

# A National Surveillance System for West Nile Virus in Zoological Institutions

Dominic Travis, Lincoln Park Zoo

Tracey McNamara, Wildlife Conservation Society

Amy Glaser, Animal Health Diagnostic Lab, Cornell University

Roy Campbell, DVBID, CDC

Duane Gubler, DVBID, CDC

# Background

- June 2001 meeting
  - The National West Nile virus Zoological Surveillance system working group
- Guidelines: *Surveillance for West Nile Virus in Zoological Institutions*
- One-year pilot study for national zoological surveillance (Aug '01 – Aug '02)
- Objectives
  - Affordable/reliable testing for zoos
  - Novel data source for national surveillance
  - Increase relationships btw PH and Zoos/Vet Med

# Sampling Scheme

## Phase I:

- Begun August 2001 – current
- Solicited tissue/blood samples from **ill/dead** at-risk animals found on zoo grounds (captive/local wildlife)
- At-risk animals = any animal (any taxa) housed outdoors and at risk of mosquito exposure

# Sampling Scheme (cont.)

## Phase II:

- **Serosurvey of at-risk animals**
  - Entire country w/emphasis on emerging region
- Archived serum/plasma 6 months prior to first known positive in each state
- Opportunistically collected blood samples from **healthy** animals
  - Seropositive follow-up to evaluate long term sequelae of WNV exposure
  - Data on varying susceptibilities and rates of WNV seroconversion in zoo species and indigenous wildlife.

# Diagnostics

- Animal Health Diagnostic Laboratory at Cornell University, College of Veterinary Medicine performs all tests required for the project.
- Tissues are tested by RT-PCR and virus isolation
- Serum and plasma tested for virus and/or antibody, depending on history

# Looking for Virus

- RT-PCR for WNV
  - Positives confirmed with virus isolation or second independent RT-PCR
- All tissues processed for virus isolation
  - Identification of all viruses observed
    - Avian reovirus

# Looking for Antibody

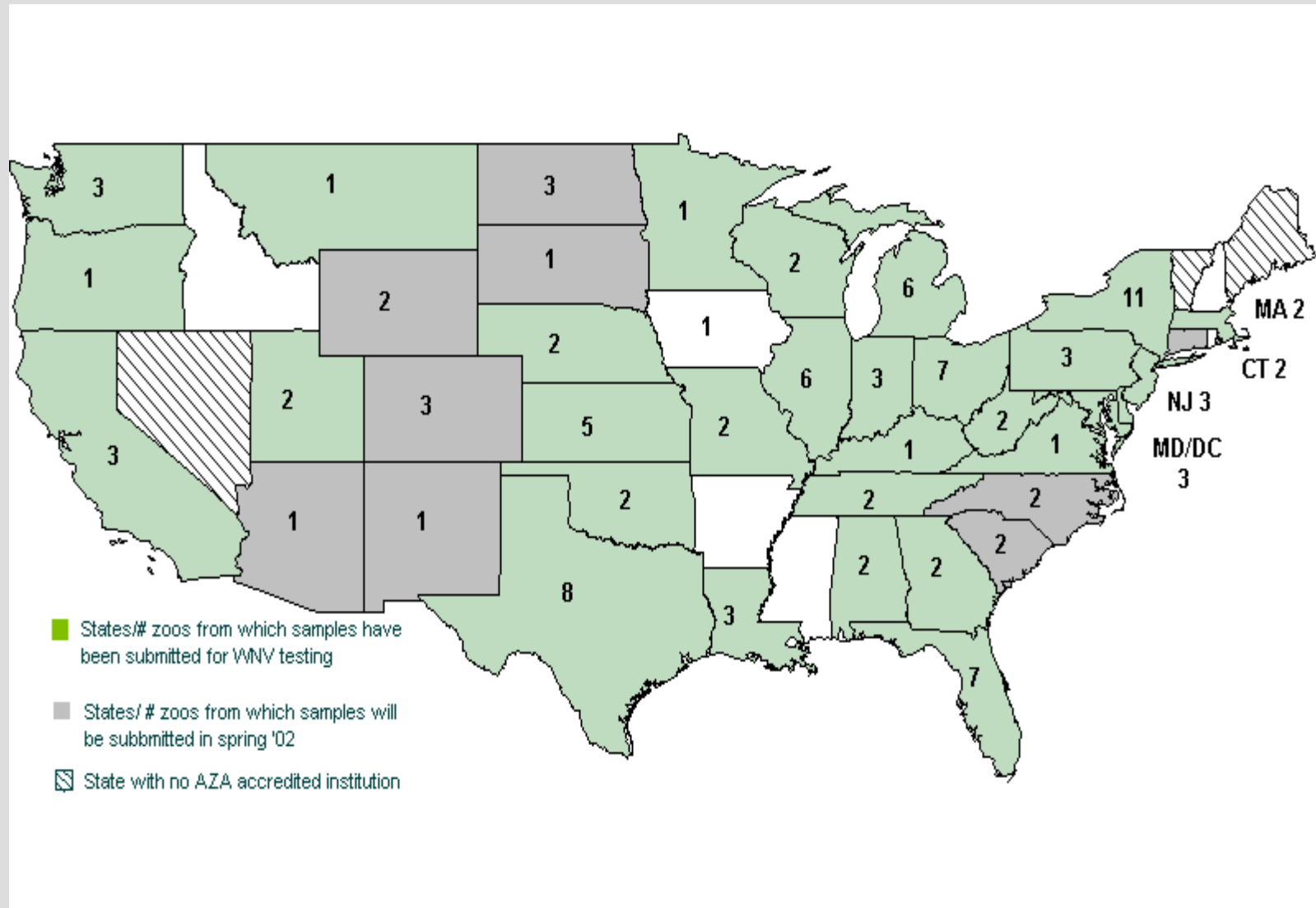
- Screen serum or plasma at 1:40 by PRNT
- Positives are titrated to a dilution of 1:640 against WNV and SLE

