 (as used by WHO 1990), results in a higher MRL.

*Volume of blood in body (V), and body weight.* Blood volume is assumed to be 7% of body weight, with an increase to about 9% during pregnancy (Best 1961). Data for the body weight of the Seychelles Islands women were not found. Assuming an average body weight of 60 kg for women, the blood volume is 4.2 L (60 kg x 0.07 L/kg).

### Calculation of Exposure Dose

The concentration of mercury in hair is assumed to be 250 times the concentration in blood. Using the mean total mercury level of 15.3 ppm in maternal hair taken at parturition to represent a NOAEL in the 66-month Seychelles testing (Davidson et al. 1998), the corresponding methylmercury concentration in blood would be:  $1/250 \times 15.3 \mu\text{g/g} \times 1 \text{ mg}/1,000 \mu\text{g} \times 1,000 \text{ g/L} = 0.061 \text{ mg/L}$ .

### Calculation of Daily Intake from Blood Concentration

$$C = \frac{f(d)}{b(V)} \cdot \frac{A_D(A_B(d))}{b(V)}$$

Using the above equation to relate the concentration in blood (C, in  $\mu\text{g/L}$ ) to daily intake (d, in  $\mu\text{g/day}$ ): where C = (percent of ingested dose absorbed through the GI tract x percent of that dose absorbed into the blood x the daily amount ingested) divided by (elimination constant x blood volume in a 60 kg female)

that is,

$$\begin{aligned} C &= (0.95 \times 0.05 \times d) / (0.014 \times 4.2) \\ C &= 0.81 d \\ 0.061 \text{ mg/L} &= 0.81 d \\ d &= 0.075 \text{ mg/day} \end{aligned}$$

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Using the assumed body weight of 60 kg for women, the estimated dose that would result in a hair level of 15.3 ppm is  $0.075/60 \text{ kg} = 0.0013 \text{ mg/kg/day}$ . Therefore, the NOAEL derived from the highest exposure group ( $n = 95$ ) at 66 months is  $0.0013 \text{ mg/kg/day}$ .

Dose and end point used for MRL derivation: 0.0013 mg/kg/day NOAEL

NOAEL  LOAEL

Uncertainty and Modifying Factors used in MRL derivation:

1  3  10 (for use of a minimal LOAEL)  
 1  3  10 (for extrapolation from animals to humans)  
 1  3  10 (for human pharmacokinetic and pharmacodynamic variability)  
 1.5  3  10 (Modifying factor to account for domain-specific findings in Faroe study)

Consideration of Uncertainty

The standard/traditional areas of uncertainty addressed in any duration-specific MRL are: (1) interspecies variability (i.e., cross-species extrapolation of a NOAEL or LOAEL); (2) intra-human variability (i.e., differences in susceptibility to a substance or effect within the human population); (3) use of an LOAEL for MRL derivation when an NOAEL for the critical effect is not available; and (4) extrapolation from subchronic to chronic duration. In addition, a modifying factor may also be used when special circumstances exist that may contribute to, or introduce, uncertainty into the calculated health guidance value (MRL) in an area not typically covered by the traditional uncertainty factor approach.

The NOAEL of 15.3 ppm mercury in maternal hair from Davidson et al. (1998) used as the starting point for MRL derivation was based upon an unusually large study cohort of the population considered most sensitive to the neurodevelopmental effects of methylmercury, i.e., pregnant women and their developing fetuses. The negative results of this study are strongly supported by the BMD NOAEL range of 13 to 21 ppm calculated for the New Zealand cohort of 237 mother-child pairs (Crump et al. 1998). Consequently, much of the uncertainty normally present in the MRL derivation process does not exist in the case of methylmercury. Nonetheless, in view of the nature of the most susceptible group (developing fetuses) and some questions raised in the vast human data base for this chemical, an aggregate value of 4.5 was employed.

This value (4.5) was based upon three separate components, two of which are interrelated and the other independent. For the Seychelles data, a value of 1.5 was used to address the variability in hair-to-blood ratios among women and fetuses in the U.S. population, as determined by pharmacokinetic modeling of actual data by Clewell et al. (1998); a second value of 1.5 was applied to address the remainder of any inter-individual variability (i.e., pharmacodynamics) in the U.S. population. A third, and independent, factor of 1.5 was employed to account for the possibility that the domain-specific tests, as employed extensively in the Faroe Islands, but not the Seychelles (which used primarily neurobehavioral tests of global function) might be able to detect very subtle neurological effects not tested for in the 66-month Seychelles cohort.

The World Health Organization (WHO, 1993, 1996) has defined the -kinetic and -dynamic components of intrahuman variability as being equal contributors to, and collectively constituting the total of, human variability. In order to ensure a conservative approach, these two interdependent components were added to give a composite uncertainty factor of three (i.e.,  $1.5 + 1.5 = 3$ ) to account for the full range of variability attributable to mercury in the Seychelles study. A modifying factor of 1.5 was also used to account for the

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possibility of domain-specific effects, as were seen in the Faroe study, being attributable to mercury. Since these effects were considered to be entirely separate or “independent” events, this modifying factor of 1.5 was multiplied by the uncertainty factor of 3.0 (for uncertainty attributable solely to the Seychelles study) to yield an aggregate uncertainty of 4.5 for chronic oral exposure to methylmercury.

While domain-specific tests from the Seychelles were reviewed at the North Carolina meeting in November 1998 and the results failed to demonstrate effects, the tests do not represent the full range of domain-specific tests that were administered in the Faroe Islands. For these reasons, and based on our consultation with our Board of Scientific Counselors about concerns for “missing” data sets (i.e., in relation to the Executive Order of children’s health and the agency’s efforts to protect the health of children, including the developing fetus), ATSDR determined that an additional factor of 1.5 should be used since the full range of domain-specific neuropsychological test results from the Seychelles are not yet available. When these results become available and if they fail to show domain-specific effects, this additional factor of 1.5 would no longer be needed. At that time ATSDR will re-evaluate its MRL, as well as all other relevant data, in compliance with the agency’s mandates and authorities.

Therefore, in the calculation of the chronic oral MRL for methylmercury, the NOAEL of 0.0013 mg/kg/day from the 66-month study (Davidson et al. 1998) is divided by 4.5, giving a chronic oral MRL for methylmercury of 0.0003 mg/kg/day [0.0013 mg/kg/day / 4.5 (UF) = 0.0003 mg/kg/day].

If an inhalation study in animals, list the conversion factors used in determining human equivalent concentration (HEC): Not applicable.

Additional studies or pertinent information which lend support to this MRL:

Crump et al. (1998) conducted benchmark dose (BMD) calculations and additional regression analyses of data collected in a study in which a series of scholastic and psychological tests were administered to children whose mothers had been exposed to methylmercury during pregnancy. Hair samples were collected from 10,970 new mothers in New Zealand in 1977 and 1978. High hair mercury levels were considered to be those over 6 ppm, which was the hair level predicted to result at steady state from consumption of mercury at the WHO/FAO Provisional Tolerable Weekly Intake of 0.3 mg total mercury/week and 0.2 mg methylmercury/week. By this criterion, 73 of approximately 1,000 mothers who had consumed fish more than three times/week during pregnancy were determined to have high hair mercury levels. In 1985, when the children were 6 to 7 years of age, 61 children (1 set of twins) of the 73 mothers in the high hair mercury group were located, and constituted the high exposure group, which was matched with three control groups (one with 3-6 ppm maternal hair mercury levels, one with 0-3 ppm whose mothers had been high fish consumers, and one with 0-3 ppm whose mothers had not been high fish consumers). The entire study cohort consisted of 237 children. A battery of 26 psychological and scholastic tests were administered to the children at school during the year 1985. Mothers were interviewed at the time of test administration to obtain additional data on social and environmental factors. In the high exposure group of children, one boy’s mother had a hair mercury level of 86 ppm, which was more than four times higher than the next highest hair mercury level of 20 ppm. BMDs (10% response rate) calculated from five tests ranged from 32 to 73 ppm, when the 86 ppm mother’s child was included. This corresponded to a BMDL range of 17 to 24 ppm. Although none of the 86 ppm child’s test scores was an outlier according to the definition used in the analyses, his scores were significantly influential in the analyses. When this child was omitted from the analyses, BMDs ranged from 13 to 21, with corresponding BMDLs of 7.4 to 10 ppm.

