

IRAC Public Health Team



August 25th , 2006

1st Meeting

August 25th, 2006



WHO Headquarters, Geneva, Switzerland



IRAC Public Health Team

August 25th, 2006

Members

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Observer

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Issues to be covered

- Disease vector resistance, e.g. mosquitoes**
- Professional use**
 - Resistance issues in**
 - Cockroaches**
 - Houseflies**
 - Fleas**
 - Bed bugs**
- Resistance risk assessment in public health**
- Resistance management issues non-ag pests**



IRAC Public Health Team: 1st Meeting

WHO Geneva, 25/08/2006, Room C202

Agenda

- 08.30 - 9.00 Opening session (R. Nauen, P. Guillet, A. Kochi, Director, Global Malaria Programme), introduction of participants.
- 09.00 - 09.45 Finalization of the Vector Manual and next steps (All)
- 09.45 - 10.15 Insecticide mode of action in agriculture vs public health, IRAC's classification scheme and mechanisms of resistance with special reference to vector control (R. Nauen)
- 10.15 -10.30 Coffee break**



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- 10.30 - 11.00 Vector control program of the Gates Foundation (K. Aultman)
- 11.00 - 11.30 Resistance as a field issue: consequences of vector ecology, behaviour, population dynamics and population genetics in development and spread of insecticide resistance. Current status of vector resistance and regional networks (P. Guillet).
- 11.30 - 12.00 Past and current problems faced in monitoring vector resistance (test kits, collection, validation and dissemination of information) (P. Guillet)
- 12.00 - 12.45 Discussions
- 12.45 - 14.00 Lunch Break**



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- 14.00 - 15.00 Team discussion on selected points raised during concalls (all)
- Views on industry production of insecticide impregnated filter papers?
 - Individual reports on industry supported monitoring programs
 - How to arrange for a freeze storage stability test for impregnated filter papers?
 - Regional vs global RM programs – Joint efforts between all parties on research initiatives (e.g. topics, supervision, funding etc.)
 - Reference centers
- 15.00 -16.00 Short-term and long-term goals of the IRAC Public Health Team (all)
- 16.00 – 16.30 Joining forces in monitoring and management of vector resistance (all participants, tea served in the room)
- 16.30 – 17.00 AOB (next concall)
- 17.00 Departure to airport**



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Check of completeness – Vector Manual

Avoiding the Development of Insecticide Resistance in Vectors and Pests of Public Health Importance

Pierre & Kate: to review Ch 2 & 10

Ralf: to review Ch 1,3,4 & 5 (done)

Mark/Phil: to review Ch 6 (done)

Robin/John: to review Ch 7 & 8 (done)

Janet/Ralf: to review Ch 9 (done)

Next steps...

IRAC Public Health Team



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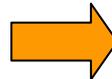


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EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION (EPPO)

EPPO standard on Resistance Risk Analysis (PP 1/213)



Specific scope

This standard describes how the risk of resistance to plant protection products can be assessed and, if appropriate, systems for risk management can be proposed, in the context of official registration of plant protection products.

1. *Bemisia tabaci*
2. *Trialeurodes vaporariorum*
3. *Aphis gossypii*
4. *Myzus persicae*
5. *Leptinotarsa decemlineata*
6. *Spodoptera exigua*
7. *Frankliniella occidentalis*
8. *Tetranychus urticae*
9. *Panonychus ulmi*
10. *Cydia pomonella*
11. *Phorodon humuli*

Examples of species in the EPPO region which have developed resistance and for which sensitivity data should normally be provided



Key issues – resistance risk assessment

Monitoring



Mechanisms



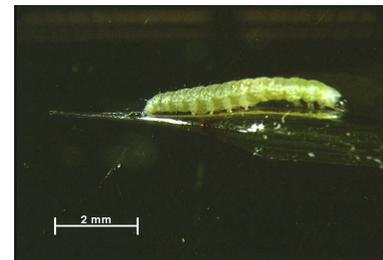
Management



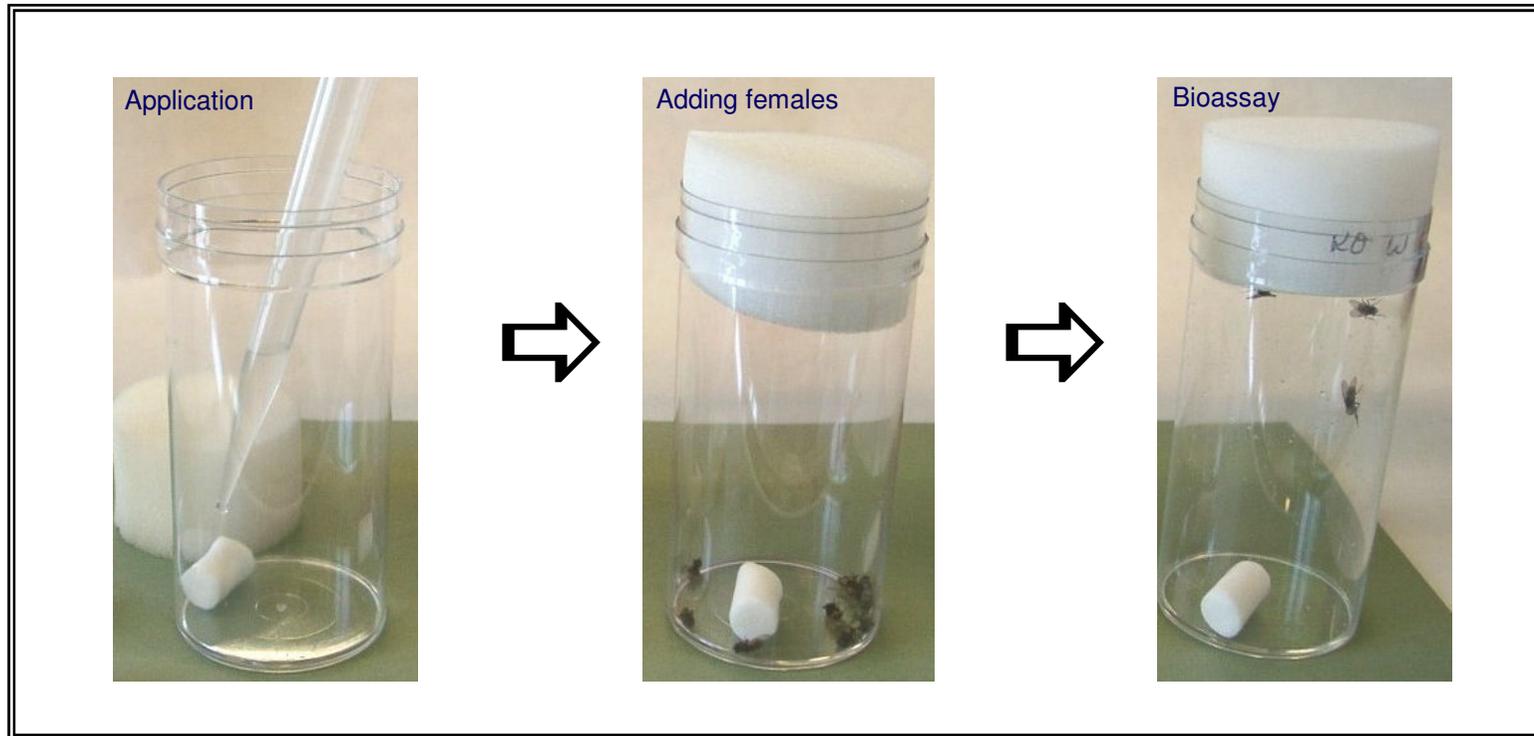
Baseline studies

Bioassay methods are as diverse as the functional morphology of pest species and depend on the physicochemical a.i. properties

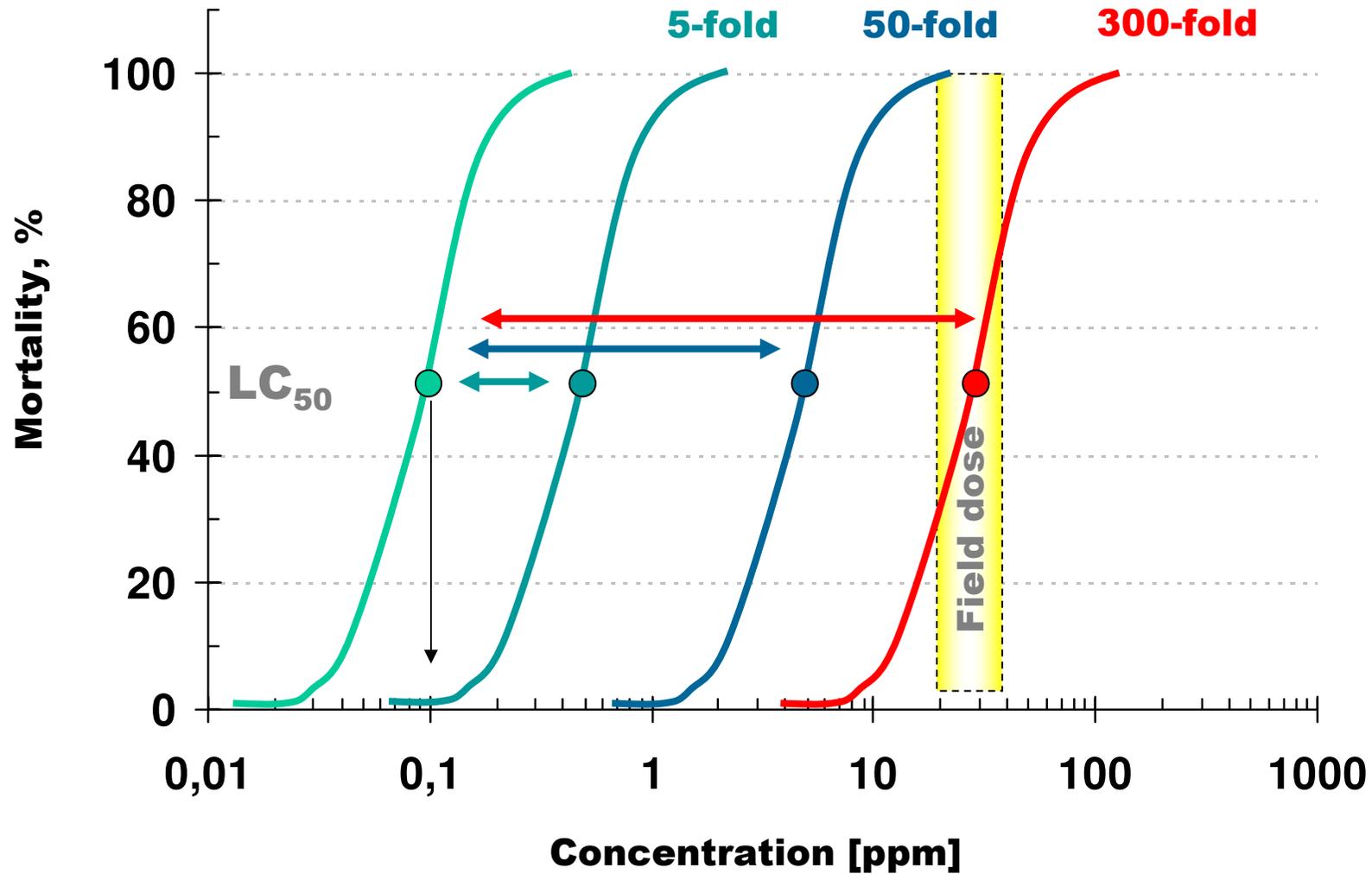
- Topical application
- Insect/leaf-dip tests
- Artificial diet tests
- Spray bioassays
- Filter paper assays
- Adult vial tests
- Systemic bioassays