# Results Aug 1, 2001 – Feb 28, 2002

- Over 1,450 animals tested
  - 967 birds (195 species)
  - 40 equids (8 species)
  - 436 other mammals (110 species)
  - 20 reptiles (6 species)
- 64 participating institutions ( 20 more will submit in spring '02)
- 30 states including the District of Colombia



# Sample Distribution



State	Avian	Equine	Other Mammal	Herptiles	Total	% of Total
AL	6	0	0	0	6	0.4
CA	28	0	12	0	40	2.8
DC	46	0	1	0	47	3.3
DE	0	0	1	0	1	0.1
FL	93	0	6	5	104	7.2
GA	98	0	26	5	129	8.9
IL	14	0	4	2	20	1.4
IN	28	0	19	0	47	3.3
KS	64	0	0	0	64	4.4
KY	12	0	0	0	12	0.8
LA	34	0	15	0	49	3.4
MA	75	4	48	0	127	8.8
MD	9	1	4	1	15	1.0
MI	1	0	0	0	1	0.1
MN	12	0	0	0	12	0.8
MO	38	0	26	0	64	4.4
MT	1	0	0	0	1	0.1
NE	29	0	2	0	31	2.1
NY	171	2	78	3	254	17.6
OH	25	1	39	4	69	4.8
OK	4	0	0	0	4	0.3
OR	4	0	0	0	4	0.3
PA	74	0	67	0	141	9.8
TN	13	7	18	0	38	2.6
TX	47	8	60	0	115	8.0
UT	7	0	0	0	7	0.5
VA	1	0	0	0	1	0.1
WA	1	0	0	0	1	0.1
WI	27	0	10	0	37	2.6
WV	5	0	0	0	5	0.3
SUM	967	23	436	20	1446	100.0
% of total	66.9	1.6	30.2	1.4	100.0	