Developing fetuses in the SCDS were exposed through maternal fish ingestion before and during pregnancy. Each child was evaluated at 19 months and again at 29 months ( $\pm 2$  weeks) for infant intelligence (Bayley

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Scales of Infant Development [BSID] Mental and Psychomotor Scales), with a modified version of the BSID Infant Behavior Record to measure adaptive behaviors at 29 months (Davidson et al. 1995b). Testing was performed by a team of Seychellois nurses extensively trained in administration of the BSID. Maternal hair concentrations, measured in hair segments that corresponded to pregnancy, ranged from 0.5 to 26.7 ppm, with a median exposure of 5.9 ppm for the entire study group. The mean BSID Mental Scale Indices determined at both 19 and 29 months were found to be comparable to the mean performance of U.S. children. The BSID Psychomotor Scale Indices at both measurement intervals were two standard deviation units above U.S. norms, but were still consistent with previous findings of motor precocity in children reared in African countries. The study found no effect that could be attributed to mercury on the BSID scores obtained at either the 19- or 29-month measurement/testing interval. The 29-month cohort represented 94% of the 779 mother-infant pairs initially enrolled in the study, and approximately 50% of all live births in the Seychelles in 1989.

The only observation in the 29-month testing that might be attributable to prenatal mercury exposure was a slight decrease in the activity level in boys (but not girls) as determined by the Bayley Infant Behavior Record (subjective observation). Whereas this decrease was significant in males ( $p = 0.0004$ ), it was not statistically significant in females ( $p = 0.87$ ). When the subjective activity scores for male and female children were evaluated collectively, no statistically significant or remarkable decrease in activity was apparent outside the  $>12$  ppm maternal hair concentration group. The affect on activity level in boys is not considered an adverse effect by the authors of the study.

Grandjean et al. (1997b, 1998) reported another epidemiological study of methylmercury exposure for a population in the Faroe Islands. Although the Faroese are a fishing culture, the major source of methylmercury exposure for this population is pilot whale meat, which is intermittently consumed as part of the cultural tradition. The initial study cohort consisted of 1,022 singleton births occurring in a 21-month window during 1986-1987. At approximately 7 years of age, neurobehavioral testing was conducted on 917 of the remaining cohort members. No abnormalities attributable to mercury were found during clinical examinations or neurophysiological testing. A neuropsychological test battery was also conducted, which included the following: Finger Tapping; Hand-Eye Coordination; reaction time on a Continuous Performance Test; Wechsler Intelligence Scale for Children - Revised Digit Spans, Similarities, and Block Designs; Bender Visual Motor Gestalt Test; Boston Naming Test; and California Verbal Learning Test (Children). Neuropsychological tests emphasized motor coordination, perceptual-motor performance, and visual acuity. Pattern reversal visual evoked potentials (VEP) with binocular full-field stimulation, brain stem auditory evoked potentials (BAEP), postural sway, and the coefficient of variation for R-R inter-peak intervals (CVR) on the electrocardiogram were all measured. The neuropsychological testing indicated mercury-related dysfunction in the domains of language, attention, memory, and visuospatial and motor function (to a lesser extent), which the authors considered to remain after the children of women with maternal hair mercury concentrations above  $10 \mu\text{g/g}$  (10 ppm) were excluded. While this study represents a significant contribution to the human database for methylmercury exposure and effects, a number of potentially influential factors not fully considered as possible covariates somewhat cloud the interpretation of the results.

These differences between the neuropsychological effects observed in the Faroe Island cohort and the absence of effects reported in the Seychelles Island cohort might result from a variety of factors. The Faroe Island children were older (7–8 years versus 5.5 in the SCDS). Some of the measurement instruments (i.e., the neuropsychological test administered) were also different. Since the first neuropsychological testing in the Faroe study was not conducted until 7 years of age, it is not known whether the observed effects might have been apparent at an earlier age. Ongoing and planned future testing of the Seychelles population will provide additional information on the progression of any observed effects. Further examination of the

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Seychelles population using the neuropsychological test that showed positive results in the Faroe Islands population will also allow a more direct comparison of results.

The diet in the two studies was also considerably different. The majority of the mercury exposure to the Faroe Island population came from whale meat (estimated at about 3 ppm in muscle tissue) with a relatively small portion coming from fish. Some of the mercury in whale meat is in the form of inorganic mercury. In the Seychelles study, all of the mercury came from fish as methylmercury with concentrations of around 0.3 ppm. Whale meat blubber is widely consumed in the Faroe Islands and also contains polychlorinated biphenyls (PCBs). Grandjean et al. (1995b) estimated a daily intake of 200 µg of PCB. This value can be compared to the Tolerable Daily Intake of PCBs established by the FDA, of 60–70 µg/day for an adult. Further statistical analysis of the possible influence of PCBs on the observed study results needs to be conducted (see the discussion below on Peer Panel 1 Review of Key Studies for additional comments).

The primary biomarker used to estimate mercury exposure was also different between the two studies. The Faroe Island analysis used cord blood, and the Seychelles study used maternal hair level. The use of mercury in cord blood has the advantage of being a more direct measure of exposure to the fetus, but the levels at term may not reflect exposures at earlier developmental stages. While Grandjean et al. (1997) did report maternal hair mercury levels, the mean hair level for the interquartile range of 2.6–7.7 ppm was reported only as a geometric average (4.27 ppm). In contrast, the Seychellois study reported only an arithmetic mean level for the entire study population (6.8 ppm). While both are valid measures, a direct comparison of “average” values for the two studies is not possible without further statistical analysis of both data sets.

In the case of the Faroe study, there were no data presented in the peer-reviewed publications to address variability of food/whale meat or blubber intake among the Faroe Islanders, making it difficult to evaluate the possibility of peak intake levels during critical development phases. Consumption data was reported only as <1 pilot whale meat meal/month and 1-2 fish meals per week. In contrast, the Seychelles dietary habits provide a relatively stable intake, and a high degree of correlation was found between mean hair levels in samples covering each trimester versus levels in samples for the entire pregnancy (Cernichiari et al. 1995a). Cernichiari et al. (1995b) also report a good correlation between levels of total mercury in neonatal brain and levels in the corresponding maternal hair. While the contribution of continued mercury exposure through breast feeding or post-weaning diet was not fully addressed in the Seychellois study reports (Davidson 1995, 1998), that is not considered a significant drawback with the study, since no effects on neurobehavioral/neuropsychological testing were seen at any maternal hair level. In the Faroese assessment of latent neuropsychological effects from an *in utero* exposure to mercury, however, the role of continuing postnatal exposure to mercury either from breast milk or from ingestion of methylmercury-containing foods (e.g., pilot whale meat) is less clear. Specifically, it is not known what proportion, if any, of the neuropsychological effects reported in the Faroe Island population could be attributed to seven years of postnatal exposure to methylmercury in food. The variability and magnitude of this postnatal exposure should, therefore, be further evaluated.