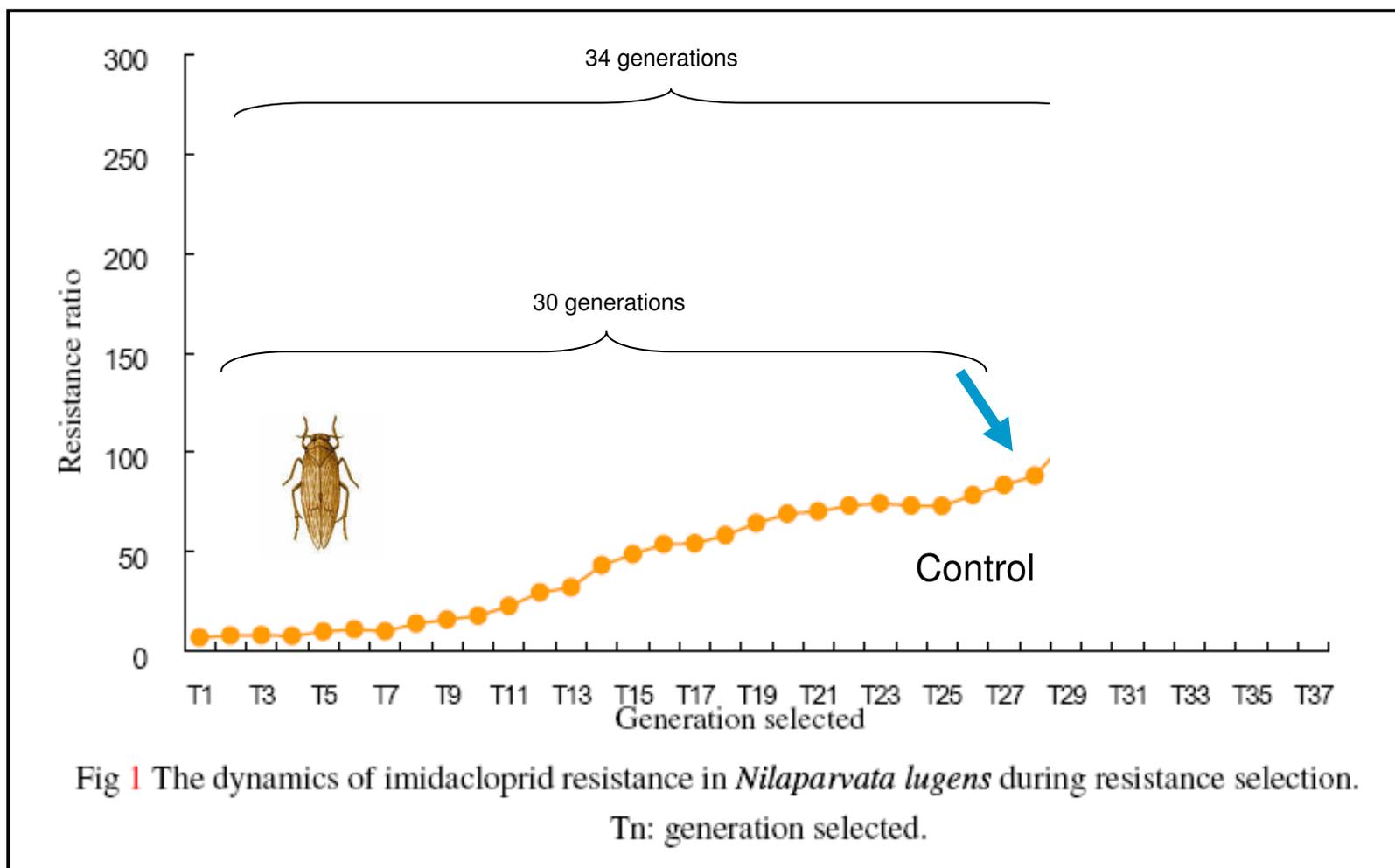
Feeding bioassay to monitor imidacloprid susceptibility in *Musca domestica*



Why to monitor for resistance?



Selection for imidacloprid resistance in BPH*



Liu & Han, *Pest Manag Sci*, 2006

*BPH = Brown planthopper, *Nilaparvata lugens*

Case study

Pollen beetle resistance to pyrethroid insecticides

Winter oilseed rape acreage Germany: 1,300,000 ha
Registered products for beetle control: 8
Mode of action classes: 1 (!)
Generations per season: 1



Meligethes aeneus

- most important pest in oilseed rape
- damage of the flowering structures
- reduction in number of buds

Insecticidal products in the German oilseed-rape market

➔ only pyrethroids!



Active ingredient:

1. Fastac 100 SC
2. Karate Zeon 100 CS
3. Trafo 50 WG
4. Decis flüssig 25 EC
5. Ripcord 100 EC
6. Sumicidin Alpha 50 EC
7. Baythroid 50 EC
8. Bulldock 25 EC

Alpha-Cypermethrin
Lambda-Cyhalothrin
Lambda-Cyhalothrin
Deltamethrin
Cypermethrin
Esfenvalerate
Cyfluthrin
Beta-Cyfluthrin

IRAC

Mode of Action Classification
The key to Insecticide Resistance Management

Group 1: Acetylcholine esterase Inhibitors*

1A: Carbamates

1B: Organophosphates

Group 2: GABA-gated chloride channel antagonists

2A: Cyclopyrimetholides

2B: Flupyrifur (or flupyrifluorol)

Group 3: Sodium channel modulators*

3A: Pyrethroids

Group 4: Nicotinic Acetylcholine receptor agonists/antagonists

4A: Neonicotinoids

4B

4C: Imidacloprid & acetamiprid

Group 5: Nicotinic Acetylcholine Receptor Agonists (not 4)

Group 6: Chloride Channel activators

Group 7: Juvenile hormone mimics

7A: Analogs

7B: Fenoxycarb

7C: Pyriproxyfen

Group 8: Unknown or non-specific - fumigants

8A

8B

8C

Group 9: Unknown or Non-specific - selective feeding blockers

9A

9B

9C

Group 10: Unknown or Non-specific - Mite Growth Inhibitors

10A

10B

Group 11: Microbial disruptors of insect mid-gut membrane - includes transgenic crops expressing Bt toxin, Bt. Var. known

11A1

11A2

11B1

11B2

11C

Group 12: Inhibitors of oxidative phosphorylation, disruptors of ATP formation (inhibitors of ATP synthase)

12A

12B

12C

Group 13: Uncouplers of oxidative phosphorylation via disruption of H⁺ proton gradient

Group 14: Unallocated

Group 15: Inhibitors of chitin biosynthesis, type 0, Lipid transfer

Group 16: Inhibitors of chitin biosynthesis, type 1, Homocysteine

Group 17: Moulting disruptor, Chitinase

Group 18: Ecdysone agonists Moulting disruptors

18A: Diacylglycerols

18B

Group 19: Octadecanoyl agonists

Group 20: Coupling site II electron transport inhibitors (Complex III)

20A

20B

20C

Group 21: Coupling site I electron transport inhibitors

21A: METI acaricides & Rotenone

Group 22: Voltage dependent Sodium Channel blockers

Group 23: Inhibitors of lipid synthesis

23A: Trehalose diuretics

Group 24: Mitochondrial complex IV electron transport inhibitors

24A

24B

24C

Group 25: Neuronal Inhibitors (unknown mode of action)

Group 26: Acetylcholinesterase inhibitors

Group 27: Synergists

27A: Esterase Inhibitors

27B: P450 mono-oxygenase inhibitors

Group 28: Muscarinic receptor modulator

Group UN: Compounds with unknown mode of action

UNA

UNB

UNC

UND

Group NS: Miscellaneous non-specific (multi-site) inhibitors

NSA

NSB

More information on the Insecticide Resistance Action Committee and the Mode of Action Classification is available from www.irac-online.org or from Alan Porter e-mail: sporber@ntra.sipin.com

Structures are reproduced from the Pesticide Manual with permission from the British Crop Protection Council

* Only major representatives of the groups are shown

IRAC Mode of Action Classification v5.1, September 2005 ¹

| Main Group and Primary Site of Action | Chemical Sub-group or exemplifying Active Ingredient | Active Ingredients |
|---------------------------------------|--|---|
| 3 Sodium channel modulators | DDT Methoxychlor | DDT Methoxychlor |
| | Pyrethroids | Acrinathrin, Allethrin, d-cis-trans Allethrin, d-trans Allethrin, Bifenthrin , Bioallethrin, Bioallethrin S-cyclopentenyl , Bioresmethrin, Cycloprothrin, Cyfluthrin, beta-Cyfluthrin , Cyhalothrin, lambda-Cyhalothrin , gamma-Cyhalothrin, Cypermethrin , alpha-Cypermethrin, beta-Cypermethrin, theta-cypermethrin, zeta-Cypermethrin, Cyphenothrin , (1 <i>R</i>)- <i>trans</i> -isomers], Deltamethrin , Empenthrin , (E <i>Z</i>)- (1 <i>R</i>)- isomers], Esfenvalerate, Etofenprox , Fenpropathrin, Fenvalerate, Flucythrinate, Flumethrin, tau-Fluvalinate , Halfenprox, Imiprothrin, Permethrin, Phenothrin [(1 <i>R</i>)- <i>trans</i> - isomer], Prallethrin, Resmethrin, RU 15525, Silafluofen, Tefluthrin, Tetramethrin, Tetramethrin [(1 <i>R</i>)-isomers], Tralomethrin, Transfluthrin, ZXI 8901 |

This document has been prepared using the most up-to-date information available to IRAC. It is provided to user groups, grower organisations, extension personnel, regulatory authorities such as the US EPA and all those involved in resistance management, as an agreed definitive statement by the agrochemical industry on the mode of action of insecticides currently in use.