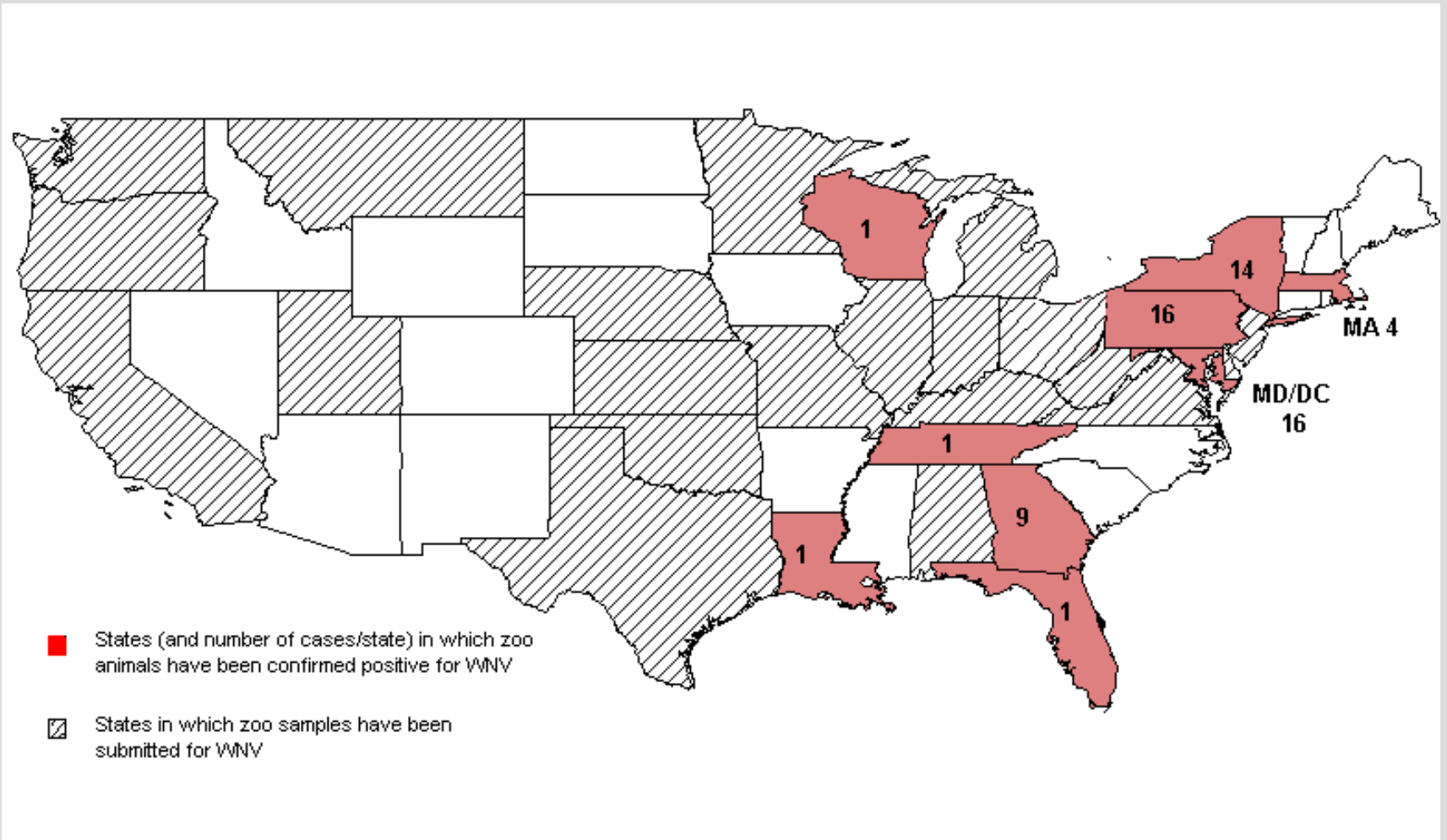
# Confirmed Results

- 30 WNV antibody-positive birds (1 wild, 29 captive)
- 29 WNV virus-positive birds (16 wild, 13 captive)
- 1 WNV antibody-positive reptile (captive crocodile monitor)
- 1 captive bird was antibody-positive for St. Louis encephalitis virus (SLE) in GA
- 13 animals (10 avian, 2 mammal, 1 reptile) screened antibody-positive and are currently undergoing confirmatory testing.

# Distribution of WNV Positives

- Antibody positive animals - zoos from 9 states
  - FL, GA, LA, MA, MD/DC, NY, PA, TN, WI
- Virus-positive animals - zoos from 4 states
  - MD/DC, GA, NY, PA
- All (antibody and virus) were from known WNV-endemic areas in the United States
- No positive zoo sample has predated the first positive predated the first event in any area