#### Peer Panel Review of Key Studies

In addition to the traditional peer review process that precedes publication in most scientific journals, the studies considered by ATSDR for use in estimating a chronic oral MRL for methylmercury underwent two stringent reviews by recognized experts in the environmental health field.

On July 20 and 21, 1998, ATSDR assembled a panel of 18 experts from the scientific and medical communities to review current issues and the relevant literature on mercury and its compounds, including



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methylmercury (ATSDR 1999). Several members of each of the respective research teams that conducted the Iraqi, Seychelles, Faroe, and Madeira studies were included among the expert panelists, and provided extensive overviews of their studies. The presentations were followed by an open, wide-ranging scientific discussion of the merits and interpretations of the currently available studies. Topics of significant discussion included the relative merits of the respective study populations, exposure regimens, sensitivity of neurobehavioral measures, and determination of an uncertainty factor. While it was unanimously agreed that the Seychelles and Faroe studies were both excellent studies that provided a significant contribution to the human database for methylmercury exposure and effects, a number of factors that could have contributed to the study results, but were not considered as possible statistical covariates, were discussed. In the case of the Faroe study, the consumption of whale blubber, which is known to be contaminated with PCBs, DDT, and possibly other organochlorines, introduces a potentially significant influence on the study results. Weihe et al. (1996) reported that the PCB and DDT concentrations in blubber of pilot whales taken in Faroese waters are about 30 ppm and 20 ppm, respectively. In contrast, the Seychellois population does not eat marine mammals at all. In addition, the Faroe study did not address other possible statistical covariates, such as the dietary and nutritional status of the study population and the use of tobacco during pregnancy, further complicating the interpretation of the neuropsychological test results.

On November 18–20, 1998, a workshop on Scientific Issues Relevant to the Assessment of Health Effects from Exposure to Methylmercury was conducted in Raleigh, North Carolina. Jointly sponsored by the U.S. Department of Health and Human Services, the National Institute of Environmental Health Sciences (NIEHS), the Centers for Disease Control and Prevention (CDC), the Food and Drug Administration (FDA), the U.S. Environmental Protection Agency (EPA), the National Oceanic and Atmospheric Administration (NOAA), the Office of Science and Technology Policy (OSTP), the Office of Management and Budget (OMB), and ATSDR, the purpose of this workshop was to discuss and evaluate the major epidemiologic studies that associated methylmercury exposure and the results of an array of developmental measures in children. These studies monitored and evaluated exposed populations in Iraq, the Seychelles Islands, the Faroe Islands, and the Amazon River Basin. A number of animal studies were also considered in support of a human health risk assessment. Presentation of these studies by the research team that conducted the study was followed by an expert panel evaluation that examined each study, taking into consideration the exposure data, experimental design and statistical analysis, potential confounders and variables, and neurobehavioral endpoints evaluated. A fifth panel evaluated the results of relevant animal studies. Significant issues that were discussed included the use of umbilical cord blood mercury levels vs. hair mercury concentrations as an index of methylmercury exposure during pregnancy, the patterns of exposure, the dietary/health status of study populations, other potentially relevant exposures, other confounding influences, and the adjustments made for statistical covariates. All five panels at this workshop commended the efforts of the investigators and respective staffs of the Seychelles and Faroe studies for conducting highly sophisticated investigations under difficult conditions. However, specific findings of several of the panels raise issues that, at present, preclude the Faroe data from consideration as a starting point for MRL derivation.

In their addressal of the potential influence of concurrent PCB exposure on the Faroe results, the Confounders and Variables (Epidemiology) panel indicated that with respect to four of the pre-natal outcomes (related primarily to verbal and memory performance), when PCBs were included in the model, only one of these outcomes is specifically related to mercury exposure. Concerning this matter, the panel wrote that "... the most likely explanation is that both (mercury and PCBs)... affect these three outcomes, but their relative contributions cannot be determined given their concurrence in this population." The Neurobehavioral Endpoints Panel also looked at this issue, and noted that "PCB exposure might act as an effect modifier, increasing the susceptibility to MeHg."; however, this panel further indicated that it did not believe that the effects seen in the Faroe Islands were due to uncontrolled confounding by PCBs. A third

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panel that addressed the issue of concurrent PCB exposures, the Statistics/Design Panel, noted that only 3 of 208 PCB congeners were measured in the Faroe study, and stated that it “seems likely that mercury was measured more accurately than the biologically relevant PCB exposure. Consequently even if the neurological effects seen in this study were caused entirely by PCBs, it is possible that mercury would still be more highly correlated with these effects than PCBs.” The Statistics/Design Panel also said that “the best method to deal with this problem would be to study a population where exposure to PCBs is not an issue.” This statement points directly to the Seychelles study as the study most appropriate for MRL derivation.

Another issue raised at Raleigh workshop concerned the taking of hair samples for determining pre-natal exposure. In the Seychelles, hair samples were collected 6 months post-partum, and segments corresponding to pregnancy were selected for analysis. In the case of the Faroese, hair samples were taken at the scalp. Regarding that, the Confounders and Variables (Epidemiology) panel stated that “Given the time it takes the Hg to be excreted into the hair, we can assume that samples collected at parturition do not cover the last 6 weeks of gestation, during which critically important neuronal proliferation and differentiation is taking place.”

Regarding both the Seychelles and Faroe studies, the Neurobehavioral Endpoints Panel found “no specific neurobehavioral signature injury from MeHg” in the data from either study (Seychelles or Faroe). The same panel also noted that episodic exposure in the Faroe Islands (1–2 fish meals/week and <1 pilot whale meal/month) “may reduce the likelihood of detecting a consistent ‘neurobehavioral signature injury’ specific to MeHg and may account for different observations in children with the same average exposure.”

Based upon the discussions at the Raleigh workshop and the individual panel findings, as well as the aforementioned Atlanta expert panel review, ATSDR has determined the Seychellois study to represent the most appropriate and reliable data base currently available for calculation of a chronic oral MRL from a population exposed only to methylmercury by a relevant route of exposure for the overall U.S. population.

[It should be emphasized that the Seychelles study and the Faroe study represent credible scientific contributions by widely respected research teams. Similarly, both studies extend our knowledge base well beyond that provided by the Iraqi study and make significant contributions to our understanding of the effects of low-level exposure to methylmercury by an exposure route and vehicle (i.e., food) relevant to U.S. populations. The continuing monitoring and evaluation of the Seychellois and Faroese populations with more comparable neurobehavioral indices should help strengthen our understanding of the effects of low level chronic methylmercury exposure and should reduce the uncertainty regarding the public health implications of exposure.]