Resistance monitoring bioassay

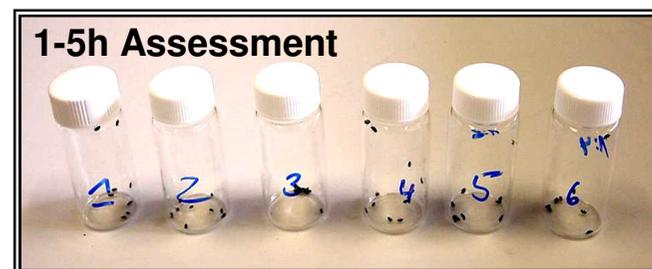
- Baseline-susceptibility
- Monitoring



Pyrethroids

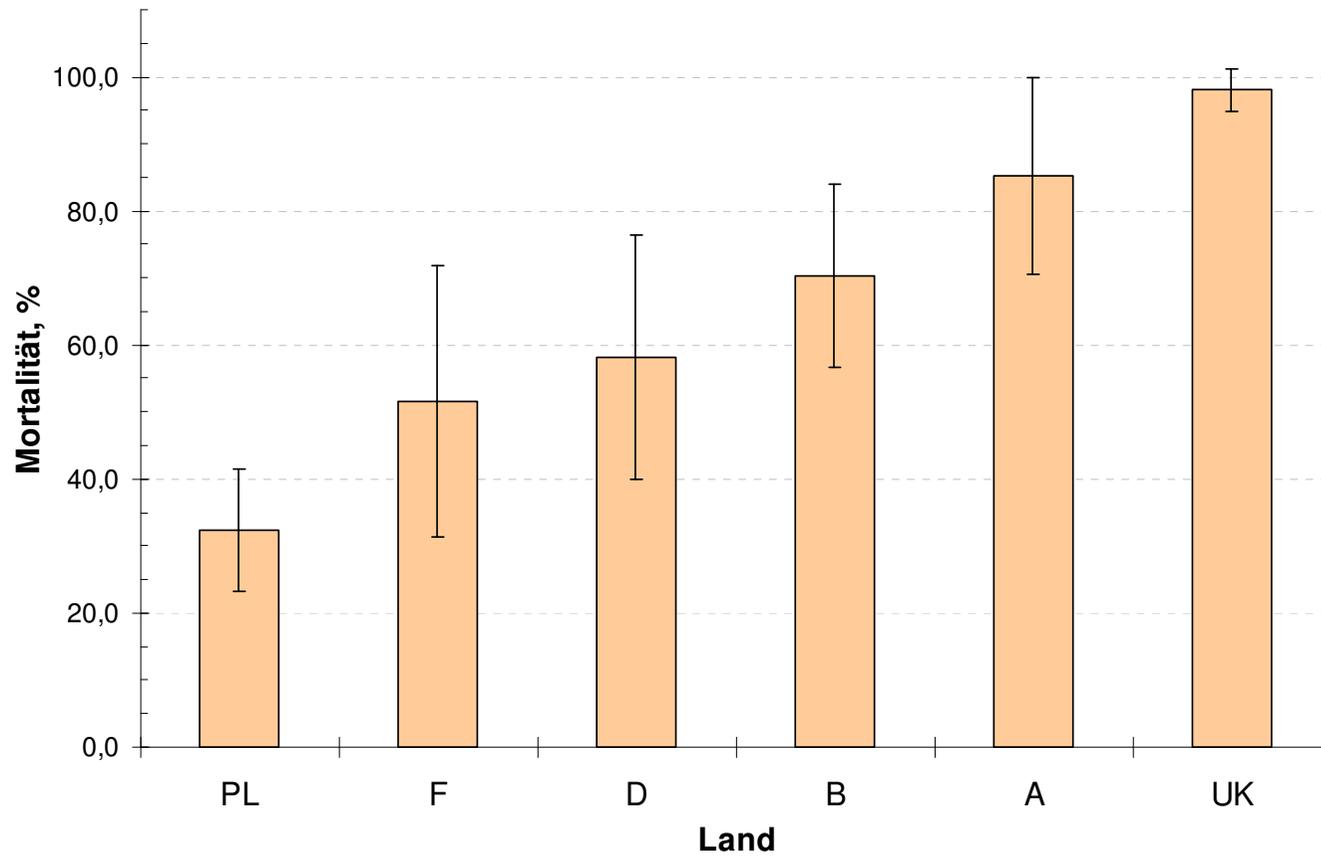
Organophosphates

AVT
(Adult vial test)



Pollen beetle pyrethroid resistance monitoring in different European countries – mean values \pm SD of all populations checked for each country in 2006

AVT - lambda-Cyhalothrin (20% of the field rate, 24h)



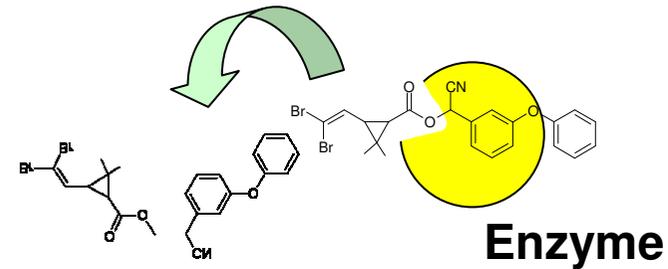
Resistance mechanisms

Some important examples leading to field failure of insecticides under practical conditions in agriculture

Resistance mechanisms

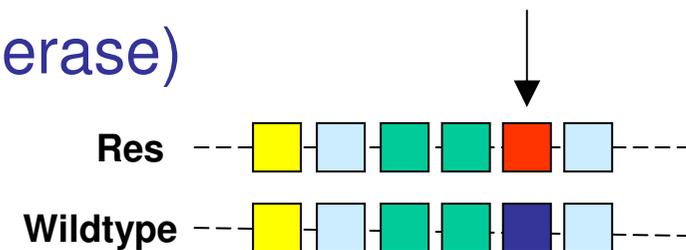
Metabolic Resistance (enzymatic cleavage)

- Esterases
- Monooxygenases
- Glutathione S-transferases



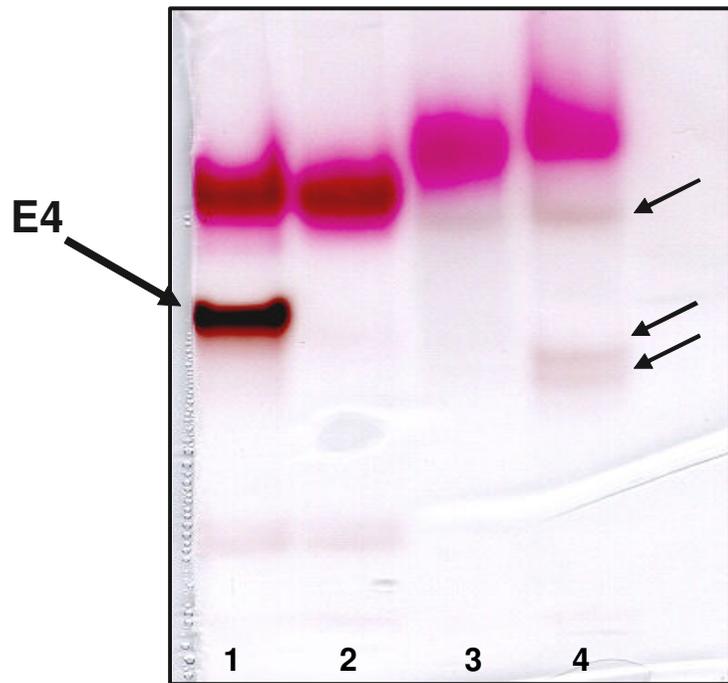
Target site resistance

- MACE (Modified acetylcholinesterase)
- Kdr („Knock-down-resistance“)
- Rdl („Resistance to dieldrin“)



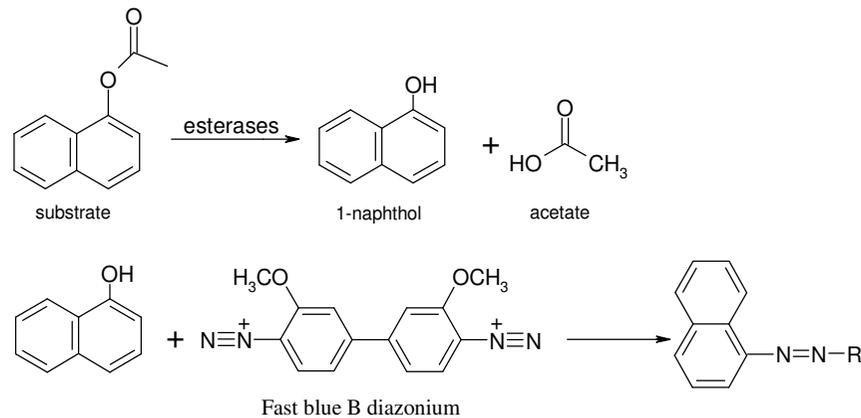
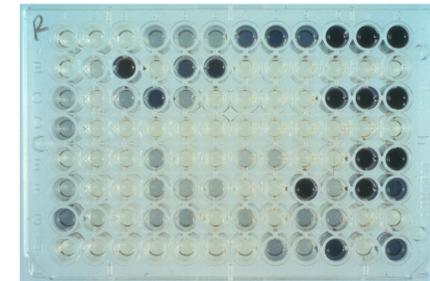
Reduced penetration

Esterases confer OP, carbamate and pyrethroid resistance in aphids such as *Myzus persicae*