# Confirmed Positive Distribution



State	Spp.	W/D	WNV Pos type	End point
DC/MD	1 Crow	W	Antibody	160
DC/MD	12 Crow	W	Virus	
DC/MD	1 Crocodile Monitor	D	Antibody	$\geq 640$
DC/MD	1 Crow	W	Antibody	20
DC/MD	1 Magpie	D	Virus	
FL	1 Crowned crane	D	Antibody	40
GA	4 Chilean Flamingo	D	Virus	
GA	1 Tawny Owl	D	Virus	
GA	1 Turaco	D	Antibody	$\geq 640$
GA	2 Wattled Crane	D	Antibody	320
GA	1 Wattled Curassow	D	Antibody	320
LA	1 Marabou Stork	D	Antibody	320
MA	1 Bald Eagle	D	Antibody	$\geq 640$
MA	1 Barnacle Goose	D	Antibody	320
MA	1 Saddle Billed Stork	D	Antibody	$\geq 640$

State	Spp.	W/D	WNV Pos type	End point
NY	1 American Kestrel	D	Virus	
NY	1 Chilean Flamingo	D	Virus	
NY	3 Crow	W	Virus	
NY	1 Flamingo	D	Antibody	80
NY	1 Golden Eagle	D	Antibody	360
NY	2 Pelican	D	Antibody	80
NY	3 Penguin	D	Virus	
NY	1 Snowy Owl	D	Antibody	40
NY	1 Snowy Owl	D	Virus	
PA	1 Bald Eagle	D	Antibody	320
PA	2 Flamingo	D	Antibody	40, >=640
PA	1 Goose spp.	D	Antibody	40
PA	1 Greater Magellan goose	D	Antibody	80
PA	2 Humboldt Penguin	D	Antibody	320
PA	9 Humboldt Penguin	D	Antibody	>=640
TN	West African Crowned Crane	D	Antibody	>=640
WI	Demoiselle crane	D	Antibody	>=640

# Clinical Illness

- 6 (21%) WNV virus-positive captive animals exhibited clinical signs of illness
  - 23 (79%) were found dead in exhibits or on zoo grounds.
- 6 (20%) WNV antibody-positive animals presented with neurologic clinical signs (including one wild crow and the crocodile monitor)
  - 24 (80%) were found through serosurvey.



# Inconclusive Results

- 7 captive animals (6 birds, 1 African wild dog) were WNV antibody-positive upon screening but could not be conclusively classified as WNV or SLE upon end-point titration
- 4 captive birds WNV virus-positive by PCR were not confirmed by virus isolation
- 1 captive bird that was WNV antibody-negative but WNV virus-positive

# Other Interesting Findings

- 6 wild crows found culture-positive for an unidentified virus of the family Reoviridae
- 6 birds were shipped to one institution from Africa with weakly positive antibody titers to WNV found in serum samples archived during quarantine.

# Reporting Structure

- Results reported to submitting institution and central zoo database
- Submitting institutions required to report to local PH
  - Local PH code and incorporate data into national system
- Validation survey in progress
  - 30 institutions validated

# Conclusions

- Phase I and II have:
  - Created/strengthened relationships between zoos and local/state health officials for the detection/reporting of a zoonotic disease threat.
  - Provided data to the public health system
  - Given zoos an avenue for testing endangered species
- Future Directions:
  - This project created a framework that may be applied to other biologic threats of concern.
  - Regional diagnostic centers
  - Vaccination

# Working Group Members

- **Bruce Akey**, VA Dept. Agriculture
- **Wilbur Amand**, American Association of Zoo Veterinarians
- **Robyn Barbiers**, Lincoln Park Zoo
- **Bobby Brown**, CDC
- **Grant Campbell**, CDC, DVBID
- **Pamela Diaz**, Chicago Department of Public Health
- **Cindy Driscoll**, MD Dept. Nat. Res.
- **Ed DuBovi**, Cornell College of Veterinary Medicine
- **Millicent Eidson**, NY State Dept. of Health
- **Amy Glaser**, Cornell College of Veterinary Medicine
- **Thomas Gomez**, USDA, APHIS, VS
- **Duane Gubler**, CDC, DVBID
- **Nicholas Komar**, CDC, DVBID
- **Laura Kramer**, NY State Dept. of Health
- **Bob McLean**, USGS, National Wildlife Health Center
- **Rita McManamon**, Zoo Atlanta
- **Tracey McNamara**, Wildlife Conservation Society
- **Hayley Murphy**, Zoo New England
- **Eileen Ostlund**, USDA, NVSL
- **Mary Grace Stobierski**, MI Dept. of Community Health
- **Scott Terrell**, Disney's Animal Kingdom
- **Kristin Vehrs**, American Zoo and Aquarium Association