Other epidemiology studies were also considered by the workshop panels. Lebel et al. (1997) evaluated a fish-eating populations in the Amazon River Basin with a neurofunctional test battery and clinical manifestations of nervous system dysfunction in relation to hair mercury concentrations. The villagers examined live along the Tapajos River, a tributary of the Amazon. The study population consisted of 91 adult inhabitants 15-31 years of age. Hair mercury levels were below 50 µg/g (ppm). Clinical examinations were essentially normal, although persons displaying disorganized movements on an alternating movement task and those with restricted visual fields generally had higher hair mercury levels. Near visual contrast, sensitivity, and manual dexterity (adjusted for age) were found to decrease significantly with increasing mercury levels, while a tendency for muscular fatigue and decreasing strength were observed in women. The authors suggested that dose-dependent nervous system alterations might be associated with hair mercury levels below 50 ppm. This study, however, also had a number of potentially confounding factors. The impact of parasitic and other diseases endemic to the study area is of primary

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concern in the interpretation of the Lebel et al. (1997) results. In addition, the overall nutritional status of the study population was not known or reported, and the use of neuroactive drugs (from local herbs, plants, roots, or mushrooms) was not considered as a potential confounder or covariate. The previous mercury exposure history of the study cohort was also unclear. This is of particular importance because gold mining procedures that use metallic mercury have been commonly practiced along the Amazon Basin for decades. Finally, the endpoints of the Lebel et al. (1977) study evaluated adult toxicity and not effects in the developing fetus or the newborn (i.e., the most sensitive human population).

The panel also reviewed the Iraqi study. Cox et al. (1989) and WHO (1990) reported delayed onset of walking in offspring in Iraqi children whose mothers were exposed to methylmercury through the consumption of seed grain treated with methylmercury as a fungicide (Al-Mufti et al. 1976; Bakir et al. 1973; Cox et al. 1989; Marsh et al. 1981, 1987). Exposure to methylmercury from other sources (e.g., fish or meat) was probably very low or nonexistent (Al-Mufti et al. 1976). It is likely that the children were exposed both prenatally through the placenta and postnatally through the mother's milk. A maternal exposure level of 0.0012 mg/kg/day, corresponding to the hair level of 14 ppm, was estimated using a simple, one-compartment pharmacokinetic model.

Myers et al. (1997) evaluated the population of the SCDS for developmental milestones similar to those determined in Iraq. As part of this ongoing study, cohort children were evaluated at 6.5, 19, 29, and 66 months of age. At 19 months care-givers were asked at what age the child walked (n=720 out of 738) and talked (n=680). Prenatal mercury exposure was determined by atomic absorption analysis of maternal hair segments corresponding to hair growth during the pregnancy. The median mercury level in maternal hair for the cohort in this analysis was 5.8 ppm, with a range of 0.5–26.7 ppm. The mean age (in months) at walking was 10.7 (SD=1.9) for females and 10.6 (SD=2.0) for males. The mean age for talking (in months) was 10.5 (SD=2.6) for females, and 11.0 (SD=2.9) for males. After adjusting for covariates and statistical outliers, no association was found between the age at which Seychellois children walked or talked and prenatal exposure to mercury. The ages for achievement of the developmental milestones were normal for walking and talking in the Seychellois toddlers following prenatal exposure to methylmercury from a maternal fish diet. The 5.8 ppm NOAEL of this study is considerably below the one estimated from the dose-response analysis of the data for the Iraqi methylmercury poisonings (10 ppm).

Clarkson (1995) raised some interesting issues concerning whether it is reasonable to apply health effects data based on an acute exposure to methylmercury fungicide eaten in homemade bread (in the 1971–1972 Iraq incident) to fish-eating populations having chronic exposure to much lower concentrations of methylmercury. Clarkson (1995) addressed two specific issues. The first regards the body's "defense mechanisms" that serve to mitigate the potential damage from mercury. One such mechanism in the case of methylmercury involves an enterohepatic cycling process in which methylmercury from dietary sources absorbed through the intestine is carried to the liver, where substantial quantities are secreted back into the bile and returned to the intestinal tract. During the residence time in the gut, microflora break the carbon-mercury bond, converting methylmercury into inorganic mercury, which in turn is poorly absorbed and is excreted in the feces. This creates an effective detoxification pathway for low-dose dietary exposures to methylmercury, but probably not for acute, high-dose exposures, such as occurred in Iraq. Secondly, the transport of methylmercury into brain tissue is inhibited by the presence of many amino acids, including leucine, methionine, and phenylalanine. Thus, it is possible that the rising plasma concentrations of amino acids from ingestion of fish protein may serve to depress the uptake of methylmercury by the brain.

While both of these issues need further laboratory/clinical investigation, they do raise appropriate questions concerning the relevance of the relatively short-term (i.e., about six weeks), high-level contaminated grain exposure scenario encountered in Iraq to the dietary methylmercury exposure scenarios encountered in many

## APPENDIX A

fish-eating populations (e.g., the Seychelles Islanders, Faroe Islanders, Peruvian villagers, and Inuit native people of Greenland). This position is supported by Cicmanec (1996), who reviewed data from the Iraqi study, as well as data from studies of fish-consuming populations in the Faroe Islands, Seychelles Islands, and Peruvian fishing villages. Cicmanec concluded that the Iraqi population does not represent a sensitive subpopulation within a perinatal group; rather, the relative lower threshold identified in that study was the result of confounders. Crump et al. (1995) reanalyzed the dose-response data from the Cox et al. (1989) report of the Iraqi incident and found the results to be potentially skewed by inadequacies in the study design and data-collection methods. Shortcomings or potentially confounding factors include: (1) the retrospective recall of developmental milestones by mothers and other family members; (2) the lack of precision in the determination of birth and other milestone dates; (3) and the possible biasing of the dose-response analysis by variation in symptom reporting and infant sex composition in the two study subcohorts. Crump et al. (1995) noted that perhaps the most serious limitation of the Iraqi study is the inability to assess the potential effects of low-level chronic-duration exposure to methylmercury, as these particular data are based on very high intake levels over a relatively brief period of time.

No increase in the frequency of neurodevelopmental abnormalities in early childhood was observed in a cohort of 131 infant-mother pairs in Mancora, Peru (Marsh et al. 1995b). The mean concentration of mercury in maternal hair was determined to be 8.3 ppm (range, 1.2–30 ppm), and the source of the mercury was believed to be from consumption of marine fish. Similarly, a study of 583 Faroe Island infants for the first 12 months after birth found no decrease in the age of attainment of sitting, creeping (crawling), and standing developmental milestones (Grandjean et al. 1995a). The age at which a child reached a particular developmental milestone was not only not found to be associated with prenatal mercury exposure, but infants that reached a milestone early were found to have significantly higher mercury concentrations in their hair at 12 months of age. It was also found that early milestone attainment was clearly associated with breast-feeding, which was in turn related to higher infant hair mercury levels. The authors (Grandjean et al. 1995a) concluded that the beneficial effects associated with breast-feeding seemed to overrule, or to compensate for, any neurotoxic effects on milestone development that could be due to the presence of contaminants (e.g., mercury) in human milk.

Additional studies have shown developmental toxicity after oral exposure of humans and animals to organic mercury compounds (Amin-Zaki et al. 1974; Bakir et al. 1973; Bornhausen et al. 1980; Cagianò et al. 1990; Elsner 1991; Engleson and Herner 1952; Fowler and Woods 1977; Guidetti et al. 1992; Harada 1978; Hughes and Annau 1976; Ilback et al. 1991; Inouye and Kajiwara 1988; Khera and Tabacova 1973; Lindstrom et al. 1991; McKeown-Eyssen et al. 1983; Nolen et al. 1972; Olson and Boush 1975; Rice 1992; Rice and Gilbert 1990; Snyder and Seelinger 1976; Stoltenburg-Didinger and Markwort 1990).