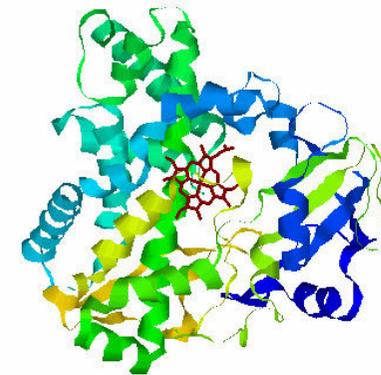
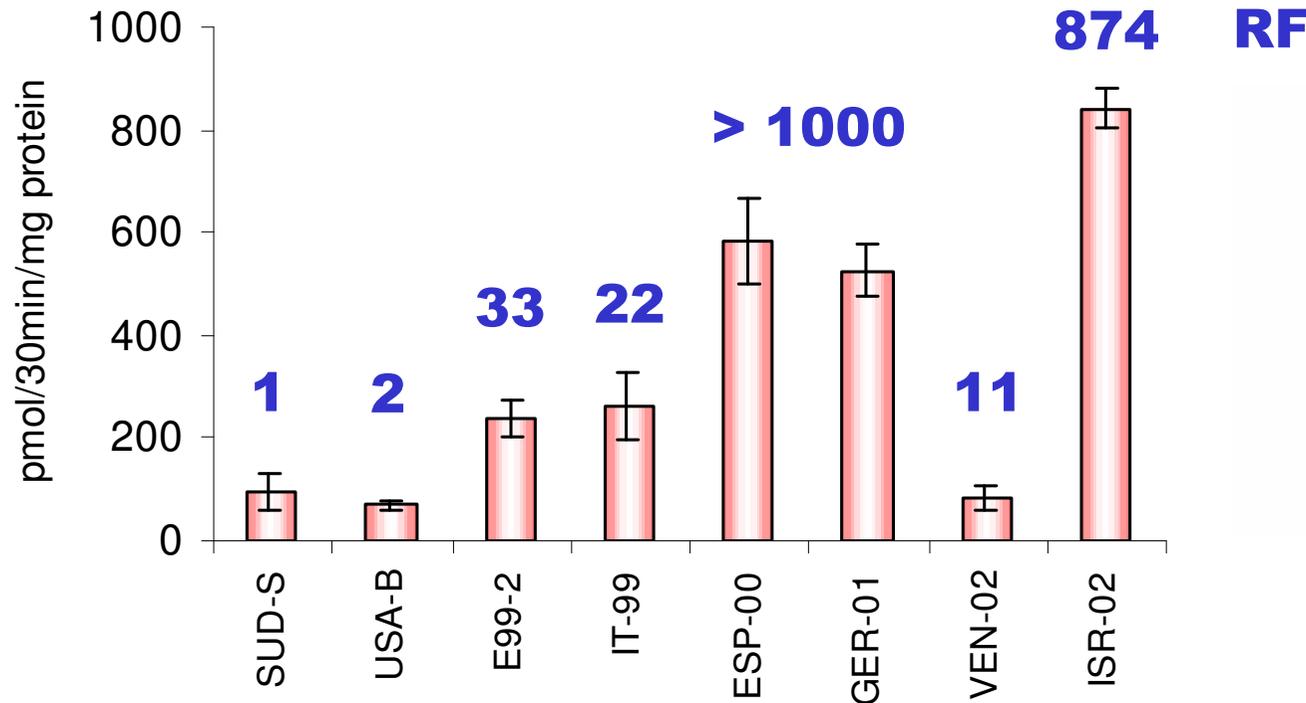
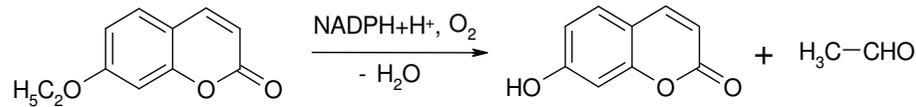
Native PAGE of aphid homogenates

- | | | |
|---|-------------------|--------|
| 1 | <i>M.persicae</i> | JR |
| 2 | | NS |
| 3 | <i>P.humuli</i> | H2-99 |
| 4 | | H08-03 |



Nauen et al. (1996) Bull, Entomol. Res. 86

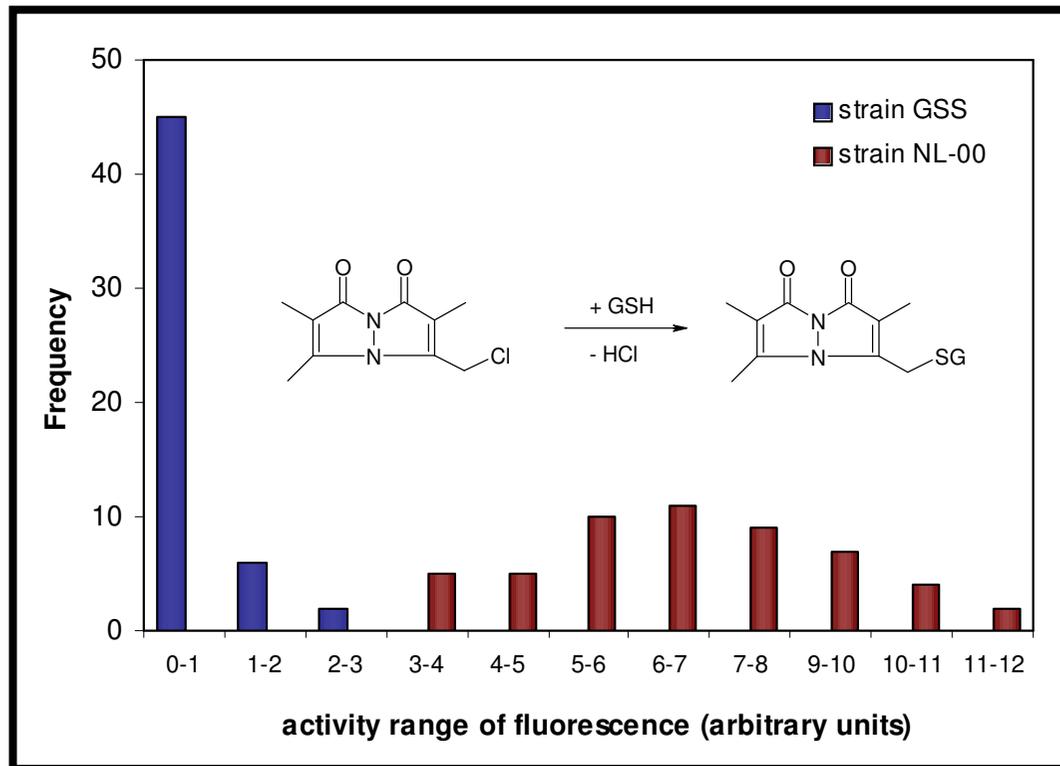
Monoxygenases confer neonicotinoid resistance in cotton whiteflies, *Bemisia tabaci*



Rauch & Nauen (2003) Arch. Insect Biochem. Physiol. 54

7-Ethoxycoumarin-O-deethylase activity is a biochemical marker linked to neonicotinoid resistance in *Bemisia tabaci*

Elevated **glutathione S-transferase** levels in abamectin resistant two-spotted spider mites, *T. urticae*

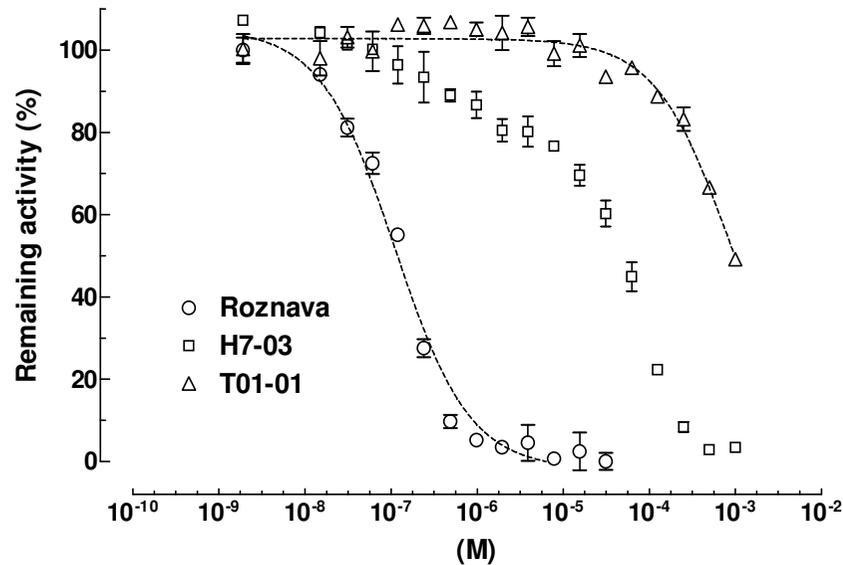


Strain NL-00 exhibited a >50-fold resistance to abamectin

| Strain | K_m (SD) μM | V_{max} (SD) mOD/min/equivalent |
|------------|-----------------------------|--------------------------------------|
| WI | 79 (3,0) | 46 (0,6) |
| GSS | 75 (5,8) | 25 (0,7) |
| NL-00 | 90 (5,5) | 57 (1,2) |
| WI | 591 (45,8) | 49 (1,1) |
| GSS | 648 (32,4) | 35 (0,5) |
| NL-00 | 669 (45,9) | 96 (2,0) |
| | μM | Fluorescence/equivalent |
| MCB | | |
| WI | 64 (9,3) | 1243 (69,2) |
| GSS | 60 (13,1) | 1016 (83,4) |
| NL-00 | 86 (9,7) | 5114 (245,3) |

Nauen & Stumpf (2002) Anal. Biochem. 303
Rauch & Nauen (2002) Pestic. Biochem. Physiol. 72