The accumulation of mercury is greater in larger fish and in fish higher in the food chain. The tendency for increased mercury concentration with increasing fish body weight is particularly noticeable in carnivorous fish species. Malm et al. (1995) analyzed mercury concentrations in 16 species of carnivorous fish from the Tapajos River basin in Brazil and hair samples from local populations who regularly ate such fish. Mercury levels in the fish averaged 0.55 ppm (range, 0.04–3.77 ppm), and the mercury levels in the hair of the affected fish-eating populations averaged approximately 25 ppm. In one population that consumed higher quantities of large carnivorous fish at the end of the local rainy season, 8 of 29 persons evaluated had hair mercury levels above 40 ppm, and one individual had a hair mercury concentration of 151 ppm. Some villages along the river can have per capita daily fish consumption rates around 200 g or more, which would greatly impact the human body burden and hair levels of mercury in such populations.

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Alternative Derivations of the MRL

To ensure a health guidance value based upon the best use of the Seychelles study data (widely considered the most relevant data available), ATSDR evaluated alternate MRL derivation methods for methylmercury.

One such method was a physiologically based pharmacokinetic approach using the mean total mercury level of 6.8 ppm in maternal hair for the entire Seychellois study cohort. Using the same formula as in the previous MRL calculation,

$$\begin{aligned}C &= (0.95 \times 0.05 \times d) / (0.014 \times 4.2) \\C &= 0.81 \text{ d} \\(1/250 \times 6.8) &= 0.027 \\0.027 \text{ mg/L} &= 0.81 \text{ d} \\d &= 0.034 \text{ mg/day} \\0.034 \text{ mg/day} / 60 \text{ kg} &= 0.0006 \text{ mg/kg/day}\end{aligned}$$

In consideration of uncertainty factors for this MRL approach, multiple factors also apply. In this case, the mean value of 6.8 ppm for the NOAEL is for the entire study cohort at 66 months ( $n = 711$ ). An uncertainty factor of 1.5 was used to account for the pharmacokinetically based variability of hair-to-blood ratios (95% confidence level) in pregnant women and fetuses in the U.S. population (Clewell et al. 1998, 1999). The extremely large size of the study population ( $n=711$ ), in combination with an uncertainty factor of 1.5, is considered adequate to encompass the full range of pharmacokinetic and pharmacodynamic variability within the human population. An independent modifying factor of 1.5 was also used to take into consideration the positive results of the domain-specific tests administered in the Faroe study (Grandjean et al. 1997, 1998). The uncertainty factor of 1.5, multiplied by the modifying factor of 1.5, yields a total aggregate value of 2.25. Applying the factor of 2.25 to the daily intake calculated from the 6.8 ppm NOAEL yields a chronic oral MRL value of 0.0003 mg/kg/day for methylmercury (0.0006 mg/kg/day divided by 2.25 = 0.0003 mg/kg/day).

A third approach to deriving a health guidance value is the use of bench mark dose (BMD) modeling. Clewell et al. (1998) used a benchmark dose analysis to determine a reference dose (RfD, a health guidance value used by the Environmental Protection Agency and, in some ways, the equivalent of ATSDR's chronic oral MRL). Clewell et al. (1998) used the data from the 29-month test in the Seychellois population (Davidson et al. 1995b) for their analysis (i.e., the 66-month study had not been published at the time of their benchmark dose analysis). The BMD is calculated by fitting a mathematical dose-response model to dose-response data. The bench mark dose level (BMDL) is a lower statistical confidence bound on the BMD and replaces the NOAEL in the calculation of a health guidance value. The BMD approach has been proposed as superior to the use of "average" or "grouped" exposure estimates when dose-response information is available, as is the case for the Seychelles study. Clewell et al. (1998) note that the Faroe Islands study reported by Grandjean et al. (1997b) could not be used for dose-response modeling due to inadequate reporting of the data and the confounding influence of co-exposure to PCBs.

For the 29-month Seychelles data, Clewell et al. (1998) used the 95% lower bound on the 10% benchmark dose level (BMDL), which represents a conservative estimate of the traditional NOAEL. The benchmark dose modeling over the entire range of neurological endpoints reported by Davidson et al. (1995b) yielded a lowest BMDL<sub>10</sub> of 21 ppm methylmercury in maternal hair. This BMDL<sub>10</sub> was then converted to an expected distribution of daily ingestion rates across a population of U.S. women of child-bearing age by using a Monte Carlo analysis with a physiologically based pharmacokinetic (PBPK) model of methylmercury developed by Gearhart et al. (1995). This analysis addresses the impact of interindividual

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pharmacokinetic variability on the relationship between ingestion rate and hair concentration for methylmercury. The resulting distribution had a geometric mean value of 0.00160 mg/kg/day (S.D. 0.00133). The 1st, 5th, and 10th percentiles of that distribution were 0.00086, 0.00104, and 0.00115 mg/kg/day, respectively. Clewell et al. (1998) suggested that the 5th percentile of 0.00104 mg/kg/day provides a scientifically based, conservative basis that incorporates the pharmacokinetic variability across the U.S. population of child-bearing women and that no other uncertainty factor for interindividual variability would be needed. To the benchmark-estimated NOAEL of 21 ppm derived from the Seychelles 29-month data, Clewell et al. (1998) applied an uncertainty factor of 3 to account for data base limitations. (Note: The 66-month Seychelles data was not yet published at the time; hence the reliance on the 29-month Seychelles data for the benchmark analysis.) Consequently, Clewell et al. (1998) concluded that using a NOAEL of 7 ppm (21 ppm / 3 (UF) provides additional protection against the possibility that effects could occur at lower concentrations in some populations. Based upon this reasoning, they recommended a health guidance value (i.e., an RfD) of 0.0004 mg/kg/day. If a modifying factor of 1.5 is used to further address the domain-specific findings in the Faroe study, a final MRL of 0.3 µg/kg/day results.

The above benchmark analysis of 29-month data from the Seychelles Child Development Study strongly supports the MRL of 0.0003 mg/kg/day calculated by ATSDR in this profile. Similarly, addressing the Seychellois 66-month data from the perspective of using the mean value (15.3 ppm) of the highest exposure group in the study, a method prescribed in ATSDR's published guidance for MRL development (Chou et al. 1998), also results in an identical MRL. ATSDR therefore has high confidence that this level is protective of the health of all potentially exposed human populations.

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## APPENDIX B

### USER'S GUIDE

#### Chapter 1

##### Public Health Statement

This chapter of the profile is a health effects summary written in non-technical language. Its intended audience is the general public especially people living in the vicinity of a hazardous waste site or chemical release. If the Public Health Statement were removed from the rest of the document, it would still communicate to the lay public essential information about the chemical.

The major headings in the Public Health Statement are useful to find specific topics of concern. The topics are written in a question and answer format. The answer to each question includes a sentence that will direct the reader to chapters in the profile that will provide more information on the given topic.

#### Chapter 2

##### Tables and Figures for Levels of Significant Exposure (LSE)

Tables (2-1, 2-2, and 2-3) and figures (2-1 and 2-2) are used to summarize health effects and illustrate graphically levels of exposure associated with those effects. These levels cover health effects observed at increasing dose concentrations and durations, differences in response by species, minimal risk levels (MRLs) to humans for noncancer end points, and EPA's estimated range associated with an upper-bound individual lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. Use the LSE tables and figures for a quick review of the health effects and to locate data for a specific exposure scenario. The LSE tables and figures should always be used in conjunction with the text. All entries in these tables and figures represent studies that provide reliable, quantitative estimates of No-Observed-Adverse-Effect Levels (NOAELs), Lowest-Observed-Adverse-Effect Levels (LOAELs), or Cancer Effect Levels (CELs).

The legends presented below demonstrate the application of these tables and figures. Representative examples of LSE Table 2-1 and Figure 2-1 are shown. The numbers in the left column of the legends correspond to the numbers in the example table and figure.

##### LEGEND

###### See LSE Table 2-1

- (1) Route of Exposure One of the first considerations when reviewing the toxicity of a substance using these tables and figures should be the relevant and appropriate route of exposure. When sufficient data exists, three LSE tables and two LSE figures are presented in the document. The three LSE tables present data on the three principal routes of exposure, i.e., inhalation, oral, and dermal (LSE Table 2-1, 2-2, and 2-3, respectively). LSE figures are limited to the inhalation (LSE Figure 2-1) and oral (LSE Figure 2-2) routes. Not all substances will have data on each route of exposure and will not therefore have all five of the tables and figures.

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- (2) Exposure Period Three exposure periods - acute (less than 15 days), intermediate (15–364 days), and chronic (365 days or more) are presented within each relevant route of exposure. In this example, an inhalation study of intermediate exposure duration is reported. For quick reference to health effects occurring from a known length of exposure, locate the applicable exposure period within the LSE table and figure.
- (3) Health Effect The major categories of health effects included in LSE tables and figures are death, systemic, immunological, neurological, developmental, reproductive, and cancer. NOAELs and LOAELs can be reported in the tables and figures for all effects but cancer. Systemic effects are further defined in the "System" column of the LSE table (see key number 18).
- (4) Key to Figure Each key number in the LSE table links study information to one or more data points using the same key number in the corresponding LSE figure. In this example, the study represented by key number 18 has been used to derive a NOAEL and a Less Serious LOAEL (also see the 2 "18r" data points in Figure 2-1).
- (5) Species The test species, whether animal or human, are identified in this column. Section 2.5, "Relevance to Public Health," covers the relevance of animal data to human toxicity and Section 2.3, "Toxicokinetics," contains any available information on comparative toxicokinetics. Although NOAELs and LOAELs are species specific, the levels are extrapolated to equivalent human doses to derive an MRL.
- (6) Exposure Frequency/Duration The duration of the study and the weekly and daily exposure regimen are provided in this column. This permits comparison of NOAELs and LOAELs from different studies. In this case (key number 18), rats were exposed to 1,1,2,2-tetrachloroethane via inhalation for 6 hours per day, 5 days per week, for 3 weeks. For a more complete review of the dosing regimen refer to the appropriate sections of the text or the original reference paper, i.e., Nitschke et al. 1981.
- (7) System This column further defines the systemic effects. These systems include: respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, and dermal/ocular. "Other" refers to any systemic effect (e.g., a decrease in body weight) not covered in these systems. In the example of key number 18, 1 systemic effect (respiratory) was investigated.
- (8) NOAEL A No-Observed-Adverse-Effect Level (NOAEL) is the highest exposure level at which no harmful effects were seen in the organ system studied. Key number 18 reports a NOAEL of 3 ppm for the respiratory system which was used to derive an intermediate exposure, inhalation MRL of 0.005 ppm (see footnote "b").
- (9) LOAEL A Lowest-Observed-Adverse-Effect Level (LOAEL) is the lowest dose used in the study that caused a harmful health effect. LOAELs have been classified into "Less Serious" and "Serious" effects. These distinctions help readers identify the levels of exposure at which adverse health effects first appear and the gradation of effects with increasing dose. A brief description of the specific endpoint used to quantify the adverse effect accompanies the LOAEL. The respiratory effect reported in key number 18 (hyperplasia) is a Less serious LOAEL of 10 ppm. MRLs are not derived from Serious LOAELs.
- (10) Reference The complete reference citation is given in chapter 8 of the profile.



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- (11) CEL A Cancer Effect Level (CEL) is the lowest exposure level associated with the onset of carcinogenesis in experimental or epidemiologic studies. CELs are always considered serious effects. The LSE tables and figures do not contain NOAELs for cancer, but the text may report doses not causing measurable cancer increases.
- (12) Footnotes Explanations of abbreviations or reference notes for data in the LSE tables are found in the footnotes. Footnote "b" indicates the NOAEL of 3 ppm in key number 18 was used to derive an MRL of 0.005 ppm.

**LEGEND****See Figure 2-1**

LSE figures graphically illustrate the data presented in the corresponding LSE tables. Figures help the reader quickly compare health effects according to exposure concentrations for particular exposure periods.

- (13) Exposure Period The same exposure periods appear as in the LSE table. In this example, health effects observed within the intermediate and chronic exposure periods are illustrated.
- (14) Health Effect These are the categories of health effects for which reliable quantitative data exists. The same health effects appear in the LSE table.
- (15) Levels of Exposure concentrations or doses for each health effect in the LSE tables are graphically displayed in the LSE figures. Exposure concentration or dose is measured on the log scale "y" axis. Inhalation exposure is reported in mg/m<sup>3</sup> or ppm and oral exposure is reported in mg/kg/day.
- (16) NOAEL In this example, 18r NOAEL is the critical endpoint for which an intermediate inhalation exposure MRL is based. As you can see from the LSE figure key, the open-circle symbol indicates to a NOAEL for the test species-rat. The key number 18 corresponds to the entry in the LSE table. The dashed descending arrow indicates the extrapolation from the exposure level of 3 ppm (see entry 18 in the Table) to the MRL of 0.005 ppm (see footnote "b" in the LSE table).
- (17) CEL Key number 38r is 1 of 3 studies for which Cancer Effect Levels were derived. The diamond symbol refers to a Cancer Effect Level for the test species-mouse. The number 38 corresponds to the entry in the LSE table.
- (18) Estimated Upper-Bound Human Cancer Risk Levels This is the range associated with the upper-bound for lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. These risk levels are derived from the EPA's Human Health Assessment Group's upper-bound estimates of the slope of the cancer dose response curve at low dose levels ( $q_1^*$ ).
- (19) Key to LSE Figure The Key explains the abbreviations and symbols used in the figure.

**SAMPLE**

1 6

**TABLE 2-1. Levels of Significant Exposure to [Chemical x] – Inhalation**

Key to figure <sup>a</sup>	Species	Exposure frequency/duration	System	NOAEL (ppm)	LOAEL (effect)		Reference
					Less serious (ppm)	Serious (ppm)	
<b>INTERMEDIATE EXPOSURE</b>							
2 6	5	6	7	8	9		10
3 6	Systemic	9	9	9	9		9
4 6	18	Rat	13 wk 5d/wk 6hr/d	Resp	3 <sup>b</sup>	10 (hyperplasia)	Nitschke et al. 1981
<b>CHRONIC EXPOSURE</b>							
						11	
	Cancer					9	
38	Rat	18 mo 5d/wk 7hr/d				20	(CEL, multiple organs) Wong et al. 1982
39	Rat	89–104 wk 5d/wk 6hr/d				10	(CEL, lung tumors, nasal tumors) NTP 1982
40	Mouse	79–103 wk 5d/wk 6hr/d				10	(CEL, lung tumors, hemangiosarcomas) NTP 1982