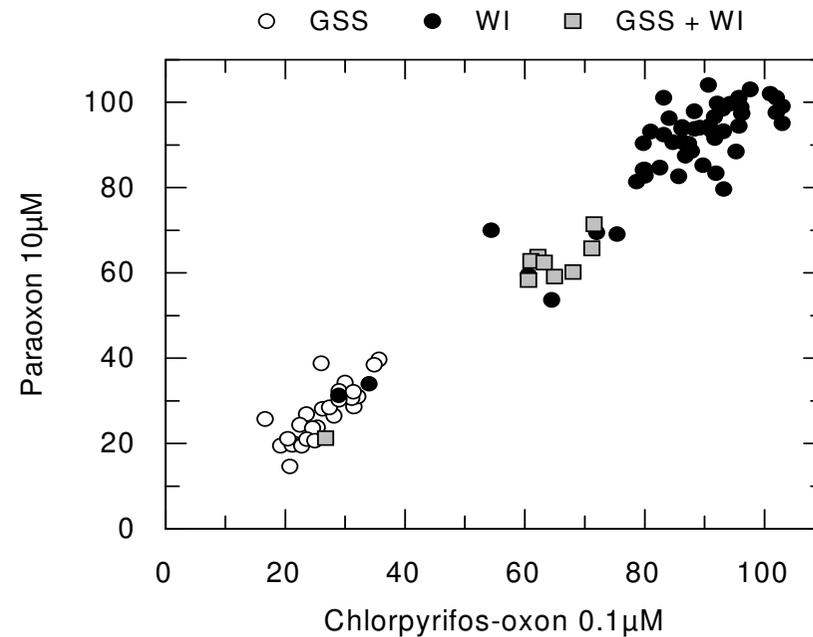
In insensitive acetylcholinesterase confers OP and carbamate resistance in several pest species



Pirimicarb resistance in *Phorodon humuli* due to MACE



OP resistance in *T. urticae* due to insensitive AChE

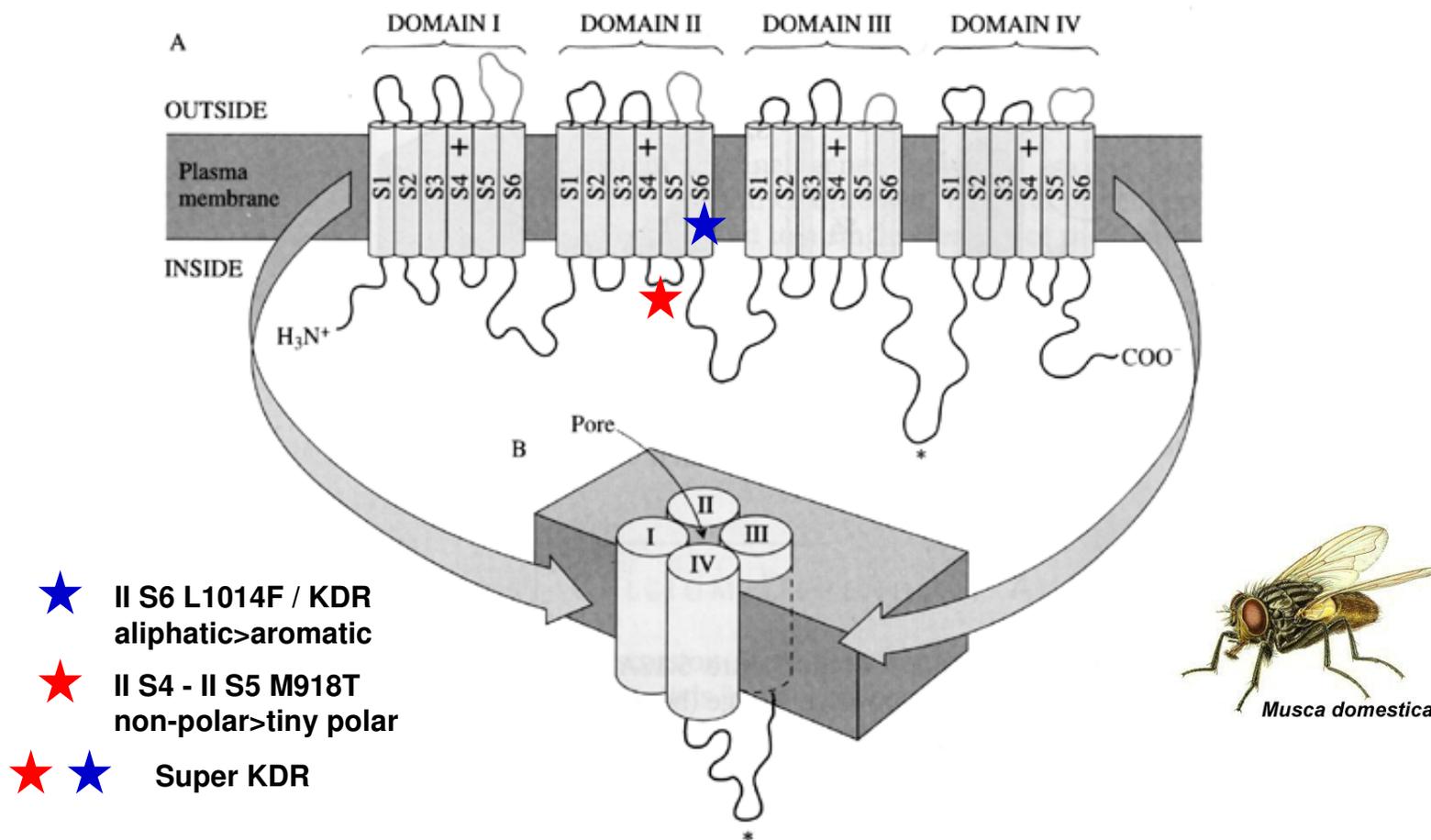


Stumpf et al. (2001) Pestic Biochem Physiol 69

Benting & Nauen (2004) Pest Manag Sci
Nauen et al. (2006) Pestic Biochem Physiol, submitted

Knock down resistance / pyrethroids and DDT

Target: voltage-gated Sodium Channel, transduction of nerve activity



Mutations of the voltage-gated sodium channel in pest insects

CTT
 CTC
 CTA
 CTG
 TTA
 TTG

Leucin = susceptible

TTT
 TTC

Phenylalanin = KDR

CAT
 CAC

Histidin = KDR
H. virescens

TCA
 TCG

Serin = KDR
Mosquitos

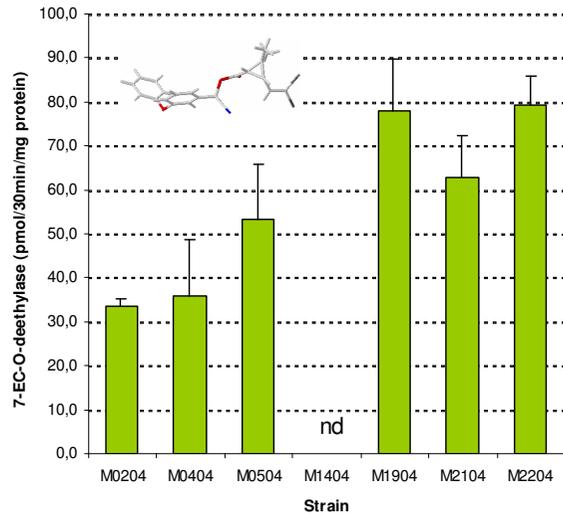
| | |
|------------------------|---|
| M. domestica (2987) | TGGGCGATGTCAGCTGTATACCCCTTCTTCTTGGCCACGGTCGTGATAGGCAATCTTGTGGTCTTAACTTTCTTAGCTT |
| B. germanica (2924) | TTGGAGACTGGTCCTGCATCCCGTTCTTCTTGGCCACTGTCGTCAATGGAAACTTGGTTGTGTGAACTCTTCTTGGCC |
| D. melanogaster (3301) | TGGGCGATGTCCTGTGCATTCCTTCTTCTTGGCCACCGTGTGTCATCGGCAATCTTGTGGTACTTAACCTTTTCTTAGCC |
| P. humuli (259) | TCGGAGAACCACGTGTATACCAATCTTCTTGGCTACTGTGTGTCATGGTAACCTCGTGGTCTCAACTCTTTCTTGGCG |
| P. xylostella (269) | TGGGGGACGTCCTGTATCCCTTCTTCTTGGCCACCGTGTGTCATGGCAACTTTGTGGTCTCAACTTGTTC----- |
| M. persicae (269) | TCGGAGAACCAACGTGTATACCAATCTTCTTGGCTACTGTGTGTCATGGTAACCTCGTGGTCTCAACCTCTTCTT----- |
| L. decemlineata (2217) | TTGGAGATGTATCCTGTATCCCAATTTCTTCTTAGCCACAGTGTGTCATGGCAATCTTGTGTGTTGAACTCTTCTTGGCC |
| A. gossypii (275) | TCGGAGAACCAACGTGTATACCAATCTTCTTGGCATCTGTGTGTCATCGGTAACCTTGTGGTACTTAATCTTTCTTGGCG |
| H. virescens (2354) | TCGGAGATGTCCTTGTATACCCCTTCTTCTTGGCTACCGTGTGTCATGGCAATCTTGTGGTACTTAACCTTTTCTTGGCC |
| B. microplus (404) | TCTCAGGCTGGCCCTGCATCCCTTCTTCTTGGCTACTGTAGTGTGTCATCGGGAACCTTGTGGTGCTCAACCTTTTCTCGCC |
| B. tabaci (275) | TTGGTGTATGTGTCCTGTATTCCTTTTCTTCTTGGCCACTGTGTTAICGGTTACCTTGTAGTTTAAATCTTTCTTAGCG |
| A. gambiae (257) | TCGGTGTATGTATCCTGCATACCAATTTCTTCTTGGCCACTGTAGTGATAGGAATTTAAGTCGTGCTTAACCTTTTCTTAGCC |
| Consensus(3330) | TCGGAGATGT TCCTGTATACC TTCTTCTTGGCCACTGT GTCATTGG AACCTTGTGGT CT AACCTTTTCTT GCC |

So what confers pyrethroid resistance in European populations of pollen beetles?