12 6

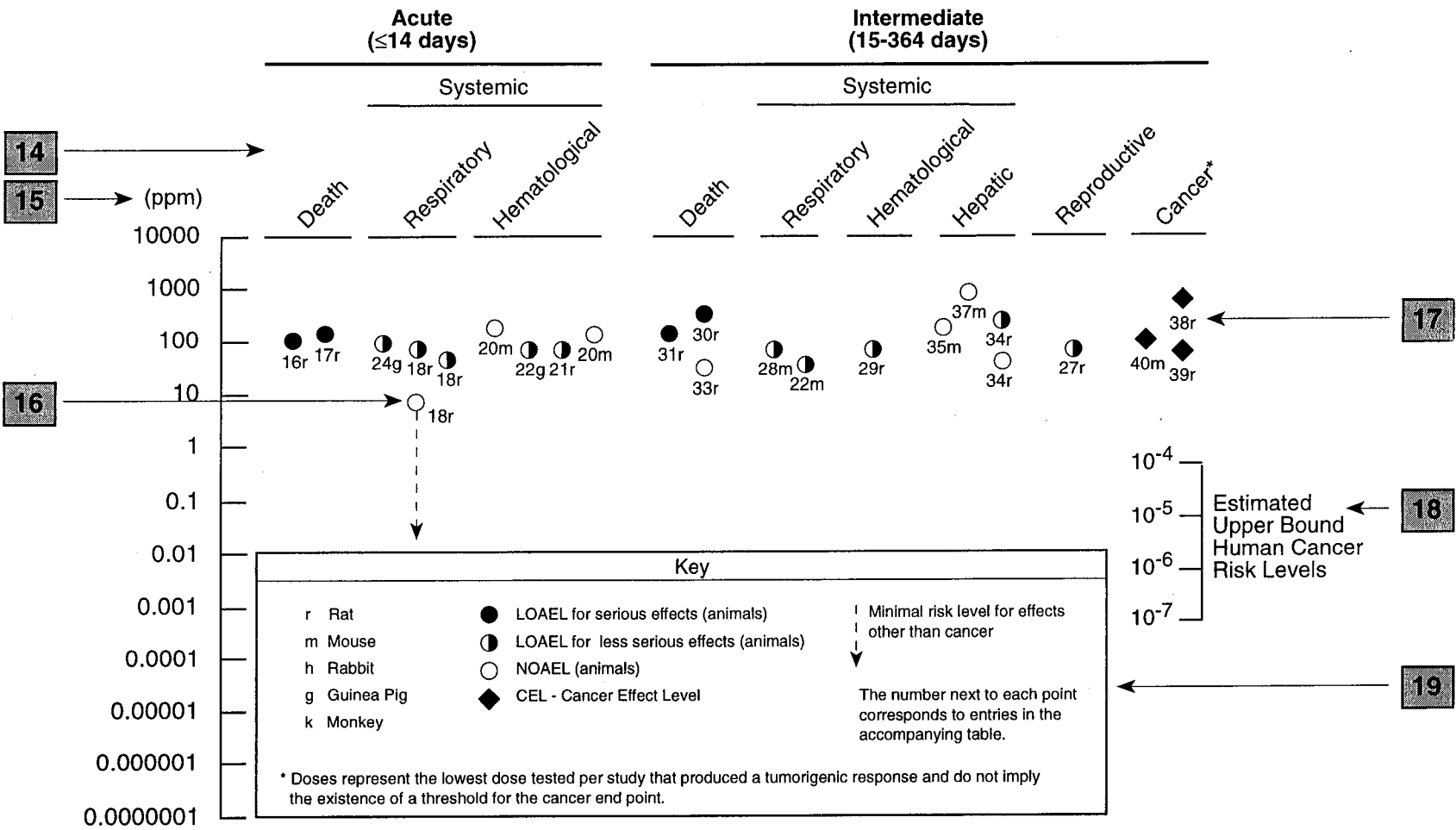
<sup>a</sup> The number corresponds to entries in Figure 2-1.

<sup>b</sup> Used to derive an intermediate inhalation Minimal Risk Level (MRL) of 5x10<sup>-3</sup> ppm; dose adjusted for intermittent exposure and divided by an uncertainty factor of 100 (10 for extrapolation from animal to humans, 10 for human variability).

CEL = cancer effect level; d = days(s); hr = hour(s); LOAEL = lowest-observed-adverse-effect level; mo = month(s); NOAEL = no-observed-adverse-effect level; Resp = respiratory; wk = week(s)

# SAMPLE

**13** → **Figure 2-1. Levels of Significant Exposure to [Chemical X] – Inhalation**



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**Chapter 2 (Section 2.5)****Relevance to Public Health**

The Relevance to Public Health section provides a health effects summary based on evaluations of existing toxicologic, epidemiologic, and toxicokinetic information. This summary is designed to present interpretive, weight-of-evidence discussions for human health end points by addressing the following questions.

1. What effects are known to occur in humans?
2. What effects observed in animals are likely to be of concern to humans?
3. What exposure conditions are likely to be of concern to humans, especially around hazardous waste sites?

The section covers end points in the same order they appear within the Discussion of Health Effects by Route of Exposure section, by route (inhalation, oral, dermal) and within route by effect. Human data are presented first, then animal data. Both are organized by duration (acute, intermediate, chronic). *In vitro* data and data from parenteral routes (intramuscular, intravenous, subcutaneous, etc.) are also considered in this section. If data are located in the scientific literature, a table of genotoxicity information is included.

The carcinogenic potential of the profiled substance is qualitatively evaluated, when appropriate, using existing toxicokinetic, genotoxic, and carcinogenic data. ATSDR does not currently assess cancer potency or perform cancer risk assessments. Minimal risk levels (MRLs) for noncancer end points (if derived) and the end points from which they were derived are indicated and discussed.

Limitations to existing scientific literature that prevent a satisfactory evaluation of the relevance to public health are identified in the Data Needs section.

**Interpretation of Minimal Risk Levels**

Where sufficient toxicologic information is available, we have derived minimal risk levels (MRLs) for inhalation and oral routes of entry at each duration of exposure (acute, intermediate, and chronic). These MRLs are not meant to support regulatory action; but to acquaint health professionals with exposure levels at which adverse health effects are not expected to occur in humans. They should help physicians and public health officials determine the safety of a community living near a chemical emission, given the concentration of a contaminant in air or the estimated daily dose in water. MRLs are based largely on toxicological studies in animals and on reports of human occupational exposure.

MRL users should be familiar with the toxicologic information on which the number is based. Chapter 2.5, "Relevance to Public Health," contains basic information known about the substance. Other sections such as 2.8, "Interactions with Other Substances," and 2.9, "Populations that are Unusually Susceptible" provide important supplemental information.

MRL users should also understand the MRL derivation methodology. MRLs are derived using a modified version of the risk assessment methodology the Environmental Protection Agency (EPA) provides (Barnes and Dourson 1988) to determine reference doses for lifetime exposure (RfDs).

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To derive an MRL, ATSDR generally selects the most sensitive endpoint which, in its best judgement, represents the most sensitive human health effect for a given exposure route and duration. ATSDR cannot make this judgement or derive an MRL unless information (quantitative or qualitative) is available for all potential systemic, neurological, and developmental effects. If this information and reliable quantitative data on the chosen endpoint are available, ATSDR derives an MRL using the most sensitive species (when information from multiple species is available) with the highest NOAEL that does not exceed any adverse effect levels. When a NOAEL is not available, a lowest-observed-adverse-effect level (LOAEL) can be used to derive an MRL, and an uncertainty factor (UF) of 10 must be employed. Additional uncertainty factors of 10 must be used both for human variability to protect sensitive subpopulations (people who are most susceptible to the health effects caused by the substance) and for interspecies variability (extrapolation from animals to humans). In deriving an MRL, these individual uncertainty factors are multiplied together. The product is then divided into the inhalation concentration or oral dosage selected from the study. Uncertainty factors used in developing a substance-specific MRL are provided in the footnotes of the LSE Tables.



**APPENDIX C****ACRONYMS, ABBREVIATIONS, AND SYMBOLS**

ACGIH	American Conference of Governmental Industrial Hygienists
ADI	Acceptable Daily Intake
ADME	Absorption, Distribution, Metabolism, and Excretion
AFID	alkali flame ionization detector
AFOSH	Air Force Office of Safety and Health
AML	acute myeloid leukemia
AOAC	Association of Official Analytical Chemists
atm	atmosphere
ATSDR	Agency for Toxic Substances and Disease Registry
AWQC	Ambient Water Quality Criteria
BAT	Best Available Technology
BCF	bioconcentration factor
BEI	Biological Exposure Index
BSC	Board of Scientific Counselors
C	Centigrade
CAA	Clean Air Act
CAG	Cancer Assessment Group of the U.S. Environmental Protection Agency
CAS	Chemical Abstract Services
CDC	Centers for Disease Control and Prevention
CEL	Cancer Effect Level
CELDS	Computer-Environmental Legislative Data System
CERCLA	Comprehensive Environmental Response, Compensation, and Liability Act
CFR	Code of Federal Regulations
Ci	curie
CL	ceiling limit value
CLP	Contract Laboratory Program
cm	centimeter
CML	chronic myeloid leukemia
CNS	central nervous system
CPSC	Consumer Products Safety Commission
CWA	Clean Water Act
d	day
Derm	dermal
DHEW	Department of Health, Education, and Welfare
DHHS	Department of Health and Human Services
DNA	deoxyribonucleic acid
DOD	Department of Defense
DOE	Department of Energy
DOL	Department of Labor
DOT	Department of Transportation
DOT/UN/ NA/IMCO	Department of Transportation/United Nations/ North America/International Maritime Dangerous Goods Code
DWEL	Drinking Water Exposure Level

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ECD	electron capture detection
ECG/EKG	electrocardiogram
EEG	electroencephalogram
EEGL	Emergency Exposure Guidance Level
EPA	Environmental Protection Agency
F	Fahrenheit
F <sub>1</sub>	first-filial generation
FAO	Food and Agricultural Organization of the United Nations
FDA	Food and Drug Administration
FEMA	Federal Emergency Management Agency
FIFRA	Federal Insecticide, Fungicide, and Rodenticide Act
FPD	flame photometric detection
fpm	feet per minute
ft	foot
FR	<i>Federal Register</i>
g	gram
GC	gas chromatography
Gd	gestational day
gen	generation
GLC	gas liquid chromatography
GPC	gel permeation chromatography
HPLC	high-performance liquid chromatography
hr	hour
HRGC	high resolution gas chromatography
HSDB	Hazardous Substance Data Bank
IDLH	Immediately Dangerous to Life and Health
IARC	International Agency for Research on Cancer
ILO	International Labor Organization
in	inch
IRIS	Integrated Risk Information System
K <sub>d</sub>	adsorption ratio
kg	kilogram
kgg	metric ton
K <sub>oc</sub>	organic carbon partition coefficient
K <sub>ow</sub>	octanol-water partition coefficient
L	liter
LC	liquid chromatography
LC <sub>Lo</sub>	lethal concentration, low
LC <sub>50</sub>	lethal concentration, 50% kill
LD <sub>Lo</sub>	lethal dose, low
LD <sub>50</sub>	lethal dose, 50% kill
LT <sub>50</sub>	lethal time, 50% kill
LOAEL	lowest-observed-adverse-effect level
LSE	Levels of Significant Exposure
m	meter
MA	<i>trans,trans</i> -muconic acid
MAL	Maximum Allowable Level
mCi	millicurie
MCL	Maximum Contaminant Level



## APPENDIX C

MCLG	Maximum Contaminant Level Goal
mg	milligram
min	minute
mL	milliliter
mm	millimeter
mm Hg	millimeters of mercury
mmol	millimole
mo	month
mppcf	millions of particles per cubic foot
MRL	Minimal Risk Level
MS	mass spectrometry
NAAQS	National Ambient Air Quality Standard
NAS	National Academy of Science
NATICH	National Air Toxics Information Clearinghouse
NATO	North Atlantic Treaty Organization
NCE	normochromatic erythrocytes
NCI	National Cancer Institute
NIEHS	National Institute of Environmental Health Sciences
NIOSH	National Institute for Occupational Safety and Health
NIOSH TIC	NIOSH's Computerized Information Retrieval System
NFPA	National Fire Protection Association
ng	nanogram
NLM	National Library of Medicine
nm	nanometer
NHANES	National Health and Nutrition Examination Survey
nmol	nanomole
NOAEL	no-observed-adverse-effect level
NOES	National Occupational Exposure Survey
NOHS	National Occupational Hazard Survey
NPD	nitrogen phosphorus detection
NPDES	National Pollutant Discharge Elimination System
NPL	National Priorities List
NR	not reported
NRC	National Research Council
NS	not specified
NSPS	New Source Performance Standards
NTIS	National Technical Information Service
NTP	National Toxicology Program
ODW	Office of Drinking Water, EPA
OERR	Office of Emergency and Remedial Response, EPA
OHM/TADS	Oil and Hazardous Materials/Technical Assistance Data System
OPP	Office of Pesticide Programs, EPA
OPPTS	Office of Prevention, Pesticides and Toxic Substances, EPA
OPPT	Office of Pollution Prevention and Toxics, EPA
OSHA	Occupational Safety and Health Administration
OSW	Office of Solid Waste, EPA
OTS	Office of Toxic Substances
OW	Office of Water
OWRS	Office of Water Regulations and Standards, EPA

## APPENDIX C

PAH	Polycyclic Aromatic Hydrocarbon
PBPD	Physiologically Based Pharmacodynamic
PBPK	Physiologically Based Pharmacokinetic
PCE	polychromatic erythrocytes
PEL	permissible exposure limit
PID	photo ionization detector
pg	picogram
pmol	picomole
PHS	Public Health Service
PMR	proportionate mortality ratio
ppb	parts per billion
ppm	parts per million
ppt	parts per trillion
PSNS	Pretreatment Standards for New Sources
REL	recommended exposure level/limit
RfC	Reference Concentration
RfD	Reference Dose
RNA	ribonucleic acid
RTECS	Registry of Toxic Effects of Chemical Substances
RQ	Reportable Quantity
SARA	Superfund Amendments and Reauthorization Act
SCE	sister chromatid exchange
sec	second
SIC	Standard Industrial Classification
SIM	selected ion monitoring
SMCL	Secondary Maximum Contaminant Level
SMR	standard mortality ratio
SNARL	Suggested No Adverse Response Level
SPEGL	Short-Term Public Emergency Guidance Level
STEL	short-term exposure limit
STORET	Storage and Retrieval
TD <sub>50</sub>	toxic dose, 50% specific toxic effect
TLV	threshold limit value
TOC	Total Organic Compound
TPQ	Threshold Planning Quantity
TRI	Toxics Release Inventory
TSCA	Toxic Substances Control Act
TRI	Toxics Release Inventory
TWA	time-weighted average
U.S.	United States
UF	uncertainty factor
VOC	Volatile Organic Compound
yr	year
WHO	World Health Organization
wk	week
>	greater than
≥	greater than or equal to
=	equal to

## APPENDIX C

<	less than
≤	less than or equal to
%	percent
α	alpha
β	beta
γ	gamma
δ	delta
μm	micrometer
μg	microgram
q <sub>1</sub> *	cancer slope factor
-	negative
+	positive
(+)	weakly positive result
(-)	weakly negative result