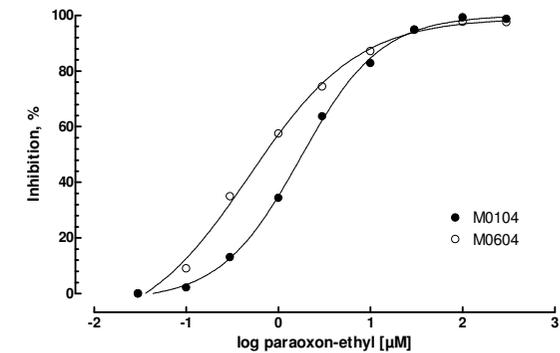
Pyrethroid-resistance pollen beetles is conferred by elevated levels of monooxygenases, but not esterases and kdr

1)

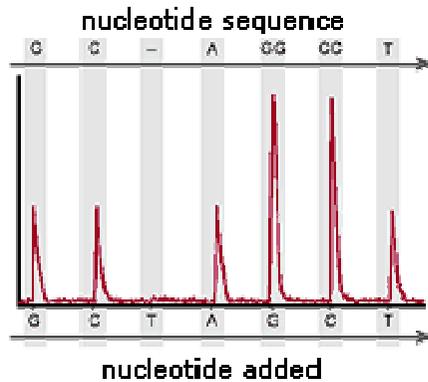


- | | | |
|----|------------------|---|
| 1) | P450 | + |
| 2) | Kdr | - |
| 3) | MACE | - |
| 4) | Esterases | - |

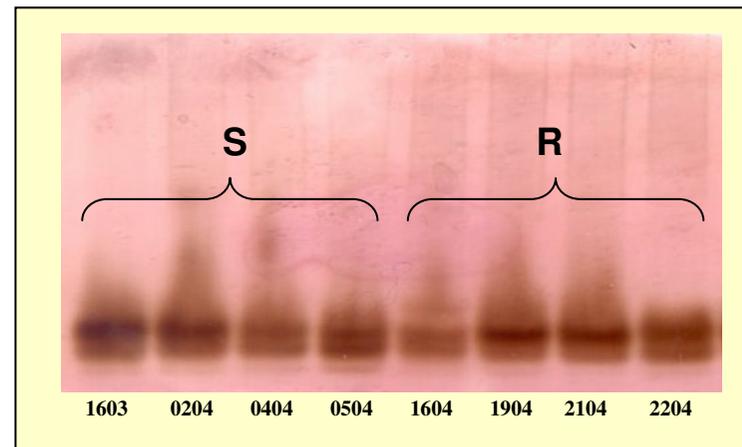
3)



2)



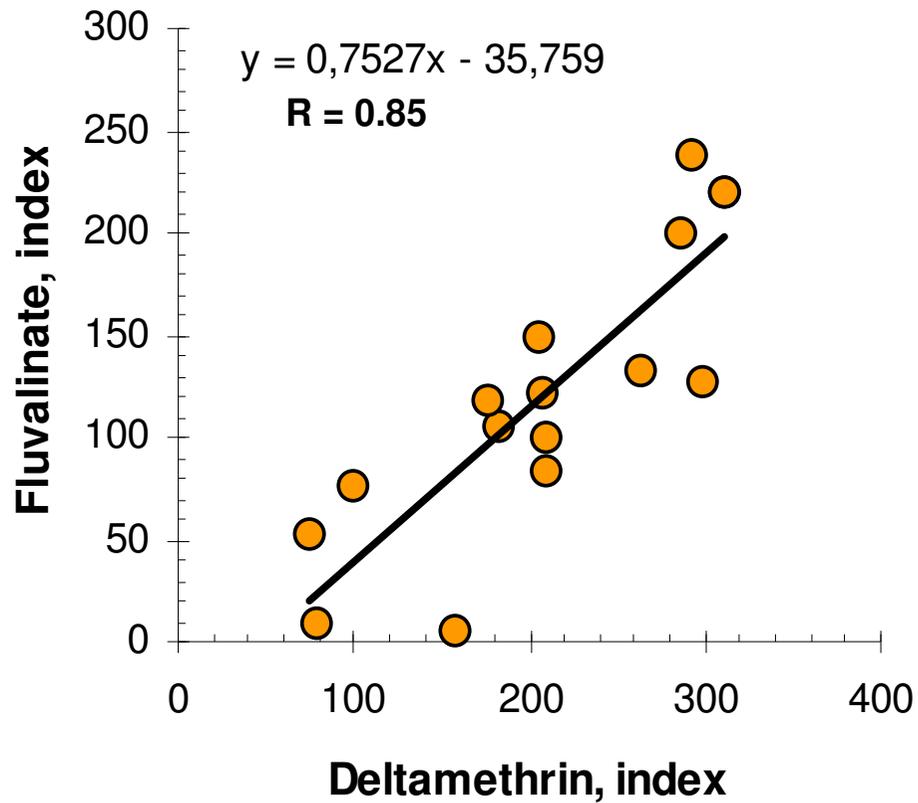
4)



Substrate: alpha-naphthylacetate (gradient PAGE)

Pyrethroid cross resistance in pollen beetles

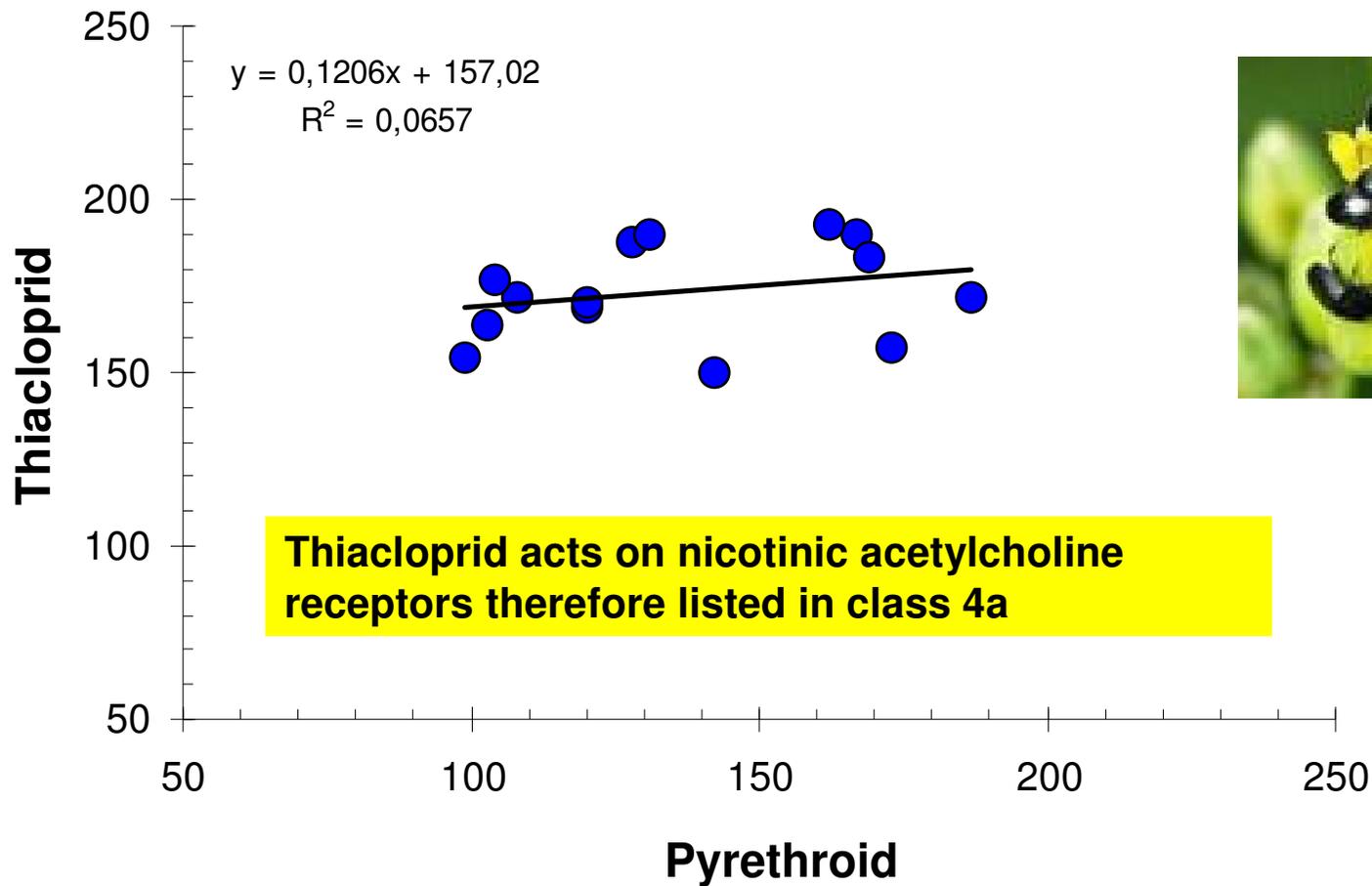
Monitoring



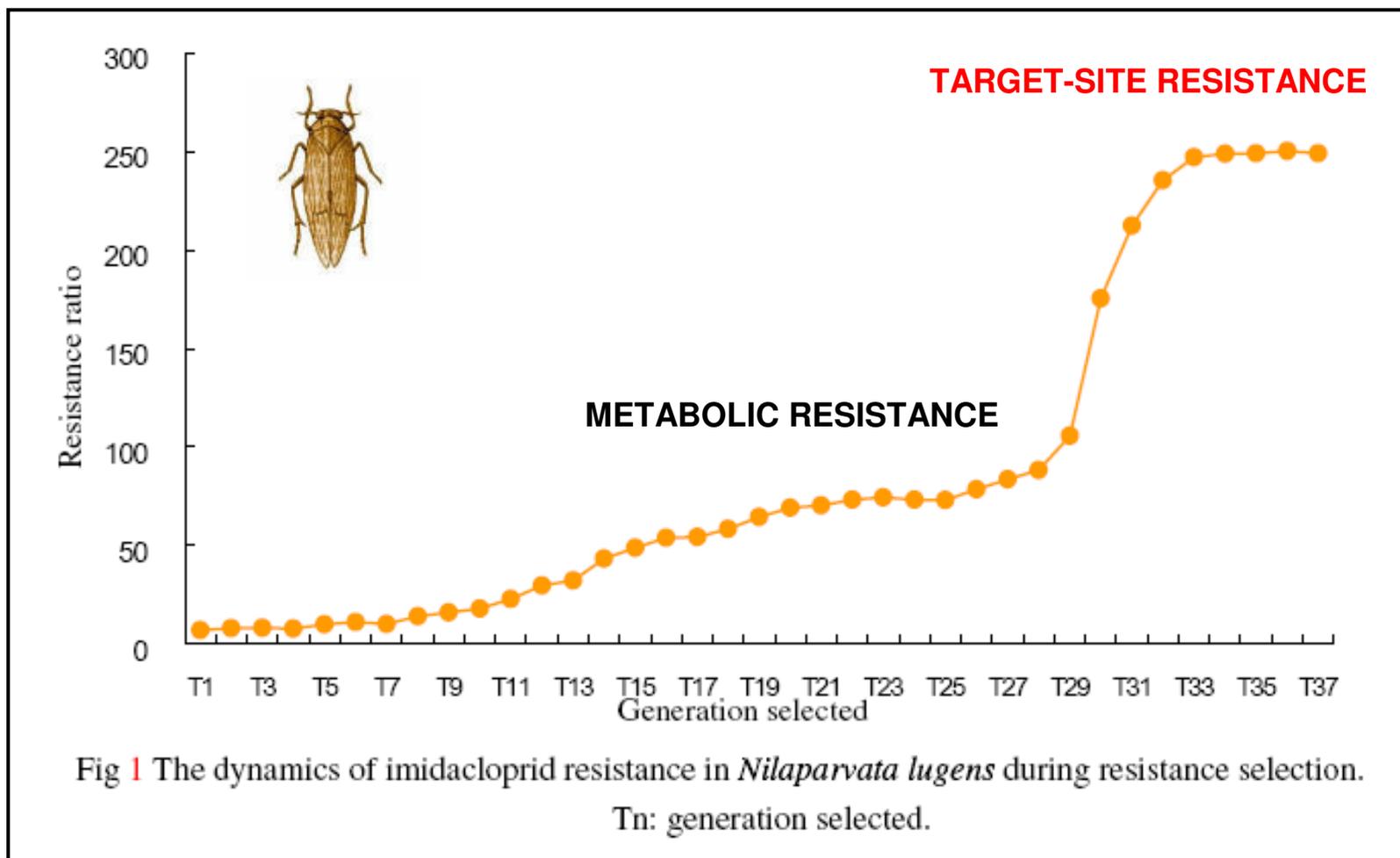
Correlation between deltamethrin and tau-fluvalinate efficacy points to pyrethroid cross-resistance in pollen beetles



No correlation between thiacloprid (IRAC class 4A insecticide) and pyrethroid (class 3 insecticides) efficacy on pollen beetles



Remember the brown planthopper example?



Liu & Han, *Pest Manag Sci*, 2006

A nicotinic acetylcholine receptor mutation conferring target-site resistance to imidacloprid in *Nilaparvata lugens* (brown planthopper)

Zewen Liu^a, Martin S. Williamson^a, Stuart J. Lansdell^b, Ian Denholm^a, Zhaojun Han^{a,b}, and Neil S. Millar^a

^aKey Laboratory of Monitoring and Management of Plant Diseases and Insects, Ministry of Agriculture, Nanjing Agricultural University, Nanjing 210095, China; ^bRothamsted Research, Harpenden, Hertfordshire AL5 2JQ, United Kingdom; and ^cDepartment of Pharmacology, University College London,

| | | | | |
|----|----|--------|---|---------|
| NI | 1S | KFGSWT | Y | DGNHVDL |
| NI | 1R | KFGSWT | S | DGNHVDL |
| NI | 3S | KFGSWT | Y | NGAQVDL |
| NI | 3R | KFGSWT | S | NGAQVDL |

↑
Y151S

review September 24, 2004
 been slow to emerge but now involves a number of insect pests (10). In the best-studied example of resistance, involving the whitefly *Bemisia tabaci*, attributable to enhanced oxidative detoxification of by overexpressed monooxygenases, rather than changes in nAChRs (11). For other major insect-both target-site modifications and enhanced de- been identified as being important resistance

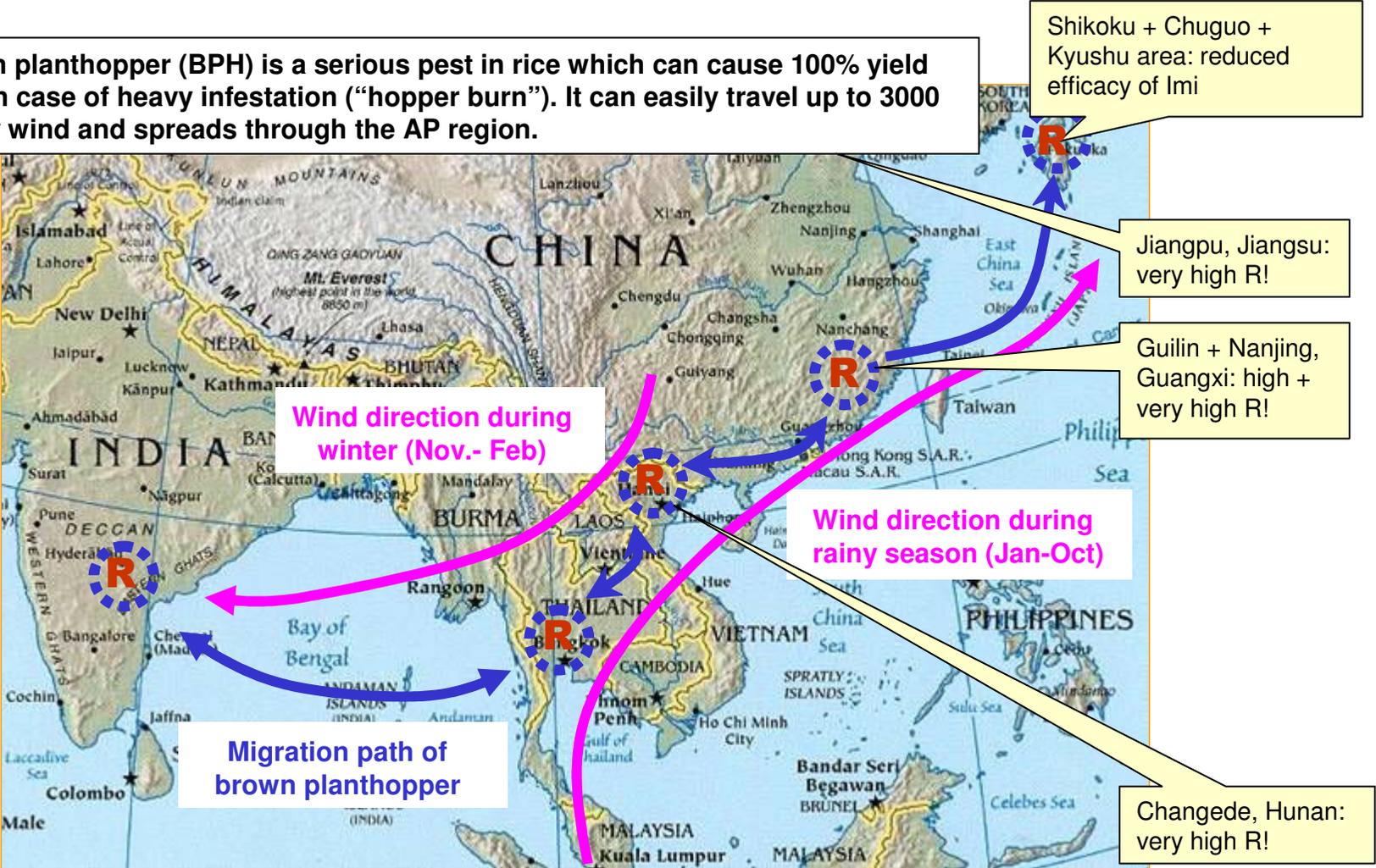
standing the molecular basis of imidacloprid resistance, five nAChR subunits (Nla1–Nla4 and Nlβ1) have been cloned from *N. lugens*. A comparison of nAChR subunit genes from imidacloprid-sensitive and imidacloprid-resistant populations has identified a single point mutation at a conserved position (Y151S) in two nAChR subunits, Nla1 and Nla3. A strong correlation between the frequency of the Y151S point mutation and the level of resistance to imidacloprid has been demonstrated by allele-specific PCR. By expression of hybrid nAChRs containing *N. lugens* α and rat β2 subunits, evidence was obtained that demonstrates that mutation Y151S is responsible for a substantial reduction in specific [³H]imidacloprid binding. This study provides direct evidence for the occurrence of target-site resistance to a neonicotinoid insecticide.



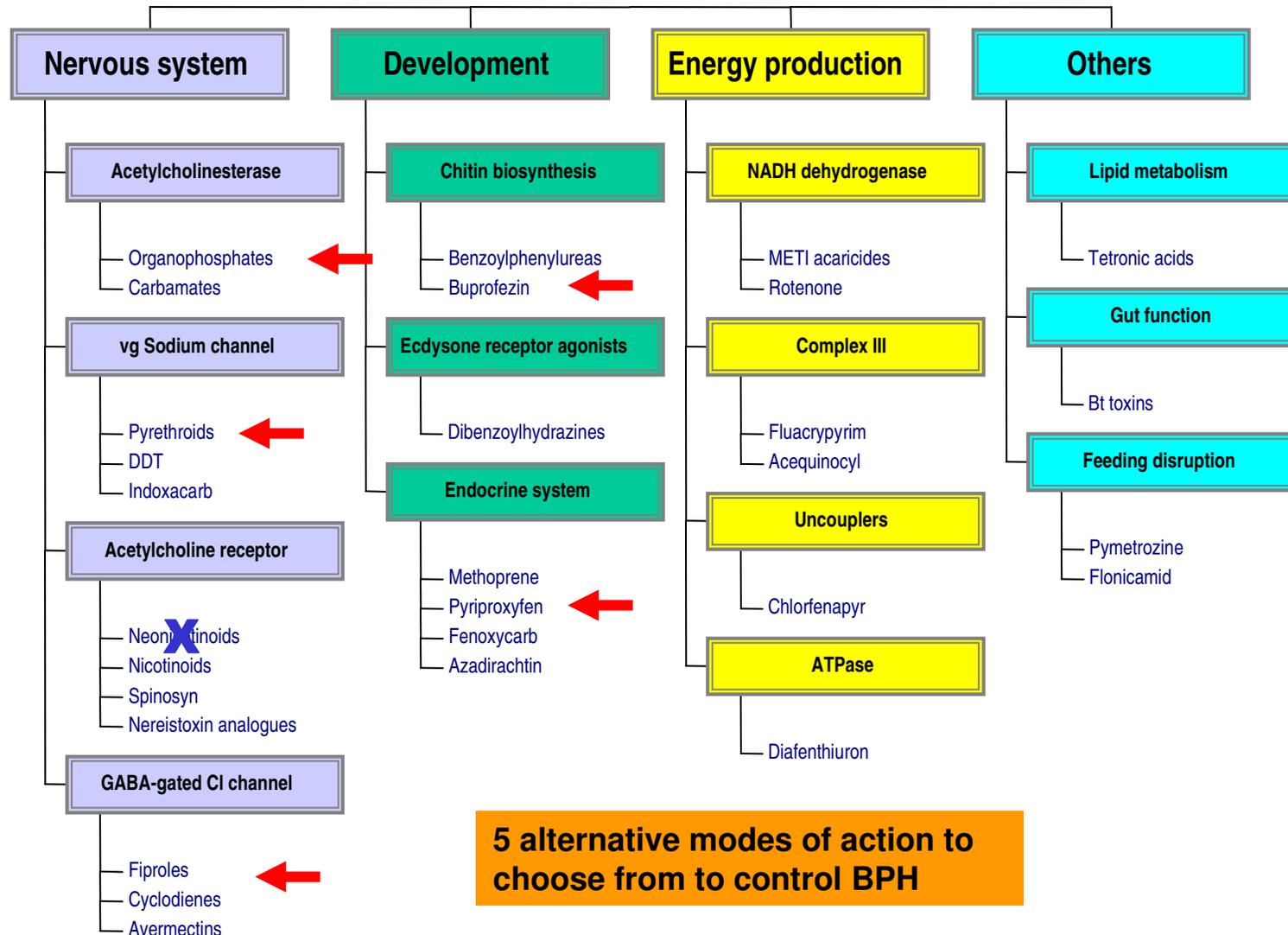
Imidacloprid-susceptible and resistant *N. lugens* strains, a mutation

Migration of BPH populations across Asia

Brown planthopper (BPH) is a serious pest in rice which can cause 100% yield loss in case of heavy infestation (“hopper burn”). It can easily travel up to 3000 km by wind and spreads through the AP region.



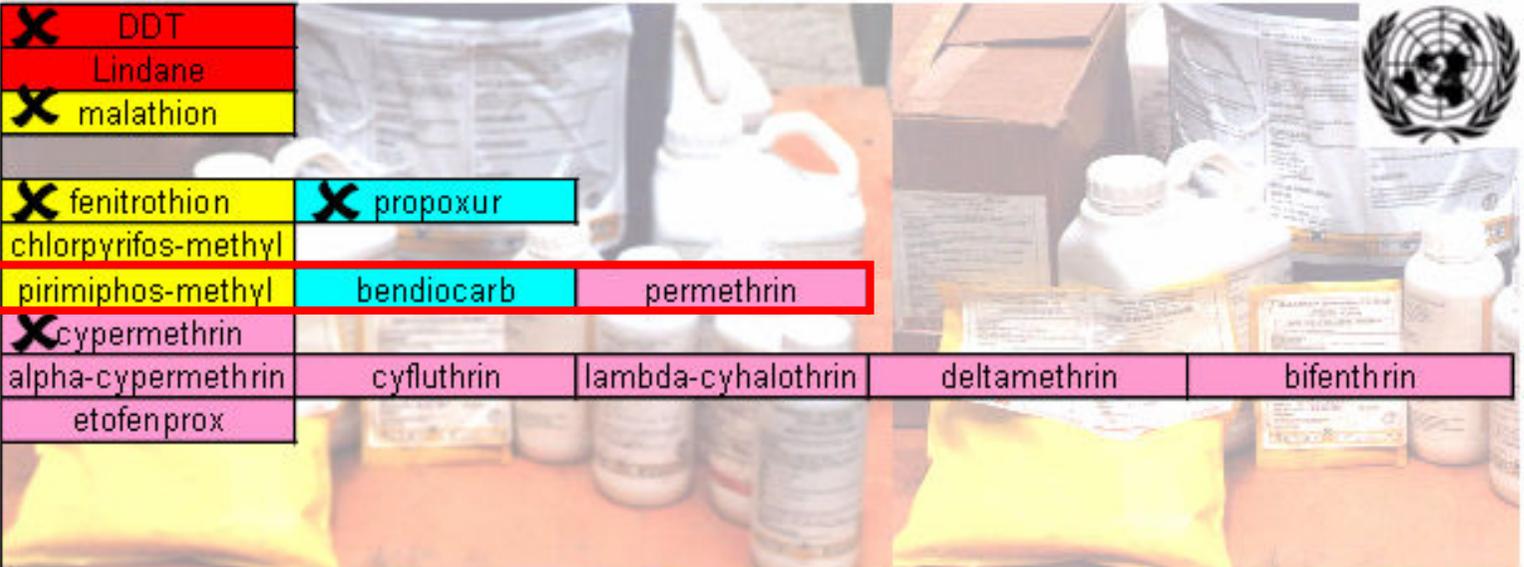
Chemical options for RM in brown planthoppers



**It is possible to manage resistance properly –
but tool box diversity is necessary**



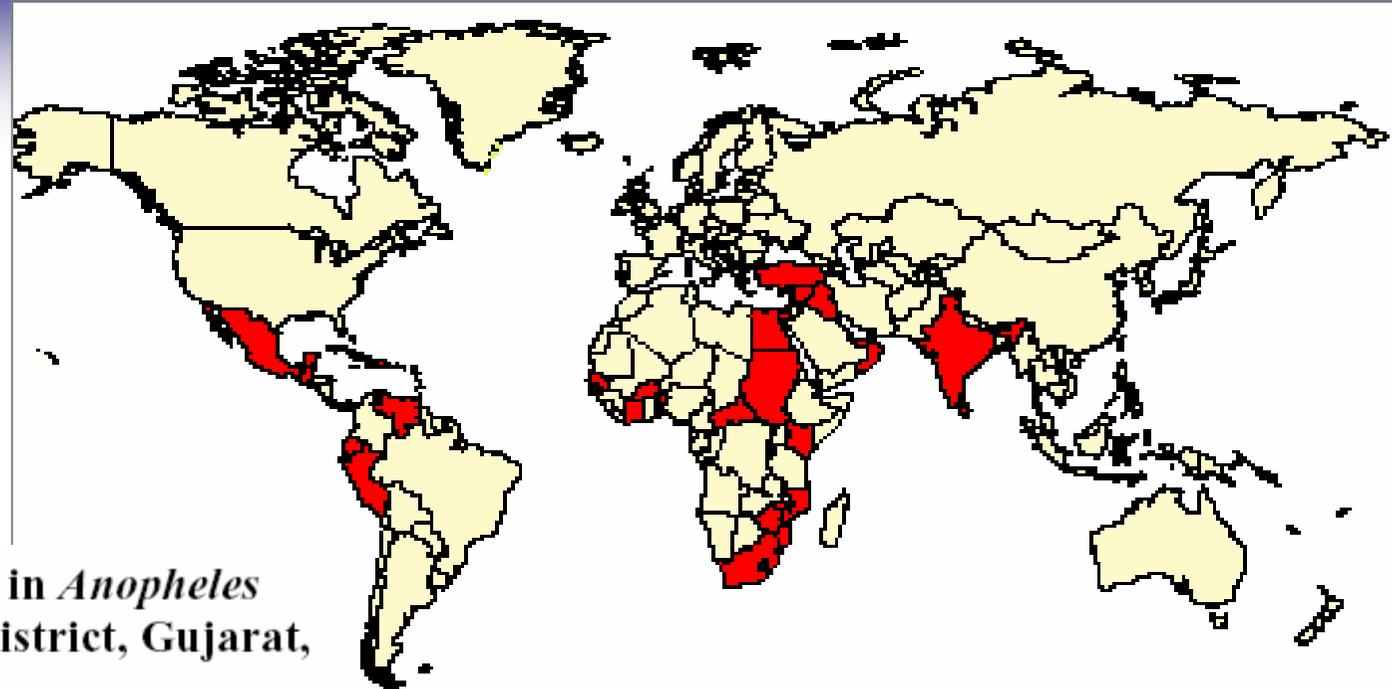
No new insecticidal mode of action class introduced for adult mosquito control since more than 30 years !



| | | | | | |
|---------|---------------------------|-----------------------|--------------------|--------------|------------|
| 1940-45 | X DDT | | | | |
| 1945-50 | Lindane | | | | |
| 1950-55 | X malathion | | | | |
| 1955-60 | | | | | |
| 1960-65 | X fenitrothion | X propoxur | | | |
| 1965-70 | chlorpyrifos-methyl | | | | |
| 1970-75 | pirimiphos-methyl | bendiocarb | permethrin | | |
| 1975-80 | X cypermethrin | | | | |
| 1980-85 | alpha-cypermethrin | cyfluthrin | lambda-cyhalothrin | deltamethrin | bifenthrin |
| 1985-90 | etofenprox | | | | |
| 1990-95 | | | | | |
| 1995-00 | | | | | |
| 2000-05 | | | | | |

Loss of insecticides due to:

1. Resistance issues
2. Regulatory hurdles
3. Economic reasons



TRENDS in Parasitology

Pyrethroid resistance in *Anopheles culicifacies* in Surat district, Gujarat, west India

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Malaria Research Centre (ICMR), 22 Sham Nath Marg, Delhi 110 054, India. December 2001.

A focus of deltamethrin resistance in *Anopheles culicifacies*, the major vector of malaria in India, was identified in Surat district, Gujarat, western coast of India, where two synthetic pyrethroids, deltamethrin and cyfluthrin are being used under the public health programme since 1996, as a selective vector control measure. The per cent mortalities in *An. culicifacies* after one-hour exposure to 0.05% deltamethrin varied from 60 to 78.

The development of pyrethroid resistance in *An. culicifacies* is of great concern to the malaria control programme because SPs are being used in public health programmes to control multiple-resistant vectors and tackle epidemic outbreaks. Also, this is the only group of insecticides currently used for bed-net impregnation for malaria control.

Major biochemical mechanisms conferring resistance to important classes of insecticides in adult mosquitoes



| | Biochemical mechanism of resistance | | | | |
|------------------|-------------------------------------|----------------|--------------------|-------------|------|
| | Metabolic | | | Target-site | |
| | Esterases | Monooxygenases | GSH S-Transferases | kdr | MACE |
| Pyrethroids | ● | ● | | ● | |
| DDT | | ● | ● | ● | |
| Carbamates | ● | | | | ● |
| Organophosphates | ● | ● | | | ● |

WHO urged for new chemicals

Alternative insecticides: an urgent need

WHO investigation in all the Pesticide Industries in 2001: No new compounds collected

Morteza Zaim and Pierre Guillet

Most insecticides used against pests and vectors of human disease (e.g. fleas, flies and mosquitoes) are spin-offs from agrochemical research and development. The arsenal of safe and cost-effective public health insecticides is being depleted by restrictions for various reasons (e.g. insecticide resistance, unacceptable side effects and non re-registration) and the number of new products launched is dwindling. Mobilizing public resources and establishment of partnerships to support research and development of public health insecticides is crucial in the post-DDT and post-pyrethroid era.

Thanks for your attention!

Time for coffee...

